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Peer Reviewer of World Journal of Cardiology, Ahed Jumah Alkhatib, MD, PhD, Doctor, Research Scientist, Department of Legal Medicine, Toxicology and Forensic Medicine, Jordan University of Science & Technology, Jordan, Irbid 22110, Irbid, Jordan. ajalkhatib@just.edu.jo

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EDITORIAL

Mechanistic insights into fasting-induced autophagy in the aging heart

Hannaneh Parvaresh, Katarzyna Paczek, Md Abdul Alim Al-Bari, Nabil Eid

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Hannaneh Parvaresh, Department of Biology, Faculty of Science, Ferdowsi University of Mashhad, Mashhad 9177948974, Iran

Katarzyna Paczek, Department of Chiropractic, International Medical University, Kuala Lumpur 57000, Malaysia

Md Abdul Alim Al-Bari, Department of Pharmacy, University of Rajshahi, Rajshahi 6205, Bangladesh

Nabil Eid, Department of Anatomy, Division of Human Biology, School of Medicine, International Medical University, Kuala Lumpur 57000, Malaysia

Corresponding author: Nabil Eid, MD, PhD, Academic Editor, Associate Professor, Lecturer, Department of Anatomy, Division of Human Biology, School of Medicine, International Medical University, Bukit Jalil, Kuala Lumpur 57000, Malaysia. nabilsaleheid@imu.edu.my

Abstract

Autophagy is a prosurvival mechanism for the clearance of accumulated abnormal proteins, damaged organelles, and excessive lipids within mammalian cells. A growing body of data indicates that autophagy is reduced in aging cells. This reduction leads to various diseases, such as myocardial hypertrophy, infarction, and atherosclerosis. Recent studies in animal models of an aging heart showed that fasting-induced autophagy improved cardiac function and longevity. This improvement is related to autophagic clearance of damaged cellular components via either bulk or selective autophagy (such as mitophagy). In this editorial, we summarize the mechanisms of autophagy in normal and aging hearts. In addition, the protective effect of fasting-induced autophagy in cardiac aging has been highlighted.

Key Words: Aging; Autophagy; Heart; Fasting; Mitophagy

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Core Tip: Autophagy is an essential mechanism for the clearance of harmful cellular components, which accumulate with age. However, autophagic machinery decreases with age, resulting in various diseases, such as cardiac hypertrophy. Recently, fasting-induced autophagy has been reported to improve cardiac function in animal models of aging via normalization of defective autophagic machinery. Therefore, autophagy is an important target for the prevention of cardiac pathologies in the geriatric population.

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INTRODUCTION

Cardiac aging

Improvements in treatment procedures have contributed to increased life expectancy and growth in the aged population, especially in industrialized countries[1]. Aging is associated with a structural and functional decline in multiple organs, such as the heart. A sedentary lifestyle can also accelerate the incidence of aging-related diseases, including cardiovascular disease (CVD)[2-5].

Cardiovascular aging affects both the heart and the blood circulation system through slow and progressive alterations that can result in the development of left ventricular hypertrophy, diastolic dysfunction, coronary artery disease, stroke, hypertension, atherosclerosis, atrial fibrillation, and heart failure [6-9]. Aortic valve sclerosis is a valvulopathy associated with aging and is characterized by myxomatous degeneration, collagen deposition, and progression to aortic stenosis (AS)[10]. AS is an indicator of increased CVD risk and is mainly defined as increased leaflet calcification and decreased leaflet mobility[11]. Moreover, approximately 13%-16% of elderly people suffer from aortic regurgitation[12], which results in left ventricular dilation and dysfunction over time. Another valvular change related to aging is mitral annular calcification, which usually accompanies aortic valve sclerosis[13].

The free radical theory of aging and the mitochondrial theory have been suggested to explain the cellular deterioration observed in aging and suggest that the age-related decline in mitochondrial function and structure is a major driver of cardiomyocyte senescence, which causes endothelial dysfunction, alteration in the vasculature, and/or vascular injury

Cellular senescence is activated following multiple stressors, including the elevation of reactive oxygen species (ROSs); proinflammatory cytokines; and metabolic, mechanical, and chemical toxicity. Cellular senescence impairs the repair and regeneration of damaged cells in cardiovascular tissues [15-17]. Cellular senescence is characterized by genome instability, telomere attrition, and mitochondrial dysfunction[18].

Dysfunctional mitochondria produce less ATP while also generating increased amounts of ROS[19], exposing aged cardiomyocytes to high levels of oxidative stress. Autophagic and proteasomal degradation are the main mechanisms for the removal of damaged mitochondria and abnormal proteins in aged postmitotic cardiomyocytes. However, these mechanisms decline with age [20]. Eventually, when these mechanisms are unable to compensate for the accumulated cellular damage, stem-cell exhaustion and altered intercellular communication occur, further contributing to aging [18].

Autophagy in cardiac aging: Reduced autophagy accelerates cardiac aging

Autophagy activity is usually reduced with age[21]. A decrease in autophagy in the hearts of aged flies[22] and aged C57BL/6 mice (20-26 months old) has been reported[23,24].

Autophagy is a protective housekeeping mechanism critical for cellular homeostasis and survival. Long-lived, damaged, and dysfunctional organelles; misfolded proteins; and invading pathogens are eliminated through this degradation process, providing building components for cellular renovation to effectively adapt cells to stressful conditions, such as nutrient deprivation, hypoxia, or oxidative stress[25,26].

Autophagy can be selective or nonselective. Under starvation conditions, the protein and any cytoplasmic content can be non-selectively targeted for catabolic recycling to maintain cellular energy production. However, there are also selective forms of autophagy that specifically target damaged organelles. For instance, mitophagy is a type of autophagy that selectively removes damaged mitochondria [27]. Mitochondria play a substantial role in cellular functions as well as cellular death. Thus, mitochondrial dysfunction is a crucial determinant of lifespan across species [28,29].

Three types of autophagy have been recognized: Macroautophagy, microautophagy, and chaperone-mediated autophagy, all of which lead to the turnover of intracellular components via various mechanisms. "Autophagy" is a term that generally refers to macroautophagy, which is the most prevalent form of autophagy[30,31].

Molecular machinery of autophagy

Autophagy is initiated when several autophagy-related gene products (Atg1-Atg12) and other proteins are organized to form a phagophore. These proteins consist of at least five molecular components that mediate fusion between autophagosome (AP) and lysosomes: (1) The Atg1/unc-51-like kinase complex; (2) the Beclin 1/class III phosphatidylinositol 3kinase (PI3K) complex; (3) Atg9 and vacuole membrane protein 1; (4) two ubiquitin-like proteins (Atg12 and Atg8/LC3)



conjugation systems; and (5) proteins that mediate fusion between APs and lysosomes[25,32].

The initial step of AP formation starts with Beclin1 (Atg6) and class III PI3K, which play crucial roles in vesicle isolation. Other Atg proteins are involved in Beclin-1-mediated formation of the Class III PI3K complex. In the next step, the AP undergoes elongation *via* two conjugation systems. First, Atg12 is conjugated to Atg5 with the help of Atg7 and Atg10[33,34], followed by the conjugation of phosphatidylethanolamine to microtubule-associated protein 1 LC3 *via* Atg4, Atg7 and Atg3. Consequently, the cytoplasmic LC3 (LC3-I) is converted to membranous (LC3-II) form, which is responsible for formation and maturation of the AP[35]. In the end, fusion of APs and lysosomes occurs with the formation of autolysosome (AL) for degradation and recycling[36].

The protein kinases mammalian target of rapamycin (mTOR) and AMP-activated protein kinase (AMPK) are implicated in the regulatory mechanisms of autophagy. Autophagy is inhibited by the mTOR. Phosphorylation of Unc-51 Like autophagy activating kinase-1 (ULK1) by AMPK is involved in autophagy promotion, although mTOR represses this process[37]. Figure 1 demonstrates the various mechanisms of autophagy in mammalian cells.

Autophagy in the heart

Accumulating evidence reveals that autophagy plays essential homeostatic roles in the heart under normal physiological conditions and during the aging process; additionally, it has an essential role in improving the immune response and reducing inflammation[38]. Consequently, any perturbations to this process in the cardiovascular system can elicit harmful effects on health.

Autophagy attenuates with age and has serious implications for heart structure and function. A decrease in autophagy causes the development of heart failure, hypertension, atherosclerosis, and ischemic heart disease[39].

Mitophagy is the selective autophagic clearance of damaged mitochondria and is crucial for the bioenergetics of the cardiovascular system; thus, mitophagy dysfunction is generally accompanied by cardiac disorders[27,40,41]. In addition, studies have suggested that autophagic degradation of damaged mitochondria decelerates cardiovascular senescence and has a positive effect on the healthy lifespan of animals[42-44].

Age-induced impairment of autophagy

Cardiomyocytes undergo age-related changes in proteostasis pathways, resulting in calcium homeostasis impairment, ROSs induction, hypertrophy and fibrosis, and eventual structural damage and diminished cardiac function. Moreover, with age, the MTOR-1 complex is significantly upregulated, and the AMPK pathway is downregulated. In addition, transcription factors involved in autophagy and lysosomal proteins such as TFEB and Forkhead transcription factor (FOXO) 3 are deactivated with advanced aging, resulting in reduced expression of autophagy genes [28-31].

Any defect in the autophagy process accelerates aging; likewise, aging is suppressed when autophagy is stimulated. Deletion of atg5, a cardiac-specific autophagy-related gene, in adult mice leads to an accelerated aging phenotype, including the development of cardiac hypertrophy, left ventricular dilatation, and contractile dysfunction [20,45].

Mutations in the atg4c gene increase the risk of heart disease in elderly patients and eventually death[46]. Cardiomyocyte-specific deletion of glycogen synthase kinase-3 in mice reduced basal autophagy levels and accelerated cardiac aging[47]. Dysfunction of autophagy with age slows the turnover of damaged proteasomes and contributes to age-associated CVD and cardiomyocyte senescence[48]. Mitophagy is impaired in aged mice, and mitophagy induction improves mitochondrial function and reduces arterial wall stiffness[49].

Acyl-coenzyme A binding protein (ACBP), which is encoded by a diazepambinding inhibitor (DBI), acts as an extracellular feedback inhibitor of autophagy[50]. It appears that high ACBP/DBI values correlate with future cardiovascular events (such as heart surgery, myocardial infarction, and stroke), suggesting that ACBP/DBI is indeed a biomarker of biological aging[39].

Mechanisms underlying age-related cardiac remodeling: involvement of autophagy

Although there are many potential causes underlying the decline in cardiovascular function with age, a major determinant of the aging process is likely the progressive loss of quality control due to reduced autophagy.

Hyperactivation of mTOR and reduced AMPK activity[51] in old age can directly inhibit autophagy by inactivating the pro-autophagic ULK1 complex[52], contributing to the downregulation of autophagy activity.

It is conceivable that exposure to excessive ROS during aging promotes the accumulation of oxidized proteins, mitochondrial DNA mutations, and protein misfolding[53]. Additionally, several cytosolic and mitochondrion-localized proteins involved in autophagy regulation become dysfunctional, thus contributing to abnormal mitochondrial turnover and the removal of damaged mitochondria[54]. This chain of events results in impaired autophagy due to exhaustion of the aged autophagic machinery.

In addition, it has been proposed that a hallmark of aging in postmitotic cells, such as cardiomyocytes, is the aggregation of nondegradable structures inside lysosomes, termed lipofuscin, which impedes lysosomal function and therefore can likely inhibit autophagy[55].

It has been shown that intracellular calcium has a key regulatory effect on cardiomyocyte autophagy. Inositol 1,4,5-trisphosphate (IP3) receptors mediate calcium release and transfer to mitochondria. This process inhibits autophagy by suppressing AMPK activation[56]. Since evidence has shown that IP3 receptors are upregulated in the aged, hypertrophied, and failing myocardium of rodents[57] and humans[58], increased IP3 receptor-mediated calcium signaling likely exacerbates autophagy in the aging heart[59].

FOXO and sirtuin proteins are also major metabolic regulators that mediate age-related vascular changes, particularly endothelial dysfunction[9].

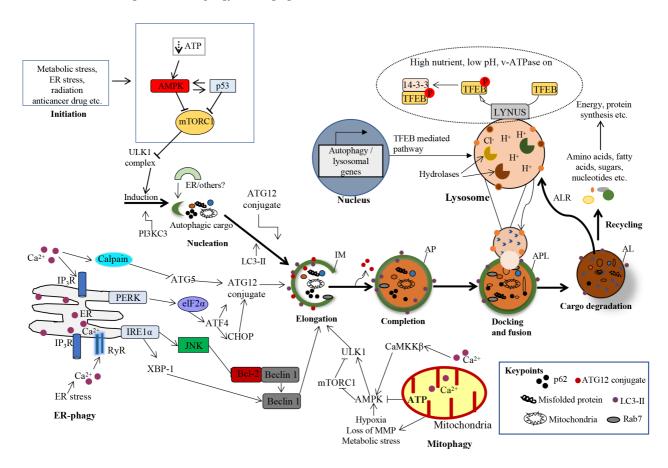


Figure 1 Molecular mechanisms of various stages of autophagy. Autophagy is activated in response to various cellular stresses and is triggered by a decrease in rapamycin complex 1 (mTORC1) activity due to the activation of AMP-activated protein kinase (AMPK) or p53 signaling. mTORC1 suppresses the activity of Unc-51-like autophagy activating kinase 1 (ULK1) complex. Therefore, inhibition of mTORC1 causes the initialization of the ULK1-mediated formation of the isolation (autophagosomal) membrane (IM) in association with the class III phosphatidylinositide 3-kinase complex. The IM expands into an autophagosome (AP) with a double-layer membrane, which can engulf any cellular component, including proteins, damaged organelles, and lipid droplets. The AP merges with the lysosome (via LAMP-1, 2), forming autophagolysosome or autolysosome (AL), and resulting in the degradation of the cargo by cathepsins and the autophagic lysosome reformation. The nucleation, elongation and maturation of the IM are dependent on two ubiquitin-like conjugation systems (ATG12 and ATG8), which involve multiple autophagy proteins, including Beclin1, ATG5, ATG16 and MT-associated protein 1 LC3. The AL provides an acidic milieu for hydrolytic enzymes to digest the engulfed components. Nuclear localization of transcription factor EB is critical to the formation of lysosomes and to the enhanced expression of autophagy proteins. Importantly, autophagy could be selective of mitochondria (mitophagy) or ER (ER-phagy). However, the detailed mechanisms of this selected autophagy are beyond the scope of this study[28]. AMPK: AMP-activated protein kinase; PI3KC3: Phosphatidylinositide 3-kinase complex; APL: Autophagolysosome; AL: Autolysosome; ALR: Autophagic lysosome reformation; IM: Isolation (autophagosomal) membrane; TFEB: Transcription factor EB; mTORC1: Rapamycin complex 1. Citation: Al-Bari MAA, Ito Y, Ahmed S, Radwan N, Ahmed HS, Eid N. Targeting Autophagy with Natural Products as a Potential Therapeutic Approach for Cancer. Int J Mol Sci 2021; 22: 9807. Copyright ©The Author(s) 2021. Published by MDPI.

Dietary activation of autophagy in the heart via caloric restriction or fasting

Dietary interventions involving caloric restriction (CR) and fasting are among several stress stimuli that can induce autophagy in response to food deprivation[60-62]. CR was defined as a reduction in caloric intake using a diet containing adequate amounts of protein, vitamins, and minerals[63]. CR is a potent inducer of autophagy in the heart[64], and its positive impacts on health and lifespan in various model organisms, primates and humans have been studied [65-67]. CR is the most potent physiological stimulus of autophagy and ameliorates cardiac dysfunction (systolic and diastolic) and attenuates myocardial hypertrophy and fibrosis at the cardiomyocyte level. CR reduces mitochondrial damage, lipid accumulation, oxidative stress, apoptosis, telomere shortening, senescence marker levels, and circulating proinflammatory cytokine levels[68].

Autophagy plays an important role in CR-mediated longevity [69] via clearance of damaged mitochondria, reduction of oxidative stress, improvement of insulin sensitivity and suppression of inflammatory responses[61,62].

Short-term CR for 10 wk in mice rejuvenated symptoms of the aging heart, such as significant improvement in diastolic function and regression of age-dependent cardiac hypertrophy[70]. Moreover, CR reversed age-dependent cardiac proteome remodeling and mitigated oxidative damage and ubiquitination in these mice.

In aged animals, hypertrophy, and fibrosis, as well as systolic and diastolic dysfunctions, improved after CR[68,71]. The beneficial effects of CR observed in cardiomyocytes include enhanced mitochondrial fitness and reduced oxidative stress, apoptotic cell death, inflammation, and importantly, senescence[68]. In vasculature, CR helps improve endothelial cell function and attenuates collagen deposition, elastin remodeling, and oxidative stress; as a result, CR reduces arterial stiffness[72]. Another study revealed improvements in numerous markers of cardiovascular health in humans after short-

term periodic fasting, which is also a pro-autophagic dietary regimen [73].

Intermittent fasting (IF) has attracted the attention of researchers as a dietary intervention associated with better compliance and long-term adherence than CR in recent years[74]. IF consists of regular cycles of times with no or minimal caloric intake interrupted by periods of normal food consumption. Alternate day fasting delays cardiac aging in rats, as determined by reduced hypertrophy and fibrosis[75,76] and extended lifespan[77]. The advantageous effects of life-long alternate-day fasting were attributed to reduced phosphoinositide 3-kinase signaling, which was associated with reduced myocardial collagen deposition, oxidative stress, inflammatory markers, and B-type natriuretic peptide levels[75,78].

A fasting-mimicking diet (FMD) is considered another form of dietary intervention in which individuals consume low amounts of calories, sugars, and proteins but high amounts of unsaturated fats. Studies of FMD effects in mice have shown improved cognitive function and a rejuvenated immune system, in addition to promoting lifespan and health factors by reducing cancer incidence, obesity, and inflammation[79]. FMD was investigated in humans, and the findings showed reduced age-related CVD risk factors, including reduced blood pressure, body mass index, fasting glucose, and inflammation, as well as an improved lipid profile[80].

The efficacy of fasting on autophagy in the heart was assessed in male FBN rats by randomly dividing them into different groups of equal amounts of protein, vitamin, and mineral intake, while the CR groups received 20% less food from a 125% fortified diet for six weeks. Additionally, in addition to one simple CR group, two other CR groups were given 5 or 50 mg/kg/day resveratrol. Compared with AL group, a marked reduction of expression of p62 (autophagy substrate) in the left ventricle was observed in the CR and Resv-50 rats, indicating enhanced cardiac autophagy in the CR group. Similarly, a significant overexpression of Beclin-1 was found in the Resv-50 and CR animals. The CR + Resv-50 group of rats showed dramatically attenuated doxorubicin-induced damage, which can be due to enhanced autophagy [81]. Another study investigated the autophagic response of CR on diabetic rat hearts. Diabetic and nondiabetic rats were exposed to a CR diet (30% energy reduction) for 32 wk. Compared with those of diabetic AL rats, diabetic CR rats exhibited an increase in the hepatic and cardiac LC3-II/LC3-I ratio (indicating enhanced autophagy)[82].

A high-fat diet (HFD) (fat 60% kcal/100 kcal fat) was given to the FVBN male mice for 4-20 wk, after which they were subjected to overnight fasting to study the mechanisms of fasting-induced autophagy in the fatty mice heart. After 24 h of fasting, there was a significant conversion of LC3-I conversion to LC3-II in lean mice heart but was not associated with a change in diet-induced obesity (DIO) mice. Furthermore, fasting suppressed mTOR in both lean and DIO mice, as indicated by increased AMPK phosphorylation and enhanced dephosphorylation of S6. Interestingly, mTOR inhibition was greater in obese mice. Taken together, these findings indicate that fasting activates autophagy in the hearts of lean mice[83].

Godar *et al*[84] investigated the impacts of IF on the autophagy-lysosome machinery in the myocardium. The authors studied the effects of fasting after 24 h, followed by 24 h of refeeding or 24 and 48 h of fasting for six weeks. The AP abundance increased dramatically after 48 h of fasting. Treatment with chloroquine (an autophagy inhibitor) was associated with a significant increase in LC3-II and SQSTM1/p62 after 24 h of fasting but not in fed mice. Thus, fasting induces autophagy in cardiomyocytes; however, autophagy returns to basal levels on gestational days.

The effects of IF on right ventricular (RV) function in a rat model of pulmonary arterial hypertension (characterized by RV mitochondrial dysfunction and resultant lipotoxicity and microbiome dysbiosis) were explored. IF improved RV systolic and diastolic function and decreased RV cardiomyocyte hypertrophy and fibrosis, which was likely mediated by autophagy activation[85]. These protective effects could be related to autophagy activation.

Recent findings from studies also show that cardiometabolic parameters (*e.g.*, adiposity, insulin sensitivity, and cardiac function) can be influenced by the time of day at which food is consumed[86]. To test the hypothesis that fasting during the sleep period elicits beneficial adaptation effects on cardiac function, wild-type mice were fasted for 24 h or for either the 12-h light/sleep phase or the 12-h dark/awake phase. Repression of myocardial p-mTOR and protein synthesis occurred during the dark phase; both parameters remained elevated in the hearts of fasted mice during the light phase. In contrast, markers of autophagy (*e.g.*, LC3-II) exhibited peak responses to fasting during the light phase. Collectively, these data show that the responsiveness of the heart to fasting is temporally partitioned[86].

IF alleviated HFD-induced obesity cardiomyopathy in male C57BL/6J mice by improving cardiac functional and structural impairment and serum lipid metabolic disorders induced by HFD through decreasing lipid deposition, apoptosis and m6A methylation in the heart[87].

Researchers compared the effects of alternate day fasting on elderly (aged 24 months) and young (aged 6 months) male rats. The results of this study indicated that alternate day fasting protected against inflammation and fibrosis in the heart during aging by inhibiting oxidative damage and NF-kB activation[76]. Other studies have shown that fasting preconditioning activates AMPK, induces autophagy, decreases ROS levels, and inhibits NF-kB signaling in the cardiac tissues of rats[88]. In addition, compared with fasting controls, IF in human subjects resulted in autophagy upregulation and reduced levels of proinflammatory cytokines, indicating the protective effects of fasting on the vascular system. This effect is most likely mediated by the anti-inflammatory effects of autophagy[89]. We investigated fasting-induced autophagy among large groups of population in the UAE during Ramadan (the holy Islamic fasting month). The results of this study will be published shortly in specific journals. Furthermore, these results were presented in part at the Sharjah First International Conference on Fasting, February 28-29, 2024, at Sharjah University, United Arab Emirates[90].

CONCLUSION

In conclusion, fasting-induced autophagy is beneficial for ensuring cardiac function, preventing disease, and improving longevity. However, additional studies *in vivo* in animal models of cardiac aging are needed to determine the specific

molecular mechanisms involved in normalizing autophagy by fasting. In addition, large-scale studies on humans are needed. Ramadan fasting, a type of IF (a common religious practice) in Islamic countries, could be investigated in large groups of geriatric people with or without cardiac diseases. Importantly, further in vitro research should be directed toward human cardiac tissues to better understand the molecular mechanisms of fasting-induced autophagy and its beneficial effects on longevity pathways and prevention of CVDs.

FOOTNOTES

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ORCID number: Md Abdul Alim Al-Bari 0000-0002-1777-3662; Nabil Eid 0000-0002-2938-2618.

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REFERENCES

- Lutz W, Sanderson W, Scherbov S. The coming acceleration of global population ageing. Nature 2008; 451: 716-719 [PMID: 18204438 DOI: 10.1038/nature06516]
- 2 Mattson MP. Lifelong brain health is a lifelong challenge: from evolutionary principles to empirical evidence. Ageing Res Rev 2015; 20: 37-45 [PMID: 25576651 DOI: 10.1016/j.arr.2014.12.011]
- 3 Mattson MP, Arumugam TV. Hallmarks of Brain Aging: Adaptive and Pathological Modification by Metabolic States. Cell Metab 2018; 27: 1176-1199 [PMID: 29874566 DOI: 10.1016/j.cmet.2018.05.011]
- Evans MA, Sano S, Walsh K. Cardiovascular Disease, Aging, and Clonal Hematopoiesis. Annu Rev Pathol 2020; 15: 419-438 [PMID: 4 31689371 DOI: 10.1146/annurev-pathmechdis-012419-032544]
- Virani SS, Alonso A, Benjamin EJ, Bittencourt MS, Callaway CW, Carson AP, Chamberlain AM, Chang AR, Cheng S, Delling FN, Djousse L, Elkind MSV, Ferguson JF, Fornage M, Khan SS, Kissela BM, Knutson KL, Kwan TW, Lackland DT, Lewis TT, Lichtman JH, Longenecker CT, Loop MS, Lutsey PL, Martin SS, Matsushita K, Moran AE, Mussolino ME, Perak AM, Rosamond WD, Roth GA, Sampson UKA, Satou GM, Schroeder EB, Shah SH, Shay CM, Spartano NL, Stokes A, Tirschwell DL, VanWagner LB, Tsao CW; American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. Heart Disease and Stroke Statistics-2020 Update: A Report From the American Heart Association. Circulation 2020; 141: e139-e596 [PMID: 31992061 DOI: 10.1161/CIR.0000000000000757]
- Cai Y, Liu H, Song E, Wang L, Xu J, He Y, Zhang D, Zhang L, Cheng KK, Jin L, Wu M, Liu S, Qi D, Lopaschuk GD, Wang S, Xu A, Xia Z. Deficiency of telomere-associated repressor activator protein 1 precipitates cardiac aging in mice \emph{via} p53/PPAR α signaling. $\emph{Theranostics}$ 2021; 11: 4710-4727 [PMID: 33754023 DOI: 10.7150/thno.51739]
- 7 Hu C, Zhang X, Teng T, Ma ZG, Tang QZ. Cellular Senescence in Cardiovascular Diseases: A Systematic Review. Aging Dis 2022; 13: 103-128 [PMID: 35111365 DOI: 10.14336/AD.2021.0927]
- Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a "set up" for 8 vascular disease. Circulation 2003; 107: 139-146 [PMID: 12515756 DOI: 10.1161/01.cir.0000048892.83521.58]
- North BJ, Sinclair DA. The intersection between aging and cardiovascular disease. Circ Res 2012; 110: 1097-1108 [PMID: 22499900 DOI: 9 10.1161/CIRCRESAHA.111.246876]
- Otto CM. Why is aortic sclerosis associated with adverse clinical outcomes? J Am Coll Cardiol 2004; 43: 176-178 [PMID: 14736433 DOI: 10 10.1016/j.jacc.2003.10.027]
- Dai DF, Chen T, Johnson SC, Szeto H, Rabinovitch PS. Cardiac aging: from molecular mechanisms to significance in human health and 11 disease. Antioxid Redox Signal 2012; 16: 1492-1526 [PMID: 22229339 DOI: 10.1089/ars.2011.4179]
- Nassimiha D, Aronow WS, Ahn C, Goldman ME. Association of coronary risk factors with progression of valvular aortic stenosis in older 12 persons. Am J Cardiol 2001; 87: 1313-1314 [PMID: 11377366 DOI: 10.1016/s0002-9149(01)01531-4]
- 13 Jeon DS, Atar S, Brasch AV, Luo H, Mirocha J, Naqvi TZ, Kraus R, Berman DS, Siegel RJ. Association of mitral annulus calcification, aortic valve sclerosis and aortic root calcification with abnormal myocardial perfusion single photon emission tomography in subjects age < or =65 years old. J Am Coll Cardiol 2001; 38: 1988-1993 [PMID: 11738305 DOI: 10.1016/s0735-1097(01)01678-3]
- Zhao RZ, Jiang S, Zhang L, Yu ZB. Mitochondrial electron transport chain, ROS generation and uncoupling (Review). Int J Mol Med 2019; 14 44: 3-15 [PMID: 31115493 DOI: 10.3892/ijmm.2019.4188]
- Colavitti R, Finkel T. Reactive oxygen species as mediators of cellular senescence. IUBMB Life 2005; 57: 277-281 [PMID: 16036611 DOI: 15 10.1080/15216540500091890]
- Olivieri F, Prattichizzo F, Grillari J, Balistreri CR. Cellular Senescence and Inflammaging in Age-Related Diseases. Mediators Inflamm 2018;



- **2018**: 9076485 [PMID: 29849499 DOI: 10.1155/2018/9076485]
- Lewis-McDougall FC, Ruchaya PJ, Domenjo-Vila E, Shin Teoh T, Prata L, Cottle BJ, Clark JE, Punjabi PP, Awad W, Torella D, Tchkonia T, 17 Kirkland JL, Ellison-Hughes GM. Aged-senescent cells contribute to impaired heart regeneration. Aging Cell 2019; 18: e12931 [PMID: 30854802 DOI: 10.1111/acel.12931]
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell 2013; 153: 1194-1217 [PMID: 23746838 DOI: 18 10.1016/j.cell.2013.05.039]
- Lesnefsky EJ, Chen Q, Hoppel CL. Mitochondrial Metabolism in Aging Heart. Circ Res 2016; 118: 1593-1611 [PMID: 27174952 DOI: 19 10.1161/CIRCRESAHA.116.307505]
- 20 Taneike M, Yamaguchi O, Nakai A, Hikoso S, Takeda T, Mizote I, Oka T, Tamai T, Oyabu J, Murakawa T, Nishida K, Shimizu T, Hori M, Komuro I, Takuji Shirasawa TS, Mizushima N, Otsu K. Inhibition of autophagy in the heart induces age-related cardiomyopathy. Autophagy 2010; **6**: 600-606 [PMID: 20431347 DOI: 10.4161/auto.6.5.11947]
- Russ DW, Boyd IM, McCoy KM, McCorkle KW. Muscle-specificity of age-related changes in markers of autophagy and sphingolipid 21 metabolism. Biogerontology 2015; 16: 747-759 [PMID: 26296420 DOI: 10.1007/s10522-015-9598-4]
- Chang C, Kang P, Liu Y, Huang K, Taylor E, Sagona AP, Nezis IP, Bodmer R, Ocorr K, Bai H. Activin Signaling Regulates Autophagy and 22 Cardiac Aging through mTORC2. *BioRxiv* 2017; 139360 [DOI: 10.1101/139360]
- Ren J, Yang L, Zhu L, Xu X, Ceylan AF, Guo W, Yang J, Zhang Y. Akt2 ablation prolongs life span and improves myocardial contractile 23 function with adaptive cardiac remodeling: role of Sirt1-mediated autophagy regulation. Aging Cell 2017; 16: 976-987 [PMID: 28681509 DOI: 10.1111/acel.126161
- Linton PJ, Gurney M, Sengstock D, Mentzer RM Jr, Gottlieb RA. This old heart: Cardiac aging and autophagy. J Mol Cell Cardiol 2015; 83: 24 44-54 [PMID: 25543002 DOI: 10.1016/j.yjmcc.2014.12.017]
- Kroemer G, Mariño G, Levine B. Autophagy and the integrated stress response. Mol Cell 2010; 40: 280-293 [PMID: 20965422 DOI: 25 10.1016/j.molcel.2010.09.023]
- He C, Bassik MC, Moresi V, Sun K, Wei Y, Zou Z, An Z, Loh J, Fisher J, Sun Q, Korsmeyer S, Packer M, May HI, Hill JA, Virgin HW, 26 Gilpin C, Xiao G, Bassel-Duby R, Scherer PE, Levine B. Exercise-induced BCL2-regulated autophagy is required for muscle glucose homeostasis. Nature 2012; **481**: 511-515 [PMID: 22258505 DOI: 10.1038/nature10758]
- Alim Al-Bari A, Ito Y, Thomes PG, Menon MB, García-Macia M, Fadel R, Stadlin A, Peake N, Faris ME, Eid N, Klionsky DJ. Emerging 27 mechanistic insights of selective autophagy in hepatic diseases. Front Pharmacol 2023; 14: 1149809 [PMID: 37007026 DOI: 10.3389/fphar.2023.1149809]
- 28 Al-Bari MAA, Ito Y, Ahmed S, Radwan N, Ahmed HS, Eid N. Targeting Autophagy with Natural Products as a Potential Therapeutic Approach for Cancer. Int J Mol Sci 2021; 22 [PMID: 34575981 DOI: 10.3390/ijms22189807]
- Eid N, Ito Y, Otsuki Y. The autophagic response to alcohol toxicity: the missing layer. J Hepatol 2013; 59: 398 [PMID: 23624249 DOI: 29 10.1016/j.jhep.2013.03.038]
- Parzych KR, Klionsky DJ. An overview of autophagy: morphology, mechanism, and regulation. Antioxid Redox Signal 2014; 20: 460-473 30 [PMID: 23725295 DOI: 10.1089/ars.2013.5371]
- 31 Koutouroushis C, Sarkar O. Role of Autophagy in Cardiovascular Disease and Aging. Cureus 2021; 13: e20042 [PMID: 34873555 DOI: 10.7759/cureus.20042]
- Wang L, Ye X, Zhao T. The physiological roles of autophagy in the mammalian life cycle. Biol Rev Camb Philos Soc 2019; 94: 503-516 32 [PMID: 30239126 DOI: 10.1111/brv.12464]
- Meijer AJ, Codogno P. Regulation and role of autophagy in mammalian cells. Int J Biochem Cell Biol 2004; 36: 2445-2462 [PMID: 15325584 33 DOI: 10.1016/j.biocel.2004.02.002]
- Sun Q, Fan W, Chen K, Ding X, Chen S, Zhong Q. Identification of Barkor as a mammalian autophagy-specific factor for Beclin 1 and class 34 III phosphatidylinositol 3-kinase. Proc Natl Acad Sci U S A 2008; 105: 19211-19216 [PMID: 19050071 DOI: 10.1073/pnas.0810452105]
- Nishida K, Kyoi S, Yamaguchi O, Sadoshima J, Otsu K. The role of autophagy in the heart. Cell Death Differ 2009; 16: 31-38 [PMID: 35 19008922 DOI: 10.1038/cdd.2008.163]
- Gatica D, Chiong M, Lavandero S, Klionsky DJ. Molecular mechanisms of autophagy in the cardiovascular system. Circ Res 2015; 116: 456-36 467 [PMID: 25634969 DOI: 10.1161/CIRCRESAHA.114.303788]
- Russell RC, Tian Y, Yuan H, Park HW, Chang YY, Kim J, Kim H, Neufeld TP, Dillin A, Guan KL. ULK1 induces autophagy by 37 phosphorylating Beclin-1 and activating VPS34 lipid kinase. Nat Cell Biol 2013; 15: 741-750 [PMID: 23685627 DOI: 10.1038/ncb2757]
- Rubinsztein DC, Mariño G, Kroemer G. Autophagy and aging. Cell 2011; 146: 682-695 [PMID: 21884931 DOI: 10.1016/j.cell.2011.07.030] 38
- Sasaki Y, Ikeda Y, Iwabayashi M, Akasaki Y, Ohishi M. The Impact of Autophagy on Cardiovascular Senescence and Diseases. Int Heart J 39 2017; **58**: 666-673 [PMID: 28966332 DOI: 10.1536/ihj.17-246]
- Bravo-San Pedro JM, Kroemer G, Galluzzi L. Autophagy and Mitophagy in Cardiovascular Disease. Circ Res 2017; 120: 1812-1824 [PMID: 40 28546358 DOI: 10.1161/CIRCRESAHA.117.311082]
- Nicolás-Ávila JA, Lechuga-Vieco AV, Esteban-Martínez L, Sánchez-Díaz M, Díaz-García E, Santiago DJ, Rubio-Ponce A, Li JL, 41 Balachander A, Quintana JA, Martínez-de-Mena R, Castejón-Vega B, Pun-García A, Través PG, Bonzón-Kulichenko E, García-Marqués F, Cussó L, A-González N, González-Guerra A, Roche-Molina M, Martin-Salamanca S, Crainiciuc G, Guzmán G, Larrazabal J, Herrero-Galán E, Alegre-Cebollada J, Lemke G, Rothlin CV, Jimenez-Borreguero LJ, Reyes G, Castrillo A, Desco M, Muñoz-Cánoves P, Ibáñez B, Torres M, Ng LG, Priori SG, Bueno H, Vázquez J, Cordero MD, Bernal JA, Enríquez JA, Hidalgo A. A Network of Macrophages Supports Mitochondrial Homeostasis in the Heart. Cell 2020; 183: 94-109.e23 [PMID: 32937105 DOI: 10.1016/j.cell.2020.08.031]
- Zaglia T, Milan G, Ruhs A, Franzoso M, Bertaggia E, Pianca N, Carpi A, Carullo P, Pesce P, Sacerdoti D, Sarais C, Catalucci D, Krüger M, 42 Mongillo M, Sandri M. Atrogin-1 deficiency promotes cardiomyopathy and premature death via impaired autophagy. J Clin Invest 2014; 124: 2410-2424 [PMID: 24789905 DOI: 10.1172/JCI66339]
- Gong G, Song M, Csordas G, Kelly DP, Matkovich SJ, Dorn GW 2nd. Parkin-mediated mitophagy directs perinatal cardiac metabolic maturation in mice. Science 2015; 350: aad2459 [PMID: 26785495 DOI: 10.1126/science.aad2459]
- Eisenberg T, Abdellatif M, Schroeder S, Primessnig U, Stekovic S, Pendl T, Harger A, Schipke J, Zimmermann A, Schmidt A, Tong M, Ruckenstuhl C, Dammbrueck C, Gross AS, Herbst V, Magnes C, Trausinger G, Narath S, Meinitzer A, Hu Z, Kirsch A, Eller K, Carmona-Gutierrez D, Büttner S, Pietrocola F, Knittelfelder O, Schrepfer E, Rockenfeller P, Simonini C, Rahn A, Horsch M, Moreth K, Beckers J, Fuchs H, Gailus-Durner V, Neff F, Janik D, Rathkolb B, Rozman J, de Angelis MH, Moustafa T, Haemmerle G, Mayr M, Willeit P, von Frieling-Salewsky M, Pieske B, Scorrano L, Pieber T, Pechlaner R, Willeit J, Sigrist SJ, Linke WA, Mühlfeld C, Sadoshima J, Dengjel J, Kiechl S,



- Kroemer G, Sedej S, Madeo F. Cardioprotection and lifespan extension by the natural polyamine spermidine. Nat Med 2016; 22: 1428-1438 [PMID: 27841876 DOI: 10.1038/nm.4222]
- 45 Nakai A, Yamaguchi O, Takeda T, Higuchi Y, Hikoso S, Taniike M, Omiya S, Mizote I, Matsumura Y, Asahi M, Nishida K, Hori M, Mizushima N, Otsu K. The role of autophagy in cardiomyocytes in the basal state and in response to hemodynamic stress. Nat Med 2007; 13: 619-624 [PMID: 17450150 DOI: 10.1038/nm1574]
- Walter S, Atzmon G, Demerath EW, Garcia ME, Kaplan RC, Kumari M, Lunetta KL, Milaneschi Y, Tanaka T, Tranah GJ, Völker U, Yu L, Arnold A, Benjamin EJ, Biffar R, Buchman AS, Boerwinkle E, Couper D, De Jager PL, Evans DA, Harris TB, Hoffmann W, Hofman A, Karasik D, Kiel DP, Kocher T, Kuningas M, Launer LJ, Lohman KK, Lutsey PL, Mackenbach J, Marciante K, Psaty BM, Reiman EM, Rotter JI, Seshadri S, Shardell MD, Smith AV, van Duijn C, Walston J, Zillikens MC, Bandinelli S, Baumeister SE, Bennett DA, Ferrucci L, Gudnason V, Kivimaki M, Liu Y, Murabito JM, Newman AB, Tiemeier H, Franceschini N. A genome-wide association study of aging. Neurobiol Aging 2011; 32: 2109.e15-2109.e28 [PMID: 21782286 DOI: 10.1016/j.neurobiolaging.2011.05.026]
- Zhou J, Force T. Focusing the spotlight on GSK-3 in aging. Aging (Albany NY) 2013; 5: 388-389 [PMID: 23804600 DOI: 10.18632/aging.1005681
- Korolchuk VI, Menzies FM, Rubinsztein DC. A novel link between autophagy and the ubiquitin-proteasome system. Autophagy 2009; 5: 862-48 863 [PMID: 19458478 DOI: 10.4161/auto.8840]
- LaRocca TJ, Hearon CM Jr, Henson GD, Seals DR. Mitochondrial quality control and age-associated arterial stiffening. Exp Gerontol 2014; **58**: 78-82 [PMID: 25034910 DOI: 10.1016/j.exger.2014.07.008]
- Bravo-San Pedro JM, Sica V, Martins I, Anagnostopoulos G, Maiuri C, Kroemer G. Cell-autonomous, paracrine and neuroendocrine 50 feedback regulation of autophagy by DBI/ACBP (diazepam binding inhibitor, acyl-CoA binding protein): the obesity factor. Autophagy 2019; **15**: 2036-2038 [PMID: 31470770 DOI: 10.1080/15548627.2019.1662585]
- Johnson SC, Rabinovitch PS, Kaeberlein M. mTOR is a key modulator of ageing and age-related disease. Nature 2013; 493: 338-345 [PMID: 51 23325216 DOI: 10.1038/nature11861]
- Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. Nat Cell Biol 2011; 13: 132-141 [PMID: 21258367 DOI: 10.1038/ncb2152]
- Dai DF, Rabinovitch PS, Ungvari Z. Mitochondria and cardiovascular aging. Circ Res 2012; 110: 1109-1124 [PMID: 22499901 DOI: 10.1161/CIRCRESAHA.111.246140]
- Ikeda Y, Sciarretta S, Nagarajan N, Rubattu S, Volpe M, Frati G, Sadoshima J. New insights into the role of mitochondrial dynamics and 54 autophagy during oxidative stress and aging in the heart. Oxid Med Cell Longev 2014; 2014: 210934 [PMID: 25132912 DOI: 10.1155/2014/210934]
- Brunk UT, Terman A. Lipofuscin: mechanisms of age-related accumulation and influence on cell function. Free Radic Biol Med 2002; 33: 55 611-619 [PMID: 12208347 DOI: 10.1016/s0891-5849(02)00959-0]
- 56 Cárdenas C, Miller RA, Smith I, Bui T, Molgó J, Müller M, Vais H, Cheung KH, Yang J, Parker I, Thompson CB, Birnbaum MJ, Hallows KR, Foskett JK. Essential regulation of cell bioenergetics by constitutive InsP3 receptor Ca2+ transfer to mitochondria. Cell 2010; 142: 270-283 [PMID: 20655468 DOI: 10.1016/j.cell.2010.06.007]
- Wu X, Zhang T, Bossuyt J, Li X, McKinsey TA, Dedman JR, Olson EN, Chen J, Brown JH, Bers DM. Local InsP3-dependent perinuclear 57 Ca2+ signaling in cardiac myocyte excitation-transcription coupling. J Clin Invest 2006; 116: 675-682 [PMID: 16511602 DOI:
- 58 Yamda J, Ohkusa T, Nao T, Ueyama T, Yano M, Kobayashi S, Hamano K, Esato K, Matsuzaki M. Up-regulation of inositol 1,4,5 trisphosphate receptor expression in atrial tissue in patients with chronic atrial fibrillation. J Am Coll Cardiol 2001; 37: 1111-1119 [PMID: 11263617 DOI: 10.1016/s0735-1097(01)01144-5]
- Decuypere JP, Welkenhuyzen K, Luyten T, Ponsaerts R, Dewaele M, Molgó J, Agostinis P, Missiaen L, De Smedt H, Parys JB, Bultynck G. 59 Ins(1,4,5)P3 receptor-mediated Ca2+ signaling and autophagy induction are interrelated. Autophagy 2011; 7: 1472-1489 [PMID: 22082873] DOI: 10.4161/auto.7.12.17909]
- Aris JP, Alvers AL, Ferraiuolo RA, Fishwick LK, Hanvivatpong A, Hu D, Kirlew C, Leonard MT, Losin KJ, Marraffini M, Seo AY, Swanberg V, Westcott JL, Wood MS, Leeuwenburgh C, Dunn WA Jr. Autophagy and leucine promote chronological longevity and respiration proficiency during calorie restriction in yeast. Exp Gerontol 2013; 48: 1107-1119 [PMID: 23337777 DOI: 10.1016/j.exger.2013.01.006]
- Libert S, Guarente L. Metabolic and neuropsychiatric effects of calorie restriction and sirtuins. Annu Rev Physiol 2013; 75: 669-684 [PMID: 61 23043250 DOI: 10.1146/annurev-physiol-030212-183800]
- Rickenbacher A, Jang JH, Limani P, Ungethüm U, Lehmann K, Oberkofler CE, Weber A, Graf R, Humar B, Clavien PA. Fasting protects 62 liver from ischemic injury through Sirt1-mediated downregulation of circulating HMGB1 in mice. J Hepatol 2014; 61: 301-308 [PMID: 24751831 DOI: 10.1016/j.jhep.2014.04.010]
- Most J, Tosti V, Redman LM, Fontana L. Calorie restriction in humans: An update. Ageing Res Rev 2017; 39: 36-45 [PMID: 27544442 DOI: 63 10.1016/j.arr.2016.08.005]
- Wohlgemuth SE, Julian D, Akin DE, Fried J, Toscano K, Leeuwenburgh C, Dunn WA Jr. Autophagy in the heart and liver during normal 64 aging and calorie restriction. Rejuvenation Res 2007; 10: 281-292 [PMID: 17665967 DOI: 10.1089/rej.2006.0535]
- Roth GS, Mattison JA, Ottinger MA, Chachich ME, Lane MA, Ingram DK. Aging in rhesus monkeys: relevance to human health 65 interventions. Science 2004; 305: 1423-1426 [PMID: 15353793 DOI: 10.1126/science.1102541]
- Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in 66 humans. Proc Natl Acad Sci U S A 2004; 101: 6659-6663 [PMID: 15096581 DOI: 10.1073/pnas.0308291101]
- Postnikoff SDL, Johnson JE, Tyler JK. The integrated stress response in budding yeast lifespan extension. Microb Cell 2017; 4: 368-375 67 [PMID: 29167799 DOI: 10.15698/mic2017.11.597]
- 68 Sheng Y, Lv S, Huang M, Lv Y, Yu J, Liu J, Tang T, Qi H, Di W, Ding G. Opposing effects on cardiac function by calorie restriction in different-aged mice. Aging Cell 2017; 16: 1155-1167 [PMID: 28799249 DOI: 10.1111/acel.12652]
- 69 Ntsapi C, Loos B. Caloric restriction and the precision-control of autophagy: A strategy for delaying neurodegenerative disease progression. Exp Gerontol 2016; 83: 97-111 [PMID: 27473756 DOI: 10.1016/j.exger.2016.07.014]
- Dai DF, Karunadharma PP, Chiao YA, Basisty N, Crispin D, Hsieh EJ, Chen T, Gu H, Djukovic D, Raftery D, Beyer RP, MacCoss MJ, 70 Rabinovitch PS. Altered proteome turnover and remodeling by short-term caloric restriction or rapamycin rejuvenate the aging heart. Aging Cell 2014; 13: 529-539 [PMID: 24612461 DOI: 10.1111/acel.12203]
- Shinmura K, Tamaki K, Sano M, Murata M, Yamakawa H, Ishida H, Fukuda K. Impact of long-term caloric restriction on cardiac senescence:



- caloric restriction ameliorates cardiac diastolic dysfunction associated with aging. J Mol Cell Cardiol 2011; 50: 117-127 [PMID: 20977912 DOI: 10.1016/j.vimcc.2010.10.0181
- Donato AJ, Walker AE, Magerko KA, Bramwell RC, Black AD, Henson GD, Lawson BR, Lesniewski LA, Seals DR. Life-long caloric restriction reduces oxidative stress and preserves nitric oxide bioavailability and function in arteries of old mice. Aging Cell 2013; 12: 772-783 [PMID: 23714110 DOI: 10.1111/acel.12103]
- Stekovic S, Hofer SJ, Tripolt N, Aon MA, Royer P, Pein L, Stadler JT, Pendl T, Prietl B, Url J, Schroeder S, Tadic J, Eisenberg T, Magnes C, Stumpe M, Zuegner E, Bordag N, Riedl R, Schmidt A, Kolesnik E, Verheyen N, Springer A, Madl T, Sinner F, de Cabo R, Kroemer G, Obermayer-Pietsch B, Dengjel J, Sourij H, Pieber TR, Madeo F. Alternate Day Fasting Improves Physiological and Molecular Markers of Aging in Healthy, Non-obese Humans. Cell Metab 2019; 30: 462-476.e6 [PMID: 31471173 DOI: 10.1016/j.cmet.2019.07.016]
- Johnstone A. Fasting for weight loss: an effective strategy or latest dieting trend? Int J Obes (Lond) 2015; 39: 727-733 [PMID: 25540982 74 DOI: 10.1038/ijo.2014.214]
- 75 Castello L, Maina M, Testa G, Cavallini G, Biasi F, Donati A, Leonarduzzi G, Bergamini E, Poli G, Chiarpotto E. Alternate-day fasting reverses the age-associated hypertrophy phenotype in rat heart by influencing the ERK and PI3K signaling pathways. Mech Ageing Dev 2011; **132**: 305-314 [PMID: 21741396 DOI: 10.1016/j.mad.2011.06.006]
- Castello L, Froio T, Maina M, Cavallini G, Biasi F, Leonarduzzi G, Donati A, Bergamini E, Poli G, Chiarpotto E. Alternate-day fasting 76 protects the rat heart against age-induced inflammation and fibrosis by inhibiting oxidative damage and NF-kB activation. Free Radic Biol Med 2010; 48: 47-54 [PMID: 19818847 DOI: 10.1016/j.freeradbiomed.2009.10.003]
- 77 Goodrick CL, Ingram DK, Reynolds MA, Freeman JR, Cider NL. Effects of intermittent feeding upon growth and life span in rats. Gerontology 1982; 28: 233-241 [PMID: 7117847 DOI: 10.1159/000212538]
- 78 Inuzuka Y, Okuda J, Kawashima T, Kato T, Niizuma S, Tamaki Y, Iwanaga Y, Yoshida Y, Kosugi R, Watanabe-Maeda K, Machida Y, Tsuji S, Aburatani H, Izumi T, Kita T, Shioi T. Suppression of phosphoinositide 3-kinase prevents cardiac aging in mice. Circulation 2009; 120: 1695-1703 [PMID: 19822807 DOI: 10.1161/CIRCULATIONAHA.109.871137]
- Brandhorst S, Choi IY, Wei M, Cheng CW, Sedrakyan S, Navarrete G, Dubeau L, Yap LP, Park R, Vinciguerra M, Di Biase S, Mirzaei H, 79 Mirisola MG, Childress P, Ji L, Groshen S, Penna F, Odetti P, Perin L, Conti PS, Ikeno Y, Kennedy BK, Cohen P, Morgan TE, Dorff TB, Longo VD. A Periodic Diet that Mimics Fasting Promotes Multi-System Regeneration, Enhanced Cognitive Performance, and Healthspan. Cell Metab 2015; 22: 86-99 [PMID: 26094889 DOI: 10.1016/j.cmet.2015.05.012]
- Wei M, Brandhorst S, Shelehchi M, Mirzaei H, Cheng CW, Budniak J, Groshen S, Mack WJ, Guen E, Di Biase S, Cohen P, Morgan TE, Dorff T, Hong K, Michalsen A, Laviano A, Longo VD. Fasting-mimicking diet and markers/risk factors for aging, diabetes, cancer, and cardiovascular disease. Sci Transl Med 2017; 9 [PMID: 28202779 DOI: 10.1126/scitranslmed.aai8700]
- 81 Dutta D, Xu J, Dirain ML, Leeuwenburgh C. Calorie restriction combined with resveratrol induces autophagy and protects 26-month-old rat hearts from doxorubicin-induced toxicity. Free Radic Biol Med 2014; 74: 252-262 [PMID: 24975655 DOI: 10.1016/j.freeradbiomed.2014.06.011]
- Makino N, Oyama J, Maeda T, Koyanagi M, Higuchi Y, Tsuchida K. Calorie restriction increases telomerase activity, enhances autophagy, 82 and improves diastolic dysfunction in diabetic rat hearts. Mol Cell Biochem 2015; 403: 1-11 [PMID: 25662949 DOI: 10.1007/s11010-015-2327-0]
- Andres AM, Kooren JA, Parker SJ, Tucker KC, Ravindran N, Ito BR, Huang C, Venkatraman V, Van Eyk JE, Gottlieb RA, Mentzer RM Jr. 83 Discordant signaling and autophagy response to fasting in hearts of obese mice: Implications for ischemia tolerance. Am J Physiol Heart Circ Physiol 2016; **311**: H219-H228 [PMID: 27199111 DOI: 10.1152/ajpheart.00041.2016]
- 84 Godar RJ, Ma X, Liu H, Murphy JT, Weinheimer CJ, Kovacs A, Crosby SD, Saftig P, Diwan A. Repetitive stimulation of autophagylysosome machinery by intermittent fasting preconditions the myocardium to ischemia-reperfusion injury. Autophagy 2015; 11: 1537-1560 [PMID: 26103523 DOI: 10.1080/15548627.2015.1063768]
- 85 Prisco SZ, Eklund M, Moutsoglou DM, Prisco AR, Khoruts A, Weir EK, Thenappan T, Prins KW. Intermittent Fasting Enhances Right Ventricular Function in Preclinical Pulmonary Arterial Hypertension. J Am Heart Assoc 2021; 10: e022722 [PMID: 34747187 DOI: 10.1161/JAHA.121.022722]
- Brewer RA, Collins HE, Berry RD, Brahma MK, Tirado BA, Peliciari-Garcia RA, Stanley HL, Wende AR, Taegtmeyer H, Rajasekaran NS, Darley-Usmar V, Zhang J, Frank SJ, Chatham JC, Young ME. Temporal partitioning of adaptive responses of the murine heart to fasting. Life Sci 2018; 197: 30-39 [PMID: 29410090 DOI: 10.1016/j.lfs.2018.01.031]
- 87 Xu Z, Qin Y, Lv B, Tian Z, Zhang B. Intermittent Fasting Improves High-Fat Diet-Induced Obesity Cardiomyopathy via Alleviating Lipid Deposition and Apoptosis and Decreasing m6A Methylation in the Heart. Nutrients 2022; 14 [PMID: 35057432 DOI: 10.3390/nu14020251]
- Yue XY, Wang XB, Zhao RZ, Jiang S, Zhou X, Jiao B, Zhang L, Yu ZB. Fasting improves tolerance to acute hypoxia in rats. Biochem Biophys 88 Res Commun 2021; **569**: 161-166 [PMID: 34252588 DOI: 10.1016/j.bbrc.2021.06.099]
- Malinowski B, Zalewska K, Węsierska A, Sokołowska MM, Socha M, Liczner G, Pawlak-Osińska K, Wiciński M. Intermittent Fasting in 89 Cardiovascular Disorders-An Overview. Nutrients 2019; 11 [PMID: 30897855 DOI: 10.3390/nu11030673]
- Eid N, Al-Bari MAA, Menon MB. Fasting-induced autophagy in health and disease: history, mechanisms, and benefits. Sharjah First 90 International Conference on Fasting, 2024. Available from: https://www.sharjah.ac.ae/en/Media/Conferences/1FR/Pages/default.aspx



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