

World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2022 September 15; 14(9): 1604-1890



REVIEW

- 1604** Advances in postoperative adjuvant therapy for primary liver cancer
Zeng ZM, Mo N, Zeng J, Ma FC, Jiang YF, Huang HS, Liao XW, Zhu GZ, Ma J, Peng T
- 1622** Immunotherapy for nonalcoholic fatty liver disease-related hepatocellular carcinoma: Lights and shadows
Costante F, Airola C, Santopaolo F, Gasbarrini A, Pompili M, Ponziani FR
- 1637** Emerging role of caldesmon in cancer: A potential biomarker for colorectal cancer and other cancers
Alnuaimi AR, Nair VA, Malhab LJB, Abu-Gharbieh E, Ranade AV, Pintus G, Hamad M, Busch H, Kirfel J, Hamoudi R, Abdel-Rahman WM

MINIREVIEWS

- 1654** Liquid biopsy to detect resistance mutations against anti-epidermal growth factor receptor therapy in metastatic colorectal cancer
Valenzuela G, Burotto M, Marcelain K, González-Montero J
- 1665** Implication of gut microbiome in immunotherapy for colorectal cancer
Koustas E, Trifylli EM, Sarantis P, Papadopoulos N, Aloizos G, Tsagarakis A, Damaskos C, Garmpis N, Garmpi A, Papavassiliou AG, Karamouzis MV

ORIGINAL ARTICLE**Basic Study**

- 1675** Potential of six-transmembrane epithelial antigen of the prostate 4 as a prognostic marker for colorectal cancer
Fang ZX, Li CL, Chen WJ, Wu HT, Liu J

Case Control Study

- 1689** Inverse relations between *Helicobacter pylori* infection and risk of esophageal precancerous lesions in drinkers and peanut consumption
Pan D, Sun GJ, Su M, Wang X, Yan QY, Song G, Wang YY, Xu DF, Wang NN, Wang SK

Retrospective Cohort Study

- 1699** Prognostic impact of tumor deposits on overall survival in colorectal cancer: Based on Surveillance, Epidemiology, and End Results database
Wu WX, Zhang DK, Chen SX, Hou ZY, Sun BL, Yao L, Jie JZ
- 1711** Consolidation chemotherapy with capecitabine after neoadjuvant chemoradiotherapy in high-risk patients with locally advanced rectal cancer: Propensity score study
Sheng XQ, Wang HZ, Li S, Zhang YZ, Geng JH, Zhu XG, Quan JZ, Li YH, Cai Y, Wang WH

Retrospective Study

- 1727 Efficacy and safety of computed tomography-guided microwave ablation with fine needle-assisted puncture positioning technique for hepatocellular carcinoma
Hao MZ, Hu YB, Chen QZ, Chen ZX, Lin HL
- 1739 Clinicopathological characterization of ten patients with primary malignant melanoma of the esophagus and literature review
Zhou SL, Zhang LQ, Zhao XK, Wu Y, Liu QY, Li B, Wang JJ, Zhao RJ, Wang XJ, Chen Y, Wang LD, Kong LF
- 1758 Endoscopic debulking resection with additive chemoradiotherapy: Optimal management of advanced inoperable esophageal squamous cell carcinoma
Ren LH, Zhu Y, Chen R, Shrestha Sachin M, Lu Q, Xie WH, Lu T, Wei XY, Shi RH
- 1771 Nomogram for predicting the prognosis of tumor patients with sepsis after gastrointestinal surgery
Chen RX, Wu ZQ, Li ZY, Wang HZ, Ji JF
- 1785 Efficacy and safety of laparoscopic radical resection following neoadjuvant therapy for pancreatic ductal adenocarcinoma: A retrospective study
He YG, Huang XB, Li YM, Li J, Peng XH, Huang W, Tang YC, Zheng L

Observational Study

- 1798 To scope or not - the challenges of managing patients with positive fecal occult blood test after recent colonoscopy
Rattan N, Willmann L, Aston D, George S, Bassan M, Abi-Hanna D, Anandabaskaran S, Ermerak G, Ng W, Koo JH
- 1808 Clinical implications of interleukins-31, 32, and 33 in gastric cancer
Liu QH, Zhang JW, Xia L, Wise SG, Hambly BD, Tao K, Bao SS
- 1823 Construction and analysis of an ulcer risk prediction model after endoscopic submucosal dissection for early gastric cancer
Gong SD, Li H, Xie YB, Wang XH
- 1833 Percutaneous insertion of a novel dedicated metal stent to treat malignant hilar biliary obstruction
Cortese F, Acquafredda F, Mardighian A, Zurlo MT, Ferraro V, Memeo R, Spiliopoulos S, Inchingolo R

EVIDENCE-BASED MEDICINE

- 1844 Prediction of gastric cancer risk by a polygenic risk score of *Helicobacter pylori*
Wang XY, Wang LL, Liang SZ, Yang C, Xu L, Yu MC, Wang YX, Dong QJ

META-ANALYSIS

- 1856 Dissecting novel mechanisms of hepatitis B virus related hepatocellular carcinoma using meta-analysis of public data
Aljabban J, Rohr M, Syed S, Cohen E, Hashi N, Syed S, Khorfan K, Aljabban H, Borkowski V, Segal M, Mukhtar M, Mohammed M, Boateng E, Nemer M, Panahiazar M, Hadley D, Jalil S, Mumtaz K
- 1874 Prognostic and clinicopathological value of Twist expression in esophageal cancer: A meta-analysis
Song WP, Wang SY, Zhou SC, Wu DS, Xie JY, Liu TT, Wu XZ, Che GW

LETTER TO THE EDITOR

1886 Nutrition deprivation affects the cytotoxic effect of CD8 T cells in hepatocellular carcinoma

Zhang CY, Liu S, Yang M

ABOUT COVER

Editorial Board Member of *World Journal of Gastrointestinal Oncology*, Luigi Marano, MD, PhD, Associate Professor, Department of Medicine, Surgery, and Neurosciences, University of Siena, Siena 53100, Italy. luigi.marano@unisi.it

AIMS AND SCOPE

The primary aim of *World Journal of Gastrointestinal Oncology* (*WJGO*, *World J Gastrointest Oncol*) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, *etc.*

INDEXING/ABSTRACTING

The *WJGO* is now abstracted and indexed in PubMed, PubMed Central, Science Citation Index Expanded (SCIE, also known as SciSearch®), Journal Citation Reports/Science Edition, Scopus, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 edition of Journal Citation Reports® cites the 2021 impact factor (IF) for *WJGO* as 3.404; IF without journal self cites: 3.357; 5-year IF: 3.250; Journal Citation Indicator: 0.53; Ranking: 162 among 245 journals in oncology; Quartile category: Q3; Ranking: 59 among 93 journals in gastroenterology and hepatology; and Quartile category: Q3. The *WJGO*'s CiteScore for 2021 is 3.6 and Scopus CiteScore rank 2021: Gastroenterology is 72/149; Oncology is 203/360.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Ying-Yi Yuan*; Production Department Director: *Xiang Li*; Editorial Office Director: *Jia-Ru Fan*.

NAME OF JOURNAL

World Journal of Gastrointestinal Oncology

ISSN

ISSN 1948-5204 (online)

LAUNCH DATE

February 15, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Monjur Ahmed, Florin Burada

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/1948-5204/editorialboard.htm>

PUBLICATION DATE

September 15, 2022

COPYRIGHT

© 2022 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Implication of gut microbiome in immunotherapy for colorectal cancer

Evangelos Koustas, Eleni-Myrto Trifylli, Panagiotis Sarantis, Nikolaos Papadopoulos, Georgios Aloizos, Ariadne Tsagarakis, Christos Damaskos, Nikolaos Garmpis, Anna Garmpi, Athanasios G Papavassiliou, Michalis V Karamouzis

Specialty type: Oncology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0

Grade B (Very good): B, B

Grade C (Good): 0

Grade D (Fair): 0

Grade E (Poor): 0

P-Reviewer: Manojlovic N, Serbia; Popovic LS, Serbia

Received: March 20, 2022

Peer-review started: March 20, 2022

First decision: April 17, 2022

Revised: May 9, 2022

Accepted: July 31, 2022

Article in press: July 31, 2022

Published online: September 15, 2022



Evangelos Koustas, Eleni-Myrto Trifylli, Panagiotis Sarantis, Athanasios G Papavassiliou, Michalis V Karamouzis, Department of Biological Chemistry, Medical School, National and Kapodistrian University of Athens, Athens 11527, Greece

Nikolaos Papadopoulos, Georgios Aloizos, 1st Department of Internal Medicine, 417 Army Share Fund Hospital of Athens, Athens 11521, Attica, Greece

Ariadne Tsagarakis, Beth Israel Deaconess Medical Center, Boston, MA 02215, United States

Christos Damaskos, N.S. Christeas Laboratory of Experimental Surgery and Surgical Research, Medical School, National and Kapodistrian University of Athens, Athens 11527, Greece

Nikolaos Garmpis, Second Department of Propedeutic Surgery, Laiko General Hospital, Medical School, National and Kapodistrian University of Athens, Athens 11527, Greece

Anna Garmpi, First Department of Pathology, Medical School, National and Kapodistrian University of Athens, Athens 11527, Greece

Corresponding author: Evangelos Koustas, MD, PhD, Doctor, Senior Research Fellow, Department of Biological Chemistry, Medical School, National and Kapodistrian University of Athens, 75 M. Asias Street, Athens 11527, Greece. vang.koustas@gmail.com

Abstract

Colorectal cancer (CRC) constitutes the third most frequently reported malignancy in the male population and the second most common in women in the last two decades. Colon carcinogenesis is a complex, multifactorial event, resulting from genetic and epigenetic aberrations, the impact of environmental factors, as well as the disturbance of the gut microbial ecosystem. The relationship between the intestinal microbiome and carcinogenesis was relatively undervalued in the last decade. However, its remarkable effect on metabolic and immune functions on the host has been in the spotlight as of recent years. There is a strong relationship between gut microbiome dysbiosis, bowel pathogenicity and responsiveness to anti-cancer treatment; including immunotherapy. Modifications of bacteriome consistency are closely associated with the immunologic response to immunotherapeutic agents. This condition that implies the necessity of gut microbiome manipulation. Thus, creating an optimal response for CRC patients to immunotherapeutic agents. In this paper, we will review the current literature

observing how gut microbiota influence the response of immunotherapy on CRC patients.

Key Words: Colorectal cancer; Gut microbiome; Immunotherapy; Checkpoint inhibitors; Tumor micro-environment

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Colorectal cancer (CRC) constitutes the third most frequent malignancy. CRC is a complex, multistep process. The impact of environmental factors as well as the disturbance of the gut microbial ecosystem is associated with CRC development. There is a strong relationship between the gut microbiome and resistance to immunotherapy.

Citation: Koustas E, Trifylli EM, Sarantis P, Papadopoulos N, Aloizos G, Tsagarakis A, Damaskos C, Garmpis N, Garmpi A, Papavassiliou AG, Karamouzis MV. Implication of gut microbiome in immunotherapy for colorectal cancer. *World J Gastrointest Oncol* 2022; 14(9): 1665-1674

URL: <https://www.wjgnet.com/1948-5204/full/v14/i9/1665.htm>

DOI: <https://dx.doi.org/10.4251/wjgo.v14.i9.1665>

INTRODUCTION

Colorectal cancer (CRC) constitutes the third most frequently reported malignancy in the male population and the second most common in women in the last several decades, based off GLOBACAN epidemiological data[1]. Colon carcinogenesis is a complex, multifactorial event composed of genetic and epigenetic aberrations, which additionally causes the disturbance of gut homeostasis resulting from gut microbiota modifications[2]. The microbiome constitutes a multiplex ecosystem of microorganisms located in the gastrointestinal tract of many species, including humans[3].

The relationship between the intestinal microbiome and disease development, including carcinogenesis, was relatively undervalued in the last decade. However, the interrelation of gut microbiota with the main functions of the host has recently been in the spotlight[4]. The digestive tract contains the largest amount of microbiota colonization among other anatomical regions, accounting for approximately 70% of the human microbiota make-up[5], including viral and bacterial microorganisms, archaea and fungi[6,7]. The proximal parts of the GI tract, including the stomach and small intestine, present few microbiota species whereas the distal part, the colon, presents the largest number of species (microorganisms) in the colonic substance[7]. The six main phyla of the gut microbiome (90% of the population) include[8]: Bacteroidetes, Actinobacteria, Firmicutes, Proteobacteria, Verrucomicrobia, and Euryarchaeota[9]. Of all the genera found in the human gut, Bacteroides makes up the majority of the population (30%)[10], implying its significant effect on the human functional system. Additionally, many genera from the Firmicutes phylum compose a high amount of the intestinal substance, such as lactobacillus, Clostridium, Faecalibacterium, Eubacterium and Ruminococcus[11]. The application of metagenomics on fecal specimens has given the opportunity for microbiome quantification and analysis, and potentially its use as a potent diagnostic tool[12].

LITERATURE SEARCH

PubMed was searched to identify studies on gut microbiome, immunotherapy and CRC. PubMed and Reference Citation Analysis (<https://www.referencecitationanalysis.com/>) were searched to identify studies on gut microbiome, immunotherapy and CRC. The literature review was completed on February 28, 2022. The following search terms were applied: "Colorectal cancer", "Immunotherapy", "Checkpoint inhibitors," "Tumor microenvironment," and "Gut microbiome". The reference lists of all related articles were screened for other potentially relevant studies. The search citation analysis is presented in the reference list. Finally, the authors similarly reviewed the reference lists of eligible articles to identify further eligible articles, books and other forms of publication. Publications that are written in any other language other than English were excluded. Publications of abstracts were also excluded.

THE FUNCTIONAL ROLE OF THE GUT MICROBIOME

Gut microbiota exhibits diverse functions in the human organism and are responsible for many metabolic processes and biosynthesis. Vitamin synthesis constitutes one of the key roles of gut microbiota, such as riboflavin, vitamin B1, biotin, vitamin K and cobalamin[13]. They also have a crucial role in non-digestible carbohydrate metabolism; to transform them into short-chain fatty acids (SCFAs), such as butyric acid, acetic acid and propionic[14], which are produced by the main phyla of bacteriome, this includes Bacteroidetes and Firmicutes[15]. Alteration of the above metabolic process leads to modification of the fatty acid production and overall metabolic imbalance[16]. Along with their involvement in vitamin and short fatty acids synthesis, they take part in bile acid production[17]. Neuromodulators are also produced by gut microbiota, with a significant implication for the gut-brain axis, which includes the peripheral and central nervous systems as well as the enteric nervous system [18]. Many neurological and psychiatric disorders are closely connected with the gut microbiome. This can occur because they are responsible for synthesizing many pro-inflammatory cytokines, amyloids and liposaccharides[18]. Based on metagenomics, genome disturbance and dysbiotic flora can cause a predisposition to develop a number of malignancies[19], including non-neoplastic disorders, such as atopy, functional intestinal disturbances, like irritable bowel syndrome (IBS), inflammatory bowel disease (IBD) and metabolic syndrome[20,21].

There is a strong relationship between gut microbiome dysbiosis and bowel pathogenicity. In the case of the bowel, functional disorders such as IBS have many studies illustrating an altered consistency of the bacteriome, with both an increase or decrease in the quantity of many bacteria. It is specifically observed as an aberrant increase of *Ruminococcus*, *Firmicutes*, and *Clostridium spp.* with an abnormal decrease of *Ruminococcus albus* and *callidus*, *Bacteroides fragilis* and *bulgatus*[18]. Additionally, the overproduction of SCFAs that deregulate the secretion of serotonin from the enteroendocrine cells leads to increased bowel movements and fermentation. This causes the symptomatology associated with meteorism[22]. Patients who suffer from organic bowel diseases, such as IBD, Ulcerative colitis and Crohn's disease (CD) have been observed to have an altered microbiome. The modification of the gut microbiome is closely associated to dietary habits[23]. Patients with CD specifically demonstrate increased amounts of *Neisseria caecorrodens*, *E. coli* and proteobacteria[24], while enhanced amounts of fungal species such as *Candida albicans*, *Cyberlindnera jadinii* and *Saccharomyces cerevisiae* can also be observed[25]. In addition, a decreased number of some bacterial taxa, such as Firmicutes, *Faecalibacterium prausnitzii*, Bacteroidetes and Roseburia, is observed[26]. Dietary habits that include a high amount of fruit and vegetable consumption can lower the risk for developing CD[27].

Intestinal epithelial cells are closely interrelated with the immune system *via* the existence of goblet and Paneth cells and their products. Goblet cells are located in intestinal mucosa and have a crucial role in producing mucus. Paneth cells are located in the crypts of Lieberkühn, secreting various immunomodulatory peptides with antimicrobial qualities[28]. Moreover, bacterial metabolites also take place in immune responses *via* the production of SCFAs and are closely associated with innate immunity and antibody production[29].

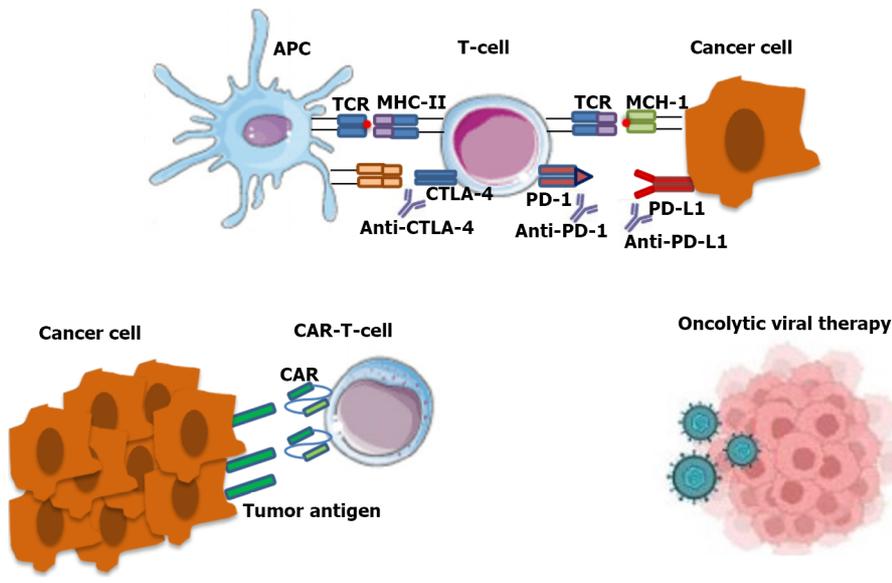
Immunotherapy constitutes a significant therapeutic option, including immune checkpoint inhibitors, cancer vaccines and chimeric antigen receptor-T cells[30]. This treatment modality makes use of the immune responses to create an anti-neoplastic effect. The main therapeutic agents include the following monoclonal antibodies: (1) Anti-cytotoxic T-lymphocyte antigen-4 (anti-CTLA-4); (2) Anti-programmed cell death 1 ligand 1 (anti-PD-L1); and (3) Anti-programmed cell death protein 1 (anti-PD-1)[28,31]. The principal advantage of immunotherapeutic agents includes their aimed action on malignant cells appears in [Figure 1](#).

This therapeutic modality is currently selected as an anti-cancer treatment specifically in cases of tumors that are characterized by high microsatellite instability (MSI-H)[32]. Tumors that present MSI-H arise from a defective DNA mismatch repair (MMR) mechanism that leads to the accumulation of genetic mutations. This can be seen in the case of mutant MSH2, PMS2, MSH6 and MLH1. Or by epigenetic aberration, such as genome hyper-methylation[33]. There are many reports that gut microbiota influences the response to anti-cancer treatment including immunotherapy[34]. It is observed that a significant number of CRC patients that lacked a specific taxa in their bacteriome, presented a limited response to immunotherapy agents such as anti-PD1. This condition implies the use for more personalized anti-cancer treatments that can prove to be potent. In this paper, we review the current literature on how gut microbiota influences the response of CRC patients to immunotherapy[35].

THE ROLE OF MICROBIOME IN COLON CARCINOGENESIS

There are many studies about the implication of gut microbiota in immunotherapeutic agents including immune checkpoint inhibitors for melanoma. Fewer studies exist about its role in CRC treatment management.

Modifications in the gut microbiome and microbial metabolites have been involved in many pathological processes and diseases, including colon carcinogenesis[36]. Many intestinal bacterial



DOI: 10.4251/wjgo.v14.i9.1665 Copyright ©The Author(s) 2022.

Figure 1 Mechanism of action of both anti anti-cytotoxic T-lymphocyte antigen-4 and anti-programmed cell death protein 1/programmed cell death 1 ligand 1 check point inhibitors. In the tumor microenvironment, antigen-presenting cells (APCs), such as dendritic cells processed specific tumor peptides (TAAs) and complexed them to major histocompatibility complex (MHC) molecules. Then, APC migrated to T cell-dependent areas of tumor presented TAA to naïve or quiescent T cells. Checkpoint inhibitor, such as anti-programmed cell death protein 1 (anti-PD-1)/anti-programmed cell death 1 ligand 1 (anti-PD-L1) and/or anti-cytotoxic T-lymphocyte antigen-4 (anti-CTLA-4) on tumor cells, lead to re-activation of immune responses. The anti-PD-1 or anti-PD-L1 blocking by monoclonal antibodies (as nivolumab, pembrolizumab for PD-1 or atezolizumab for PD-L1) ipilimumab restore CD28 pro-activity signaling and restore effective anti-tumor T lymphocyte responses. The anti-CTLA-4 blocking by monoclonal antibodies as ipilimumab restore CD28 pro-activity signaling and result in effective anti-tumor T lymphocyte responses. The binding of PD-L1 to PD-1 and CTLA-4 to B7 keeps T cells from killing tumor cells in the body. Blocking the binding with an immune checkpoint inhibitor allows the T cells to kill tumor cells (upper panel). Chimeric antigen receptor (CAR) T cells are T cells that have been genetically engineered to produce an artificial T cell receptor for use in immunotherapy. CARs are receptor proteins that have been engineered to give T cells the new ability to target a specific protein (lower panel).

products have been implicated in malignant states in the intestinal tract[37]. Several studies demonstrate the presence of an altered microbiome either in CRC patients’ fecal specimens or in malignant tissues compared to healthy patients[38]. These alterations in the microbiome which take place in the initial steps of CRC development can be utilized as predictive biomarkers as well as microbial diagnostic gene markers. This can be utilized in patients with an increased risk of developing colon adenomas that can potentially lead to CRC[39].

Environmental factors have a high influence on the gut microbiome along with idiosyncratic factors [40] which subsequently induce carcinogenesis and CRC development *via* the overgrowth of particular microbial species in the flora[41]. The formulation of colonic microbial substances is closely related to modifiable factors such as eating behavior and style of living[42]. While there is a key role in the metabolism of nutrients[43], there is also a diversity of environmental risk factors that are associated with colorectal carcinogenesis such as obesity, tobacco use, alcohol consumption and prepared meat products[44].

Many studies demonstrate the implication of specific bacterial taxa in carcinogenesis, such as *Enterococcus faecalis*, *Helicobacter hepaticus*, *Bacteroides fragilis* and *Fusobacterium nucleatum*. The products of the previously mentioned microbes lead to genomic alterations[45]. While in the case of the *Fusobacterium nucleatum*, the carcinogenesis indirectly occurs *via* the perpetual secretion of pro-inflammatory cytokines [46]. This phenomenon implies the close interrelation of the microbiome with immune response and metabolic processes[47].

There is a notable reduction of genera from the Firmicutes phylum, which produce a significant metabolite, the alleged butyrate. An enhanced reproduction of specific phyla, such as *Bacteroides fragilis*, *Peptostreptococcus stomatis* as well as *Tarvi monas micra*, *Fusobacterium nucleatum*[48] and *Solobacterium moorei*[49]. Additionally, there are reports that show an increased amount of *Enterococcus*, *Escherichia coli*, *Klebsiella* and *Streptococcus*, as well as a decrease in *Rothia*[2].

There is considerable evidence that CRC development is closely associated with the presence of Fusobacteriaceae family members, such as *Fusobacterium nucleatum*, *necrophorum* and *mortiferum*[37] *via* a mechanism that was reportedly observed in mice[50].

Generally, dysbiosis which includes the modification of microbial taxa in the gut ecosystem leads either to a limited variety of microbiota or the overgrowth of microbes. This can further lead to the development of opportunistic infections[51], destruction of the intestinal epithelial barrier, bacterial translocation to the mesenteric lymph nodes or the circulatory system, ultimately leading to a local and

systemic inflammatory response[52].

Recruitment of T lymphocytes is observed in CRC malignant tissues[53] *via* the secretion of chemotactic cytokines. This is further related to an abundance in proteobacteria Ruminococcaceae, *B. fragilis* and *E.coli*. Alternatively, a high number of Fusobacterias is associated with a dismal prognosis. In *in vitro* it has been observed to express an increased number of recruited T cells and inflammatory modulators [interleukin (IL)-6, IL-8, IL-1][54], an inhibitory effect on Natural killer cells, as well as tumor-infiltrating lymphocytes[55]. Although *Fusobacterium nucleatum* is normally associated with a worse prognosis, it constitutes a promoter for differentiation in regulatory T cells leading to a decrease in expression of scurfin or forkheadbox P3 which is correlated to prolonged survival[56].

IMMUNOTHERAPY IN CRC

The therapeutic management of CRC is considered quite challenging due to the complex molecular basis including genetic and epigenetic alterations[57]. In recent years, immunotherapeutic agents are utilized for tumors that present high MSI-H which results from a defective DNA MMR or epigenetic modification[33]. An epigenetic aberration is genome hyper-methylation in addition to mutant genes such as PMS2, MLH1 as well as MSH2 and MSH6[58]. In the case of MSI-H colorectal tumors, there is evident methylation of CpG islands in the promoter of the BRAF proto-oncogene[59]. It is observed that patients with BRAF and RAS genetic mutations present resistance to immunotherapeutic treatments with a limited enhancement of survival[60]. It can occur in cases of epidermal growth factor receptor inhibitors, like cetuximab, as well as Panitumumab[61]. In comparison with MSI tumors, the microsatellite stable tumors present a more aggressive phenotype and poor prognosis[62]. Immunotherapeutic agents, such as pembrolizumab are commonly used in cases of chemo-resistant advanced colorectal malignant tumors despite the existence or lack of either MMR or MSI-H based off the KEYNOTE 028 clinical trial[63]. For tumors with MMR phenotype, the utilization of nivolumab alone or with ipilimumab is highly recommended[47]. The administration of cancer vaccines in CRC is still under study and it is limited solely to cases of end-stage CRC[64]. Talimogene laherparepvec vaccine uses Herpes virus type-1 as a vector which targets the *GM-CSF* gene. The combination of systemic use of atezolizumab (anti-PD-L1 immunotherapeutic agent) with the above vaccine is currently under assessment for tumors with microsatellite stability[63] or as a monotherapy in secondary liver cancer [65].

Tumor microenvironment and microbiome in CRC

Tumor microenvironment (TME) includes multiple types of cells, such as fibroblasts, immune cells, endothelial and stromal cells[66]. TME demonstrates a significant role in immune responses, particularly in CRC, and constitutes as a therapeutic target for many anti-cancer agents[67]. The stroma around the tumor has a key role in resistance to chemotherapy due to the fact that it includes a heterogeneous population of cells with various levels of differentiation. This contributes to invasive tumor behavior and dissemination. This is shown in the case of tumor-associated macrophages and cancer-associated fibroblasts. Both of these are related to a dismal prognosis and neoangiogenesis[68,69], as well as Myeloid-derived suppressor cells which are also implicated in tumor progression and invasion. Their effect is under the regulation of tumoral products like chemokine (C-C motif) ligand 2 and 5 (CCL2 and CCL5)[70].

It was previously stated that the gut microbiota exhibited various effects on the differentiation mechanism and tumor development. While they influence the tumor response to immunotherapeutics [71], the existence of intra-tumoral bacteria is reported in many solid tumors, especially in breast cancer. It was demonstrated that the microbiome is particular for each kind of malignant tumor presenting distinct metabolic functions[72]. Based on data that was collected by whole-transcriptome analysis, there is a distinct microbiome correlated with different malignant tumors, implying a specific microbial profile for each type of cancer[73]. Additionally, TME has a crucial role in the existence and multiplication of intra-tumoral bacteria[74]. Many studies illustrate the close relationship between immunotherapy and gut microbiota, and their implication in the anti-tumor mechanism such as immune-checkpoint inhibitors[72].

THE IMPLICATION OF GUT MICROBIOME IN IMMUNOTHERAPY

Resistance to immunotherapy is difficult to overcome in clinical practice[31]. Manipulation of gut microbiota constitutes a promising method for reducing the resistance to therapeutic agents. This is implied by the notable effect of intestinal microbial products on the malignant tumor where they could also be considered cancer-driving molecules[75].

Experimental studies on mice have shown that bacteria have a crucial role in the anti-cancer immune response. While the response was limited in the case of germ-free mice[28], it was primarily reported that intestinal microbiota have a significant role in the response especially to immune checkpoint

inhibitors. However, the previous observation was also demonstrated in humans when an immune checkpoint blockade was applied[28]. In mouse-model studies, fecal microbial transplantation (FMT) from mice that presented immune-responsive microbiota, to germ-free mice, provided a better anti-neoplastic response and tumor growth management. This result is associated with an increased amount of cytotoxic T lymphocytes (CD8+) in TME[76]. Whereas the transfer of fecal samples, including microbiota prone for carcinogenesis, provides the opposite results to physiological mice[77]. However, the correlation of the anti-tumor response with external factors must be taken into consideration.

Alterations in the consistency of bacteriome were reported in cases of patients with an active response to PD-1 inhibitors. More specifically, these patients presented a higher amount of *Enterococcus faecium*, *Bifidobacterium longum* and *Collinsella aerofaciens*. Fecal specimens that presented the above microbial taxa were characterized as “responder” stool samples and were transferred *via* FMT to germ-free mice. Subsequently, the germ-free mice started to express the stool phenotype of the responders[28].

Based on various human and animal-model cohort studies, intestinal microbiota could not only have been beneficial but also toxic effects on immune checkpoint inhibition[78]. Reduced toxicity was observed in specimens where Bacteroidetes genera were in abundance. Although they relate to unresponsiveness to immune checkpoint inhibitors (ICIs), in contrast to Firmicutes, and especially in the case of Ruminococaceae, they were not only responsive to ICIs but also presented toxic effects. In cases of overgrown *Faecalibacterium prausnitzii*, patients had an increased risk of presenting colitis related with CTLA-4 inhibitors[79,80].

Manipulation of intestinal microbiota for immunotherapy-response improvement

Based on all the characteristics of the intestinal microbiota, they can either promote the anti-neoplastic response or induce inflammation and carcinogenesis[81]. A reduced anti-cancer response in the host was observed in germ-free mice or with antibiotic administration (broad-spectrum)[28,35]. In cases with urinary tract malignancies and lung cancer, antibiotics had a harmful effect on anti-PD1/PD-L1 treatment[35] in comparison to cyclophosphamide which presented a promoting effect on the overgrowth of *Barnesiella intestine hominis* in the intestinal tract and a stimulatory effect on anti-cancer immune response[82].

However, the manipulation of microbiota and utilization of antibiotics for the killing of bacteria is detrimental to the response to immunotherapeutic agents. This method includes the risk of killing favorable bacterial species. To avoid the non-elective effect of antibiotics, bacteriophage therapy is administered which permits a selective elimination of unfavorable bacteria[83].

Lastly, environmental and lifestyle habits could potentially alter the gut microbiome. These include physical exercise, proper dietary habits, sleep patterns, as well as *via* the utilization of FMT[84]. Bacteriotherapy or FMT includes the transferring of beneficial bacterial species such as Bacteroides, Bifidobacteria, *E. hirae* and *Akkermansia mucini philia*[85].

CONCLUSION

The relationship between the intestinal microbiome and disease development, such as carcinogenesis, was underestimated in the last decades. Nevertheless, the crucial role of intestinal microbiota has been in the spotlight as of recent years. Not only for their significant influence on the main metabolic functions of the host but also on the immune and anti-tumor responses. Immunotherapeutic agents are commonly used specifically for cases with chemo-resistant advanced colorectal malignant tumors. The implication of gut microbiota in the anti-cancer immune response is still under research. However, there are many reports supporting that the lack of specific bacterial taxa in CRC patients leads to a limited response to immunotherapy or complete unresponsiveness with the presence of specific phyla that could promote the anti-cancer response. Based on various human and animal-model cohort studies, intestinal microbiota could not only have beneficial effects on immune checkpoint inhibition but also have detrimental effects. The aforementioned phenomenon illustrates the necessity for the manipulation of intestinal microbiota. Specifically for the highest anti-neoplastic immune response, either *via* bacteriophage therapy or lifestyle habits modifications as well as FMT. Further research regarding the implication of gut microbiome on immunotherapy responses is needed for the identification of additional druggable targets, along with the manipulation of intestinal microbiota to achieve an optimal therapeutic response personalized for each patient.

FOOTNOTES

Author contributions: All authors participated in the writing and editing of the manuscript.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by

external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: Greece

ORCID number: Evangelos Koustas 0000-0003-0583-0540; Eleni-Myrto Trifylli 0000-0002-0080-9032; Panagiotis Sarantis 0000-0001-5848-7905; Nikolaos Papadopoulos 0000-0002-8702-1685; Georgios Aloizos 0000-0002-0017-0718; Christos Damaskos 0000-0002-5454-2564; Nikolaos Garmpis 0000-0001-9980-0056; Anna Garmpi 0000-0003-0258-5965; Athanasios G Papavassiliou 0000-0001-5803-4527; Michalis V Karamouzis 0000-0003-1369-8201.

S-Editor: Fan JR

L-Editor: Filipodia

P-Editor: Fan JR

REFERENCES

- 1 **Song M**, Chan AT, Sun J. Influence of the Gut Microbiome, Diet, and Environment on Risk of Colorectal Cancer. *Gastroenterology* 2020; **158**: 322-340 [PMID: 31586566 DOI: 10.1053/j.gastro.2019.06.048]
- 2 **Quigley EM**. Gut bacteria in health and disease. *Gastroenterol Hepatol (N Y)* 2013; **9**: 560-569 [PMID: 24729765]
- 3 **Gagnière J**, Raisch J, Veziat J, Barnich N, Bonnet R, Buc E, Bringer MA, Pezet D, Bonnet M. Gut microbiota imbalance and colorectal cancer. *World J Gastroenterol* 2016; **22**: 501-518 [PMID: 26811603 DOI: 10.3748/wjg.v22.i2.501]
- 4 **Tilg H**, Adolph TE, Gerner RR, Moschen AR. The Intestinal Microbiota in Colorectal Cancer. *Cancer Cell* 2018; **33**: 954-964 [PMID: 29657127 DOI: 10.1016/j.ccell.2018.03.004]
- 5 **Saus E**, Iraola-Guzmán S, Willis JR, Brunet-Vega A, Gabaldón T. Microbiome and colorectal cancer: Roles in carcinogenesis and clinical potential. *Mol Aspects Med* 2019; **69**: 93-106 [PMID: 31082399 DOI: 10.1016/j.mam.2019.05.001]
- 6 **Passos MDCF**, Moraes-Filho JP. Intestinal microbiota in digestive diseases. *Arq Gastroenterol* 2017; **54**: 255-262 [PMID: 28723981 DOI: 10.1590/S0004-2803.201700000-31]
- 7 **Shapira M**. Gut Microbiotas and Host Evolution: Scaling Up Symbiosis. *Trends Ecol Evol* 2016; **31**: 539-549 [PMID: 27039196 DOI: 10.1016/j.tree.2016.03.006]
- 8 **Feng Q**, Chen WD, Wang YD. Gut Microbiota: An Integral Moderator in Health and Disease. *Front Microbiol* 2018; **9**: 151 [PMID: 29515527 DOI: 10.3389/fmicb.2018.00151]
- 9 **Gao R**, Gao Z, Huang L, Qin H. Gut microbiota and colorectal cancer. *Eur J Clin Microbiol Infect Dis* 2017; **36**: 757-769 [PMID: 28063002 DOI: 10.1007/s10096-016-2881-8]
- 10 **Jandhyala SM**, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Nageshwar Reddy D. Role of the normal gut microbiota. *World J Gastroenterol* 2015; **21**: 8787-8803 [PMID: 26269668 DOI: 10.3748/wjg.v21.i29.8787]
- 11 **Rinninella E**, Raoul P, Cintoni M, Franceschi F, Miggiano GAD, Gasbarrini A, Mele MC. What is the Healthy Gut Microbiota Composition? *Microorganisms* 2019; **7** [PMID: 30634578 DOI: 10.3390/microorganisms7010014]
- 12 **Zeller G**, Tap J, Voigt AY, Sunagawa S, Kultima JR, Costea PI, Amiot A, Böhm J, Brunetti F, Habermann N, Hercog R, Koch M, Luciani A, Mende DR, Schneider MA, Schrotz-King P, Tournigand C, Tran Van Nhieu J, Yamada T, Zimmermann J, Benes V, Kloor M, Ulrich CM, von Knebel Doeberitz M, Sobhani I, Bork P. Potential of fecal microbiota for early-stage detection of colorectal cancer. *Mol Syst Biol* 2014; **10**: 766 [PMID: 25432777 DOI: 10.15252/msb.20145645]
- 13 **LeBlanc JG**, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 2013; **24**: 160-168 [PMID: 22940212 DOI: 10.1016/j.copbio.2012.08.005]
- 14 **Makki K**, Deehan EC, Walter J, Bäckhed F. The Impact of Dietary Fiber on Gut Microbiota in Host Health and Disease. *Cell Host Microbe* 2018; **23**: 705-715 [PMID: 29902436 DOI: 10.1016/j.chom.2018.05.012]
- 15 **Pushpanathan P**, Mathew GS, Selvarajan S, Seshadri KG, Srikanth P. Gut microbiota and its mysteries. *Indian J Med Microbiol* 2019; **37**: 268-277 [PMID: 31745030 DOI: 10.4103/ijmm.IJMM_19_373]
- 16 **Perry RJ**, Peng L, Barry NA, Cline GW, Zhang D, Cardone RL, Petersen KF, Kibbey RG, Goodman AL, Shulman GI. Acetate mediates a microbiome-brain-β-cell axis to promote metabolic syndrome. *Nature* 2016; **534**: 213-217 [PMID: 27279214 DOI: 10.1038/nature18309]
- 17 **Fiorucci S**, Carino A, Baldoni M, Santucci L, Costanzi E, Graziosi L, Distrutti E, Biagioli M. Bile Acid Signaling in Inflammatory Bowel Diseases. *Dig Dis Sci* 2021; **66**: 674-693 [PMID: 33289902 DOI: 10.1007/s10620-020-06715-3]
- 18 **Gomaa EZ**. Human gut microbiota/microbiome in health and diseases: a review. *Antonie Van Leeuwenhoek* 2020; **113**: 2019-2040 [PMID: 33136284 DOI: 10.1007/s10482-020-01474-7]
- 19 **Nishida A**, Inoue R, Inatomi O, Bamba S, Naito Y, Andoh A. Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin J Gastroenterol* 2018; **11**: 1-10 [PMID: 29285689 DOI: 10.1007/s12328-017-0813-5]
- 20 **Temraz S**, Nassar F, Nasr R, Charafeddine M, Mukherji D, Shamseddine A. Gut Microbiome: A Promising Biomarker for Immunotherapy in Colorectal Cancer. *Int J Mol Sci* 2019; **20** [PMID: 31450712 DOI: 10.3390/ijms20174155]
- 21 **Dabke K**, Hendrick G, Devkota S. The gut microbiome and metabolic syndrome. *J Clin Invest* 2019; **129**: 4050-4057 [PMID: 31573550 DOI: 10.1172/JCI129194]
- 22 **Kadooka Y**, Sato M, Imaizumi K, Ogawa A, Ikuyama K, Akai Y, Okano M, Kagoshima M, Tsuchida T. Regulation of

- abdominal adiposity by probiotics (*Lactobacillus gasseri* SBT2055) in adults with obese tendencies in a randomized controlled trial. *Eur J Clin Nutr* 2010; **64**: 636-643 [PMID: 20216555 DOI: 10.1038/ejcn.2010.19]
- 23 **Younis N**, Zarif R, Mahfouz R. Inflammatory bowel disease: between genetics and microbiota. *Mol Biol Rep* 2020; **47**: 3053-3063 [PMID: 32086718 DOI: 10.1007/s11033-020-05318-5]
- 24 **Zhu W**, Winter MG, Byndloss MX, Spiga L, Duerkop BA, Hughes ER, Büttner L, de Lima Romão E, Behrendt CL, Lopez CA, Sifuentes-Dominguez L, Huff-Hardy K, Wilson RP, Gillis CC, Tükel Ç, Koh AY, Burstein E, Hooper LV, Bäumlner AJ, Winter SE. Precision editing of the gut microbiota ameliorates colitis. *Nature* 2018; **553**: 208-211 [PMID: 29323293 DOI: 10.1038/nature25172]
- 25 **Lane ER**, Zisman TL, Suskind DL. The microbiota in inflammatory bowel disease: current and therapeutic insights. *J Inflamm Res* 2017; **10**: 63-73 [PMID: 28652796 DOI: 10.2147/JIR.S116088]
- 26 **Sokol H**, Seksik P, Furet JP, Firmesse O, Nion-Larmurier I, Beaugerie L, Cosnes J, Corthier G, Marteau P, Doré J. Low counts of *Faecalibacterium prausnitzii* in colitis microbiota. *Inflamm Bowel Dis* 2009; **15**: 1183-1189 [PMID: 19235886 DOI: 10.1002/ibd.20903]
- 27 **Torres J**, Mehandru S, Colombel JF, Peyrin-Biroulet L. Crohn's disease. *Lancet* 2017; **389**: 1741-1755 [PMID: 27914655 DOI: 10.1016/S0140-6736(16)31711-1]
- 28 **Li W**, Deng Y, Chu Q, Zhang P. Gut microbiome and cancer immunotherapy. *Cancer Lett* 2019; **447**: 41-47 [PMID: 30684593 DOI: 10.1016/j.canlet.2019.01.015]
- 29 **Pabst O**. New concepts in the generation and functions of IgA. *Nat Rev Immunol* 2012; **12**: 821-832 [PMID: 23103985 DOI: 10.1038/nri3322]
- 30 **Dahiya DS**, Kichloo A, Singh J, Albosta M, Lekkala M. Current immunotherapy in gastrointestinal malignancies A Review. *J Investig Med* 2021; **69**: 689-696 [PMID: 33443046 DOI: 10.1136/jim-2020-001654]
- 31 **Jacob JB**, Jacob MK, Parajuli P. Review of immune checkpoint inhibitors in immuno-oncology. *Adv Pharmacol* 2021; **91**: 111-139 [PMID: 34099106 DOI: 10.1016/bs.apha.2021.01.002]
- 32 **Lichtenstern CR**, Ngu RK, Shalapur S, Karin M. Immunotherapy, Inflammation and Colorectal Cancer. *Cells* 2020; **9** [PMID: 32143413 DOI: 10.3390/cells9030618]
- 33 **Gologan A**, Sepulveda AR. Microsatellite instability and DNA mismatch repair deficiency testing in hereditary and sporadic gastrointestinal cancers. *Clin Lab Med* 2005; **25**: 179-196 [PMID: 15749237 DOI: 10.1016/j.cll.2004.12.001]
- 34 **Roy S**, Trinchieri G. Microbiota: a key orchestrator of cancer therapy. *Nat Rev Cancer* 2017; **17**: 271-285 [PMID: 28303904 DOI: 10.1038/nrc.2017.13]
- 35 **Routy B**, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, Fluckiger A, Messaoudene M, Rauber C, Roberti MP, Fidelle M, Flament C, Poirier-Colame V, Opolon P, Klein C, Iribarren K, Mondragón L, Jacquilot N, Qu B, Ferrere G, Clémenson C, Mezquita L, Masip JR, Naltet C, Brosseau S, Kaderbhai C, Richard C, Rizvi H, Levenez F, Galleron N, Quinquis B, Pons N, Ryffel B, Minard-Colin V, Gonin P, Soria JC, Deutsch E, Loriot Y, Ghiringhelli F, Zalcman G, Goldwasser F, Escudier B, Hellmann MD, Eggermont A, Raoult D, Albiges L, Kroemer G, Zitvogel L. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* 2018; **359**: 91-97 [PMID: 29097494 DOI: 10.1126/science.aan3706]
- 36 **Ashktorab H**, Kupfer SS, Brim H, Carethers JM. Racial Disparity in Gastrointestinal Cancer Risk. *Gastroenterology* 2017; **153**: 910-923 [PMID: 28807841 DOI: 10.1053/j.gastro.2017.08.018]
- 37 **Yachida S**, Mizutani S, Shiroma H, Shiba S, Nakajima T, Sakamoto T, Watanabe H, Masuda K, Nishimoto Y, Kubo M, Hosoda F, Rokutan H, Matsumoto M, Takamaru H, Yamada M, Matsuda T, Iwasaki M, Yamaji T, Yachida T, Soga T, Kurokawa K, Toyoda A, Ogura Y, Hayashi T, Hatakeyama M, Nakagama H, Saito Y, Fukuda S, Shibata T, Yamada T. Metagenomic and metabolomic analyses reveal distinct stage-specific phenotypes of the gut microbiota in colorectal cancer. *Nat Med* 2019; **25**: 968-976 [PMID: 31171880 DOI: 10.1038/s41591-019-0458-7]
- 38 **Mima K**, Cao Y, Chan AT, Qian ZR, Nowak JA, Masugi Y, Shi Y, Song M, da Silva A, Gu M, Li W, Hamada T, Kosumi K, Hanyuda A, Liu L, Kostic AD, Giannakis M, Bullman S, Brennan CA, Milner DA, Baba H, Garraway LA, Meyerhardt JA, Garrett WS, Huttenhower C, Meyerson M, Giovannucci EL, Fuchs CS, Nishihara R, Ogino S. *Fusobacterium nucleatum* in Colorectal Carcinoma Tissue According to Tumor Location. *Clin Transl Gastroenterol* 2016; **7**: e200 [PMID: 27811909 DOI: 10.1038/ctg.2016.53]
- 39 **Yu J**, Feng Q, Wong SH, Zhang D, Liang QY, Qin Y, Tang L, Zhao H, Stenvang J, Li Y, Wang X, Xu X, Chen N, Wu WK, Al-Aama J, Nielsen HJ, Kiilerich P, Jensen BA, Yau TO, Lan Z, Jia H, Li J, Xiao L, Lam TY, Ng SC, Cheng AS, Wong VW, Chan FK, Yang H, Madsen L, Datz C, Tilg H, Wang J, Brünner N, Kristiansen K, Arumugam M, Sung JJ. Metagenomic analysis of faecal microbiome as a tool towards targeted non-invasive biomarkers for colorectal cancer. *Gut* 2017; **66**: 70-78 [PMID: 26408641 DOI: 10.1136/gutjnl-2015-309800]
- 40 **Song M**, Chan AT. Environmental Factors, Gut Microbiota, and Colorectal Cancer Prevention. *Clin Gastroenterol Hepatol* 2019; **17**: 275-289 [PMID: 30031175 DOI: 10.1016/j.cgh.2018.07.012]
- 41 **Clay SL**, Fonseca-Pereira D, Garrett WS. Colorectal cancer: the facts in the case of the microbiota. *J Clin Invest* 2022; **132** [PMID: 35166235 DOI: 10.1172/JCI155101]
- 42 **Rothschild D**, Weissbrod O, Barkan E, Kurilshikov A, Korem T, Zeevi D, Costea PI, Godneva A, Kalka IN, Bar N, Shilo S, Lador D, Vila AV, Zmora N, Pevsner-Fischer M, Israeli D, Kosower N, Malka G, Wolf BC, Avnit-Sagi T, Lotan-Pompan M, Weinberger A, Halpern Z, Carmi S, Fu J, Wijmenga C, Zernakova A, Elinav E, Segal E. Environment dominates over host genetics in shaping human gut microbiota. *Nature* 2018; **555**: 210-215 [PMID: 29489753 DOI: 10.1038/nature25973]
- 43 **Thaiss CA**, Zmora N, Levy M, Elinav E. The microbiome and innate immunity. *Nature* 2016; **535**: 65-74 [PMID: 27383981 DOI: 10.1038/nature18847]
- 44 **Hills RD Jr**, Pontefract BA, Mishcon HR, Black CA, Sutton SC, Theberge CR. Gut Microbiome: Profound Implications for Diet and Disease. *Nutrients* 2019; **11** [PMID: 31315227 DOI: 10.3390/nu11071613]
- 45 **Fan Y**, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol* 2021; **19**: 55-71 [PMID: 32887946 DOI: 10.1038/s41579-020-0433-9]

- 46 **Chattopadhyay I**, Dhar R, Pethusamy K, Seethy A, Srivastava T, Sah R, Sharma J, Karmakar S. Exploring the Role of Gut Microbiome in Colon Cancer. *Appl Biochem Biotechnol* 2021; **193**: 1780-1799 [PMID: [33492552](#) DOI: [10.1007/s12010-021-03498-9](#)]
- 47 **Cheng Y**, Ling Z, Li L. The Intestinal Microbiota and Colorectal Cancer. *Front Immunol* 2020; **11**: 615056 [PMID: [33329610](#) DOI: [10.3389/fimmu.2020.615056](#)]
- 48 **Loftus M**, Hassouneh SA, Yooseph S. Bacterial community structure alterations within the colorectal cancer gut microbiome. *BMC Microbiol* 2021; **21**: 98 [PMID: [33789570](#) DOI: [10.1186/s12866-021-02153-x](#)]
- 49 **Yang Y**, Cai Q, Shu XO, Steinwandel MD, Blot WJ, Zheng W, Long J. Prospective study of oral microbiome and colorectal cancer risk in low-income and African American populations. *Int J Cancer* 2019; **144**: 2381-2389 [PMID: [30365870](#) DOI: [10.1002/ijc.31941](#)]
- 50 **Yang Y**, Weng W, Peng J, Hong L, Yang L, Toiyama Y, Gao R, Liu M, Yin M, Pan C, Li H, Guo B, Zhu Q, Wei Q, Moyer MP, Wang P, Cai S, Goel A, Qin H, Ma Y. Fusobacterium nucleatum Increases Proliferation of Colorectal Cancer Cells and Tumor Development in Mice by Activating Toll-Like Receptor 4 Signaling to Nuclear Factor- κ B, and Up-regulating Expression of MicroRNA-21. *Gastroenterology* 2017; **152**: 851-866.e24 [PMID: [27876571](#) DOI: [10.1053/j.gastro.2016.11.018](#)]
- 51 **Frosali S**, Pagliari D, Gambassi G, Landolfi R, Pandolfi F, Cianci R. How the Intricate Interaction among Toll-Like Receptors, Microbiota, and Intestinal Immunity Can Influence Gastrointestinal Pathology. *J Immunol Res* 2015; **2015**: 489821 [PMID: [26090491](#) DOI: [10.1155/2015/489821](#)]
- 52 **Levy M**, Kolodziejczyk AA, Thaiss CA, Elinav E. Dysbiosis and the immune system. *Nat Rev Immunol* 2017; **17**: 219-232 [PMID: [28260787](#) DOI: [10.1038/nri.2017.7](#)]
- 53 **Ruan H**, Leibowitz BJ, Zhang L, Yu J. Immunogenic cell death in colon cancer prevention and therapy. *Mol Carcinog* 2020; **59**: 783-793 [PMID: [32215970](#) DOI: [10.1002/mc.23183](#)]
- 54 **Proença MA**, Biselli JM, Succì M, Severino FE, Berardinelli GN, Caetano A, Reis RM, Hughes DJ, Silva AE. Relationship between *Fusobacterium nucleatum*, inflammatory mediators and microRNAs in colorectal carcinogenesis. *World J Gastroenterol* 2018; **24**: 5351-5365 [PMID: [30598580](#) DOI: [10.3748/wjg.v24.i47.5351](#)]
- 55 **Gur C**, Ibrahim Y, Isaacson B, Yamin R, Abed J, Gamliel M, Enk J, Bar-On Y, Stanietsky-Kaynan N, Copenhagen-Glazer S, Shussman N, Almogy G, Cuapio A, Hofer E, Mevorach D, Tabib A, Ortenberg R, Markel G, Miklič K, Jonjic S, Brennan CA, Garrett WS, Bachrach G, Mandelboim O. Binding of the Fap2 protein of *Fusobacterium nucleatum* to human inhibitory receptor TIGIT protects tumors from immune cell attack. *Immunity* 2015; **42**: 344-355 [PMID: [25680274](#) DOI: [10.1016/j.immuni.2015.01.010](#)]
- 56 **Saito T**, Nishikawa H, Wada H, Nagano Y, Sugiyama D, Atarashi K, Maeda Y, Hamaguchi M, Ohkura N, Sato E, Nagase H, Nishimura J, Yamamoto H, Takiguchi S, Tanoue T, Suda W, Morita H, Hattori M, Honda K, Mori M, Doki Y, Sakaguchi S. Two FOXP3(+)/CD4(+) T cell subpopulations distinctly control the prognosis of colorectal cancers. *Nat Med* 2016; **22**: 679-684 [PMID: [27111280](#) DOI: [10.1038/nm.4086](#)]
- 57 **Sideris M**, Papagrigoriadis S. Molecular biomarkers and classification models in the evaluation of the prognosis of colorectal cancer. *Anticancer Res* 2014; **34**: 2061-2068 [PMID: [24778007](#)]
- 58 **Fishel R**. Mismatch repair. *J Biol Chem* 2015; **290**: 26395-26403 [PMID: [26354434](#) DOI: [10.1074/jbc.R115.660142](#)]
- 59 **Li WQ**, Kawakami K, Ruszkiewicz A, Bennett G, Moore J, Iacopetta B. BRAF mutations are associated with distinctive clinical, pathological and molecular features of colorectal cancer independently of microsatellite instability status. *Mol Cancer* 2006; **5**: 2 [PMID: [16403224](#) DOI: [10.1186/1476-4598-5-2](#)]
- 60 **Thiel A**, Ristimäki A. Toward a Molecular Classification of Colorectal Cancer: The Role of BRAF. *Front Oncol* 2013; **3**: 281 [PMID: [24298448](#) DOI: [10.3389/fonc.2013.00281](#)]
- 61 **Koustas E**, Karamouzis MV, Mihailidou C, Schizas D, Papavassiliou AG. Co-targeting of EGFR and autophagy signaling is an emerging treatment strategy in metastatic colorectal cancer. *Cancer Lett* 2017; **396**: 94-102 [PMID: [28323034](#) DOI: [10.1016/j.canlet.2017.03.023](#)]
- 62 **Koustas E**, Papavassiliou AG, Karamouzis MV. The role of autophagy in the treatment of BRAF mutant colorectal carcinomas differs based on microsatellite instability status. *PLoS One* 2018; **13**: e0207227 [PMID: [30427914](#) DOI: [10.1371/journal.pone.0207227](#)]
- 63 **André T**, Shiu KK, Kim TW, Jensen BV, Jensen LH, Punt C, Smith D, Garcia-Carbonero R, Benavides M, Gibbs P, de la Fouchardiere C, Rivera F, Elez E, Bendell J, Le DT, Yoshino T, Van Cutsem E, Yang P, Farooqui MZH, Marinello P, Diaz LA Jr; KEYNOTE-177 Investigators. Pembrolizumab in Microsatellite-Instability-High Advanced Colorectal Cancer. *N Engl J Med* 2020; **383**: 2207-2218 [PMID: [33264544](#) DOI: [10.1056/NEJMoa2017699](#)]
- 64 **Ganesh K**, Stadler ZK, Cercek A, Mendelsohn RB, Shia J, Segal NH, Diaz LA Jr. Immunotherapy in colorectal cancer: rationale, challenges and potential. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 361-375 [PMID: [30886395](#) DOI: [10.1038/s41575-019-0126-x](#)]
- 65 **Raman SS**, Hecht JR, Chan E. Talimogene laherparepvec: review of its mechanism of action and clinical efficacy and safety. *Immunotherapy* 2019; **11**: 705-723 [PMID: [31045464](#) DOI: [10.2217/imt-2019-0033](#)]
- 66 **Wang DK**, Zuo Q, He QY, Li B. Targeted Immunotherapies in Gastrointestinal Cancer: From Molecular Mechanisms to Implications. *Front Immunol* 2021; **12**: 705999 [PMID: [34447376](#) DOI: [10.3389/fimmu.2021.705999](#)]
- 67 **Grizzi F**, Basso G, Borroni EM, Cavalleri T, Bianchi P, Stifter S, Chiriva-Internati M, Malesci A, Laghi L. Evolving notions on immune response in colorectal cancer and their implications for biomarker development. *Inflamm Res* 2018; **67**: 375-389 [PMID: [29322204](#) DOI: [10.1007/s00011-017-1128-1](#)]
- 68 **Koustas E**, Sarantis P, Kyriakopoulou G, Papavassiliou AG, Karamouzis MV. The Interplay of Autophagy and Tumor Microenvironment in Colorectal Cancer-Ways of Enhancing Immunotherapy Action. *Cancers (Basel)* 2019; **11** [PMID: [31013961](#) DOI: [10.3390/cancers11040533](#)]
- 69 **Yang X**, Li Y, Zou L, Zhu Z. Role of Exosomes in Crosstalk Between Cancer-Associated Fibroblasts and Cancer Cells. *Front Oncol* 2019; **9**: 356 [PMID: [31131261](#) DOI: [10.3389/fonc.2019.00356](#)]
- 70 **Qian BZ**, Li J, Zhang H, Kitamura T, Zhang J, Campion LR, Kaiser EA, Snyder LA, Pollard JW. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature* 2011; **475**: 222-225 [PMID: [21654748](#) DOI: [10.1038/nature09923](#)]

- 10.1038/nature10138]
- 71 **Ge Y**, Wang X, Guo Y, Yan J, Abuduwaili A, Aximujiang K, Wu M. Correction to: Gut microbiota influence tumor development and Alter interactions with the human immune system. *J Exp Clin Cancer Res* 2021; **40**: 334 [PMID: 34696779 DOI: 10.1186/s13046-021-02131-1]
 - 72 **Nejman D**, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, Rotter-Maskowitz A, Weiser R, Mallel G, Gigi E, Meltser A, Douglas GM, Kamer I, Gopalakrishnan V, Dadosh T, Levin-Zaidman S, Avnet S, Atlan T, Cooper ZA, Arora R, Cogdill AP, Khan MAW, Ologun G, Bussi Y, Weinberger A, Lotan-Pompan M, Golani O, Perry G, Rokah M, Bahar-Shany K, Rozeman EA, Blank CU, Ronai A, Shaoul R, Amit A, Dorfman T, Kremer R, Cohen ZR, Harnof S, Siegal T, Yehuda-Shnaidman E, Gal-Yam EN, Shapira H, Baldini N, Langille MGI, Ben-Nun A, Kaufman B, Nissan A, Golan T, Dadiani M, Levanon K, Bar J, Yust-Katz S, Barshack I, Peeper DS, Raz DJ, Segal E, Wargo JA, Sandbank J, Shental N, Straussman R. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Science* 2020; **368**: 973-980 [PMID: 32467386 DOI: 10.1126/science.aay9189]
 - 73 **Poore GD**, Kopylova E, Zhu Q, Carpenter C, Fraraccio S, Wandro S, Kosciolk T, Janssen S, Metcalf J, Song SJ, Kanbar J, Miller-Montgomery S, Heaton R, Mckay R, Patel SP, Swafford AD, Knight R. Microbiome analyses of blood and tissues suggest cancer diagnostic approach. *Nature* 2020; **579**: 567-574 [PMID: 32214244 DOI: 10.1038/s41586-020-2095-1]
 - 74 **Qiu Q**, Lin Y, Ma Y, Li X, Liang J, Chen Z, Liu K, Huang Y, Luo H, Huang R, Luo L. Exploring the Emerging Role of the Gut Microbiota and Tumor Microenvironment in Cancer Immunotherapy. *Front Immunol* 2020; **11**: 612202 [PMID: 33488618 DOI: 10.3389/fimmu.2020.612202]
 - 75 **Pitt JM**, Vétizou M, Daillère R, Roberti MP, Yamazaki T, Routy B, Lepage P, Boneca IG, Chamaillard M, Kroemer G, Zitvogel L. Resistance Mechanisms to Immune-Checkpoint Blockade in Cancer: Tumor-Intrinsic and -Extrinsic Factors. *Immunity* 2016; **44**: 1255-1269 [PMID: 27332730 DOI: 10.1016/j.immuni.2016.06.001]
 - 76 **Matson V**, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre ML, Luke JJ, Gajewski TF. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. *Science* 2018; **359**: 104-108 [PMID: 29302014 DOI: 10.1126/science.aao3290]
 - 77 **Dzutsev A**, Badger JH, Perez-Chanona E, Roy S, Salcedo R, Smith CK, Trinchieri G. Microbes and Cancer. *Annu Rev Immunol* 2017; **35**: 199-228 [PMID: 28142322 DOI: 10.1146/annurev-immunol-051116-052133]
 - 78 **Zhou CB**, Zhou YL, Fang JY. Gut Microbiota in Cancer Immune Response and Immunotherapy. *Trends Cancer* 2021; **7**: 647-660 [PMID: 33674230 DOI: 10.1016/j.trecan.2021.01.010]
 - 79 **Chaput N**, Lepage P, Coutzac C, Soularue E, Le Roux K, Monot C, Boselli L, Routier E, Cassard L, Collins M, Vaysse T, Marthey L, Eggermont A, Asvatourian V, Lanoy E, Mateus C, Robert C, Carbonnel F. Baseline gut microbiota predicts clinical response and colitis in metastatic melanoma patients treated with ipilimumab. *Ann Oncol* 2019; **30**: 2012 [PMID: 31408090 DOI: 10.1093/annonc/mdz224]
 - 80 **Frankel AE**, Coughlin LA, Kim J, Froehlich TW, Xie Y, Frenkel EP, Koh AY. Metagenomic Shotgun Sequencing and Unbiased Metabolomic Profiling Identify Specific Human Gut Microbiota and Metabolites Associated with Immune Checkpoint Therapy Efficacy in Melanoma Patients. *Neoplasia* 2017; **19**: 848-855 [PMID: 28923537 DOI: 10.1016/j.neo.2017.08.004]
 - 81 **Alexander JL**, Wilson ID, Teare J, Marchesi JR, Nicholson JK, Kinross JM. Gut microbiota modulation of chemotherapy efficacy and toxicity. *Nat Rev Gastroenterol Hepatol* 2017; **14**: 356-365 [PMID: 28270698 DOI: 10.1038/nrgastro.2017.20]
 - 82 **Fong W**, Li Q, Yu J. Gut microbiota modulation: a novel strategy for prevention and treatment of colorectal cancer. *Oncogene* 2020; **39**: 4925-4943 [PMID: 32514151 DOI: 10.1038/s41388-020-1341-1]
 - 83 **Zhang J**, Hong Y, Harman NJ, Das A, Ebner PD. Genome sequence of a salmonella phage used to control salmonella transmission in Swine. *Genome Announc* 2014; **2** [PMID: 25212610 DOI: 10.1128/genomeA.00521-14]
 - 84 **Schwartz DJ**, Rebeck ON, Dantas G. Complex interactions between the microbiome and cancer immune therapy. *Crit Rev Clin Lab Sci* 2019; **56**: 567-585 [PMID: 31526274 DOI: 10.1080/10408363.2019.1660303]
 - 85 **Wang JW**, Kuo CH, Kuo FC, Wang YK, Hsu WH, Yu FJ, Hu HM, Hsu PI, Wang JY, Wu DC. Fecal microbiota transplantation: Review and update. *J Formos Med Assoc* 2019; **118** Suppl 1: S23-S31 [PMID: 30181015 DOI: 10.1016/j.jfma.2018.08.011]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-3991568
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>

