

• RAPID COMMUNICATION •

Effects of *Helicobacter pylori* infection on gastric epithelial cell kinetics in patients with chronic renal failure

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CONCLUSION: In gastric epithelial cells, expression of both the pre-apoptotic protein Bax and the proliferation marker PCNA increase with *H pylori* infection. This increase is more evident in patients with uremia. These findings suggest that uremia accelerates apoptosis and proliferation in gastric epithelial cells.

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Key words: *Helicobacter pylori*; Chronic renal failure; Bax; Proliferating cell nuclear antigen

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Abstract

AIM: To evaluate the effects of *Helicobacter pylori* infection on gastric epithelial cell kinetics in patients with chronic renal failure (CRF).

METHODS: Forty-four patients were enrolled in this study and divided into four groups with respect to their *Helicobacter pylori* (*H pylori*) and CRF status. Groups were labeled as follows: 1a: normal renal function, *H pylori* negative ($n = 12$), 1b: normal renal function, *H pylori* positive ($n = 11$), 2a: CRF, *H pylori* negative ($n = 10$), 2b: CRF, *H pylori* positive ($n = 11$). Upper gastrointestinal endoscopy was done in all the patients involved in the study. During endoscopic investigation, antral biopsy specimens were taken from each patient. In order to evaluate the cell apoptosis and proliferation in gastric epithelial cells, Bax and proliferating cell nuclear antigen (PCNA) labeling indexes (LI) were assessed with immunohistochemical staining method.

RESULTS: For groups 1a, 1b, 2a, and 2b, mean Bax LI was identified as 34.4 ± 13.7 , 44.1 ± 16.5 , 46.3 ± 20.5 , 60.7 ± 13.8 , respectively and mean PCNA LI was identified as 36.2 ± 17.2 , 53.6 ± 25.6 , 59.5 ± 25.6 , 67.2 ± 22 , respectively. When the one-way ANOVA test was applied, statistically significant differences were detected between the groups for both Bax LI ($P = 0.004 < 0.01$) and PCNA LI ($P = 0.009 < 0.01$). When groups were compared further in terms of Bax LI and PCNA LI with Tukey's HSD test for multiple pairwise comparisons, statistically significant difference was observed only between groups 1a and 2b ($P = 0.006 < 0.01$).

INTRODUCTION

Helicobacter pylori (*H pylori*) is the most common chronic infection in human beings. *H pylori* infection has been related to peptic ulcer disease, gastric lymphoma and stomach cancers^[1,2]. It is an important problem for public health as it can cause several diseases.

Gastrointestinal mucosa is characterized with rapid epithelial cell turnover and homeostasis that is mainly provided by apoptosis^[3]. Therefore, it can be considered that the impaired apoptosis may have a role in the pathogenesis of many gastrointestinal system diseases.

In many studies, it has been shown that apoptotic cells are increased after colonization of *H pylori* in the stomach^[4,7], and those increased apoptotic cell levels decrease to normal levels after *H pylori* eradication^[5,8,9]. Increase in apoptotic cell rates causes compensatory hyperproliferative response in order to maintain gastric mucosal tissue mass^[10-12].

The mechanisms by which *H pylori* induces apoptosis of gastric epithelial cells have remained unclear. It has been suggested that many bacterial products might be inducing apoptosis^[5,13-15]. One of the factors that has been responsible for apoptosis of gastric epithelial cells is ammonia, which is produced via the degradation of urea by the urease enzyme of *H pylori*^[5,16]. Because of high intragastric urea concentrations in patients with chronic renal failure (CRF), levels of ammonia produced by *H pylori* are significantly high^[16]. Therefore, disorders associated with excessive ammonia production are more

prevalent^[17,18].

Although the relationship of *H pylori* with apoptosis and proliferation in gastric epithelial cells has been mostly studied in cases with normal renal functions, gastric epithelial cell kinetics in patients with CRF still remains unknown. This study aims to investigate the effect of *H pylori* on gastric epithelial cell kinetics in patients with CRF.

MATERIALS AND METHODS

Subjects

The study included 44 patients, of whom 23 with normal renal function and 21 with CRF who were on hemodialysis treatment.

Those with gastric and/or duodenal ulcers as confirmed by upper gastrointestinal endoscopy were excluded from the study. None of the patients included had a history of gastric surgery. Patients who had been treated for *H pylori* or who had used any antibiotics, proton-pump inhibitor, H₂ receptor blocker or any compound that includes bismuth during the last one month were also excluded from the study.

The CRF patients were on a thrice-weekly hemodialysis program. The dialysis prescription included 4-5 h of bicarbonate hemodialysis with standard cuprophane membranes (hollow fiber 1-1.2 m² surface area, Gambro, Sweden).

We divided the patients into four groups based on their *H pylori* and renal function status:

Group 1a: normal renal function, *H pylori* negative (12 patients)

Group 1b: normal renal function, *H pylori* positive (11 patients)

Group 2a: CFR, *H pylori* negative (10 patients)

Group 2b: CRF, *H pylori* positive (11 patients)

Endoscopy method

All patients were examined by using the upper gastrointestinal endoscopy (video-endoscope Olympus GIP Q240) after premedication with intravenous midazolam (2.5-7.5 mg), after an overnight fast.

During upper gastrointestinal endoscopy, multiple biopsy materials were taken from the antral region of the stomach. The biopsy specimens were transferred to the pathology laboratory in 10% buffered formalin. This study was conducted after getting permission from the local ethics committee of the Baskent University School of Medicine, Ankara, Turkey.

Identification of *H pylori* infection

H pylori was detected under microscopy on the histological sections stained with Giemsa staining method.

Laboratory methods

After a 12-h fasting period, venous blood samples were taken from the forearm-superficial veins of the patients in the morning. In the HD patients, the blood samples were taken before HD session. Serum levels of creatinine (normal <1.2 mg/dL) and blood urea nitrogen (BUN, normal <20 mg/dL) were measured in the central laboratory of our hospital by using routine automated

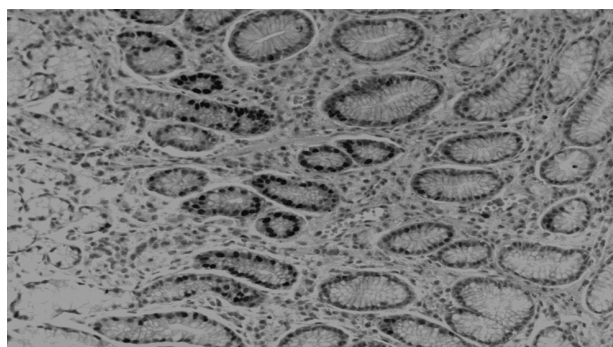


Figure 1 Proliferating cell nuclear antigen (PCNA) immunostaining of the gastric mucosa, original magnification $\times 250$.

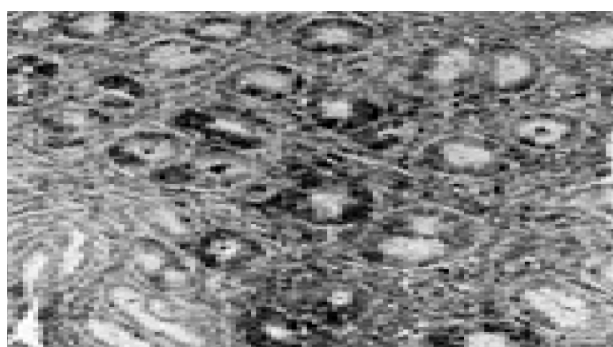


Figure 2 Bax immunostaining of the gastric mucosa, original magnification $\times 400$.

techniques.

Immunohistochemical staining for proliferating cell nuclear antigen and Bax

All biopsies were fixed in formalin, embedded in paraffin and processed routinely. Briefly, 4-mm-thick sections were deparaffinized and mounted on poly-L-lysine-coated slides. The sections in a citrate buffer (0.01 mol/L, pH 6) were heated in a microwave oven for 15 min at a maximum power (700 W), and then cooled at room temperature for 20 min. A standard three-step immunoperoxidase avidin-biotin peroxidase complex (ABC) technique was used to identify the proliferating cell nuclear antigen (PCNA) (PC10, Neomarkers, CA, USA) (Figure 1). A catalyzed signal amplification (CSA) system was used to detect the polyclonal antibody Bax (K1500, Dako, Denmark) (Figure 2). PCNA and Bax positive cells were counted in the glandular neck region, which corresponds to the area of cell proliferation^[19,20]. The field to be counted was chosen under $\times 400$ magnification from the well-labeled area. About 1 000 cells were counted in each case to determine the PCNA and Bax labeling indexes (LI). The percentage of positively stained cells over total cells counted was then calculated and used as a labeling index for PCNA and Bax expression^[21,22]. All histological slides were reviewed by the same experienced pathologist. The pathologist who rated the PCNA and Bax LI was unaware of the *H pylori* and renal function status of the patients.

Statistical analysis

Statistical analyses were carried out using the program SPSS 9.0 for Windows. To compare the groups, we used Student's *t*-test, Mann-Whitney *U* test, one-way ANOVA and Tukey's HSD test when appropriate. *P* values <0.05 were considered statistically significant.

RESULTS

The number of patients, their mean ages and HD duration are shown in Table 1 for each group separately. There were no statistically significant differences between the groups with regard to mean ages, gender, and HD duration (*P*>0.05).

Creatinine levels were 0.7 ± 0.2 , 0.8 ± 0.2 , 7.3 ± 1.1 , 7.1 ± 1.3 mg/dL, and BUN levels were 13.8 ± 3.5 , 15.4 ± 4.1 , 98.7 ± 12.2 , 96.1 ± 11.5 mg/dL in groups 1a, 1b, 2a, 2b, respectively (Table 2). Considering the creatinine and BUN levels, there were no statistically significant differences between the groups 1a and 1b (*P*>0.05), and between the groups 2a and 2b (*P*>0.05) but levels were higher in groups 2a and 2b than 1a and 1b (*P*<0.001).

Table 1 Number, gender, mean age, and hemodialysis duration of patient groups

	Group 1a	Group 1b	Group 2a	Group 2b
Patients (<i>n</i>)	12	11	10	11
Sex (M/F)	7/5	6/5	5/5	6/5
Mean age (yr)	37±7	39±7	35±8	36±11
Hemodialysis duration (mo)	-	-	10.4±4.5	9.3±4.4

Group 1a: normal renal function, *H. pylori* negative; group 1b: normal renal function, *H. pylori* positive; group 2a: CRF, *H. pylori* negative; group 2b: CRF, *H. pylori* positive.

For groups 1a, 1b, 2a, 2b, mean Bax LI was identified as 34.4 ± 13.7 , 44.1 ± 16.5 , 46.3 ± 20.5 , 60.7 ± 13.8 , respectively, and mean PCNA LI was identified as 36.2 ± 17.2 , 53.6 ± 25.6 , 59.5 ± 25.6 , 67.2 ± 22 , respectively.

When the one-way ANOVA test was applied, statistically significant differences were detected between the groups for both Bax LI (*P* = 0.004 <0.01) and PCNA

Table 2 Creatinine, BUN, Bax, and PCNA LI in different patient groups

	Group 1a	Group 1b	Group 2a	Group 2b
Creatinine	0.7 ± 0.2	0.8 ± 0.2	$7.3 \pm 1.1^{a,b}$	$7.1 \pm 1.3^{c,d}$
BUN	13.8 ± 3.5	15.4 ± 4.1	$98.7 \pm 12.2^{a,b}$	$96.1 \pm 11.5^{c,d}$
Bax labeling index	34.4 ± 13.7	44.1 ± 16.5	46.3 ± 20.5	60.7 ± 13.8^e
PCNA labeling index	36.2 ± 17.2	53.6 ± 25.6	59.5 ± 25.6	67.2 ± 22.1^e

Group 1a: normal renal function, *H. pylori* negative; group 1b: normal renal function, *H. pylori* positive; group 2a: CRF, *H. pylori* negative; group 2b: CRF, *H. pylori* positive. ^a*P*<0.001 group 1a vs 2a; ^b*P*<0.001 group 1b vs 2a; ^c*P*<0.001 group 1a vs 2b; ^d*P*<0.001 group 1b vs 2b; ^e*P*<0.01 group 1a vs 2b.

LI (*P* = 0.009 <0.01). When groups were compared further in terms of Bax LI and PCNA LI with Tukey's HSD test for multiple pairwise comparisons, statistically significant difference was observed only between groups 1a and 2b (*P* = 0.006 <0.01).

In patients with *H. pylori* infection and CRF, Bax LI and PCNA LI were found to be increased. However, the increase was more prominent in patients with CRF compared to patients with *H. pylori* infection.

DISCUSSION

This is the first study where cell turnover of gastric epithelial cells is investigated in patients with CRF. We have shown that both *H. pylori* infection and uremia cause increases in Bax and PCNA LI in gastric epithelial cells. Other studies in the literature that show *H. pylori* increases the proliferation and apoptosis in gastric epithelial cells in cases with normal renal function support the results of our study. In addition, we also noted that both apoptosis and proliferation in gastric epithelial cells are more increased in patients with CRF.

Maintaining gastric mucosal integrity is a complex biological process^[23]. This subject is provided with the balance between programmed cell death, which is also called apoptosis, and epithelial cell proliferation^[23–26]. Apoptosis has been accepted as a physiological form of death and is a genetically programmed process where the cell commits suicide^[27,28]. Unlike necrosis, dead cells do not cause inflammatory response in apoptosis^[28]. Deranged apoptosis has been implicated in carcinogenesis, autoimmune diseases, and various infectious diseases including *H. pylori* infection^[5,23,28,29]. Regulation of apoptosis is a complex process. This process includes the activation of various apoptosis-related proteins such as the Bcl-2 family, p53, Fas and its ligand (FasL), and the interleukin-1b-related concerning enzyme (ICE) family^[23,30].

There are several methods for evaluating cell division^[31]. PCNA is a co-factor of DNA polymerase and mainly determined in late G₁ or S phase. There are several evidences that PCNA assessment is a useful tool to evaluate cell proliferation and previous studies have shown that PCNA index correlates with S phase fraction of tumor cells determined by DNA flow cytometry^[32–35]. Furthermore, it has been found that PCNA immunostaining correlates with Ki67, the latter marking cells in G₁ and G₂ phases in addition to those in S phase and also with thymidine labeling index^[36] and with bromodeoxyuridine uptake in cancer cell lines^[20,37].

Many genes regulate the balance between apoptosis and cell proliferation. The pro-apoptotic protein, Bax, is a member of Bcl-2 gene family and is one of the genes that regulate apoptosis. Bax gene is a tumor suppressor gene and induces apoptosis via encoding Bax protein. It has been suggested that proliferation and apoptosis in the antral mucosa caused by *H. pylori* is associated with the alterations in genes that regulate the process^[8,22,38,39]. Konturek et al^[23] showed that apoptosis induced by *H. pylori* in gastric epithelial cells is associated with the upregulation

of proapoptotic Bax.

In many studies, it has been shown that the number of apoptotic cells in the stomach increase after *H pylori* colonization^[4-6]. In addition, it has also been shown that the apoptotic cell number in stomach decreases to normal after *H pylori* eradication^[5,8,9]. This increase in apoptotic rate causes compensatory hyperproliferative response in order to maintain gastric mucosa tissue mass^[11].

In the literature, there are some data which suggest that apoptosis is associated with both bacterial and host factors^[13-15,23]. However, the mechanism of apoptosis in *H pylori* infection is still not clear.

Some of the factors accused within the apoptosis are lipopolysaccharides, ammonia, and monochloramine (NH₂Cl), which is a highly toxic substance generated as a result of the reaction of ammonia with neutrophil-derived free radicals^[13-15]. There are still debates on the role of cytotoxic agents such as *H pylori*-CagA and *H pylori*-VacA in apoptosis. In recent *in vivo* and *in vitro* studies, it has been found that *H pylori* strains can induce apoptosis without any dependency on Cag-A and Vac-A genotype^[23]. In addition to the bacterial factors in infections of *H pylori*, characteristics of the host can also play important roles. Evidences are increasing about the inflammatory reactions that are induced by *H pylori* in gastric mucosa, especially cytokines such as nitric oxide and TNF α might assist to induce apoptosis^[6].

One of the major factors that are responsible for *H pylori* derived apoptosis is ammonia. *H pylori* has a strong urease activity and the hydrolysis of urea causes production of high level of ammonia concentration in mucous gel layer^[40]. *In vitro* studies have shown that *H pylori* derived ammonia causes lesions in insulated gastric epithelial cells. Ammonia concentrations, determined in *H pylori* infected subjects, cause gastric mucosal lesions^[40-42], delay in mucosal improvement^[43], and induce apoptosis in gastric epithelial cells^[13,44].

Ammonia level, which is produced by *H pylori*, is controlled by the existence of urea in gastric juice. Patients with CRF have high intragastric urea concentrations and levels of ammonia produced by *H pylori* are significantly high^[16]. Therefore, disorders associated with excessive ammonia production may be more prevalent in patients with uremia. In our cases, increased apoptosis and proliferation rate that are observed in uremia patients can be associated with excessive ammonia production by *H pylori* infection in uremia patients compared with cases that have normal renal functions.

In conclusion, in gastric epithelial cells, expression of both the pre-apoptotic protein Bax and the proliferation marker PCNA increase with *H pylori* infection. This increase is more evident in patients with uremia. These findings suggest that uremia accelerates apoptosis and proliferation in gastric epithelial cells.

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