

Randomized Controlled Trial

Changes in urinary excretion of water and sodium transporters during amiloride and bendroflumethiazide treatment

Janni M Jensen, Frank H Mose, Anna-Ewa O Kulik, Jesper N Bech, Robert A Fenton, Erling B Pedersen

Janni M Jensen, Frank H Mose, Anna-Ewa O Kulik, Jesper N Bech, Erling B Pedersen, University Clinic in Nephrology and Hypertension, Department of Medical Research, Holstebro Hospital, Regional Hospital Jutland West and Aarhus University, 7500 Holstebro, Denmark

Robert A Fenton, Department of Biomedicine, Aarhus University, 8000 Aarhus, Denmark

Author contributions: All authors had contributed to the manuscript; Jensen JM, Mose FH and Pedersen EB designed the project; Jensen JM, Mose FH and Kulik AEO performed the experiments and statistical analyzes; Fenton RA performed the NKCC2 antibody characterization; Jensen JM, Mose FH, Bech JN, Fenton RA and Pedersen EB wrote and edited the manuscript; all authors read and approved the final manuscript.

Supported by Grants from The Lundbeck Foundation, Aase and Ejnar Danielsens Foundation, Helen and Ejnar Bjoernows Foundation and Region Midjuttlands Research Fund.

Ethics approval statement: This study was review and approved by the Regional Committees on Health Research Ethics, Skottenborg 26, 8600 Viborg, Denmark (j.no 1-10-72-178-12).

Clinical trial registration: This study is registered at clinical trials. The registration identification number is: NCT 01635231 <http://clinicaltrials.gov/ct2/show/NCT01635231>.

Informed consent statement: All study participants provided informed written consent prior to study enrolment.

Conflict-of-interest statement: The authors declare that they have no competing interests.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on

different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Janni M Jensen, PhD, University Clinic in Nephrology and Hypertension, Department of Medical Research, Holstebro Hospital, Regional Hospital Jutland West and Aarhus University, Laegaardvej 12, 7500 Holstebro, Denmark. jannimaj@gmail.com
 Telephone: +45-7843-6588
 Fax: +45-7843-6582

Received: November 13, 2014

Peer-review started: November 18, 2014

First decision: February 7, 2015

Revised: March 8, 2015

Accepted: April 28, 2015

Article in press: April 30, 2015

Published online: July 6, 2015

Abstract

AIM: To quantify changes in urinary excretion of aquaporin2 water channels (u-AQP2), the sodium-potassium-chloride co-transporter (u-NKCC2) and the epithelial sodium channels (u-ENaC) during treatment with bendroflumethiazide (BFTZ), amiloride and placebo.

METHODS: In a randomized, double-blinded, placebo-controlled, 3-way crossover study we examined 23 healthy subjects on a standardized diet and fluid intake. The subjects were treated with amiloride 5 mg, BFTZ 1.25 mg or placebo twice a day for 4.5 d before each examination day. On the examination day, glomerular filtration rate was measured by the constant infusion clearance technique with ⁵¹Cr-EDTA as reference substance. To estimate the changes in water transport *via* AQP2 and sodium transport *via* NKCC2 and ENaC, u-NKCC2, the gamma fraction of ENaC (u-ENaC_γ), and

u-AQP2 were measured at baseline and after infusion with 3% hypertonic saline. u-NKCC2, u-ENaC γ , u-AQP2 and plasma concentrations of vasopressin (p-AVP), renin (PRC), angiotensin II (p-ANG II) and aldosterone (p-Aldo) were measured, by radioimmunoassay. Central blood pressure was estimated by applanation tonometry and body fluid volumes were estimated by bio-impedance spectroscopy. General linear model with repeated measures or related samples Friedman's two-way analysis was used to compare differences. Post hoc Bonferroni correction was used for multiple comparisons of post infusion periods to baseline within each treatment group.

RESULTS: At baseline there were no differences in u-NKCC2, u-ENaC γ and u-AQP2. PRC, p-Ang II and p-Aldo were increased during active treatments ($P < 0.001$). After hypertonic saline, u-NKCC2 increased during amiloride ($6\% \pm 34\%$; $P = 0.081$) and increased significantly during placebo ($17\% \pm 24\%$; $P = 0.010$). U-AQP2 increased significantly during amiloride ($31\% \pm 22\%$; $P < 0.001$) and placebo ($34\% \pm 27\%$; $P < 0.001$), while u-NKCC2 and u-AQP2 did not change significantly during BFTZ ($-7\% \pm 28\%$; $P = 0.257$ and $5\% \pm 16\%$; $P = 0.261$). U-ENaC γ increased in all three groups ($P < 0.050$). PRC, Ang II and p-Aldo decreased to the same extent, while AVP increased, but to a smaller degree during BFTZ ($P = 0.048$). cDBP decreased significantly during BFTZ ($P < 0.001$), but not during amiloride or placebo. There were no significant differences in body fluid volumes.

CONCLUSION: After hypertonic saline, u-NKCC2 and u-AQP2 increased during amiloride, but not during BFTZ. Lower p-AVP during BFTZ potentially caused less stimulation of NKCC2 and AQP2 and subsequent lower reabsorption of water and sodium.

Key words: Amiloride; Thiazide; Sodium-potassium-chloride co-transporter; Aquaporin2; Epithelial sodium channels; Sodium; Water; Sodium transporters; Hypertonic saline; Urine

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Measurements of urinary sodium-potassium chloride co-transporter (NKCC2), epithelial sodium channel (ENaC) and aquaporin2 (AQP2) can be used as biomarkers of water- and sodium transport in the nephron. However, it has never been studied to what extent the function of NKCC2, ENaC and AQP2 is simultaneously affected in response to diuretics. The present study showed that infusion of 3% saline increased u-NKCC2 and u-AQP2 during amiloride and placebo, while u-NKCC2 and u-AQP2 remained unchanged during bendroflumethiazide. Therefore, in contrast to amiloride, bendroflumethiazide caused the absence of a compensatory reabsorption of sodium *via* NKCC2 and water *via* AQP2.

Pedersen EB. Changes in urinary excretion of water and sodium transporters during amiloride and bendroflumethiazide treatment. *World J Nephrol* 2015; 4(3): 423-437 Available from: URL: <http://www.wjgnet.com/2220-6124/full/v4/i3/423.htm> DOI: <http://dx.doi.org/10.5527/wjn.v4.i3.423>

INTRODUCTION

During normal conditions, approximately 60% of filtered sodium is absorbed in the proximal tubules and 30% of sodium is absorbed in the kidneys *via* the sodium-potassium chloride co-transporter (NKCC2) in the thick ascending limb of Henle's loop (TAL). The distal convoluted tubules are responsible for 5%-10% of sodium reabsorption *via* the sodium chloride co-transporter (NCC)^[1]. Thiazides inhibit NCC in distal tubules and decrease sodium reabsorption^[2]. In the collecting duct the epithelial sodium channel (ENaC) is responsible for the reabsorption of 3%-5% of filtered sodium^[1]. Amiloride is a potassium sparing selective inhibitor of ENaC channels^[3]. Water is predominantly reabsorbed in the proximal tubules and thin descending limb of Henle's loop^[1]. In the collecting ducts water absorption depends on passive transport *via* AQP2 water channels and is regulated by vasopressin (AVP)^[4]. AQP2 can be excreted into urine^[5,6], and may be used as a biomarker of collecting duct water transport^[7-9]. Similarly, urinary excretion of beta ENaC correlates with changes in urinary sodium excretion^[10]. Recently, our group documented changes in transport of water *via* AQP2 and sodium *via* ENaC in healthy subjects after infusion of isotonic glucose or hypertonic and isotonic saline, by measurements of urinary excretion of AQP2 (u-AQP2) and gamma ENaC (u-ENaC γ)^[11] and abnormal urinary excretion of NKCC2 (u-NKCC2) and u-AQP2 in patients with chronic kidney disease^[12].

In the present randomised, placebo-controlled study present study in healthy young subjects, we hypothesize that excretion of NKCC2 will not be effected but compensatory increases in distal transporter activity will occur during thiazide treatment but not during amiloride. This is compared to placebo both at baseline and in response to a saline load. Therefore, the aim was to quantify changes in urinary excretion of NKCC2, u-ENaC γ and u-AQP2 as estimates of tubular water and sodium handling at baseline conditions and after 3% saline infusion, during treatment with bendroflumethiazide (BFTZ), amiloride and placebo. In addition, changes in renal tubular function, vasoactive hormones, body fluid volumes and blood pressure were measured. The novelty of this study is due to; measurements of u-NKCC2 and the interplay with ENaC, AQP2 and the regulating mechanisms involved in water and sodium homeostasis, while simultaneously antagonizing NCC with BFTZ and ENaC with amiloride. Quantification of sodium- and water channel proteins in urine during different conditions may provide important information of the mechanisms involved in water and

sodium balance in the nephron.

MATERIALS AND METHODS

Design

The trial was conducted as a randomized, double-blinded, placebo-controlled, 3-way crossover study. Subjects were randomized to tablet BFTZ, amiloride or placebo for 4.5 d. Each treatment period was followed by an examination day, separated by at least 3 wk.

Participants

Eligible participants were healthy non-smoking men and women aged between 18-45 years. Exclusion criteria were clinical signs or history of heart, lung, kidney, endocrine or malignant disease; abnormal findings in ECG, urine dipstick or biochemistry [blood cell count, plasma concentrations of glucose, bilirubin, alanine aminotransferase, alkaline phosphatase, sodium, potassium, creatinine, albumin, cholesterol and haemoglobin; arterial hypertension (24 h ambulatory BP > 130/80 mmHg); medical treatment (except oral contraceptives); alcohol and substance abuse, *i.e.*, more than 21 alcoholic drinks per week for males and 14 drinks for females]; current smoking; pregnancy, breast feeding; donation of blood within one month prior to the study and obesity (BMI > 32 kg/m²). Withdrawal criteria were, development of conditions given in exclusion criteria during the study, withdrawal of informed consent and poor compliance.

Participants were recruited through advertisement at public institutions in Holstebro, Denmark.

Study settings

The study took place at Department of Medical Research, University Clinic of Nephrology and Hypertension, Regional Hospital Holstebro, Denmark, from 1st of August 2012 until 13th of September 2013.

Ethics

This study was reviewed and approved by the Regional Committees on Health Research Ethics, Skottenborg 26, Viborg, Denmark (j.no 1-10-72-178-12) and was carried out in accordance with the Helsinki Declaration. All study participants provided informed written consent prior to study enrolment.

Effect variables

The main effect variable was u-NKCC2. Secondary effect variables were: u-AQP2, u-ENaC_γ, glomerular filtration rate (GFR), free water clearance (C_{H₂O}), urine output (UO), urinary excretion of sodium (u-Na) and potassium (u-K), fractional excretion of sodium (FE_{Na}) and potassium (FE_K), plasma sodium (p-Na) and potassium (p-K), plasma osmolality (p-osm) and plasma albumin (p-alb), plasma concentration of renin (PRC), angiotensin II (p-Ang II), aldosterone (p-Aldo), vasopressin (p-AVP), extracellular fluid volume (ECV),

intracellular fluid volume (ICV), total body water (TBW), brachial systolic- (bSBP) and diastolic blood pressure (bDBP), pulse wave velocity (PWV) and central systolic- (cSBP) and diastolic blood pressure (cDBP).

Number of subjects

Using a significance level of 5% and a power of 80% it was calculated that the number of subjects should be 16, when the minimal relevant difference in u-NKCC2 was 0.3 ng/min and SD was 0.3 ng/min. In this study, incomplete voiding during study days was expected in some subjects; therefore, 20 subjects were included as a minimum.

Randomisation

Subjects were randomized to treatment using block randomization conducted at www.randomization.com. Aarhus Hospital Pharmacy, Denmark, generated the randomization sequence into five blocks of six from 01-30 and labeled the bottles. Five days prior to each examination day, participants received a numbered bottle containing BFTZ, amiloride or placebo tablets. BFTZ, amiloride and placebo were capsulated in grey DB Caps[®] size B (Capsugel) with click effect to obtain blinding. The randomization code was kept at Aarhus Hospital Pharmacy during the trial. Individual randomization codes were kept in sealed envelopes at Department of Medical Research if necessary for the investigator to know the given treatment. Investigators, participants and other study personnel were blinded to treatment assignment for the duration of the study.

Experimental procedures

Experimental procedure prior to the study day:

Four days prior to each study day, subjects consumed a standardized diet regarding calories, sodium and fluid. The diet consisted of 11000 (kJ/d) with an energy distribution of 55% carbohydrates, 30% fat and 15% protein in accordance to general dietary guidelines. The sodium content was approximately 120-150 mmol pr. day. The subjects were asked to drink 2500 mL/d. No alcohol or soft drink consumption was allowed while on the standardized diet. A maximum of two cups (6 oz.) of coffee or tea was allowed daily. Subjects were instructed to keep their usual physical activity during the experiments but to abstain from hard training the day prior to the examination. A 24-h urine collection from 7:00 AM to 7:00 AM on the examination day was used to assess water and sodium balance. A 24-h ambulatory BP measurement was performed to evaluate the effect of the intervention on blood pressure (Table 1).

Interventions

During the four-day diet and the morning of the examination day, participants were randomized to capsules containing either 1.25 mg BFTZ, 5 mg amiloride or matching placebo twice daily at 7 AM and 6 PM.

Table 1 Experimental procedures

Periods	Before the study day				On the study day												
	Day-4	Day-3	Day-2	Day-1	6:00-8:00	8:00-8:30	8:30-09:00	9:00-9:30	9:30-10:00	10:00-10:30	10:30-11:00	11:00-11:30	11:30-12:00	12:00-12:30	12:30-13:00	13:00-13:30	
								0	30	Baseline	60	90	Infusion 120	150	180	210	240
Time																	
Diet	x	x	x	x													
Study drug	xx	xx	xx	xx	x												
24-h BP																	
24-h urine																	
IV access																	
Weight																	
Water load																	
Urine sample																	
Blood samples																	
Blood pressure																	
⁵¹ Cr-EDTA																	
IV. fluid																	
App. Ton																	
BIS																	

24-h BP: 24-h ambulatory blood pressure measurements; 24-h urine: 24-h urine collection; App.Ton: Applanation tonometry; BIS: Bioimpedance spectroscopy.

Experimental procedure on the study day

Table 1 shows the time points of study interventions. Following an overnight fast, subjects arrived at our research facility at 8:00 AM. Two indwelling catheters for blood sampling and administration of ⁵¹Cr-EDTA and fluid were placed in both cubital veins. Every 30 min, starting at arrival, participants received an oral water load of 175 mL. Urine was collected in standing or sitting position. Otherwise, subjects were kept in a supine position in a quiet temperature-controlled room (22 °C-25 °C). At 9:00 AM a priming dose of ⁵¹Cr-EDTA was administered, followed by sustained infusion. Three 30-min baseline clearance periods were obtained from 9:30 AM to 11:00 AM. The baseline periods were followed by an infusion period from 11:00 AM to 12:00 PM during which a sustained infusion of 3% hypertonic saline was administered. The post infusion period consisted of three 30-min periods from 12:00 PM to 1:30 PM. Blood and urine samples were collected every 30 min from 8:30 AM to 1:30 PM.

Blood samples were drawn and analyzed for ⁵¹Cr-EDTA, p-sodium, p-potassium, p-albumin and p-osmolality. Analysis of PRC, p-Aldo II, p-Aldo and p-AVP were conducted from blood samples drawn at 11:00 AM, 12:00 PM and 1:30 PM.

Urine samples were analyzed for u-⁵¹Cr-EDTA, u-sodium, u-creatinine and u-osmolality. Analysis of u-AQP2, u-NKCC2 and u-ENaCγ was conducted from the 24-h urine collection and clearance period 10:30-11:00 AM (basal); 11:00-12:00 AM (fluid infusion), 12:00-12:30 PM (30 min after cessation of fluid infusion) and 1:00-1:30 PM (90 min after cessation of fluid infusion). For data analysis, the 30-min periods from 9:30 AM to 1:30 PM were subdivided into: baseline (0-90 min), infusion period (90-150 min) and three post infusion period 150-180 min, 180-210 min and 210-240 min).

Measurements of PWV, augmentation index (Aix) and cBP were performed at 11:00 AM (before infusion) and 12:00 AM (after infusion). Body composition was measured at 8:30 AM, 11:00 AM, 12:00 PM and 1:30 PM (end of examination day).

Measurements

Renal function: Glomerular filtration rate was measured by the constant infusion clearance technique with ⁵¹Cr-EDTA as reference substance. More than 15% variation in GFR between the three baseline periods led to the exclusion of clearance related analysis.

Fractional excretion of sodium and potassium was calculated as: $[\text{Sodium/potassium clearance } (C_{\text{Na/K}}) / \text{GFR} \times 100\%]$. Free water clearance was calculated as: $[\text{Urine output (UO)} - \text{osmolar clearance } (C_{\text{OSM}})]$. C_{OSM} was calculated as: $[\text{Urine osmolality/plasma osmolality} \times \text{UO}]$.

Blood samples: were centrifuged for 10 min at 2200 $\times g$ at 4 °C. Plasma hormone samples were kept frozen at -20 °C (Ang II) and -80 °C (PRC, Aldo, and AVP) until assayed. Renin in plasma was determined using an immunoradiometric assay (CIS Bio International, Gif-Sur-Yvette Cedex, France). Minimal detection level was 1 pg/mL. The coefficients of variation were 14.5% (interassay) and 4.5% (intra assay). Aldosterone in plasma was determined by radioimmunoassay (Demeditec Diagnostics Systems Laboratories Inc., Webster, TX, United States). Minimal detection level was 22 pmol/L. The coefficients of variation were 8.2% (inter-assay) and 3.9% (intra-assay). Arginine vasopressin and Angiotensin II were extracted from plasma with C₁₈ Sep-Pak (Water associates, Milford, MA, United States) and subsequently measured using radioimmunoassay as previously described^[13]. The antibody against angiotensin II was obtained from the Department of Clinical Physiology, Glostrup Hospital, Glostrup, Denmark. Minimal detection level was 2 pmol/L. The coefficients of variation were 12% (inter-assay) and 8% (intra-assay). The antibody against AVP was a gift from Professor Jacques Dürr (Miami, FL, United States). Minimal detection level was 0.2 pmol/L. The coefficients of variation were 13% (inter-assay) and 9% (intra-assay).

Generation of NKCC2 specific antibody: A novel rabbit polyclonal antiserum against human NKCC2 (*Slc12a2*) was generated against the following peptide: CNITKTPKKDGSIN by Genscript® (New Jersey, United States). The N-terminal cysteine was added for conjugation to carrier protein and for attaching the peptide to the affinity purification column. The immune serum from two rabbits (#593 and #594) was affinity purified using immunizing peptides, resulting in NKCC2-specific antibodies. NKCC2 antibody characterization has previously been described^[12].

Urine sample immunoassays: Urines were stored frozen at -20 °C until assayed.

U-NKCC2 was measured in urine by a newly developed radioimmunoassay^[12]. Antibodies were raised in rabbits against human NKCC2 (*Slc12a2*) against the peptide CNITKTPKKDGSIN. The N-terminal cysteine was added for conjugation to carrier protein and affinity purification. Minimal detection level was 0.5 ng/tube. The coefficients of variation were 14% (inter-assay) and 6.8% (intra-assay).

U-AQP2 was measured by radioimmunoassay as previously described^[9,14]. Antibodies were raised in rabbits to a synthetic peptide corresponding to the 15 COOH-

terminal amino acids in human AQP2 to which was added an NH₂-terminal cysteine for conjugation and affinity purification. Minimal detection level was 34 pg/tube per tube. The coefficients of variation were 11.7% (inter-assay) and 5.9% (intra-assay).

U-ENaC γ was measured by radioimmunoassay as previously described^[15,16]. Antibodies were raised against a synthetic ENaC γ peptide in rabbits and affinity purified^[17]. Minimal detection level was 48 pg/tube. The coefficients of variation were 14% (inter-assay) and 6.7% (intra-assay).

Blood pressure measurement: Twenty-four hours BP was measured using Kivex TM-2430 (Kivex, Hoersholm, Denmark). Measurements were taken every 15 min during daytime and every 30 min overnight. Brachial blood pressure was recorded using a semiautomatic oscillometric device (Omron 705IT, Omron Matsusaka, Japan).

Plasma and urine: Concentrations of sodium, potassium, creatinine and albumin were measured using routine methods at the Department of Clinical Biochemistry, Holstebro Hospital.

Plasma and urine osmolality was measured by freezing point depression (Advanced Model 3900 multisampling osmometer).

Bioimpedance spectroscopy: Was performed at 50 frequencies, from 5 to 1000 kHz using the Fresenius Body Composition Monitor and the Fluid Management Tool, version 3.

Applanation tonometry: Recordings of PWA and carotid-femoral PWV were obtained by applanation tonometry (SphygmoCor® CPV system®, AtCor Medical, Sydney, Australia) as double-recordings by a trained observer. Only duplicate recording meeting the quality requirements were included in the final analysis. An operator index of 80 or more was required to accept recordings of a peripheral pulse-wave form^[18].

Study drug

Bendroflumethiazide [Tablet Salures 2.5 mg (1/2 tablet)] were obtained from Pfizer AB, Sollentuna, Sweden. Amiloride (Tablet Amilorid Mylan 5 mg) were obtained from Mylan AB, Stockholm, Sweden via Tjellesen Max Jenne A/S, Medilink A/S, Roedovre, Denmark.

Statistical methods

Statistical analyses were performed by the authors using IBM SPSS statistics version 20.0.0 (IBM Corp.; Armonk, NY, United States).

As clearance data from the three baseline periods were very similar, single baseline values were obtained by taking the average of the measurements from the three baseline periods. Parametric data are presented as

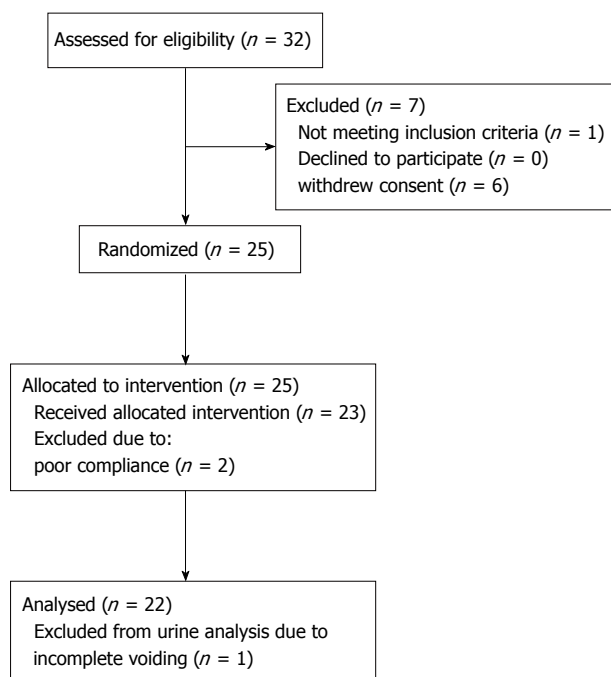


Figure 1 Flow chart.

means \pm SD and nonparametric data as medians with 25th and 75th percentiles. General linear model (GLM) with repeated measures was performed, with time as within-subject factor and intervention as between subject factor, to test for differences within and between groups. One-way ANOVA was used for comparison of means between groups when differences were found. For non-parametric data, related samples Friedman's two-way analysis was used. Post hoc Bonferroni correction was used for multiple comparisons of post infusion periods to baseline within each treatment group. Statistical significance was defined as $P < 0.050$ in all analyses.

RESULTS

Demographics

Thirty-two healthy women and men were assessed for eligibility. Nine were excluded due to: withdrawal of informed consent (6), non-compliance (2) or 24-h BP above 130/80 mmHg (1). Thus, 23 were initially allocated to and completed the study. One was not able to void satisfactorily during baseline clearance experiments and was excluded from urine analysis (Figure 1).

The 23 subjects (8 males; 15 females) who completed the trial had a mean age of 26 ± 4 years, BMI 24 ± 3 kg/m², 24-h BP $117/69 \pm 7/4$ mmHg. Screening blood values were b-haemoglobin 8.4 ± 0.7 mmol/L, p-sodium 141 ± 1 mmol/L, p-potassium 3.7 ± 0.4 mmol/L, p-creatinine 79 ± 14 μ mol/L, eGFR 93 ± 16 mL/min, p-albumin 43 ± 3 g/L, p-glucose 5 ± 1 mmol/L, p-alanine transaminase 25 ± 19 U/L and p-cholesterol 4.4 ± 0.8 mmol/L. Baseline values did

not differ between males and females, apart from p-crea (males: 91 ± 15 μ mol/L vs females: 73 ± 7 μ mol/L, $P < 0.012$), b-haemoglobin (males: 8.9 ± 0.6 mmol/L vs females: 8.2 ± 0.5 mmol/L, $P < 0.016$) and p-albumin (males: 45 ± 2 g/L vs females: 42 ± 3 g/L, $P < 0.003$).

Effects of BFTZ and amiloride on 24-h urine and ambulatory BP

UO, u-osm, C_{H₂O}, Creatinine-Clearance, u-Na, u-AQP2 and ENaC γ in 24-h urine were not significantly different between treatments. During BFTZ treatment u-NKCC2 and u-K were significant higher than both amiloride and placebo treatment (Table 2). Twenty-four hour ambulatory bSBP did not differ between treatments, however there was a small but significant lower bDBP during amiloride treatment (Table 2).

Effects of BFTZ and amiloride on u-NKCC2, u-ENaC γ and u-AQP2

Figure 2 shows the changes in urinary excretion of AQP2, NKCC2 and ENaC γ during basal, infusion and post-infusion periods.

At baseline, u-NKCC2 did not differ between groups. U-NKCC2 decreased during the infusion period and increased during the first post infusion period in all three treatments. U-NKCC2 increased further during amiloride ($6\% \pm 34\%$; $P = 0.081$) and placebo ($17\% \pm 24\%$; $P = 0.010$), whereas u-NKCC2 declined in the BFTZ treated group ($-7\% \pm 28\%$; $P = 0.257$), during the two last post infusion periods. By the end of the examination day there was a significant difference between BFTZ vs amiloride ($P < 0.001$) and vs placebo ($P = 0.033$). There was no significant difference between amiloride and placebo groups ($P = 0.407$).

At baseline, u-ENaC γ was similar. In response to 3% saline, u-ENaC γ increased significant to a maximum after the first post infusion period. Although u-ENaC γ tended to be lower during amiloride treatment, there was no statistical difference between the three treatment groups throughout the examination day.

There was no significant difference in u-AQP2 at baseline. In response to 3% saline, u-AQP2 increased significantly and similarly during amiloride ($31\% \pm 22\%$; $P < 0.001$) and placebo treatment ($34 \pm 27\%$; $P < 0.001$), but did not change during BFTZ ($5\% \pm 16\%$; $P = 0.261$). By the end of the examination day there was a significant difference between BFTZ vs amiloride and placebo ($P < 0.001$), but there was no difference between amiloride and placebo.

Divided by gender, the creatinine adjusted excretion of u-AQP2, u-NKCC2 and u-ENaC γ tended to be higher in females compared to males in all three treatment groups, but the difference is attributed to a lower urinary excretion of creatinine in females (data not shown).

Effects of BFTZ and amiloride on GFR and tubular function

Table 3 shows the absolute values of C_{H₂O}, UO, FE_{Na},

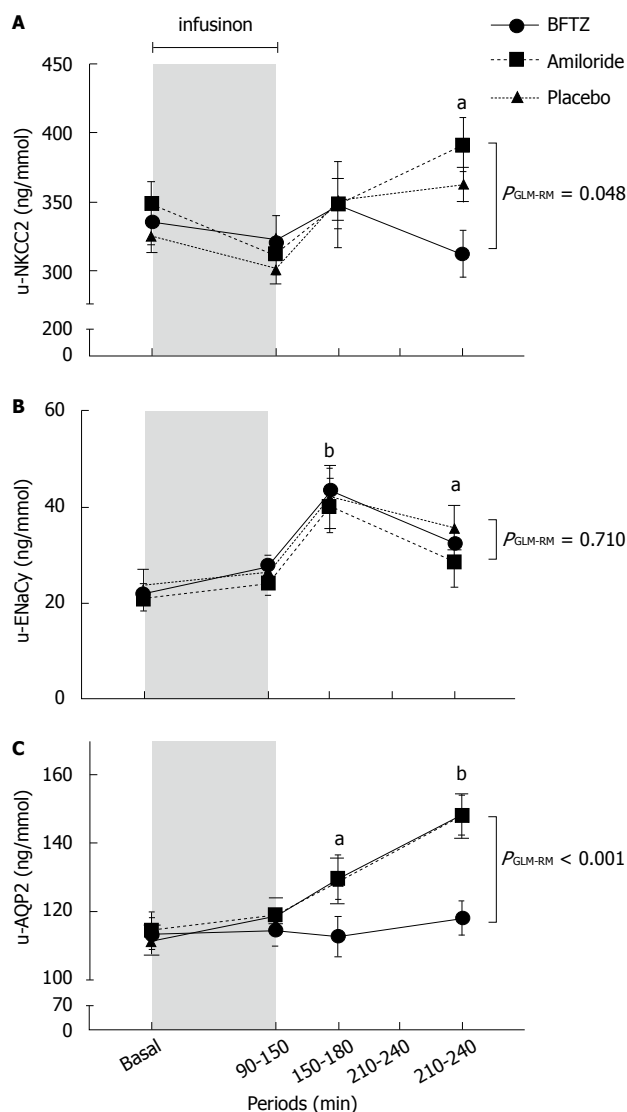


Figure 2 Effects of 3% hypertonic saline on urinary excretion of sodium-potassium-2chloride co-transporter (A), gamma fraction of epithelial sodium channels (B) and aquaporin2 (C) in 22 healthy subjects treated with bendroflumethiazide, amiloride or placebo. Values are means \pm SEM. General linear model with repeated measurements (GLM-RM) was performed to test for differences between groups. A: U-NKCC2 increased during amiloride ($P = 0.081$) and placebo ($P = 0.010$) treatments. The increase in u-NKCC2 was however only significant during placebo. U-NKCC2 did not change during BFTZ; B: U-ENaC increased significantly and to the same extent during all three treatments; C: U-AQP2 increased significantly during amiloride and placebo ($P < 0.001$), but not during BFTZ. Paired t-test was used for comparison of post-infusion periods vs baseline. ^a $P < 0.050$; ^b $P < 0.001$. AQP2: Aquaporin2; U-NKCC2: Urinary excretion of sodium-potassium-2chloride co-transporter; ENaC: Epithelial sodium channels; BFTZ: Bendroflumethiazide.

u-Na, FE_K , u-K and ^{51}Cr -EDTA clearance.

CH_2O and UO decreased significantly in all three treatments. At baseline, CH_2O was lower during BFTZ and showed an attenuated decrease at post infusion period 210-240 min compared to amiloride ($P = 0.207$) and placebo ($P = 0.005$).

At baseline, FE_{Na} and u-Na were higher during amiloride compared to BFTZ and placebo. After 3% saline infusion there was a significant increase in u-Na and FE_{Na} in all three treatments, but less pronounced

during BFTZ ($P = 0.001$).

There was no difference in u-K at baseline. In response to 3% saline, u-K and FE_K decreased during BFTZ and increased during amiloride compared to placebo. There was a significant difference between all three treatments ($P = 0.001$). GFR did not change significantly.

Effects of BFTZ and amiloride on plasma hormones

Figure 3 shows the changes in PRC, Ang II, p-Aldo and p-AVP during the examination day. PRC, Ang II and p-Aldo were significantly increased during active treatment compared to placebo. PRC and p-Ang II were highest during BFTZ treatment ($P < 0.001$), whereas p-Aldo was highest during amiloride treatment ($P < 0.001$). PRC, Ang II and p-Aldo declined significantly in response to 3% saline, in all three treatments, with no relative differences between treatments.

P-AVP was similar at baseline. P-AVP increased in all three groups, in response to 3% saline. Although, p-AVP was lower during BFTZ at 150 min ($P = 0.048$), the relative increase in p-AVP, after 3% saline, was not significantly different between BFTZ vs placebo ($82\% \pm 100\%$ vs $116\% \pm 67\%$; $P = 0.072$).

Effects of BFTZ and amiloride on plasma

Table 4 shows the absolute values of p-Na, p-K, p-Osm and p-Alb during basal-, infusion- and post infusion periods. During baseline conditions p-osm and p-Na were significantly lower during BFTZ and amiloride compared to placebo. P-K was higher in the amiloride group compared to placebo and BFTZ, and p-K was lower during BFTZ compared to placebo.

In response to 3% saline infusion, p-Na and p-osm increased to the same extent in all three treatments, but remained highest in the placebo group. P-K decreased significant in the amiloride group compared to BFTZ and placebo. P-alb decreased significantly in all three treatments, in response to 3% saline.

Effects of BFTZ and amiloride on blood pressure

Table 5 shows the absolute values of bSBP, bDBP, pulse rate, cSBP, cDBP, PWV and AIX. At baseline there was no difference in bSBP or bDBP. In response to 3% hypertonic saline, bSBP increased and bDBP decreased. At the end of the day the decrease in bDBP was more pronounced during BFTZ ($-6\% \pm 6\%$) compared to amiloride ($-2\% \pm 6\%$; $P = 0.030$) and placebo ($-2\% \pm 5\%$; $P = 0.021$).

There was no difference in cSBP at baseline or in response to 3% saline between treatments. At baseline cDBP was the same in all three treatments, however cDBP decreased significant in the BFTZ group ($P < 0.001$) but not during amiloride and placebo. PWV followed the same pattern, however the decrease during BFTZ treatment was not significant.

Effects of BFTZ and amiloride on body fluid volumes

Figure 4 shows the changes in ICV, ECV and TBW during the examination day.

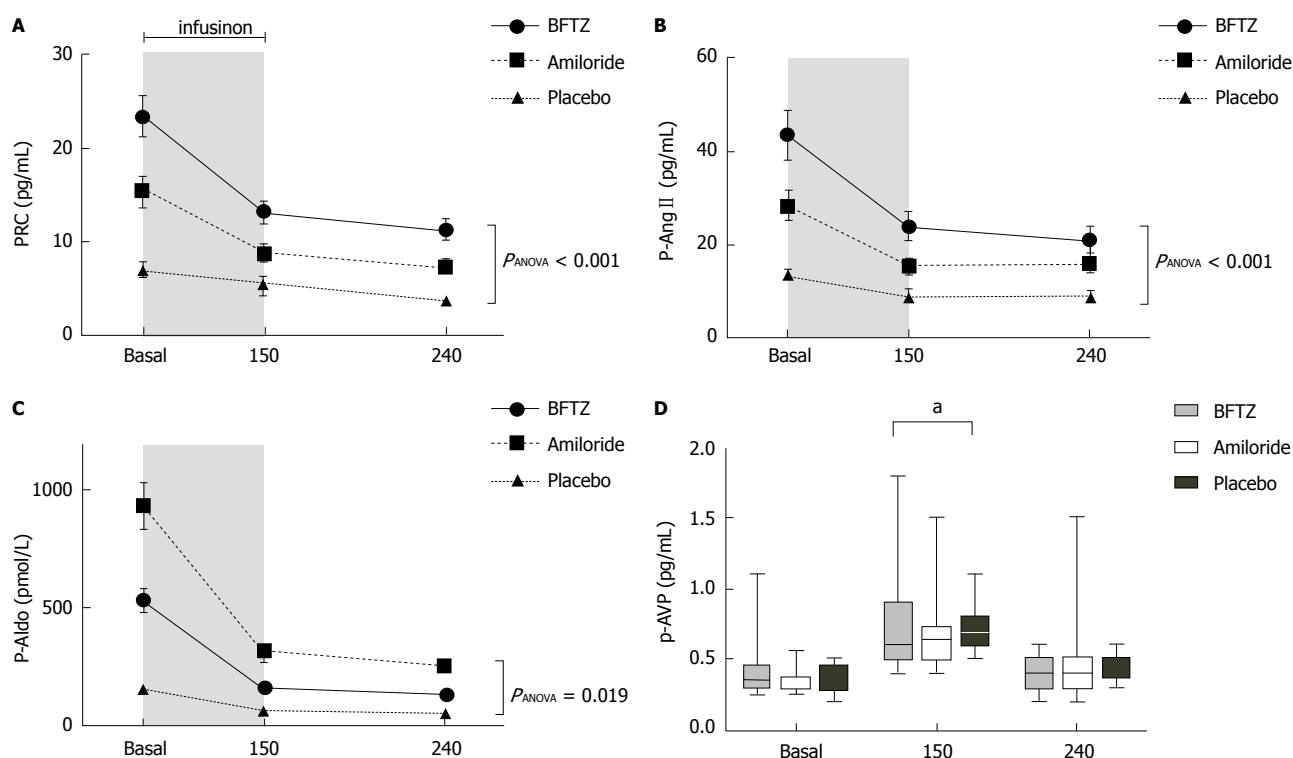


Figure 3 Effects of 3% hypertonic saline on plasma concentrations of renin (A), angiotensin II (B), aldosterone (C) and arginine vasopressin (D) in 23 healthy subjects pre-treated with bendroflumethiazide, amiloride or placebo. A-C: There was a significant difference between PRC, p-Ang II and p-Aldo plasma levels throughout the study day. Values are means \pm SEM. One-way ANOVA was used to test for differences between treatments; D: P-AVP increased significantly at 150 min with a borderline significant difference between treatments ($^aP = 0.048$). Values are medians with upper and lower limits. Friedman's test was used to test for differences between treatments. BFTZ: Bendroflumethiazide; PRC: Plasma renin concentration.

Table 2 Twenty-four hours brachial blood pressure and urine collection with fluid deprivation (12 PM to 8.00 AM) in 23 healthy subjects

	Examination day			P (ANOVA)
	Thiazide	Amilorid	Placebo	
Urine output (mL/24 h)	2527 \pm 728	2418 \pm 469	2316 \pm 700	0.481
u-osm (mosm/24 h)	865 \pm 158	835 \pm 176	761 \pm 187	0.087
C _{H2O} (mL/min)	-0.40 \pm 0.33	-0.37 \pm 0.47	-0.24 \pm 0.53	0.534
Cr.Cl (mL/min per m ²)	113 \pm 25	118 \pm 32	110 \pm 26	0.549
u-NKCC (ng/mmol)	0.35 \pm 0.07	0.30 \pm 0.05	0.32 \pm 0.06	0.025
u-AQP2 (ng/mmol)	113.2 \pm 39.2	103.3 \pm 25.5	98.5 \pm 17.4	0.244
u-ENaC γ (ng/mmol)	37.8 \pm 26.7	30.8 \pm 17.3	32.7 \pm 15.6	0.867
u-Na (mmol/24 h)	108 \pm 34	121 \pm 27	106 \pm 37	0.263
u-K (mmol/24 h)	80 \pm 20	64 \pm 21	60 \pm 19	0.002
bSBP (mmHg)	119 \pm 6	114 \pm 7	116 \pm 7	0.213
bDBP (mmHg)	72 \pm 4	69 \pm 3	70 \pm 4	0.034

Values are means \pm SD. One-way ANOVA was used for comparison between groups. *P*-values represent the possibility of a difference between groups. u-osm: Urine output, urine osmolality; C_{H2O}: Free water clearance; Cr.Cl: Creatinin clearance; u-NKCC2: Urinary NKCC2; u-AQP2: Urinary excretion of AQP2; u-ENaC γ : ENaC γ excretion adjusted for creatinin; u-Na: Urinary excretion of sodium; u-K: Potassium; bSBP: Brachial systolic blood pressure; bDBP: Brachial diastolic blood pressure.

At baseline, ECV and TBW tended to be lower during amiloride ($P = 0.515$) and BFTZ ($P = 0.951$) compared to placebo. However, it did not reach statistical significance. ICV did not differ between treatments. As expected, after administering 3% saline, ICV decreased while ECV and TBW increased reaching a maximum at the end of the study day. Although there was a tendency

towards a lower ECV and TBW in the two diuretic groups there were no statistically significant differences in volume status between the three treatments.

DISCUSSION

In the present study, the aim was to investigate the

Table 3 Effect of 3% hypertonic saline on urinary parameters in 22 healthy subjects treated with bendroflumethiazide or amiloride

Periods	Baseline	Infusion	Post infusion			<i>P</i> _{GLM RM}
	0-90 min	90-150 min	150-180 min	180-210 min	210-240 min	
<i>C</i> _{H2O}						< 0.001
BFTZ	3.1 ± 1.6 ^a	-0.2 ± 0.5 ^d	-1.6 ± 0.6 ^d	-1.6 ± 0.7 ^d	-0.6 ± 1.6 ^d	
Amiloride	3.7 ± 0.9	-0.6 ± 0.7 ^d	-2.1 ± 0.6 ^d	-2.4 ± 0.8 ^d	-1.4 ± 1.7 ^d	
Placebo	4.4 ± 1.1	-0.6 ± 0.6 ^d	1.8 ± 0.7 ^d	-2.5 ± 1.0 ^d	-2.0 ± 0.5 ^d	
<i>P</i> _{GLM between subjects}				0.061		
<i>P</i> _{ANOVA}	0.006	NS	NS	0.001	0.007	
UO (mL/min)						0.029
BFTZ	6.1 ± 1.6	2.6 ± 0.6 ^d	1.4 ± 0.5 ^d	1.7 ± 1.0 ^d	2.7 ± 1.9 ^d	
Amiloride	6.8 ± 1.3	2.5 ± 0.6 ^d	1.6 ± 0.5 ^d	2.0 ± 0.7 ^d	3.2 ± 1.2 ^d	
Placebo	7.3 ± 1.2	2.3 ± 0.9 ^d	1.7 ± 0.8 ^d	2.2 ± 1.2 ^d	2.4 ± 1.1 ^d	
<i>P</i> _{GLM between subjects}				0.245		
<i>P</i> _{ANOVA}	0.019	NS	NS	NS	NS	
u-Na (mmol/min)						< 0.001
BFTZ	1.4 ± 0.4	1.7 ± 0.4 ^b	1.9 ± 0.7 ^b	2.1 ± 0.6 ^d	2.0 ± 0.6 ^b	
Amiloride	1.6 ± 0.5	2.0 ± 0.7	2.5 ± 1.0 ^b	2.9 ± 1.2 ^d	3.0 ± 1.0 ^d	
Placebo	1.3 ± 0.3	1.9 ± 1.1 ^a	2.5 ± 1.3 ^b	3.3 ± 1.7 ^d	3.0 ± 1.0 ^d	
<i>P</i> _{GLM between subjects}				0.020		
<i>P</i> _{ANOVA}	0.028	NS	NS	0.007	<0.001	
FENa (%)						< 0.001
BFTZ	1.5 ± 0.4	1.8 ± 0.5 ^a	2.0 ± 0.6 ^b	2.2 ± 0.6 ^d	2.1 ± 0.6 ^d	
Amiloride	1.8 ± 0.6	2.1 ± 0.6 ^a	2.6 ± 1.0 ^b	2.9 ± 1.2 ^d	3.0 ± 1.1 ^d	
Placebo	1.4 ± 0.4	2.1 ± 1.0 ^b	2.7 ± 1.2 ^d	3.0 ± 1.1 ^d	3.1 ± 1.1 ^d	
<i>P</i> _{GLM between subjects}				0.036		
<i>P</i> _{ANOVA}	0.022	NS	NS	0.019	0.001	
u-K (mmol/min)						< 0.001
BFTZ	20.3 ± 6.7	17.7 ± 5.3	15.8 ± 4.4 ^b	14.2 ± 5.6 ^b	13.3 ± 6.3 ^b	
Amiloride	18.5 ± 8.4	15.7 ± 9.3	18.6 ± 11.1	22.4 ± 12.0	23.3 ± 10.2 ^a	
Placebo	22.3 ± 9.3	15.6 ± 6.9 ^d	16.4 ± 8.5 ^a	22.3 ± 12.3	20.4 ± 7.9	
<i>P</i> _{GLM between subjects}				0.255		
<i>P</i> _{ANOVA}	NS	NS	NS	0.015	0.001	
FEK (%)						< 0.001
BFTZ	21.7 ± 7.4	20.0 ± 6.6	16.1 ± 4.8 ^b	15.0 ± 5.9 ^b	14.4 ± 6.9 ^b	
Amiloride	20.8 ± 9.8	18.7 ± 10.8	20.5 ± 12.0	23.6 ± 12.4	25.0 ± 11.0 ^a	
Placebo	23.7 ± 9.3	18.1 ± 8.2 ^b	17.5 ± 8.9 ^a	20.6 ± 9.9	21.4 ± 8.8	
<i>P</i> _{GLM between subjects}				0.254		
<i>P</i> _{ANOVA}	NS	NS	NS	0.018	0.001	
⁵¹ Cr-EDTA (mL/min per 1.73m ²)						0.271
BFTZ	92.1 ± 10.8	91.3 ± 11.9	96.0 ± 16.7	96.9 ± 16.8	96.5 ± 21.2	
Amiloride	92.7 ± 13.7	93.2 ± 12.8	94.1 ± 12.1	96.7 ± 14.1	100.2 ± 15.6	
Placebo	96.5 ± 9.5	90.0 ± 13.3	94.7 ± 15.1	102.4 ± 14.3	98.1 ± 15.9	
<i>P</i> _{GLM between subjects}				0.887		

Free water clearance (*C*_{H2O}), urinary output (OU), excretion of sodium (u-Na) and fractional excretion of sodium (FENa), urinary excretion of potassium (u-K) and fractional excretion of potassium (FEK) and ⁵¹Cr-EDTA clearance in a randomized, placebo-controlled, crossover study of 23 healthy subjects. Values are mean ± SD. General linear model (GLM) with repeated measures was performed for comparison within the group and intervention as between subjects factor. One-way ANOVA was performed when differences were found between interventions. Post hoc Bonferroni correction was used for multiple comparisons of post infusion periods to baseline within each treatment group. ^a*P* < 0.05; ^b*P* < 0.01; ^d*P* < 0.001.

effect of five days BFTZ and amiloride treatment on the urinary excretion of NKCC2, ENaC_γ and AQP2 during baseline conditions and after an acute intravenous volume load of 3% hypertonic saline in healthy subjects. To our knowledge, this study is the first randomized, placebo-controlled trial that measured the changes in u-NKCC2, u-ENaC_γ and u-AQP2 during inhibition of the NCC cotransporter with BFTZ and ENaC with amiloride in humans.

This study showed that, in response to 3% saline, u-NKCC2, u-ENaC_γ and u-AQP2 increased to the same

extent during amiloride and placebo treatment, but neither u-NKCC2 nor u-AQP2 changed significantly during BFTZ.

Sodium and tubular sodium transporters

Thiazides predominantly inhibit NCC along the distal convoluted tubules^[2]. Animal studies have shown that when thiazide was administered chronically, urinary sodium returned to normal within 2-3 d^[19]. This is in accordance with our findings in 24 h urine collection. Further, a study documented that longer term NCC

Table 4 Effect of 3% hypertonic saline on plasma in 23 healthy subjects treated with bendroflumethiazide or amiloride

Time	Baseline	Infusion	Post infusion			<i>P</i> _{GLM-RM}
	0-90 min	150 min	180 min	210 min	240 min	
p-Na						0.281
BFTZ	137 ± 2	141 ± 2 ^d	141 ± 2 ^d	139 ± 2 ^d	139 ± 2 ^d	
Amilorid	137 ± 2	141 ± 2 ^d	141 ± 2 ^d	140 ± 2 ^d	139 ± 2 ^d	
Placebo	139 ± 1	43 ± 2 ^d	142 ± 1 ^d	141 ± 1 ^d	140 ± 1 ^d	
<i>P</i> _{GLM between subjects}			0.003			
<i>P</i> _{ANOVA}	0.001	0.019	0.004	0.007	0.005	
p-K						0.001
BFTZ	3.35 ± 0.22	3.32 ± 0.23	3.43 ± 0.26	3.42 ± 0.21	3.40 ± 0.20	
Amilorid	4.32 ± 0.30	4.17 ± 0.23 ^d	4.26 ± 0.27	4.27 ± 0.25	4.20 ± 0.20 ^a	
Placebo	3.89 ± 0.18	3.83 ± 0.22	3.97 ± 0.23	3.94 ± 0.22	3.92 ± 0.20	
<i>P</i> _{GLM between subjects}			< 0.0001			
<i>P</i> _{ANOVA}	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	
p-Osm						0.600
BFTZ	281 ± 5	288 ± 4 ^d	288 ± 6 ^d	286 ± 5 ^d	284 ± 4 ^d	
Amilorid	283 ± 4	290 ± 4 ^d	290 ± 3 ^d	287 ± 3 ^d	285 ± 3 ^a	
Placebo	286 ± 3	294 ± 4 ^d	292 ± 4 ^d	290 ± 4 ^d	289 ± 3 ^b	
<i>P</i> _{GLM between subjects}			< 0.0001			
<i>P</i> _{ANOVA}	< 0.001	< 0.001	0.005	0.002	< 0.001	
p-Alb (g/L)						0.007
BFTZ	40.7 ± 3.1	35.5 ± 2.2 ^d	35.9 ± 2.7 ^d	36.0 ± 2.8 ^d	36.0 ± 2.7 ^d	
Amilorid	40.9 ± 2.9	35.5 ± 2.3 ^d	36.4 ± 2.6 ^d	36.4 ± 2.7 ^d	36.3 ± 2.5 ^d	
Placebo	39.0 ± 2.4	34.5 ± 2.0 ^d	35.1 ± 2.2 ^d	35.1 ± 2.3 ^d	35.3 ± 2.4 ^d	
<i>P</i> _{GLM between subjects}			0.203			

Plasma concentrations of sodium (p-Na), potassium (p-K) and albumin (p-Alb) and plasma osmolality (p-osm). Values are mean ± SD. General linear model (GLM) with repeated measures was performed for comparison within the group and intervention as between subjects factor. One-way ANOVA was performed when differences were found between interventions. Bonferroni correction was used for multiple comparisons between study-periods *vs* baseline. ^a*P* < 0.05; ^b*P* < 0.01; ^d*P* < 0.001.

inhibition might cause a structural adaption, which will activate ENaC and cause increased distal sodium reabsorption and kaliuresis^[20]. Twenty-four hours urine collections, demonstrated a small, but significantly increased u-K, and increased u-ENaC_γ during BFTZ compared to amiloride and placebo; which supports the theory of a compensatory increase in sodium reabsorption *via* ENaC during longer-term thiazide treatment.

In this present study, 3% hypertonic saline induced an increase in u-NKCC2 when subjects were treated with amiloride and placebo. It was probably related to a counter regulatory mechanism to compensate for temporarily impaired lower fractional sodium reabsorption in proximal tubules during volume expansion^[21-23]. It has previously been described in healthy humans that u-NKCC2 decreased after 3% saline^[12]. However, the subjects' average age was approximately 35 years older in the aforementioned study. Tian *et al.*^[24] showed a blunting in the up regulation of sodium transport proteins in response to water restriction in aged rats, which seemed to be particularly apparent with regards to NKCC2. The age difference might explain the discrepancy in the u-NKCC2 response between the two studies.

During BFTZ treatment, u-NKCC2 ceased to increase. In rats, chronic thiazide treatment produces a compensatory fractional increased reabsorption of sodium in proximal tubules^[19], which might explain why

u-NKCC2 ceased to increase in the late post infusion periods during BFTZ. Thus, during BFTZ, there was no need of a compensatory reabsorption *via* NKCC, which is supported by the relative lower increase in FE_{Na} during BFTZ compared to both amiloride and placebo, in response to 3% saline. In animals, AVP has been demonstrated to increase NKCC2 activity, mediated by V2 receptors *via* adenylate-cyclase-6 to facilitate phosphorylation and trafficking of NKCC2 to the apical membrane^[25,26]. As p-AVP was lower during BFTZ treatment, it cannot be excluded that the decline in u-NKCC2 might also reflect a lack of stimulation from AVP.

Thus, our findings reflect a more profound change in glomerular tubular balance during BFTZ treatment, than the more distal acting diuretic, amiloride.

In the collecting ducts, sodium transport occurs *via* the ENaC located in the luminal membrane of principal cells^[27,28]. ENaC can be regulated by aldosterone^[29,30], but is also regulated by AVP, that binds to the V2 receptors and induces a rapid change in channel activity *via* ENaC opening^[31-36]. Recently our group demonstrated an increased u-ENaC_γ after hypertonic saline infusion in healthy young subjects. The increase in u-ENaC_γ was explained by an increased sodium load to the distal tubules caused by a decrease in renal sodium absorption in the proximal tubules^[11,21,22]. In the present study, we measured a similar increase during all three treatments. As amiloride inhibits

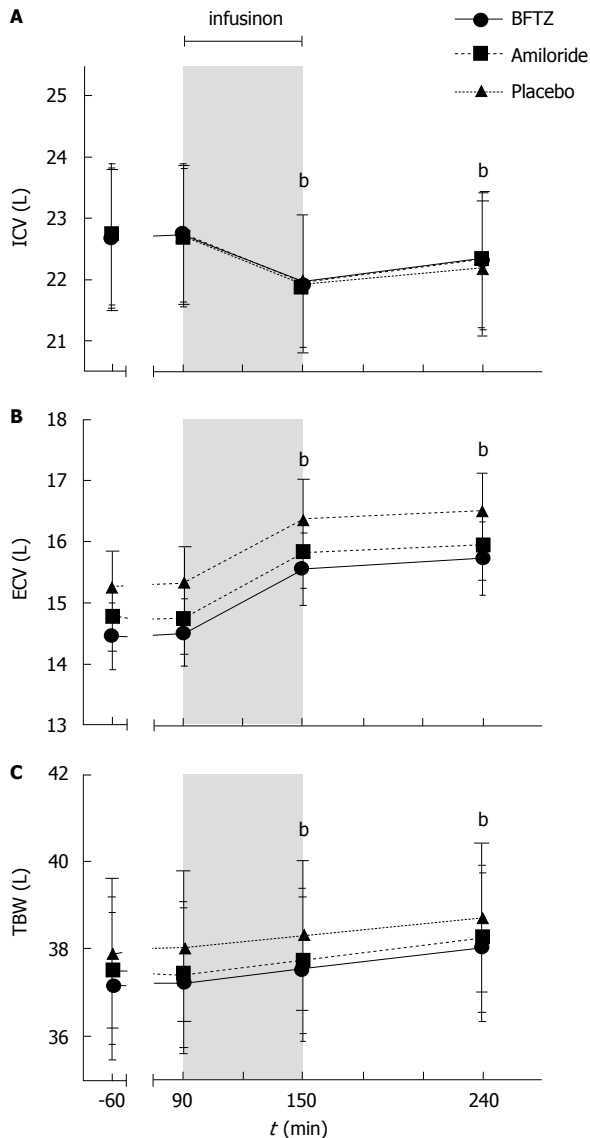


Figure 4 Effects of 3% hypertonic saline on (A) intracellular and (B) extracellular volume and (C) total body water in 23 healthy subjects pretreated with bendroflumethiazide, amiloride or placebo. Values are means \pm SEM. General linear model with repeated measures was non-significant between treatments. Paired *t*-test was used for comparison of post infusion periods vs baseline ^b*P* < 0.001. BFTZ: Bendroflumethiazide; ECV: Extracellular volume; ICV: Intracellular volume; TBW: Total body water.

ENaC, we expected to find a decrease in u-ENaC_f both at baseline and in response to 3% saline treatment during amiloride treatment, especially as we had also found an increased FE_{Na} at baseline. There are several possible explanations: Firstly, as ENaC only controls as little as 2%-5% of sodium reabsorption, perhaps a small decrease in the fractional reabsorption of sodium *via* ENaC would cause a significant rise in excretion of sodium. Secondly, p-Aldo was highest during amiloride treatment and its stimulation on the principal cells might have increased the amount of ENaC within the apical membrane, and thus antagonized the effect of amiloride. Thirdly, amiloride treatment has been shown to increase whole cell channel abundance caused by an intracellular sodium feedback mechanism^[37,38]. These

intracellular counter regulatory mechanisms might also be involved.

As expected, during the acute hypertonic sodium load, PRC, p-Ang II and p-Aldo decreased and urinary sodium excretion increased^[9,11,39]. The increase in urinary sodium excretion is preceded by a decrease in ENaC expression and activity^[40]. Meanwhile, p-AVP increased due to increased p-osm, and likely caused ENaC channels to be inserted in the apical membrane and thus increased reabsorption of sodium^[41].

Thus, despite increased FE_{Na} at baseline during amiloride treatment, u-ENaC_f did not differ significantly between treatments neither at baseline nor after 3% saline. These findings do not appear to be dependent on aldosterone, but more to reflect a compensatory role of ENaC to adjust for the decreased reabsorption of sodium in proximal parts of the nephron during an acute sodium load.

Water and AQP2

Vasopressin (AVP) regulates AQP2 function by binding to V2 receptors in the basolateral membrane of principal cells, increasing the delivery of intracellular vesicles containing AQP2 to the apical membrane and thus increasing water reabsorption^[42,43]. AQP2 is also excreted into urine^[5-9,44-46]. Volume expansion with 3% hypertonic saline increases plasma osmolality, p-AVP, reabsorption of water and u-AQP2^[11,14,47]. In the present study, there was an increase in u-AQP2, in response to 3% saline, during amiloride and placebo treatment, but not during BFTZ. The changes in u-AQP2 during BFTZ correspond to the attenuated decrease in CH₂O. Different explanations include: (1) an increased water load to the collecting tubules due to inhibition of NCC in distal collecting ducts, resulting in higher water excretion; (2) Decreased p-osm and p-AVP during BFTZ treatment and thus a reduced effect on V2R in the collecting ducts; and (3) A reduced need for counter regulatory water reabsorption in the collecting ducts due to the lower reabsorption of sodium *via* NKCC2.

Potassium

Potassium is freely excreted in glomerulus and it is reabsorbed and secreted across the nephron^[48]. Intracellular signalling networks, volume status, p-K status and aldosterone tightly regulate the balance of potassium excretion^[49,50]. Thiazides do not affect potassium transport directly, but induce adaption primarily along the connecting and collecting tubules where enhanced sodium reabsorption stimulates potassium secretion *via* renal outer medullary K⁺ (ROMK) and large-conductance K⁺ (BK) channels^[20,51].

It has recently been shown that angiotensin II directly inhibits ROMK in potassium-depleted animals, and thereby contributes to potassium conservation^[52,53]. In this present study, p-Ang II was highest during BFTZ treatment, and may have inhibited ROMK, and explains the increased sodium reabsorption during 3% saline

Table 5 Effect of 3% hypertonic saline on brachial blood pressure, central blood pressure and pulse wave velocity in 23 healthy subjects treated with bendroflumethiazide or amiloride

Periods	Baseline	Infusion	Post infusion			<i>P</i> _{GLM RM}
	0-90 min	150 min	180 min	210 min	240 min	
bSBP						0.825
BFTZ	112 ± 9	118 ± 9 ^d	116 ± 10	116 ± 11	117 ± 13	
Amilorid	110 ± 8	115 ± 9 ^d	114 ± 10 ^d	114 ± 10 ^b	114 ± 10 ^b	
Placebo	113 ± 10	117 ± 10 ^d	115 ± 9	115 ± 8	115 ± 10	
<i>P</i> _{GLM between subjects}			0.679			
bDBP						0.055
BFTZ	66 ± 7	63 ± 7 ^b	62 ± 6 ^d	62 ± 7 ^d	62 ± 7 ^b	
Amilorid	64 ± 4	62 ± 5	60 ± 5 ^b	61 ± 5	62 ± 6	
Placebo	64 ± 6	63 ± 5	63 ± 5	62 ± 6	63 ± 6	
<i>P</i> _{GLM between subjects}			0.695			
Pulse Rate						0.782
BFTZ	58 ± 10	61 ± 11 ^a	61 ± 11 ^b	62 ± 10 ^b	62 ± 10 ^b	
Amilorid	57 ± 10	61 ± 11 ^d	60 ± 10 ^d	61 ± 11 ^d	61 ± 11 ^d	
Placebo	55 ± 10	59 ± 12 ^a	59 ± 12 ^b	59 ± 12 ^b	59 ± 13 ^a	
<i>P</i> _{GLM between subjects}			0.712			
cSBP						NS
BFTZ	99 ± 7	98 ± 7				
Amilorid	96 ± 5	97 ± 7				
Placebo	98 ± 6	98 ± 8				
<i>P</i> _{ANOVA}	NS	NS				
cDBP						< 0.001
BFTZ	67 ± 5	63 ± 6				
Amilorid	65 ± 5	65 ± 6				
Placebo	65 ± 5	64 ± 5				
<i>P</i> _{ANOVA}	NS	NS				
PWV						0.055
BFTZ	5.5 ± 0.6	5.3 ± 0.4				
Amilorid	5.3 ± 0.5	5.3 ± 0.5				
Placebo	5.3 ± 0.7	5.3 ± 0.6				
<i>P</i> _{ANOVA}	NS	NS				
AI						0.034
BFTZ	-2.2 ± 14.6	-5.9 ± 17.7				
Amilorid	0.4 ± 12.5	-1.4 ± 13.4				
Placebo	-1.6 ± 14.6	-4.9 ± 18.6				
<i>P</i> _{ANOVA}	NS	NS				

Values are mean ± SD. General linear model (GLM) with repeated measures was performed for comparison within the group and intervention as between subjects factor for brachial systolic (bSBP), brachial diastolic blood pressure (bDBP) and pulse rate. Bonferroni correction was used for multiple comparisons between study-periods *vs* baseline. ^a*P* < 0.05; ^b*P* < 0.001; ^d*P* < 0.001; Paired *t*-test was used for comparison between post infusion *vs* baseline for central systolic (cSBP), central diastolic (cDBP), pulse wave velocity (PWV) and augmentation index (AIx).

while potassium secretion decreased.

During amiloride treatment there was a decrease in u-K at baseline. Amiloride exerts a direct effect on potassium excretion due to the blocking of ENaC. If the influx of sodium does not occur, there will be no lumen negative potential to drive potassium excretion^[48]. In response to 3% hypertonic saline however, the excretion of potassium increased most during amiloride. This phenomenon might have been due to prolonged effect of aldosterone to increase sodium reabsorption *via* ENaC and secretion *via* ROMK, despite current amiloride blockage.

Blood pressure

Thiazide decreases ECV and peripheral vascular resistance^[2,54]. Amiloride is an antihypertensive that exhibits its effects by significant natriuresis^[55]. In this study the lack of difference in 24 h ambulatory blood

pressure might partly be explained by the fact that the subjects were young and healthy with normal BP before entering the study. Moreover, BFTZ and amiloride are relatively weak antihypertensives and a very negligible blood pressure lowering effect was expected in these normohypertensive subjects.

Data showed that bDBP decreased significant in all three treatments, but bDBP decreased relatively more during BFTZ treatment compared to amiloride and placebo. As brachial office BP was also used to calibrate the SphygmoCor cDBP this may explain the reduction in cDBP during BFTZ. A negative augmentation index (AIx) has been reported in healthy young subjects, but is of limited use due to normal cardiovascular elasticity in this age group^[56].

Body fluid volumes

The determination of body fluid volumes *via* bioimpedance

spectroscopy (BIS) is an accurate method for estimating total body water and the distribution of water between the intracellular and extracellular spaces^[57]. In this present study, we measured no statistical difference between the groups, but as expected TBV was lower during both diuretic treatments compared to placebo, due to a decrease in ECV followed by sodium deficit. This reduction in ECV, during BFTZ treatment, is in agreement with current knowledge^[2]. We did not expect a major decrease in ECV after amiloride, being a weak diuretic agent^[55]. However the decrease in ECV was very similar to BFTZ. A significant difference in body fluid volumes between treatment groups was not detected in the present study, possibly due to the small number of subjects in each group.

Strengths and limitations

The major strength of this study was the design as a randomized, placebo controlled, double-blinded crossover study with a homogenous group of healthy young men and women. The test conditions were very well defined regarding diet, sodium and fluid intake. Thus, the results are not confounded by differences in sodium or water intake. However, as the study group was healthy humans the conclusions is limited to this population group and may not be extracted to patients with disturbances in water and sodium balance. Also the excretion of NCC was not measured, which would have provided us with even more information on renal handling of sodium.

Conclusion

In this study of healthy humans, amiloride and placebo clearly increased u-NKCC2, u-ENaC_γ and u-AQP2 in response to 3% hypertonic saline, while u-NKCC2 and u-AQP2 were unchanged during BFTZ. In contrast to amiloride, BFTZ treatment seemed to have changed glomerular-tubular balance, which caused the absence of a compensatory reabsorption of sodium *via* NKCC2 after hypertonic saline. It is possible that the lower p-AVP during BFTZ treatment resulted in a relatively less stimulation of NKCC2 and AQP2, with subsequent reduced transport of sodium and water *via* the transporters. During all three treatments, the increase in u-ENaC_γ might reflect a compensatory reabsorption to adjust for the decreased reabsorption of sodium in the proximal part of the nephron.

ACKNOWLEDGEMENTS

The authors greatly acknowledge the skilful assistance of our laboratory technicians: Inger-Merete Paulsen, Anne Mette Ravn, Kirsten Nygaard, Henriette Vorup Simonsen and Susan Milton Rasmussen.

COMMENTS

Background

The discovery that urine contains proteins from renal epithelia of proximal

tubule, Henle's loop, distal convoluted tubule and the collecting ducts has provided us with urinary biomarkers as a tool to investigate physiological and pathophysiological processes in renal sodium and water homeostasis.

Research frontiers

Urinary excretion of the aquaporin2 water channel (u-AQP2) is a biomarker that has been investigated in numerous studies of various water-balance disorders. It has also been demonstrated that the urinary excretion of epithelial sodium channels (u-ENaC) can be used as biomarkers of sodium transport *via* excretion of epithelial sodium channels (ENaC). However the exact physiological mechanisms are still unknown, and studies are needed to address the complete physiological handling of sodium and water in humans.

Innovations and breakthroughs

In animals, volume expansion and diuretics changes proximal water and sodium reabsorption and the expression of AQP2 and sodium transporters along the nephron. In addition, changes in transport activity of the sodium-potassium-2chloride cotransporter (NKCC2); ENaC and AQP2 may also be involved in the abnormal tubular function in patients with chronic kidney disease. However, it has never been studied to what extent the function of NKCC2, ENaC and AQP2 is simultaneously affected in response to amiloride and bendroflumethiazide (BFTZ) in humans. In the present study, the u-NKCC2 and u-AQP2 increased during amiloride and placebo, while u-NKCC2 and u-AQP2 remained unchanged during BFTZ, in response to infusion of 3% saline.

Applications

Thus, measurements of water- and sodium transporters in urine, as biomarkers of water-and sodium transport *via* NKCC2, ENaC and AQP2 may provide important information of the mechanisms involved in water and sodium balance in the kidney.

Terminology

AQP2, NKCC2 and ENaC are transporters in the nephron that play essential roles in regulating water and sodium homeostasis, extracellular volume and controlling blood pressure by reabsorbing water and sodium. BFTZ is a diuretic that inhibit the sodium-chloride co-transporter and amiloride is a diuretic that block the ENaC channels. These diuretics, which were developed empirically to treat patients with edema and hypertension, can be used as tools to characterize sodium transport pathways.

Peer-review

This is an interesting paper.

REFERENCES

- 1 **Esteva-Font C**, Ballarin J, Fernández-Llama P. Molecular biology of water and salt regulation in the kidney. *Cell Mol Life Sci* 2012; **69**: 683-695 [PMID: 21997386 DOI: 10.1007/s00018-011-0858-4]
- 2 **Duarte JD**, Cooper-DeHoff RM. Mechanisms for blood pressure lowering and metabolic effects of thiazide and thiazide-like diuretics. *Expert Rev Cardiovasc Ther* 2010; **8**: 793-802 [PMID: 20528637 DOI: 10.1586/erc.10.27]
- 3 **Kleyman TR**, Cragoe EJ. Amiloride and its analogs as tools in the study of ion transport. *J Membr Biol* 1988; **105**: 1-21 [PMID: 2852254]
- 4 **Nielsen S**, Agre P. The aquaporin family of water channels in kidney. *Kidney Int* 1995; **48**: 1057-1068 [PMID: 8569067]
- 5 **Kanno K**, Sasaki S, Hirata Y, Ishikawa S, Fushimi K, Nakanishi S, Bichet DG, Marumo F. Urinary excretion of aquaporin-2 in patients with diabetes insipidus. *N Engl J Med* 1995; **332**: 1540-1545 [PMID: 7537863 DOI: 10.1056/NEJM199506083322303]
- 6 **Rai T**, Sekine K, Kanno K, Hata K, Miura M, Mizushima A, Marumo F, Sasaki S. Urinary excretion of aquaporin-2 water channel protein in human and rat. *J Am Soc Nephrol* 1997; **8**: 1357-1362 [PMID: 9294826]
- 7 **Elliot S**, Goldsmith P, Knepper M, Haughey M, Olson B. Urinary excretion of aquaporin-2 in humans: a potential marker of collecting duct responsiveness to vasopressin. *J Am Soc Nephrol* 1996; **7**: 403-409 [PMID: 8704105]
- 8 **Saito T**, Ishikawa SE, Sasaki S, Nakamura T, Rokkaku K, Kawakami A, Honda K, Marumo F, Saito T. Urinary excretion of aquaporin-2 in the diagnosis of central diabetes insipidus. *J Clin Endocrinol Metab* 1997; **82**: 1823-1827 [PMID: 9177390]
- 9 **Pedersen RS**, Bentzen H, Bech JN, Pedersen EB. Effect of water

- deprivation and hypertonic saline infusion on urinary AQP2 excretion in healthy humans. *Am J Physiol Renal Physiol* 2001; **280**: F860-F867 [PMID: 11292629]
- 10 **Lauridsen TG**, Vase H, Starklint J, Graffe CC, Bech JN, Nielsen S, Pedersen EB. Increased renal sodium absorption by inhibition of prostaglandin synthesis during fasting in healthy man. A possible role of the epithelial sodium channels. *BMC Nephrol* 2010; **11**: 28 [PMID: 21029429 DOI: 10.1186/1471-2369-11-28]
 - 11 **Jensen JM**, Mose FH, Bech JN, Nielsen S, Pedersen EB. Effect of volume expansion with hypertonic- and isotonic saline and isotonic glucose on sodium and water transport in the principal cells in the kidney. *BMC Nephrol* 2013; **14**: 202 [PMID: 24067081 DOI: 10.1186/1471-2369-14-202]
 - 12 **Jensen JM**, Mose FH, Kulik AE, Bech JN, Fenton RA, Pedersen EB. Abnormal urinary excretion of NKCC2 and AQP2 in response to hypertonic saline in chronic kidney disease: an intervention study in patients with chronic kidney disease and healthy controls. *BMC Nephrol* 2014; **15**: 101 [PMID: 24970686 DOI: 10.1186/1471-2369-15-101]
 - 13 **Pedersen EB**, Eiskjaer H, Madsen B, Danielsen H, Egeblad M, Nielsen CB. Effect of captopril on renal extraction of renin, angiotensin II, atrial natriuretic peptide and vasopressin, and renal vein renin ratio in patients with arterial hypertension and unilateral renal artery disease. *Nephrol Dial Transplant* 1993; **8**: 1064-1070 [PMID: 8272217]
 - 14 **Graffe CC**, Bech JN, Pedersen EB. Effect of high and low sodium intake on urinary aquaporin-2 excretion in healthy humans. *Am J Physiol Renal Physiol* 2012; **302**: F264-F275 [PMID: 21993890 DOI: 10.1152/ajprenal.00442.2010]
 - 15 **Lauridsen TG**, Vase H, Starklint J, Bech JN, Pedersen EB. Protein-enriched diet increases water absorption via the aquaporin-2 water channels in healthy humans. *Nephrol Dial Transplant* 2010; **25**: 2502-2510 [PMID: 20237060 DOI: 10.1093/ndt/gfq111]
 - 16 **Matthesen SK**, Larsen T, Vase H, Lauridsen TG, Jensen JM, Pedersen EB. Effect of amiloride and spironolactone on renal tubular function and central blood pressure in patients with arterial hypertension during baseline conditions and after furosemide: a double-blinded, randomized, placebo-controlled crossover trial. *Clin Exp Hypertens* 2013; **35**: 313-324 [PMID: 22966789 DOI: 10.3109/10641963.2012.721843]
 - 17 **Hager H**, Kwon TH, Vinnikova AK, Masilamani S, Brooks HL, Frøkjaer J, Knepper MA, Nielsen S. Immunocytochemical and immunoelectron microscopic localization of alpha-, beta-, and gamma-ENaC in rat kidney. *Am J Physiol Renal Physiol* 2001; **280**: F1093-F1106 [PMID: 11352848]
 - 18 **Matthesen SK**, Larsen T, Vase H, Lauridsen TG, Pedersen EB. Effect of potassium supplementation on renal tubular function, ambulatory blood pressure and pulse wave velocity in healthy humans. *Scand J Clin Lab Invest* 2012; **72**: 78-86 [PMID: 22149452 DOI: 10.3109/00365513.2011.635216]
 - 19 **Walter SJ**, Shirley DG. The effect of chronic hydrochlorothiazide administration on renal function in the rat. *Clin Sci (Lond)* 1986; **70**: 379-387 [PMID: 3698514]
 - 20 **Hunter RW**, Craigie E, Homer NZ, Mullins JJ, Bailey MA. Acute inhibition of NCC does not activate distal electrogenic Na⁺ reabsorption or kaliuresis. *Am J Physiol Renal Physiol* 2014; **306**: F457-F467 [PMID: 24402096 DOI: 10.1152/ajprenal.00339.2013]
 - 21 **Dirks JH**, Cirksema WJ, Berliner RW. The effects of saline infusion on sodium reabsorption by the proximal tubule of the dog. *J Clin Invest* 1965; **44**: 1160-1170 [PMID: 14328393 DOI: 10.1172/JCI105223]
 - 22 **Svensén CH**, Waldrop KS, Edsberg L, Hahn RG. Natriuresis and the extracellular volume expansion by hypertonic saline. *J Surg Res* 2003; **113**: 6-12 [PMID: 12943804]
 - 23 **Häberle DA**, von Baeyer H. Characteristics of glomerulotubular balance. *Am J Physiol* 1983; **244**: F355-F366 [PMID: 6340526]
 - 24 **Tian Y**, Riaz S, Khan O, Klein JD, Sugimura Y, Verbalis JG, Ecelbarger CA. Renal ENaC subunit, Na-K-2Cl and Na-Cl cotransporter abundances in aged, water-restricted F344 x Brown Norway rats. *Kidney Int* 2006; **69**: 304-312 [PMID: 16408120 DOI: 10.1038/sj.ki.5000076]
 - 25 **Rieg T**, Tang T, Uchida S, Hammond HK, Fenton RA, Vallon V. Adenylyl cyclase 6 enhances NKCC2 expression and mediates vasopressin-induced phosphorylation of NKCC2 and NCC. *Am J Pathol* 2013; **182**: 96-106 [PMID: 23123217 DOI: 10.1016/j.ajpath.2012.09.014]
 - 26 **Giménez I**, Forbush B. Short-term stimulation of the renal Na-K-Cl cotransporter (NKCC2) by vasopressin involves phosphorylation and membrane translocation of the protein. *J Biol Chem* 2003; **278**: 26946-26951 [PMID: 12732642 DOI: 10.1074/jbc.M303435200]
 - 27 **Edinger RS**, Bertrand CA, Rondandino C, Apodaca GA, Johnson JP, Butterworth MB. The epithelial sodium channel (ENaC) establishes a trafficking vesicle pool responsible for its regulation. *PLoS One* 2012; **7**: e46593 [PMID: 23029554 DOI: 10.1371/journal.pone.0046593]
 - 28 **Löffing J**, Korbmacher C. Regulated sodium transport in the renal connecting tubule (CNT) via the epithelial sodium channel (ENaC). *Pflugers Arch* 2009; **458**: 111-135 [PMID: 19277701 DOI: 10.1007/s00424-009-0656-0]
 - 29 **Garty H**, Palmer LG. Epithelial sodium channels: function, structure, and regulation. *Physiol Rev* 1997; **77**: 359-396 [PMID: 9114818]
 - 30 **Masilamani S**, Kim GH, Mitchell C, Wade JB, Knepper MA. Aldosterone-mediated regulation of ENaC alpha, beta, and gamma subunit proteins in rat kidney. *J Clin Invest* 1999; **104**: R19-R23 [PMID: 10510339 DOI: 10.1172/JCI7840]
 - 31 **Bankir L**, Fernandes S, Bardoux P, Bouby N, Bichet DG. Vasopressin-V2 receptor stimulation reduces sodium excretion in healthy humans. *J Am Soc Nephrol* 2005; **16**: 1920-1928 [PMID: 15888562 DOI: 10.1681/ASN.2004121079]
 - 32 **Ecelbarger CA**, Kim GH, Wade JB, Knepper MA. Regulation of the abundance of renal sodium transporters and channels by vasopressin. *Exp Neurol* 2001; **171**: 227-234 [PMID: 11573975 DOI: 10.1006/exnr.2001.7775]
 - 33 **Bugaj V**, Pochynyuk O, Stockand JD. Activation of the epithelial Na⁺ channel in the collecting duct by vasopressin contributes to water reabsorption. *Am J Physiol Renal Physiol* 2009; **297**: F1411-F1418 [PMID: 19692483 DOI: 10.1152/ajprenal.00371.2009]
 - 34 **Ecelbarger CA**, Kim GH, Terris J, Masilamani S, Mitchell C, Reyes I, Verbalis JG, Knepper MA. Vasopressin-mediated regulation of epithelial sodium channel abundance in rat kidney. *Am J Physiol Renal Physiol* 2000; **279**: F46-F53 [PMID: 10894786]
 - 35 **Perucca J**, Bichet DG, Bardoux P, Bouby N, Bankir L. Sodium excretion in response to vasopressin and selective vasopressin receptor antagonists. *J Am Soc Nephrol* 2008; **19**: 1721-1731 [PMID: 18596120 DOI: 10.1681/ASN.2008010021]
 - 36 **Stockand JD**. Vasopressin regulation of renal sodium excretion. *Kidney Int* 2010; **78**: 849-856 [PMID: 20736986 DOI: 10.1038/ki.2010.276]
 - 37 **Patel AB**, Frindt G, Palmer LG. Feedback inhibition of ENaC during acute sodium loading in vivo. *Am J Physiol Renal Physiol* 2013; **304**: F222-F232 [PMID: 23171553 DOI: 10.1152/ajprenal.00596.2012]
 - 38 **Frindt G**, Silver RB, Windhager EE, Palmer LG. Feedback regulation of Na channels in rat CCT. II. Effects of inhibition of Na entry. *Am J Physiol* 1993; **264**: F565-F574 [PMID: 8384418]
 - 39 **Andersen LJ**, Jensen TU, Bestle MH, Bie P. Isotonic and hypertonic sodium loading in supine humans. *Acta Physiol Scand* 1999; **166**: 23-30 [PMID: 10372975]
 - 40 **Pácha J**, Frindt G, Antonian L, Silver RB, Palmer LG. Regulation of Na channels of the rat cortical collecting tubule by aldosterone. *J Gen Physiol* 1993; **102**: 25-42 [PMID: 8397276]
 - 41 **Butterworth MB**. Regulation of the epithelial sodium channel (ENaC) by membrane trafficking. *Biochim Biophys Acta* 2010; **1802**: 1166-1177 [PMID: 20347969 DOI: 10.1016/j.bbdis.2010.03.010]
 - 42 **DiGiovanni SR**, Nielsen S, Christensen EI, Knepper MA. Regulation of collecting duct water channel expression by vasopressin in Brattleboro rat. *Proc Natl Acad Sci USA* 1994; **91**: 8984-8988 [PMID: 7522327]
 - 43 **Nielsen S**, Chou CL, Marples D, Christensen EI, Kishore BK, Knepper MA. Vasopressin increases water permeability of kidney

- collecting duct by inducing translocation of aquaporin-CD water channels to plasma membrane. *Proc Natl Acad Sci USA* 1995; **92**: 1013-1017 [PMID: 7532304]
- 44 **Wen H**, Frokiaer J, Kwon TH, Nielsen S. Urinary excretion of aquaporin-2 in rat is mediated by a vasopressin-dependent apical pathway. *J Am Soc Nephrol* 1999; **10**: 1416-1429 [PMID: 10405197]
- 45 **Pisitkun T**, Shen RF, Knepper MA. Identification and proteomic profiling of exosomes in human urine. *Proc Natl Acad Sci USA* 2004; **101**: 13368-13373 [PMID: 15326289 DOI: 10.1073/pnas.0403453101]
- 46 **Pedersen RS**, Bentzen H, Bech JN, Nyvad O, Pedersen EB. Urinary aquaporin-2 in healthy humans and patients with liver cirrhosis and chronic heart failure during baseline conditions and after acute water load. *Kidney Int* 2003; **63**: 1417-1425 [PMID: 12631357 DOI: 10.1046/j.1523-1755.2003.00858.x]
- 47 **Starklint J**, Bech JN, Pedersen EB. Down-regulation of urinary AQP2 and unaffected response to hypertonic saline after 24 hours of fasting in humans. *Kidney Int* 2005; **67**: 1010-1018 [PMID: 15698439 DOI: 10.1111/j.1523-1755.2005.00164.x]
- 48 **Castañeda-Bueno M**, Arroyo JP, Gamba G. Independent regulation of Na⁺ and K⁺ balance by the kidney. *Med Princ Pract* 2012; **21**: 101-114 [PMID: 22042004 DOI: 10.1159/000332580]
- 49 **Arroyo JP**, Ronzaud C, Lagnaz D, Staub O, Gamba G. Aldosterone paradox: differential regulation of ion transport in distal nephron. *Physiology* (Bethesda) 2011; **26**: 115-123 [PMID: 21487030 DOI: 10.1152/physiol.00049.2010]
- 50 **Hoorn EJ**, Ellison DH. WNK kinases and the kidney. *Exp Cell Res* 2012; **318**: 1020-1026 [PMID: 22405999 DOI: 10.1016/j.yexcr.2012.02.029]
- 51 **Ellison DH**, Loffing J. Thiazide effects and adverse effects: insights from molecular genetics. *Hypertension* 2009; **54**: 196-202 [PMID: 19564550 DOI: 10.1161/HYPERTENSIONAHA.109.129171]
- 52 **Wei Y**, Zamilowicz B, Satlin LM, Wang WH. Angiotensin II inhibits the ROMK-like small conductance K channel in renal cortical collecting duct during dietary potassium restriction. *J Biol Chem* 2007; **282**: 6455-6462 [PMID: 17194699 DOI: 10.1074/jbc.M607477200]
- 53 **Yue P**, Sun P, Lin DH, Pan C, Xing W, Wang W. Angiotensin II diminishes the effect of SGK1 on the WNK4-mediated inhibition of ROMK1 channels. *Kidney Int* 2011; **79**: 423-431 [PMID: 20927043 DOI: 10.1038/ki.2010.380]
- 54 **Jessup JA**, Brosnihan KB, Gallagher PE, Chappell MC, Ferrario CM. Differential effect of low dose thiazides on the Renin Angiotensin system in genetically hypertensive and normotensive rats. *J Am Soc Hypertens* 2008; **2**: 106-115 [PMID: 19343087 DOI: 10.1016/j.jash.2007.10.005]
- 55 **Bull MB**, Laragh JH. Amiloride. A potassium-sparing natriuretic agent. *Circulation* 1968; **37**: 45-53 [PMID: 5634728]
- 56 **Hughes AD**, Park C, Davies J, Francis D, McG Thom SA, Mayet J, Parker KH. Limitations of augmentation index in the assessment of wave reflection in normotensive healthy individuals. *PLoS One* 2013; **8**: e59371 [PMID: 23544061 DOI: 10.1371/journal.pone.0059371]
- 57 **Moissl UM**, Wabel P, Chamney PW, Bosaeus I, Levin NW, Bosy-Westphal A, Korth O, Müller MJ, Ellegård L, Malmros V, Kaitwatcharachai C, Kuhlmann MK, Zhu F, Fuller NJ. Body fluid volume determination via body composition spectroscopy in health and disease. *Physiol Meas* 2006; **27**: 921-933 [PMID: 16868355 DOI: 10.1088/0967-3334/27/9/012]

P- Reviewer: McNally RJQ, Su M S- Editor: Ji FF

L- Editor: A E- Editor: Yan JL





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

