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**Understanding wound healing in obesity**

Cotterell A *et al*. Wound healing in obesity

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**Abstract**

Obesity has become more prevalent in the global population. It is associated with the development of several diseases including diabetes mellitus, coronary heart disease, and metabolic syndrome. There are a multitude of factors impacted by obesity that may contribute to poor wound healing outcomes. With millions worldwide classified as obese, it is imperative to understand wound healing in these patients. Despite advances in the understanding of wound healing in both healthy and diabetic populations, much is unknown about wound healing in obese patients. This review examines the impact of obesity on wound healing and several animal models that may be used to broaden our understanding in this area. As a growing portion of the population identifies as obese, understanding the underlying mechanisms and how to overcome poor wound healing is of the utmost importance.

**Key Words:** Obesity; Wound healing; Adipokines; Tissue fibrosis; Diabetes; Preclinical animal models; Hypertrophic skin scarring; Wound tension; Metabolic syndrome

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**Core Tip:** Obesity induces a chronic low-grade inflammatory state through increased release of adipokines, cytokines, and chemokines from excess adipose tissue. The chronic low-grade inflammation is thought to contribute to a dampened immune response during the inflammatory phase of wound healing leading to delayed wound healing. While there are several animal models used to study wound healing, they have not been widely applied to studying the effects of obesity on wound healing leading to a gap in the literature on this topic.

**INTRODUCTION**

Obesity has become more common over the past 40 years, with approximately 33% of the population being classified as overweight or obese[1]. In adults, obesity is defined as a body mass index of 30.0 kg/m2 or greater[2-4]. Obesity accounts for up to 7% of total healthcare costs in developed nations, classifying it as a significant expenditure of national healthcare budgets[3]. Obesity is associated with the development of several diseases including diabetes mellitus, coronary heart disease, hypertension, and certain forms of cancer, and has been associated with a decreased lifetime expectancy[2-4]. Delayed wound healing seen in patients with diabetes mellitus, commonly associated with obesity, can be attributed to changes in the macro- and microvasculature, decreased production of growth factors, and poor quality of granulation tissue[5,6]. Impaired wound healing can be caused by changes in the four phases of wound healing – hemostasis, inflammation, proliferation, and remodeling (Figure 1).

Due to the significant rise in obesity worldwide, it is important to understand the role that that this disease may have on wound healing. The purpose of this review is to highlight the impact of obesity on cutaneous wound healing and discuss future studies that must be performed to advance our understanding of the subject. We also aim to review the phases of wound healing and how they are impacted in obese states.

***Physiologic changes in the human body due to obesity***

There are several physiologic changes that occur in the body as the result of obesity. In the respiratory system, impaired diaphragmatic relaxation and chest expansion due to additional adipose tissue result in hyperventilation as well as decreased vital capacity and tidal volumes[7,8]. Fibroblasts, which require partial pressure of oxygen greater than 15 mmHg for proper function, are unable to produce collagen in the wound edges in obese patients where the pressure of arterial oxygen (PaO2) is near 0 mmHg[8]. In the cardiovascular system, there is excess workload on the heart to supply oxygen to all tissues of the body. Patients with longstanding obesity may eventually develop heart failure, resulting in decreased cardiac output, reduced blood volume, and impaired circulation. It is known that adipose tissue is not well vascularized and is more susceptible to ischemia and hypoxia when compared to the epidermis[7]. It is important to explore these changes in vascularity and asses how they may contribute to wound healing outcomes.

***Changes in vascularity***

In addition to alterations in respiratory physiology and cardiac function, macro- and microvasulature is also impacted in the obese state. It is well documented that adipose tissue has decreased vascularity when compared to other tissues in the body[7,9]. This decrease in vascularity may contribute to the poor wound healing outcomes seen in this population. The increase in adipose tissue is also negatively correlated with angiogenesis[9]. Glucocorticoids have been well documented as inhibitors of angiogenesis. Elevated levels of 11β-hydroxysteroid dehydrogenase type 1, a glucocorticoid-amplifying enzyme, has been associated with obesity[9]. In addition to affecting angiogenesis, glucocorticoids have also been associated with altering immune responses.

***Changes in immune responses***

It is crucial to consider changes in immune function, as alterations at baseline may have significant impact on homeostasis. A number of studies have been published elucidating the connections between obesity and a pro-inflammatory state[10]. These alterations in immune function are mediated by adipocyte hypertrophy in conjunction with infiltration of pro-inflammatory cell types including CD8+ T-cells, CD4+ T-cells, and M1 macrophages[10]. In addition to changes in immune cell activation, elevated levels of glucocorticoids are seen in both obese humans and mice, likely due to increased stress on the body[11]. This chronic stress on the body is also associated with chronic low-grade inflammation[9]. Studies in obese mice have demonstrated progressive increases in proinflammatory cytokines due to activation of invariant natural killer T cells by excess lipids[9]. Elevated cytokines include tumor necrosis factor alpha (TNF-α), leptin, interleukin (IL)-6, and transforming growth factor beta (TGF-β) which are all involved at varying stages of the wound healing process[9].

**PHASES OF WOUND HEALING**

Given the physiologic changes seen in obesity, it is possible that they may have direct impact on the wound healing process. To understand possible mechanisms of interaction, one must be familiar with the wound healing process under normal physiologic conditions. Briefly, the first phase of wound healing, hemostasis, occurs when there is damage to endothelial cells. Following hemostasis, the inflammatory phase begins in which there is edema and an influx of inflammatory cells. The inflammatory phase is then followed by both proliferation and remodeling, the latter of which can take place for over two years[12].

***Hemostasis***

Hemostasis, the first phase of wound healing, occurs due to endothelial damage and can last from minutes to hours. Immediately upon injury, damaged blood vessels will vasoconstrict and both the intrinsic and extrinsic coagulation cascades are activated by nearby platelets and endothelial cells[13]. The thrombus that forms is rich in platelets, collagen, fibronectin, and thrombin; it serves a scaffold for neutrophils, monocytes, and other invading cells leading to the release of cytokines, growth factors, and local vasoconstrictors such as serotonin[13]. Following hemostasis, release of histamine induces migration of inflammatory cells to the site of injury, thus marking the beginning of the inflammatory phase.

***The inflammatory phase***

The inflammatory phase generally begins shortly after initial injury and the hemostasis phase[14]. The inflammatory phase is critical to the wound healing process as it marks recruitment of the innate immune system. Within the first 48 to 96 h after injury, monocytes are recruited from the surrounding tissue and transform into macrophages[13]. These activated macrophages are necessary for the transition from the inflammatory phase to the proliferative phase. Macrophages release vascular endothelial growth factor, fibroblast growth factor, and TNF-α to stimulate angiogenesis in addition to TGF-β, epidermal growth factor, and platelet-derived growth factor for fibroplasia[13]. Neutrophils, the predominant cell type during this phase, are recruited to the site of injury via IL-8 released from platelets during degranulation, and secrete IL-1, TNF-α, and TGF-β[13]. In the skin following injury, toll-like receptors are expressed on host cells leading to the activation of two distinct pathways-the nuclear factor kappa beta and mitogen-activated protein kinase pathways. The activation of these pathways is the hallmark of the inflammatory phase[15]. As the inflammatory phase resolves, the body begins the proliferative phase. Healing may begin only after the inflammatory phase is done[16].

***The proliferative phase***

During the proliferative phase, the body prioritizes restoring the local vascular network and re-epithelializing the wound surface[15]. Keratinocytes begin migrating from the edges of the wound bed while epithelial stem cells begin proliferating in reactions influenced by both chemical and mechanical signals from both anti- and pro-inflammatory cells, inducing fibroplasia. During fibroplasia, fibroblasts become activated, transition to myofibroblasts and secrete components of the extracellular matrix that promote wound contraction, contributing to formation of a persistent scar[17]. These signals lead to the development of granulation tissue, which is largely comprised of collagen III, new blood vessels, and fibroblasts. Fibroblasts are the predominant cell type in granulation tissue and respond to cytokines released from macrophages to induce re-epithelialization[18]. Fibroblasts release keratinocyte growth factor 1 and 2 in addition to IL-6, and these cytokines stimulate local keratinocytes to migrate, proliferate, and differentiate in the wound bed[13]. Research has shown that wounds deficient in IL-6 have decreased collagen deposition, epithelialization, and angiogenesis[18]. Given the intricacies involved in the four phases of wound healing, it is important to note how these phases of wound healing are affected by obesity.

**FACTORS AFFECTING WOUND HEALING IN OBESITY**

There are several factors that may contribute to poor wound healing found in obese populations (Figure 2). Patients with obesity are in a persistent inflammatory state; because of this, these patients have a prolonged inflammatory phase contributing to poor wound healing outcomes (Figure 3)[16].

***Adipokines***

Adipokines are cytokines produced by adipose tissue that affect metabolism, reproduction, and satiety. Currently known adipokines include leptin, adiponectin, and resistin[19]. Leptin, the most studied, regulates food intake and energy expenditure via the central nervous system. It has been shown to be effective in improving metabolic dysfunction in patients with either lipodystrophy or congenital leptin deficiencies[20,21]. Leptin has also been shown to be structurally similar to IL-2 and growth hormone 1, suggesting that it may have pro-inflammatory activity; it has been shown to increase the production of TNF-α and IL-6 by monocytes[22]. Interestingly, leptin levels also increase in serum in response to pro-inflammatory stimuli including TNF-α[22].

Leptin is not the only proinflammatory adipokine that has been extensively studied in recent decades. Resistin, another proinflammatory adipokine, has been shown to induce insulin resistance in mice. However, it is unclear if these effects exist in humans as well[21]. In mice, it has been shown that deficiencies of this adipokine in ob/ob mice lead to increase obesity despite improved glucose tolerance and insulin sensitivity[23].

In addition to pro-inflammatory adipokines, there are also anti-inflammatory adipokines of which adiponectin is one that has been relatively well studied. Adiponectin, almost exclusively produced and secreted by adipocytes, has become well studied for its anti-inflammatory, anti-apoptotic, and insulin-sensing properties[21]. It has been shown to protect against several disorders associated with obesity; adiponectin expression has also been found to have decreased levels in patients with obesity[18,21]. In addition to the impact of adipokines on wound healing, cytokines and chemokines have also been implicated in the pathogenesis of chronic wounds in obese and pre-diabetic populations.

***Cytokines and chemokines***

Adipose tissue is also an important source of cytokines, and there are many cytokines and chemokines that influence wound healing in obesity. Proinflammatory cytokines including monocyte chemotactic protein-1, TNF-α, IL-1, IL-6, and IL-8, are notably increased in obesity (Table 1)[21,24,25]. Patients and mice with increased percentages of adipose tissue produce more of these pro-inflammatory cytokines at baseline leading to a state of chronic low-grade inflammation. Alterations in serum levels of these cytokines, chemokines, and adipokines may drive the underlying physiologic processes that contribute to poor wound healing outcomes in obese patients.

***Wound tension, tissue pressure, and hematoma formation***

In addition to chemical mediators impacting wound healing, a number of mechanical forces also impact wound healing in obesity. Due to excess adipose tissue, there is increased tension on wounds; this increased tension is frequently associated with hypertrophic scarring and stretched scars[16,26]. Increased tissue pressure is also seen due to excess adipose tissue, both in unwounded and wounded cutaneous skin[16,27]. This increased pressure is associated with reduced perfusion and vascularity[27]. Furthermore, hematoma formation is a complication frequently seen in patients due to excess tissue pressure. Given the multitude of complications associated with obesity, it is imperative to understand what impact obesity plays in the wound healing process.

**OBESITY AND IMPACT ON WOUND HEALING**

As previously discussed, patients with obesity have persistently elevated levels of pro-inflammatory cytokines in serum. This elevated immune response at baseline may likely cause a dampened immune response during the inflammatory phase of wound healing resulting in the chronic, slow-healing wounds frequently seen in this population.

**ANIMAL MODELS USED TO STUDY WOUND HEALING**

To study wound healing, it is essential to create animal models that mimic wound healing in human skin. There are three main models to study this process – the hypertrophic wound model, the wound-induced hair follicle neogenesis (WIHN) model, and the excisional wound model (Figure 4). Although many animal species are used to study healing, this review will only discuss advantages and disadvantages of murine models. Mice and rats are popular for these animal models as they are widely available and relatively inexpensive. However, skin in these animals contain myofibroblasts, which allow their wounds to heal *via* contraction rather than through re-epithelialization and granulation as seen in human skin.

***Hypertrophic skin model***

Cutaneous incisional mouse wounds rapidly heal with minimal fibrosis and will not result in hypertrophic scarring during the normal wound healing process; thus, strategies have been developed to induce hypertrophic scar formation. Hypertrophic scars are associated with excessive scarring due to severe trauma and delayed wound healing. These scars may result in skin disfiguration and restriction of joint mobility[28,29]. In humans, hypertrophic scars develop within 4-8 wk following wound closure and growth may persist for 6 months before gradual recession[29,30]. Understanding the pathophysiology of these scars is imperative for the development of restorative treatments. Therefore, the Incisional wound model in mice has been adapted to generate pathologic scarring with mechanical tension devices, and termed the Hypertrophic skin model. Advantages of this model include low cost and reproducible production of a hypertrophic scar. Disadvantages include generation of a hypertrophic scar thinner than humans and requirement of a specially designed device to generate mechanical tension.

In this model, a 2-cm full thickness incisional wound is made on the mouse dorsal surface, and a loading device is sutured on either side of the wound bed to create tension[29,31]. Due to this external mechanical tension, a hypertrophic scar that is more similar in both histology and morphology to human cutaneous scars is created in mouse skin. Scars studied using this model demonstrate epidermal thickening with loss of adnexal structures and hair follicles, mast cell infiltrate (similar to what is seen in human HTS), hypervascularity, collagen whorls, and increased cellularity[31]. The increased cellularity observed is accompanied by a significant decrease in cellular apoptosis mediated by upregulation of Akt, a pro-survival marker[31].

***WIHN model***

Humans regenerate neither terminal nor vellus hair following large full-thickness wounds, presenting a significant clinical issue. Hair follicle regeneration is a complex process that requires coordination between multiple tissues including epidermis, dermis, muscles, and nerves. WIHN is a powerful tool used to study de novo skin regeneration following large full-thickness trauma[32]. Murine primary hair follicles are analogous to human terminal hair, while secondary hair follicles are analogous to human vellus hair, making mice a prime animal to study changes in hair growth following large full-thickness trauma. Advantages of this model include low cost, the ability to study regeneration following adult skin wounding, and development of a model to test therapeutics to activate regenerative wound healing[33]. Disadvantages of this model include variability between mouse strains, environmental conditions, age of mice, and need for creation of large wound size for the animal[33].

This model has shown that wound stiffness modulates hair follicle neogenesis and is partially regulated through mechanotransduction pathways[32]. WIHN observed using this model shows decreased focal adhesion kinase, α-smooth muscle actin, extracellular matrix expression, and cytoskeletal signaling with increased cell survival and increased levels of phospho-signal transducer and activator of transcription 3 and ephrin tyrosine kinase A3 in the central wound area[32]. These characteristics provide a wound environment that is optimal for promoting tissue regeneration following injury occurs only in the central wound area[32-34].

***Excisional wound model***

Through the use of splints, excisional wound models have been used in mice to create an animal model that more closely mimics the wound healing process in human skin[34]. The full-thickness excisional wound model is the most commonly used model to study wound healing. These wounds extend through the panniculosus carnosus, and a silicone splint is fixed around the wound to minimize contraction. Splinting increases time to complete wound closure and the amount of granulation tissue produced with no significant changes to the capacity for epithelialization[35]. The use of splinting in mice to study the wound healing process has been widely used and has contributed significantly to what we know about the wound healing process[35-37]. Advantages of this model include ease of access to the wound bed to study the effects of pharmaceuticals, biomaterials, and other agents to augment the wound healing process and assess cellular populations in granulation tissue at various stages of healing using histological and immunofluorescence techniques[35-37].

All of these models have their own unique set of advantages and disadvantages (Table 2). Nevertheless, each has contributed greatly to what we know about wound healing in healthy animals, yet there is a lack of studies using these models in understanding the effects of obesity on wound healing. Before applying these tools, we must first understand existing models to study obesity.

**ANIMAL MODELS TO STUDY OBESITY**

Several models have been developed in mice to study the impact of obesity on varying physiological processes including the use of genetic mutations and diet-induced obesity models. As described above, these animal models can be used in conjunction with existing wound healing models to examine the influence obesity has on wound healing under varying conditions. First, we will take a look at diet-induced obesity models.

***Diet-induced obesity in mice***

Human studies have also shown that high-fat diet diets with over 30% kilocalories (kcal) from fat can easily induce obesity[38]. Therefore, it is important to study obesity in this context as the dramatic rise in worldwide obesity over such a short period of time cannot be attributed to genetics[38]. In mice, a positive relationship has been found between dietary fat consumption and both body weight and fat gain[39,40]. To produce dietary-induced obesity in mice, diets contain between 40% and 60% kcal from fat. Similarly to human studies, the amount of dietary fat in the diet is associated with reduced glucose tolerance and increased white adipose tissue, blood glucose levels, and body weight over a 12-wk study period[39]. As previously mentioned, an advantage of diet-induced obesity models is that they may mimic development of obesity similarly to what is observed in humans. In addition to the low costs associated with this model, potential disadvantages of this model include length of time required to induce obesity. Diet-induced obesity models also display leptin insensitivity, which may be caused by persistently elevated leptin levels[41]. To further determine the effects of leptin on obesity, models have been developed to study changes in leptin metabolism.

***Genetic models of obesity***

The use of ob/ob mice (leptin-deficient due to obese gene mutation) and db/db mice (leptin-resistant due to diabetes gene mutation in the leptin receptor) have allowed researchers to investigate the pathogenesis of both obesity and type 2 diabetes mellitus[41]. Although the mechanisms by which these mutations cause obesity are different, mice display similar phenotypes including hyperphagia, hyperglycemia, obesity, and decreased metabolism[42,43]. Ob/ob mice display altered lipid metabolism in the liver associated with higher incidence of inflammatory infiltrate and hepatic steatosis, while db/db mice display lower hepatic inflammation with increased inflammatory tone in adipose tissue[43]. These genetic mouse models of obesity are easily replicated as the strains are readily available for purchase and are useful to study genetic impact on obesity, however they are limited in their ability to study obesity that arises due to environmental factors.

**ANIMAL STUDIES OF OBESE WOUND HEALING**

While studies using the hypertrophic skin, WIHN, and excisional splinted wound models in ob/ob, db/db, or diet-induced obesity mice have not been widely published to date, there have been studies in both mice and rats investigating the effects of obesity on cutaneous wound repair[44-47]. These data have shown significant changes in inflammatory cell infiltrate, tissue organization, and gene expression in cutaneous skin wounds[44-47]. While these studies give insight on the wound healing process in obesity, one limitation is the lack of splinting in excisional wounds. Without splinting, there is reduced mechanical tension on wounds, which reduces granulation tissue in the wound similar to what is seen in human wounds[35-37]. In an analysis between genetic and diet-induced obesity, researchers found that genetic murine models displayed wound healing capacity similar to that of diabetes mellitus in humans with reduced tissue responses in diet-induced mice with reduced serum levels of TGF-β and elevated TNF-α[47]. These results suggest that diet-induced obesity models may be better suited to study the effects of obesity on wound healing.

**FUTURE DIRECTIONS**

The wound healing process in healthy patients is well documented and is conserved between both humans and animal models used to study wound healing[12-14,17,48,49]. The effects of diabetes mellitus on wound healing have been well documented in both human and animal studies. Despite the growing body of literature surrounding wound healing, there is a gap in the literature investigating the effects of obesity on wound healing and how best to augment it in this population. Understanding the wound healing process in obesity is imperative as care requires knowledge of the physiologic demands of the body[7-9].

While there are a number of models used to study wound healing, these powerful tools have not yet been applied to study wound healing in the context of obesity. Studies using either the WIHN, excisional wound, or hypertrophic wound models should be applied to existing animal models of obesity to deepen our knowledge of the effects of obesity on wound healing. Obesity induces a chronic low-grade inflammatory response with increased production of pro-inflammatory adipokines, cytokines, and chemokines that contribute to poor wound healing phenomena seen in this population[8,11,21,44,47]. Much is known about these important proteins in the context of both obesity and normal wound healing. With an increasing number of studies assessing alterations in the both inflammatory responses and wound healing in obese mice and patients, it is imperative to continue these investigations. Future experimentation must center on altering levels of these proteins to determine if it is possible to augment wound healing outcomes in obese and overweight populations.

**CONCLUSION**

Approximately one third of the global population is classified as obese or overweight. With millions of patients sustaining cutaneous wounds annually, it is imperative to investigate the impact of obesity on wound healing and how to augment cutaneous wound healing in this growing population. Dysregulation of inflammatory responses due to production of pro-inflammatory mediators at baseline may contribute to poor wound healing outcomes.

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**REFERENCES**

1 **Chooi YC**, Ding C, Magkos F. The epidemiology of obesity. *Metabolism* 2019; **92**: 6-10 [PMID: 30253139 DOI: 10.1016/j.metabol.2018.09.005]

2 **Ogden CL**, Yanovski SZ, Carroll MD, Flegal KM. The epidemiology of obesity. *Gastroenterology* 2007; **132**: 2087-2102 [PMID: 17498505 DOI: 10.1053/j.gastro.2007.03.052]

3 **Kopelman PG**. Obesity as a medical problem. *Nature* 2000; **404**: 635-643 [PMID: 10766250 DOI: 10.1038/35007508]

4 **Pi-Sunyer X**. The medical risks of obesity. *Postgrad Med* 2009; **121**: 21-33 [PMID: 19940414 DOI: 10.3810/pgm.2009.11.2074]

5 **Greenhalgh DG**. Wound healing and diabetes mellitus. *Clin Plast Surg* 2003; **30**: 37-45 [PMID: 12636214 DOI: 10.1016/s0094-1298(02)00066-4]

6 **Brem H**, Tomic-Canic M. Cellular and molecular basis of wound healing in diabetes. *J Clin Invest* 2007; **117**: 1219-1222 [PMID: 17476353 DOI: 10.1172/jci32169]

7 **Wilson JA**, Clark JJ. Obesity: impediment to wound healing. *Crit Care Nurs Q* 2003; **26**: 119-132 [PMID: 12744592 DOI: 10.1097/00002727-200304000-00006]

8 **Groszek DM**. Promoting wound healing in the obese patient. *AORN J* 1982; **35**: 1132-1138 [PMID: 6179475 DOI: 10.1016/s0001-2092(07)62477-6]

9 **Pierpont YN**, Dinh TP, Salas RE, Johnson EL, Wright TG, Robson MC, Payne WG. Obesity and surgical wound healing: a current review. *ISRN Obes* 2014; **2014**: 638936 [PMID: 24701367 DOI: 10.1155/2014/638936]

10 **Kalupahana NS**, Moustaid-Moussa N, Claycombe KJ. Immunity as a link between obesity and insulin resistance. *Mol Aspects Med* 2012; **33**: 26-34 [PMID: 22040698 DOI: 10.1016/j.mam.2011.10.011]

11 **Dallman MF**, Pecoraro NC, La Fleur SE, Warne JP, Ginsberg AB, Akana SF, Laugero KC, Houshyar H, Strack AM, Bhatnagar S, Bell ME. Glucocorticoids, chronic stress, and obesity. *Prog Brain Res* 2006; **153**: 75-105 [PMID: 16876569 DOI: 10.1016/s0079-6123(06)53004-3]

12 **Gurtner GC**, Werner S, Barrandon Y, Longaker MT. Wound repair and regeneration. *Nature* 2008; **453**: 314-321 [PMID: 18480812 DOI: 10.1038/nature07039]

13 **Broughton G 2nd**, Janis JE, Attinger CE. The basic science of wound healing. *Plast Reconstr Surg* 2006; **117**: 12S-34S [PMID: 16799372 DOI: 10.1097/01.prs.0000225430.42531.c2]

14 **Eming SA**, Martin P, Tomic-Canic M. Wound repair and regeneration: mechanisms, signaling, and translation. *Sci Transl Med* 2014; **6**: 265sr6 [PMID: 25473038 DOI: 10.1126/scitranslmed.3009337]

15 **Landén NX**, Li D, Ståhle M. Transition from inflammation to proliferation: a critical step during wound healing. *Cell Mol Life Sci* 2016; **73**: 3861-3885 [PMID: 27180275 DOI: 10.1007/s00018-016-2268-0]

16 **Guo S**, Dipietro LA. Factors affecting wound healing. *J Dent Res* 2010; **89**: 219-229 [PMID: 20139336 DOI: 10.1177/0022034509359125]

17 **Zangooei MH**, Jalili S. Protein fold recognition with a two-layer method based on SVM-SA, WP-NN and C4.5 (TLM-SNC). *Int J Data Min Bioinform* 2013; **8**: 203-223 [PMID: 24010268 DOI: 10.1504/ijdmb.2013.055507]

18 **Lin ZQ**, Kondo T, Ishida Y, Takayasu T, Mukaida N. Essential involvement of IL-6 in the skin wound-healing process as evidenced by delayed wound healing in IL-6-deficient mice. *J Leukoc Biol* 2003; **73**: 713-721 [PMID: 12773503 DOI: 10.1189/jlb.0802397]

19 **Fasshauer M**, Blüher M. Adipokines in health and disease. *Trends Pharmacol Sci* 2015; **36**: 461-470 [PMID: 26022934 DOI: 10.1016/j.tips.2015.04.014]

20 **Oral EA**, Simha V, Ruiz E, Andewelt A, Premkumar A, Snell P, Wagner AJ, DePaoli AM, Reitman ML, Taylor SI, Gorden P, Garg A. Leptin-replacement therapy for lipodystrophy. *N Engl J Med* 2002; **346**: 570-578 [PMID: 11856796 DOI: 10.1056/nejmoa012437]

21 **Ouchi N**, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* 2011; **11**: 85-97 [PMID: 21252989 DOI: 10.1038/nri2921]

22 **Santos-Alvarez J**, Goberna R, Sánchez-Margalet V. Human leptin stimulates proliferation and activation of human circulating monocytes. *Cell Immunol* 1999; **194**: 6-11 [PMID: 10357875 DOI: 10.1006/cimm.1999.1490]

23 **Qi Y**, Nie Z, Lee YS, Singhal NS, Scherer PE, Lazar MA, Ahima RS. Loss of resistin improves glucose homeostasis in leptin deficiency. *Diabetes* 2006; **55**: 3083-3090 [PMID: 17065346 DOI: 10.2337/db05-0615]

24 **Wang T**, He C. Pro-inflammatory cytokines: The link between obesity and osteoarthritis. *Cytokine Growth Factor Rev* 2018; **44**: 38-50 [PMID: 30340925 DOI: 10.1016/j.cytogfr.2018.10.002]

25 **Vendrell J**, Broch M, Vilarrasa N, Molina A, Gómez JM, Gutiérrez C, Simón I, Soler J, Richart C. Resistin, adiponectin, ghrelin, leptin, and proinflammatory cytokines: relationships in obesity. *Obes Res* 2004; **12**: 962-971 [PMID: 15229336 DOI: 10.1038/oby.2004.118]

26 **Meyer M**, McGrouther DA. A study relating wound tension to scar morphology in the pre-sternal scar using Langers technique. *Br J Plast Surg* 1991; **44**: 291-294 [PMID: 2059787 DOI: 10.1016/0007-1226(91)90074-t]

27 **Fleischmann E**, Kurz A, Niedermayr M, Schebesta K, Kimberger O, Sessler DI, Kabon B, Prager G. Tissue oxygenation in obese and non-obese patients during laparoscopy. *Obes Surg* 2005; **15**: 813-819 [PMID: 15978153 DOI: 10.1381/0960892054222867]

28 **Kloeters O**, Tandara A, Mustoe TA. Hypertrophic scar model in the rabbit ear: a reproducible model for studying scar tissue behavior with new observations on silicone gel sheeting for scar reduction. *Wound Repair Regen* 2007; **15 Suppl 1**: S40-S45 [PMID: 17727466 DOI: 10.1111/j.1524-475X.2007.00224.x]

29 **Li J**, Wang J, Wang Z, Xia Y, Zhou M, Zhong A, Sun J. Experimental models for cutaneous hypertrophic scar research. *Wound Repair Regen* 2020; **28**: 126-144 [PMID: 31509318 DOI: 10.1111/wrr.12760]

30 **Gauglitz GG**, Korting HC, Pavicic T, Ruzicka T, Jeschke MG. Hypertrophic scarring and keloids: pathomechanisms and current and emerging treatment strategies. *Mol Med* 2011; **17**: 113-125 [PMID: 20927486 DOI: 10.2119/molmed.2009.00153]

31 **Aarabi S**, Bhatt KA, Shi Y, Paterno J, Chang EI, Loh SA, Holmes JW, Longaker MT, Yee H, Gurtner GC. Mechanical load initiates hypertrophic scar formation through decreased cellular apoptosis. *FASEB J* 2007; **21**: 3250-3261 [PMID: 17504973 DOI: 10.1096/fj.07-8218com]

32 **Harn HI**, Chiu PY, Lin CH, Chen HY, Lai YC, Yang FS, Wu CC, Tang MJ, Chuong CM, Hughes MW. Topological Distribution of Wound Stiffness Modulates Wound-Induced Hair Follicle Neogenesis. *Pharmaceutics* 2022; **14** [PMID: 36145674 DOI: 10.3390/pharmaceutics14091926]

33 **Xue Y**, Lim CH, Plikus MV, Ito M, Cotsarelis G, Garza LA. Wound-Induced Hair Neogenesis Model. *J Invest Dermatol* 2022; **142**: 2565-2569 [PMID: 36153062 DOI: 10.1016/j.jid.2022.07.013]

34 **Harn HI**, Wang SP, Lai YC, Van Handel B, Liang YC, Tsai S, Schiessl IM, Sarkar A, Xi H, Hughes M, Kaemmer S, Tang MJ, Peti-Peterdi J, Pyle AD, Woolley TE, Evseenko D, Jiang TX, Chuong CM. Symmetry breaking of tissue mechanics in wound induced hair follicle regeneration of laboratory and spiny mice. *Nat Commun* 2021; **12**: 2595 [PMID: 33972536 DOI: 10.1038/s41467-021-22822-9]

35 **Wong VW**, Sorkin M, Glotzbach JP, Longaker MT, Gurtner GC. Surgical approaches to create murine models of human wound healing. *J Biomed Biotechnol* 2011; **2011**: 969618 [PMID: 21151647 DOI: 10.1155/2011/969618]

36 **Lintel H**, Abbas DB, Lavin CV, Griffin M, Guo JL, Guardino N, Churukian A, Gurtner GC, Momeni A, Longaker MT, Wan DC. Transdermal deferoxamine administration improves excisional wound healing in chronically irradiated murine skin. *J Transl Med* 2022; **20**: 274 [PMID: 35715816 DOI: 10.1186/s12967-022-03479-4]

37 **Mascharak S**, desJardins-Park HE, Davitt MF, Griffin M, Borrelli MR, Moore AL, Chen K, Duoto B, Chinta M, Foster DS, Shen AH, Januszyk M, Kwon SH, Wernig G, Wan DC, Lorenz HP, Gurtner GC, Longaker MT. Preventing Engrailed-1 activation in fibroblasts yields wound regeneration without scarring. *Science* 2021; **372** [PMID: 33888614 DOI: 10.1126/science.aba2374]

38 **Hariri N**, Thibault L. High-fat diet-induced obesity in animal models. *Nutr Res Rev* 2010; **23**: 270-299 [PMID: 20977819 DOI: 10.1017/S0954422410000168]

39 **Takahashi M**, Ikemoto S, Ezaki O. Effect of the fat/carbohydrate ratio in the diet on obesity and oral glucose tolerance in C57BL/6J mice. *J Nutr Sci Vitaminol (Tokyo)* 1999; **45**: 583-593 [PMID: 10683810 DOI: 10.3177/jnsv.45.583]

40 **Bourgeois F**, Alexiu A, Lemonnier D. Dietary-induced obesity: effect of dietary fats on adipose tissue cellularity in mice. *Br J Nutr* 1983; **49**: 17-26 [PMID: 6821685 DOI: 10.1079/bjn19830006]

41 **Lin S**, Thomas TC, Storlien LH, Huang XF. Development of high fat diet-induced obesity and leptin resistance in C57Bl/6J mice. *Int J Obes Relat Metab Disord* 2000; **24**: 639-646 [PMID: 10849588 DOI: 10.1038/sj.ijo.0801209]

42 **Coleman DL**. Obese and diabetes: two mutant genes causing diabetes-obesity syndromes in mice. *Diabetologia* 1978; **14**: 141-148 [PMID: 350680 DOI: 10.1007/BF00429772]

43 **Suriano F**, Vieira-Silva S, Falony G, Roumain M, Paquot A, Pelicaen R, Régnier M, Delzenne NM, Raes J, Muccioli GG, Van Hul M, Cani PD. Novel insights into the genetically obese (ob/ob) and diabetic (db/db) mice: two sides of the same coin. *Microbiome* 2021; **9**: 147 [PMID: 34183063 DOI: 10.1186/s40168-021-01097-8]

44 **Min KK**, Neupane S, Adhikari N, Sohn WJ, An SY, Kim JY, An CH, Lee Y, Kim YG, Park JW, Lee JM, Kim JY, Suh JY. Effects of resveratrol on bone-healing capacity in the mouse tooth extraction socket. *J Periodontal Res* 2020; **55**: 247-257 [PMID: 31797379 DOI: 10.1111/jre.12710]

45 **Slavkovsky R**, Kohlerova R, Tkacova V, Jiroutova A, Tahmazoglu B, Velebny V, Rezačová M, Sobotka L, Kanta J. Zucker diabetic fatty rat: a new model of impaired cutaneous wound repair with type II diabetes mellitus and obesity. *Wound Repair Regen* 2011; **19**: 515-525 [PMID: 21649785 DOI: 10.1111/j.1524-475X.2011.00703.x]

46 **Nascimento AP**, Costa AM. Overweight induced by high-fat diet delays rat cutaneous wound healing. *Br J Nutr* 2006; **96**: 1069-1077 [PMID: 17181882 DOI: 10.1017/bjn20061955]

47 **Seitz O**, Schürmann C, Hermes N, Müller E, Pfeilschifter J, Frank S, Goren I. Wound healing in mice with high-fat diet- or ob gene-induced diabetes-obesity syndromes: a comparative study. *Exp Diabetes Res* 2010; **2010**: 476969 [PMID: 21318183 DOI: 10.1155/2010/476969]

48 **Talbott HE**, Mascharak S, Griffin M, Wan DC, Longaker MT. Wound healing, fibroblast heterogeneity, and fibrosis. *Cell Stem Cell* 2022; **29**: 1161-1180 [PMID: 35931028 DOI: 10.1016/j.stem.2022.07.006]

49 **Diegelmann RF**, Evans MC. Wound healing: an overview of acute, fibrotic and delayed healing. *Front Biosci* 2004; **9**: 283-289 [PMID: 14766366 DOI: 10.2741/1184]

**Footnotes**

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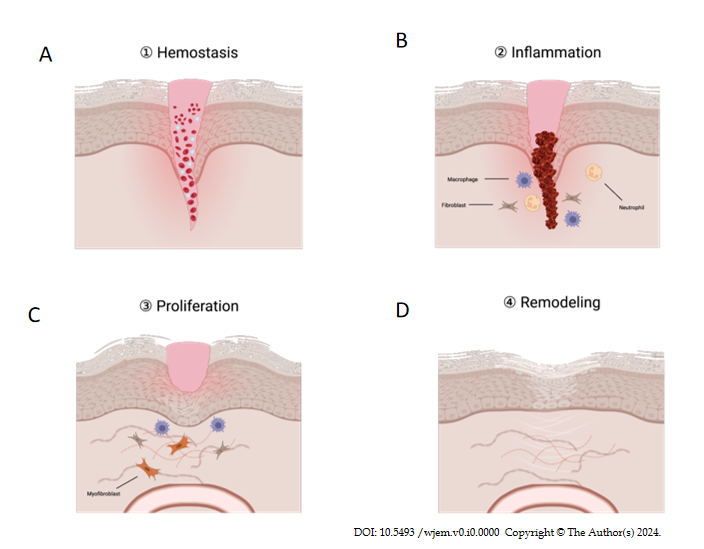
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Grade D (Fair): 0

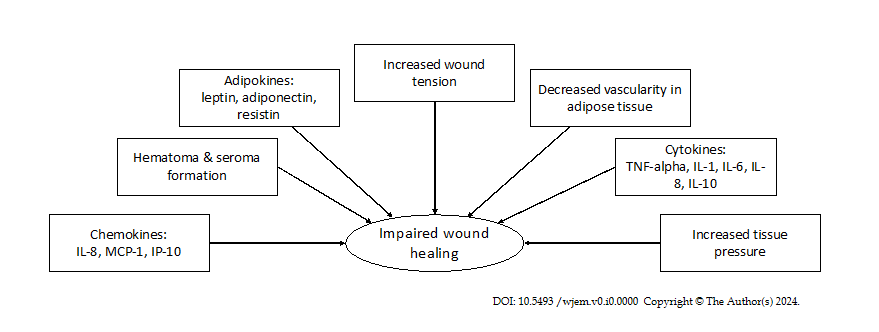
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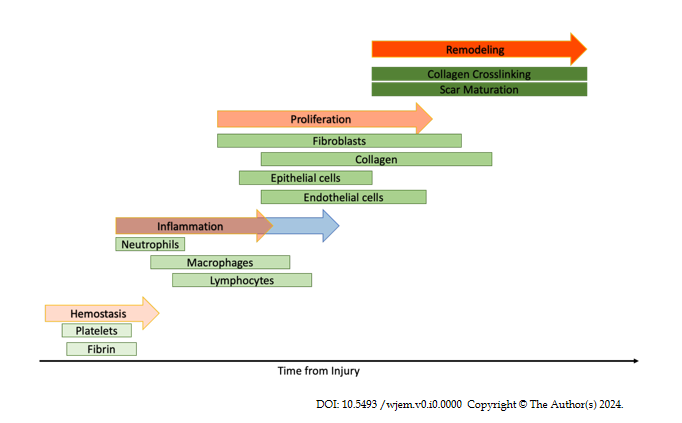
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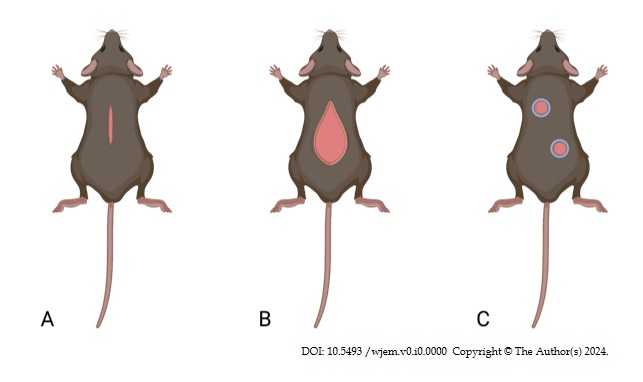
**Figure 1 The four phases of wound healing. Wound healing is divided into 4 distinct phases that have overlap.** A: Hemostasis, B: Inflammation, C: Proliferation, D: Remodeling. Cell-to-cell interactions mediated by both cytokines and chemokines are imperative for the transitions between phases. During hemostasis, platelets and fibrin function to form a plug to stop bleeding. Hemostasis is followed by the inflammatory phase which is characterized by release of cytokines, most notably from macrophages, that are imperative for induction of the proliferative phase. During proliferation, the body is focused on neovascularization and re-epithelialization of the wound surface. The final phase, remodeling, takes place over months to years and is currently not well characterized.



**Figure 2 Factors related to wound healing impairment in obesity.** Obesity is characterized by a chronic inflammatory state that is associated with changes mediated by varying levels of adipokines, chemokines, and cytokines. In addition to these chemical signals, physiologic changes including increased tissue pressure and decreased vascularity of adipose tissue also contribute to poor wound healing outcomes. MCP-1: Monocyte chemoattractant protein-1; IP-10: Interferon-gamma-inducible protein 10; TNF-α: Tumor necrosis factor alpha; IL: Interleukin.



**Figure 3 A timeline of the four phases of wound healing.** Each phase of wound healing is mediated by a distinct population of cells. While the phases have significant overlap, alterations in levels of cytokines or cell types in any phase may cause delayed wound healing. The inflammatory phase is notably extended in obese patients (blue arrow), this is thought to contribute to poor healing outcomes in this population.



**Figure 4 Schematic of three major models of wound healing.** A: Hypertrophic wound model. This model allows for use of a device to produce constant tension in the wound bed to produce healing with a hypertrophic scar similar to what is seen in areas of high tension on the body; B: Wound-induced hair follicle neogenesis model. This model is used to investigate regeneration in the setting of large full-thickness trauma; C: Excisional wound model. This model is the most commonly used model to study wound healing and is popular for its ability to investigate the role of various therapeutics to augment wound healing in a similar manner to what is seen in human skin.

**Table 1 Cytokines produced by adipose tissue that may contribute to low-grade inflammation**

|  |  |
| --- | --- |
| TNF-α | Secreted by macrophages, natural killer cells, and lymphocytes[13]. Crucial in formation and maintenance of granulomas[30] |
| IL-1 | Produced by various cell types including macrophages, fibroblasts, and epithelial cells[13]. Stimulates fibroblast and keratinocyte growth and collagen synthesis by fibroblasts |
| IL-6 | Secreted by various cell types including macrophages and adipocytes[13]. An important mediator of the acute phase response and regulator of glucose homeostasis in obesity |
| IL-8 | Produced by various cell types including macrophages, epithelial cells, and endothelial cells[13]. Primarily recruits neutrophils and other granulocytes to sites of tissue injury |
| MCP-1 | Also known as CCL2. Primarily secreted by monocytes, macrophages, and dendritic cells[13]. Attracts monocytes, memory T cells, and dendritic cells to sites of inflammation produced by either tissue injury or inflammation |

TNF-α: Tumor necrosis factor alpha; IL: Interleukin; MCP-1: Monocyte chemoattractant protein-1.

**Table 2 Animal models used in wound healing, advantages and disadvantage**s

|  |  |  |
| --- | --- | --- |
| Animal model | Advantages | Disadvantages |
| Hypertrophic wound model | Low cost | Hypertrophic scar different from human[27,29] |
|  | Allows for production of hypertrophic scar[28,29] | Requires special device[28,29] |
|  |  | Labor intensive: Frequent care to maintain tension and device placement[31] |
| Wound-induced hair follicle neogenesis (WIHN) model | Low cost |  |
|  | Regeneration following adult wounding with minimal recovery of hair follicles at the scar center[32,48] | High variability: mouse strains, environmental conditions, age of mice, and wound size)[33] |
|  | Test therapeutics to activate regenerative wound healing[33] | May not translate to human injury[48] |
| Excisional wound model | Low cost | Labor intensive: frequent dressing changes to maintain tension[35] |
|  | Wound healing similar to human[35] |  |
|  | Allows for fibroblast lineage tracing[49] |  |