

## Expression and significance of tumor-related genes in HCC

Zi-Li Lü, Dian-Zhong Luo, Jian-Ming Wen

Zi-Li Lü, Dian-Zhong Luo, Department of Pathology, Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Jian-Ming Wen, Department of Pathology, Zhongshan Medical College, Sun Yat-Sen University, Guangzhou 510080, Guangdong Province, China

Supported by the National Natural Science Foundation of China, No. 39860079

Correspondence to: Dr. Dian-Zhong Luo, Department of Pathology, Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China. luodianzhong@yahoo.com.cn  
Telephone: +86-771-5356534

Received: 2004-08-18 Accepted: 2004-11-24

### Abstract

**AIM:** To investigate the expression and clinical significance of *DEK*, *cyclin D1*, insulin-like growth factor II (*IGF-II*), glypican 3 (*GPC3*), ribosomal phosphoprotein 0 (*rpP0*) mRNA in hepatocellular carcinoma (HCC) and its paraneoplastic tissues.

**METHODS:** The expression of mRNAs of *DEK*, *cyclin D1*, *IGF-II*, *GPC3* and *rpP0* mRNA was detected in HCC and its paraneoplastic tissues by multiplex RT-PCR.

**RESULTS:** By the simplex RT-PCR, the overexpression of mRNAs of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNA in HCC and its paraneoplastic tissues was 78.1%, 87.5%, 87.5%, 75.0%, 81.3% and 15.6%, 40.6%, 37.5%, 21.9%, 31.3% respectively ( $P < 0.05$ ). By the multiplex RT-PCR, at least one of the mRNAs was detected in all HCC samples and in 75.0% of paraneoplastic samples ( $P > 0.05$ ). However, all these five mRNAs were found in 68.8% of HCC samples, but only in 9.4% of paraneoplastic tissues ( $P < 0.05$ ). The positive expression of mRNAs of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* in well- and poorly-differentiated HCC was 89.0%, 66.7%, 66.7%, 66.7%, 77.8% and 73.9%, 95.7%, 95.7%, 82.6%, respectively ( $P > 0.05$ ). The expression of these genes in HCCs with  $\alpha$ -feto protein (AFP) negative and positive was 90.0%, 80.0%, 90.0%, 90.0%, 90.0% and 72.7%, 86.3%, 77.3%, 90.9%, 68.2% respectively ( $P > 0.05$ ).

**CONCLUSION:** The expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNA in HCC is much higher in HCC than in its paraneoplastic tissues. Multiplex RT-PCR assay is an effective, sensitive, accurate, and cost-effective diagnostic method of HCC.

© 2005 The WJG Press and Elsevier Inc. All rights reserved.

**Key words:** HCC; *DEK*; *Cyclin D1*; *IGF-II*; *GPC3*; *rpP0*

Lü ZL, Luo DZ, Wen JM. Expression and significance of tumor-related genes in HCC. *World J Gastroenterol* 2005; 11(25): 3850-3854

<http://www.wjgnet.com/1007-9327/11/3850.asp>

### INTRODUCTION

Hepatocellular carcinoma (HCC) is an aggressive malignancy with poor prognosis and one of the most common tumors in humans. The development of HCC is a chronic process and involves many factors, including infection of hepatitis virus and contamination with aflatoxin B1<sup>[1]</sup>. Recent advances in molecular genetics indicate that some tumor suppressor genes, oncogenes, and growth factors may play an important role in hepatocarcinogenesis<sup>[2]</sup>.

Several methods such as DDRT-PCR, cDNA screening<sup>[3]</sup> are used to identify differential expression of mRNA in tumor and non-tumor tissues. It is reported that some genes, such as *DEK*, *rpP0*<sup>[4]</sup>, *cyclin D1*<sup>[5]</sup>, *IGF-II*<sup>[6]</sup>, and *GPC3*<sup>[7]</sup>, are overexpressed in HCC tissue. The expression of *DEK*, *IGF-II* and *rpP0* is higher in HCC than non-tumor tissues in our previous study.

RT-PCR is widely used to analyze gene expression in HCC and paraneoplastic liver tissue. However, the overexpressed genes are only relatively higher in HCC than in paraneoplastic liver tissue and the positive detection rate is relatively low. Moreover, RT-PCR can detect only a single gene once in the past, and the results obtained are fluctuant and less useful in clinics. In order to find a method to enhance the specificity and positive rate, multiplex PCR was used to detect several genes, such as *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0*.

### MATERIALS AND METHODS

#### Tissue samples and patients

HCC and corresponding paraneoplastic tissues were obtained with informed consent from 32 patients who underwent hepatectomy at the First Affiliated Hospital of Guangxi Medical University and Guangxi Tumor Hospital. The profiles were obtained from medical records of 20 male and 12 female patients with an average age of 42.5 years. Twenty-two patients were positive for  $\alpha$ -feto protein (AFP). HCC and paraneoplastic tissues were enucleated separately and immediately frozen in liquid nitrogen. Histological classification was performed according to the Edmondson's grading criteria.

#### Multiplex RT-PCR

Total RNA was isolated from 100 mg of frozen tissue according to the manufacturer's instructions using TRIzol

kit (Sagon Company, Shanghai, China), and then dissolved in water that was treated by DEPC. Four micrograms of total RNA was used to produce cDNA using oligo(dT) primer and MuLV reverse transcription (MBI Company) in a final volume of 20  $\mu$ L at 42  $^{\circ}$ C for 1 h. The reaction was terminated by incubation at 75  $^{\circ}$ C for 10 min. One microgram of products was PCR amplified with multiple primer sets (0.5  $\mu$ mol/L, Sagon Company, Shanghai, China), 0.75 units of Taq DNA polymerase (MBI), 0.5  $\mu$ L 10 mmol/L dNTPs, 2.5  $\mu$ L 10 $\times$  buffer. The primers were as follows: *DEK*: 5'-AGG CAC TGT GTC CTC ATT AA and 5'-TCT GAC AGA ATT TCA GGA CA (332 bp); *cyclin D1*: 5'-TAT TTG CAT AAC CCT GAG CG and 5'-GTG ACT ACA TGC ATA TGA GC (350 bp); *IGF*: 5'-AGG AGC TCC TGG ATA ATT TC and 5'-AAT ATT TCA CGT GAC AGA AC (421 bp); *GPC3*: 5'-TGG ACA TCA ATG AGT GCC TC and 5'-CAC ATT CTG GTG AGC ATT CG (204 bp); *rpP0*: 5'-ATG TGA AGT CAC TGT GCC AG and 5'-CTT GGC TTC AAC CTT AGC TG (549 bp). GAPDH was used as control, the primers were 5'-TGA GTA CGT CGT GGA GTC CA and 5'-CAA AGT TGT CAT GGA TGA CC (230 bp). The conditions were initial denaturation at 94  $^{\circ}$ C for 5 min, 30 cycles of amplification, each cycle consisting of denaturation at 94  $^{\circ}$ C for 45 s, annealing at 58  $^{\circ}$ C for 30 s, extension at 72  $^{\circ}$ C for 30 s, final extension at 72  $^{\circ}$ C for 7 min. Because of the limitation by the length of PCR products, the primers of *cyclin D1* and *GPC3* were used together in a single reaction and the primers of *DEK*, *IGF-II*, and *rpP0* were used together in a single reaction.

### Analysis by electrophoresis

The amplified products were electrophoresed on 1.2% agarose gel to detect the expression of the genes in HCC and paraneoplastic tissues. The images were analyzed by Quantity One software.

### Statistical analysis

Results were analyzed by  $\chi^2$  test to compare the differences between the groups.  $P < 0.05$  was considered statistically significant.

## RESULTS

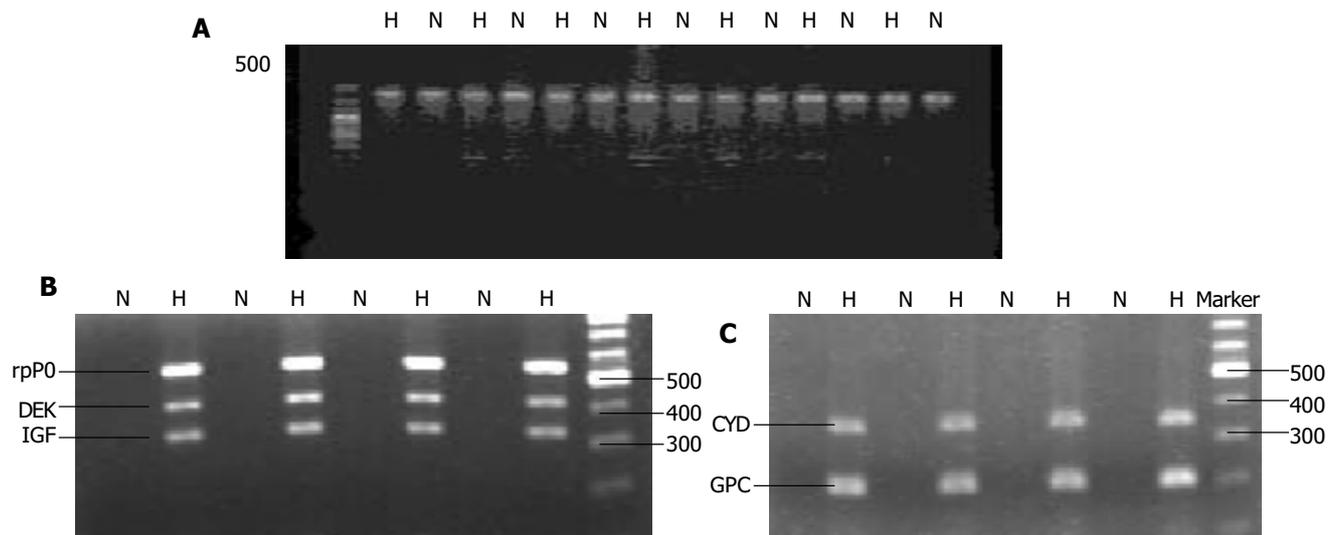
### Expression of GAPDH mRNA

The expression of *GAPDH* mRNA was detected in all HCC and non-HCC tissues. There was no difference between the two groups (Figure 1A).

### Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC and adjacent noncancerous liver tissues

The expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC and adjacent noncancerous liver tissues was 78.1%, 87.5%, 87.5%, 75.0%, 81.3%, and 15.6%, 40.6%, 37.5%, 21.9%, 31.3%, respectively ( $P < 0.05$ , Table 1), which were significantly higher in HCC tissues than in adjacent nontumorous tissue (Figures 1B and C). The density of the bands was also higher in HCC than in adjacent noncancerous liver tissues.

The expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in well- and poorly-differentiated HCC was 89.0%,



**Figure 1** Expression of GAPDH (A), mRNA of *rpP0*, *DEK*, and *IGF-II* (B), mRNA of *cyclin D1* and *GPC3* (C) in HCC tissue (H) and adjacent nontumorous tissue (N).

**Table 1** Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC and adjacent noncancerous liver tissues

	Cyclin D1 <sup>a</sup>		GPC3 <sup>c</sup>		DEK <sup>e</sup>		IGF <sup>g</sup>		rpP0 <sup>i</sup>	
	T	N	T	N	T	N	T	N	T	N
Positive	28	13	24	7	25	5	28	12	26	10
Negative	4	19	8	25	7	27	4	20	6	22

T, HCC tumor tissue; N, adjacent nontumorous tissue.  $\chi^2$ -test: <sup>a</sup> $P < 0.05$ , <sup>c</sup> $P < 0.05$ , <sup>e</sup> $P < 0.05$ , <sup>g</sup> $P < 0.05$ , <sup>i</sup> $P < 0.05$  vs others.

66.7%, 66.7%, 66.7%, 77.8% and 73.9%, 95.7%, 95.7%, 95.7%, 82.6%, respectively ( $P>0.05$ , Table 2).

The expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC with AFP negative and positive was 90.0%, 80.0%, 90.0%, 90.0%, 90.0% and 72.7%, 86.3%, 77.3%, 90.9%, 68.2%, respectively ( $P>0.05$ , Table 3).

### Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC and adjacent noncancerous liver tissues shown by multiplex RT-PCR

By multiplex RT-PCR, at least one of the mRNAs could be detected in all HCC tissues and in 75.0% of paraneoplastic tissues ( $P>0.05$ ) (Table 4). However, all these five mRNAs were found in 68.8% of HCC tissue, but only in 9.4% of paraneoplastic tissues ( $P<0.05$ , Table 4).

## DISCUSSION

The growth of cells depends on the regulation by many factors, including oncogenes, tumor suppressor genes, growth factors, signal transduction factors, and apoptosis factors, *etc.* The origin of tumor is related to the modification of these genes. Genes such as *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0*, are involved in the initiation and development of HCC, and the possible markers for the diagnosis of HCC in clinic. To identify whether these genes were generally involved in hepatocarcinogenesis, multiplex PCR was used in the present study. We found that these genes had an upregulated expression in HCC and multiplex PCR could enhance the detective positive rate.

### *Cyclin D1*

Cyclin, cyclin-dependent kinases, and tumor suppressor gene

products interact and regulate the normal cell cycle. *Cyclin D1* and cyclin-dependent kinases are required for completion of the G<sub>1</sub>/S transition in normal mammalian cells<sup>[8]</sup>. *Cyclin D1* is located on chromosome 11q13 and exhibits many characteristics of cellular oncogenes. Overexpression of *cyclin D1* may be associated with actual gene amplification or transcriptional dysregulation in cancer. *Cyclin D1* is overexpressed in hyperplastic lesions, such as endometrioid adenocarcinoma<sup>[9]</sup>, mantle cell lymphoma<sup>[10]</sup>, and ovarian carcinoma<sup>[11]</sup>. The results in our study showed that the expression of *cyclin D1* was significantly higher in HCC than in adjacent nontumorous tissue. The mechanism of *cyclin D1* dysregulation in HCC is not clear, but it is likely that the dysregulation contributes to increasing the proportion of cells in transition from G<sub>1</sub> to S phase. The overexpression of *cyclin D1* may be one of the several mechanisms involved in hyperplasia of liver cells.

### *DEK*

*DEK* is a 43-ku phosphoprotein that was first isolated as part of a fusion protein expressed in a subtype of acute myeloid leukemias with t(6;9) chromosomal translocations<sup>[12]</sup>. *DEK* was lately identified as an autoimmune antigen in patients with pauciarticular onset juvenile rheumatoid arthritis, systemic lupus erythematosus<sup>[13]</sup>, and other autoimmune diseases. In recent study, it was demonstrated that *DEK* was a site-specific DNA binding protein that was involved likely in transcriptional regulation and signal transduction and it had implications not only for HIV-2 transcription<sup>[14]</sup> but also for multiple cellular processes involving with *DEK*. Recent data demonstrated that the major portion of *DEK* is associated with chromatin

**Table 2** Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in well- and poorly-differentiated HCC

	Cyclin D1		GPC3		DEK		IGF-II		rpP0	
	+	-	+	-	+	-	+	-	+	-
Well-differentiated HCC	6	3	6	3	8	1	6	3	7	2
Poor-differentiated HCC	22	1	18	5	17	6	22	1	19	4
Total	28	4	24	8	25	7	28	4	26	6

**Table 3** Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC with AFP negative and positive HCC tissue

	<i>n</i>	Cyclin D1+	GPC3+	DEK+	IGF-II+	rpP0+
AFP(-)	10	8	9	9	9	9
AFP(+)	22	20	15	16	19	17
Total	32	28	24	25	28	26

$\chi^2$  test,  $P>0.05$ , the group of APF(+) vs the group of APF(-).

**Table 4** Expression of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* mRNAs in HCC and adjacent noncancerous liver tissues by multiplex RT-PCR

Groups	<i>n</i>	Positive reaction		Negative reaction	
		Anyone of the five mRNAs (%)	All of the five mRNAs (%)	Anyone of the five mRNAs (%)	All of the five mRNAs (%)
T	32	32 (100)	22 (68.8)	10 (31.3)	0 (0)
N	32	24 (75.0)	3 (9.4)	29 (90.6)	8 (25.0)
		$P>0.05$	$P>0.05$	$P>0.05$	$P>0.05$

*in vivo* and suggested that it might play a role in chromatin architecture<sup>[15]</sup>. Our present experiment shows that the percentage of overexpression of mRNA of *DEK* (78.1%) in HCC was higher than in adjacent nontumorous tissues (15.6%). It indicates the overexpression of *DEK* may be involved in the transformation from normal liver tissue to HCC, perhaps by activating the oncogenes.

### GPC3

*GPC3* gene is located at Xq26, and mutated in the Simpson-Golabi-Behmel syndrome<sup>[16]</sup>. It may be regulated by methylation of the inactive X in expressing tissues, and encodes a developmentally regulated heparin sulfate proteoglycan that is bound to the cell surface through a glycosylphosphatidylinositol anchor. Based on their localization on the cell surface, such glypicans are thought to regulate interactions between growth factor and their receptors. It is associated with apoptosis and cell signal transduction. It is reported that *GPC3* is a tissue-specific gene in breast tumor<sup>[17]</sup>, ovarian tumor<sup>[18]</sup>, and malignant mesothelioma<sup>[19]</sup> in which it is downregulated by aberrant methylation of the *GPC3* promoter region, and upregulated in HCC. In the present study, *GPC3* was overexpressed in HCC tissue and the positive expression rate was 75.0%, which was higher than that in adjacent nontumorous tissue (21.9%). Although the role of overexpression of *GPC3* in the development of HCC is not known, it may break the balance between cell growth and death.

### IGF-II

It was reported that IGF-II is positive in some benign neoplastic nodules and HCC<sup>[20]</sup>. In the present study, the levels of IGF-II was higher in HCC than in adjacent nontumorous tissue, suggesting that the growth factor may act as an autocrine regulation of cell proliferation, *GPC3* may act as a positive regulator of IGF-II, although we have not detected a direct interaction between *GPC3* and IGF-II. It is possible that *GPC3* positively regulates IGF-II activity by interacting with the components of its signaling system.

### rpP0

Ribosome acts as a place for protein synthesis. It is composed of rRNA and ribosomal phosphoproteins. In the family of ribosomal phosphoprotein, there are five members: P0, P1 $\alpha$ , P1 $\beta$ , P2 $\alpha$ , and P2 $\beta$ . Three functional domains can be defined in the rpP0: one involved in binding to rRNA, one connected to P1/P2 protein interaction, and one associated with elongation factors<sup>[21]</sup>. It was reported that rpP0 expression increases in colon carcinoma cells<sup>[22]</sup>. In the present study, rpP0 was overexpressed in HCC tumor tissue, suggesting that upregulation of rpP0 is associated with HCC, which may be a signal for increasing protein synthesis.

### Multiplex RT-PCR and HCC

In this study, we used multiplex RT-PCR to detect the expression of mRNAs of *DEK*, *cyclin D1*, *IGF-II*, *GPC3*, *rpP0* in HCC and adjacent nontumorous liver tissue. We found that at least one of the mRNAs could be detected in all HCC tissues and 68.8% of HCC tissues expressed all these five mRNAs, while only 9.4% of paraneoplastic tissues

expressed all of them ( $P < 0.05$ ), suggesting that multiplex RT-PCR enhances the detective sensitivity and specificity when combining several specific primers. The expression of anyone of these mRNAs in liver tissue could be regarded as a risk factor for HCC. The higher the expression of these mRNAs the greater the risk. When all these mRNAs are negative in tissues, the possibility of HCC is lower. Multiplex RT-PCR provides an easy and quick method to detect the expression of these genes in liver tissues.

### ACKNOWLEDGMENTS

The authors thank Dr. Dang-Rong Li for practical and productive technical advice regarding the RT-PCR technique and Dr. Le-Qun Li for providing useful tissues.

### REFERENCES

- 1 Park US, Su JJ, Ban KC, Qin L, Lee EH, Lee YI. Mutations in the p53 tumor suppressor gene in tree shrew hepatocellular carcinoma associated with hepatitis B virus infection and intake of aflatoxin B1. *Gene* 2000; **251**: 73-80
- 2 Park DY, Sol MY, Suh KS, Shin EC, Kim CH. Expressions of transforming growth factor (TGF)-beta1 and TGF-beta type II receptor and their relationship with apoptosis during chemical hepatocarcinogenesis in rats. *Hepatol Res* 2003; **27**: 205-213
- 3 Sell S. Mouse models to study the interaction of risk factors for human liver cancer. *Cancer Res* 2003; **63**: 7553-7562
- 4 Kondoh N, Wakatsuki T, Ryo A, Hada A, Aihara T, Horiuchi S, Goseki N, Matsubara O, Takenaka K, Shichita M, Tanaka K, Shuda M, Yamamoto M. Identification and characterization of genes associated with human hepatocellular carcinoma. *Cancer Res* 1999; **59**: 4990-4996
- 5 Azechi H, Nishida N, Fukuda Y, Nishimura T, Minata M, Katsuma H, Kuno M, Ito T, Komeda T, Kita R, Takahashi R, Nakao K. Disruption of the p16/cyclin D1/retinoblastoma protein pathway in the majority of human hepatocellular carcinoma. *Oncology* 2001; **60**: 346-354
- 6 Tsai JF, Jeng JE, Chuang LY, You HL, Ho MS, Lai CS, Wang LY, Hsieh MY, Chen SC, Chuang WL, Lin ZY, Yu ML, Dai CY. Serum insulin-like growth factor-II and alpha-fetoprotein as tumor markers of hepatocellular carcinoma. *Tumour Biol* 2003; **24**: 291-298
- 7 Sung YK, Hwang SY, Park MK, Farooq M, Han IS, Bae HI, Kim JC, Kim M. Glypican-3 is overexpressed in human hepatocellular carcinoma. *Cancer Sci* 2003; **94**: 259-262
- 8 Jirstrom K, Ringberg A, Ferno M, Anagnostaki L, Landberg G. Tissue microarray analyses of G1/S-regulatory proteins in ductal carcinoma *in situ* of the breast indicate that low cyclin D1 is associated with local recurrence. *Br J Cancer* 2003; **89**: 1920-1926
- 9 Ruhul Quddus M, Latkovich P, Castellani WJ, James Sung C, Steinhoff MM, Briggs RC, Miranda RN. Expression of cyclin D1 in normal, metaplastic, hyperplastic endometrium and endometrioid carcinoma suggests a role in endometrial carcinogenesis. *Arch Pathol Lab Med* 2002; **126**: 459-463
- 10 Howe JG, Crouch J, Cooper D, Smith BR. Real-time quantitative reverse transcription-PCR for cyclin D1 mRNA in blood, marrow, and tissue specimens for diagnosis of mantle cell lymphoma. *Clin Chem* 2004; **50**: 80-87
- 11 Raju U, Nakata E, Mason KA, Ang KK, Milas L. Flavopiridol, a cyclin-dependent kinase inhibitor, enhances radiosensitivity of ovarian carcinoma cells. *Cancer Res* 2003; **63**: 3263-3267
- 12 Maeda T, Kosugi S, Ujiie H, Osumi K, Fukui T, Yoshida H, Kashiwagi H, Ishikawa J, Tomiyama Y, Matsuzawa Y. Localized relapse in bone marrow in a posttransplantation patient with t(6;9) acute myeloid leukemia. *Int J Hematol* 2003; **77**: 522-525
- 13 Wichmann I, Respaldiza N, Garcia-Lozano JR, Montes M,

- Sanchez-Roman J, Nunez-Roldan A. Autoantibodies to DEK oncoprotein in systemic lupus erythematosus (SLE). *Clin Exp Immunol* 2000; **119**: 530-532
- 14 **Faulkner NE**, Hilfinger JM, Markovitz DM. Protein phosphatase 2A activates the HIV-2 promoter through enhancer elements that include the pTet site. *J Biol Chem* 2001; **276**: 25804-25812
- 15 **Kappes F**, Burger K, Baack M, Fackelmayer FO, Gruss C. Subcellular localization of the human proto-oncogene protein DEK. *J Biol Chem* 2001; **276**: 26317-26323
- 16 **Veugelers M**, Cat BD, Muyldermans SY, Reekmans G, Delande N, Frints S, Legius E, Fryns JP, Schrandt-Stumpel C, Weidle B, Magdalena N, David G. Mutational analysis of the GPC3/GPC4 glypican gene cluster on Xq26 in patients with Simpson-Golabi-Behmel syndrome: identification of loss-of-function mutations in the GPC3 gene. *Hum Mol Genet* 2000; **9**: 1321-1328
- 17 **Xiang YY**, Ladeda V, Filmus J. Glypican-3 expression is silenced in human breast cancer. *Oncogene* 2001; **20**: 7408-7412
- 18 **Lin H**, Huber R, Schlessinger D, Morin PJ. Frequent silencing of the GPC3 gene in ovarian cancer cell lines. *Cancer Res* 1999; **59**: 807-810
- 19 **Murthy SS**, Shen T, De Rienzo A, Lee WC, Ferriola PC, Jhanwar SC, Mossman BT, Filmus J, Testa JR. Expression of GPC3, an X-linked recessive overgrowth gene, is silenced in malignant mesothelioma. *Oncogene* 2000; **19**: 410-416
- 20 **Tsai JF**, Jeng JE, Chuang LY, You HL, Ho MS, Lai CS, Wang LY, Hsieh MY, Chen SC, Chuang WL, Lin ZY, Yu ML, Dai CY. Serum insulin-like growth factor-II and alpha-Fetoprotein as tumor markers of hepatocellular carcinoma. *Tumour Biol* 2003; **24**: 291-298
- 21 **Rodriguez-Gabriel MA**, Remacha M, Ballesta JP. The RNA interacting domain but not the protein interacting domain is highly conserved in ribosomal protein P0. *J Biol Chem* 2000; **275**: 2130-2136
- 22 **Barnard GF**, Staniunas RJ, Bao S, Mafune K, Steele GD Jr, Gollan JL, Chen LB. Increased expression of human ribosomal phosphoprotein P0 messenger RNA in hepatocellular carcinoma and colon carcinoma. *Cancer Res* 1992; **52**: 3067-3072

Science Editor Wang XL and Guo SY Language Editor Elsevier HK