

Markers of bile duct tumors

Giulia Malaguarnera, Maria Giordano, Isabella Paladina, Alessandra Rando, Mario Uccello, Francesco Basile, Antonio Biondi, Santo Carnazzo, Innocenza Alessandria, Clorinda Mazzarino

Giulia Malaguarnera, Clorinda Mazzarino, Department of Bio-medical Science, University of Catania, via Androne 83, 95124 Catania, Italy

Maria Giordano, Isabella Paladina, Alessandra Rando, Mario Uccello, Departments of Senescence, Urological, and Neurological Sciences, University of Catania, via Messina 829, 95126 Catania, Italy

Francesco Basile, Antonio Biondi, Department of General Surgery, University Medical School of Catania, Hospital Vittorio Emanuele, via Plebiscito 628, 95124 Catania, Italy

Santo Carnazzo, Department of General Surgery, University Medical School of Catania, Policlinico via Santa Sofia 86, 95126 Catania, Italy

Innocenza Alessandria, Department of Internal Medicine, University Medical School of Catania, Hospital Cannizzaro, via Messina 829, 95126 Catania, Italy

Author contributions: Malaguarnera G, Giordano M, Paladina I, Rando A, Uccello M and Alessandria I evaluated the importance of serum markers in the diagnosis and the follow up of bile duct tumors and their usefulness in the choice of a correct therapeutic approach; Basile F, Biondi A and Carnazzo S noticed the importance of serum markers to establish a prognosis and evaluate the possibility of a surgical approach; Mazzarino C coordinated the work.

Correspondence to: Maria Giordano, MD, Departments of Senescence, Urological, and Neurological Sciences, University of Catania, via Messina 829, 95126 Catania, Italy. maria-gior@hotmail.it

Telephone: +39-95-7262008 Fax: +39-95-7262011

Received: August 6, 2010 Revised: February 23, 2011

Accepted: March 2, 2011

Published online: April 15, 2011

Abstract

Biliary tract carcinomas are relatively rare, representing less than 1% of cancers. However, their incidence has increased in Japan and in industrialized countries like the USA. Biliary tract tumors have a poor prognosis and a high mortality rate because they are usually detected late in the course of the disease; therapeutic treatment options are often limited and of minimal utility. Recent studies have shown the importance of serum and molecular

markers in the diagnosis and follow up of biliary tract tumors. This review aims to introduce the main features of the most important serum and molecular markers of biliary tree tumors. Some considerable tumor markers are cancer antigen 125, carbohydrate antigen 19-9, carcinoembryonic antigen, chromogranin A, mucin 1, mucin 5, alpha-fetoprotein, claudins and cytokeratins.

© 2011 Baishideng. All rights reserved.

Key words: Bile duct tumors; Cholangiocarcinoma; Tumor markers; Carbohydrate antigen 19-9; Chromogranin A

Peer reviewer: Sunil Krishnan, MD, Assistant Professor, Anderson Cancer Center, 1515 Holcombe Blvd, Houston, TX 77030, United States

Malaguarnera G, Giordano M, Paladina I, Rando A, Uccello M, Basile F, Biondi A, Carnazzo S, Alessandria I, Mazzarino C. Markers of bile duct tumors. *World J Gastrointest Oncol* 2011; 3(4): 49-59 Available from: URL: <http://www.wjgnet.com/1948-5204/full/v3/i4/49.htm> DOI: <http://dx.doi.org/10.4251/wjgo.v3.i4.49>

INTRODUCTION

Biliary tract carcinomas are relatively rare, representing less than 1% of cancers^[1]. However, an increase in their incidence and mortality in Japan and industrialized countries like the USA has been noted^[2,3].

Gallbladder adenocarcinoma and cholangiocarcinoma (CCA) account for 4% and 3% of all gastrointestinal cancers respectively^[4,5]. These malignancies are highly fatal with 1- and 2-year survival rates of 25% and 13% respectively^[6]. CCAs can be anatomically classified into intrahepatic (including hilar) or extrahepatic tumors. 60%-70% of bile duct tumors arise in the bifurcation of the hepatic ducts (Klatskin tumors), 20%-30% in the distal common bile duct while 5%-10% of CCAs arise within the intrahepatic ducts of the liver parenchyma^[7].

Intrahepatic cholangiocarcinomas (HI-CCAs) originate in the small bile ducts and tend to be grouped with hepatocellular carcinoma (HCC) as primary liver tumors^[8,9].

In general, most malignant bile duct tumors present with painless obstructive symptoms which include pale stools, dark urine and jaundice. Increased symptoms in the course of the pathology such as right upper quadrant abdominal pain, fever and rigors are indicative of superimposed cholangitis^[10].

The diagnosis of biliary tract disease is realized by ultrasonography (US), computed tomography (CT) or magnetic resonance imaging (MRI) of the liver and magnetic resonance cholangiopancreatography (MRCP) or endoscopic retrograde cholangiopancreatography (ERCP). Intrahepatic CCA still remains an unfortunate liver tumor with a high mortality rate as many patients diagnosed with CCA cannot be offered a curative treatment^[11]. Recent studies have also shown the utility of serum and molecular markers in the diagnosis and follow up of biliary tract tumors. Since these cancers are usually detected late in the course of the disease, treatment options are often limited and of minimal utility. Thus, research efforts have concentrated on identifying potential markers of neoplasia that can be incorporated into diagnostic tests and therapeutic modalities for use in individuals at risk for these lethal malignancies. This review aims to introduce the main features of the most important serum and molecular markers of biliary tree tumors.

CANCER ANTIGEN 125

Cancer antigen 125 (CA 125) is elevated in 58% of patients with gallbladder cancer and in 40%-50% of cholangiocarcinoma patients^[12] but it has a low specificity; in fact, its serum levels can increase in other gastrointestinal or gynaecological malignancies and several cholangiopathies^[13]. It tends to increase in patients with ascites and is considered a possible indicator of peritoneal involvement.

CA 125 is more specific than carbohydrate antigen 19-9 (CA 19-9) in detecting CCA and in distinguishing between benign and malignant causes of bile duct obstruction. It is not easily influenced by bile duct inflammation or calculi while CA 19-9 is increased in cases of cholangitis and hepatolithiasis.

Carbohydrate antigen 19-9

CA 19-9 is a glycolipid synthesized by the pancreatic, biliary, gastric, colonic cells, endometrial and salivary epithelia. CA 19-9 is used as a screening tool for CCA in patients with primary sclerosing cholangitis (PSC). Elevated concentrations of serum CA 19-9 in bile duct cancers have been frequently reported: a rate of 97% was found in CCA patients by Torzilli *et al*^[14], Hultcrantz *et al*^[15] reported a rate of 76% and Caturelli *et al*^[16] a rate of 68%. Nichols *et al*^[17] showed that a serum CA 19-9 value greater than 100 U/mL predicted the presence of CCA in patients with a sensitivity of 89% and a specificity of 86%. CA 19-9 has an increasing role in the differential diagnosis of benign and

malignant hepatobiliary and pancreatic conditions. Benign conditions of the hepatobiliary and pancreatic tract, such as Mirizzi syndrome, autoimmune pancreatitis, benign biliary stenosis secondary to PSC and pancreatic exocrine dysfunction, have been associated with abnormal elevation of CA 19-9^[18-21]. If bile flow is blocked by biliary obstruction in benign conditions such as choledocolitiasis, epithelial cells will be impaired by inflammation and will proliferate at the same time^[22]. As a result, more CA 19-9 may be released into the bloodstream. In such conditions, the increase of CA 19-9 is associated with hyperbilirubinemia secondary to biliary obstruction, and subsequent values of CA 19-9 tends to normalize after the restoration of biliary drainage^[20,21]. The increase of CA19-9 is also useful to predict resectability of CCA^[23,24]. A marked elevation of serum CA 19-9 is associated with advanced and unresectable biliary cancers. Elevated levels of CA 19-9 are correlated with advanced disease and poor prognosis and high preoperative values correlate with poor survival.

Carcinoembryonic antigen

Carcinoembryonic antigen (CEA) is a highly glycosylated cell surface glycoprotein with a molecular weight of 180000 and is considered a colon-specific oncofetal protein^[25]. CEA is often found in patients with malignant tumors of the digestive system such stomach, colon, biliary tract and pancreas cancer. Qin *et al*^[26] reported a significant increase of CEA serum levels in CCA patients. In their study, the sensitivity and specificity rates of serum CEA were 68.57% and 81.52% respectively. Similar results (63.3% and 78.4%) were shown by Ramage *et al*^[27]. Various studies have found a higher sensitivity of CA 19-9 in comparison with CEA values in diagnosis of CCA but the combination of these two markers increases the sensitivity and specificity. The following formula is suggested as a screening test for CCA patients with PSC: CA 19-9 + (CEA × 40). An index greater than 400 U/mL has a positive predictive value of 100%, a specificity of 100% and a sensitivity of 67%. Recently, Chalasani *et al*^[28] demonstrated that an index greater than 400 U/mL was no better than a 100 U/mL one in predicting the presence of CCA.

CHROMOGRANIN A

Chromogranin A (CgA) is an acidic glycoprotein contained in the secretory granules of neuroendocrine (NE) cells^[29]. It is widely used as an immunohistochemical marker of neuroendocrine tumors (NETs). It may also serve as a serum marker and is co-secreted with the amines and peptides of the neurosecretory granules^[30]. It is a very sensitive, specific marker of NETs. The most common site of origin for carcinoid tumors is the gastrointestinal system, followed by the bronchopulmonary system (25%). Apart from those arising from the ampulla of Vater, intrahepatic bile duct, gallbladder and cystic duct, carcinoid tumors of the extrahepatic bile duct are extremely rare. Primary carcinoid tumors may arise from endocrine cells of the body and fundus of gallbladder or from pre-existing endocrine cells present in the

mucus membrane of the gallbladder neck.

Elevated serum levels of CgA are also reported in patients with colon, lung, breast, liver and prostate cancers^[51-55] and are related to a tumor NE differentiation which has a prognostic importance in several malignancies^[36]. CgA is also considered a significantly independent, negative prognostic marker for CCA patients^[37].

MUCIN-1 AND MUCIN-5AC

Mucins are heavily O-glycosylated proteins, mainly expressed by ductal and glandular epithelial tissues. The human mucin family is divided into secreted and transmembrane forms. The secreted forms are mucin-2 (MUC2), mucin-5AC (MUC5AC), mucin-5AB (MUC5AB) and mucin-6 (MUC6) that are released from the apical membrane in order to form a physical barrier that protects the epithelial layer from adverse conditions such as exposure to commensally bacteria, ingested toxins and reactive oxygen species (ROS). Among the secreted mucins, MUC2 is the most correlated with inflammation and cancer. MUC2 suppresses inflammation and inhibits the development of intestinal tumors^[38,39]. A recent study has shown that the loss of MUC2 expression in mice is associated with an augmented proliferation and survival of intestinal epithelial cells responsible for an increased exposure to luminal contents and induction of inflammation^[40]. Transmembrane mucins include mucin-1 (MUC1), mucin-4 (MUC4), mucin-13 (MUC13) and mucin-16 (MUC16) which also contribute to forming physiological barriers and transmitting growth and survival signal to the cells. Mucin genes are expressed in several cells and tissues: in particular, specific MUC2 and MUC3 are expressed in bowel^[41] while MUC5AC and MUC6 in gastric tissues^[42]. Alterations of quantity and quality of mucins occur in cancer tissues, like in the case of CCA^[43,44]. Various degrees of MUC1 glycosylation between normal and tumor cells have been demonstrated in several tumors.

MUC1 is a transmembrane glycoprotein frequently found in the developing intrahepatic bile ducts of fetal liver^[45] but not in the normal adult intrahepatic biliary tract^[46,47]. For this reason, it is an oncofetal antigen in the intrahepatic biliary tree^[45]. An increased expression of MUC1 has often been demonstrated in many tumors, including CCA^[48,49], and its over expression is correlated with poor prognosis and unfavourable survival. High MUC1 immunoreactivity is associated with vascular invasion and malignant progression in three different ways. Firstly, tumor cells expressing high levels of MUC1 can repel each other^[50] favoring the development of metastases. Secondly, vascular invasion involves the binding between ligand (E-selectin) of endothelial cells and epitopes (sialyl Lewis) on tumor cells^[51,52]. In fact sialyl Lewis, a carbohydrate epitope which is present at high levels in tumor cells and serum of CCA patients, has been identified as an epitope of MUC1^[53]. Thirdly, high MUC1 levels on the membrane of tumor cells suppress the immunity of patients, inhibiting the interaction between cytotoxic lymphocytes

and tumor cells^[54]. The expression of MUC1 can help the development of metastases, particularly in the liver. Therefore, it is often difficult to distinguish a metastatic nodule from HCC. Xu *et al.*^[55] noticed that KL-6 mucin, a type of MUC1 mucin, is expressed in intrahepatic CCA but not in HCC, demonstrating its usefulness in differential diagnosis. Sasaki *et al.*^[56] have found that the expression of MUC1 apomucin in small bile ducts is correlated with biliary epithelial damage and the consecutive immune-mediated processes. In fact, cytotoxic T cells recognize MUC1, preferentially expressed on epithelial tumors.

While MUC1 expression reflects histological differentiation, increased proliferative activity and invasiveness, MUC2 expression is correlated with a decrease in proliferative activity.

MUC5AC is a secreted gel-forming mucin aberrantly expressed in CCA tissues; indeed, it is considered an excellent biomarker for CCA. High MUC5AC expression is correlated with tumor size and metastases which are responsible for the poor outcome of CCA patients who have positive serum MUC5AC status. Moreover, increased MUC5AC expression is related to neural invasion.

In conclusion, the expression of MUC1 and MUC5AC mucins is associated with metastasis and is considered a useful prognostic marker for a poor outcome in CCA patients.

PROMYELOCYTIC LEUKEMIA, P53 AND DPC4

Promyelocytic leukemia (PML) is a tumor suppressor gene implicated in the pathogenesis of several malignancies like leukaemia and plays a role in the regulation of apoptosis, cell growth and DNA repair^[57]. It is considered a novel molecular cancer marker. PML protein expression is reduced in many tumors such as prostate, colon, breast, lung carcinomas and lymphomas. PML has a prognostic significance in human gallbladder cancer (GBC). In this case, the loss of PML expression is related to poor prognosis, metastatic lymphatic invasion and stage of GBC, suggesting that the protein may be involved in GBC progression. Patients with normal PML and p53 expression have a better prognosis than those with abnormal expression of PML and/or p53^[58].

Many studies have shown a relationship between p53 mutations and GBC, reporting that the rate of p53 over expression varied from 30.6% to 72.6%^[59-61]. P53 mutations are correlated to histological type, tumor grade and survival time^[62]. Various studies suggest that biliary tract carcinomas have a different frequency of p53 immunoreactivity depending on their site of origin (gallbladder, intrahepatic, proximal and distal common bile duct), perhaps reflecting different pathogenic factors in their respective etiologies^[62-64].

DPC4 is another tumor suppressor gene localized on the long arm of chromosome 18. It is inactivated in 55% of pancreatic adenocarcinomas and in bile duct carcinomas. Compared with their more proximal (intrahepatic

and perihilar) counterparts, distal common bile duct carcinomas show a higher frequency of loss of DPC4 gene expression and of p53 over expression.

Ki67

Ki67 antigen is exclusively expressed in proliferating cells (in the G1, S, G2 and M phases) and not in quiescent cells (G0 phase). Ki67 is considered a useful marker for clinical use because of its reliability^[65]. High Ki67 serum values are associated with poor prognosis in some tumors such as breast and oesophageal cancers and lymphomas^[66].

Biliary tract cancer patients with higher Ki67 index tend to have a low survival rate since such high levels lead to an increased speed in the growth of tumor cells.

Transmembrane carbonic anhydrase isoenzyme IX

Carbonic anhydrase (CA) plays a crucial role in various physiological processes. It catalyses the interconversion of carbon dioxide in bicarbonate. Eleven isoenzymes, differing in their tissue distribution and enzymatic activity, have been identified in mammals^[67-70] and several isoenzymes have found in the hepatobiliary tract. Biliary epithelial cells express cytoplasmic carbonic anhydrase II (CA II), an apical membrane-associated carbonic anhydrase IV (CA IV) and a basolateral transmembrane carbonic anhydrase isoenzyme IX (MN/CA IX)^[71,72], while in hepatocytes the most frequent isoenzyme is the mitochondrial carbonic anhydrase VA (CA VA)^[73,74].

The expression of MN/CA IX is confined to the basolateral surfaces of normal biliary epithelial cells, whereas no positive reaction is found in hepatocytes.

Most of biliary epithelial tumors express MN/CA IX while only few HCCs show positive immunoreactions, suggesting that MN/CA IX can be considered a useful biomarker for the differential diagnosis of hepatobiliary neoplasms.

MATRIX METALLOPROTEINASES

Tumors invade the basement membrane through the secretion of enzymes that digest the extracellular matrix (ECM) proteins and allow angiogenesis. These enzymes are matrix metalloproteinases (MMPs). They belong to family of zinc-dependent endopeptidases, responsible for the degradation of all components of the ECM^[75]. MMPs-related factors also increase the proliferation of tumor cells promoting the mitosis. MMPs activity is balanced by tissue inhibitors of metalloproteinases (TIMPs)^[76,77]. In particular, matrix metalloproteinase 2 (MMP-2) and matrix metalloproteinase 9 (MMP-9) focalize their action on collagen IV, the major component of the basement membrane^[76-78]. *In vitro* studies have shown that microvascular endothelial cells do not constitutively secrete MMP-9; however, when exposed to an angiogenic stimulus, like tumor necrosis factor- α (TNF- α), MMP-9 production is up-regulated^[79]. Furthermore, MMP-9 plays a fundamental role in the catalytic activity of tumor cell invasion, metastasis and in the regulation of angiogenesis^[80,81].

It seems that the up-regulation of MMP-9 is linked with cyclooxygenase-2 (COX-2) expression which is induced by TNF- α and increased in all inflammatory states, thus in an organism affected by cancer^[82]. Recent studies have noticed that MMPs and cyclooxygenases (COXs) are over expressed in CCA cells. However, Leelawat *et al*^[83] did not show significant differences in MMP-9 levels between CCA patients and the controls, as in the case of serum matrix metalloproteinase 7 (MMP-7) values. In fact, the presence of higher serum MMP-7 levels in CCA patients than those with benign biliary tract disease was demonstrated.

MMP-7 is expressed by epithelial cells^[84]. Therefore, it cannot be considered as a specific marker of bile duct tumors but its expression in CCA is an unfavorable post-operative prognostic factor^[85].

Another important member of this family is represented by MMP-2. Kirimlioğlu *et al*^[86] analyzed the role of MMPs in every type of biliary tract cancer and reported the presence of MMP-2 expression in 75% of the distal part of the biliary ducts, and also GBC, distal CCA and ampullary carcinoma expressed MMP-2 in 30%, 37% and 40% of the cases respectively. They also showed that MMP-2 and MMP-9 levels were higher in subjects with neural invasion although they demonstrated no correlation between the expression of MMPs and tumor differentiation and the presence of metastasis.

Erb-B

The Erb-B family consists of four distinct receptors: ErbB1 (EGFR), ErbB2, ErbB3 and ErbB4. All these receptors are composed of three parts: an extracellular ligand-binding domain, a transmembrane lipophilic domain and a conserved cytoplasmic tyrosine kinase domain^[87,88].

In particular, C-erb-B2 is an oncogene situated on chromosome 17 and is also known as neu or HER-2^[89,90]. The protein expressed by this gene serves as receptor of the epidermal growth factor (EGF) and so it plays an important role in the angiogenesis. EGFR is involved in different human cancers such as breast, ovarian, skin, kidney, pancreas, lung, salivary glands and digestive tract tumors^[91]. Zheng *et al*^[92] also showed an increase of C-erb-B2 expression in extrahepatic cholangiocarcinoma (EH-CCA). Particularly, a significant difference of the expression of Erb-B in correlation with the grade and differentiation of the tumor has been found. The comparison between tumor grading III-IV and I and the highly and less differentiated tumors revealed a significant difference, suggesting that C-erb-B-2 could be involved in the processes of development, invasion of tumor and metastasis.

ALPHA-FETOPROTEIN

Alpha-fetoprotein (AFP) is a fetal glycoprotein with a molecular weight of about 72 kDa. It was first described in human fetus in 1956 and successfully assumed a key role in the diagnosis and follow up of HCC^[93,94]. Under physiological conditions, it is synthesized by fetal hepato-

cytes, yolk sac cells and gastrointestinal cells. Some days after birth, AFP serum levels begin to decrease until they gradually get to a level lower than 10 ng/mL. The development of radioimmunoassay for AFP has increased its sensibility^[95-100] and nowadays it is considered a useful marker for embryonal cell carcinoma and liver diseases. However, some studies have demonstrated the possible use of this marker in the primary neoplasm of the gastrointestinal tract. McIntire *et al*^[101] showed an increase of AFP values in patients with pancreas, biliary tract and stomach carcinomas in comparison with those affected by colon, esophagus and small bowel carcinomas. AFP is not only an indicator of cell de-differentiation but also an important sign of hepatic stem cells^[102]. Jalanko *et al*^[103] reported a slight increase of serum AFP concentrations only in a small percent of patients.

AFP is the main serum biomarker of HCC but its variant, lectin-reactive AFP (AFP-L3), has been demonstrated to be useful in the diagnosis of intrahepatic CCA. Okuda *et al*^[104] noticed that AFP-positive patients presented HCC features which were very different from those of classical intrahepatic cholangiocarcinoma (IH-CCA) whose patient were seropositive to CA 19-9. They supposed that the IH-CCA seropositive for AFP-L3, HCC and CCA might have originated from hepatic precursor cells with the characteristic of both hepatocytes and bile duct cells.

N-CADHERIN

E and N-cadherins are expressed on hepatocytes and HCC cells membrane^[105,106]. N-cadherin expression is liver-specific. Hepatocytes and intrahepatic biliary epithelial cells express this marker on their plasma membrane while the extrahepatic bile ducts do not present this marker. This finding could be explained through the different embryological origins of extrahepatic and intrahepatic ducts. Mosnier *et al*^[107] demonstrated that N- and E-cadherins are present on the membrane of IH-CCA tumor cells with a rate of 66% and 45% respectively. N-cadherin expression is more frequent in peripheral CCAs than in hilar ones while EH-CCAs are negative for N-cadherin expression. The N-cadherin membranous expression plays an important role in distinguishing intrahepatic and perihilar CCAs from most HCCs and digestive tract tumors such as pancreatic adenocarcinoma.

VASCULAR ENDOTHELIAL GROWTH FACTOR-C

Vascular endothelial growth factor-C (VEGF-C) is considered a lymphangiogenic factor acting *via* VEGF receptor-3 (VEGFR-3). It is expressed by lymphatic endothelial cells and stimulates the growth of tumor-associated lymphatic vessels^[108,109].

One of the main features of CCA is its lymph node metastasis. The lymphatic invasion and lymph node metas-

tasis are important prognostic factors for IH-CCA^[110,111]. Aishima *et al*^[112] demonstrated that the density of the lymphatic vessels is lower in the center than in the tumor periphery and the tumor invasion into the lymphatic vessels is detected in the peritumoral area. Poorly differentiated tumors present higher density of lymphatic vessels in the tumor periphery and peritumoral area than in the tumor central area. VEGF-C expression of cancer cells is correlated with lymph node metastasis in IH-CCA^[113]. The lymphatic invasion in IH-CCA does not relate to the lymphangiogenesis but another mechanism is involved in the lymphatic spread. In fact, the preexisting lymphatic vessels in the tumor margin are sufficient for lymphatic metastasis. Recent studies have shown that VEGF-C is associated with lymphonodal metastasis and supports the entry of cancer cells into peritumoral lymphatic vessels^[114].

CLAUDINS

Claudins are transmembrane proteins and indispensable components of tight junctions^[115,116]. The tight junctions play an important role in the maintenance of epithelial cell polarity^[117,118]; they function as a barrier against the paracellular diffusion of solutes and also seem to regulate the growth and differentiation of cultured cells^[119-123]. Therefore, modifications of their constituents could lead to alterations of their functions.

Claudine is a family of 24 members, differentially expressed in various types of tissues. Németh *et al*^[124] showed that claudins are differently expressed in various compartments of the biliary tract. They analyzed the different types of Claudine in six sample groups (normal intrahepatic and extrahepatic bile ducts, normal gallbladder and carcinomas in the correspondent locations). Significant results have been reported for claudin-2 and in claudin-4. In particular, claudin-2 expression was stronger in GBC than in intrahepatic and extrahepatic bile duct cancers but a higher presence of claudin-4 has been found in extrahepatic bile duct cancers. Previous studies had already demonstrated the importance of claudin-4 for its ability to differentiate biliary tract cancers from hepatocellular tumors^[125,126]. On the contrary, claudin-2 causes leakiness to the tight junction complex which becomes less restrictive for ions^[127,128]. Therefore it plays a key role in bile concentration and in the processes of reabsorption in the proximal tubule and in the thin descending limb of Henle's loop^[125]. Németh *et al*^[124] also found a stronger expression of claudin-1 and claudin-10 in intrahepatic bile duct cancer than in extrahepatic bile duct cancer and in GBC.

THROMBOSPONDIN-1

Cancer growth is made possible by angiogenesis, regulated by angiogenic and antiangiogenic factors such as thrombospondin-1 (TSP-1), interferon alpha/beta, platelet factor-4, angiostatin and endostatin^[129,130]. In such a way, TSP-1 is a multifunctional matrix protein which is highly expressed, like its receptors, in the desmoplastic

stroma and the basement membrane, and is associated with tumors such as breast cancer and fibroadenoma^[131]. Nevertheless, its over expression in endothelial cells has been demonstrated to inhibits tumor genesis^[132]. Such results have also been shown by Kawahara *et al*^[133] who compared mRNA levels of TSP-1 and VEGF in CCA and HCC tumor cells in order to find a possible correlation between their expression and the different vascularization. They demonstrated that the up-regulation of TSP-1 and the down-regulation of VEGF in tumor cells may have a role in the hypovascularity of CCA when compared with HCC which is hypervascularized by the up-regulation of VEGF^[133].

CYTOKERATINS

Cytokeratins (CKs) are a complex subclass of the intermediate filaments gene family, made up of more than 20 different polypeptides. They are classified in type A or class I (CK9- CK20) and type B or class II (CK1-CK8)^[134-136]. The expression of CKs is generally confined to epithelia and their neoplasms^[137,138]. They are not specific tumor markers but the presence of distinct expression patterns of CKs in the various pathways of epithelial differentiation has been shown^[137,139,140]. In human cells, we can find from 2 to 10 types of CKs and, especially in the human liver, there is a different distribution in hepatocytes and in bile duct cells. CK8 and 18 are present in hepatocytes whereas CK7, 8, 17, 18 and 19 characterize bile duct cells. Among them, CK7 and CK19 can be considered as markers of this tissue and they could be used to differentiate CCA from HCC^[134,135]. Stroescu *et al*^[141] reported that the association of CKs and CEA makes the differential diagnosis between HCC and CCA more sensible. Cytokeratin immunophenotype of an adenocarcinoma can also help in the identification of its primary site. Several studies have shown the usefulness of CK17 as antibody for the identification of pancreaticobiliary adenocarcinoma and in the distinction of pancreaticobiliary and gastric adenocarcinomas when the primary site of the tumor is occult^[142].

SERUM CYTOKERATIN 19 FRAGMENT

Cytokeratins are intermediate filaments which are part of the cytoskeleton of epithelium. Serum cytokeratin 19 fragment (CYFRA 21-1) is considered a useful marker for non-small-cell lung cancer^[143] and a prognostic factor for many tumors such as cervical^[144], esophageal^[145], breast^[146] and gastric^[147] cancers.

Uenishi *et al*^[148] showed the association between CYFRA 21-1 serum concentrations and the tumor stage of CCA. Serum CYFRA 21-1 high values are correlated to tumor progression and poor post operative outcomes in CCA patients. The treatment of these patients includes not only surgery but also adjuvant therapy with gemcitabine combined with 5-fluorouracil or platinum, considered as first-line chemotherapy for advanced biliary tract carcinomas^[149,150].

Moreover, CYFRA 21-1 is considered a useful marker in the differential diagnosis between CCA and HCC. Thus, hepatocytes and bile duct cells show a different cytokeratin pattern^[151] that is maintained during malignant transformation.

Other markers

It is often very difficult to distinguish biliary disorders from bile duct and pancreatic cancers. However, Alvaro^[152] analyzed the possibility of discriminating CCA from benign biliary disorders and pancreatic cancers through the analysis of the biliary concentration of insulin-like growth factor 1. Furthermore, Tangkijvanich *et al*^[153] recently demonstrated the usefulness of combined use of AFP and glypican-3 to differentiate HCC from other liver cancers, like IH-CCA, because of the complementary role of these two markers. Such markers could be useful to discriminate benign from malignant neoplasm or other pathologies, but the possible presence of bile duct cancers could also be indicated by immune system alteration and its specific markers.

Enjoji *et al*^[154] noticed that RCAS1, a tumor associated antigen present in immune diseases, can be expressed in biliary tract carcinomas. This protein has a defensive role against the immune attack inducing apoptosis of immune cells. RCAS1 high levels lead to a rapid development of the neoplasia and its expression may be a specific event taking place in immune-mediated diseases. Its role in immune diseases is useful to distinguish malignant diseases from non-immune inflammatory biliary diseases such as drug-induced cholestasis and cholelithiasis.

Another proof that the immune system plays a fundamental role in the development of biliary tract cancers is the increase of heat shock protein expression, chaperone molecules that are present in several stress conditions^[155,156]. In particular, Romani *et al*^[157] noticed that the immunohistochemical expression of heat shock protein 27-kDa (HSP27) is correlated with patient survival and thus it could be considered a prognostic factor in IH-CCA. This study demonstrated that HSP27 acts to inhibit apoptotic cell death while HSP72 is correlated with the presence of necrosis and lymphoid infiltration.

CONCLUSION

Biliary tract cancers are relatively uncommon malignancies. Although the entire biliary tract is potentially at risk, the perihilar region is the most involved site, accounting for about 60% of all these tumors^[158]. However, an increase in the incidence of intrahepatic cancer in the US and the UK has also been shown^[159].

Biliary tract cancers are characterized by slow growth but, because of the late presentation of their symptoms, they are usually diagnosed in advanced stages when the majority of therapeutic choices are not curative^[158]. This leads to integrating the use of markers in order to formulate a better diagnosis and therapeutic approach.

In general, the first approach is a complete surgical resection that is possible with acceptable morbidity and

mortality^[160]. The possibility of applying this therapeutic choice is influenced by the tumor mass. Several studies have suggested a correlation between the size of the tumor and tumor markers levels. Particularly, Gerhardt *et al*^[161] found that initial levels of CA 19-9 in perihilar CCA patients who underwent to resection were lower than in those affected by unresectable disease. Similar results were found by Kau *et al*^[162] in relation to periampullary carcinoma. The levels of tumor markers are also important to indicate a possible prognosis. Fernández-Ruiz *et al*^[163] showed a worse prognosis in patients with elevated serum levels of CA 19-9 at the time of diagnosis for values > 270 IU/L.

Chemotherapy is another possible treatment option in patients with advanced biliary tract cancer. Patients with a good performance status can benefit from a combined chemotherapy consisting of two of the following drugs: gemcitabine, 5-FU/FA or capecitabine or a platinum analog^[164-166]. New discoveries about the role of markers, such as VEGF and ErbB, have led towards targeted therapy. However, trials with HER2-neu or EGFR inhibitors such as lapatinib or erlotinib have shown little or no activity in advanced biliary system adenocarcinomas^[167,168].

REFERENCES

- 1 Alison MR, Poulson R, Forbes SJ. Update on hepatic stem cells. *Liver* 2001; **21**: 367-373
- 2 Ikai I, Arii S, Kojima M, Ichida T, Makuuchi M, Matsuyama Y, Nakanuma Y, Okita K, Omata M, Takayasu K, Yamaoka Y. Reevaluation of prognostic factors for survival after liver resection in patients with hepatocellular carcinoma in a Japanese nationwide survey. *Cancer* 2004; **101**: 796-802
- 3 Shaib YH, El-Serag HB, Davila JA, Morgan R, McGlynn KA. Risk factors of intrahepatic cholangiocarcinoma in the United States: a case-control study. *Gastroenterology* 2005; **128**: 620-626
- 4 de Groen PC, Gores GJ, LaRusso NF, Gunderson LL, Nagorney DM. Biliary tract cancers. *N Engl J Med* 1999; **341**: 1368-1378
- 5 Fong Y, Malhotra S. Gallbladder cancer: recent advances and current guidelines for surgical therapy. *Adv Surg* 2001; **35**: 1-20
- 6 Lazcano-Ponce EC, Miquel JF, Muñoz N, Herrero R, Ferrer C, Wistuba II, Alonso de Ruiz P, Aristi Urista G, Nervi F. Epidemiology and molecular pathology of gallbladder cancer. *CA Cancer J Clin* 2001; **51**: 349-364
- 7 Nakeeb A, Pitt HA, Sohn TA, Coleman J, Abrams RA, Piantadosi S, Hruban RH, Lillemoe KD, Yeo CJ, Cameron JL. Cholangiocarcinoma. A spectrum of intrahepatic, perihilar, and distal tumors. *Ann Surg* 1996; **224**: 463-473; discussion 473-475
- 8 Khan SA, Davidson BR, Goldin R, Pereira SP, Rosenberg WM, Taylor-Robinson SD, Thillainayagam AV, Thomas HC, Thursz MR, Wasan H. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. *Gut* 2002; **51** Suppl 6: VI1-VI9
- 9 Blechacz B, Gores GJ. Tumor-specific marker genes for intrahepatic cholangiocarcinoma: utility and mechanistic insight. *J Hepatol* 2008; **49**: 160-162
- 10 Khan SA, Davidson BR, Goldin R, Pereira SP, Rosenberg WM, Taylor-Robinson SD, Thillainayagam AV, Thomas HC, Thursz MR, Wasan H. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. *Gut* 2002; **51** Suppl 6: VI1-VI9
- 11 Khan SA, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005; **366**: 1303-1314
- 12 Chaube A, Tewari M, Singh U, Shukla HS. CA 125: a potential tumor marker for gallbladder cancer. *J Surg Oncol* 2006; **93**: 665-669
- 13 Chen CY, Shiesh SC, Tsao HC, Lin XZ. The assessment of biliary CA 125, CA 19-9 and CEA in diagnosing cholangiocarcinoma--the influence of sampling time and hepatolithiasis. *Hepatogastroenterology* 2002; **49**: 616-620
- 14 Torzilli G, Makuuchi M, Ferrero A, Takayama T, Hui AM, Abe H, Inoue K, Nakahara K. Accuracy of the preoperative determination of tumor markers in the differentiation of liver mass lesions in surgical patients. *Hepatogastroenterology* 2002; **49**: 740-745
- 15 Hultcrantz R, Olsson R, Danielsson A, Järnerot G, Lööf L, Ryden BO, Wahren B, Broomé U. A 3-year prospective study on serum tumor markers used for detecting cholangiocarcinoma in patients with primary sclerosing cholangitis. *J Hepatol* 1999; **30**: 669-673
- 16 Caturelli E, Bisceglia M, Villani MR, de Maio G, Siena DA. CA 19-9 production by a cystadenoma with mesenchymal stroma of the common hepatic duct: a case report. *Liver* 1998; **18**: 221-224
- 17 Nichols JC, Gores GJ, LaRusso NF, Wiesner RH, Nagorney DM, Ritts RE Jr. Diagnostic role of serum CA 19-9 for cholangiocarcinoma in patients with primary sclerosing cholangitis. *Mayo Clin Proc* 1993; **68**: 874-879
- 18 Robertson AG, Davidson BR. Mirizzi syndrome complicating an anomalous biliary tract: a novel cause of a hugely elevated CA19-9. *Eur J Gastroenterol Hepatol* 2007; **19**: 167-169
- 19 Toomey DP, Swan N, Torreggiani W, Conlon KC. Autoimmune pancreatitis. *Br J Surg* 2007; **94**: 1067-1074
- 20 Murray MD, Burton FR, Di Bisceglie AM. Markedly elevated serum CA 19-9 levels in association with a benign biliary stricture due to primary sclerosing cholangitis. *J Clin Gastroenterol* 2007; **41**: 115-117
- 21 Uygur-Bayramicli O, Dabak R, Orbay E, Dolapcioglu C, Sargin M, Kilicoglu G, Guleryuzlu Y, Mayadagli A. Type 2 diabetes mellitus and CA 19-9 levels. *World J Gastroenterol* 2007; **13**: 5357-5359
- 22 Malaguarnera G, Giordano M, Paladina I, Berretta M, Capellani A, Malaguarnera M. Serum markers of hepatocellular carcinoma. *Dig Dis Sci* 2010; **55**: 2744-2755
- 23 Dorandeu A, Raoul JL, Siriser F, Leclercq-Rioux N, Gosselin M, Martin ED, Ramée MP, Launois B. Carcinoma of the ampulla of Vater: prognostic factors after curative surgery: a series of 45 cases. *Gut* 1997; **40**: 350-355
- 24 Nakamura S, Suzuki S, Sakaguchi T, Serizawa A, Konno H, Baba S, Baba S, Muro H. Surgical treatment of patients with mixed hepatocellular carcinoma and cholangiocarcinoma. *Cancer* 1996; **78**: 1671-1676
- 25 Gold P, Freedman SO. Specific carcinoembryonic antigens of the human digestive system. *J Exp Med* 1965; **122**: 467-481
- 26 Qin XL, Wang ZR, Shi JS, Lu M, Wang L, He QR. Utility of serum CA19-9 in diagnosis of cholangiocarcinoma: in comparison with CEA. *World J Gastroenterol* 2004; **10**: 427-432
- 27 Ramage JK, Donaghy A, Farrant JM, Iorns R, Williams R. Serum tumor markers for the diagnosis of cholangiocarcinoma in primary sclerosing cholangitis. *Gastroenterology* 1995; **108**: 865-869
- 28 Chalasani N, Baluyut A, Ismail A, Zaman A, Sood G, Ghalib R, McCashland TM, Reddy KR, Zervos X, Anbari MA, Hoen H. Cholangiocarcinoma in patients with primary sclerosing cholangitis: a multicenter case-control study. *Hepatology* 2000; **31**: 7-11
- 29 Deftos LJ. Chromogranin A: its role in endocrine function and as an endocrine and neuroendocrine tumor marker. *Endocr Rev* 1991; **12**: 181-187
- 30 O'Connor DT, Deftos LJ. Secretion of chromogranin A by peptide-producing endocrine neoplasms. *N Engl J Med* 1986;

- 314: 1145-1151
- 31 **Ranno S**, Motta M, Rampello E, Risino C, Bennati E, Malaguarnera M. The chromogranin-A (CgA) in prostate cancer. *Arch Gerontol Geriatr* 2006; **43**: 117-126
- 32 **Malaguarnera M**, Cristaldi E, Cammalleri L, Colonna V, Lipari H, Capici A, Cavallaro A, Beretta M, Alessandria I, Luca S, Motta M. Elevated chromogranin A (CgA) serum levels in the patients with advanced pancreatic cancer. *Arch Gerontol Geriatr* 2009; **48**: 213-217
- 33 **Tropea F**, Baldari S, Restifo G, Fiorillo MT, Surace P, Herberg A. Evaluation of chromogranin A expression in patients with non-neuroendocrine tumours. *Clin Drug Investig* 2006; **26**: 715-722
- 34 **Spadaro A**, Ajello A, Morace C, Zirilli A, D'arrigo G, Luigianno C, Martino F, Bene A, Migliorato D, Turiano S, Ferrau O, Freni MA. Serum chromogranin-A in hepatocellular carcinoma: diagnostic utility and limits. *World J Gastroenterol* 2005; **11**: 1987-1990
- 35 **Malaguarnera M**, Vacante M, Fichera R, Cappellani A, Cristaldi E, Motta M. Chromogranin A (CgA) serum level as a marker of progression in hepatocellular carcinoma (HCC) of elderly patients. *Arch Gerontol Geriatr* 2010; **51**: 81-85
- 36 **Erickson LA**, Lloyd RV. Practical markers used in the diagnosis of endocrine tumors. *Adv Anat Pathol* 2004; **11**: 175-189
- 37 **Hong SM**, Kim MJ, Pi DY, Jo D, Yu E, Ro JY. Neuroendocrine differentiation in extrahepatic bile duct carcinomas and its prognostic significance. *Hum Pathol* 2005; **36**: 732-740
- 38 **Wendeler MW**, Drenckhahn D, Gessner R, Baumgartner W. Intestinal LI-cadherin acts as a Ca²⁺-dependent adhesion switch. *J Mol Biol* 2007; **370**: 220-230
- 39 **Gessner R**, Tauber R. Intestinal cell adhesion molecules. Liver-intestine cadherin. *Ann N Y Acad Sci* 2000; **915**: 136-143
- 40 **Velcich A**, Yang W, Heyer J, Fragale A, Nicholas C, Viani S, Kucherlapati R, Lipkin M, Yang K, Augenlicht L. Colorectal cancer in mice genetically deficient in the mucin Muc2. *Science* 2002; **295**: 1726-1729
- 41 **Chang SK**, Dohrman AF, Basbaum CB, Ho SB, Tsuda T, Toribara NW, Gum JR, Kim YS. Localization of mucin (MUC2 and MUC3) messenger RNA and peptide expression in human normal intestine and colon cancer. *Gastroenterology* 1994; **107**: 28-36
- 42 **Buisine MP**, Devisme L, Maunoury V, Deschodt E, Gosselin B, Copin MC, Aubert JP, Porchet N. Developmental mucin gene expression in the gastroduodenal tract and accessory digestive glands. I. Stomach. A relationship to gastric carcinoma. *J Histochem Cytochem* 2000; **48**: 1657-1666
- 43 **Jass JR**, Walsh MD. Altered mucin expression in the gastrointestinal tract: a review. *J Cell Mol Med* 2001; **5**: 327-351
- 44 **Pereira MB**, Dias AJ, Reis CA, Schmitt FC. Immunohistochemical study of the expression of MUC5AC and MUC6 in breast carcinomas and adjacent breast tissues. *J Clin Pathol* 2001; **54**: 210-213
- 45 **Sasaki M**, Nakanuma Y, Terada T, Kim YS. Biliary epithelial expression of MUC1, MUC2, MUC3 and MUC5/6 apomucins during intrahepatic bile duct development and maturation. An immunohistochemical study. *Am J Pathol* 1995; **147**: 574-579
- 46 **Sasaki M**, Nakanuma Y. Expression of mucin core protein of mammary type in primary liver cancer. *Hepatology* 1994; **20**: 1192-1197
- 47 **Yamashita K**, Yonezawa S, Tanaka S, Shirahama H, Sakoda K, Imai K, Xing PX, McKenzie IF, Hilken J, Kim YS. Immunohistochemical study of mucin carbohydrates and core proteins in hepatolithiasis and cholangiocarcinoma. *Int J Cancer* 1993; **55**: 82-91
- 48 **Sasaki M**, Nakanuma Y, Kim YS. Characterization of apomucin expression in intrahepatic cholangiocarcinomas and their precursor lesions: an immunohistochemical study. *Hepatology* 1996; **24**: 1074-1078
- 49 **Matsumura N**, Yamamoto M, Aruga A, Takasaki K, Nakano M. Correlation between expression of MUC1 core protein and outcome after surgery in mass-forming intrahepatic cholangiocarcinoma. *Cancer* 2002; **94**: 1770-1776
- 50 **Wesseling J**, van der Valk SW, Vos HL, Sonnenberg A, Hilken J. Episialin (MUC1) overexpression inhibits integrin-mediated cell adhesion to extracellular matrix components. *J Cell Biol* 1995; **129**: 255-265
- 51 **Regimbald LH**, Pilarski LM, Longenecker BM, Reddish MA, Zimmermann G, Hugh JC. The breast mucin MUC1 as a novel adhesion ligand for endothelial intercellular adhesion molecule 1 in breast cancer. *Cancer Res* 1996; **56**: 4244-4249
- 52 **Nath D**, Hartnell A, Happerfield L, Miles DW, Burchell J, Taylor-Papadimitriou J, Crocker PR. Macrophage-tumour cell interactions: identification of MUC1 on breast cancer cells as a potential counter-receptor for the macrophage-restricted receptor, sialoadhesin. *Immunology* 1999; **98**: 213-219
- 53 **Yamashita Y**, Ho JJ, Farrelly ER, Hirakawa K, Sowa M, Kim YS. Forskolin and phorbol ester have opposite effects on the expression of mucin-associated sialyl-Lewis(a) in pancreatic cancer cells. *Eur J Cancer* 2000; **36**: 113-120
- 54 **van de Wiel-van Kemenade E**, Ligtenberg MJ, de Boer AJ, Buijs F, Vos HL, Melief CJ, Hilken J, Figdor CG. Episialin (MUC1) inhibits cytotoxic lymphocyte-target cell interaction. *J Immunol* 1993; **151**: 767-776
- 55 **Xu H**, Inagaki Y, Tang W, Guo Q, Wang F, Seyama Y, Midorikawa Y, Gai R, Kokudo N, Sugawara Y, Nakata M, Makuuchi M. Elevation of serum KL-6 mucin levels in patients with cholangiocarcinoma. *Hepatogastroenterology* 2008; **55**: 2000-2004
- 56 **Sasaki M**, Nakanuma Y. Frequent expression of MUC1 apomucin on biliary epithelial cells of damaged small bile ducts in primary biliary cirrhosis and chronic viral hepatitis: an immunohistochemical study. *Hepatology* 1996; **23**: 1313-1317
- 57 **Gurrieri C**, Capodice P, Bernardi R, Scaglioni PP, Nafa K, Rush LJ, Verbel DA, Cordon-Cardo C, Pandolfi PP. Loss of the tumor suppressor PML in human cancers of multiple histologic origins. *J Natl Cancer Inst* 2004; **96**: 269-279
- 58 **Malaguarnera L**, Cristaldi E, Malaguarnera M. The role of immunity in elderly cancer. *Crit Rev Oncol Hematol* 2010; **74**: 40-60
- 59 **Ajiki T**, Onoyama H, Yamamoto M, Asaka K, Fujimori T, Maeda S, Saitoh Y. P53 protein expression and prognosis in gallbladder carcinoma and premalignant lesions. *Hepatogastroenterology* 1996; **43**: 521-526
- 60 **Washington K**, Gottfried MR. Expression of p53 in adenocarcinoma of the gallbladder and bile ducts. *Liver* 1996; **16**: 99-104
- 61 **Shrestha ML**, Miyake H, Kikutsuji T, Tashiro S. Prognostic significance of Ki-67 and p53 antigen expression in carcinomas of bile duct and gallbladder. *J Med Invest* 1998; **45**: 95-102
- 62 **Diamantis I**, Karamitopoulou E, Perentes E, Zimmermann A. p53 protein immunoreactivity in extrahepatic bile duct and gallbladder cancer: correlation with tumor grade and survival. *Hepatology* 1995; **22**: 774-779
- 63 **Teh M**, Wee A, Raju GC. An immunohistochemical study of p53 protein in gallbladder and extrahepatic bile duct/ampullary carcinomas. *Cancer* 1994; **74**: 1542-1545
- 64 **Lee YC**, Song SY, Chung JB, Kang JK, Park IS. p53 protein expression in extrahepatic bile duct cancer. *Yonsei Med J* 1996; **37**: 112-117
- 65 **Hitchcock CL**. Ki-67 staining as a means to simplify analysis of tumor cell proliferation. *Am J Clin Pathol* 1991; **96**: 444-446
- 66 **Wintzer HO**, Zipfel I, Schulte-Mönting J, Hellerich U, von Kleist S. Ki-67 immunostaining in human breast tumors and its relationship to prognosis. *Cancer* 1991; **67**: 421-428
- 67 **Lönnnerholm G**, Selking O, Wistrand PJ. Amount and distribution of carbonic anhydrases CA I and CA II in the gastrointestinal tract. *Gastroenterology* 1985; **88**: 1151-1161
- 68 **Sly WS**, Hu PY. Human carbonic anhydrases and carbonic anhydrase deficiencies. *Annu Rev Biochem* 1995; **64**: 375-401
- 69 **Parkkila S**, Parkkila AK. Carbonic anhydrase in the aliment-

- tary tract. Roles of the different isozymes and salivary factors in the maintenance of optimal conditions in the gastrointestinal canal. *Scand J Gastroenterol* 1996; **31**: 305-317
- 70 **Fleming RE**, Parkkila S, Parkkila AK, Rajaniemi H, Waheed A, Sly WS. Carbonic anhydrase IV expression in rat and human gastrointestinal tract regional, cellular, and subcellular localization. *J Clin Invest* 1995; **96**: 2907-2913
- 71 **Parkkila S**, Parkkila AK, Juvonen T, Waheed A, Sly WS, Saarnio J, Kaunisto K, Kellokumpu S, Rajaniemi H. Membrane-bound carbonic anhydrase IV is expressed in the luminal plasma membrane of the human gallbladder epithelium. *Hepatology* 1996; **24**: 1104-1108
- 72 **Parkkila AK**, Herva R, Parkkila S, Rajaniemi H. Immunohistochemical demonstration of human carbonic anhydrase isoenzyme II in brain tumours. *Histochem J* 1995; **27**: 974-982
- 73 **Nagao Y**, Srinivasan M, Platero JS, Svendrowski M, Waheed A, Sly WS. Mitochondrial carbonic anhydrase (isozyme V) in mouse and rat: cDNA cloning, expression, subcellular localization, processing, and tissue distribution. *Proc Natl Acad Sci USA* 1994; **91**: 10330-10334
- 74 **Shah GN**, Hewett-Emmett D, Grubb JH, Migas MC, Fleming RE, Waheed A, Sly WS. Mitochondrial carbonic anhydrase CA VB: differences in tissue distribution and pattern of evolution from those of CA VA suggest distinct physiological roles. *Proc Natl Acad Sci USA* 2000; **97**: 1677-1682
- 75 **McCawley LJ**, Matrisian LM. Matrix metalloproteinases: multifunctional contributors to tumor progression. *Mol Med Today* 2000; **6**: 149-156
- 76 **Stetler-Stevenson WG**, Kruttsch HC, Liotta LA. TIMP-2: identification and characterization of a new member of the metalloproteinase inhibitor family. *Matrix Suppl* 1992; **1**: 299-306
- 77 **Ponton A**, Coulombe B, Skup D. Decreased expression of tissue inhibitor of metalloproteinases in metastatic tumor cells leading to increased levels of collagenase activity. *Cancer Res* 1991; **51**: 2138-2143
- 78 **Liotta LA**, Stetler-Stevenson WG. Metalloproteinases and cancer invasion. *Semin Cancer Biol* 1990; **1**: 99-106
- 79 **Jackson CJ**, Nguyen M. Human microvascular endothelial cells differ from macrovascular endothelial cells in their expression of matrix metalloproteinases. *Int J Biochem Cell Biol* 1997; **29**: 1167-1177
- 80 **Björklund M**, Koivunen E. Gelatinase-mediated migration and invasion of cancer cells. *Biochim Biophys Acta* 2005; **1755**: 37-69
- 81 **Westermarck J**, Kähäri VM. Regulation of matrix metalloproteinase expression in tumor invasion. *FASEB J* 1999; **13**: 781-792
- 82 **Itatsu K**, Sasaki M, Yamaguchi J, Ohira S, Ishikawa A, Ikeda H, Sato Y, Harada K, Zen Y, Sato H, Ohta T, Nagino M, Nimura Y, Nakanuma Y. Cyclooxygenase-2 is involved in the up-regulation of matrix metalloproteinase-9 in cholangiocarcinoma induced by tumor necrosis factor- α . *Am J Pathol* 2009; **174**: 829-841
- 83 **Leelawat K**, Sakchinabut S, Narong S, Wannaprasert J. Detection of serum MMP-7 and MMP-9 in cholangiocarcinoma patients: evaluation of diagnostic accuracy. *BMC Gastroenterol* 2009; **9**: 30
- 84 **Folgueras AR**, Pendás AM, Sánchez LM, López-Otín C. Matrix metalloproteinases in cancer: from new functions to improved inhibition strategies. *Int J Dev Biol* 2004; **48**: 411-424
- 85 **Itatsu K**, Zen Y, Yamaguchi J, Ohira S, Ishikawa A, Ikeda H, Sato Y, Harada K, Sasaki M, Sasaki M, Sakamoto H, Nagino M, Nimura Y, Ohta T, Nakanuma Y. Expression of matrix metalloproteinase 7 is an unfavorable postoperative prognostic factor in cholangiocarcinoma of the perihilar, hilar, and extrahepatic bile ducts. *Hum Pathol* 2008; **39**: 710-719
- 86 **Kirimlioglu H**, Türkmen I, Başsüllü N, Dirican A, Karadağ N, Kirimlioglu V. The expression of matrix metalloproteinases in intrahepatic cholangiocarcinoma, hilar (Klatskin tumor), middle and distal extrahepatic cholangiocarcinoma, gallbladder cancer, and ampullary carcinoma: role of matrix metalloproteinases in tumor progression and prognosis. *Turk J Gastroenterol* 2009; **20**: 41-47
- 87 **Roskoski R Jr.** The ErbB/HER receptor protein-tyrosine kinases and cancer. *Biochem Biophys Res Commun* 2004; **319**: 1-11
- 88 **Linggi B**, Carpenter G. ErbB receptors: new insights on mechanisms and biology. *Trends Cell Biol* 2006; **16**: 649-656
- 89 **Semba K**, Kamata N, Toyoshima K, Yamamoto T. A v-erbB-related protooncogene, c-erbB-2, is distinct from the c-erbB-1/epidermal growth factor-receptor gene and is amplified in a human salivary gland adenocarcinoma. *Proc Natl Acad Sci USA* 1985; **82**: 6497-6501
- 90 **Toyoshima K**, Semba K, Akiyama T, Ikawa S, Yamamoto T. The c-erbB-2 gene encodes a receptor-like protein with tyrosine kinase activity. *Cold Spring Harb Symp Quant Biol* 1986; **51 Pt 2**: 977-982
- 91 **Pelosi G**, Del Curto B, Dell'Orto P, Pasini F, Veronesi G, Spaggiari L, Maisonneuve P, Iannucci A, Terzi A, Lonardon A, Viale G. Lack of prognostic implications of HER-2/neu abnormalities in 345 stage I non-small cell carcinomas (NSCLC) and 207 stage I-III neuroendocrine tumours (NET) of the lung. *Int J Cancer* 2005; **113**: 101-108
- 92 **Zheng J**, Zhu YM. Expression of c-erbB-2 proto-oncogene in extrahepatic cholangiocarcinoma and its clinical significance. *Hepatobiliary Pancreat Dis Int* 2007; **6**: 412-415
- 93 **Bergstrand CG**, Czar B. Demonstration of a new protein fraction in serum from the human fetus. *Scand J Clin Lab Invest* 1956; **8**: 174
- 94 **Tatarinov IuS.** [Detection of embryo-specific alpha-globulin in the blood serum of a patient with primary liver cancer]. *Vopr Med Khim* 1964; **10**: 90-91
- 95 **Chayvialle JA**, Ganguli PC. Radioimmunoassay of alpha-fetoprotein in human plasma. *Lancet* 1973; **1**: 1355-1357
- 96 **Hirai H**, Nishi S, Watabe H. Radioimmunoassay of α -fetoprotein. Protides of the biological fluids. Pergamon Press, 1973; **20**: 579-587
- 97 **Purves LR**, Purves M. Serum alpha-feto-protein. VI. The radio-immunoassay evidence for the presence of AFP in the serum of normal people and during pregnancy. *S Afr Med J* 1972; **46**: 1290-1297
- 98 **Ruoslahti E**, Seppala M. Studies of carcino-fetal proteins. 3. Development of a radioimmunoassay for -fetoprotein. Demonstration of -fetoprotein in serum of healthy human adults. *Int J Cancer* 1971; **8**: 374-383
- 99 **Silver HK**, Gold P, Feder S, Freedman SO, Shuster J. Radioimmunoassay for human alpha 1 -fetoprotein. *Proc Natl Acad Sci USA* 1973; **70**: 526-530
- 100 **Waldmann TA**, McIntire KR. Serum-alpha-fetoprotein levels in patients with ataxia-telangiectasia. *Lancet* 1972; **2**: 1112-1115
- 101 **McIntire KR**, Waldmann TA, Moertel CG, Go VL. Serum alpha-fetoprotein in patients with neoplasms of the gastrointestinal tract. *Cancer Res* 1975; **35**: 991-996
- 102 **Takahashi H**, Oyamada M, Fujimoto Y, Satoh MI, Hattori A, Dempo K, Mori M, Tanaka T, Watabe H, Masuda R. Elevation of serum alpha-fetoprotein and proliferation of oval cells in the livers of LEC rats. *Jpn J Cancer Res* 1988; **79**: 821-827
- 103 **Jalanko H**, Kuusela P, Roberts P, Sipponen P, Haglund CA, Mäkelä O. Comparison of a new tumour marker, CA 19-9, with alpha-fetoprotein and carcinoembryonic antigen in patients with upper gastrointestinal diseases. *J Clin Pathol* 1984; **37**: 218-222
- 104 **Okuda H**, Shiratori K, Yamamoto M, Takasaki K, Nakano M. Clinicopathologic features of patients with intrahepatic cholangiocarcinoma who are seropositive for alpha-fetoprotein-L3 and those with combined hepatocellular and cholangiocarcinoma. *J Gastroenterol Hepatol* 2006; **21**: 869-873
- 105 **Yamaoka K**, Nouchi T, Tazawa J, Hiranuma S, Marumo F, Sato C. Expression of gap junction protein connexin 32 and E-cadherin in human hepatocellular carcinoma. *J Hepatol* 1995; **22**: 536-539

- 106 **Wei Y**, Van Nhieu JT, Prigent S, Srivatanakul P, Tiollais P, Buendia MA. Altered expression of E-cadherin in hepatocellular carcinoma: correlations with genetic alterations, beta-catenin expression, and clinical features. *Hepatology* 2002; **36**: 692-701
- 107 **Mosnier JF**, Kandel C, Cazals-Hatem D, Bou-Hanna C, Gournay J, Jarry A, Laboisse CL. N-cadherin serves as diagnostic biomarker in intrahepatic and perihilar cholangiocarcinomas. *Mod Pathol* 2009; **22**: 182-190
- 108 **Wong SY**, Haack H, Crowley D, Barry M, Bronson RT, Hynes RO. Tumor-secreted vascular endothelial growth factor-C is necessary for prostate cancer lymphangiogenesis, but lymphangiogenesis is unnecessary for lymph node metastasis. *Cancer Res* 2005; **65**: 9789-9798
- 109 **Skobe M**, Hawighorst T, Jackson DG, Prevo R, Janes L, Velasco P, Riccardi L, Alitalo K, Claffey K, Detmar M. Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. *Nat Med* 2001; **7**: 192-198
- 110 **Inoue K**, Makuuchi M, Takayama T, Torzilli G, Yamamoto J, Shimada K, Kosuge T, Yamasaki S, Konishi M, Kinoshita T, Miyagawa S, Kawasaki S. Long-term survival and prognostic factors in the surgical treatment of mass-forming type cholangiocarcinoma. *Surgery* 2000; **127**: 498-505
- 111 **Kawarada Y**, Yamagiwa K, Das BC. Analysis of the relationships between clinicopathologic factors and survival time in intrahepatic cholangiocarcinoma. *Am J Surg* 2002; **183**: 679-685
- 112 **Aishima S**, Nishihara Y, Iguchi T, Taguchi K, Taketomi A, Maehara Y, Tsuneyoshi M. Lymphatic spread is related to VEGF-C expression and D2-40-positive myofibroblasts in intrahepatic cholangiocarcinoma. *Mod Pathol* 2008; **21**: 256-264
- 113 **Park BK**, Paik YH, Park JY, Park KH, Bang S, Park SW, Chung JB, Park YN, Song SY. The clinicopathologic significance of the expression of vascular endothelial growth factor-C in intrahepatic cholangiocarcinoma. *Am J Clin Oncol* 2006; **29**: 138-142
- 114 **Schneider M**, Büchler P, Giese N, Giese T, Wiltng J, Büchler MW, Friess H. Role of lymphangiogenesis and lymphangiogenic factors during pancreatic cancer progression and lymphatic spread. *Int J Oncol* 2006; **28**: 883-890
- 115 **Furuse M**, Fujita K, Hiiiragi T, Fujimoto K, Tsukita S. Claudin-1 and -2: novel integral membrane proteins localizing at tight junctions with no sequence similarity to occludin. *J Cell Biol* 1998; **141**: 1539-1550
- 116 **Morita K**, Furuse M, Fujimoto K, Tsukita S. Claudin multi-gene family encoding four-transmembrane domain protein components of tight junction strands. *Proc Natl Acad Sci USA* 1999; **96**: 511-516
- 117 **Rodriguez-Boulan E**, Nelson WJ. Morphogenesis of the polarized epithelial cell phenotype. *Science* 1989; **245**: 718-725
- 118 **Miyoshi J**, Takai Y. Molecular perspective on tight-junction assembly and epithelial polarity. *Adv Drug Deliv Rev* 2005; **57**: 815-855
- 119 **Gumbiner BM**. Breaking through the tight junction barrier. *J Cell Biol* 1993; **123**: 1631-1633
- 120 **Anderson JM**, Van Itallie CM. Tight junctions and the molecular basis for regulation of paracellular permeability. *Am J Physiol* 1995; **269**: G467-G475
- 121 **Ivanov AI**, Nusrat A, Parkos CA. Endocytosis of the apical junctional complex: mechanisms and possible roles in regulation of epithelial barriers. *Bioessays* 2005; **27**: 356-365
- 122 **Balda MS**, Matter K. Tight junctions. *J Cell Sci* 1998; **111** (Pt 5): 541-547
- 123 **Tsukita S**, Furuse M, Itoh M. Structural and signalling molecules come together at tight junctions. *Curr Opin Cell Biol* 1999; **11**: 628-633
- 124 **Németh Z**, Szász AM, Tátrai P, Németh J, Gyorffy H, Somorácz A, Szijártó A, Kupcsulik P, Kiss A, Schaff Z. Claudin-1, -2, -3, -4, -7, -8, and -10 protein expression in biliary tract cancers. *J Histochem Cytochem* 2009; **57**: 113-121
- 125 **Lódi C**, Szabó E, Holczbauer A, Batmunkh E, Szijártó A, Kupcsulik P, Kovalszky I, Paku S, Illyés G, Kiss A, Schaff Z. Claudin-4 differentiates biliary tract cancers from hepatocellular carcinomas. *Mod Pathol* 2006; **19**: 460-469
- 126 **Nishino R**, Honda M, Yamashita T, Takatori H, Minato H, Zen Y, Sasaki M, Takamura H, Horimoto K, Ohta T, Nakanuma Y, Kaneko S. Identification of novel candidate tumour marker genes for intrahepatic cholangiocarcinoma. *J Hepatol* 2008; **49**: 207-216
- 127 **Amasheh S**, Meiri N, Gitter AH, Schöneberg T, Mankertz J, Schulzke JD, Fromm M. Claudin-2 expression induces cation-selective channels in tight junctions of epithelial cells. *J Cell Sci* 2002; **115**: 4969-4976
- 128 **Kiuchi-Saishin Y**, Gotoh S, Furuse M, Takasuga A, Tano Y, Tsukita S. Differential expression patterns of claudins, tight junction membrane proteins, in mouse nephron segments. *J Am Soc Nephrol* 2002; **13**: 875-886
- 129 **Hanahan D**, Folkman J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell* 1996; **86**: 353-364.
- 130 **Folkman J**. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat Med* 1995; **1**: 27-31
- 131 **Tuszynski GP**, Nicosia RF. The role of thrombospondin-1 in tumor progression and angiogenesis. *Bioessays* 1996; **18**: 71-76
- 132 **Sheibani N**, Frazier WA. Thrombospondin 1 expression in transformed endothelial cells restores a normal phenotype and suppresses their tumorigenesis. *Proc Natl Acad Sci USA* 1995; **92**: 6788-6792
- 133 **Kawahara N**, Ono M, Taguchi K, Okamoto M, Shimada M, Takenaka K, Hayashi K, Mosher DF, Sugimachi K, Tsuneyoshi M, Kuwano M. Enhanced expression of thrombospondin-1 and hypovascularity in human cholangiocarcinoma. *Hepatology* 1998; **28**: 1512-1517
- 134 **Karsten U**, Papsdorf G, Roloff G, Stolley P, Abel H, Walther I, Weiss H. Monoclonal anti-cytokeratin antibody from a hybridoma clone generated by electrofusion. *Eur J Cancer Clin Oncol* 1985; **21**: 733-740
- 135 **Balaton AJ**, Nehama-Sibony M, Gotheil C, Callard P, Baviera EE. Distinction between hepatocellular carcinoma, cholangiocarcinoma, and metastatic carcinoma based on immunohistochemical staining for carcinoembryonic antigen and for cytokeratin 19 on paraffin sections. *J Pathol* 1988; **156**: 305-310
- 136 **Hurlimann J**, Gardiol D. Immunohistochemistry in the differential diagnosis of liver carcinomas. *Am J Surg Pathol* 1991; **15**: 280-288
- 137 **Cooper D**, Schermer A, Sun TT. Classification of human epithelia and their neoplasms using monoclonal antibodies to keratins: strategies, applications, and limitations. *Lab Invest* 1985; **52**: 243-256
- 138 **Moll R**, Franke WW, Schiller DL, Geiger B, Krepler R. The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. *Cell* 1982; **31**: 11-24
- 139 **Moll R**, Lowe A, Laufer J, Franke WW. Cytokeratin 20 in human carcinomas. A new histodiagnostic marker detected by monoclonal antibodies. *Am J Pathol* 1992; **140**: 427-447
- 140 **Wu YJ**, Rheinwald JG. A new small (40 kd) keratin filament protein made by some cultured human squamous cell carcinomas. *Cell* 1981; **25**: 627-35
- 141 **Stroescu C**, Herlea V, Dragnea A, Popescu I. The diagnostic value of cytokeratins and carcinoembryonic antigen immunostaining in differentiating hepatocellular carcinomas from intrahepatic cholangiocarcinomas. *J Gastrointest Liver Dis* 2006; **15**: 9-14
- 142 **Sarbia M**, Fritze F, Gedder H, von Weyhern C, Rosenberg R, Gellert K. Differentiation between pancreaticobiliary and upper gastrointestinal adenocarcinomas: is analysis of cytokeratin 17 expression helpful? *Am J Clin Pathol* 2007; **128**: 255-259
- 143 **Pujol JL**, Molinier O, Ebert W, Daurès JP, Barlesi F, Buccheri

- G, Paesmans M, Quoix E, Moro-Sibilot D, Szturmowicz M, Bréchet JM, Muley T, Grenier J. CYFRA 21-1 is a prognostic determinant in non-small-cell lung cancer: results of a meta-analysis in 2063 patients. *Br J Cancer* 2004; **90**: 2097-2105
- 144 **Dowek I**, Barak M, Uri N, Greenberg E. The prognostic value of the tumour marker Cyfra 21-1 in carcinoma of head and neck and its role in early detection of recurrent disease. *Br J Cancer* 2000; **83**: 1696-701
- 145 **Brockmann JG**, St Nottberg H, Glodny B, Heinecke A, Senninger NJ. CYFRA 21-1 serum analysis in patients with esophageal cancer. *Clin Cancer Res* 2000; **6**: 4249-4252
- 146 **Nakata B**, Takashima T, Ogawa Y, Ishikawa T, Hirakawa K. Serum CYFRA 21-1 (cytokeratin-19 fragments) is a useful tumour marker for detecting disease relapse and assessing treatment efficacy in breast cancer. *Br J Cancer* 2004; **91**: 873-878
- 147 **Nakata B**, Chung YS, Kato Y, Ogawa M, Ogawa Y, Inui A, Maeda K, Sawada T, Sowa M. Clinical significance of serum CYFRA 21-1 in gastric cancer. *Br J Cancer* 1996; **73**: 1529-1532
- 148 **Uenishi T**, Yamazaki O, Tanaka H, Takemura S, Yamamoto T, Tanaka S, Nishiguchi S, Kubo S. Serum cytokeratin 19 fragment (CYFRA21-1) as a prognostic factor in intrahepatic cholangiocarcinoma. *Ann Surg Oncol* 2008; **15**: 583-589
- 149 **André T**, Tournigand C, Rosmorduc O, Provent S, Maindrault-Goebel F, Avenin D, Selle F, Paye F, Hannoun L, Houry S, Gayet B, Lotz JP, de Gramont A, Louvet C. Gemcitabine combined with oxaliplatin (GEMOX) in advanced biliary tract adenocarcinoma: a GERCOR study. *Ann Oncol* 2004; **15**: 1339-1343
- 150 **Eckel F**, Schmid RM. Chemotherapy in advanced biliary tract carcinoma: a pooled analysis of clinical trials. *Br J Cancer* 2007; **96**: 896-902
- 151 **Van Eyken P**, Desmet VJ. Cytokeratins and the liver. *Liver* 1993; **13**: 113-122
- 152 **Alvaro D**. Serum and bile biomarkers for cholangiocarcinoma. *Curr Opin Gastroenterol* 2009; **25**: 279-284
- 153 **Tangkijvanich P**, Chanmee T, Komtong S, Mahachai V, Wisedopas N, Pothacharoen P, Kongtawelert P. Diagnostic role of serum glypican-3 in differentiating hepatocellular carcinoma from non-malignant chronic liver disease and other liver cancers. *J Gastroenterol Hepatol* 2010; **25**: 129-137
- 154 **Enjoji M**, Nakashima M, Nishi H, Choi I, Oimomi H, Sugimoto R, Kotoh K, Taguchi K, Nakamuta M, Nawata H, Watanabe T. The tumor-associated antigen, RCAS1, can be expressed in immune-mediated diseases as well as in carcinomas of biliary tract. *J Hepatol* 2002; **36**: 786-792
- 155 **Motta M**, Ferlito L, Malaguarnera L, Vinci E, Bosco S, Maugeri D, Malaguarnera M. Alterations of the lymphocytic set-up in elderly patients with cancer. *Arch Gerontol Geriatr* 2003; **36**: 7-14
- 156 **Malaguarnera L**, Ferlito L, Di Mauro S, Imbesi RM, Scalia G, Malaguarnera M. Immunosenescence and cancer: a review. *Arch Gerontol Geriatr* 2001; **32**: 77-93
- 157 **Romani AA**, Crafa P, Desenzani S, Graiani G, Lagrasta C, Sianesi M, Soliani P, Borghetti AF. The expression of HSP27 is associated with poor clinical outcome in intrahepatic cholangiocarcinoma. *BMC Cancer* 2007; **7**: 232
- 158 **Khan SA**, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005; **366**: 1303-1314
- 159 **Patel T**. Increasing incidence and mortality of primary intrahepatic cholangiocarcinoma in the United States. *Hepatology* 2001; **33**: 1353-1357
- 160 **Jarnagin WR**, Fong Y, DeMatteo RP, Gonen M, Burke EC, Bodniewicz BS J, Youssef BA M, Klimstra D, Blumgart LH. Staging, resectability, and outcome in 225 patients with hilar cholangiocarcinoma. *Ann Surg* 2001; **234**: 507-517; discussion 517-519
- 161 **Gerhardt T**, Milz S, Schepke M, Feldmann G, Wolff M, Sauerbruch T, Dumoulin FL. C-reactive protein is a prognostic indicator in patients with perihilar cholangiocarcinoma. *World J Gastroenterol* 2006; **12**: 5495-5500
- 162 **Kau SY**, Shyr YM, Su CH, Wu CW, Lui WY. Diagnostic and prognostic values of CA 19-9 and CEA in periampullary cancers. *J Am Coll Surg* 1999; **188**: 415-420
- 163 **Fernández-Ruiz M**, Guerra-Vales JM, Colina-Ruizdelgado F. Comorbidity negatively influences prognosis in patients with extrahepatic cholangiocarcinoma. *World J Gastroenterol* 2009; **15**: 5279-5286
- 164 **Kornek GV**, Schuell B, Laengle F, Gruenberger T, Penz M, Karall K, Depisch D, Lang F, Scheithauer W. Mitomycin C in combination with capecitabine or biweekly high-dose gemcitabine in patients with advanced biliary tract cancer: a randomised phase II trial. *Ann Oncol* 2004; **15**: 478-483
- 165 **Ducieux M**, Van Cutsem E, Van Laethem JL, Gress TM, Jeziorski K, Rougier P, Wagener T, Anak O, Baron B, Nordlinger B. A randomised phase II trial of weekly high-dose 5-fluorouracil with and without folinic acid and cisplatin in patients with advanced biliary tract carcinoma: results of the 40955 EORTC trial. *Eur J Cancer* 2005; **41**: 398-403
- 166 **Rao S**, Cunningham D, Hawkins RE, Hill ME, Smith D, Daniel F, Ross PJ, Oates J, Norman AR. Phase III study of 5FU, etoposide and leucovorin (FELV) compared to epirubicin, cisplatin and 5FU (ECF) in previously untreated patients with advanced biliary cancer. *Br J Cancer* 2005; **92**: 1650-1654
- 167 **Philip PA**, Mahoney MR, Allmer C, Thomas J, Pitot HC, Kim G, Donehower RC, Fitch T, Picus J, Erlichman C. Phase II study of erlotinib in patients with advanced biliary cancer. *J Clin Oncol* 2006; **24**: 3069-3074
- 168 **Ramanathan RK**, Belani CP, Singh DA, Tanaka M, Lenz HJ, Yen Y, Kindler HL, Iqbal S, Longmate J, Mack PC, Lurje G, Gandour-Edwards R, Dancey J, Gandara DR. A phase II study of lapatinib in patients with advanced biliary tree and hepatocellular cancer. *Cancer Chemother Pharmacol* 2009; **64**: 777-783

S- Editor Wang JL L- Editor Roemmele A E- Editor Ma WH