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Clinical and Translational Research

CD93 serves as a potential biomarker of gastric cancer and correlates with the tumor microenvironment

Li Z *et al*. CD93-A biomarker of gastric cancer

Zheng Li, Xiao-Jie Zhang, Chong-Yuan Sun, He Fei, Ze-Feng Li, Dong-Bing Zhao

Abstract

BACKGROUND

The tumor microenvironment (TME) plays an important role in the growth and expansion of gastric cancer (GC). Studies have identified that CD93 is involved in abnormal tumor angiogenesis, which may be related to the regulation of TME.

AIM

To determine the role of CD93 in GC.

METHODS

Transcriptomic data of GC was investigated in a cohort from The Cancer Genome Atlas. Additionally, RNA-seq data sets from Gene Expression Omnibus (GSE118916, GSE52138, GSE79973, GSE19826, and GSE84433) were applied to validate the results. We performed the immune infiltration analyses using ESTIMATE, CIBERSORT, and ssGSEA. Furthermore, weighted gene co-expression network analysis (WGCNA) was conducted to identify the immune-related genes.

RESULTS

Compared to normal tissues, CD93 significantly enriched in tumor tissues (t = 4.669, 95%CI: 0.342-0.863, P < 0.001). Higher expression of CD93 was significantly associated with shorter overall survival (hazard ratio = 1.62, 95%CI: 1.09-2.4, P = 0.017), less proportion of CD8 T and activated natural killer cells in the TME (P < 0.05), and lower tumor mutation burden (t = 4.131, 95%CI: 0.721-0.256, P < 0.001). Genes co-expressed with CD93 were mainly enriched in angiogenesis. Moreover, 11 genes were identified with a strong relationship between CD93 and the immune microenvironment using WGCNA.

CONCLUSION

CD93 is a novel prognostic and diagnostic biomarker for GC, which is closely related to the immune infiltration in TME. Although this retrospective study made a comprehensive analysis, the prospective cohort studies are preferred to further confirm these conclusions.

Key Words: Gastric cancer; CD93; Tumor microenvironment; Immunotherapy; Prognosis; Biomarker

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Core Tip: Gastric cancer (GC) is an aggressive malignancy, with a 5-year survival rate lower than 20%. The disease burden caused by GC remains heavy worldwide. In this study, various analyses were performed using transcriptomic profiles from the Gene Expression Omnibus databases and The Cancer Genome Atlas. Finally, enrichment analysis and protein-protein interaction network were constructed. CD93 is identified as a diagnostic and prognostic biomarker of GC, which is closely related to the immune

infiltration in the tumor microenvironment. Then, Immune-related gene modules were identified to further reveal the relationship between CD93 and immune characteristics.

INTRODUCTION

As a common malignant digestive tract tumor, gastric cancer (GC) is the fourth leading cause of cancer-related mortality worldwide, and the 5-year survival rate of GC is lower than 20%[1]. Treatments for GC include endoscopic resection, surgery (D2 lymphadenectomy), perioperative or adjuvant chemotherapy, targeted therapy, immunotherapy, and so on. Among them, immunotherapy for GC has attracted more attention these years[2,3]. Tumor microenvironment (TME) refers to tumor cells and surrounding cellular matrix, including blood vessels, and immune cells, and is an important factor influencing the effect of immunotherapy for GC[4]. Targeting and suppressing the immunosuppressive properties of the TME can enhance the overall response rate (ORR) levels of immunotherapy, including immune checkpoint inhibitors (ICIs)[5-7]. CD93 is known as a C1q receptor shown to be involved in a variety of biological processes such as inflammatory response, tumor angiogenesis, matrix regulation, innate lymphoid cell function, and so on[8-10], which suggests that CD93 may participate in the regulation of TME. A recent study has found that blocking the CD93 pathway contributes to drug transport and immunotherapy by normalizing the vasculature of tumors. Blockade of the CD93 pathway can improve the efficacy of chemotherapy and immunotherapy[11]. Although the value of CD93 in some tumors has been explored, the specific role of CD93 in GC is still unclear.

Given these considerations, we were deeply interested in the relationship between CD93 and immune infiltration in the TME and the value of CD93 in the diagnosis, prognosis, and immunotherapy of GC. Therefore, transcriptome profiling RNA-seq of stomach adenocarcinoma (STAD) was investigated from The Cancer Genome Atlas (TCGA) and Gene Expression Omnibus (GEO). We performed a pan-cancer analysis at first to identify the general significance of CD93 in cancers. Then, we divided patients with GC in this study into two groups by the expression of CD93 and compared their differences in immune cell infiltration, gene mutation landscape, tumor mutational burden, and so on.

MATERIALS AND METHODS

Data sources and processing

Gene expression RNA-seq (HTSeq-Counts, HTSeq-FPKM) and survival data in a cohort of TCGA Stomach Cancer were downloaded from UCSC Xena[12]. HTSeq-FPKM of 375 tumor samples and 32 normal samples were used for further analysis. HTSeq-Counts data were used to identify the differential expressed genes (DEGs) of low and high CD93 expression groups using the R package DESeq2[13] ($|log2FoldChange| > 1$ and adjusted $P < 0.05$). Expression profiling data in 5 datasets (GSE118916, GSE52138, GSE79973, GSE19826, and GSE84433) from GEO[14] were downloaded as validation sets.

Mutation data (MuTect2) including 414 patients with STAD from TCGA were processed using R package maftools[15]. The waterfall plot was used to show the genetic mutation using the R package ComplexHeatmap[16].

Pan-cancer analysis

To understand the general significance of CD93 in cancers, we compared the expression levels of CD93 between tumor tissues and normal tissues in various cancer types using TIMER2.0[17]. Additionally, TISIDB[18] was also used to obtain the relationship between CD93 expression and the OS of these cancer types.

Diagnostic and prognostic value analysis

We compared CD93 expression levels of GC and normal tissues in both unpaired and paired samples and visualize outcomes using R package ggplot2. Receiver operating characteristic (ROC) curves and Kaplan-Meier survival analysis were conducted to further explore the diagnostic and prognostic value of CD93, and R packages pROC[19] and survminer were used for visualization, respectively. Univariate and multivariate COX proportional hazards models were established for better understanding. In addition, Immunohistochemistry and Immunofluorescence of CD93 were obtained from Human Protein Atlas[20].

Functional enrichment analysis

Genes that were significantly positively or negatively related to CD93 were identified using LinkedOmics[21]. Heatmaps were used to show the top 50 positively and the top 50 negatively correlated genes. Then, we constructed a protein-protein interaction (PPI) network of positively correlated genes *via* GeneMANIA[22]. Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses of these genes were performed using the R package clusterProfiler[23].

Immune infiltration analysis

A plot from the TIMER[24] was used to show the correlations between the CD93 expression level and B cell, CD8+T Cell, CD4+T cell, macrophage, neutrophil, and dendritic cell. ESTIMATE is a method to identify the proportions of stromal and immune cells, which can bring the in-depth exploration of TME. We evaluated the immune score (immune component), stromal score (stromal component), and ESTIMATE score (comprehensive score of immunity and matrix) of each sample from TCGA and GEO using the R package estimate[25]. CIBERSORT[26] is a tool to characterize the cell composition of various tissues. We calculate the proportion of 22 immune cells in each sample with STAD using this method. Then, we conducted the ssGSEA to evaluate the infiltration level of 28 immune cell types based on the published immune gene sets[27] using the R package GSVA[28].

Weighted gene co-expression network analysis

Weighted gene co-expression network analysis (WGCNA) was applied to identify the module genes related to CD93 and the immunity of patients with STAD using the R package WGCNA[29] (softPower = 4). Nine modules were obtained to calculate their relationships with stromal score, immune score, ESTIMATE score, and tumor purity. Finally, we identified 11 hub genes based on the value of module membership (MM) > 0.80 and gene significance (GS) > 0.85.

Analysis of hub genes

The PPI network and GO enrichment analysis of 11 hub genes were performed using the R package clusterProfiler and STRING[30], respectively. Then, we calculated Spearman's correlation of 11 hub genes, hub gene-ESTIMATE, and hub gene-ssGSEA using the R package corrplot.

Statistical analysis

All statistical analyses mentioned in this article were conducted by R (version 4.2.0) and SPSS (version 25.0). Weltch't test and Spearman's coefficient were used for box plots and correlation analysis, respectively. We evaluated statistical significance using two-sided t-tests and defined it as [a]$P < 0.05$, [b]$P < 0.01$, and [c]$P < 0.001$.

RESULTS

Pan-cancer analysis of CD93

The expression of CD93 between tumor and normal tissues in various cancers was compared using TIMER2.0, suggesting that CD93 expression was significantly different between tumor and normal tissues in various types of cancers ($P < 0.05$) (Figure 1A). We further investigated the effect of CD93 on OS across human cancers *via* the TISIDB. It showed that high CD93 expression led to shorter overall survival in STAD, kidney renal papillary cell carcinoma, brain lower-grade glioma, lung squamous cell carcinoma, and uveal melanoma, while leading to a longer one in kidney renal clear cell carcinoma (Figure 1B).

Diagnostic value for GC of CD93

The CD93 expression levels of unpaired and paired samples from TCGA were both significantly higher in GC than in normal tissues ($t = 4.669$, 95%CI: 0.342-0.863, $P < 0.001$; $t = 3.238$, 95%CI: 0.196-0.877, $P = 0.003$) (Figure 2A and B). In addition, expression data obtained from GEO (GSE118916, GSE52138, GSE79973, and GSE19826) was applied for verification (Figure 2H-K). All datasets from TCGA and GEO showed a significantly

higher expression of CD93 in GC tissues than in normal tissues ($P < 0.05$). Immunohistochemistry indicated higher CD93 expression in GC tissues than in normal tissues from the protein level (Figure 2E). Immunofluorescence indicated that CD93 mainly expressed in vesicles, plasma membrane, and toggle channels (Figure 2F). Furthermore, ROC curves were performed using the datasets mentioned above to evaluate the diagnostic value of CD93, the area under the curve was 0.695, 0.876, 0.806, 0.750, and 0.771, respectively (Figure 2C and L-O).

Prognostic value for GC of CD93

Samples from the TCGA STAD dataset were divided into two groups by the CD93 expression level, including the low CD93 expression group (low, n = 114) and the high CD93 expression group (high, n = 220). Kaplan-Meier analysis of two groups was conducted, suggesting that patients with high expression of CD93 had significantly shorter OS [hazard ratio (HR) = 1.60, 95%CI: 1.09-2.35, P = 0.017] (Figure 2D). In addition, we set GSE84433 with 357 GC patients as an external independent validation dataset and divided these patients into low-CD93 (low, n = 178) and high-CD93 (high, n = 179) expression groups. A similar result could be drawn from the Kaplan–Meier analysis that patients in the high-CD93 expression group had a shorter OS (HR: 1.74, 95%CI: 1.29-2.35, P < 0.001) (Figure 2G). Univariate and multivariate COX regression analysis were conducted as Table 1, indicating CD93 was a significant independent prognostic risk factor for GC (HR = 1.62, 95%CI: 1.09-2.40, P = 0.017). The baseline patient characteristics were summarized in Table 2

Identification and enrichment analysis of correlation genes

Correlation analysis with CD93 based on the Pearson test was conducted using LinkedOmics. The result was visualized by a volcano plot (Figure 3A). We obtained 7026 significantly positively correlated genes (red dots) and 5308 significantly negatively ones (green dots), respectively. In addition, we showed heatmaps of the top 50 positively correlated and the top 50 negatively correlated genes (Figure 3B and C). PPI network of

the top 10 positive CD93 co-expressed genes was further constructed using GeneMANIA, suggesting functions of "angiogenesis", "endothelium development", and "regulation of angiogenesis" (Figure 3D). GO enrichment analysis of positive CD93 co-expressed genes mainly enriched in "ameboidal-type cell migration" (biological process), "collagen-containing extracellular matrix" (cell component), and "growth factor binding" (molecular function) (Figure 3E). KEGG pathway enrichment analysis indicated that these genes mainly participated in the "PI3K-Akt signaling pathway", "Focal adhesion", and "Pathways in cancer" (Figure 3F).

Immune-related analysis in the TME

The expression of CD93 was positively proportional to CD8+T Cell, CD4+T cell, macrophage, neutrophil, and dendritic cell ($P < 0.05$) (Figure 4A). Furthermore, CD93 expression tended to have a positive correlation with ESTIMATE results (immune score, stromal score, and ESTIMATE score) ($P < 0.05$) (Figure 4B-D). Then, we performed the CIBERSORT for determining the proportion of 22 immune cells in each sample with STAD (Figure 4E). The proportion of 22 immune cells in two groups was compared using CIBERSORT, suggesting the proportion of CD8 T cells, follicular helper T cells, and activated NK cells in the high CD93 expression group was significantly lower than that in the low CD93 expression group, whereas Monocytes, Dendritic cells resting, and Mast cells resting had just the reverse ($P < 0.05$) (Figure 4F). ssGSEA showed that 24 types of immune cell (such as activated B cell, activated CD8 T cell, activated dendritic cell, central memory CD4 T cell, and central memory CD8 T cell) had a significantly higher expression in the high CD93 expression group, while CD56 bright natural killer (NK) cell had a lower expression in this group ($P < 0.05$) (Figure 4G).

Gene mutation analysis and tumor mutation burden comparison

Gene mutation of GC is closely related to its therapeutic efficacy. Accordingly, a waterfall plot was used to identify the top 15 significant gene mutations (such as *TTN*, *MUC16*, and *LRP1B*) between two groups ($P < 0.05$) (Figure 5A). Furthermore, we made a

comparison of tumor mutation burden (TMB) between low and high CD93 expression groups, which indicated that patients with high CD93 expression had a lower TMB ($t =$ 4.131, 95%CI: 0.721-0.256, $P < 0.001$) (Figure 5B). The mutation rate of CD93 in GC ranked fourth in pan-cancer (Figure 5C).

Identification of hub genes in the immune microenvironment of GC

A total of 1679 DEGs (966 upregulated and 713 downregulated) were obtained between low and high CD93 expression groups. Then, we visualize the results using a volcano plot (Figure 6A). WGCNA was conducted to identify a module related to CD93 expression and immune infiltration (Figure 6B-D). The Yellow module was screened out because of its high correlation with immunity ($r = 0.89$, $P = 5 \times 10^{-122}$), hence we acquired 11 hub genes (*MPEG1*, *IL10RA*, *SRGN*, *SLA*, *DOCK2*, *NCKAP1L*, *IKZF1*, *PTPRC*, *SIGLEC10*, *PLEK*, *P2RY10*) from the yellow module based on MM > 0.80 and GS > 0.85 (Figure 6E).

Analysis of 11 hub genes

GO enrichment analysis identified these genes were mainly enriched in "positive regulation of phagocytosis" (biological process), "cytoplasmic side of plasma membrane" (cell component), and "interleukin-10 binding" (molecular function) (Figure 7A). Furthermore, we constructed a PPI network and made a correlation analysis of these genes (Figure 7B and C). Correlation analysis between these genes and TME (ESTIMATE and ssGSEA) suggested that these genes were closely related to both stromal components and immune infiltration in GC (Figure 7D and E).

Validation of immune-related characteristics in TME

GSE84433 (357 samples) was set as an external independent validation dataset. We performed ESTIMATE, CIBERSORT, and ssGSEA to evaluate the immune-related characteristics of CD93 in TME again. Then, several similar results as before could be obtained. CD93 expression was positively correlated with the ESTIMATE results (Figure

8A-C). CIBERSORT showed the proportion of various immune cell types in each sample (Figure 8D). The proportion of follicular helper T cells and activated NK cells in the high-CD93 expression group was lower compared to that in the low-CD93 expression group (Figure 8E). ssGSEA indicated that 24 immune cell types (including activated B cell, activated CD8 T cell, and activated dendritic cell) expressed significantly higher in the high-CD93 expression group (Figure 8F). Accordingly, CD93 was identified to be closely related to immune infiltration in TME of GC.

DISCUSSION

A In this study, we applied bioinformatics technology to determine the specific role of CD93 in GC. Various analytical methods identified that CD93 is a biomarker for the diagnosis and prognosis of GC. Concerning the potential mechanisms of CD93, enrichment analysis was performed. Consistent with previous studies[11], CD93 was found to be involved in the formation of tumor blood vessels in GC. Such disordered, immature, and impermeable blood vessels can lead to poor tumor blood perfusion. The resulting hypoxic microenvironment can promote the production of more aggressive tumor cells and limit the killing effect of immune cells[31]. In addition to regulating angiogenesis, GO enrichment analysis suggested that CD93 is involved in matrix formation including cell junction, focal adhesion, and regulation of cytokine production, which further demonstrates the important status of CD93 in TME. Also worth noting is that CD93 plays a critical role in the PI3K-Akt signaling pathway. The PI3K-Akt pathway is constantly found to be activated in various cancers and has been considered a promising target for therapy. Multiple activators of this pathway have been proved to possess oncogenic potentials *in vivo* and *in vitro* with diverse mechanisms, including stimulation of metabolic reprogramming, proliferation, and so on[32]. These tend to be part of the reasons for the poor prognosis of patients with GC induced by CD93.

As important components of the TME, immune cells can inhibit or promote tumor progression by interacting with tumor cells[33]. The investigation of immune components in TME brings a deeper understanding of the biological characteristics, prognosis, and

other information of tumors. Based on ESTIMATE, CIBERSORT, and ssGSEA, a comparison of immune cell infiltration between low and high CD93 expression groups was conducted. According to the result of ESTIMATE, we found that CD93 was significantly proportional to immunity. Previous studies could give reasonable explanations for this result. The blood vessel wall mainly consists of endothelial cells, pericytes, and smooth muscle cells. On the one hand, these cells can activate T cells by expressing MHCI, MHCII, and some costimulatory factors such as CD80, and CD86 to participate in the immune response[34]. On the other hand, a variety of immune cell subsets, including NK cells, T helper 17 cells, regulatory T lymphocytes, and functional subsets of macrophages can act as regulators of arteriogenesis[35]. The crosstalk between the vascular system and immunity explains the high correlation between CD93 and immunity.

From the proportion of immune cell expression, CD8 T cells, Follicular helper T cells, and activated NK cells showed a lower proportion in the high CD93 expression group, while monocytes, resting dendritic cells, and resting mast cells had just the reverse. This is probably caused by local microenvironment hypoxia and accumulation of metabolic end-products induced by abnormal vascular proliferation due to high expression of CD93. CD8 T cells and NK cells are important effector cells involved in anti-tumor immune response in TME and are related to tumor progression and prognosis[36,37]. At present, CD8 T cells have been described as a variety of subtypes, including Tc1, Tc2, Tc9, Tc17, and Tc22, each with different cytotoxicity and effects. Among these cell subtypes, Tc17 and Tc22 are the main T cell subtypes in gastric tissue. Tc17 has no cytotoxicity, and its high expression is negatively correlated with the survival time of GC, while Tc22 is just the opposite. Besides, follicular helper T cells are the key to the production of germinal center formation[38]. They interact with tumor-specific B cells to enhance the anti-tumor effect of CD8 T cells. In summary, the reduced proportion of these important immune cells in the TME may be the main reason for the poor prognosis caused by CD93. However, analysis of the expression of immune cells in two groups suggested that various types of immune cells were highly expressed in the high CD93 expression group.

Although blood vessels are conducive to tissue growth and immune response, they can contribute to inflammation and malignant diseases. Abnormal angiogenesis induced by CD93 in TME can promote tumor growth and form an immune-hostile microenvironment[39], and this effect exceeds its immune enhancement effect, which makes the prognosis of the high CD93 expression group with high immune infiltration still poor.

In recent years, the ICIs represented by PD-1, PD-L1, and CTLA-4 bring considerable disease relief to tumor patients, playing an important role in tumor immunotherapy. However, not all patients can benefit from ICIs. A series of studies have shown that TMB is a potential biomarker for predicting the response to ICIs and patients with high TMB possess a better immunotherapeutic effect of ICIs[40,41]. In this study, we made a comparison of gene mutational landscape and TMB between two groups. The results showed that GC patients with high expression of CD93 had a lower TMB, indicating that the effect of immunotherapy in GC patients with high expression of CD93 is poor. Then, we performed WGCNA to identify the key genes related to CD93 in the tumor immune microenvironment of GC. We obtained 11 genes from the yellow module. Among them, SRGN overexpression has been previously shown to promote colorectal cancer metastasis and predict a poor prognosis of hepatocellular carcinoma[42,43]. This time, the identification of these 11 genes can help us further understand the immune microenvironment of GC and suggest potential methods for immunotherapy of GC in the future.

Although we have taken a variety of methods to obtain a comprehensive understanding of the relationship between CD93 and GC, we use 5 cohorts (GSE118916, GSE52138, GSE79973, GSE19826, and GSE84433) as external validation sets, some limitations of this study should be recognized. First, this is a retrospective study. Selection bias, loss of follow-up bias, recall bias, and other biases exist in the study. Thus, a prospective study is required to avoid these biases. Furthermore, limited by TCGA and GEO, we only performed research and analysis from the genetic level. A study that can

demonstrate CD93 expression from the protein level or reveal the direct mechanism needs to be conducted in the future.

CONCLUSION

All in all, comprehensive analyses were applied using transcriptomic profiles and survival information from the GEO and TCGA databases, suggesting that CD93 is a biomarker of diagnosis and prognosis for GC, which closely correlates with immune infiltration in TME. These data help us further comprehend the role of CD93 in the immune microenvironment and may suggest potential strategies for immunotherapy of GC in the future.

ARTICLE HIGHLIGHTS

Research background

Gastric cancer (GC) is a common malignancy with poor 5-year survival rate. Tumor microenvironment (TME) containing intricate interaction between immune and non-immune cells produces significant impact of the survival of GC. Additionally, CD93 was proved to be associated with abnormal angiogenesis, which could be involved in TME of GC.

Research motivation

This study was conducted to determine the specific role of CD93 in GC in order to provide insights for the discovery of novel therapeutic target of GC in the feature.

Research objectives

Cohorts data of GC patients was investigated from The Cancer Genome Atlas and Gene Expression Omnibus (GSE118916, GSE52138, GSE79973, GSE19826, and GSE84433).

Research methods

We performed a series of immune infiltration analyses using ESTIMATE, CIBERSORT, and ssGSEA. Furthermore, weighted gene co-expression network analysis was conducted to identify the immune-related genes.

Research results

CD93 significantly enriched in tumor tissues. Additionally, higher expression of CD93 was significantly associated with shorter overall survival, less proportion of CD8 T and activated nature killer cells in the TME, and lower tumor mutational burden.

Research conclusions

CD93 is a novel prognostic and diagnostic biomarker for GC, which is closely related to the immune infiltration in TME.

Research perspectives

CD93 can serve as a potential therapeutic target for the immunotherapy of GC in the feature.

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