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REVIEW

Holistic perspective of the role of gut microbes in diabetes mellitus and its management

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Alagiakrishnan K and Halverson T contributed equally by reviewing the literature and drafting the manuscript; both the authors have read and approved the final version.

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Abstract

The gut microbiota (GM) plays a role in the development and progression of type 1 and type 2 diabetes mellitus (DM) and its complications. Gut dysbiosis contributes to the pathogenesis of DM. The GM has been shown to influence the efficacy of different antidiabetic medications. Intake of gut biotics, like prebiotics, probiotics and synbiotics, can improve the glucose control as well as the metabolic profile associated with DM. There is some preliminary evidence that it might even help with the cardiovascular, ophthalmic, nervous, and renal complications of DM and even contribute to the prevention of DM. More large-scale research studies are needed before wide spread use of gut biotics in clinical practice as an adjuvant therapy to the current management of DM.

Key Words: Probiotics; Prebiotics; Synbiotics; Diabetes mellitus; Microbial dysbiosis; Antidiabetic drugs

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Core Tip: The emerging role of the gut microbiome on diabetes development, progression as well as prevention has been discussed in this manuscript. The significance of gut dysbiosis in the aetiopathogenesis of diabetes mellitus and its complications has been reviewed. A bidirectional relationship exists between the antidiabetic drugs and the gut microbiome. Faecal transplantation, and bariatric surgery, typically used to treat morbid obesity, have also been shown to improve commensal gut microbiota changes. Diabetic outcomes and management can improve with better understanding of the drug-gut microbiome interactions. There is emerging evidence pointing out that gut biotics can be an add-on therapy with the antidiabetic management. To our knowledge, there is no evidence about the role of gut microbes of diabetic patients who had pancreatic cell transplantation, as well as the role of gut

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biotics influencing the management in this group.

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INTRODUCTION

Globally diabetes mellitus (DM) is a common medical disorder and is seen in pandemic proportions[1] with the global prevalence in adult subjects is roughly 10% [2]. The International Diabetes Federation projected by 2035, there will be 592 million cases of diabetes in the world[3]. DM type 1 is secondary to auto-immune- mediated loss of beta-cell function and is seen in 5% of the diabetic population. DM type 2 is mainly due to insulin resistance and is seen in 95% of diabetic subjects[4]. The 2016 US National Health Interview Survey data showed roughly 8.58% of the population had type 2 DM and 0.55% had type 1 DM[5].

Various research has been done in the last decade since the study of the human microbiome in 2012[6,7]. Microbes contribute to 2% of human body weight and the bacterial genomes exceeds human genes by a factor of 150[8,9]. Gut microbiota (GM) varies with age, diet, geographical location, life style, and the use of xenobiotics[10-12]. In the recent years there have been more focus on the GM in the development, progression, and distant organ complications due to DM[13]. Many studies have shown the role of the gut microbiome in DM[14-17].

The gut microbiome starts to develop with the mode of birth and it is influenced by environmental factors, diet, as well as certain medications, including antibiotics [18]. There are differences between the gut microbes seen between non-diabetic and diabetic subjects[20] (Table 1). Gut dysbiosis plays a role in numerous diseases including DM. Both altered GM and endocrine disrupters can influence the development of DM[21]. In this literature review, we analyzed the evidence for the role of GM in the development, pathogenesis, complications, management, and prevention of DM.

LITERATURE SEARCH

A literature search was performed using the electronic databases MEDLINE (1966-February 2021), EMBASE and SCOPUS (1965-February 2021), and DARE (1966-February 2021). The main search items were gut bacteria, GM, intestinal flora, gut dysbiosis, type 1 DM, type 2 DM, diabetic retinopathy, diabetic neuropathy, diabetic nephropathy, probiotics, prebiotics, synbiotics, bariatric surgery, and faecal transplantation. Non-English articles were excluded.

GM IN TYPE 1 DM

Studies have shown that Firmicutes/Bacteroides ratio is altered in type 1 DM[22]. In the study by Huang et al[23] (2018) negative association was seen with gut microbe Faecalibacterium and Ruminococcacea and hemoglobin A1c (HbA1c), whereas in the study by Fassatoui et al[24]. (2019) a negative association was seen between HbA1c and Akkermansia muciciniphia. A systematic review of studies done in Hispanic populations showed that patients with newly diagnosed type 1 DM have high levels of Bacteroides with a reduced proportion of Prevotella, Megamonas, and Acidaminococcus. With the initiation of insulin treatment these subjects showed an increase of Prevotella levels. Prior to the development of type 1 DM, inverse relationship of Firmicutes/ Bacteroidetes ratio has been reported [25].

Table 1 Changes in the iniciobiolite in type 1 and type 2 diabetes memitas					
Location	Change in microbiome				
Type 1 diabetes					
Gastrointestinal tract	(1) Decreased: Prevotella; Megamona; Acidaminococcus; and (2) Increased: Bacteriodes	Elena <i>et al</i> [25], 2019			
Gastrointestinal tract	(1) Decreased: Bifidobacterium adolescentis; Bifidobacteria; and (2) Increased: Clostridium perfingens; Bacteroides	De Goffau <i>et al</i> [122], 2013			
Gastrointestinal tract	Increased: Leptotrichia goodfellowii; Bacillus cerus; Enterobacter mori LMG 25706	Tai <i>et al</i> [123], 2016			
Gastrointestinal tract	Increased: Bacteroidetes/Firmicutes	Giongo <i>et al</i> [124], 2011			
Gastrointestinal tract	(1) Decreased: Faecalibacterium prausnitzii; and (2) Increased: Bacteroides dorei; Bacteroides vulgatus	De Goffa <i>et al</i> [125], 2014			
Gastrointestinal tract	(1) Decreased: Prevotella; Akkermansia; Bifidobacterium adolescentis; Roseburia faecis; Faecalibacterium prausnitzii; and (2) Increased: Dialister invisus; Gemella sanguinis; Difidobacterium longum	Brown <i>et al</i> [126], 2011			
Type 2 diabetes					
Gastrointestinal tract	(1) Decreased: Clostridium coccoides; Clostridium leptum; and (2) Increased: Lactobacillus	Chen <i>et al</i> [28], 2019			
Gastrointestinal tract	(1) Decreased: Bifidobacterium; Bacteroides; Faecalibacterium; Akkermansia; Roseburia; and (2) Increased: Ruminococcus; Fusobacterium; Blautia	Gurung et al[30], 2020			

(1) Decreased: Aquabacterium; Xanthomonas; Pseudonocardia; and (2) Increased: Actinotalea; Alishewanella;

GM IN TYPE 2 DM

(1) Decreased: Bifidobacterium; Akkermansia; and (2) Increased: Dorea

(1) Decreased: Bifdobacterium; and (2) Increased: Lactobacillus

Seiminibacterium; Pseudoclavibacter

Table 1 Changes in the microbiome in type 1 and type 2 diabetes mellitus

The type of gut microbes and the changes seen with them influence the development of DM. The prominent GM seen in the intestine are the gram-positive Firmicutes and gram-negative Bacteroidetes and it is influenced by dietary changes [26]. A change in the ratio of Bacteroidetes to Firmicutes is associated with DM[27]. A case-control study done by Chen et al[28] (2019) in newly diagnosed type 2 DM subjects, Lactobacillus faecal count was significantly higher whereas Clostridium coccoides and Clostridium leptum was lower, and these changes in the microbes was positively correlated with glycated hemoglobin with higher Lactobacillus count subjects, and negatively correlated with decreased Clostridium count subjects when compared with healthy controls. Another study found that patients with DM showed an affiliation with the following phyla of bacteria: Firmicutes, Bacteroidetes, Proteobacteria and Actinobacteria^[29]. Alterations in the gut microbe population may be related to DM, and gut microbes Ruminococcus and Fusobacterium has been shown with the development of type 2 DM, when compared to healthy adults[30]. A study by Sedighi et al[31] (2017) found that patients with type 2 DM has increased levels of Lactobacillus, while healthy controls showed increased Bifidobacterium. With respect to the Lactobacillus genus, there are various mixed results suggesting its association with type 2 DM. Certain strain such as L. acidophilus, L. gasseri, and L. salivarius have been increased where as L. amyloyorus has been decreased[30]. However, many species from this genus, such as L. plantarum, L. casei, and L. rhamnosus are often involved in probiotic preparation and have shown to be beneficial in diabetic mice models[30]. Overall, it looks that there may be a strain-specific association with DM.

Further changes in the microbiome in patients with DM are listed in Table 1. Nutrient imbalance by affecting the GM can influence the development of type 2 DM. With newly diagnosed type 2 DM different measurement parameters like age, blood lipids, body-mass index, blood pressure, and dietary nutrient intake was related to the gut microbiome composition[32].

Gastrointestinal

Gastrointestinal

tract

tract Blood Li et al[127], 2020

Sedighi et al[31], 2017

Qiu et al[38], 2019

RELATIONSHIP BETWEEN GUT AND BLOOD MICROBIOME AND ITS **ASSOCIATION WITH TYPE 2 DM**

Cani et al[33] (2008) in their animal study showed lipopolysaccharide produced by gram negative intestinal bacteria can translocate into systemic circulation through a leaky gut and can result in endotoxemia causing metabolic dysfunction and obesity. Recent evidence points out in addition to gut microbiome, the blood microbiome plays a role in DM. Blood is usually considered to be sterile, but the research suggests the presence of a microbe or microbial component in healthy humans is known as a blood microbiome. The evidence for blood human microbiome is slowly growing [34-36].

In a study by Sato et al[37] (2014) with Japanese type 2 DM subjects, blood microbiome translocation from gut microbiome was detected at a higher rate (28%) in type 2 diabetic subjects when compared with healthy controls (4%) (P < 0.01). A recent nested case control study by Qiu et al[38] (2019) showed the blood microbe Sediminibacterium is associated with increased risk of type 2 DM [Odd ratio (OR) = 14.098, 95%CI: 1.358-146.330] where as the microbe *Bacteroides* in blood have a reduced risk for type 2 DM (OR = 0.367, 95%CI: 0.151-0.894).

GM AS A COMPLEX ENDOCRINE ORGAN

The regulation of the GI system is done by short-chain fatty acids (SCFA) derived from the metabolism of carbohydrates, and GM plays a role in this function. In addition, the gut microbes produce hormone like chemicals that can act at distant targets. Neuroactive compounds like tryptophan and neurotransmitters like serotonin, dopamine, noradrenaline, GABA, and hormones like leptin, ghrelin and glucagon-like peptide 1 (GLP-1) are indirectly regulated by SCFAs via enteroendocrine cells. Overall, the gut microbes produce several substances of a hormonal nature into the circulation which act as distant sites. Because of the GM's ability to influence distant organs and systems as mentioned above it is considered as an endocrine organ. Overall GM functions as an autonomous endocrine organ and plays a role in bodily endocrine actions including neuroendocrine and immunoendocrine regulations[39-42].

DM AND GUT DYSBIOSIS

Gut dysbiosis, is a state of increased or altered prevalence of gut bacteria which might in turn result in many disorders such as gastrointestinal, obesity, DM, immunological, and neurobehavioral diseases[43]. Shifts in the GM's composition with more pathogenic species and phyla can contribute to the above-mentioned diseases. Hyperglycemia was associated with changes of microbiota composition, preferring the non-commensal ones, on the detriment of beneficial phyla such as Bacteroidetes, Proteobacteria, and Actinobacteria. The ratio of Firmicutes/Bacteroidetes has been found to be correlated with plasma glucose concentration. Microbiota are capable to ferment undigested carbohydrates, fiber, and other dietary and xenobiotic compounds to produce SCFAs, which through their ubiquitous receptor play an important role in host glucose metabolism[37,44,45]. The Human Microbiome plays a role in gut permeability, modification of bile acids, glucose breakdown and in the absorption of nutrients[46,47].

Normal commensal bacteria are helpful in maintaining the gut wall integrity, innate immunity, insulin sensitivity, metabolism, and in communication with the brain functions, as well as help to prevent the penetration of harmful microorganisms in the bowel. Bidirectional relationship exists between the GM and the brain. This chain of communication depends on the interaction of gut microbe through immune and neuroendocrine system with the central nervous system. Short-chain fatty acids, such as butyrates, acetates and propionates, produced by the GM are beneficial to different metabolic processes. The imbalance between the microbiome and host organism lead to dysbiosis. Gut microbiome dysbiosis through inflammation and metabolic dysregulation increases insulin resistance and influence the development of type 2 DM[48] (Figure 1).

Microbial dysbiosis can also be the result of nutritional imbalance which can lead to a low-grade inflammatory state, obesity, and other metabolic disorders[49]. Gut microbes affect gut permeability, glucose and lipid metabolism, energy homeostasis, and insulin sensitivity. Like any other medical conditions, gut microbes play a role in

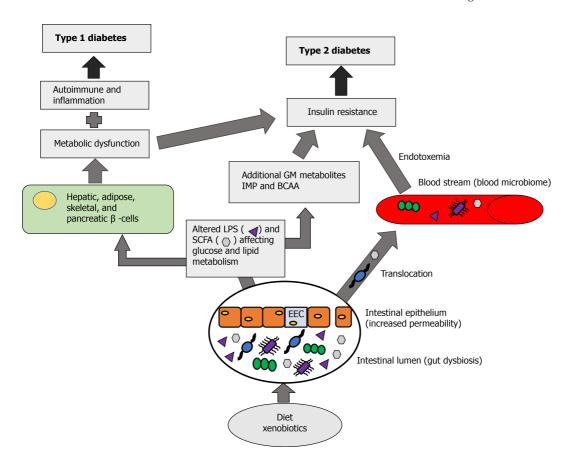


Figure 1 The role of gut dysbiosis in diabetes mellitus. The ingestion of a diet rich in carbohydrates and fats along with certain xenobiotics can lead to a disruption of the gut microbiome (dysbiosis). Under normal conditions, the gut bacteria produce metabolic products such as short chain fatty acids (SCFA) (Hexagons) that act locally and have a positive benefit on metabolism. Under conditions of dysbiosis there can be a disruption to the enteroendocrine cells and lead to gut permeability. This can lead to an increase in these metabolic products as well as bacterial translocation to the bloodstream, leading to endotoxemia resulting in metabolic dysfunction and insulin resistance contributing to type 2 diabetes. Gut dysbiosis also results in altered production of SCFA and release of lipopolysaccharides (LPS) (Triangles) and an increase production of other metabolites such as imidazole propionate and bacteria derived amino acids. These metabolites can act directly to affect insulin resistance. Excess SCFA and LPS can act on hepatic, skeletal, adipose, and pancreatic cells leading to metabolic dysfunction, altered inflammation and immune response which can influence insulin resistance. These factors can contribute to the development of type 1 and type 2 diabetes. SCFA: Short chain fatty acids; EEC: Enteroendocrine cells; LPS: Lipopolysaccharides; BCAA: Bacteria derived amino acids; IMP: Imidazole propionate; GM: Gut microbiota.

inflammation and immunity[50]. A diet rich in fat and sugar may lead to an abundance of lipopolysaccharide (LPS) release from GM and this LPS, by entering into systemic circulation, can affect β -cells, leading to decreased insulin release, and thereby altering systemic insulin sensitivity, resulting in insulin resistance, and potentially leading to DM[51].

Diets rich in carbohydrates and fat as well as xenobiotics (medications affecting the gut microbes) can cause gut dysbiosis. Normally GM produces metabolic products like SCFA, acetate, butyrate and propionate which acts locally leading to beneficial effects on different metabolic process. When there is gut dysbiosis, it can affect the enteroendocrine L-type cells in the intestinal epithelium and increase the gut permeability (leaky gut) causing these metabolic products to enter into the systemic circulation, as well as translocation of the gut microbiome into the circulation leading to the formation of the blood microbiome. This blood microbiome can cause endotoxemia and affect both metabolic dysfunction and insulin resistance. Gut dysbiosis results in excessive production of SCFA and LPS, as well as additional GM metabolites like imidazole propionate (IMP), derived from histidine, and bacteria derived amino acids. Excessive SCFA and LPS by acting on hepatic, adipose, skeletal and pancreatic cells causes metabolic dysfunction, inflammation and altered immune response. When there is a metabolic dysfunction due to gut dysbiosis combined with inflammatory and altered immune response it can cause type 1 DM, and when combined with insulin resistance due to gut dysbiosis as well as the effect of blood microbiome it can lead to the development of type 2 DM (Figure 1).

GUT MICROBES AND METABOLIC NETWORKS

The human gut contains a wide variety of microbial communities that carry out a wide range of biochemical functions that can influence the human body through metabolite production, physiological regulation, and interacting with the host's cellular response and immunity[52]. It has also been found that the host's own genetics can influence the composition of their gut microbiome, making each host a unique ecosystem[53]. Dynamic changes in the gut microbiome have been seen within individuals often in various disease states, such as obesity, and DM[19,54-56]. The GM has been found to cause enhanced transcriptional changes in the intestinal cells and protein biosynthesis in the crypts within the intestine [57].

SCFAs produced by GM can serve as signaling molecules that can influence the host's lipid and glucose levels, liver, skeletal muscle, and even immunity[52]. When there is a disruption of the gut microbiome, the altered mixture of SCFA may influence obesity, insulin sensitivity, weight gain and other comorbidities[58,59]. Obese individuals with type 2 DM have shown changes in the GM that are distinct, from non-diabetic subjects. It was found that individuals with type 2 DM showed an increase level of *Proteobacteria* and *Bacteroides* with a decreased level of *Firmicutes* [19].

The GM has been found to influence the host's metabolism and show great adaptability to the changing environment within the intestines based on diet, genetics, and various physiological cues from the host [52]. The human gut microbiome can modulate absorption as well as nutrient availability within the host. This can be achieved through gene expression changes, alteration of hormones and immunity [52].

ASSOCIATION BETWEEN MICROBIOME, OBESITY AND DM

Microbial diversity and the production of SCFA as well as products such as butyrate, propionate, and acetate have been found to have a protective role against obesity and insulin resistance [60,61]. SCFAs are able to act as signaling molecule that can activate a variety of pathways that are involved in cholesterol, lipid, and glucose metabolism [58]. Modifications of the microbiome can influence metabolic parameters, in particular when there is a higher abundance of Firmicutes leading to a higher Firmicutes/Bacteroidetes ratio that may be linked to obesity[62]. This may in part be due to the fact that Firmicutes are more efficient at promoting the nutrient absorption leading to subsequent weight gain compared to Bacteroidetes[63].

A study showed the GM composition is different in obese subjects with and without type 2 DM[20]. A recent study also showed for the first time in subjects with type 2 DM the relationship between body composition and GM[64]. Faecal microbiota of obese subjects without DM had increased numbers of SCFA producing microbes, whereas obese subjects with type 2 DM had less beneficial SCFA butyrate producing microbes[65].

ROLE OF GUT MICROBES IN THE PROGRESSION OF DM

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The progression of DM is seen as macrovascular [66] and microvascular complications like retinopathy, nephropathy, and neuropathy[67]. Gut microbes seem to play a role in the progression of DM and also shown to play a role in these complications. Diet induced diabetic animal models helps to study these complications[68]. Studies have shown that subjects with DM and eye complications have higher bacterial conjunctival flora when compared to subjects without DM[69-72]. Beli et al[73] (2018) in their animal study showed the association between the GM and diabetic retinopathy (Table 2). More research is needed to understand the mechanism how GM causes diabetic retinopathy[74].

Diabetic neuropathy is seen as autonomic neuropathy as well as distal sensory and motor neuropathy and correlate with diabetic control, and GM also seems to play a role[75]. In a human study with early diabetic nephropathy, Barrios et al[76] (2015) showed an increase in colonic GM, whereas with end-stage renal disease patients microbes producing urease, uricase, p-cresol and indole-forming enzymes were seen [77]. The proposed mechanisms for progression of kidney disease could be due to GM imbalance, metabolic shifts, immunosuppression, inflammation, as well as accumulation of uremic toxins[78].

Table 2 Selected animal studies showing the effect of various interventions on the gut microbiome and the role of gut microbiota in diabetes mellitus management

Intervention	Organism	Health benefit	Change in microbiome	Ref.
Intermittent fasting	Mice	Protection from diabetic retinopathy by increasing Tauroursodeoxycholate (a neuroprotective bile acid) producing microbes	Increased Firmicutes and decreased Bacteroidetes and Verrucomicrobia in diabetic mice undergoing intermittent fasting	Beli <i>et al</i> [73], 2018
Antibiotic treatment (ampicillin, metronidazole, neomycin, vancomycin, or their cocktail)	Mice	Reduction in fasting glucose. Change in glucose tolerance (seen with ampicillin, vancomycin, or cocktail)	Alterations in the α - and β - diversity. An association with $Akkermansia$ $mucinipjila$ with decrease fasting glucose. The effect is mediated through systemic changes in glucose metabolism	Rodrigues <i>et al</i> [94], 2017
Prebiotic: Acorn and sago	Mice	Mice fed acorn and sago derived prebiotics had an amelioration of the glucose intolerance and insulin resistance induced by a high-fat diet feeding. Intake of both novel prebiotics as well as inulin increases SCFAs levels in the mouse gut		Ahmadi <i>et al</i> [103], 2019
Combination of a functional fibre [PolyGlycopleX (PGX) with metformin (MET) or sitagliptin and metformin (S/MET)]	Mice	PGX + MET and PGX + S/MET showed reduced glycemia compared to controls and single treatment ($P = 0.001$). HbA1c was lower in PGX + S/MET compared to all other treatments ($P = 0.001$)		Reimer <i>et al</i> [93], 2014
Artificial sweetener (Neotame)	Mice	Decreased butyrate synthetic genes in Neotame group. Higher concentrations of cholesterol ($P < 0.05$) and fatty acids ($P < 0.05$) in Neotame treated mice feces	Reduction in α -diversity and altered β -diversity. Reduced Firmicutes ($P < 0.01$) and increased Bacteroides ($P < 0.01$)	Chi et al [85], 2018
Combination of metformin and a prebiotic [konjac mannan-oligosaccharides (MOS)]	Mice	Combination of metformin and MOS help ameliorate insulin resistance and improved glycemic control ($P < 0.05$) and repair islet and hepatic histology	Metformin and MOS change the microbiome (<i>P</i> < 0.0001) with: Decreased: Rikenellaceae and Clostridiales; Increased: <i>Akkermansia muciniphila</i> and <i>Bifidobacterium pseudolongum</i>	Zheng <i>et al</i> [96], 2018

MANAGEMENT

In DM, normal GM can be restored using diet, gut biotics, faecal transplantation, and bariatric surgery, which may help with the proper management of DM.

Faecal transplant, bariatric surgery

There is some evidence from human studies, that both faecal transplant and bariatric surgery improved the glucose and metabolic parameters by altering the GM[48]. A meta-analysis done by Magouliotis et al[79] (2017) showed some discrepancy between the human studies and the benefits witnessed from bariatric surgery. Another study looking at obese insulin resistant subjects who received allogenic faecal transplants from a lean insulin sensitive donor show improved insulin sensitivity for a short period of 6 weeks, however the benefit was not seen past 12 weeks[80] (Table 3).

Nutritional therapy

Diet can modulate the GM and play a role in the management of DM by preventing gut dysbiosis[81] (Table 2). Fruits and vegetables contain polyphenols which can increase beneficial GM like A. muciniphila, Lactobacilli and Bifidobacteria [82]. Unbalanced dietary intake can affect the structure and abundance of GM which can play a role in the development of DM[83].

Artificial sweeteners

Artificial sweeteners are no-calorie sugar substitutes, may induce glucose intolerance by affecting the gut microbes. In an animal study with saccharin-fed mice showed an increase in Bacteroides and a reduction in Lactobacillus reuteri leading to GM dysbiosis and glucose intolerance [84]. Similar effects were seen in another study by Chi et al [85] (2018) using the artificial sweetener, Neotame. In a cross-sectional human study by Frankenfeld et al [86] (2015), showed sweeteners like aspartame or acesulfame-K found no effect on gut bacterial abundance. A recent randomized-blinded crossover study in healthy participants did not demonstrate measurable changes in the GM or in SCFAs after 14 d daily intake of aspartame and sucralose[87]. These preliminary observations

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Table 3 Selected human studies showing the effect of diet, gut biotics, faecal transplantation and bariatric surgery on gut microbiome and the role of gut microbiota in diabetes mellitus management

Intervention	Organism	Health benefit	Change in microbiome	Ref.
Probiotics	Human	Decreased fasting blood glucose and HbA1c levels. Increased HDL levels, however no significant effect on BMI and LDL levels were found		Kocsis <i>et al</i> [112], 2020
Artificial sweeteners (aspartame and acesulfame-K)	Human		Compared to controls, aspartame and acesulfame-K had different bacterial diversity ($P < 0.01$, $P = 0.03$ respectively), compared to controls	Frankenfeld <i>et al</i> [86], 2015
Probiotics, Prebiotics, or synbiotics	Human (meta- analysis)	The use of probiotics, prebiotics, or synbiotics showed a decrease in FBG ($P < 0.01$), total cholesterol ($P = 0.02$), triacylglycerols ($P = 0.01$) and insulinaemia ($P < 0.01$), as well as increased HDL-cholesterol levels ($P < 0.01$. Even though HbA1c reduction is seen it is not statistically significant. No effect on LDL-cholesterol was seen		Bock et al[115], 2020
Laparoscopic sleeve gastrectomy	Human	Decreased weight and BMI. Restored insulin tolerance and type 2 DM remission	Increased: <i>Bacteroidetes/Firmicutes</i> ratio at 1- and 3-months post surgery. <i>Lactobacillales</i>	Kikuchi <i>et al</i> [128], 2018;Li <i>et al</i> [129], 2013
Roux-en-Y gastric bypass	Human	Type 2 DM remission and improved BMI and weight loss. Improved gastric emptying and bile acid metabolism	Decreased: Bacteroidetes/Firmicutes ratio. Improved probiotic supplementation effects due to lowered pH environment	Selber-Hnatiw <i>et al</i> [52], 2020; Li <i>et al</i> [129], 2013

BMI: Body mass index; HbA1c: Hemoglobin A1c; FBG: Fasting blood glucose; HDL: High-density lipoprotein; LDL: Low-density lipoprotein.

needed to be established in future human research studies.

ALTERATION OF GM BY ANTIDIABETIC DRUGS AND ITS ROLE IN DM **MANAGEMENT**

Antidiabetic drugs can influence the gut microbiome by affecting the drug microbiome interface, whereas the gut microbiome also influences the metabolism and play a role in the efficacy of antidiabetic drugs. The interactions of antidiabetic drugs and microbiota is getting more attention as it may play a role in the management of DM [88]. Antidiabetic agents cause alteration of the specific gut microbes. Metformin increases the population of Akkermansia muciniphila by 18-fold, enhancing the digestion of mucin and increasing SCFA[89]. Metformin, in addition to Akkermasia, causes increase in Lactobacillus and Bifidobacterium, whereas insulin increase Fusobacterium [90]. This first line antidiabetic agent in type 2 DM modifies the GM, alter the bile acid circulation and thereby a possibility that primary site of action may be gut and the GM [91].

Understanding the pharmacokinetics, pharmacodynamics and pharmacomicrobiomics of antidiabetic medications and gut microbes can help to understand druggut microbiome and its potential benefit with antidiabetic drugs. Overall, it may help to better manage the DM management[92].

Antidiabetic drugs have been shown to affect the different gut microbes and their metabolic effects through the medication-microbiome-metabolism axis. GM can influence the pharmacokinetics of various antidiabetic drugs such as drug absorption, drug metabolism which can affect the potency of these medications. Overall, there is a bidirectional relationship exist between antidiabetic drugs and gut microbes[88].

Different combinations of antidiabetic drugs are used to better control DM. The commonly used combination is metformin with sulphonylureas, thiazolidinediones, DPP-4 inhibitors and insulin. One animal study showed some delay in the progression of DM when sitagliptin/metformin combination given with a prebiotic fibre[93]. Currently, there is a need for more research of different combination therapies on GM.

GUT BIOTICS AND DM

Animal studies

Several animal studies have showed that gut biotics, like prebiotics and probiotics, can improve the efficacy of antidiabetic drugs. Treatment with individual or a cocktail of antibiotics reduced dysbiosis and decrease fasting glucose but did not affect body weight, as well as antibiotic treatment also changed gene expression in the ileum and liver, and shifted the alpha (α) and beta (β) diversities of GM[94]. In an animal study with mice, combining probiotics and/or prebiotics with antidiabetic medications showed an improvement in glycemic control and insulin sensitivity [95]. A study by Reimer et al [93] (2014) found that using a combination of sitagliptin and metformin with a functional fiber can delay DM progression. In an animal study usage of mannan-oligosaccharides by altering the GM increased the hypoglycemic effects of metformin[96]. Yang et al[97] (2020) found that Genistein found in soybeans and soy derived foods (prebiotic) helped to improve glucose and lipid metabolism by altering GM composition[97]. In another animal study, certain GM like Bacteroides fragilis, A. muciniphila, L. plantarum, L. casei can induce interleukin 10 (IL-10), which has been shown to improve both insulin resistance and glucose metabolism[98] (Table 2).

Human studies

Many gut microbes have been shown to have antidiabetic effect in humans by different mechanisms including effect on insulin sensitivity [99]. Roseburia intestinalis can improve insulin sensitivity by increasing IL-22 production [100]. Some strains of Lactobacilli act like acarbose and have been shown to inhibit alpha glucosidase[101]. Prebiotics can feed the gut microbiome and increase the population of L-cells in the intestine and thereby increase the amount of GLP-1[102] and prevent high fat diet induced insulin resistance[103]. In the recent PREMOTE randomized control trial (RTC) study, probiotics showed antidiabetic effect by altering metabolic homeostasis [104]. Thus, GM may be useful in the management of DM[105]. Jafarnejad et al[106] (2015) and Asemi et al[107] (2014), in their two studies showed multi-probiotic supplement as well as synbiotic (L. sporogenes plus inulin) product helps to reduce glucose and other metabolic parameters. Tonucci et al[108] (2015) in their double-blind RCT study comparing fermented milk containing L. acidophilus (LA-5) plus B. animalis (Lactis BB-12) with plain fermented milk in 45 type 2 DM subjects showed decreased in HbA1c as well as low-density lipoproteins cholesterol and inflammatory cytokines. Multiple RTCs and the meta-analysis of these RCT's with different gut microbes demonstrated antidiabetic effect as well as effect on different metabolic parameters [109-111] (Table 3).

A recent meta-analysis of 14 RCTs showed significant decrease in HbA1c in the probiotic group compared to placebo controls, weighted mean difference (WMD) is -0.33%, 95%CI -0.53 to -0.13, P = 0.001. In this meta-analysis, probiotics significantly reduced fasting blood glucose, insulin, lipid profile and inflammatory marker in addition to blood pressure levels[112]. Another meta-analysis showed similar result with reduction in HbA1c% (WMD = -0.24, 95%CI: -0.44 to -0.04, P = 0.02), fasting blood glucose (WMD = -0.44 mmol/L, 95%CI: -0.74 to -0.15, P = 0.003)[113,114]. A meta-analysis study done in 2021 with probiotics, prebiotics or synbiotics on type 2 DM also showed significant improvement in glucose and other metabolic parameters [115]. Prebiotic inulin improves glycemic control in young adults with type 1 DM [116]. Certain specific species of probiotic microbes as well as certain prebiotics by altering the GM was shown to improve the auto-immune condition, which plays a major role in the pathogenesis of type 1 DM[117].

A study by Didari et al[118] (2014) looked at the safety of probiotics and synbiotics and found that certain populations, such as patients who are immunocompromised, with cardiac valvular disease, having a central venous catheter, or those with shortbowel syndrome may have an increased risk for systemic infections. Thus, caution may be warranted when using these products in diabetic patients and a risk-benefit analysis should be considered.

GUT MICROBES AND THE PREVENTION OF DM

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Some preliminary evidence in animal studies indicates altering GM may help to prevent DM[119,120]. A recent study by Gurung et al[30] (2020) showed with certain gut microbes like Bifidobacterium, Bacteroides, Faecalibacterium, Akkermansia and Roseburia have a negative association with DM and appears to be protective. In 42 healthy adults, GM Lactobacillus johnsonii seems to reduce the risk of type 1 DM[121].

CONCLUSION

Gut dysbiosis plays a role in the development and progression of DM. The current evidence also points out that the GM can play a role in DM related complications. Modulation of the gut bacteria or dysbiosis can be corrected by fibre, diet, antidiabetic medications, and by using gut biotics like prebiotics, probiotics, and synbiotics as well as by bariatric surgery and faecal transplantation. The interaction between gut microbes and antidiabetic agents is a promising field that may change the landscape of DM management in the future. There is some preliminary evidence to show that GM may play a role in the prevention of DM. More research is needed on a large scale to confirm these findings.

REFERENCES

- Toniolo A, Cassani G, Puggioni A, Rossi A, Colombo A, Onodera T, Ferrannini E. The diabetes pandemic and associated infections: suggestions for clinical microbiology. Rev Med Microbiol 2019; **30**: 1-17 [PMID: 30662163 DOI: 10.1097/MRM.0000000000000155]
- NCD Risk Factor Collaboration (NCD-RisC). Worldwide trends in diabetes since 1980: a pooled analysis of 751 population-based studies with 4.4 million participants. Lancet 2016; 387: 1513-1530 [PMID: 27061677 DOI: 10.1016/S0140-6736(16)00618-8]
- 3 Ayadurai S, Hattingh HL, Tee LB, Md Said SN. A Narrative Review of Diabetes Intervention Studies to Explore Diabetes Care Opportunities for Pharmacists. J Diabetes Res 2016; 2016: 5897452 [PMID: 27247949 DOI: 10.1155/2016/5897452]
- 4 American Diabetes Association. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes-2018. Diabetes Care 2018; 41: S13-S27 [PMID: 29222373 DOI: 10.2337/dc18-S0021
- Bullard KM, Cowie CC, Lessem SE, Saydah SH, Menke A, Geiss LS, Orchard TJ, Rolka DB, Imperatore G. Prevalence of Diagnosed Diabetes in Adults by Diabetes Type - United States, 2016. MMWR Morb Mortal Wkly Rep 2018; **67**: 359-361 [PMID: 29596402 DOI: 10.15585/mmwr.mm6712a21
- 6 Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. Nature 2012; 486: 207-214 [PMID: 22699609 DOI: 10.1038/nature11234]
- Chattopadhyay A, Mythili S. The journey of gut microbiome an introduction and its influence on metabolic disorders. Front Biol 2018; 5: 327-341 [DOI: 10.1007/s11515-018-1490-6]
- Molina DK, DiMaio VJ. Normal organ weights in men: part I-the heart. Am J Forensic Med Pathol 2012; **33**: 362-367 [PMID: 22182983 DOI: 10.1097/PAF.0b013e31823d298b]
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Jian M, Zhou Y, Li Y, Zhang X, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J; MetaHIT Consortium, Bork P, Ehrlich SD, Wang J. A human gut microbial gene catalogue established by metagenomic sequencing. Nature 2010; 464: 59-65 [PMID: 20203603 DOI: 10.1038/nature08821]
- Conlon MA, Bird AR. The impact of diet and lifestyle on gut microbiota and human health. Nutrients 2014; 7: 17-44 [PMID: 25545101 DOI: 10.3390/nu7010017]
- He Y, Wu W, Zheng HM, Li P, McDonald D, Sheng HF, Chen MX, Chen ZH, Ji GY, Zheng ZD, Mujagond P, Chen XJ, Rong ZH, Chen P, Lyu LY, Wang X, Wu CB, Yu N, Xu YJ, Yin J, Raes J, Knight R, Ma WJ, Zhou HW. Regional variation limits applications of healthy gut microbiome reference ranges and disease models. Nat Med 2018; 24: 1532-1535 [PMID: 30150716 DOI: 10.1038/s41591-018-0164-x]
- 12 Maier L, Pruteanu M, Kuhn M, Zeller G, Telzerow A, Anderson EE, Brochado AR, Fernandez KC, Dose H, Mori H, Patil KR, Bork P, Typas A. Extensive impact of non-antibiotic drugs on human gut bacteria. Nature 2018; 555: 623-628 [PMID: 29555994 DOI: 10.1038/nature25979]
- Schroeder BO, Bäckhed F. Signals from the gut microbiota to distant organs in physiology and disease. Nat Med 2016; 22: 1079-1089 [PMID: 27711063 DOI: 10.1038/nm.4185]
- Barlow GM, Yu A, Mathur R. Role of the Gut Microbiome in Obesity and Diabetes Mellitus. Nutr Clin Pract 2015; **30**: 787-797 [PMID: 26452391 DOI: 10.1177/0884533615609896]
- Bouter KE, van Raalte DH, Groen AK, Nieuwdorp M. Role of the Gut Microbiome in the Pathogenesis of Obesity and Obesity-Related Metabolic Dysfunction. Gastroenterology 2017; 152: 1671-1678 [PMID: 28192102 DOI: 10.1053/j.gastro.2016.12.048]
- Chen X, Devaraj S. Gut Microbiome in Obesity, Metabolic Syndrome, and Diabetes. Curr Diab Rep 2018; 18: 129 [PMID: 30338410 DOI: 10.1007/s11892-018-1104-3]

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- Vallianou NG, Stratigou T, Tsagarakis S. Microbiome and diabetes: Where are we now? Diabetes Res Clin Pract 2018; 146: 111-118 [PMID: 30342053 DOI: 10.1016/j.diabres.2018.10.008]
- Purkait D, Hameed S, Fatima Z. Chapter 15 gut microbiome: Current development, challenges, and perspectives. In: Rastegari AA, Yadav AN, Yadav N. New and future developments in microbial biotechnology and bioengineering: Elsevier, 2020: 227-241
- Larsen N, Vogensen FK, van den Berg FW, Nielsen DS, Andreasen AS, Pedersen BK, Al-Soud WA, Sørensen SJ, Hansen LH, Jakobsen M. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. PLoS One 2010; 5: e9085 [PMID: 20140211 DOI: 10.1371/journal.pone.0009085]
- Thingholm LB, Rühlemann MC, Koch M, Fuqua B, Laucke G, Boehm R, Bang C, Franzosa EA, Hübenthal M, Rahnavard A, Frost F, Lloyd-Price J, Schirmer M, Lusis AJ, Vulpe CD, Lerch MM, Homuth G, Kacprowski T, Schmidt CO, Nöthlings U, Karlsen TH, Lieb W, Laudes M, Franke A, Huttenhower C. Obese Individuals with and without Type 2 Diabetes Show Different Gut Microbial Functional Capacity and Composition. Cell Host Microbe 2019; 26: 252-264.e10 [PMID: 31399369 DOI: 10.1016/j.chom.2019.07.004]
- Nowak K, Jabłońska E, Ratajczak-Wrona W. Immunomodulatory effects of synthetic endocrine disrupting chemicals on the development and functions of human immune cells. Environ Int 2019; 125: 350-364 [PMID: 30743143 DOI: 10.1016/j.envint.2019.01.078]
- Mrozinska S, Kapusta P, Gosiewski T, Sroka-Oleksiak A, Ludwig-Słomczyńska AH, Matejko B, Kiec-Wilk B, Bulanda M, Malecki MT, Wolkow PP, Klupa T. The Gut Microbiota Profile According to Glycemic Control in Type 1 Diabetes Patients Treated with Personal Insulin Pumps. Microorganisms 2021; 9 [PMID: 33445500 DOI: 10.3390/microorganisms9010155]
- 23 Huang Y, Li SC, Hu J, Ruan HB, Guo HM, Zhang HH, Wang X, Pei YF, Pan Y, Fang C. Gut microbiota profiling in Han Chinese with type 1 diabetes. Diabetes Res Clin Pract 2018; 141: 256-263 [PMID: 29733871 DOI: 10.1016/j.diabres.2018.04.032]
- Fassatoui M, Lopez-Siles M, Díaz-Rizzolo DA, Jmel H, Naouali C, Abdessalem G, Chikhaoui A, Nadal B, Jamoussi H, Abid A, Gomis R, Abdelhak S, Martinez-Medina M, Kefi R. Gut microbiota imbalances in Tunisian participants with type 1 and type 2 diabetes mellitus. Biosci Rep 2019; 39 [PMID: 31147456 DOI: 10.1042/BSR20182348]
- Elena RM, Gabriela GD, Arnulfo GC, Enrique CA. Studying the Gut Microbiome of Latin America and Hispanic/Latino Populations. Insight into Obesity and Diabetes: Systematic Review. Curr Diabetes Rev 2019; 15: 294-301 [PMID: 30058496 DOI: 10.2174/1573399814666180730124817]
- Walker AW, Ince J, Duncan SH, Webster LM, Holtrop G, Ze X, Brown D, Stares MD, Scott P, Bergerat A, Louis P, McIntosh F, Johnstone AM, Lobley GE, Parkhill J, Flint HJ. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. ISME J 2011; 5: 220-230 [PMID: 20686513 DOI: 10.1038/ismej.2010.118]
- Blandino G, Inturri R, Lazzara F, Di Rosa M, Malaguarnera L. Impact of gut microbiota on diabetes mellitus. Diabetes Metab 2016; 42: 303-315 [PMID: 27179626 DOI: 10.1016/j.diabet.2016.04.004]
- Chen PC, Chien YW, Yang SC. The alteration of gut microbiota in newly diagnosed type 2 diabetic patients. Nutrition 2019; 63-64: 51-56 [PMID: 30933725 DOI: 10.1016/j.nut.2018.11.019]
- Das T, Jayasudha R, Chakravarthy S, Prashanthi GS, Bhargava A, Tyagi M, Rani PK, Pappuru RR, Sharma S, Shivaji S. Alterations in the gut bacterial microbiome in people with type 2 diabetes mellitus and diabetic retinopathy. Sci Rep 2021; 11: 2738 [PMID: 33531650 DOI: 10.1038/s41598-021-82538-0]
- Gurung M, Li Z, You H, Rodrigues R, Jump DB, Morgun A, Shulzhenko N. Role of gut microbiota in type 2 diabetes pathophysiology. EBioMedicine 2020; 51: 102590 [PMID: 31901868 DOI: 10.1016/j.ebiom.2019.11.051]
- Sedighi M, Razavi S, Navab-Moghadam F, Khamseh ME, Alaei-Shahmiri F, Mehrtash A, Amirmozafari N. Comparison of gut microbiota in adult patients with type 2 diabetes and healthy individuals. *Microb Pathog* 2017; **111**: 362-369 [PMID: 28912092 DOI: 10.1016/j.micpath.2017.08.038]
- Nuli R, Cai J, Kadeer A, Zhang Y, Mohemaiti P. Integrative Analysis Toward Different Glucose Tolerance-Related Gut Microbiota and Diet. Front Endocrinol (Lausanne) 2019; 10: 295 [PMID: 31191448 DOI: 10.3389/fendo.2019.00295]
- 33 Cani PD, Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, Burcelin R. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. Diabetes 2008; 57: 1470-1481 [PMID: 18305141 DOI: 10.2337/db07-1403]
- 34 Païssé S, Valle C, Servant F, Courtney M, Burcelin R, Amar J, Lelouvier B. Comprehensive description of blood microbiome from healthy donors assessed by 16S targeted metagenomic sequencing. Transfusion 2016; 56: 1138-1147 [PMID: 26865079 DOI: 10.1111/trf.13477]
- Olde Loohuis LM, Mangul S, Ori APS, Jospin G, Koslicki D, Yang HT, Wu T, Boks MP, Lomen-Hoerth C, Wiedau-Pazos M, Cantor RM, de Vos WM, Kahn RS, Eskin E, Ophoff RA. Transcriptome analysis in whole blood reveals increased microbial diversity in schizophrenia. Transl Psychiatry 2018; 8: 96 [PMID: 29743478 DOI: 10.1038/s41398-018-0107-9]
- Whittle E, Leonard MO, Harrison R, Gant TW, Tonge DP. Multi-Method Characterization of the Human Circulating Microbiome. Front Microbiol 2018; 9: 3266 [PMID: 30705670 DOI: 10.3389/fmicb.2018.03266]
- Sato J, Kanazawa A, Ikeda F, Yoshihara T, Goto H, Abe H, Komiya K, Kawaguchi M, Shimizu T, Ogihara T, Tamura Y, Sakurai Y, Yamamoto R, Mita T, Fujitani Y, Fukuda H, Nomoto K,

- Takahashi T, Asahara T, Hirose T, Nagata S, Yamashiro Y, Watada H. Gut dysbiosis and detection of "live gut bacteria" in blood of Japanese patients with type 2 diabetes. Diabetes Care 2014; 37: 2343-2350 [PMID: 24824547 DOI: 10.2337/dc13-2817]
- 38 Qiu J, Zhou H, Jing Y, Dong C. Association between blood microbiome and type 2 diabetes mellitus: A nested case-control study. J Clin Lab Anal 2019; 33: e22842 [PMID: 30714640 DOI: 10.1002/jcla.22842]
- Evans JM, Morris LS, Marchesi JR. The gut microbiome: the role of a virtual organ in the endocrinology of the host. J Endocrinol 2013; 218: R37-R47 [PMID: 23833275 DOI: 10.1530/JOE-13-0131]
- 40 Lyte M. Probiotics function mechanistically as delivery vehicles for neuroactive compounds: Microbial endocrinology in the design and use of probiotics. Bioessays 2011; 33: 574-581 [PMID: 21732396 DOI: 10.1002/bies.201100024]
- 41 Hampl R, Stárka L. Endocrine disruptors and gut microbiome interactions. Physiol Res 2020; 69: S211-S223 [PMID: 33094620 DOI: 10.33549/physiolres.934513]
- 42 Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG. Minireview: Gut microbiota: the neglected endocrine organ. Mol Endocrinol 2014; 28: 1221-1238 [PMID: 24892638 DOI: 10.1210/me.2014-1108]
- Rosenfeld CS. Gut Dysbiosis in Animals Due to Environmental Chemical Exposures. Front Cell Infect Microbiol 2017; 7: 396 [PMID: 28936425 DOI: 10.3389/fcimb.2017.00396]
- Velmurugan G, Ramprasath T, Gilles M, Swaminathan K, Ramasamy S. Gut Microbiota, Endocrine-Disrupting Chemicals, and the Diabetes Epidemic. Trends Endocrinol Metab 2017; 28: 612-625 [PMID: 28571659 DOI: 10.1016/j.tem.2017.05.001]
- Needell JC, Zipris D. The Role of the Intestinal Microbiome in Type 1 Diabetes Pathogenesis. Curr Diab Rep 2016; **16**: 89 [PMID: 27523648 DOI: 10.1007/s11892-016-0781-z]
- Singer-Englar T, Barlow G, Mathur R. Obesity, diabetes, and the gut microbiome: an updated review. Expert Rev Gastroenterol Hepatol 2019; 13: 3-15 [PMID: 30791839 DOI: 10.1080/17474124.2019.1543023
- Yu F, Han W, Zhan G, Li S, Jiang X, Wang L, Xiang S, Zhu B, Yang L, Luo A, Hua F, Yang C. Abnormal gut microbiota composition contributes to the development of type 2 diabetes mellitus in db/db mice. Aging (Albany NY) 2019; 11: 10454-10467 [PMID: 31760385 DOI: 10.18632/aging.102469]
- 48 Halmos T, Suba I. [Physiological patterns of intestinal microbiota. The role of dysbacteriosis in obesity, insulin resistance, diabetes and metabolic syndrome]. Orv Hetil 2016; 157: 13-22 [PMID: 26708682 DOI: 10.1556/650.2015.30296]
- Fallucca F, Porrata C, Fallucca S, Pianesi M. Influence of diet on gut microbiota, inflammation and type 2 diabetes mellitus. First experience with macrobiotic Ma-Pi 2 diet. Diabetes Metab Res Rev 2014; **30** Suppl 1: 48-54 [PMID: 24532292 DOI: 10.1002/dmrr.2518]
- Aw W, Fukuda S. Understanding the role of the gut ecosystem in diabetes mellitus. J Diabetes Investig 2018; 9: 5-12 [PMID: 28390093 DOI: 10.1111/jdi.12673]
- Acosta-Montaño P, Rodríguez-Velázquez E, Ibarra-López E, Frayde-Gómez H, Mas-Oliva J, Delgado-Coello B, Rivero IA, Alatorre-Meda M, Aguilera J, Guevara-Olaya L, García-González V. Fatty Acid and Lipopolysaccharide Effect on Beta Cells Proteostasis and its Impact on Insulin Secretion. Cells 2019; 8 [PMID: 31412623 DOI: 10.3390/cells8080884]
- 52 Selber-Hnatiw S, Sultana T, Tse W, Abdollahi N, Abdullah S, Al Rahbani J, Alazar D, Alrumhein NJ, Aprikian S, Arshad R, Azuelos JD, Bernadotte D, Beswick N, Chazbey H, Church K, Ciubotaru E, D'Amato L, Del Corpo T, Deng J, Di Giulio BL, Diveeva D, Elahie E, Frank JGM, Furze E, Garner R, Gibbs V, Goldberg-Hall R, Goldman CJ, Goltsios FF, Gorjipour K, Grant T, Greco B, Guliyev N, Habrich A, Hyland H, Ibrahim N, Iozzo T, Jawaheer-Fenaoui A, Jaworski JJ, Jhajj MK, Jones J, Joyette R, Kaudeer S, Kelley S, Kiani S, Koayes M, Kpata AJAL, Maingot S, Martin S, Mathers K, McCullogh S, McNamara K, Mendonca J, Mohammad K, Momtaz SA, Navaratnarajah T, Nguyen-Duong K, Omran M, Ortiz A, Patel A, Paul-Cole K, Plaisir PA, Porras Marroquin JA, Prevost A, Quach A, Rafal AJ, Ramsarun R, Rhnima S, Rili L, Safir N, Samson E, Sandiford RR, Secondi S, Shahid S, Shahroozi M, Sidibé F, Smith M, Sreng Flores AM, Suarez Ybarra A, Sénéchal R, Taifour T, Tang L, Trapid A, Tremblay Potvin M, Wainberg J, Wang DN, Weissenberg M, White A, Wilkinson G, Williams B, Wilson JR, Zoppi J, Zouboulakis K, Gamberi C. Metabolic networks of the human gut microbiota. Microbiology (Reading) 2020; 166: 96-119 [PMID: 31799915 DOI: 10.1099/mic.0.000853]
- Gordon JI, Hooper LV, McNevin MS, Wong M, Bry L. Epithelial cell growth and differentiation. III. Promoting diversity in the intestine: conversations between the microflora, epithelium, and diffuse GALT. Am J Physiol 1997; 273: G565-G570 [PMID: 9316460 DOI: 10.1152/aipgi.1997.273.3.G5651
- 54 Karlsson F, Tremaroli V, Nielsen J, Bäckhed F. Assessing the human gut microbiota in metabolic diseases. Diabetes 2013; 62: 3341-3349 [PMID: 24065795 DOI: 10.2337/db13-0844]
- Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. Nature 2012; 489: 242-249 [PMID: 22972297 DOI: 10.1038/nature11552]
- Ussar S, Griffin NW, Bezy O, Fujisaka S, Vienberg S, Softic S, Deng L, Bry L, Gordon JI, Kahn CR. Interactions between Gut Microbiota, Host Genetics and Diet Modulate the Predisposition to Obesity and Metabolic Syndrome. Cell Metab 2015; 22: 516-530 [PMID: 26299453 DOI: 10.1016/j.cmet.2015.07.007]

- 57 **El Aidy S**, Hooiveld G, Tremaroli V, Bäckhed F, Kleerebezem M. The gut microbiota and mucosal homeostasis: colonized at birth or at adulthood, does it matter? *Gut Microbes* 2013; **4**: 118-124 [PMID: 23333858 DOI: 10.4161/gmic.23362]
- den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res* 2013; 54: 2325-2340 [PMID: 23821742 DOI: 10.1194/jlr.R036012]
- 59 Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol* 2015; 11: 577-591 [PMID: 26260141 DOI: 10.1038/nrendo.2015.128]
- 60 Lin HV, Frassetto A, Kowalik EJ Jr, Nawrocki AR, Lu MM, Kosinski JR, Hubert JA, Szeto D, Yao X, Forrest G, Marsh DJ. Butyrate and propionate protect against diet-induced obesity and regulate gut hormones via free fatty acid receptor 3-independent mechanisms. PLoS One 2012; 7: e35240 [PMID: 22506074 DOI: 10.1371/journal.pone.0035240]
- 61 Gao Z, Yin J, Zhang J, Ward RE, Martin RJ, Lefevre M, Cefalu WT, Ye J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* 2009; 58: 1509-1517 [PMID: 19366864 DOI: 10.2337/db08-1637]
- 62 Rácz B, Dušková M, Stárka L, Hainer V, Kunešová M. Links between the circadian rhythm, obesity and the microbiome. *Physiol Res* 2018; 67: S409-S420 [PMID: 30484668 DOI: 10.33549/physiolres.934020]
- 63 Koliada A, Syzenko G, Moseiko V, Budovska L, Puchkov K, Perederiy V, Gavalko Y, Dorofeyev A, Romanenko M, Tkach S, Sineok L, Lushchak O, Vaiserman A. Association between body mass index and Firmicutes/Bacteroidetes ratio in an adult Ukrainian population. *BMC Microbiol* 2017; 17: 120 [PMID: 28532414 DOI: 10.1186/s12866-017-1027-1]
- 64 Hung WC, Hung WW, Tsai HJ, Chang CC, Chiu YW, Hwang SJ, Kuo MC, Chen SC, Dai CY, Tsai YC. The Association of Targeted Gut Microbiota with Body Composition in Type 2 Diabetes Mellitus. Int J Med Sci 2021; 18: 511-519 [PMID: 33390820 DOI: 10.7150/ijms.51164]
- Meijnikman AS, Gerdes VE, Nieuwdorp M, Herrema H. Evaluating Causality of Gut Microbiota in Obesity and Diabetes in Humans. *Endocr Rev* 2018; 39: 133-153 [PMID: 29309555 DOI: 10.1210/er.2017-00192]
- 66 Leustean AM, Ciocoiu M, Sava A, Costea CF, Floria M, Tarniceriu CC, Tanase DM. Implications of the Intestinal Microbiota in Diagnosing the Progression of Diabetes and the Presence of Cardiovascular Complications. *J Diabetes Res* 2018; 2018: 5205126 [PMID: 30539026 DOI: 10.1155/2018/5205126]
- 67 Zhao S, Jang S, Lee YK, Kim DG, Jin Z, Koh HJ. Genetic Basis of Tiller Dynamics of Rice Revealed by Genome-Wide Association Studies. *Plants (Basel)* 2020; 9 [PMID: 33276582 DOI: 10.3390/nu12123719]
- 68 Preguiça I, Alves A, Nunes S, Gomes P, Fernandes R, Viana SD, Reis F. Diet-Induced Rodent Models of Diabetic Peripheral Neuropathy, Retinopathy and Nephropathy. *Nutrients* 2020; 12 [PMID: 31963709 DOI: 10.3390/nu12010250]
- 69 Ham B, Hwang HB, Jung SH, Chang S, Kang KD, Kwon MJ. Distribution and Diversity of Ocular Microbial Communities in Diabetic Patients Compared with Healthy Subjects. *Curr Eye Res* 2018; 43: 314-324 [PMID: 29172724 DOI: 10.1080/02713683.2017.1406528]
- 70 Rowan S, Taylor A. The Role of Microbiota in Retinal Disease. Adv Exp Med Biol 2018; 1074: 429-435 [PMID: 29721973 DOI: 10.1007/978-3-319-75402-4 53]
- 71 **Baim AD**, Movahedan A, Farooq AV, Skondra D. The microbiome and ophthalmic disease. *Exp Biol Med (Maywood)* 2019; **244**: 419-429 [PMID: 30463439 DOI: 10.1177/1535370218813616]
- 72 **Floyd JL**, Grant MB. The Gut-Eye Axis: Lessons Learned from Murine Models. *Ophthalmol Ther* 2020; **9**: 499-513 [PMID: 32617914 DOI: 10.1007/s40123-020-00278-2]
- 73 Beli E, Yan Y, Moldovan L, Vieira CP, Gao R, Duan Y, Prasad R, Bhatwadekar A, White FA, Townsend SD, Chan L, Ryan CN, Morton D, Moldovan EG, Chu FI, Oudit GY, Derendorf H, Adorini L, Wang XX, Evans-Molina C, Mirmira RG, Boulton ME, Yoder MC, Li Q, Levi M, Busik JV, Grant MB. Restructuring of the Gut Microbiome by Intermittent Fasting Prevents Retinopathy and Prolongs Survival in db/db Mice. Diabetes 2018; 67: 1867-1879 [PMID: 29712667 DOI: 10.2337/db18-0158]
- 74 St Leger AJ, Caspi RR. Visions of Eye Commensals: The Known and the Unknown About How the Microbiome Affects Eye Disease. *Bioessays* 2018; 40: e1800046 [PMID: 30289987 DOI: 10.1002/bies.201800046]
- 75 Grasset E, Burcelin R. The gut microbiota to the brain axis in the metabolic control. Rev Endocr Metab Disord 2019; 20: 427-438 [PMID: 31656993 DOI: 10.1007/s11154-019-09511-1]
- 76 Barrios C, Beaumont M, Pallister T, Villar J, Goodrich JK, Clark A, Pascual J, Ley RE, Spector TD, Bell JT, Menni C. Gut-Microbiota-Metabolite Axis in Early Renal Function Decline. *PLoS One* 2015; 10: e0134311 [PMID: 26241311 DOI: 10.1371/journal.pone.0134311]
- 77 Wong J, Piceno YM, DeSantis TZ, Pahl M, Andersen GL, Vaziri ND. Expansion of urease- and uricase-containing, indole- and p-cresol-forming and contraction of short-chain fatty acid-producing intestinal microbiota in ESRD. Am J Nephrol 2014; 39: 230-237 [PMID: 24643131 DOI: 10.1159/000360010]
- Stavropoulou E, Kantartzi K, Tsigalou C, Konstantinidis T, Romanidou G, Voidarou C, Bezirtzoglou E. Focus on the Gut-Kidney Axis in Health and Disease. Front Med (Lausanne) 2020; 7: 620102 [PMID: 33553216 DOI: 10.3389/fmed.2020.620102]

- Magouliotis DE, Tasiopoulou VS, Sioka E, Chatedaki C, Zacharoulis D. Impact of Bariatric Surgery on Metabolic and Gut Microbiota Profile: a Systematic Review and Meta-analysis. Obes Surg 2017; 27: 1345-1357 [PMID: 28265960 DOI: 10.1007/s11695-017-2595-8]
- 80 Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JF, Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druesne A, Van Hylckama Vlieg JE, Bloks VW, Groen AK, Heilig HG, Zoetendal EG, Stroes ES, de Vos WM, Hoekstra JB, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. Gastroenterology 2012; 143: 913-6.e7 [PMID: 22728514 DOI: 10.1053/j.gastro.2012.06.031]
- Candela M, Biagi E, Soverini M, Consolandi C, Quercia S, Severgnini M, Peano C, Turroni S, Rampelli S, Pozzilli P, Pianesi M, Fallucca F, Brigidi P. Modulation of gut microbiota dysbioses in type 2 diabetic patients by macrobiotic Ma-Pi 2 diet. Br J Nutr 2016; 116: 80-93 [PMID: 27151248 DOI: 10.1017/S0007114516001045]
- Quddus MR, Lawrence WD. A Seminal Observation! Int J Surg Pathol 2016; 24: 47 [PMID: 26338717 DOI: 10.1177/1066896915604025]
- 83 Krezalek MA, Yeh A, Alverdy JC, Morowitz M. Influence of nutrition therapy on the intestinal microbiome. Curr Opin Clin Nutr Metab Care 2017; 20: 131-137 [PMID: 27997410 DOI: 10.1097/MCO.00000000000003481
- Suez J, Korem T, Zeevi D, Zilberman-Schapira G, Thaiss CA, Maza O, Israeli D, Zmora N, Gilad S, Weinberger A, Kuperman Y, Harmelin A, Kolodkin-Gal I, Shapiro H, Halpern Z, Segal E, Elinav E. Artificial sweeteners induce glucose intolerance by altering the gut microbiota. Nature 2014; 514: 181-186 [PMID: 25231862 DOI: 10.1038/nature13793]
- 85 Chi L, Bian X, Gao B, Tu P, Lai Y, Ru H, Lu K. Effects of the Artificial Sweetener Neotame on the Gut Microbiome and Fecal Metabolites in Mice. Molecules 2018; 23 [PMID: 29425148 DOI: 10.3390/molecules23020367]
- Frankenfeld CL, Sikaroodi M, Lamb E, Shoemaker S, Gillevet PM. High-intensity sweetener consumption and gut microbiome content and predicted gene function in a cross-sectional study of adults in the United States. Ann Epidemiol 2015; 25: 736-42.e4 [PMID: 26272781 DOI: 10.1016/j.annepidem.2015.06.083]
- 87 Ahmad SY, Friel J, Mackay D. The Effects of Non-Nutritive Artificial Sweeteners, Aspartame and Sucralose, on the Gut Microbiome in Healthy Adults: Secondary Outcomes of a Randomized Double-Blinded Crossover Clinical Trial. Nutrients 2020; 12 [PMID: 33171964 DOI: 10.3390/nu12113408]
- Whang A, Nagpal R, Yadav H. Bi-directional drug-microbiome interactions of anti-diabetics. EBioMedicine 2019; 39: 591-602 [PMID: 30553752 DOI: 10.1016/j.ebiom.2018.11.046]
- Wu H, Esteve E, Tremaroli V, Khan MT, Caesar R, Mannerås-Holm L, Ståhlman M, Olsson LM, Serino M, Planas-Fèlix M, Xifra G, Mercader JM, Torrents D, Burcelin R, Ricart W, Perkins R, Fernandez-Real JM, Bäckhed F. Metformin alters the gut microbiome of individuals with treatmentnaive type 2 diabetes, contributing to the therapeutic effects of the drug. Nat Med 2017; 23: 850-858 [PMID: 28530702 DOI: 10.1038/nm.4345]
- Zhang F, Wang M, Yang J, Xu Q, Liang C, Chen B, Zhang J, Yang Y, Wang H, Shang Y, Wang Y, Mu X, Zhu D, Zhang C, Yao M, Zhang L. Response of gut microbiota in type 2 diabetes to hypoglycemic agents. *Endocrine* 2019; **66**: 485-493 [PMID: 31410749 DOI: $10.1007/s12020\hbox{-}019\hbox{-}02041\hbox{-}5]$
- Buse JB, DeFronzo RA, Rosenstock J, Kim T, Burns C, Skare S, Baron A, Fineman M. The Primary Glucose-Lowering Effect of Metformin Resides in the Gut, Not the Circulation: Results From Shortterm Pharmacokinetic and 12-Week Dose-Ranging Studies. Diabetes Care 2016; 39: 198-205 [PMID: 26285584 DOI: 10.2337/dc15-0488]
- 92 Rizkallah MR, Saad R, Aziz RK. The human microbiome project, personalized medicine and the birth of pharmacomicrobiomics. Curr Pharmacogenomics Person Med 2010; 8: 182-193 [DOI: 10.2174/187569210792246326]
- Reimer RA, Grover GJ, Koetzner L, Gahler RJ, Lyon MR, Wood S. Combining sitagliptin/metformin with a functional fiber delays diabetes progression in Zucker rats. J Endocrinol 2014; 220: 361-373 [PMID: 24389593 DOI: 10.1530/JOE-13-0484]
- Rodrigues RR, Greer RL, Dong X, DSouza KN, Gurung M, Wu JY, Morgun A, Shulzhenko N. Antibiotic-Induced Alterations in Gut Microbiota Are Associated with Changes in Glucose Metabolism in Healthy Mice. Front Microbiol 2017; 8: 2306 [PMID: 29213261 DOI: 10.3389/fmicb.2017.023061
- 95 Stenman LK, Waget A, Garret C, Briand F, Burcelin R, Sulpice T, Lahtinen S. Probiotic B420 and prebiotic polydextrose improve efficacy of antidiabetic drugs in mice. Diabetol Metab Syndr 2015; 7: 75 [PMID: 26366205 DOI: 10.1186/s13098-015-0075-7]
- 96 Zheng J, Li H, Zhang X, Jiang M, Luo C, Lu Z, Xu Z, Shi J. Prebiotic Mannan-Oligosaccharides Augment the Hypoglycemic Effects of Metformin in Correlation with Modulating Gut Microbiota. J Agric Food Chem 2018; 66: 5821-5831 [PMID: 29701959 DOI: 10.1021/acs.jafc.8b00829]
- Yang R, Jia Q, Mehmood S, Ma S, Liu X. Genistein ameliorates inflammation and insulin resistance through mediation of gut microbiota composition in type 2 diabetic mice. Eur J Nutr 2021; 60: 2155-2168 [PMID: 33068158 DOI: 10.1007/s00394-020-02403-0]
- Dagdeviren S, Jung DY, Friedline RH, Noh HL, Kim JH, Patel PR, Tsitsilianos N, Inashima K, Tran DA, Hu X, Loubato MM, Craige SM, Kwon JY, Lee KW, Kim JK. IL-10 prevents aging-

1476

- associated inflammation and insulin resistance in skeletal muscle. FASEB J 2017; 31: 701-710 [PMID: 27811060 DOI: 10.1096/fj.201600832R]
- Andreasen AS, Larsen N, Pedersen-Skovsgaard T, Berg RM, Møller K, Svendsen KD, Jakobsen M, Pedersen BK. Effects of Lactobacillus acidophilus NCFM on insulin sensitivity and the systemic inflammatory response in human subjects. Br J Nutr 2010; 104: 1831-1838 [PMID: 20815975 DOI: 10.1017/S0007114510002874]
- 100 Wang X, Ota N, Manzanillo P, Kates L, Zavala-Solorio J, Eidenschenk C, Zhang J, Lesch J, Lee WP, Ross J, Diehl L, van Bruggen N, Kolumam G, Ouyang W. Interleukin-22 alleviates metabolic disorders and restores mucosal immunity in diabetes. Nature 2014; 514: 237-241 [PMID: 25119041 DOI: 10.1038/nature13564]
- Panwar H, Calderwood D, Grant IR, Grover S, Green BD. Lactobacillus strains isolated from infant faeces possess potent inhibitory activity against intestinal alpha- and beta-glucosidases suggesting anti-diabetic potential. Eur J Nutr 2014; 53: 1465-1474 [PMID: 24414142 DOI: 10.1007/s00394-013-0649-9]
- Napolitano A, Miller S, Nicholls AW, Baker D, Van Horn S, Thomas E, Rajpal D, Spivak A, Brown JR, Nunez DJ. Novel gut-based pharmacology of metformin in patients with type 2 diabetes mellitus. PLoS One 2014; 9: e100778 [PMID: 24988476 DOI: 10.1371/journal.pone.0100778]
- Ahmadi S, Nagpal R, Wang S, Gagliano J, Kitzman DW, Soleimanian-Zad S, Sheikh-Zeinoddin M, Read R, Yadav H. Prebiotics from acorn and sago prevent high-fat-diet-induced insulin resistance via microbiome-gut-brain axis modulation. J Nutr Biochem 2019; 67: 1-13 [PMID: 30831458 DOI: 10.1016/i.inutbio.2019.01.0111
- 104 Zhang Y, Gu Y, Ren H, Wang S, Zhong H, Zhao X, Ma J, Gu X, Xue Y, Huang S, Yang J, Chen L, Chen G, Qu S, Liang J, Qin L, Huang Q, Peng Y, Li Q, Wang X, Kong P, Hou G, Gao M, Shi Z, Li X, Qiu Y, Zou Y, Yang H, Wang J, Xu G, Lai S, Li J, Ning G, Wang W. Gut microbiome-related effects of berberine and probiotics on type 2 diabetes (the PREMOTE study). Nat Commun 2020; 11: 5015 [PMID: 33024120 DOI: 10.1038/s41467-020-18414-8]
- Adeshirlarijaney A, Gewirtz AT. Considering gut microbiota in treatment of type 2 diabetes mellitus. Gut Microbes 2020; 11: 253-264 [PMID: 32005089 DOI: 10.1080/19490976.2020.1717719]
- Jafarnejad S, Saremi S, Jafarnejad F, Arab A. Effects of a Multispecies Probiotic Mixture on Glycemic Control and Inflammatory Status in Women with Gestational Diabetes: A Randomized Controlled Clinical Trial. J Nutr Metab 2016; 2016: 5190846 [PMID: 27429803 DOI: 10.1155/2016/5190846]
- Asemi Z, Khorrami-Rad A, Alizadeh SA, Shakeri H, Esmaillzadeh A. Effects of synbiotic food consumption on metabolic status of diabetic patients: a double-blind randomized cross-over controlled clinical trial. Clin Nutr 2014; 33: 198-203 [PMID: 23786900 DOI: 10.1016/j.clnu.2013.05.015]
- Tonucci LB. Olbrich Dos Santos KM, Licursi de Oliveira L, Rocha Ribeiro SM, Duarte Martino HS. Clinical application of probiotics in type 2 diabetes mellitus: A randomized, double-blind, placebo-controlled study. Clin Nutr 2017; 36: 85-92 [PMID: 26732026 DOI: 10.1016/j.clnu.2015.11.011]
- Zhang Q, Wu Y, Fei X. Effect of probiotics on glucose metabolism in patients with type 2 diabetes mellitus: A meta-analysis of randomized controlled trials. Medicina (Kaunas) 2016; 52: 28-34 [PMID: 26987497 DOI: 10.1016/j.medici.2015.11.008]
- Yao K, Zeng L, He Q, Wang W, Lei J, Zou X. Effect of Probiotics on Glucose and Lipid Metabolism in Type 2 Diabetes Mellitus: A Meta-Analysis of 12 Randomized Controlled Trials. Med Sci Monit 2017; 23: 3044-3053 [PMID: 28638006 DOI: 10.12659/msm.902600]
- Kasińska MA, Drzewoski J. Effectiveness of probiotics in type 2 diabetes: a meta-analysis. Pol Arch Med Wewn 2015; 125: 803-813 [PMID: 26431318 DOI: 10.20452/pamw.3156]
- Kocsis T, Molnár B, Németh D, Hegyi P, Szakács Z, Bálint A, Garami A, Soós A, Márta K, Solymár M. Probiotics have beneficial metabolic effects in patients with type 2 diabetes mellitus: a meta-analysis of randomized clinical trials. Sci Rep 2020; 10: 11787 [PMID: 32678128 DOI: 10.1038/s41598-020-68440-1]
- Tao YW, Gu YL, Mao XQ, Zhang L, Pei YF. Correction to: Effects of probiotics on type II diabetes mellitus: a meta-analysis. J Transl Med 2020; 18: 105 [PMID: 32111256 DOI: 10.1186/s12967-020-02274-31
- 114 Tao YW, Gu YL, Mao XQ, Zhang L, Pei YF. Effects of probiotics on type II diabetes mellitus: a meta-analysis. J Transl Med 2020; 18: 30 [PMID: 31952517 DOI: 10.1186/s12967-020-02213-2]
- 115 Bock PM, Telo GH, Ramalho R, Sbaraini M, Leivas G, Martins AF, Schaan BD. The effect of probiotics, prebiotics or synbiotics on metabolic outcomes in individuals with diabetes: a systematic review and meta-analysis. Diabetologia 2021; 64: 26-41 [PMID: 33047170 DOI: 10.1007/s00125-020-05295-1]
- Ho J, Reimer RA, Doulla M, Huang C. Effect of prebiotic intake on gut microbiota, intestinal permeability and glycemic control in children with type 1 diabetes: study protocol for a randomized controlled trial. Trials 2016; 17: 347 [PMID: 27456494 DOI: 10.1186/s13063-016-1486-y]
- Mishra SP, Wang S, Nagpal R, Miller B, Singh R, Taraphder S, Yadav H. Probiotics and Prebiotics for the Amelioration of Type 1 Diabetes: Present and Future Perspectives. Microorganisms 2019; 7 [PMID: 30832381 DOI: 10.3390/microorganisms7030067]
- 118 Didari T, Solki S, Mozaffari S, Nikfar S, Abdollahi M. A systematic review of the safety of

- probiotics. Expert Opin Drug Saf 2014; 13: 227-239 [PMID: 24405164 DOI: 10.1517/14740338.2014.872627]
- Matsuzaki T, Nagata Y, Kado S, Uchida K, Kato I, Hashimoto S, Yokokura T. Prevention of onset in an insulin-dependent diabetes mellitus model, NOD mice, by oral feeding of Lactobacillus casei. APMIS 1997; 105: 643-649 [PMID: 9298104 DOI: 10.1111/j.1699-0463.1997.tb05066.x]
- Xie Y, Xiao M, Ni Y, Jiang S, Feng G, Sang S, Du G. Alpinia oxyphylla Miq. Extract Prevents Diabetes in Mice by Modulating Gut Microbiota. J Diabetes Res 2018; 2018: 4230590 [PMID: 29967794 DOI: 10.1155/2018/4230590]
- Marcial GE, Ford AL, Haller MJ, Gezan SA, Harrison NA, Cai D, Meyer JL, Perry DJ, Atkinson MA, Wasserfall CH, Garrett T, Gonzalez CF, Brusko TM, Dahl WJ, Lorca GL. Lactobacillus johnsonii N6.2 Modulates the Host Immune Responses: A Double-Blind, Randomized Trial in Healthy Adults. Front Immunol 2017; 8: 655 [PMID: 28659913 DOI: 10.3389/fimmu.2017.00655]
- de Goffau MC, Luopajärvi K, Knip M, Ilonen J, Ruohtula T, Härkönen T, Orivuori L, Hakala S, Welling GW, Harmsen HJ, Vaarala O. Fecal microbiota composition differs between children with β -cell autoimmunity and those without. Diabetes 2013; 62: 1238-1244 [PMID: 23274889 DOI: 10.2337/db12-05261
- 123 Tai N, Peng J, Liu F, Gulden E, Hu Y, Zhang X, Chen L, Wong FS, Wen L. Microbial antigen mimics activate diabetogenic CD8 T cells in NOD mice. J Exp Med 2016; 213: 2129-2146 [PMID: 27621416 DOI: 10.1084/jem.20160526]
- Giongo A, Gano KA, Crabb DB, Mukherjee N, Novelo LL, Casella G, Drew JC, Ilonen J, Knip M, Hyöty H, Veijola R, Simell T, Simell O, Neu J, Wasserfall CH, Schatz D, Atkinson MA, Triplett EW. Toward defining the autoimmune microbiome for type 1 diabetes. ISME J 2011; 5: 82-91 [PMID: 20613793 DOI: 10.1038/ismej.2010.92]
- de Goffau MC, Fuentes S, van den Bogert B, Honkanen H, de Vos WM, Welling GW, Hyöty H, Harmsen HJ. Aberrant gut microbiota composition at the onset of type 1 diabetes in young children. Diabetologia 2014; 57: 1569-1577 [PMID: 24930037 DOI: 10.1007/s00125-014-3274-0]
- Brown CT, Davis-Richardson AG, Giongo A, Gano KA, Crabb DB, Mukherjee N, Casella G, Drew JC, Ilonen J, Knip M, Hyöty H, Veijola R, Simell T, Simell O, Neu J, Wasserfall CH, Schatz D, Atkinson MA, Triplett EW. Gut microbiome metagenomics analysis suggests a functional model for the development of autoimmunity for type 1 diabetes. PLoS One 2011; 6: e25792 [PMID: 22043294 DOI: 10.1371/journal.pone.0025792]
- 127 Li Q, Chang Y, Zhang K, Chen H, Tao S, Zhang Z. Implication of the gut microbiome composition of type 2 diabetic patients from northern China. Sci Rep 2020; 10: 5450 [PMID: 32214153 DOI: 10.1038/s41598-020-62224-3]
- Kikuchi R, Irie J, Yamada-Goto N, Kikkawa E, Seki Y, Kasama K, Itoh H. The Impact of Laparoscopic Sleeve Gastrectomy with Duodenojejunal Bypass on Intestinal Microbiota Differs from that of Laparoscopic Sleeve Gastrectomy in Japanese Patients with Obesity. Clin Drug Investig 2018; **38**: 545-552 [PMID: 29508311 DOI: 10.1007/s40261-018-0638-0]
- Li JF, Lai DD, Ni B, Sun KX. Comparison of laparoscopic Roux-en-Y gastric bypass with laparoscopic sleeve gastrectomy for morbid obesity or type 2 diabetes mellitus: a meta-analysis of randomized controlled trials. Can J Surg 2013; 56: E158-E164 [PMID: 24284156 DOI: 10.1503/cjs.026912]



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