

# World Journal of *Diabetes*

*World J Diabetes* 2021 October 15; 12(10): 1587-1811



**EXPERT RECOMMENDATIONS**

- 1587** Expert opinion on the preoperative medical optimization of adults with diabetes undergoing metabolic surgery  
*Bhattacharya S, Kalra S, Kapoor N, Singla R, Dutta D, Aggarwal S, Khandelwal D, Surana V, Dhingra A, Kantroo V, Chittawar S, Deka N, Bindal V, Dutta P*

**REVIEW**

- 1622** Estrogens and the regulation of glucose metabolism  
*Alemany M*
- 1655** Role of nucleic acid sensing in the pathogenesis of type 1 diabetes  
*Badal D, Sachdeva N, Maheshwari D, Basak P*
- 1674** Interactions between diabetes and COVID-19: A narrative review  
*Sabri S, Bourron O, Phan F, Nguyen LS*

**MINIREVIEWS**

- 1693** Diabetes and gut microbiota  
*Xi Y, Xu PF*
- 1704** Tale of two kinases: Protein kinase A and Ca<sup>2+</sup>/calmodulin-dependent protein kinase II in pre-diabetic cardiomyopathy  
*Gaitán-González P, Sánchez-Hernández R, Arias-Montaño JA, Rueda A*
- 1719** Glycemic targets in critically ill adults: A mini-review  
*See KC*
- 1731** Galectin-3 possible involvement in antipsychotic-induced metabolic changes of schizophrenia: A minireview  
*Borovcanin MM, Vesic K, Jovanovic M, Mijailovic NR*

**ORIGINAL ARTICLE****Basic Study**

- 1740** Medication adherence and quality of life among type-2 diabetes mellitus patients in India  
*Mishra R, Sharma SK, Verma R, Kangra P, Dahiya P, Kumari P, Sahu P, Bhakar P, Kumawat R, Kaur R, Kaur R, Kant R*
- 1750** Metabolic and inflammatory functions of cannabinoid receptor type 1 are differentially modulated by adiponectin  
*Wei Q, Lee JH, Wu CS, Zang QS, Guo S, Lu HC, Sun Y*

**Case Control Study**

- 1765** Diabetic kidney disease: Are the reported associations with single-nucleotide polymorphisms disease-specific?

*Saracyn M, Kisiel B, Franaszczyk M, Brodowska-Kania D, Źmudzki W, Malecki R, Niemczyk L, Dyrla P, Kamiński G, Płoski R, Niemczyk S*

**Retrospective Cohort Study**

- 1778** Utility of oral glucose tolerance test in predicting type 2 diabetes following gestational diabetes: Towards personalized care

*Bayoumi RAL, Khamis AH, Tahlak MA, Elgergawi TF, Harb DK, Hazari KS, Abdelkareem WA, Issa AO, Choudhury R, Hassanein M, Lakshmanan J, Alawadi F*

**Retrospective Study**

- 1789** Diabetes patients with comorbidities had unfavorable outcomes following COVID-19: A retrospective study

*Luo SK, Hu WH, Lu ZJ, Li C, Fan YM, Chen QJ, Chen ZS, Ye JF, Chen SY, Tong JL, Wang LL, Mei J, Lu HY*

**LETTER TO THE EDITOR**

- 1809** Non-alcoholic fatty liver disease, diabetes medications and blood pressure

*Ilias I, Thomopoulos C*

**ABOUT COVER**

Editorial Board Member of *World Journal of Diabetes*, Sze M Ng, MBBS, FHEA, FRCPCH, SFFMLM, MSc, LLM, MBA, PhD, Associate Professor, University of Liverpool, Consultant Paediatric Endocrinologist, Southport & Ormskirk NHS, Ormskirk L39 2AZ, United Kingdom. may.ng@nhs.net

**AIMS AND SCOPE**

The primary aim of *World Journal of Diabetes* (*WJD*, *World J Diabetes*) is to provide scholars and readers from various fields of diabetes with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

*WJD* mainly publishes articles reporting research results and findings obtained in the field of diabetes and covering a wide range of topics including risk factors for diabetes, diabetes complications, experimental diabetes mellitus, type 1 diabetes mellitus, type 2 diabetes mellitus, gestational diabetes, diabetic angiopathies, diabetic cardiomyopathies, diabetic coma, diabetic ketoacidosis, diabetic nephropathies, diabetic neuropathies, Donohue syndrome, fetal macrosomia, and prediabetic state.

**INDEXING/ABSTRACTING**

The *WJD* is now abstracted and indexed in Science Citation Index Expanded (SCIE, also known as SciSearch®), Current Contents/Clinical Medicine, Journal Citation Reports/Science Edition, PubMed, and PubMed Central. The 2021 Edition of Journal Citation Reports® cites the 2020 impact factor (IF) for *WJD* as 3.763; IF without journal self cites: 3.684; 5-year IF: 7.348; Journal Citation Indicator: 0.64□Ranking: 80 among 145 journals in endocrinology and metabolism; and Quartile category: Q3.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: Lin-Yu Tong Wang; Production Department Director: Yu-Jie Ma; Editorial Office Director: Jia-Ping Yan.

**NAME OF JOURNAL**

*World Journal of Diabetes*

**ISSN**

ISSN 1948-9358 (online)

**LAUNCH DATE**

June 15, 2010

**FREQUENCY**

Monthly

**EDITORS-IN-CHEF**

Lu Cai, Md. Shahidul Islam, Jian-Bo Xiao, Manfredi Rizzo

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/1948-9358/editorialboard.htm>

**PUBLICATION DATE**

October 15, 2021

**COPYRIGHT**

© 2021 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION ETHICS**

<https://www.wjgnet.com/bpg/GerInfo/288>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

Submit a Manuscript: <https://www.f6publishing.com>*World J Diabetes* 2021 October 15; 12(10): 1622-1654DOI: [10.4239/wjd.v12.i10.1622](https://doi.org/10.4239/wjd.v12.i10.1622)

ISSN 1948-9358 (online)

REVIEW

## Estrogens and the regulation of glucose metabolism

Marià Alemany

**ORCID number:** Marià Alemany  
0000-0002-9783-8293.**Author contributions:** Alemany M is the only author.**Conflict-of-interest statement:** The Author declares no conflict of interests for this article.**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>**Manuscript source:** Invited manuscript**Specialty type:** Endocrinology and metabolism**Country/Territory of origin:** Spain**Peer-review report's scientific quality classification**

Grade A (Excellent): 0

Grade B (Very good): B

Grade C (Good): C

Grade D (Fair): 0

**Marià Alemany**, Faculty of Biology, University of Barcelona, Barcelona 08028, Catalonia, Spain**Corresponding author:** Marià Alemany, PhD, Emeritus Professor, Faculty of Biology, University of Barcelona, Av. Diagonal, 643, Barcelona 08028, Catalonia, Spain.  
[malemany@ub.edu](mailto:malemany@ub.edu)

### Abstract

The main estrogens: estradiol, estrone, and their acyl-esters have been studied essentially related to their classical estrogenic and pharmacologic functions. However, their main effect in the body is probably the sustained control of core energy metabolism. Estrogen nuclear and membrane receptors show an extraordinary flexibility in the modulation of metabolic responses, and largely explain gender and age differences in energy metabolism: part of these mechanisms is already sufficiently known to justify both. With regard to energy, the estrogen molecular species act essentially through four key functions: (1) Facilitation of insulin secretion and control of glucose availability; (2) Modulation of energy partition, favoring the use of lipid as the main energy substrate when more available than carbohydrates; (3) Functional protection through antioxidant mechanisms; and (4) Central effects (largely through neural modulation) on whole body energy management. Analyzing the different actions of estrone, estradiol and their acyl esters, a tentative classification based on structure/effects has been postulated. Either separately or as a group, estrogens provide a comprehensive explanation that not all their quite diverse actions are related solely to specific molecules. As a group, they constitute a powerful synergic action complex. In consequence, estrogens may be considered wardens of energy homeostasis.

**Key Words:** Estrogens; Insulin; Estrogen receptors; Energy metabolism; Glucose; Antioxidants; Metabolic syndrome

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Estrogens play a paramount and continued regulatory role, based on the synergy between the different forms of estrogen to maintain energy (and lipid/glucose) homeostasis. These functions include preventing: oxidative damage, lipid-induced inflammation, excess fat accrual and the complications of excess amino nitrogen. This short incomplete list is fairly close to a recipe for preventing the development of metabolic syndrome; abundant epidemiological and (partial) experimental data help

Grade E (Poor): 0

**Received:** January 22, 2021**Peer-review started:** January 22, 2021**First decision:** March 1, 2021**Revised:** March 10, 2021**Accepted:** April 14, 2021**Article in press:** April 14, 2021**Published online:** October 15, 2021**P-Reviewer:** Kung WM, Navarro-González JF**S-Editor:** Zhang L**L-Editor:** Webster JR**P-Editor:** Wang LY

support this assertion. We have to look more widely at estrogens (the different structural-functional types described in the text) to understand their extensive and powerful control of energy homeostasis.

---

**Citation:** Alemany M. Estrogens and the regulation of glucose metabolism. *World J Diabetes* 2021; 12(10): 1622-1654

**URL:** <https://www.wjgnet.com/1948-9358/full/v12/i10/1622.htm>

**DOI:** <https://dx.doi.org/10.4239/wjd.v12.i10.1622>

---

## INTRODUCTION

### ***The complex and growing implication of steroid hormones in homeostasis***

Steroid hormones are derived from sterols, which play a critical role in the structure of the (mainly) eukaryotic membrane[1]. Most steroid hormones regulate animal functions, especially in vertebrates. Only recently, plant steroid hormone analogs such as the brassinosterols[2] have been found to play a significant role in the metabolic regulation in the most evolved plants[3,4]. Other steroids, including estrogens have also been found to act as regulators in some plants, but the information is still relatively scarce[5,6]. In addition, a number of higher plants are able to synthesize animal steroid hormones, such as testosterone and estrogens[7], as well as structural analogs which interfere with physiological functions and vital cycles of some animals [8,9], including direct allopathic interference[10]. The steroid hormone ecdysone is critical for molting of insects and other animals[11], and plants synthesize the analogs ecdysteroids to limit insect development[12].

Nevertheless, the most studied types of steroid hormones (estrogen, androgen, corticosteroid, progesterone) are typical and characteristic of vertebrates. However, the variability in functions, regulation and even mechanism of action is considerable, since the degree of implication of these hormones in the fundamental aspects of life: reproduction, feeding-growth-metabolism, neural and metabolic regulation, fall squarely in their fine tuning of life cycles, survival and evolution. Their effects, largely gene expression modulation, are being continuously uncovered, in a way that deviates from the classical distribution of functions for human steroid hormones, often presented as mere sex-definition signals or regulators of mineral and glucose homeostasis. The implication of all of these hormones in the defense mechanisms (including a massive implication in the immune system control[13,14] and optimization of metabolic function[15,16]) has been growing in importance in parallel to their development along the evolution path[17] that brought humans to their amazing homeostatic resilience.

This linear review is focused on estrogens, one of the most important vertebrate steroid hormone types, due to their critical function on the control of core metabolic partition in addition to their fundamental immune system control and reproductive functions.

### ***The estrogens are not only “sex hormones”***

Most of the investigations of estrogen effects on metabolic regulation, irrespective of sex, are fairly recent and notably skewed (Box 1). So far we have only limited information on the major role played by different forms of physiological estrogens in the control of energy metabolism at the whole body level[16,18].

The initially intense development of research on estrogens came to a climax by the mid XX<sup>th</sup> century[19,20], and was essentially focused on their pharmacology, as part of the development of combined estrogen-progestogen preparations for safe birth control in humans[21]. The studies on steroid hormones were not limited to estrogens (and progestogens), but were extended to androgens[22] and, especially, to glucocorticoids [23] through the development of a large number of synthetic drugs, widely used and which their development continues[24]. In a way, this expansion in the pharmacology of steroid hormones also provided considerable information on their mechanisms of action[25] and metabolism[26], including an extensive analysis of some possible complications of their clinical use[27-29]. Nowadays, the natural human corticosteroids (cortisol and cortisone, but also corticosterone[30]) are seldom prescribed, despite showing often quite different effects and pharmacological profile than the

myriad of synthetic corticosteroids in use[31]. The latter may bind to most of the natural receptors[32], but basically do not share the transporter proteins[33] or the inter-organ self-regulatory mechanisms of natural hormones (e.g. the hypothalamus-pituitary axis).

The common identification of “estrogen” with 17 $\beta$ -estradiol (E2) and “androgen” with testosterone (T) is an inadequate oversimplification that helps to dismiss the regulatory and fine-tuning interrelationships of the different molecular species of estrogens, both with themselves or with androgens and other steroid hormones.

The estrogens are ancient regulatory agents, remarkably preserved along evolution. The number and structure of relevant molecules remains small and unaltered in spite of the variety and complexity of the mechanisms modulating their actions, somehow reflecting the cumulative experience (and expansion of metabolic interventions) acquired during evolution. The versatility of the nuclear receptors’ modulation and signaling pathways allow the superposition of a dense web of signals, including fail-safe, duplicate, alternative and redundant mechanisms, which often make it difficult to find answers to the direct questions relevant to the clinicians.

### **The structures of estrogens**

The principal distinguishing feature of animal estrogens is the phenolic nature of ring A, usually with a -OH in C3. No other type of steroid hormone contains a phenolic ring. The steroid nucleus of estrogens has 18 C, and lack a side chain. The main human functional estrogens are 3-hydroxy-17-keto-estrin (E1 estrone), 17 $\beta$  estradiol or 3, 17 $\beta$ -dihydroxy-estrin (E2 estradiol) and 3, 16 $\alpha$ , 17 $\beta$ -trihydroxy-estrin (E3 estriol); during fetal development[34], another estrogen should be included: 3, 15 $\alpha$ , 16 $\alpha$ , 17 $\beta$ -tetrahydroxy-estrin (E4 estetrol). Compared with all other mammalian steroid hormones, they are highly lipophilic (E1 > E2 > E3). This peculiarity facilitates their transport by lipoproteins[35] and binding to membranes[36] (including their crossing). The interactions with lipophilic entourages have been credited as a main factor for their effects on membranes[37] and mitochondrial function[38]. It is often assumed that estrogens are carried in the blood bound to proteins, largely sex hormone-binding globulin (SHBG), but the higher affinity and metabolic response to energy changes[39] of T (competing with E2 for SHBG) favors a closer dependence of the globulin levels and/or structure/affinity[40,41]. The much lower levels of E2 than T in plasma (both in women and men)[33] suggest that this dual (if real) transport of hormones may be a consequence of modulation of the molecular affinity of SHBG, in part through modification of its molecular weight[39,40,42]. The key factor is that under physiological conditions SHBG (or a varied group of SHBG isoforms) binds essentially T[39] and estrogen (almost 90% of plasma E2, but practically no E1[33]). In addition, the *in vitro* estrogenicity of E1 is considerably lower than that of E2[43]. This fact, together with the abundance of E1 in men (despite being an estrogen), and its high lipophilicity resulted in a limited pharmacological interest for this molecule and a consequent lack of literature on it, and its function as a free hormone remains obscure. E1 is the most abundant estrogen in the body (when its esterified forms are included) [44], since it is produced (and stored[44]) in large amounts in white adipose tissue (WAT)[45]. Probably because of its lipophilicity, a large portion of E1 in plasma is found esterified as sulfate, much more soluble than the free hormone[46,47], which facilitates its transport and eventual excretion. However, E1 can be made even more lipophilic by esterification with a fatty acid on C3[48] yielding acyl-estrone (AE1). In this form it has been found in lipoproteins[49] and adipose tissue[44]. AE1 are synthesized by adipocytes and modulated by leptin and insulin[50].

E2 is also esterified with fatty acids (acyl-E2 or AE2), becoming more lipophilic than E2, and thus also found in blood lipoproteins[51,52]. However, AE2 is largely esterified in C17 and not in C3 as are the AE1 esters[53]. This peculiarity of AE2 has been attributed to its higher capability to protect the lipids which surround the hormone from oxidation thanks to the unaffected phenolic -OH[51]. In any case, the highly lipophilic estrogens (through esterification with long-chain fatty acids) are a common occurrence for which no definitive function has yet been fully agreed upon, and which shows that the usual molecular species of natural estrogens, their transport in the bloodstream including their binding and physiological functions are far from being fully known. The high concentration of these varied estrogen-acyl-ester molecules in tissues, such as WAT[54] suggests a possible role of reserve or storage of preformed estrogenic molecules[53], which has been explained in part by the easiness of their synthesis by acyl-transferases, widely present in lipoproteins and tissues[55].

### Main gender differences in human estrogen function

The scant number of in-depth non-clinical or pharmacological studies may be in part a consequence of the bias against estrogens (and of their bad name, Box 1). The reasons usually presented to sustain negative opinions, (which in the end limit metabolic analyses, and the eventual therapeutic use) are based largely on two factors: their known role as promoters of some forms of cancer, mainly breast[56] and endometrial [57], and a number of risks derived from their use[58,59] other than their assumed role as “female-linked” hormones. The essentiality of estrogens has been proven (in both sexes) for many functions, such as those related with sex differentiation and reproduction, as well as bone health[60]; but the key factor is the increasing flow of data that establishes a direct implication of estrogens in the control of the basic core of energy metabolism[16,61]. This control is affected by age, sex and diet; thus, the simpler division of steroid hormones function using a strict sex-oriented focus is no longer applicable. We simply need to know more about the estrogens and their functions, in exactly the same way as any other hormone, keeping in mind the species-specificity in some of their functions when establishing comparisons with animal models (Box 2).

Women have higher circulating levels of E2 than men, from puberty to menopause, with notable variation between physiological situations[62]. Men, even at their maximal reproductive capacity age, also show fairly high blood levels of E2[63]. There are not enough data on AE2 levels and distribution to establish valid comparisons, but it is probable that the parallelism will be maintained. On the other side, seldom clear gender differences are found in E1, the most abundant estrogen (free or esterified) in human blood. E1-sulfate (SE1) is subjected to a regulative “solubility/excretion” cycle [46] comparable to that of dehydroepiandrosterone[64]. The ample abundance of AE1 in tissues (rat) shows a more marked dependence on the mass of WAT than on sex[65].

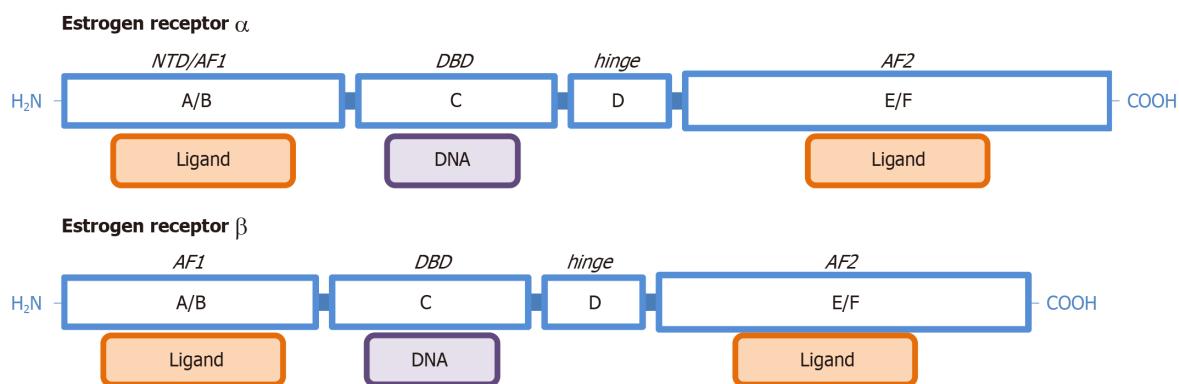
T and E2 differently influence brain development from its earliest stages, both in the setting of its functional structure and -later- its psychological orientation and focus[66, 67]. The resilience of women against insulin resistance is higher than that of men[68, 69], at least until menopause[69]. Estrogen protects bone from demineralization in women and men[70,71], a function in part shared by T (at a lower potency, however [72]). The accrual and maintenance of body protein falls largely on androgens, mainly T[73,74], acting in a synergic way with growth hormone[75] and insulin[76] and countering the proteolytic capability of glucocorticoids[77]. The contribution of free estrogens to the maintenance of body protein mass seems to be more limited[78].

### **Estradiol signaling: classical nuclear receptors. Estrogen receptors $\alpha$ and estrogen receptors $\beta$**

Thanks to their lipophilic nature, E2 (and E1) can easily cross membranes and bind specific estrogen receptors (ER) within the cell[79]. After dimerization[80] they are brought to the nucleus, where the complex E2-ER binds to deoxyribonucleic acid (DNA)[81] or to specialized proteins[82,83], eventually eliciting the expression or repression of specific genes. The nuclear-type estrogen receptors are highly complex [79,84]. Estrogen signaling, up to its final manifestation is not a fast process such as that of nervous or rapid-signaling chemical regulating agents (Box 3).

Binding to ER is, essentially, specific for the physiological estrogens[75,85], but a wide number of plant secondary metabolism compounds, synthetic non-steroidal estrogenic drugs and even some toxic industrial waste also bind the ER[86]. In humans, there are two main types of nuclear estrogenic receptors: estrogen receptors  $\alpha$  (ER $\alpha$ ) and estrogen receptors  $\beta$  (ER $\beta$ )[87]. In fact, ER $\alpha$  and ER $\beta$  are two families of related receptors, which maintain the same overall structure but not their complete sequence, the ERs being adapted, adjusted or changed for best effectiveness, in different tissues or because of changing needs[84,88].

The structures of ER $\alpha$  and ER $\beta$  are shown in Figure 1. The main dominions are marked with letters (A to F), and correspond roughly (A/B) to a zinc finger and a binding site, activation function site 1 of the ERs (AF1); C is the place for binding estrogen-response elements (ERE) and then DNA[89]; D is a shorter sequence related to the binding of chaperone proteins and to the process of dimerization; and E/F is the ligand binding domain for estrogens and other factors, AF2[84]. The main ligands are the natural estrogens of mammals (E2, E1, E3 and E4), but some drugs, phytoestrogens [10], metals and diverse chemicals (xenoestrogens) can also bind the receptors[90]. Binding to the AF1 and AF2 may result in synergistic effects[91]. The length and distribution of ER parts may change within each receptor family depending on alternate sequences and splicing[84]. The affinity of ER $\alpha$ : is maximal for E2, followed by E1, and that of ER $\beta$  is also E2, followed by E3.



**Figure 1 Functional structure of estrogen receptors  $\alpha$  and estrogen receptors  $\beta$ .** Domains names (A to F), and common denominations for the main functions are included: N-terminus dominion; DNA binding dominion, activation function site 1 of the estrogen receptors and activation function site 2 of the estrogen receptors. The graph is not drawn to scale and represents the complete (highest molecular weight) form for each of the two families of nuclear ERs. NTD: N-terminus dominion; DBD: DNA binding dominion; AF1: Activation function site 1 of the estrogen receptors; AF2: Activation function site 2 of the estrogen receptors.

The combination of affinities and the panoply of modulators and cell type-specific distribution of ERs results in an extended variety of possible effects, making widely variable the action of estrogens, in order to send specific signals to organs or groups of cells within a wide array of possibilities[92-95]. The ERs are, perhaps one of the best examples of receptor adjustment to the needs of tissues under varying conditions, attained through a considerable number of mechanisms. Both ER $\alpha$  and ER $\beta$  are dimeric and coded by different genes[96,97], with an additional abundance of polymorphisms[98,99]. Their distribution in the cells of different tissues and organs is independent for each receptor[100], as are their final gene expression effects[101,102]. They can show synergic[103] or antagonistic[104] effects, even for the same molecular species, and depend, largely, on the post-binding relationships of the E2-ER complex. This situation is further complicated by the interaction of the ER (or E2-ER) with a number of different mechanisms of modulation, such as selective estrogen receptor modulators (SERM)[105-107]; selective ER down-regulators (or degraders)[108]; and the specific ERE, which directly affect the function of ERs[109,110]. Other closely related ("orphan" up to recently) receptors participate in the regulation of critical pathways, but in many cases their relationship with the ERs remains unclear.

### Membrane estrogen receptors

In addition to the canonical nuclear ERs, estrogens also bind cell surface ERs[111-113]. As usual with ERs, the terms used to define these receptors shift between the importance given to their location, partial signaling pathway, speed of action and other considerations: membrane ER[114], non-nuclear/non-transcriptional signaling ER[115,116] non-genomic signaling ER; membrane-linked ER $\alpha$ [117,118] or both ER $\alpha$  and ER $\beta$ [112]; caveolae- or lipid-related ER[119,120]; G-protein-coupled ER (GPER) [121,122]. This list adds to the existence of alternative or non-genomic "direct" or "rapid" effects of ER stimulation in some cells, eliciting immediate responses (a shift-or development-from the main advantage of the delayed steroid hormone signaling)[123,124].

All these effects suggest that, the ER structure is essentially that described above, with two main types (families) of complete/incomplete ERs: ER $\alpha$  and ER $\beta$ , which are found in the cytosol (and nuclei) of cells, including members of the ER $\alpha$  family linked to G-proteins (GPRA1/GPR30)[122,125] attached or close to the plasma membrane.

The proven relationship of ER $\alpha$  to membrane or fat droplet-related structures may be a consequence of the adaptability of ER $\alpha$ [120]; the existence of free and fatty acid-esterified estrogen in lipid-related cell structures[126], or both. In any case, the association of ER $\alpha$  to caveolin-1[119] and then complexed with G-proteins, helps explain the presence-binding of the ER[127], and then E2 in the membrane entourage [128]. In this case, the signal may be transferred to membrane-related structures, as is the case with increasing calcium release[115]. The G protein-ER complex, upon the binding of E2 may also induce nuclear effects *via* activation of tyrosine kinases[129] and the MAP/ERK or PI3 $\alpha$ /Akt pathways[130]. The stimulated G protein-activated receptor may also signal through GPR30[131]. The activated system containing the ER also enhances adenylate cyclase activity[132] *via* phosphorylation of the cAMP response element[133]. However, these direct membrane-related mechanisms may

coexist with also faster direct translational actions of conventional ERs somewhat linked to membranes[134] or with other mechanisms hinted at but not fully disclosed yet[135,136]. This includes the presence of ER receptors within cell structures, such as mitochondria (*e.g.* ER $\beta$ )[137].

It has been established that non-genomic effects of ER bound to E2 may belong to two confluent mechanism types: direct effects elicited from the membrane and effects developed through cytosol signaling cascades and actions. Both processes are probably coincident for different cell settings. Hypothalamic inhibition of guanylate cyclase[138] and LH secretion[139], as well as increased cell migration[140] and other brain effects[141] add to the widening array of non-genomic effects of ERs. Most of these effects have lately been attributed to the ER $\alpha$  irrespective of the place of binding with E2, but the implication of ER $\beta$  has also been described[142,143].

Notwithstanding, all these receptor-related mechanisms described cannot fully explain all the biochemical effects induced by estrogen signaling[144-148], leaving ample space for the assumption of direct, *i.e.* non ER-related, involvement (largely of E2 and its C17-fatty esters, AE2)[149,150]. However, these effects have been described only in lipoproteins, other lipid masses or lipid/protein interfaces[54,151]. The direct effects of estrogens on mitochondria have been related to specific mitochondria receptors[152,153], apparently containing ER $\alpha$ , ER $\beta$ [154] and other possible binding structures[155]. Their role on mitochondrial function, however, has been found to be significant[154,156], especially in the regulation of energy providing pathways[157,158]. The possibility of estrogen direct incrustation in the lipid layer of membranes has been proposed as a way to modify their functionality[159] and enhance the E2/AE2 anti-oxidative properties in a way similar to its postulated function in lipoproteins[160,161].

### **Estrone and AE1**

E1 is a rather peculiar and resilient hormone (Box 4); we do not yet have a direct explanation for its massive synthesis and storage, since the lipophilic nature of E1 (but not that of SE1) limits its action in plasma, cell and interstitial space. Non-esterified E1 levels are related to those of E2, with E2/E1 ratios fairly stable for men (*c.* 1) and more variable for women (*c.* 1.5-2) up to menopause[162]. However, measurement of circulating estrogen is difficult, often showing poor correlations between instrumental and immunoassay results[163]. The relationship of E1 with E2 levels, in addition to sex (*and* age) is affected by diabetes/obesity[164]. Furthermore, analyses of SE1 seldom include other E1 esters nor free E1, which compartmentation (important in lipoproteins) skews E1 serum levels towards lower values. The obese show high plasma SE1 concentrations[165]. In any case, the whole-body AE1 content in rats is several orders of magnitude higher than free (and sulfate-esterified) E2[44], however, the AE1 content in obese rats is relatively lower than in normal-weight animals, despite AE1 being essentially stored in WAT[44].

The oral pharmacological administration of oleoyl-E1 to normal weight and obese rats[166,167], induces a marked decrease in fat depots[168], not dependent on the degree of obesity and diet[167,169]. The loss of fat runs parallel to the normalization of glycemia, blood lipids and other metabolic syndrome (MS) markers[170], without apparent effects of estrogenization, and irrespective of energy intake manipulation [171]. AE1 has been proposed as a ponderostat signal[170], since the excess fat is shed without accompanying metabolic disorders[170,172]. Its negative effects on humans are negligible (clinical studies, phase I, unpublished data), and the positive (*i.e.* loss of excess fat, lowered insulin resistance, absence of estrogenization) were outstanding in a single case published[172]. However, its development as a drug was abandoned because an ill-designed phase II failed to be conclusive. We have no hints as to the mechanism of AE1 signaling, other than it is synthesized in cultured adipocytes[50], and WAT stores these esters in large amounts[44,65]. AE1 treatment reduces the size of WAT lipid depots[173,174]. Natural AE1 is transported in the lipid fraction (lipoproteins) of blood[49,175]. Methodology is a critical factor for the analysis and tracing of acyl-estrogens, with disparate results; *i.e.*, it has been reported that human plasma does not contain AE1 at all[176].

The main effects of AE1 are a consequence of the structural change on the whole ester, not through the release of E1[177]. When injected, marked estrogenic effects are observed, with increased E1 and E2 levels[178]. However, oral administration of AE1 does not elicit the same signs of estrogenization[171]. A highly critical analysis of oleoyl-E1 actions on rat body weight found no significant negative effects[179]. Body protein and N balance are preserved in AE1-treated (lean and obese) rats[166,167,174,180]. There is very little information available on AE1 mechanism of action. The structure of the orally administered ester seems to be modified, with low levels in

blood plasma[49,181,182], but an unidentified derivative is present in large concentrations, maintaining the estrogen nucleus in a more hydrophilic form[181]. In liver, AE1 label can be found linked to DNA shortly after administration[181]; the effects of AE1 imply the stimulation of ER $\alpha$ [183,184]. Excess AE1 is essentially excreted as SE1 [185].

There are sufficient elements to sustain the implication of AE1 in the regulation of body weight[170], but the lack of further complete studies on its mechanism of action has prevented both its clarification and its eventual therapeutic application. No other explanation has been put forward to justify the limited estrogenic potency of E1, despite its massive synthesis in the ovary and the brain[186], and, especially (in quantitative terms) in WAT[187], with a direct relationship of its total body content and circulating levels with WAT, lowered by obesity in rodents and humans[47]. The effect of the administration of free E1 to rats induce some estrogenic effects and slightly increases body weight, effects quite different to those of its acyl derivative [177].

### ***Estrogens and the regulation of energy metabolism***

Glucose is the main energy substrate, and the main simple nutrient of human diet. Glucose is also the primary inter-organ energy substrate carried by the blood to sustain the energy needs of body cells. Carbohydrates capable of yielding glucose (or other interconvertible hexoses) are a necessary part of our diet[188,189], and for many thousands of years they have constituted the main staple of our energy intake. This role has been already addressed in depth in a previous paper[189] in which we discussed the final fate of dietary carbohydrate, protein and lipids to yield two-and three-carbon metabolites (2C, 3C) and anaplerotic four-and five-carbon (4C, 5C) molecules from proteins (when excess N could be disposed of). The common shared groups of metabolites from dietary nutrients include 2C fragments (and a smaller amount of 3C from glycerol) from fats and, essentially 3C fragments from the six-carbon (6C) hexoses. The 3C could be largely used to maintain glycemia thanks to hepatic[190], renal[191] and intestinal[192] gluconeogenesis, or simply used (pyruvate) as a source of 2C (to yield acetyl-CoA), which is largely oxidized to CO<sub>2</sub> in the mitochondria through the Krebs cycle. Most of the energy drawn from glucose is obtained from the pyruvate-lactate produced in the glycolytic pathway followed by the complete oxidation of pyruvate, as acetyl-CoA, in the Krebs cycle. The 3C fragments (essentially lactate, pyruvate, glycerol, alanine and serine) can substitute glucose as an energy substrate in many tissues, avoiding the strict control of glucose levels, and providing faster access to their energy when and if enough oxidative capability and oxygen are available[193,194]. Glucose isoforms often delay somehow the oxidation of glucose[195], and thus, the direct cell use of glucose-derived 3C fragments may speed up its catabolism. This C6→C3 massive conversion is one of the most important albeit less publicized functions of WAT[196]. The presence of excess lipids (and energy) in the diet often results in an excess of 2C fragments (mainly the result of catabolic oxidation of their polymers: fatty acids) that their oxidation becomes problematic, thus the excess of energy available facilitates their storage (often long term) as fats[189].

The inadequacy of diet composition, and especially the excess of energy from fats and carbohydrate results in the progressive metabolic disorders of MS[197] with the development of sustained hyperglycemia[198], often deriving into type 2 diabetes [199], obesity[200], altered blood lipids, with hyperlipidemia[201], deriving in endothelial inflammation[202,203] and increased cardiovascular risk[204], hepatic steatosis[205], depression[206], and increasingly functional alteration of the nervous system[207], bone[208] and practically all organ/cell systems, extended even to the microbiota[197]; and, essentially, the immune system[209,210]. The causes and effects of MS have been intensively and extensively studied, and a direct relationship has been found with diet composition and excess energy[211,212], but no effective solutions have been put forward. Medical treatment is commonly limited to increased energy expenditure and changes in type of food, and (decreased) energy intake[213-215], in most of the cases, without sufficient metabolic analyses[216]. This is complemented by the pharmacological treatment of the disorders included in the MS. The relative acceleration of the MS effects with age is more clearly observed in adult (and aging) men than in women[217,218]. This difference has been attributed to the obvious diet-driven inflammation of MS[219,220], compounded in men by the progressive decrease in the synthesis (and effects) of T, in part a consequence of aging but also by the hypoandrogenism that characterizes MS[221]. Women, from adolescence to the beginning of menopause maintain their high levels of E2 and functional hypothalamus-hypophysis-gonadal axis[62]. Menopause, aging and other causes

break this equilibrium and the levels and protective effect of estrogens wane; The E1 *vs* E2 ratio of concentrations is maintained at E2 > E1 in premenopausal adult women, changing to E1 > E2 in post-menopausal women and in men (in which there is little change with age). In both cases, E2 levels were lower in men and post-menopausal women than in adult premenopausal women[62].

In aging men, especially those with MS, treatment with T reduces to some extent cardiovascular risk[222,223] and helps maintain glycemia[224,225], but possible dangers, insufficient knowledge and scant physiological analysis have limited the extension of this therapeutic avenue[226]. Similarly, for women, substitutive estrogenization is partly effective[227-229] at menopause, but its extension has been seriously limited by the fear of possible negative consequences, as discussed in Box 1. In addition, synthetic estrogens are the most used substitute drugs despite our very limited knowledge[230] of the intricacies of their action in such complex mechanisms as those described above for E2. The case of tamoxifen (agonist/antagonist) is a clear example[231]. This generalized (albeit undeclared) ban on sex hormones extends to the use of T in women, despite the fact that both E2 and T are needed for bone[70,232] health, and T for body protein maintenance[75]. Obviously there are problems to solve, but it seems that this line of study has not been sufficiently developed for reasons not based on contrasted arguments. In the case of AF1, a line of research developed by only one research group, obtained better results than those of any previous anti-obesity drug[170,179], but the development was discontinued largely for fear of "possible" future negative findings[179].

### ***Estrogens, insulin and dietary nutrients handling***

Most of the studies on the effects of estrogen on glucose metabolism have been done using E2 (and other ER ligands). There is a very limited amount of specific information on E1 direct effects; however, SE1 was found to induce hypoglycemia in genetically obese mice *via* glucose-6-phosphatase[233]. The effects of estrogens on glucose and energy handling are mediated through four coordinated actions: (1) Protection and facilitation of insulin secretion and function in the control of glucose availability to tissues; (2) Modulation of energy partition, favoring the use of lipid as the main energy substrate when their availability is higher than that of carbohydrates; (3) Functional protection through antioxidant mechanisms; and (4) Central effects on whole body energy metabolism and homeostasis maintenance.

### ***Estrogens, insulin and glucose***

E2 protects the functionality of the pancreatic  $\beta$  cells[234,235], preventing apoptosis [236], adapting their function to insulin resistance[237], and maintaining their insulin content[238]. ER stimulation inhibits lipogenesis in the  $\beta$  cells[158], which limits the negative effects of excess lipid in the cell. The loss of the ER (nuclear and/or membrane) impairs pancreatic insulin secretion[239], which is stimulated by estrogenic signaling[240]. The lack of E2 availability also increases hepatic insulin clearance[241].

Estrogens also prevent the development of diet-induced insulin resistance[242]. The gender-dependent effects of estrogen on high-fat diet-induced insulin resistance are largely dependent on the anti-inflammatory effects of the hormone[243]. E2 increases tissue insulin sensitivity[244], and lowers insulin resistance in peripheral tissues[245], with marked differences in the effects depending on gender[243]. In female mouse adipocytes, E2 lowers inflammation (and thus insulin resistance[246]), and enhances the effects of insulin on tissues[247]. The sole activation of ER $\alpha$  AF-1 is enough to prevent obesity, liver steatosis and insulin resistance in mice[248]. However, obesity and insulin resistance seems to require E2 in addition to ER $\alpha$  and AF-2, AF1 not being essential[249].

Estrogens induce a considerable number of actions in the brain, which is also able to synthesize them[250], playing an important role in its function[251,252] and behavior [253]. E2 also interacts with serotonin to affect insulin resistance[254]. Estrogenic deprivation induces mitochondrial dysfunctions in the brain which may induce the loss of cognitive functions[255]. More complex is the long saga of the relationship of estrogen in the peculiar placing of brain insulin resistance in Alzheimer's disease[256, 257]. The neuroprotective actions of estrogens[258], added to the inhibition by E2 of  $\beta$ -amyloid production[259] and the implication of E2 in the regulation of insulin degradation in the brain[260], suggest an overall beneficial effect of estrogen limiting the development of this disease. However, Alzheimer's disease affects more women than men[261], and a number of caveats have been raised against the danger of natural estrogens being implicated in its development[262]. Right now the case is not solved, with studies showing a protective effect of ER $\beta$ [263] and others hinting at the

implication of ER $\alpha$  in its pathology[264].

Estrogens, largely E2, facilitate the uptake of glucose from the intestine[265], and its extraction from the bloodstream by activation of transporters GLUT4[266] and, at least in the brain[267] GLUT1. E2 lowers liver glucose output with no changes in glycogen during mild exercise[268], a difference due in part to a modifiable maintenance/inhibition of gluconeogenesis[269]. E2 also regulates glycolysis in endothelial cells by non-genomic pathways[270], partly by increasing insulin signaling[271]. Glucose catabolism is affected by estrogens, which stimulate glycolysis *via* phosphofructokinase[272], and the pentose phosphate pathway *via* Akt[273]. In any case, the direct incidence of ER signaling on glucose handling is relatively limited and conducted *via* modulation of insulin[271]. Probably, the main effect of estrogen may be the utilization of lipids as alternative energy substrates. This is important for humans, because of the common occurrence of excess lipids (and energy) in Westernized diets, which leads to problems in dietary substrate partition[189] and the common development of MS.

### **Lipid handling and estrogens**

**Estradiol:** Estrogens lower circulating triacylglycerols (TAG) favoring their transport with a higher expression of ApoA5[274], and protecting lipoproteins against oxidation [275]. However, the main effect of E2 on lipids is favoring the shift from lipid deposition (storage) to its oxidation as energy substrate. Perhaps this is the most critical effect of estrogens on energy partition.

Treatment with E2 decreases obesity[276], protects against hepatic steatosis[277], lowers the activity of cholesterol acyl-transferase[278] and limits fat deposition[279]. All these are -again- indirect actions aimed to decrease the storage of excess TAG, since E2 does not directly regulate lipolysis[280]. Nevertheless, estrogens decrease lipogenesis[281] in WAT; and adipogenesis is also inhibited through ER $\alpha$  activation [282].

Dietary composition directly affects the substrate partition and the regulation of substrate utilization to maintain both energy and nutrients homeostasis[283]; in rats, hyperlipidic diets induce increases in E2 levels, and are correlated with an increased use of fatty acids as energy substrate[284]. The decrease in lipogenesis/adipogenesis (and the relatively enhanced lipolysis) frees the use of excess glucose and glycolytic 3C towards 2C and its oxidation in mitochondria; thus, decreasing the synthesis and storage of fatty acids (and TAG).

The effects of E2 on lipid handling are coordinated with the actions of E2 on insulin [235,285], glycemia[286] and the use of glucose as the direct energy substrate[271] instead of using it to fuel the synthesis of fatty acids. E2 lowers insulin resistance and fat storage through the ER $\alpha$  and the FA2 binding site[249]. Estrogens also lower the insulin resistance induced by excess dietary lipids[245].

A key point of these E2-derived metabolic shifts lies on the mitochondria[287,288]. Estrogen controls mitochondrial biogenesis and function[289]. E2 deprivation induces mitochondrial dysfunction and insulin resistance, which may induce alterations in the cognitive ability of subjects[255]. E2 potentiates the oxidative capacity of mitochondria, through increases in cAMP and cytochrome C oxidase activity[290]. E2 also inhibits the synthesis of adenosine triphosphate (ATP) in the mitochondria[291], which may be related to the increase in oxygen consumption and energy expenditure elicited by E2 [292], and its postulated role enhancing heat production and thermogenesis[293,294], which imply a higher overall substrate oxidative activity.

The pyruvate dehydrogenase complex (PDH) is a critical control node, which catalyzes the irreversible conversion of 3C pyruvate to 2C acetyl-CoA in the mitochondrion. The main mechanism of PDH control is phosphorylation, mainly by (inhibiting) PDH kinase 4 (PDHK4)[295]. Insulin inhibits the expression of PDHK4 [296], which has an increased activity during starvation and diabetes[297]. The levels of PPAR $\gamma$  coactivator-1  $\alpha$  (PGC-1), an important cell energy regulator[298,299], which is also increased during diabetes and starvation, modulates the function of ERs[300]. PGC-1 increases hepatic gluconeogenesis through the expression of phosphoenolpyruvate-carboxykinase[301], and co-activates, with estrogen-related receptor (ERR), the expression of glucokinase[302]. PGC-1 increases the expression of PDHK4 [303], essentially through the activation of ERR (mainly the ERR $\alpha$  and ERR $\gamma$  isoforms) [304]. ERRs are homologous to the nuclear ERs, but they are orphan receptors, *i.e.* do not have specific ligands such as E2[305]. Recently, it has been suggested that PGC-1 could be considered, perhaps, their unique non-steroid ligand[306]. ERRs increase glycolysis and glucose uptake[307].

The activation of PDHK4 by ERRs and PGC-1 is inhibited by insulin[303]. However, E2 activates ERRs[308]. In sum: insulin activates PDH, which is inhibited by ERRs modulated by cell lipid energy conditions (PGC-1) in a way that facilitates a decrease

in insulin resistance[309] and a steady flow of 3C to 2C into the mitochondria to fuel the Krebs cycle, since lipogenesis is inhibited[281,310] and cannot absorb the newly formed acetyl-CoA.

Further stimulation of mitochondrial oxidative capacity[153,311], and the availability of 4C and 5C derived from amino acids, further speeds up the oxidation of 2C by the mitochondria of liver, WAT and specific brain sites[312]. The accessibility of amino acid hydrocarbon skeletons depends on their increased oxidation (when in excess and limited capacity of the Krebs cycle[189]) *via* the alternative oxidation of amino groups to nitrogen gas[222,313]. The presence of these anaplerotic fragments and the higher oxidative capacity markedly increase the use of acetyl-CoA (from fatty acids or glucose) as the main energy substrate. The added relative inefficiency in the production of ATP[291] further helps the estrogen-controlled metabolism of adult women to dispose (albeit partially) of unwanted excess dietary energy. This effect may account in part for the resistance of women to develop the MS in its double facet of obesity and diabetes[314].

### **Estrone and acyl-estrone**

E1 has not generated as much literature as E2, but this is probably a consequence of its limited direct effects on classical estrogenicity and energy metabolism. However, it has been found that SE1 also contributes to glucose homeostasis, inhibiting glucose-6 phosphatase under conditions of hyperglycemia[233]. SE1 also lowers the levels of lipoproteins in postmenopausal women[315]. And, obviously shows estrogenic effects when given in pharmacological doses, albeit less marked than those of E2.

The anti-obesity effects of oleoyl-estrone, an AE1 ester, were studied extensively for a short time[170], but ceased before the appearance of many key studies on estrogen function and mechanism of action cited above. Thus, these older studies have to be re-analyzed from the present-day perspective. The acyl moiety of AE1 comparatively affects only partially its slimming effects[48], thus, oleic acid (the most abundant in the rat body stores) was used as standard. The E1 moiety, surprisingly, is not essential either, since both AE2 (at pharmacological levels) and oleoyl-diethyl-stilbestrol show marked body fat slimming effects[48]; however, these compounds have not been studied further because of the marked estrogenic response (toxic at the pharmacological levels analyzed) they elicited in comparison with AE1 or even E1 alone.

AE1 is not estrogenic[171]. However, the injection of liposomes loaded with AE1 induces estrogenic effects in rats due to the large amount of E1 produced by its hydrolysis[178]. The oral administration of AE1 basically excludes most of the E1[316] formed in the intestinal hydrolysis of AE1. In any case, SE1 is, finally, the main catabolite of AE1[185] (and of E1), since the fatty moiety poses no problems to its complete oxidation. This way, the interference of estrogenic effects has been circumvented simply by oral (instead of *i.v.*) administration of AE1, allowing a more direct analysis of its effects on body lipids[167,169,173]. The administration of AE1 preserves body protein[167,169,317], but markedly decreases body fat through the maintenance of a negative energy balance[167,169,177,180,317]. The process is achieved by lower food intake and unchanged thermogenesis[180] (an effect partly shared by E2 but not by E1), as well as a shift in the management of dietary fat, from accrual to oxidation for energy[88,318], such as described for E2. The effects of additional reduction of food availability are comparable (and additive) to those due to decreased appetite elicited by AE1[317]. Circulating levels of AE1 are proportional to body fat[65,182]. However, obese rats have lower AE1 levels than their lean counterparts[44]. In sum, the main effect of AE1 (given at pharmacological levels) is to shed excess body fat, without additional metabolic interference[174]. AE1 circulating levels presumably act as an indicator of whole body fat reserves under normal (not MS) conditions[319]. The AE1-induced loss of body TAG implies the concordance (described for E2) of peripheral (especially WAT) lipolysis[320], decreased lipogenesis[321] and higher energy expenditure and lipid oxidation[318].

The effects of AE1 on glucose metabolism are comparable to those described for E2: regularize hyperglycemia[174], decrease insulin resistance[174] and an overall antidiabetic action[322]. However, these effects may be just the consequence of the normalization of energy homeostasis induced by pharmacological doses of AE1[170], with full activation of the estrogen shift explained above: increased mitochondrial oxidation of 2C (and excess 3C) instead of storage of excess 2C mainly in the form of TAG-fatty acids.

The similarities of AE1 with E2 are both quantitative and qualitative. The well-known summarization of E2 function as less estrogenic than neuroprotective[323] is not applicable to the comparison with AE1 because these esters do not show either of these functions. Nevertheless, injected AE1 label has been found in cell nuclei[181],

and AE1 binds the ER $\alpha$ [183,184], but E2 cannot displace AE1 from its binding[171]. In addition, the pharmacological effects of E2 and AE1 are not superimposable[177]. This is compounded by the lack of full inhibition of AE1 actions on rodents by tamoxifen [324] and fulvestrant[184] (in fact, tamoxifen mimicked some of the effects of AE1[324] ). These data help finally differentiate the effects of AE1 from both E1 and E2, and suggest that AE1 is, probably a SERM.

### **Functional protection through antioxidant mechanisms**

E2 and E3 (but not E1) are considered effective antioxidants[325], since they help protect structural lipids from free radicals[326]. The polarized structure of estrogens makes them ideally suited to interact in interfaces between hydrophilic and lipophilic media[327], such as membranes, including mitochondria[160]. In this aspect, perhaps the AE2 esters may be the most effective, because in addition to E2, their most common acyl moiety, linoleic acid[328], is itself a main component of membranes [329], albeit being easily oxidized by free radicals[330]. The AE2 have been described as powerful antioxidants, more effective than free E2[331]. This role includes mitochondria, closely related to estrogen action for increased numbers, oxidative capacity, metabolic function and survival[161]. In this sense, both E2 and AE2 (and, probably to a lesser extent E1), control[332] and protect mitochondria in brain, liver and other tissues[156,161,333]. The antioxidant effects of estrogens seem unrelated to the classical estrogenic activity[334].

The AE2 antioxidant function is not limited to membranes, since their presence in lipoproteins helps protect them from scavenger radicals[275], maintaining their transport and signaling function. Since acylation on C17 of E2 results in a more effective antioxidant molecular type[55], and no other estrogens seem to specifically carry out this task, its uniqueness, and the importance of the function suggests that the AE2 may constitute, by themselves, a different specialized type of estrogens carrying out a critical and specific function for which they are best suited.

Whilst AE1 do not show significant estrogenicity[171], AE2 are markedly estrogenic [126,150,244], and maintain this estrogenicity longer than the 3-acyl-E2 esters[150], which suggests that they may-precisely-retain this property when packed in lipoproteins, such as low density lipoprotein (LDL)[126] or bound to plasma proteins [335]. When taken together, these properties suggest that the AE2 may, at least, fulfill the role of transport/storage of E2 in addition to an antioxidant function.

### **Central-mediated effects of estrogens on energy homeostasis**

The main arguments for the postulated subdivision of estrogens in four separate classes, based on their structure and function is based on widely different availability of sources, but the marked differences observed suggest -at least- the existence of four groups, described in Table 1, which summarize most of the information provided in the present study.

#### **E1:**

*Structural:* Estrin nucleus, with only one phenolic-OH.

*Functional:* Mild estrogenic effect; a main precursor in the synthesis of E2; main catabolite in the excretion of estrogens as SE1; SE1 being probably the main signaling form of E1; increases growth during development; quantitatively the most abundant molecule with an estrogen nucleus in the body; possible “reserve” for rapid conversion to E2 or AE1.

*Targets:* (Generalized); WAT; reproductive-system organs.

#### **AE1:**

*Structural:* Estrin nucleus, with only the phenolic-OH esterified by a fatty acid.

*Functional:* No estrogenic effects; product of esterification (or interchange) of E1, probable active SERM for ER $\alpha$ , activates lipid catabolism, *via* lipolysis and oxidation of fatty acids; postulated as ponderostat signal, markedly lowers body fat: maintains glycemia.

*Targets:* WAT; brain; liver.

#### **E2:**

*Structural:* Estrin nucleus, with the phenolic-OH, and another-OH in C17.

*Functional:* Main estrogen; marked classical estrogenic effects, protects insulin and facilitates its secretion, maintains glycemia; indirectly activates lipolysis and inhibits lipogenesis; protects and favors the increase and oxidative function of mitochondria, lowers body fat, has antioxidant capability.

*Targets:* Brain, liver, mitochondria, reproductive-system organs, bone.

#### **AE2:**

**Table 1 Comparison of the effects/functions between the main functional types of estrogens<sup>1</sup>**

<b>Effect/ function/ action/ characteristic</b>	<b>E1</b>	<b>AE1</b>	<b>E2</b>	<b>AE2</b>
Bind the ERs at the hormone binding site	↑↑ <sup>2</sup>	X	↑↑ <sup>2</sup>	~
Bind the AF1 or AF2 sites of the ERs	X <sup>2</sup>	↑	X <sup>2</sup>	~
Bind to mitochondria (and some membranes)	~	~	↑ <sup>2</sup>	↑ <sup>2</sup>
Elicit a direct classic estrogenic response	↑ <sup>2</sup>	X	↑↑ <sup>2</sup>	↑↑ <sup>2</sup>
Induces hypoglycemic effects	~i	↑↑ <sup>2</sup>	↑ <sup>2</sup>	~
Is carried by lipoproteins	↑ <sup>2</sup>	↑↑ <sup>2</sup>	X	↑ <sup>2</sup>
Show anti-oxidative effects	~	~	↑ <sup>2</sup>	↑↑ <sup>2</sup>
Activate the 3C→2C conversion (pyruvate dehydrogenase)	~	↑2	↑↑ <sup>2</sup>	~
Increase mitochondrial oxidative activity	~	~	↑	~
Increase whole body thermogenesis	~	↑ <sup>2</sup>	↑ <sup>2</sup>	~
Show lipolytic effects	X	↑	↔	~
Show lipogenic effects	↑	↓ <sup>2</sup>	↓ <sup>2</sup>	~
Decrease WAT fat mass/ limits fat deposition	↓	↑↑ <sup>2</sup>	↑ <sup>2</sup>	~
Allow the activation of the alternative N disposal pathway	~	~	↑	~
Decrease body protein mass	↔	X <sup>2</sup>	X <sup>2</sup>	~

<sup>1</sup>Specific early development and pregnancy-related estrogen molecular species and functions not included.

<sup>2</sup>Shows a coincidence of effect/function for different estrogen types in the same row.

↑: Exerts the effect described; ↓: Exerts an effect opposite to that described; ↔: Variable/not univocal responses; X: Does not exert the effect described; ~: Absent or insufficient information available; E1: Estrone; AE1: Acyl-estrone; E2: 17β-estradiol; AE2: Acyl-E2; ERs: Estrogen receptors; AF1: Activation function site 1 of the estrogen receptors; AF2: Activation function site 2 of the estrogen receptors; WAT: White adipose tissue.

**Structural:** Estrin nucleus, with only the phenolic-OH, esterified in C17 by a fatty acid, often polyunsaturated.

**Functional:** The most effective estrogen form of antioxidant; postulated as an element of transport or reserve of E2, protects lipoproteins, membranes and cell components, marked estrogenic action.

**Targets:** Mitochondria; plasma lipoproteins.

This partial (and incomplete) classification of the main estrogens is based on both structural and functional aspects. The estimated quantitative mass of these four types of estrogen (under standard conditions) in the whole body is: AE1 > E1 > E2 > AE2.

Estrogen is a fundamental modulator of female functions (including estrogenicity *stricto sensu*), which agrees with the high levels of E2 in adult pre-menopausal women. No sufficient data are available to show other gender differences in the postulated groups of estrogens, except for AE1, which pharmacological effects in obese rats are more intense in males than in females.

Estrogens, essentially E2, are responsible for the development of the sex-dependent structures (both physiological and psychological) related with mating and reproduction. Evidently this is achieved with the collaboration of other hormones, e.g. androgens, progestogens and peptidic hormones. It is unclear whether the other three groups of estrogens depicted in Table 1 play a specific significant role in the reproductive processes (with the exception, perhaps, of AE2).

The other key places for action of estrogens are characterized by containing ERs and are found throughout the whole body[336]; but WAT[337], liver[338], muscle[339], and, essentially the brain[340,341] are the main targets for their actions, and are, at least, the best studied. ER $\beta$  globally regulates lipid homeostasis[145], its activation in obesity increases whole body metabolism and mitochondrial biogenesis as a counter-measure to excess WAT lipid storage[312]. Oophorectomy alters WAT lipid metabolism[342], the plasma levels of E2 are affected by diet[284] and determine body fat deposition[343], probably through central (brain) control mechanisms.

In the hypothalamus, where E2 interaction with serotonin has been described[254]. E2 regulates sympathetic nervous control[344]. E2 and AE1 have marked anorectic

effects[180,345]. The postulated ponderostat effect of AE1[181,319] depends on its action on brain, where blood-injected label has been found[181]. The effects of estrogens on glucose and body fat have been attributed to central actions on the brain [15,346]. This is a logical assumption, including also T[347], because the brain controls the body energy metabolism[16] and homeostasis[348], *i.e.* regulates the coordinated biological maintenance systems of the body[349,350].

### **General considerations and conclusions**

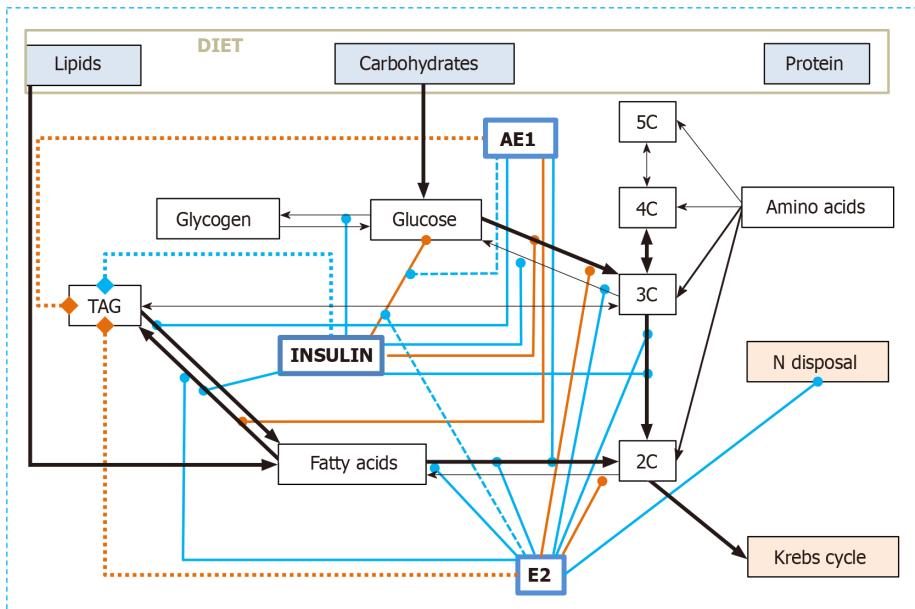
The growing number of known actions of estrogens in metabolic control cannot be fully explained by only the analysis of the “common” estrogens, essentially E1, E2 and E3 (with their sulfates). Thus, acyl-estrogen derivatives have also been included: Despite being known for a long time, and used in pharmacology, they are seldom included in general analyses of estrogens. These compounds are quite diverse. However, in practice, the studies available are limited only to acyl esters of E1 (on C3) or E2 (largely on C17); their properties are quite different, starting with estrogenicity, and continuing on to antioxidant or lipid wasting effects (Table 1).

In any case, the differences in effects induced by E1 and E2 (those of E3 seem to be closer to E2), are considerable, both in their classical estrogenic power and in their implication in regulative mechanisms: E2 being more powerful than E1 in almost any aspect related to metabolic regulation, becoming the most representative estrogen. Nevertheless, a large proportion of E2 is synthesized from E1 by widely distributed 17 OH-steroid dehydrogenases[351]. The similarities between E1 and AE1 actions are small, the latter resembling more E2 in its metabolic effects, than E2 *vs* AE2, despite the strong relationships in the main functions of AE2: antioxidant and estrogenic.

Due to the crossed coincidences of effects between E1-E2 and their acyl esters, a loose classification based on functions and structure has been developed and presented here. Either separately or as a conjoint “estrogens” block they may provide a comprehensive explanation of most of the actions of estrogens, which could not be attributed in any way solely to either one or to all the usual non-esterified estrogens as a group. It is also remarkable that these four groups constitute, together, an extensive fully synergic unit of action: antioxidant effects protect mitochondria, membranes and lipids, which are actively used for energy, limiting insulin resistance. However, the protection extends to insulin (and the pancreatic  $\beta$  cells); insulin secretion is maintained to sustain a steady glycemic response. Glucose is converted to 2C only when it is in excess, whilst dietary lipids (at the root of inflammation and development of MS) are not accrued but oxidized. Protein is preserved by strong, effective and well established mechanisms; but excess energy (and glucose) does not prevent the utilization of amino acids for energy, and, especially for efficient operation of the Krebs cycle thanks to the supply of 4C and 5C fragments. The problem of excess N disposal[189], strictly overprotected *via* the urea cycle is compensated using the direct pathway to produce  $N_2$ [189]. Lipoproteins remain functional thanks to the steroidial antioxidants, TAG transport is practically unaltered, but lipogenesis is maintained low, and lipolysis high to dispose of excess 2C energy. Thermogenesis is maintained and appetite (and food intake) are adjusted to the real needs of energy intake (plus the use of unnecessary fat reserves). A happy metabolic Arcadia under the rule of essentially one (multiple) hormonal factor: estrogens.

Figure 2 shows the main interactions of simple estrogens (essentially E2) and AE1 on the core of intermediate energy metabolism, the meeting point of carbohydrate, lipid and protein catabolism. The antioxidant effects of AE2 (and E2) have been omitted, since their main point of action lies, just from the critical step 3C $\rightarrow$ 2C and the Krebs cycle, within the mitochondrion. The coordinated implication of estrogen types in the control of this segment of substrate handling is pervasive, both by direct implication and through its modulation of insulin action, and helps clarify that the implication of estrogens (as a whole) on glucose handling (and insulin control) is very high. The control node of energy metabolism lies in the mitochondria/cytoplasm interface, and-perhaps-critically on PDH. Around this point, directly linked to oxidative function of mitochondria and their generation of ATP, the implication of estrogens is high, as are the adjustment of the supply lines of 2C and 3C, the control of glycemia and the shift of amino acid catabolism under conditions of abundant energy (and glucose) availability[189].

In addition to the need to consider the estrogens as a group of several molecular species sharing a common biochemical structure and origin, implied web-like in a large number of coordinated metabolic functions, the main conclusion of this review is, precisely, the paramount metabolic importance of the estrogens. This is based on the synergy between the different forms of estrogen to maintain energy (including, obviously, glucose homeostasis), whilst preventing oxidative damage, lipid-induced



**Figure 2 Combined regulatory effects on the utilization of dietary nutrients as energy substrates of insulin, 17 $\beta$ -estradiol and acyl-estrone.** Black arrows: Metabolic relationships; solid green lines: activation/enhancement; solid red lines: deactivation/inhibition. Dashed green lines show activation effects on regulatory processes. Thicker dot lines with final diamond symbols represent the enhancement (green) or decrement (red) of TAG reserves. For the sake of simplicity, other regulatory agents are not shown, and neither are their interactions. AE1: Acyl-estrone; E2: 17 $\beta$ -estradiol and other E2-group estrogens

inflammation, excess fat accrual (and thus, obesity), and easing nitrogen excess normalization. This short (and incomplete) list is quite similar to a prescription for preventing the development of MS. Abundant epidemiological and limited experimental data support this assertion. It seems that we have to look more openly at estrogen forms to better understand their nature and properties, and to use them to fulfill their natural purpose as wardens of energy homeostasis (Box 5).

## BOXES

### Box 1 extended negative opinions on “sex hormones” hamper their investigative study and clinical development

In the case of “sex hormones” (*i.e.*, estrogens and androgens), the intensive pharmacological development has not displaced from use the most representative natural hormones: E2 and T from the front line of pharmacotherapy. However, the abuse of synthetic drugs (anabolic steroids, for instance) for purposes not strictly medical[352, 353] have clouded the relatively recent recovery of T as a critical hormone for energy-partition[354,355].

The widely extended negative opinion against “sex hormones” continues to seriously hinder the use of T in the treatment of aging- and MS-related hypogonadism in mature and old men[225,356]. The “opinion war” against estrogen is, currently, even harder to overcome, because of its direct implication on women’s sex; and because the social, political, and even religious arguments coalesced to raise questions (real or inflated[357]) against their use for any purpose outside a few restricted and socially-conditioned gynecologic disease applications[358-360]. The fact that the natural hormone E2 continues to be the main (and effective and cheap) estrogen standard drug only adds to the widely extended negative bias against estrogens[205, 361].

### Box 2 critical methodological questions - the differences between species

Most of the problems caused by pharmacological overdosing of androgenic anabolic drugs or estrogens are probably secondary to blocking the hypothalamus-hypophysis-gonadal axis, a possibility observed many years ago[362], but seldom taken into consideration in clinical practice (and even less when the use is not medically justified). Because of this problem, continuous administration of excess anabolic androgens, can result in the loss of reproductive function[363] in addition to the derangement of their regulative metabolic (or/and psychological) functions.

Most of the metabolic studies on estrogens have been carried out in rodents for obvious reasons, but there are clear differences in the estrogen (and androgen) functions in rodents from those in humans. This includes a number of aspects, starting with the most obvious: size, metabolic rate and lifespan. The duration of the reproductive cycle, "estrus" or "heat" of rats and mice is shorter than the human ovarian cycle (which incorporates parts of the estrus cycle[364]), but their extension, phasing and physiological structure are different. The estrus is observed in most mammalian species (not in humans and apes), and is marked by changes in body temperature and energy expenditure[365]. Size affects energy expenditure (allometry) [366] and lifespan[367]. The fact that women are usually uniparous and rats normally have a two-digit number of pups (requiring a much higher energy and nutrient supply effort at the expense of the dam) is also a quantitative difference that makes uncertain the direct comparison of hormone changes and their timing, and of substrate dynamics, between different species and reproductive cycle models. Another key difference, explained above, is the E2 (and T) transport in plasma. Humans carry E2 and T bound to SHBG in high proportions of total circulating hormone, but SHBG is absent in mature rodents.

#### **Box 3 steroid hormones as medium-term signals focused to control gene expression**

Steroid hormones have longer circulating-lives than most other hormones (or other signaling molecules), which are rapidly produced (or released), then act and are inactivated, all in a short time-span. The maintenance along time of rapid-response signals is kept thanks to repeated secretion-activation/inactivation-destruction processes, which allow for rapid regulation changes, again in a short period of time. Steroid hormones, however, are produced and secreted to last for much longer periods; their prime mechanism of action is, essentially, gene translation, a process which requires more intensity of signal and longer stimulation periods. The advantage of steroids is their unbeatable stability over time in comparison to short term-effect peptidic hormones, catecholamines, etc. The target tissue specific needs are adjusted through the expression of different receptors and signaling pathways for the same steroid hormone, with additional modulation of expression, or by the numbers or proportions of molecular species, allowing for further modification of their effects under changing conditions.

#### **Box 4 estrogen structural resilience and the environment**

Estrogenic signaling is very ancient, affecting quite a number of phyla as observed by the use of estrogen analogs in the context of co-evolution of plant allopathic defense against herbivores[10,368]. The estrin nucleus is highly resistant to environmental oxidation or bacterial catabolism[369]. The non-biological disposal of human waste induces the accumulation of estrogens in continental waters[370] and its sediments [371]; they are also found in sea sediments[372]. Persistence of estrogen in the natural medium has another negative aspect, the environmental effects of waste estrogen [373], affecting both invertebrates and vertebrates[8]. Human and domestic animal overpopulation extends the increase of estrogenic waste problem to become a serious health, ecologic and economic[8,374] problem that should be understood and adequately addressed.

#### **Box 5 perspectives: the need for further advance in our knowledge of estrogens**

A critical point for the continued use of estrogens in medicine is that of the extended use of molecules designed for specific clinical applications[375]. All these patented molecules do mimic some aspects of physiological estrogen actions, but not all of them [375]. The synthesis of estradiol analogs has been oriented to increase only some effects, sought for specific clinical applications[9,376]. However, most of these compounds were designed before our knowledge of estrogen function, mechanisms of action and metabolic effects were fully known[377]. In fact, right now, our knowledge of the full list of estrogen effects is incomplete. For instance, the role of estrogens in the control of brain organization[378,379], adjustment of the immune response[380,381], or even the estrogen function in core energy metabolism regulation[15].

The proof of our limited knowledge is the lack of a clear picture of all the physiological actions carried out by estrogens. Thus, how can we expect to extend this needed knowledge to drugs devised with specific (not global) objectives? How to test their effects on functions that so far have not even been uncovered or analyzed in the classical estrogens? In fact a similar caveat should be applied to all other steroid hormones, especially corticosteroids and androgens.

We are aware that the binding modulation of the effects of estrogenic drugs is unclear, especially as to which real estrogen (as a whole) actions are carried out by each of these compounds[382,383]. In any case, during the last 70-90 years, the use of synthetic estrogens has been slowly substituted by natural estrogens[383,384], with the practical abandonment of the wonder drug diethylstilbestrol. The case of anabolic androgens (and testosterone itself) is paradigmatic: the continued use of these drugs results at least in infertility[385] and cardiovascular damage[386]. The continued use of estrogens, in particular powerful analogs, may result in unexpected, possibly negative or unexplained effects[384,387,388], simply not detected because the protean nature of estrogenic action has not permeated yet to clinical (and, especially, pharmacological) practice.

We are uncovering the proverbial tip of the iceberg of a group of steroid hormones; we need a deep analysis of estrogen functions, both from the molecular and regulatory aspects, but never forgetting that steroids act on the whole body not only on specific organs and single isolated pathways. The actual role of hormone carriers in plasma (*i.e.* SHBG), the cyclic hypothalamic-hypophysis-gonadal axis function, and the possible disorders induced by estrogenic drug substitutes, must be studied and adapted to human physiology in order to be able to resolve endocrine disorders as a whole. First: do no harm; the effects of estrogens and analog drugs under clinical conditions must be known and fully evaluated because not all estrogens and related drugs act the same way[383,384].

Last, but not least, the real and realistic implication of estrogens (as well as androgens and corticosteroids) in the regulation of the metabolic hub of energy partition should be clarified. This is an essential step to limit the ravages of aging and to understand (and correct) disorders such as the widely extended MS.

## CONCLUSION

The main conclusion of this review is, precisely, the paramount metabolic importance of the estrogens. This is based on the synergy between the different forms of estrogen to maintain energy (including, obviously, glucose homeostasis), whilst preventing oxidative damage, lipid-induced inflammation, excess fat accrual (and thus, obesity), and easing nitrogen excess normalization.

## ACKNOWLEDGEMENTS

I wish to express my gratitude to the members of my former research group (Nitrogen-Obesity, Faculty of Biology, University of Barcelona) for their help, friendship, insight, support and patience.

## REFERENCES

- 1 Dufourc EJ. Sterols and membrane dynamics. *J Chem Biol* 2008; **1**: 63-77 [PMID: 19568799 DOI: 10.1007/s12154-008-0010-6]
- 2 Tang J, Han Z, Chai J. Q&A: what are brassinosteroids and how do they act in plants? *BMC Biol* 2016; **14**: 113 [DOI: 10.1186/s12915-016-0340-8]
- 3 Heftmann E. Functions of steroids in plants. *Phytochemistry* 1974; **14**: 891-901 [DOI: 10.1016/0031-9422(75)85156-9]
- 4 Geuns JMC. Plant steroid hormones - what are they and what do they do? *Trends Biochem Sci* 1982; **7**: 7-9 [DOI: 10.1016/0968-0004(82)90053-6]
- 5 Hewitt S, Hillman JR, Knights BA. Steroidal oestrogens and plant growth and development. *New Phytol* 1980; **85**: 329-350 [DOI: 10.1111/j.1469-8137.1980.tb03172.x]
- 6 Milanesi L, Boland R. Presence of estrogen receptor (ER)-like proteins and endogenous ligands for ER in solanaceae. *Plant Sci* 2004; **166**: 397-404 [DOI: 10.1016/j.plantsci.2003.10.006]
- 7 Janeczko A, Skoczowski A. Mammalian sex hormones in plants. *Folia Histochem Cytophiol* 2005; **43**: 71-79 [PMID: 16044944]
- 8 Adeel M, Song X, Wang Y, Francis D, Yang Y. Environmental impact of estrogens on human, animal and plant life: A critical review. *Environ Int* 2017; **99**: 107-119 [PMID: 28040262 DOI: 10.1016/j.envint.2016.12.010]
- 9 Shorr E, Robinson FH, Papanicolaou GN. A clinical study of the synthetic estrogen stilbestrol. *JAMA* 1939; **113**: 2312-2318 [DOI: 10.1001/jama.1939.72800510003010]
- 10 Basu P, Maier C. Phytoestrogens and breast cancer: In vitro anticancer activities of isoflavones,

- lignans, coumestans, stilbenes and their analogs and derivatives. *Biomed Pharmacother* 2018; **107**: 1648-1666 [PMID: 30257383 DOI: 10.1016/j.bioph.2018.08.100]
- 11 **Niwa YS**, Niwa R. Transcriptional regulation of insect steroid hormone biosynthesis and its role in controlling timing of molting and metamorphosis. *Dev Growth Differ* 2016; **58**: 94-105 [PMID: 26667894 DOI: 10.1111/dgd.12248]
  - 12 **Dinan L**, Harmatha J, Volodin V, Lafont R. Phytoecdysteroids: Diversity, biosynthesis and distribution. In: Smagghe G. Ecdysone: Structures and Functions. Springer Science & Business Media B.V.: Dordrecht, 2009: 3-45
  - 13 **Bereshchenko O**, Bruscoli S, Riccardi C. Glucocorticoids, sex hormones, and immunity. *Front Immunol* 2018; **9**: 1332 [PMID: 29946321 DOI: 10.3389/fimmu.2018.01332]
  - 14 **Bouman A**, Heineman MJ, Faas MM. Sex hormones and the immune response in humans. *Hum Reprod Update* 2005; **11**: 411-423 [PMID: 15817524 DOI: 10.1093/humupd/dmi008]
  - 15 **Mauvais-Jarvis F**, Clegg DJ, Hevener AL. The role of estrogens in control of energy balance and glucose homeostasis. *Endocr Rev* 2013; **34**: 309-338 [PMID: 23460719 DOI: 10.1210/er.2012-1055]
  - 16 **Xu Y**, López M. Central regulation of energy metabolism by estrogens. *Mol Metab* 2018; **15**: 104-115 [PMID: 29886181 DOI: 10.1016/j.molmet.2018.05.012]
  - 17 **Lange IG**, Hartel A, Meyer HH. Evolution of oestrogen functions in vertebrates. *J Steroid Biochem Mol Biol* 2002; **83**: 219-226 [PMID: 12650719 DOI: 10.1016/s0960-0760(02)00225-x]
  - 18 **Simpson E**, Jones M, Misso M, Hewitt K, Hill R, Maffei L, Carani C, Boon WC. Estrogen, a fundamental player in energy homeostasis. *J Steroid Biochem Mol Biol* 2005; **95**: 3-8 [PMID: 16054355 DOI: 10.1016/j.jsbmb.2005.04.018]
  - 19 **Wilson JD**. The evolution of Endocrinology. *Clin Endocrinol* 2005; **62**: 389-396 [DOI: 10.1111/j.1365-2265.2005.02209.x]
  - 20 **O'Malley BW**. 90 years of progesterone: Reminiscing on the origins of the field of progesterone and estrogen receptor action. *J Mol Endocrinol* 2020; **65**: C1-C4 [PMID: 32599564 DOI: 10.1530/JME-20-0042]
  - 21 **Ball P**. Carl Djerassi (1923-2015). *Nature* 2015; **519**: 34 [PMID: 25739624 DOI: 10.1038/519034a]
  - 22 **Nieschlag E**, Nieschlag S. Endocrine History: The history of discovery, synthesis and development of testosterone for clinical use. *Eur J Endocrinol* 2019; **180**: R201-R212 [PMID: 30959485 DOI: 10.1530/EJE-19-0071]
  - 23 **Benedek TG**. History of the development of corticosteroid therapy. *Clin Exp Rheumatol* 2011; **29**: S-5 [PMID: 22018177]
  - 24 **Hillier SG**. Diamonds are forever: the cortisone legacy. *J Endocrinol* 2007; **195**: 1-6 [PMID: 17911391 DOI: 10.1677/JOE-07-0309]
  - 25 **Beato M**, Herrlich P, Schütz G. Steroid hormone receptors: many actors in search of a plot. *Cell* 1995; **83**: 851-857 [PMID: 8521509 DOI: 10.1016/0092-8674(95)90201-5]
  - 26 **Hanukoglu I**. Steroidogenic enzymes: structure, function, and role in regulation of steroid hormone biosynthesis. *J Steroid Biochem Mol Biol* 1992; **43**: 779-804 [PMID: 22217824 DOI: 10.1016/0960-0760(92)90307-5]
  - 27 **Liehr JG**, Avitts TA, Randerath E, Randerath K. Estrogen-induced endogenous DNA adduction: possible mechanism of hormonal cancer. *Proc Natl Acad Sci U S A* 1986; **83**: 5301-5305 [PMID: 3460092 DOI: 10.1073/pnas.83.14.5301]
  - 28 **Maguire PJ**. Estrogen replacement therapy and breast cancer. *J Reprod Med* 1993; **38**: 183-185 [PMID: 8487233]
  - 29 **Hierholzer K**, Lichtenstein I, Siebe H. Does corticosteroid metabolism in target organs affect the cardiovascular system? *J Auton Nerv Syst* 1996; **57**: 188-192 [PMID: 8964948 DOI: 10.1016/0165-1838(95)00097-6]
  - 30 **Hariharan M**, Naga S, VanNoord T, Kindt EK. Assay of human plasma cortisone by liquid chromatography: normal plasma concentrations (between 8 and 10 a.m.) of cortisone and corticosterone. *J Chromatogr* 1993; **613**: 195-201 [PMID: 8491806 DOI: 10.1016/0378-4347(93)80134-p]
  - 31 **Fietta P**, Fietta P, Delsante G. Central nervous system effects of natural and synthetic glucocorticoids. *Psychiatry Clin Neurosci* 2009; **63**: 613-622 [PMID: 19788629 DOI: 10.1111/j.1440-1819.2009.02005.x]
  - 32 **Mayer M**, Kaiser N, Milholland RJ, Rosen F. The binding of dexamethasone and triamcinolone acetonide to glucocorticoid receptors in rat skeletal muscle. *J Biol Chem* 1974; **249**: 5236-5240 [PMID: 4369267]
  - 33 **Dunn JF**, Nisula BC, Rodbard D. Transport of steroid hormones: binding of 21 endogenous steroids to both testosterone-binding globulin and corticosteroid-binding globulin in human plasma. *J Clin Endocrinol Metab* 1981; **53**: 58-68 [PMID: 7195404 DOI: 10.1210/jcem-53-1-58]
  - 34 **Coelingh Bennink HJ**, Holinka CF, Diczfalusi E. Estetrol review: profile and potential clinical applications. *Climacteric* 2008; **11** Suppl 1: 47-58 [PMID: 18464023 DOI: 10.1080/13697130802073425]
  - 35 **Tang M**, Abplanalp W, Subbiah MTR. Association of estrogens with human plasma lipoproteins: Studies using estradiol-17 $\beta$  and its hydrophobic derivative. *J Lab Clin Med* 1997; **129**: 447-452 [DOI: 10.1016/S0022-2143(97)90078-0]
  - 36 **Kow LM**, Pfaff DW. The membrane actions of estrogens can potentiate their lordosis behavior-facilitating genomic actions. *Proc Natl Acad Sci U S A* 2004; **101**: 12354-12357 [PMID: 15302933 DOI: 10.1073/pnas.0404889101]

- 37 **Jacobsohn MK**, Bauder S, Pine SR, Jacobsohn GM. Cholesterol limits estrogen uptake by liposomes and erythrocyte membranes. *Biochim Biophys Acta* 1994; **1195**: 131-140 [PMID: 7918555 DOI: 10.1016/0005-2736(94)90019-1]
- 38 **Arnold S**, Victor MB, Beyer C. Estrogen and the regulation of mitochondrial structure and function in the brain. *J Steroid Biochem Mol Biol* 2012; **131**: 2-9 [PMID: 22326731 DOI: 10.1016/j.jsbmb.2012.01.012]
- 39 **Grasa MM**, Gulfo J, Camps N, Alcalá R, Monserrat L, Moreno-Navarrete JM, Ortega FJ, Esteve M, Remesar X, Fernández-López JA, Fernández-Real JM, Alemany M. Modulation of SHBG binding to testosterone and estradiol by sex and morbid obesity. *Eur J Endocrinol* 2017; **176**: 393-404 [PMID: 28077498 DOI: 10.1530/EJE-16-0834]
- 40 **Parwanto MLE**, Suweino S, Tjahjadi D, Senjaya H, Jaya Edy H, Pakpahan A. The effect of sex hormone-binding globulin (SHBG) protein polymorphism on the levels of SHBG, testosterone, and insulin in healthy Indonesian men. *IJBMPH* 2016; **5**: 799-806 [DOI: 10.5455/ijmsph.2016.17122015293]
- 41 **Vanbillemont G**, Bogaert V, De Bacquer D, Lapauw B, Goemaere S, Toye K, Van Steen K, Taes Y, Kaufman JM. Polymorphisms of the SHBG gene contribute to the interindividual variation of sex steroid hormone blood levels in young, middle-aged and elderly men. *Clin Endocrinol (Oxf)* 2009; **70**: 303-310 [PMID: 18681858 DOI: 10.1111/j.1365-2265.2008.03365.x]
- 42 **Cousin P**, Déchaud H, Grenot C, Lejeune H, Pugeat M. Human variant sex hormone-binding globulin (SHBG) with an additional carbohydrate chain has a reduced clearance rate in rabbit. *J Clin Endocrinol Metab* 1998; **83**: 235-240 [PMID: 9435448 DOI: 10.1210/jcem.83.1.4515]
- 43 **van den Belt K**, Berckmans P, Vangenechten C, Verheyen R, Witters H. Comparative study on the in vitro/in vivo estrogenic potencies of 17beta-estradiol, estrone, 17alpha-ethynodiol and nonylphenol. *Aquat Toxicol* 2004; **66**: 183-195 [PMID: 15036873 DOI: 10.1016/j.aquatox.2003.09.004]
- 44 **Massanés RM**, Grasa MM, López-Martí J, Díaz-Silva M, Fernández-López JA, Remesar X, Alemany M. Zucker obese rats store less acyl-estrone than lean controls. *Int J Obes Relat Metab Disord* 2003; **27**: 428-432 [PMID: 12664075 DOI: 10.1038/sj.ijo.0802264]
- 45 **Siiteri PK**. Adipose tissue as a source of hormones. *Am J Clin Nutr* 1987; **45**: 277-282 [PMID: 3541569 DOI: 10.1093/ajcn/45.1.277]
- 46 **Ruder HJ**, Loriaux L, Lipsett MB. Estrone sulfate: production rate and metabolism in man. *J Clin Invest* 1972; **51**: 1020-1033 [PMID: 5014608 DOI: 10.1172/JCI106862]
- 47 **Brind J**, Strain G, Miller L, Zumoff B, Vogelman J, Orentreich N. Obese men have elevated plasma levels of estrone sulfate. *Int J Obes* 1990; **14**: 483-486 [PMID: 2401584]
- 48 **Sanchis D**, Balada F, Farrerons C, Virgili J, del Mar Grasa M, Adán C, Esteve M, Cabot C, Ardévol A, Vilà R, Fernández-López JA, Remesar X, Alemany M. Structural determinants of oleoyl-estrone slimming effects. *Life Sci* 1998; **62**: 1349-1359 [PMID: 9566777 DOI: 10.1016/s0024-3205(98)00069-1]
- 49 **Virgili J**, Casals I, Peinado-Onsurbe J, Esteve M, Julve-Gil J, Fernández-López JA, Remesar X, Alemany M. Distribution of oleoyl-estrone in rat plasma lipoproteins. *Horm Metab Res* 1999; **31**: 597-601 [PMID: 10598826 DOI: 10.1055/s-2007-978803]
- 50 **Esteve M**, Savall P, Virgili J, Fernández-López JA, Remesar X, Alemany M. Modulation by leptin, insulin and corticosterone of oleoyl-estrone synthesis in cultured 3T3L1 cells. *Biosci Rep* 2001; **21**: 755-763 [PMID: 12166825 DOI: 10.1023/a:1015580623325]
- 51 **Tikkanen MJ**, Vihma V, Jauhainen M, Höckerstedt A, Helisten H, Kaamanen M. Lipoprotein-associated estrogens. *Cardiovasc Res* 2002; **56**: 184-188 [PMID: 12393088 DOI: 10.1016/s0008-6363(02)00535-7]
- 52 **Janocko L**, Hochberg RB. Estradiol fatty acid esters occur naturally in human blood. *Science* 1983; **222**: 1334-1336 [PMID: 6419346 DOI: 10.1126/science.6419346]
- 53 **Larner JM**, MacLusky NJ, Hochberg RB. The naturally occurring C-17 fatty acid esters of estradiol are long-acting estrogens. *J Steroid Biochem* 1985; **22**: 407-413 [PMID: 3990290 DOI: 10.1016/0022-4731(85)90446-7]
- 54 **Badeau M**, Vihma V, Mikkola TS, Tuutinen A, Tikkanen MJ. Estradiol fatty acid esters in adipose tissue and serum of pregnant and pre- and postmenopausal women. *J Clin Endocrinol Metab* 2007; **92**: 4327-4331 [PMID: 17726068 DOI: 10.1210/jc.2007-1372]
- 55 **Vihma V**, Tikkanen MJ. Fatty acid esters of steroids: synthesis and metabolism in lipoproteins and adipose tissue. *J Steroid Biochem Mol Biol* 2011; **124**: 65-76 [PMID: 21277977 DOI: 10.1016/j.jsbmb.2011.01.011]
- 56 **Clemons M**, Goss P. Estrogen and the risk of breast cancer. *N Engl J Med* 2001; **344**: 276-285 [PMID: 11172156 DOI: 10.1056/NEJM200101253440407]
- 57 **Antunes CM**, Strolley PD, Rosenshein NB, Davies JL, Tonascia JA, Brown C, Burnett L, Rutledge A, Pokempner M, Garcia R. Endometrial cancer and estrogen use. Report of a large case-control study. *N Engl J Med* 1979; **300**: 9-13 [PMID: 213722 DOI: 10.1056/NEJM197901043000103]
- 58 **Henderson BE**, Ross RK, Paganini-Hill A, Mack TM. Estrogen use and cardiovascular disease. *Am J Obstet Gynecol* 1986; **154**: 1181-1186 [PMID: 3717228 DOI: 10.1016/0002-9378(86)90696-4]
- 59 **Yaffe K**, Haan M, Byers A, Tangen C, Kuller L. Estrogen use, APOE, and cognitive decline: evidence of gene-environment interaction. *Neurology* 2000; **54**: 1949-1954 [PMID: 10822435 DOI: 10.1212/wnl.54.10.1949]
- 60 **Vandenput L**, Ohlsson C. Estrogens as regulators of bone health in men. *Nat Rev Endocrinol* 2009;

- 5: 437-443 [PMID: 19528961 DOI: 10.1038/nrendo.2009.112]
- 61 **Samuel VT.** The emerging role of oestrogen-related receptor  $\gamma$  as a regulator of energy metabolism. *Diabetologia* 2014; **57**: 2440-2443 [PMID: 25257097 DOI: 10.1007/s00125-014-3377-7]
- 62 **Frederiksen H,** Johannsen TH, Andersen SE, Albrethsen J, Landersoe SK, Petersen JH, Andersen AN, Vestergaard ET, Schorring ME, Linneberg A, Main KM, Andersson AM, Juul A. Sex-specific Estrogen Levels and Reference Intervals from Infancy to Late Adulthood Determined by LC-MS/MS. *J Clin Endocrinol Metab* 2020; **105** [PMID: 31720688 DOI: 10.1210/clinem/dgz196]
- 63 **Yamamoto M,** Hibi H, Katsuno S, Miyake K. Serum estradiol levels in normal men and men with idiopathic infertility. *Int J Urol* 1995; **2**: 44-46 [PMID: 7614406]
- 64 **Yamaji T,** Ibayashi H. Plasma dehydroepiandrosterone sulfate in normal and pathological conditions. *J Clin Endocrinol Metab* 1969; **29**: 273-278 [PMID: 4303580 DOI: 10.1210/jcem-29-2-273]
- 65 **Fernández-Real JM,** Sanchis D, Ricart W, Casamitjana R, Balada F, Remesar X, Alemany M. Plasma oestrone-fatty acid ester levels are correlated with body fat mass in humans. *Clin Endocrinol (Oxf)* 1999; **50**: 253-260 [PMID: 10396370 DOI: 10.1046/j.1365-2265.1999.00669.x]
- 66 **Marrocco J,** McEwen BS. Sex in the brain: hormones and sex differences. *Dialogues Clin Neurosci* 2016; **18**: 373-383 [PMID: 28179809]
- 67 **van Goozen SH,** Cohen-Kettenis PT, Gooren LJ, Frijda NH, Van de Poll NE. Gender differences in behaviour: activating effects of cross-sex hormones. *Psychoneuroendocrinology* 1995; **20**: 343-363 [PMID: 8532819 DOI: 10.1016/0306-4530(94)00076-x]
- 68 **Høeg LD,** Sjøberg KA, Jeppesen J, Jensen TE, Frøsig C, Birk JB, Bisiani B, Hiscock N, Pilegaard H, Wojtaszewski JF, Richter EA, Kiens B. Lipid-induced insulin resistance affects women less than men and is not accompanied by inflammation or impaired proximal insulin signaling. *Diabetes* 2011; **60**: 64-73 [PMID: 20956497 DOI: 10.2337/db10-0698]
- 69 **Lejsková M,** Alušík S, Suchánek M, Zecová S, Pitha J. Menopause: clustering of metabolic syndrome components and population changes in insulin resistance. *Climacteric* 2011; **14**: 83-91 [PMID: 20443721 DOI: 10.3109/13697131003692745]
- 70 **Cauley JA.** Estrogen and bone health in men and women. *Steroids* 2015; **99**: 11-15 [PMID: 25555470 DOI: 10.1016/j.steroids.2014.12.010]
- 71 **Mohamad NV,** Ima-Nirwana S, Chin KY. Are oxidative stress and inflammation mediators of bone loss due to estrogen deficiency? *Endocr Metab Immune Disord Drug Targets* 2020; **20**: 1478-1487 [PMID: 32496996 DOI: 10.2174/1871530320666200604160614]
- 72 **Falahati-Nini A,** Riggs BL, Atkinson EJ, O'Fallon WM, Eastell R, Khosla S. Relative contributions of testosterone and estrogen in regulating bone resorption and formation in normal elderly men. *J Clin Invest* 2000; **106**: 1553-1560 [PMID: 11120762 DOI: 10.1172/JCI10942]
- 73 **Griggs RC,** Kingston W, Jozefowicz RF, Herr BE, Forbes G, Halliday D. Effect of testosterone on muscle mass and muscle protein synthesis. *J Appl Physiol (1985)* 1989; **66**: 498-503 [PMID: 2917954 DOI: 10.1152/jappl.1989.66.1.498]
- 74 **Urban RJ,** Bodenburg YH, Gilkison C, Foxworth J, Coggan AR, Wolfe RR, Ferrando A. Testosterone administration to elderly men increases skeletal muscle strength and protein synthesis. *Am J Physiol* 1995; **269**: E820-E826 [PMID: 7491931 DOI: 10.1152/ajpendo.1995.269.5.E820]
- 75 **Birzniece V,** Meinhardt UJ, Umpleby MA, Handelsman DJ, Ho KK. Interaction between testosterone and growth hormone on whole-body protein anabolism occurs in the liver. *J Clin Endocrinol Metab* 2011; **96**: 1060-1067 [PMID: 21239519 DOI: 10.1210/jc.2010-2521]
- 76 **Chow LS,** Albright RC, Bigelow ML, Toffolo G, Cobelli C, Nair KS. Mechanism of insulin's anabolic effect on muscle: measurements of muscle protein synthesis and breakdown using aminoacyl-tRNA and other surrogate measures. *Am J Physiol Endocrinol Metab* 2006; **291**: E729-E736 [PMID: 16705065 DOI: 10.1152/ajpendo.00003.2006]
- 77 **Ohtsuka A,** Hayashi K, Noda T, Tomita Y. Reduction of corticosterone-induced muscle proteolysis and growth retardation by a combined treatment with insulin, testosterone and high-protein-high-fat diet in rats. *J Nutr Sci Vitaminol (Tokyo)* 1992; **38**: 83-92 [PMID: 1629788 DOI: 10.3177/jnsv.38.83]
- 78 **Mauras N.** Estrogens do not affect whole-body protein metabolism in the prepubertal female. *J Clin Endocrinol Metab* 1995; **80**: 2842-2845 [PMID: 7559861 DOI: 10.1210/jcem.80.10.7559861]
- 79 **Kofoed EM,** Guerbadot M, Schaufele F. Structure, affinity, and availability of estrogen receptor complexes in the cellular environment. *J Biol Chem* 2010; **285**: 2428-2437 [PMID: 19926790 DOI: 10.1074/jbc.M109.045203]
- 80 **Horard B,** Castet A, Bardet PL, Laudet V, Cavailles V, Vanacker JM. Dimerization is required for transactivation by estrogen-receptor-related (ERR) orphan receptors: evidence from amphioxus ERR. *J Mol Endocrinol* 2004; **33**: 493-509 [PMID: 15525604 DOI: 10.1677/jme.1.01538]
- 81 **Schwabe JW,** Chapman L, Finch JT, Rhodes D. The crystal structure of the estrogen receptor DNA-binding domain bound to DNA: how receptors discriminate between their response elements. *Cell* 1993; **75**: 567-578 [PMID: 8221895 DOI: 10.1016/0092-8674(93)90390-c]
- 82 **Cheskin BJ,** Karathanasis S, Lytle CR. Estrogen receptor ligands modulate its interaction with DNA. *J Biol Chem* 1997; **272**: 11384-11391 [PMID: 9111047 DOI: 10.1074/jbc.272.17.11384]
- 83 **Heldring N,** Isaacs GD, Diehl AG, Sun M, Cheung E, Ranish JA, Kraus WL. Multiple sequence-specific DNA-binding proteins mediate estrogen receptor signaling through a tethering pathway. *Mol Endocrinol* 2011; **25**: 564-574 [PMID: 21330404 DOI: 10.1210/me.2010-0425]
- 84 **Fuentes N,** Silveyra P. Estrogen receptor signaling mechanisms. *Adv Protein Chem Struct Biol*

- 2019; **116**: 135-170 [PMID: 31036290 DOI: 10.1016/bs.apcsb.2019.01.001]
- 85 **Mueller-Fahrnow A**, Egner U. Ligand-binding domain of estrogen receptors. *Curr Opin Biotechnol* 1999; **10**: 550-556 [PMID: 10600690 DOI: 10.1016/s0958-1669(99)00034-8]
- 86 **Blair RM**, Fang H, Branham WS, Hass BS, Dial SL, Moland CL, Tong W, Shi L, Perkins R, Sheehan DM. The estrogen receptor relative binding affinities of 188 natural and xenochemicals: structural diversity of ligands. *Toxicol Sci* 2000; **54**: 138-153 [PMID: 10746941 DOI: 10.1093/toxsci/54.1.138]
- 87 **Decherling K**, Boersma C, Mosselman S. Estrogen receptors  $\alpha$  and  $\beta$ : Two receptors of a kind? *Curr Med Chem* 2000; **7**: 561-576 [DOI: 10.2174/0929867003375010]
- 88 **Damdimopoulos AE**, Spyrou G, Gustafsson JA. Ligands differentially modify the nuclear mobility of estrogen receptors alpha and beta. *Endocrinology* 2008; **149**: 339-345 [PMID: 17884941 DOI: 10.1210/en.2007-0198]
- 89 **Truss M**, Beato M. Steroid hormone receptors: interaction with deoxyribonucleic acid and transcription factors. *Endocr Rev* 1993; **14**: 459-479 [PMID: 8223341 DOI: 10.1210/edrv-14-4-459]
- 90 **Faroq A**. Structural and Functional diversity of estrogen receptor ligands. *Curr Top Med Chem* 2015; **15**: 1372-1384 [PMID: 25866274 DOI: 10.2174/1568026615666150413154841]
- 91 **Tora L**, White J, Brou C, Tasset D, Webster N, Scheer E, Chambon P. The human estrogen receptor has two independent nonacidic transcriptional activation functions. *Cell* 1989; **59**: 477-487 [PMID: 2805068 DOI: 10.1016/0092-8674(89)90031-7]
- 92 **Holm A**, Nilsson BO. Identification and characterization of new mechanisms in vascular oestrogen signalling. *Basic Clin Pharmacol Toxicol* 2013; **113**: 287-293 [PMID: 23953673 DOI: 10.1111/bcpt.12118]
- 93 **Srivastava DP**, Waters EM, Mermelstein PG, Kramář EA, Shors TJ, Liu F. Rapid estrogen signaling in the brain: implications for the fine-tuning of neuronal circuitry. *J Neurosci* 2011; **31**: 16056-16063 [PMID: 22072656 DOI: 10.1523/JNEUROSCI.4097-11.2011]
- 94 **Prossnitz ER**, Maggiolini M. Mechanisms of estrogen signaling and gene expression via GPR30. *Mol Cell Endocrinol* 2009; **308**: 32-38 [PMID: 19464786 DOI: 10.1016/j.mce.2009.03.026]
- 95 **Cheskins BJ**, Greger JG, Nagpal S, Freedman LP. Signaling by estrogens. *J Cell Physiol* 2007; **213**: 610-617 [PMID: 17886255 DOI: 10.1002/jcp.21253]
- 96 **Enmark E**, Pelto-Huikko M, Grandien K, Lagercrantz S, Lagercrantz J, Fried G, Nordenskjöld M, Gustafsson JA. Human estrogen receptor beta-gene structure, chromosomal localization, and expression pattern. *J Clin Endocrinol Metab* 1997; **82**: 4258-4265 [PMID: 9398750 DOI: 10.1210/jcem.82.12.4470]
- 97 **Reid G**, Denger S, Kos M, Gannon F. Human estrogen receptor-alpha: regulation by synthesis, modification and degradation. *Cell Mol Life Sci* 2002; **59**: 821-831 [PMID: 12088282 DOI: 10.1007/s00018-002-8470-2]
- 98 **Sowers MR**, Jannausch ML, McConnell DS, Kardia SR, Randolph JF Jr. Endogenous estradiol and its association with estrogen receptor gene polymorphisms. *Am J Med* 2006; **119**: S16-S22 [PMID: 16949384 DOI: 10.1016/j.amjmed.2006.07.002]
- 99 **Kjaergaard AD**, Ellervik C, Tybjaerg-Hansen A, Axelsson CK, Grønholdt ML, Grande P, Jensen GB, Nordestgaard BG. Estrogen receptor alpha polymorphism and risk of cardiovascular disease, cancer, and hip fracture: cross-sectional, cohort, and case-control studies and a meta-analysis. *Circulation* 2007; **115**: 861-871 [PMID: 17309937 DOI: 10.1161/CIRCULATIONAHA.106.615567]
- 100 **Porter JW**, Barnas JL, Welly R, Spencer N, Pitt J, Vieira-Potter VJ, Kanaley JA. Age, sex, and depot-specific differences in adipose-tissue estrogen receptors in individuals with obesity. *Obesity (Silver Spring)* 2020; **28**: 1698-1707 [PMID: 32734695 DOI: 10.1002/oby.22888]
- 101 **Barros RP**, Machado UF, Gustafsson JA. Estrogen receptors: new players in diabetes mellitus. *Trends Mol Med* 2006; **12**: 425-431 [PMID: 16890492 DOI: 10.1016/j.molmed.2006.07.004]
- 102 **Matthews J**, Gustafsson JA. Estrogen signaling: a subtle balance between ER alpha and ER beta. *Mol Interv* 2003; **3**: 281-292 [PMID: 14993442 DOI: 10.1124/mi.3.5.281]
- 103 **Moggs JG**, Orphanides G. Estrogen receptors: orchestrators of pleiotropic cellular responses. *EMBO Rep* 2001; **2**: 775-781 [PMID: 11559590 DOI: 10.1093/embo-reports/kve185]
- 104 **Shiau AK**, Barstad D, Radek JT, Meyers MJ, Nettles KW, Katzenellenbogen BS, Katzenellenbogen JA, Agard DA, Greene GL. Structural characterization of a subtype-selective ligand reveals a novel mode of estrogen receptor antagonism. *Nat Struct Biol* 2002; **9**: 359-364 [PMID: 11953755 DOI: 10.1038/nsb787]
- 105 **Xu B**, Lovre D, Mauvais-Jarvis F. Effect of selective estrogen receptor modulators on metabolic homeostasis. *Biochimie* 2016; **124**: 92-97 [PMID: 26133657 DOI: 10.1016/j.biochi.2015.06.018]
- 106 **Craig Jordan V**, McDaniel R, Agboke F, Maximov PY. The evolution of nonsteroidal antiestrogens to become selective estrogen receptor modulators. *Steroids* 2014; **90**: 3-12 [PMID: 24949934 DOI: 10.1016/j.steroids.2014.06.009]
- 107 **Blizzard TA**, Gude C, Morgan JD 2nd, Chan W, Birzin ET, Mojena M, Tudela C, Chen F, Knecht K, Su Q, Kraker B, Mosley RT, Holmes MA, Sharma N, Fitzgerald PM, Rohrer SP, Hammond ML. Androstenediol analogs as ER-beta-selective SERMs. *Bioorg Med Chem Lett* 2006; **16**: 834-838 [PMID: 16309907 DOI: 10.1016/j.bmcl.2005.11.014]
- 108 **McDonnell DP**, Wardell SE, Norris JD. Oral selective estrogen receptor downregulators (SERDs), a breakthrough endocrine therapy for breast cancer. *J Med Chem* 2015; **58**: 4883-4887 [PMID: 26039356 DOI: 10.1021/acs.jmedchem.5b00760]

- 109 **Lisse TS**, Hewison M, Adams JS. Hormone response element binding proteins: novel regulators of vitamin D and estrogen signaling. *Steroids* 2011; **76**: 331-339 [PMID: 21236284 DOI: 10.1016/j.steroids.2011.01.002]
- 110 **Bu H**, Kashireddy P, Chang J, Zhu YT, Zhang Z, Zheng W, Rao SM, Zhu YJ. ERBP, a novel estrogen receptor binding protein enhancing the activity of estrogen receptor. *Biochem Biophys Res Commun* 2004; **317**: 54-59 [PMID: 15047147 DOI: 10.1016/j.bbrc.2004.02.179]
- 111 **Wu Q**, Chambliss K, Umetani M, Mineo C, Shaul PW. Non-nuclear estrogen receptor signaling in the endothelium. *J Biol Chem* 2011; **286**: 14737-14743 [PMID: 21343284 DOI: 10.1074/jbc.R110.191791]
- 112 **Marino M**, Ascenzi P. Membrane association of estrogen receptor alpha and beta influences 17beta-estradiol-mediated cancer cell proliferation. *Steroids* 2008; **73**: 853-858 [PMID: 18206197 DOI: 10.1016/j.steroids.2007.12.003]
- 113 **Jacob J**, Sebastian KS, Devassy S, Priyadarsini L, Farook MF, Shameem A, Mathew D, Sreeja S, Thampam RV. Membrane estrogen receptors: genomic actions and post transcriptional regulation. *Mol Cell Endocrinol* 2006; **246**: 34-41 [PMID: 16423448 DOI: 10.1016/j.mce.2005.11.015]
- 114 **Pappas TC**, Gametchu B, Watson CS. Membrane estrogen receptors identified by multiple antibody labeling and impeded-ligand binding. *FASEB J* 1995; **9**: 404-410 [PMID: 7896011 DOI: 10.1096/fasebj.9.5.7896011]
- 115 **Chaban VV**, Lakhter AJ, Micevych P. A membrane estrogen receptor mediates intracellular calcium release in astrocytes. *Endocrinology* 2004; **145**: 3788-3795 [PMID: 15131017 DOI: 10.1210/en.2004-0149]
- 116 **Sak K**, Everaus H. Nongenomic effects of 17beta-estradiol--diversity of membrane binding sites. *J Steroid Biochem Mol Biol* 2004; **88**: 323-335 [PMID: 15145442 DOI: 10.1016/j.jsbmb.2004.01.004]
- 117 **Khbouz B**, de Bouronville C, Court L, Taziaux M, Corona R, Arnal JF, Lenfant F, Cornil CA. Role for the membrane estrogen receptor alpha in the sexual differentiation of the brain. *Eur J Neurosci* 2020; **52**: 2627-2645 [PMID: 31833601 DOI: 10.1111/ejn.14646]
- 118 **Pedram A**, Razandi M, Kim JK, O'Mahony F, Lee EY, Luderer U, Levin ER. Developmental phenotype of a membrane only estrogen receptor alpha (MOER) mouse. *J Biol Chem* 2009; **284**: 3488-3495 [PMID: 19054762 DOI: 10.1074/jbc.M806249200]
- 119 **Pastore MB**, Landeros RV, Chen DB, Magness RR. Structural analysis of estrogen receptors: interaction between estrogen receptors and cav-1 within the caveolae. *Biol Reprod* 2019; **100**: 495-504 [PMID: 30137221 DOI: 10.1093/biolre/iyj188]
- 120 **Márquez DC**, Chen HW, Curran EM, Welshons WV, Pietras RJ. Estrogen receptors in membrane lipid rafts and signal transduction in breast cancer. *Mol Cell Endocrinol* 2006; **246**: 91-100 [PMID: 16388889 DOI: 10.1016/j.mce.2005.11.020]
- 121 **Ribeiro M**, Sousa C, Rufino AT, Judas F, Mendes AF. Expression and function of the nonclassical estrogen receptor, GPR30, in human cartilage and chondrocytes. *J Cell Physiol* 2020; **235**: 8486-8494 [PMID: 32324271 DOI: 10.1002/jcp.29691]
- 122 **Sharma G**, Mauvais-Jarvis F, Prossnitz ER. Roles of G protein-coupled estrogen receptor GPER in metabolic regulation. *J Steroid Biochem Mol Biol* 2018; **176**: 31-37 [PMID: 28223150 DOI: 10.1016/j.jsbmb.2017.02.012]
- 123 **Ervin KS**, Lymer JM, Matta R, Clipperton-Allen AE, Kavaliers M, Choleris E. Estrogen involvement in social behavior in rodents: Rapid and long-term actions. *Horm Behav* 2015; **74**: 53-76 [PMID: 26122289 DOI: 10.1016/j.yhbeh.2015.05.023]
- 124 **Sheppard PAS**, Koss WA, Frick KM, Choleris E. Rapid actions of oestrogens and their receptors on memory acquisition and consolidation in females. *J Neuroendocrinol* 2018; **30** [PMID: 28489296 DOI: 10.1111/jne.12485]
- 125 **Prossnitz ER**, Arterburn JB, Sklar LA. GPR30: A G protein-coupled receptor for estrogen. *Mol Cell Endocrinol* 2007; **265**: 138-142 [DOI: 10.1016/j.mce.2006.12.010]
- 126 **Helisten H**, Höckerstedt A, Wähälä K, Tiitinen A, Adlercreutz H, Jauhainen M, Tikkannen MJ. Accumulation of high-density lipoprotein-derived estradiol-17beta fatty acid esters in low-density lipoprotein particles. *J Clin Endocrinol Metab* 2001; **86**: 1294-1300 [PMID: 11238523 DOI: 10.1210/jcem.86.3.7292]
- 127 **Ghaffari S**, Naderi Nabi F, Sugiyama MG, Lee WL. Estrogen inhibits LDL (low-density lipoprotein) transcytosis by human coronary artery endothelial cells via GPER (G-protein-coupled estrogen receptor) and SR-BI (scavenger receptor class B type 1). *Arterioscler Thromb Vasc Biol* 2018; **38**: 2283-2294 [PMID: 30354216 DOI: 10.1161/ATVBAHA.118.310792]
- 128 **Mizukami Y**. In vivo functions of GPR30/GPER-1, a membrane receptor for estrogen: from discovery to functions in vivo. *Endocr J* 2010; **57**: 101-107 [PMID: 19996532 DOI: 10.1507/endocrj.k09e-332]
- 129 **Prossnitz ER**, Barton M. The G-protein-coupled estrogen receptor GPER in health and disease. *Nat Rev Endocrinol* 2011; **7**: 715-726 [PMID: 21844907 DOI: 10.1038/nrendo.2011.122]
- 130 **Feldman RD**, Limbird LE. GPER (GPR30): A Nongenomic receptor (GPCR) for steroid hormones with implications for cardiovascular disease and cancer. *Annu Rev Pharmacol Toxicol* 2017; **57**: 567-584 [PMID: 27814026 DOI: 10.1146/annurev-pharmtox-010716-104651]
- 131 **Lindsey SH**, Chappell MC. Evidence that the G protein-coupled membrane receptor GPR30 contributes to the cardiovascular actions of estrogen. *Gend Med* 2011; **8**: 343-354 [PMID: 22153880 DOI: 10.1016/j.genm.2011.10.004]
- 132 **Roque C**, Baltazar G. G protein-coupled estrogen receptor 1 (GPER) activation triggers different

- signaling pathways on neurons and astrocytes. *Neural Regen Res* 2019; **14**: 2069-2070 [PMID: 31397335 DOI: 10.4103/1673-5374.262577]
- 133 **Cheong RY**, Kwakowsky A, Barad Z, Porteous R, Herbison AE, Ábrahám IM. Estradiol acts directly and indirectly on multiple signaling pathways to phosphorylate cAMP-response element binding protein in GnRH neurons. *Endocrinology* 2012; **153**: 3792-3803 [PMID: 22719057 DOI: 10.1210/en.2012-1232]
- 134 **Kow LM**, Pfaff DW. Rapid estrogen actions on ion channels: A survey in search for mechanisms. *Steroids* 2016; **111**: 46-53 [PMID: 26939826 DOI: 10.1016/j.steroids.2016.02.018]
- 135 **Lu Q**, Schnitzler GR, Ueda K, Iyer LK, Diomede OI, Andrade T, Karas RH. ER alpha rapid signaling is required for estrogen induced proliferation and migration of vascular endothelial cells. *PLoS One* 2016; **11**: e0152807 [PMID: 27035664 DOI: 10.1371/journal.pone.0152807]
- 136 **Paletta P**, Sheppard PAS, Matta R, Ervin KSJ, Choleris E. Rapid effects of estrogens on short-term memory: Possible mechanisms. *Horm Behav* 2018; **104**: 88-99 [PMID: 29847771 DOI: 10.1016/j.ybeh.2018.05.019]
- 137 **Yang SH**, Liu R, Perez EJ, Wen Y, Stevens SM Jr, Valencia T, Brun-Zinkernagel AM, Prokai L, Will Y, Dykens J, Koulen P, Simpkins JW. Mitochondrial localization of estrogen receptor beta. *Proc Natl Acad Sci U S A* 2004; **101**: 4130-4135 [PMID: 15024130 DOI: 10.1073/pnas.0306948101]
- 138 **Chu HP**, Sarkar G, Etgen AM. Estradiol and progesterone modulate the nitric oxide/cyclic gmp pathway in the hypothalamus of female rats and in GT1-1 cells. *Endocrine* 2004; **24**: 177-184 [PMID: 15347845 DOI: 10.1385/ENDO:24:2:177]
- 139 **Otsuka M**, Kadokawa H. GPR30 mediates estrone, estriol, and estradiol to suppress gonadotropin-releasing hormone-induced luteinizing hormone secretion in the anterior pituitary of heifers. *J Reprod Dev* 2017; **63**: 519-525 [PMID: 28781349 DOI: 10.1262/jrd.2017-035]
- 140 **Li Y**, Wang JP, Santen RJ, Kim TH, Park H, Fan P, Yue W. Estrogen stimulation of cell migration involves multiple signaling pathway interactions. *Endocrinology* 2010; **151**: 5146-5156 [PMID: 20861240 DOI: 10.1210/en.2009-1506]
- 141 **Kelly MJ**, Qiu J, Wagner EJ, Rønnekleiv OK. Rapid effects of estrogen on G protein-coupled receptor activation of potassium channels in the central nervous system (CNS). *J Steroid Biochem Mol Biol* 2002; **83**: 187-193 [PMID: 12650715 DOI: 10.1016/s0960-0760(02)00249-2]
- 142 **Iorga A**, Umar S, Ruffenach G, Aryan L, Li J, Sharma S, Motayagheni N, Nadadur RD, Bopassa JC, Eghbali M. Estrogen rescues heart failure through estrogen receptor Beta activation. *Biol Sex Differ* 2018; **9**: 48 [PMID: 30376877 DOI: 10.1186/s13293-018-0206-6]
- 143 **Lahm T**, Crisostomo PR, Markel TA, Wang M, Wang Y, Tan J, Meldrum DR. Selective estrogen receptor-alpha and estrogen receptor-beta agonists rapidly decrease pulmonary artery vasoconstriction by a nitric oxide-dependent mechanism. *Am J Physiol Regul Integr Comp Physiol* 2008; **295**: R1486-R1493 [PMID: 18832085 DOI: 10.1152/ajpregu.90667.2008]
- 144 **van Veen JE**, Kammel LG, Bunda PC, Shum M, Reid MS, Massa MG, Arneson D, Park JW, Zhang Z, Joseph AM, Hrncir H, Liesa M, Arnold AP, Yang X, Correa SM. Hypothalamic estrogen receptor alpha establishes a sexually dimorphic regulatory node of energy expenditure. *Nat Metab* 2020; **2**: 351-363 [PMID: 32377634 DOI: 10.1038/s42255-020-0189-6]
- 145 **Savva C**, Korach-Andre M. Estrogen receptor  $\beta$  (ER $\beta$ ) regulation of lipid homeostasis. Does sex matter? *Metabolites* 2020; **10**: 116 [DOI: 10.3390/metabo10030116]
- 146 **Raut S**, Kumar AV, Khambata K, Deshpande S, Balasinor NH. Genome-wide identification of estrogen receptor binding sites reveals novel estrogen-responsive pathways in adult male germ cells. *Biochem J* 2020; **477**: 2115-2131 [PMID: 32478811 DOI: 10.1042/BCJ20190946]
- 147 **Piperigkou Z**, Karamanos NK. Estrogen receptor-mediated targeting of the extracellular matrix network in cancer. *Semin Cancer Biol* 2020; **62**: 116-124 [PMID: 31310807 DOI: 10.1016/j.semcancer.2019.07.006]
- 148 **Hatcher KM**, Royston SE, Mahoney MM. Modulation of circadian rhythms through estrogen receptor signaling. *Eur J Neurosci* 2020; **51**: 217-228 [PMID: 30270552 DOI: 10.1111/ejn.14184]
- 149 **Janocko L**, Larner JM, Hochberg RB. The interaction of C-17 esters of estradiol with the estrogen receptor. *Endocrinology* 1984; **114**: 1180-1186 [PMID: 6705734 DOI: 10.1210/endo-114-4-1180]
- 150 **Vazquez-Alcantara MA**, Menjivar M, Garcia GA, Diaz-Zagoya JC, Garza-Flores J. Long-acting estrogenic responses of estradiol fatty acid esters. *J Steroid Biochem* 1989; **33**: 1111-1118 [PMID: 2515394 DOI: 10.1016/0022-4731(89)90417-2]
- 151 **Wang F**, Vihma V, Soronen J, Turpeinen U, Hämäläinen E, Savolainen-Pelttonen H, Mikkola TS, Naukkarinen J, Pietiläinen KH, Jauhainen M, Yki-Järvinen H, Tikkanen MJ. 17 $\beta$ -Estradiol and estradiol fatty acyl esters and estrogen-converting enzyme expression in adipose tissue in obese men and women. *J Clin Endocrinol Metab* 2013; **98**: 4923-4931 [PMID: 24081738 DOI: 10.1210/jc.2013-2605]
- 152 **Chen JQ**, Yager JD, Russo J. Regulation of mitochondrial respiratory chain structure and function by estrogens/estrogen receptors and potential physiological/pathophysiological implications. *Biochim Biophys Acta* 2005; **1746**: 1-17 [PMID: 16169101 DOI: 10.1016/j.bbamcr.2005.08.001]
- 153 **Ventura-Clapier R**, Piquereau J, Veksler V, Garnier A. Estrogens, estrogen receptors effects on cardiac and skeletal muscle mitochondria. *Front Endocrinol (Lausanne)* 2019; **10**: 557 [PMID: 31474941 DOI: 10.3389/fendo.2019.00557]
- 154 **Álvarez-Delgado C**, Mendoza-Rodríguez CA, Picazo O, Cerbón M. Different expression of  $\alpha$  and  $\beta$  mitochondrial estrogen receptors in the aging rat brain: Interaction with respiratory complex V. *Exp*

- Gerontol* 2010; **45**: 580-585 [DOI: [10.1016/j.exger.2010.01.015](https://doi.org/10.1016/j.exger.2010.01.015)]
- 155 **Rangwala SM**, Wang XM, Calvo JA, Lindsley L, Zhang YY, Deyneko G, Beaulieu V, Gao JP, Turner G, Markovits J. Estrogen-related receptor  $\gamma$  is a key regulator of muscle mitochondrial activity and oxidative capacity. *J Biol Chem* 2010; **285**: 22619-22629 [DOI: [10.1074/jbc.M110.125401](https://doi.org/10.1074/jbc.M110.125401)]
- 156 **Simpkins JW**, Yi KD, Yang SH, Dykens JA. Mitochondrial mechanisms of estrogen neuroprotection. *Biochim Biophys Acta* 2010; **1800**: 1113-1120 [PMID: [19931595](https://pubmed.ncbi.nlm.nih.gov/20091595/) DOI: [10.1016/j.bbagen.2009.11.013](https://doi.org/10.1016/j.bbagen.2009.11.013)]
- 157 **Ponnusamy S**, Tran QT, Harvey I, Smallwood HS, Thiagarajan T, Banerjee S, Johnson DL, Dalton JT, Sullivan RD, Miller DD, Bridges D, Narayanan R. Pharmacologic activation of estrogen receptor  $\beta$  increases mitochondrial function, energy expenditure, and brown adipose tissue. *FASEB J* 2017; **31**: 266-281 [PMID: [27733447](https://pubmed.ncbi.nlm.nih.gov/27733447/) DOI: [10.1096/fj.201600787RR](https://doi.org/10.1096/fj.201600787RR)]
- 158 **Zhou ZQ**, Ribas V, Rajbhandari P, Drew BG, Moore TM, Fluit AH, Reddish BR, Whitney KA, Georgia S, Vergnes L, Reue K, Liesa M, Shirihai O, van der Bliek AM, Chi NW, Mahata SK, Tiano JP, Hewitt SC, Tontonoz P, Korach KS, Mauvais-Jarvis F, Hevener AL. Estrogen receptor  $\alpha$  protects pancreatic  $\beta$ -cells from apoptosis by preserving mitochondrial function and suppressing endoplasmic reticulum stress. *J Biol Chem* 2018; **293**: 4735-4751 [DOI: [10.1074/jbc.M117.805069](https://doi.org/10.1074/jbc.M117.805069)]
- 159 **Torres MJ**, Kew KA, Ryan TE, Pennington ER, Lin CT, Buddo KA, Fix AM, Smith CA, Gilliam LA, Karvinen S, Lowe DA, Spangenburg EE, Zeczycki TN, Shaikh SR, Neufer PD. 17 $\beta$ -Estradiol Directly Lowers Mitochondrial Membrane Microviscosity and Improves Bioenergetic Function in Skeletal Muscle. *Cell Metab* 2018; **27**: 167-179.e7 [PMID: [29103922](https://pubmed.ncbi.nlm.nih.gov/29103922/) DOI: [10.1016/j.cmet.2017.10.003](https://doi.org/10.1016/j.cmet.2017.10.003)]
- 160 **Moor AN**, Gottipati S, Mallet RT, Sun J, Giblin FJ, Roque R, Cammarata PR. A putative mitochondrial mechanism for antioxidative cytoprotection by 17 $\beta$ -estradiol. *Exp Eye Res* 2004; **78**: 933-944 [PMID: [15051475](https://pubmed.ncbi.nlm.nih.gov/15051475/) DOI: [10.1016/j.exer.2004.01.001](https://doi.org/10.1016/j.exer.2004.01.001)]
- 161 **Borrás C**, Gambini J, López-Grueso R, Pallardó FV, Viña J. Direct antioxidant and protective effect of estradiol on isolated mitochondria. *Biochim Biophys Acta* 2010; **1802**: 205-211 [PMID: [19751829](https://pubmed.ncbi.nlm.nih.gov/19751829/) DOI: [10.1016/j.bbadi.2009.09.007](https://doi.org/10.1016/j.bbadi.2009.09.007)]
- 162 **Kushnir MM**, Rockwood AL, Bergquist J, Varshavsky M, Roberts WL, Yue B, Bunker AM, Meikle AW. High-sensitivity tandem mass spectrometry assay for serum estrone and estradiol. *Am J Clin Pathol* 2008; **129**: 530-539 [PMID: [18343779](https://pubmed.ncbi.nlm.nih.gov/18343779/) DOI: [10.1309/LC03BHQ5XJPJYEKG](https://doi.org/10.1309/LC03BHQ5XJPJYEKG)]
- 163 **Pauwels S**, Antonio L, Jans I, Lintermans A, Neven P, Claessens F, Decallonne B, Billen J, Vanderschueren D, Vermeersch P. Sensitive routine liquid chromatography-tandem mass spectrometry method for serum estradiol and estrone without derivatization. *Anal Bioanal Chem* 2013; **405**: 8569-8577 [PMID: [23892882](https://pubmed.ncbi.nlm.nih.gov/23892882/) DOI: [10.1007/s00216-013-7259-5](https://doi.org/10.1007/s00216-013-7259-5)]
- 164 **Jasuja GK**, Travison TG, Davda M, Rose AJ, Zhang A, Kushnir MM, Rockwood AL, Meikle W, Coville AD, D'Agostino R, Vasan RS, Bhagat S. Circulating estrone levels are associated prospectively with diabetes risk in men of the Framingham Heart Study. *Diabetes Care* 2013; **36**: 2591-2596 [PMID: [23690532](https://pubmed.ncbi.nlm.nih.gov/23690532/) DOI: [10.2337/dc12-2477](https://doi.org/10.2337/dc12-2477)]
- 165 **Brind JL**, Chervinsky K, Völgelman JH, Orentreich N. Radioimmunoassay of estrone sulfate in the serum of normal men after a non-chromatographic procedure that eliminates interference from dehydroepiandrosterone sulfate. *Steroids* 1990; **55**: 32-35 [PMID: [2137944](https://pubmed.ncbi.nlm.nih.gov/2137944/) DOI: [10.1016/0039-128x\(90\)90071-i](https://doi.org/10.1016/0039-128x(90)90071-i)]
- 166 **Sanchis D**, Balada F, Grasa MM, Virgili J, Peinado J, Monserrat C, Fernández-López JA, Remesar X, Alemany M. Oleoyl-estrone induces the loss of body fat in rats. *Int J Obes Relat Metab Disord* 1996; **20**: 588-594 [PMID: [8782737](https://pubmed.ncbi.nlm.nih.gov/8782737/)]
- 167 **López-Martí J**, Diaz-Silva M, Salas A, Grasa MM, Fernández-López J, Remesar X, Alemany M. Oleoyl-estrone induces the massive loss of body weight in Zucker *fa/fa* rats fed a high-energy hyperlipidic diet. *J Nutr Biochem* 2000; **11**: 530-535 [PMID: [11137888](https://pubmed.ncbi.nlm.nih.gov/11137888/) DOI: [10.1016/s0955-2863\(00\)00106-6](https://doi.org/10.1016/s0955-2863(00)00106-6)]
- 168 **Remesar X**, Fernández-López JA, Blay MT, Savall P, Salas A, Diaz-Silva M, Esteve M, Grasa MM, Alemany M. Effect of oral oleoyl-estrone on adipose tissue composition in male rats. *Int J Obes Relat Metab Disord* 2002; **26**: 1092-1102 [PMID: [12119575](https://pubmed.ncbi.nlm.nih.gov/12119575/) DOI: [10.1038/sj.ijo.0802056](https://doi.org/10.1038/sj.ijo.0802056)]
- 169 **Remesar X**, Guijarro P, Torregrosa C, Grasa MM, López J, Fernández-López JA, Alemany M. Oral oleoyl-estrone induces the rapid loss of body fat in Zucker lean rats fed a hyperlipidic diet. *Int J Obes Relat Metab Disord* 2000; **24**: 1405-1412 [PMID: [11126335](https://pubmed.ncbi.nlm.nih.gov/11126335/) DOI: [10.1038/sj.ijo.0801393](https://doi.org/10.1038/sj.ijo.0801393)]
- 170 **Remesar X**, Fernández-López JA, Alemany M. Oleoyl-estrone. *Med Res Rev* 2012; **32**: 1263-1291 [PMID: [21287573](https://pubmed.ncbi.nlm.nih.gov/21287573/) DOI: [10.1002/med.20240](https://doi.org/10.1002/med.20240)]
- 171 **Cabot C**, Grasa MM, Massanés RM, de Matteis R, Cinti S, Fernández-López JA, Remesar X, Alemany M. Oleoyl-estrone does not have direct estrogenic effects on rats. *Life Sci* 2001; **69**: 749-761 [PMID: [11487088](https://pubmed.ncbi.nlm.nih.gov/11487088/) DOI: [10.1016/S0024-3205\(01\)01159-6](https://doi.org/10.1016/S0024-3205(01)01159-6)]
- 172 **Alemany M**, Fernández-López JA, Petrobelli A, Granada M, Foz M, Remesar X. [Weight loss in a patient with morbid obesity under treatment with oleoyl-estrone]. *Med Clin (Barc)* 2003; **121**: 496-499 [PMID: [14588193](https://pubmed.ncbi.nlm.nih.gov/14588193/) DOI: [10.1016/s0025-7753\(03\)74000-7](https://doi.org/10.1016/s0025-7753(03)74000-7)]
- 173 **Cabot C**, Grasa MM, Fernández-López JA, Alemany M. Oleoyl-estrone treatment reduces the volume of white adipose tissue cells in the rat. *J Physiol Biochem* 2000; **56**: 369-370 [PMID: [11321531](https://pubmed.ncbi.nlm.nih.gov/11321531/) DOI: [10.1007/BF03179805](https://doi.org/10.1007/BF03179805)]
- 174 **Grasa MM**, Cabot C, Esteve M, Yubero P, Masanés RM, Blay MT, Vilà R, López-Martí J, Fernández-López JA, Remesar X, Alemany M. Daily oral oleoyl-estrone gavage induces a dose-

- dependent loss of fat in Wistar rats. *Obes Res* 2001; **9**: 202-209 [PMID: 11323446 DOI: 10.1038/oby.2001.22]
- 175 **Peinado-Onsurbe J**, Blay M, Casadomé L, Fernández-López JA, Remesar X, Alemany M. Effect of 24-h food deprivation on lipoprotein composition and oleoyl-estrone content of lean and obese Zucker rats. *Eur J Nutr* 2001; **40**: 155-160 [PMID: 11905956 DOI: 10.1007/s003940170003]
- 176 **Vihma V**, Koskela A, Turpeinen U, Hämäläinen E, Tiiainen A, Wähälä K, Tikkanen MJ, Adlercreutz H. Are there endogenous estrone fatty acyl esters in human plasma or ovarian follicular fluid? *J Steroid Biochem Mol Biol* 2011; **127**: 390-395 [PMID: 21708250 DOI: 10.1016/j.jsbmb.2011.06.007]
- 177 **Ferrer-Lorente R**, García-Peláez B, Gómez-Ollés S, Fernández-López JA, Remesar X, Alemany M. Effects of oral estrone on rat energy balance. *Steroids* 2005; **70**: 667-672 [PMID: 15885727 DOI: 10.1016/j.steroids.2005.03.007]
- 178 **Sanchis D**, Balada F, Grasa MM, Virgili J, Monserrat C, Fernández-López JA, Remesar X, Alemany M. Short-term handling of the slimming agent oleoyl-estrone in liposomes (Merlin-2) by the rat. *Mol Cell Biochem* 1997; **177**: 153-157 [PMID: 9450657 DOI: 10.1023/a:1006849128697]
- 179 **Strassburg S**, Pfluger PT, Chaudhary N, Tso P, Tschöp MH, Anker SD, Nogueiras R, Pérez-Tilve D. Action profile of the antiobesity drug candidate oleoyl-estrone in rats. *Obesity (Silver Spring)* 2010; **18**: 2260-2267 [PMID: 20339368 DOI: 10.1038/oby.2010.53]
- 180 **Sanchis D**, Balada F, Picó C, Grasa MM, Virgili J, Farrerons C, Palou A, Fernández-López JA, Remesar X, Alemany M. Rats receiving the slimming agent oleoyl-estrone in liposomes (Merlin-2) decrease food intake but maintain thermogenesis. *Arch Physiol Biochem* 1997; **105**: 663-672 [PMID: 9693713 DOI: 10.1076/apab.105.7.663.11391]
- 181 **Vilà R**, Cabot C, Villarreal L, Monegal A, Ayet E, Romero MM, Grasa MM, Esteve M, Fernández-López JA, Remesar X, Alemany M. Oleoyl-estrone is a precursor of an estrone-derived ponderostat signal. *J Steroid Biochem Mol Biol* 2011; **124**: 99-111 [PMID: 21310232 DOI: 10.1016/j.jsbmb.2011.01.017]
- 182 **Cabot C**, Masanés R, Bulló M, García-Lorda P, Fernández-López JA, Salas-Salvadó J, Alemany M. Plasma acyl-estrone levels are altered in obese women. *Endocr Res* 2000; **26**: 465-476 [PMID: 11019908 DOI: 10.3109/07435800009066180]
- 183 **Bailly J**, Raab S, Clerc R, Sebokova E, Krust A, Chambon P. The effect of oleoyl-estrone on body weight is mediated via the  $\alpha$ -estrogen receptor and not the beta estrogen receptor. *Obes Rev* 2005; **6**: 48
- 184 **Borràs M**, Guerendain M, Cabo tC, Cederroth M, Esteve M, Remesar X, Grasa MM. The estrogen receptor alpha agonist ICI 182,780 partially blocked oleoyl-estrone slimming action in C57BL6 mice fed with cafeteria diet. *Obes Rev* 2010; **11**: 158
- 185 **Cabot C**, González-Martínez D, Fernández-López JA, Alemany M. In the rat, estrone sulphate is the main serum metabolite of oral oleoyl-estrone. *J Endocrinol Invest* 2007; **30**: 376-381 [PMID: 17598968 DOI: 10.1007/BF03346313]
- 186 **Hojo Y**, Murakami G, Mukai H, Higo S, Hatanaka Y, Ogiue-Ikeda M, Ishii H, Kimoto T, Kawato S. Estrogen synthesis in the brain--role in synaptic plasticity and memory. *Mol Cell Endocrinol* 2008; **290**: 31-43 [PMID: 18541362 DOI: 10.1016/j.mce.2008.04.017]
- 187 **Killinger DW**, Strutt BJ, Roncari DA, Khalil MW. Estrone formation from dehydroepiandrosterone in cultured human breast adipose stromal cells. *J Steroid Biochem Mol Biol* 1995; **52**: 195-201 [PMID: 7873453 DOI: 10.1016/0960-0760(94)00164-h]
- 188 **Westman EC**. Is dietary carbohydrate essential for human nutrition? *Am J Clin Nutr* 2002; **75**: 951-3; author reply 953 [PMID: 11976176 DOI: 10.1093/ajcn/75.5.951]
- 189 **Remesar X**, Alemany M. Dietary energy partition: The central role of glucose. *Int J Mol Sci* 2020; **21** [PMID: 33086579 DOI: 10.3390/ijms21207729]
- 190 **Ross BD**, Hems R, Krebs HA. The rate of gluconeogenesis from various precursors in the perfused rat liver. *Biochem J* 1967; **102**: 942-951 [PMID: 16742514 DOI: 10.1042/bj1020942]
- 191 **Irias JJ**, Greenberg RE. Relationship of renal gluconeogenesis to control of ammonia formation. *Am J Physiol* 1972; **223**: 750-755 [PMID: 5075150 DOI: 10.1152/ajplegacy.1972.223.4.750]
- 192 **Habold C**, Foltzer-Jourdainne C, Le Maho Y, Lignot JH, Oudart H. Intestinal gluconeogenesis and glucose transport according to body fuel availability in rats. *J Physiol* 2005; **566**: 575-586 [PMID: 15878950 DOI: 10.1111/jphysiol.2005.085217]
- 193 **Arriarán S**, Agnelli S, Sabater D, Remesar X, Fernández-López JA, Alemany M. Evidences of basal lactate production in the main white adipose tissue sites of rats. Effects of sex and a cafeteria diet. *PLoS One* 2015; **10**: e0119572 [PMID: 25741703 DOI: 10.1371/journal.pone.0119572]
- 194 **Ho-Palma AC**, Rotondo F, Romero MM, Fernández-López JA, Remesar X, Alemany M. Use of  $^{14}$ C-glucose by primary cultures of mature rat epididymal adipocytes. Marked release of lactate and glycerol, but limited lipogenesis in the absence of external stimuli. *Adipocyte* 2018; **7**: 204-217 [PMID: 29708458 DOI: 10.1080/21623945.2018.1460020]
- 195 **Oliva L**, Fernández-López JA, Remesar X, Alemany M. The anomeric nature of glucose and its implications on its analyses and the influence of diet: Are routine glycaemia measurements reliable enough? *J Endocrinol Met* 2019; **9**: 63-70 [DOI: 10.14740/jem555]
- 196 **Rotondo F**, Ho-Palma AC, Remesar X, Fernández-López JA, Romero MM, Alemany M. Glycerol is synthesized and secreted by adipocytes to dispose of excess glucose, via glycerogenesis and increased acyl-glycerol turnover. *Sci Rep* 2017; **7**: 8983 [PMID: 28827624 DOI: 10.1038/s41598-017-09450-4]

- 197 **Santos-Marcos JA**, Pérez-Jiménez F, Camargo A. The role of diet and intestinal microbiota in the development of metabolic syndrome. *J Nutr Biochem* 2019; **70**: 1-27 [PMID: 31082615 DOI: 10.1016/j.jnutbio.2019.03.017]
- 198 **Alexander CM**, Landsman PB, Grundy SM. Metabolic syndrome and hyperglycemia: congruence and divergence. *Am J Cardiol* 2006; **98**: 982-985 [PMID: 16996888 DOI: 10.1016/j.amjcard.2006.04.046]
- 199 **Gallagher EJ**, Leroith D, Karnieli E. The metabolic syndrome--from insulin resistance to obesity and diabetes. *Med Clin North Am* 2011; **95**: 855-873 [PMID: 21855696 DOI: 10.1016/j.mcna.2011.06.001]
- 200 **Shalitin S**, Moreno LA. Obesity, Metabolic syndrome and nutrition. *World Rev Nutr Diet* 2018; **117**: 15-38 [PMID: 29393112 DOI: 10.1159/000484498]
- 201 **Monnerie S**, Comte B, Ziegler D, Morais JA, Pujos-Guillot E, Gaudreau P. Metabolomic and lipidomic signatures of metabolic syndrome and its physiological components in adults: A systematic review. *Sci Rep* 2020; **10**: 669 [PMID: 31959772 DOI: 10.1038/s41598-019-56909-7]
- 202 **Dandona P**, Aljada A, Chaudhuri A, Mohanty P, Garg R. Metabolic syndrome: a comprehensive perspective based on interactions between obesity, diabetes, and inflammation. *Circulation* 2005; **111**: 1448-1454 [PMID: 15781756 DOI: 10.1161/01.CIR.0000158483.13093.9D]
- 203 **Montecucco F**, Mach F, Pende A. Inflammation is a key pathophysiological feature of metabolic syndrome. *Mediators Inflamm* 2013; **2013**: 135984 [PMID: 23710114 DOI: 10.1155/2013/135984]
- 204 **Qiao Q**, Gao W, Zhang L, Nyamdorj R, Tuomilehto J. Metabolic syndrome and cardiovascular disease. *Ann Clin Biochem* 2007; **44**: 232-263 [PMID: 17456293 DOI: 10.1258/000456307780480963]
- 205 **Yang Z**, Hu Y, Zhang J, Xu L, Zeng R, Kang D. Estradiol therapy and breast cancer risk in perimenopausal and postmenopausal women: a systematic review and meta-analysis. *Gynecol Endocrinol* 2017; **33**: 87-92 [PMID: 27898258 DOI: 10.1080/09513590.2016.1248932]
- 206 **Pan A**, Keum N, Okereke OI, Sun Q, Kivimaki M, Rubin RR, Hu FB. Bidirectional association between depression and metabolic syndrome: a systematic review and meta-analysis of epidemiological studies. *Diabetes Care* 2012; **35**: 1171-1180 [PMID: 22517938 DOI: 10.2337/dc11-2055]
- 207 **Lambert GW**, Straznicky NE, Lambert EA, Dixon JB, Schlaich MP. Sympathetic nervous activation in obesity and the metabolic syndrome--causes, consequences and therapeutic implications. *Pharmacol Ther* 2010; **126**: 159-172 [PMID: 20171982 DOI: 10.1016/j.pharmthera.2010.02.002]
- 208 **Ornstrup MJ**, Kjær TN, Harsløf T, Stødkilde-Jørgensen H, Hougaard DM, Cohen A, Pedersen SB, Langdahl BL. Adipose tissue, estradiol levels, and bone health in obese men with metabolic syndrome. *Eur J Endocrinol* 2015; **172**: 205-216 [PMID: 25416724 DOI: 10.1530/EJE-14-0792]
- 209 **Nguyen KD**, Chawla A. Metabolic syndrome driven by macrophage interactions with the adipose tissue. *Nat Med* 2011; **17**: 43-43
- 210 **Liu R**, Nikolajczyk BS. Tissue immune cells fuel obesity-associated inflammation in adipose tissue and beyond. *Front Immunol* 2019; **10**: 1587 [PMID: 31379820 DOI: 10.3389/fimmu.2019.01587]
- 211 **Keane D**, Kelly S, Healy NP, McArdle MA, Holohan K, Roche HM. Diet and metabolic syndrome: an overview. *Curr Vasc Pharmacol* 2013; **11**: 842-857 [PMID: 24168443 DOI: 10.2174/15701611113116660173]
- 212 **Hosseini Z**, Whiting SJ, Vatanparast H. Current evidence on the association of the metabolic syndrome and dietary patterns in a global perspective. *Nutr Res Rev* 2016; **29**: 152-162 [PMID: 27955720 DOI: 10.1017/S095442241600007X]
- 213 **Bazzano LA**, Hu T, Reynolds K, Yao L, Bunol C, Liu Y, Chen CS, Klag MJ, Whelton PK, He J. Effects of low-carbohydrate and low-fat diets: a randomized trial. *Ann Intern Med* 2014; **161**: 309-318 [PMID: 25178568 DOI: 10.7326/M14-0180]
- 214 **Stange R**, Pflugbeil C, Michalsen A, Uehleke B. Therapeutic fasting in patients with metabolic syndrome and impaired insulin resistance. *Forsch Komplementmed* 2013; **20**: 421-426 [PMID: 24434756 DOI: 10.1159/000357875]
- 215 **Castellana M**, Conte E, Cignarelli A, Perrini S, Giustina A, Giovanella L, Giorgino F, Trimboli P. Efficacy and safety of very low calorie ketogenic diet (VLCKD) in patients with overweight and obesity: A systematic review and meta-analysis. *Rev Endocr Metab Disord* 2020; **21**: 5-16 [PMID: 31705259 DOI: 10.1007/s11154-019-09514-y]
- 216 **Watanabe M**, Tuccinardi D, Ernesti I, Basciani S, Mariani S, Genco A, Manfrini S, Lubrano C, Gnessi L. Scientific evidence underlying contraindications to the ketogenic diet: An update. *Obes Rev* 2020; **21**: e13053 [PMID: 32648647 DOI: 10.1111/obr.13053]
- 217 **Nael R**, Montgomery PS, Scott KJ, Blevins SM, Gardner AW. Gender differences in the prevalence and management of metabolic syndrome and its components in patients with peripheral artery disease. *Angiology* 2011; **62**: 657-661 [PMID: 21511682 DOI: 10.1177/0003319711404025]
- 218 **Laudisio A**, Marzetti E, Antonica L, Pagano F, Vetrano DL, Bernabei R, Zuccalà G. Metabolic syndrome and quality of life in the elderly: age and gender differences. *Eur J Nutr* 2013; **52**: 307-316 [PMID: 22406906 DOI: 10.1007/s00394-012-0337-1]
- 219 **Roche HM**. Dietary modulation of energy homeostasis and metabolic-inflammation. *Proc Nutr Soc* 2019; **78**: 313-318 [PMID: 30704542 DOI: 10.1017/S0029665118002872]
- 220 **Ghorabi S**, Esteghamati A, Azam K, Daneshzad E, Sadeghi O, Salari-Moghaddam A, Azadbakht L, Djafarian K. Association between dietary inflammatory index and components of metabolic

- syndrome. *J Cardiovasc Thorac Res* 2020; **12**: 27-34 [PMID: 32211135 DOI: 10.34172/jcvtr.2020.05]
- 221 **Bianchi VE**, Locatelli V. Testosterone a key factor in gender related metabolic syndrome. *Obes Rev* 2018; **19**: 557-575 [PMID: 29356299 DOI: 10.1111/obr.12633]
- 222 **Haider A**, Yassin A, Haider KS, Doros G, Saad F, Rosano GM. Men with testosterone deficiency and a history of cardiovascular diseases benefit from long-term testosterone therapy: observational, real-life data from a registry study. *Vasc Health Risk Manag* 2016; **12**: 251-261 [PMID: 27366080 DOI: 10.2147/VHRM.S108947]
- 223 **Traish AM**, Haider A, Haider KS, Doros G, Saad F. Long-term testosterone therapy improves cardiometabolic function and reduces risk of cardiovascular disease in men with hypogonadism: A real-life observational registry study setting comparing treated and untreated (control) groups. *J Cardiovasc Pharmacol Ther* 2017; **22**: 414-433 [PMID: 28421834 DOI: 10.1177/1074248417691136]
- 224 **Heufelder AE**, Saad F, Bunck MC, Gooren L. Fifty-two-week treatment with diet and exercise plus transdermal testosterone reverses the metabolic syndrome and improves glycemic control in men with newly diagnosed type 2 diabetes and subnormal plasma testosterone. *J Androl* 2009; **30**: 726-733 [PMID: 19578132 DOI: 10.2164/jandro.108.007005]
- 225 **Groti K**, Žuran I, Antonič B, Foršnarič L, Pfeifer M. The impact of testosterone replacement therapy on glycemic control, vascular function, and components of the metabolic syndrome in obese hypogonadal men with type 2 diabetes. *Aging Male* 2018; **21**: 158-169 [PMID: 29708829 DOI: 10.1080/13685538.2018.1468429]
- 226 **Hwang K**, Miner M. Controversies in testosterone replacement therapy: testosterone and cardiovascular disease. *Asian J Androl* 2015; **17**: 187-191 [PMID: 25652628 DOI: 10.4103/1008-682X.146968]
- 227 **Chalas E**. Ovaries, estrogen, and longevity. *Obstet Gynecol* 2013; **121**: 701-702 [PMID: 23635666 DOI: 10.1097/AOG.0b013e31828af732]
- 228 **Caruso S**, Cianci S, Amore FF, Ventura B, Bambili E, Spadola S, Cianci A. Quality of life and sexual function of naturally postmenopausal women on an ultralow-concentration estriol vaginal gel. *Menopause* 2016; **23**: 47-54 [PMID: 26079974 DOI: 10.1097/GME.0000000000000485]
- 229 **Viña J**, Sastre J, Pallardó FV, Gambini J, Borrás C. Modulation of longevity-associated genes by estrogens or phytoestrogens. *Biol Chem* 2008; **389**: 273-277 [PMID: 18177268 DOI: 10.1515/BC.2008.027]
- 230 **vom Saal FS**, Welshons WV. Endocrine disruptors: Manmade and natural oestrogens: opposite effects on assisted reproduction. *Nat Rev Endocrinol* 2016; **12**: 251-252 [PMID: 26988616 DOI: 10.1038/nrendo.2016.38]
- 231 **Goodsell DS**. The molecular perspective: tamoxifen and the estrogen receptor. *Stem Cells* 2002; **20**: 267-268 [PMID: 12004085 DOI: 10.1634/stemcells.20-3-267]
- 232 **Zhang Z**, Kang D, Li H. The effects of testosterone on bone health in males with testosterone deficiency: a systematic review and meta-analysis. *BMC Endocr Disord* 2020; **20**: 33 [PMID: 32145741 DOI: 10.1186/s12902-020-0509-6]
- 233 **Borthwick EB**, Houston MP, Coughtrie MW, Burchell A. The antihyperglycemic effect of estrone sulfate in genetically obese-diabetic (ob/ob) mice is associated with reduced hepatic glucose-6-phosphatase. *Horm Metab Res* 2001; **33**: 721-726 [PMID: 11753757 DOI: 10.1055/s-2001-19136]
- 234 **Liu S**, Mauvais-Jarvis F. Rapid, nongenomic estrogen actions protect pancreatic islet survival. *Islets* 2009; **1**: 273-275 [PMID: 20634925 DOI: 10.4161/isl.1.3.9781]
- 235 **Bian C**, Bai B, Gao Q, Li S, Zhao Y. 17 $\beta$ -Estradiol regulates glucose metabolism and insulin secretion in rat islet  $\beta$  cells through GPER and Akt/mTOR/GLUT2 pathway. *Front Endocrinol (Lausanne)* 2019; **10**: 531 [PMID: 31447779 DOI: 10.3389/fendo.2019.00531]
- 236 **Le May C**, Chu K, Hu M, Ortega CS, Simpson ER, Korach KS, Tsai MJ, Mauvais-Jarvis F. Estrogens protect pancreatic beta-cells from apoptosis and prevent insulin-deficient diabetes mellitus in mice. *Proc Natl Acad Sci U S A* 2006; **103**: 9232-9237 [PMID: 16754860 DOI: 10.1073/pnas.0602956103]
- 237 **Nadal A**, Alonso-Magdalena P, Soriano S, Ropero AB, Quesada I. The role of oestrogens in the adaptation of islets to insulin resistance. *J Physiol* 2009; **587**: 5031-5037 [PMID: 19687125 DOI: 10.1113/jphysiol.2009.177188]
- 238 **Alonso-Magdalena P**, Ropero AB, Carrera MP, Cederroth CR, Baquié M, Gauthier BR, Nef S, Stefani E, Nadal A. Pancreatic insulin content regulation by the estrogen receptor ER alpha. *PLoS One* 2008; **3**: e2069 [PMID: 18446233 DOI: 10.1371/journal.pone.0002069]
- 239 **Allard C**, Morford JJ, Xu B, Salwen B, Xu W, Desmoulin L, Zsombok A, Kim JK, Levin ER, Mauvais-Jarvis F. Loss of nuclear and membrane estrogen receptor- $\alpha$  differentially impairs insulin secretion and action in male and female mice. *Diabetes* 2019; **68**: 490-501 [PMID: 30305367 DOI: 10.2337/db18-0293]
- 240 **Godsland IF**. Oestrogens and insulin secretion. *Diabetologia* 2005; **48**: 2213-2220 [PMID: 16193292 DOI: 10.1007/s00125-005-1930-0]
- 241 **Santos RS**, Batista TM, Camargo RL, Morato PN, Borck PC, Leite NC, Kurauti MA, Wanschel AC, Nadal Á, Clegg DJ, Carneiro EM. Lacking of estradiol reduces insulin exocytosis from pancreatic  $\beta$ -cells and increases hepatic insulin degradation. *Steroids* 2016; **114**: 16-24 [PMID: 27192429 DOI: 10.1016/j.steroids.2016.05.002]
- 242 **Riant E**, Waget A, Cogo H, Arnal JF, Burcelin R, Gourdy P. Estrogens protect against high-fat diet-

- induced insulin resistance and glucose intolerance in mice. *Endocrinology* 2009; **150**: 2109-2117 [PMID: 19164473 DOI: 10.1210/en.2008-0971]
- 243 **Camporez JP**, Lyu K, Goldberg EL, Zhang D, Cline GW, Jurczak MJ, Dixit VD, Petersen KF, Shulman GI. Anti-inflammatory effects of oestrogen mediate the sexual dimorphic response to lipid-induced insulin resistance. *J Physiol* 2019; **597**: 3885-3903 [PMID: 31206703 DOI: 10.1111/jp.277270]
- 244 **Inada A**, Fujii NL, Inada O, Higaki Y, Furuichi Y, Nabeshima YI. Effects of 17 $\beta$ -estradiol and androgen on glucose metabolism in skeletal muscle. *Endocrinology* 2016; **157**: 4691-4705 [PMID: 27653033 DOI: 10.1210/en.2016-1261]
- 245 **Jelenik T**, Roden M. How estrogens prevent from lipid-induced insulin resistance. *Endocrinology* 2013; **154**: 989-992 [PMID: 23429711 DOI: 10.1210/en.2013-1112]
- 246 **Stubbins RE**, Najjar K, Holcomb VB, Hong J, Núñez NP. Oestrogen alters adipocyte biology and protects female mice from adipocyte inflammation and insulin resistance. *Diabetes Obes Metab* 2012; **14**: 58-66 [PMID: 21834845 DOI: 10.1111/j.1463-1326.2011.01488.x]
- 247 **Shen M**, Kumar SP, Shi H. Estradiol regulates insulin signaling and inflammation in adipose tissue. *Horm Mol Biol Clin Investig* 2014; **17**: 99-107 [PMID: 25372734 DOI: 10.1515/hmbci-2014-0007]
- 248 **Guillaume M**, Handgraaf S, Fabre A, Raymond-Letron I, Riant E, Montagner A, Vinel A, Buscato M, Smirnova N, Fontaine C, Guillou H, Arnal JF, Gourdy P. Selective activation of estrogen receptor  $\alpha$  activation function-1 is sufficient to prevent obesity, steatosis, and insulin resistance in mouse. *Am J Pathol* 2017; **187**: 1273-1287 [PMID: 28502695 DOI: 10.1016/j.ajpath.2017.02.013]
- 249 **Handgraaf S**, Riant E, Fabre A, Waget A, Burcelin R, Lière P, Krust A, Champon P, Arnal JF, Gourdy P. Prevention of obesity and insulin resistance by estrogens requires ER $\alpha$  activation function-2 (ER $\alpha$ AF-2), whereas ER $\alpha$ AF-1 is dispensable. *Diabetes* 2013; **62**: 4098-4108 [PMID: 23903353 DOI: 10.2337/db13-0282]
- 250 **Li R**, Shen Y. Estrogen and brain: synthesis, function and diseases. *Front Biosci* 2005; **10**: 257-267 [PMID: 15574366 DOI: 10.2741/1525]
- 251 **Biegan A**, Alia-Klein N, Alexoff DL, Fowler JS, Kim SW, Logan J, Pareto D, Preston-Campbell R, Wang GJ, Hildebrandt T. Relationship of estrogen synthesis capacity in the brain with obesity and self-control in men and women. *Proc Natl Acad Sci U S A* 2020; **117**: 22962-22966 [PMID: 32868418 DOI: 10.1073/pnas.2006117117]
- 252 **Sellers KJ**, Denley MCS, Saito A, Foster EM, Salgarella I, Delogu A, Kamiya A, Srivastava DP. Brain-synthesized oestrogens regulate cortical migration in a sexually divergent manner. *Eur J Neurosci* 2020; **52**: 2646-2663 [PMID: 32314480 DOI: 10.1111/ejn.14755]
- 253 **Cornil CA**, Charlier TD. Rapid behavioural effects of oestrogens and fast regulation of their local synthesis by brain aromatase. *J Neuroendocrinol* 2010; **22**: 664-673 [PMID: 20456609 DOI: 10.1111/j.1365-2826.2010.02023.x]
- 254 **Ulhaq ZS**. Estrogen – serotonin interaction and its implication on insulin resistance. *J Intern Med* 2019; **55**: 76-81 [DOI: 10.1080/20905068.2019.1670413]
- 255 **Pratchayasakul W**, Sa-Nguanmoo P, Sivasinprasasn S, Pintana H, Tawinvisan R, Sripathchandee J, Kumfu S, Chattipakorn N, Chattipakorn SC. Obesity accelerates cognitive decline by aggravating mitochondrial dysfunction, insulin resistance and synaptic dysfunction under estrogen-deprived conditions. *Horm Behav* 2015; **72**: 68-77 [PMID: 25989597 DOI: 10.1016/j.ybeh.2015.04.023]
- 256 **Berlanga-Acosta J**, Guillén-Nieto G, Rodríguez-Rodríguez N, Bringas-Vega ML, García-del-Barco-Herrera D, Berlanga-Sáez JO, García-Ojalvo A, Valdés-Sosa MJ, Valdés-Sosa PA. Insulin resistance at the crossroad of Alzheimer disease pathology: A review. *Front Endocrinol (Lausanne)* 2020; **11**: 560375 [PMID: 33224105 DOI: 10.3389/fendo.2020.560375]
- 257 **Kellar D**, Craft S. Brain insulin resistance in Alzheimer's disease and related disorders: mechanisms and therapeutic approaches. *Lancet Neurol* 2020; **19**: 758-766 [PMID: 32730766 DOI: 10.1016/S1474-4422(20)30231-3]
- 258 **Inestrosa NC**, Marzolo MP, Bonnefont AB. Cellular and molecular basis of estrogen's neuroprotection. Potential relevance for Alzheimer's disease. *Mol Neurobiol* 1998; **17**: 73-86 [PMID: 9887447 DOI: 10.1007/BF02802025]
- 259 **Greenfield JP**, Leung LW, Cai D, Kaasik K, Gross RS, Rodriguez-Boulan E, Greengard P, Xu H. Estrogen lowers Alzheimer beta-amyloid generation by stimulating trans-Golgi network vesicle biogenesis. *J Biol Chem* 2002; **277**: 12128-12136 [PMID: 11823458 DOI: 10.1074/jbc.M110009200]
- 260 **Zhao L**, Yao J, Mao Z, Chen S, Wang Y, Brinton RD. 17 $\beta$ -Estradiol regulates insulin-degrading enzyme expression via an ER $\beta$ /PI3-K pathway in hippocampus: relevance to Alzheimer's prevention. *Neurobiol Aging* 2011; **32**: 1949-1963 [PMID: 20053478 DOI: 10.1016/j.neurobiolaging.2009.12.010]
- 261 **Ma LH**, Lin GZ, Wang M. Association between estrogen and female patients with Alzheimer's disease: a meta-analysis. *Int J Clin Exp Med* 2017; **10**: 135-141
- 262 **Panidis DK**, Matalliotakis IM, Rousso DH, Kourtis AI, Koumantakis EE. The role of estrogen replacement therapy in Alzheimer's disease. *Eur J Obstet Gyn R B* 2001; **95**: 86-91 [DOI: 10.1016/S0301-2115(00)00373-0]
- 263 **Tian Z**, Fan J, Zhao Y, Bi S, Si L, Liu Q. Estrogen receptor beta treats Alzheimer's disease. *Neural Regen Res* 2013; **8**: 420-426 [PMID: 25206683 DOI: 10.3969/j.issn.1673-5374.2013.05.005]
- 264 **Tang Y**, Min Z, Xiang XJ, Liu L, Ma YL, Zhu BL, Song L, Tang J, Deng XJ, Yan Z, Chen GJ. Estrogen-related receptor alpha is involved in Alzheimer's disease-like pathology. *Exp Neurol* 2018;

- 265      **305:** 89-96 [PMID: 29641978 DOI: 10.1016/j.expneurol.2018.04.003]  
**Kimura Y**, Buddington KK, Buddington RK. The influence of estradiol and diet on small intestinal glucose transport in ovariectomized rats. *Exp Biol Med (Maywood)* 2004; **229**: 227-234 [PMID: 14988514 DOI: 10.1177/153537020422900302]
- 266      **Garrido P**, Morán J, Alonso A, González S, González C. 17 $\beta$ -estradiol activates glucose uptake via GLUT4 translocation and PI3K/Akt signaling pathway in MCF-7 cells. *Endocrinology* 2013; **154**: 1979-1989 [PMID: 23546602 DOI: 10.1210/en.2012-1558]
- 267      **Shi J**, Zhang YQ, Simpkins JW. Effects of 17 $\beta$ -estradiol on glucose transporter 1 expression and endothelial cell survival following focal ischemia in the rats. *Exp Brain Res* 1997; **117**: 200-206 [PMID: 9419067 DOI: 10.1007/s002210050216]
- 268      **Devries MC**, Hamadeh MJ, Graham TE, Tarnopolsky MA. 17 $\beta$ -estradiol supplementation decreases glucose rate of appearance and disappearance with no effect on glycogen utilization during moderate intensity exercise in men. *J Clin Endocrinol Metab* 2005; **90**: 6218-6225 [PMID: 16118338 DOI: 10.1210/jc.2005-0926]
- 269      **Yan H**, Yang W, Zhou F, Li X, Pan Q, Shen Z, Han G, Newell-Fugate A, Tian Y, Majeti R, Liu W, Xu Y, Wu C, Allred K, Allred C, Sun Y, Guo S. Estrogen improves insulin sensitivity and suppresses gluconeogenesis via the transcription factor Foxo1. *Diabetes* 2019; **68**: 291-304 [PMID: 30487265 DOI: 10.2337/db18-0638]
- 270      **Boscaro C**, Carotti M, Albiero M, Trenti A, Fadini GP, Trevisi L, Sandonà D, Cignarella A, Bolego C. Non-genomic mechanisms in the estrogen regulation of glycolytic protein levels in endothelial cells. *FASEB J* 2020; **34**: 12768-12784 [PMID: 32757462 DOI: 10.1096/fj.202001130R]
- 271      **Narasimhan A**, Sampath S, Jayaraman S, Karundevi B. Estradiol favors glucose oxidation in gastrocnemius muscle through modulation of insulin signaling molecules in adult female rats. *Endocr Res* 2013; **38**: 251-262 [PMID: 23488804 DOI: 10.3109/07435800.2013.775148]
- 272      **Imbert-Fernandez Y**, Clem BF, O'Neal J, Kerr DA, Spaulding R, Lanceta L, Clem AL, Telang S, Chesney J. Estradiol stimulates glucose metabolism via 6-phosphofructo-2-kinase (PFKFB3). *J Biol Chem* 2014; **289**: 9440-9448 [PMID: 24515104 DOI: 10.1074/jbc.M113.529990]
- 273      **Sun Y**, Gu X, Zhang E, Park MA, Pereira AM, Wang S, Morrison T, Li C, Blenis J, Gerbaudo VH, Henske EP, Yu JJ. Estradiol promotes pentose phosphate pathway addiction and cell survival via reactivation of Akt in mTORC1 hyperactive cells. *Cell Death Dis* 2014; **5**: e1231 [PMID: 24832603 DOI: 10.1038/cddis.2014.204]
- 274      **Luo F**, Guo Y, Ruan GY, Peng R, Li XP. Estrogen lowers triglyceride via regulating hepatic APOA5 expression. *Lipids Health Dis* 2017; **16**: 72 [PMID: 28376804 DOI: 10.1186/s12944-017-0463-0]
- 275      **Shwaery GT**, Vita JA, Keaney JF Jr. Antioxidant protection of LDL by physiological concentrations of 17 $\beta$ -estradiol. Requirement for estradiol modification. *Circulation* 1997; **95**: 1378-1385 [PMID: 9118503 DOI: 10.1161/01.cir.95.6.1378]
- 276      **Ting WJ**, Huang CY, Jiang CH, Lin YM, Chung LC, Shen CY, Pai P, Lin KH, Viswanadha VP, Liao SC. Treatment with 17 $\beta$ -estradiol reduced body weight and the risk of cardiovascular disease in a high-fat diet-induced animal model of obesity. *Int J Mol Sci* 2017; **18** [PMID: 28335423 DOI: 10.3390/ijms18030629]
- 277      **Tian GX**, Sun Y, Pang CJ, Tan AH, Gao Y, Zhang HY, Yang XB, Li ZX, Mo ZN. Oestradiol is a protective factor for non-alcoholic fatty liver disease in healthy men. *Obes Rev* 2012; **13**: 381-387 [PMID: 22239319 DOI: 10.1111/j.1467-789X.2011.00978.x]
- 278      **Kavanagh K**, Davis MA, Zhang L, Wilson MD, Register TC, Adams MR, Rudel LL, Wagner JD. Estrogen decreases atherosclerosis in part by reducing hepatic acyl-CoA:cholesterol acyltransferase 2 (ACAT2) in monkeys. *Arterioscler Thromb Vasc Biol* 2009; **29**: 1471-1477 [PMID: 19759374 DOI: 10.1161/ATVBAHA.109.191825]
- 279      **Litwak SA**, Wilson JL, Chen W, Garcia-Rudaz C, Khaksari M, Cowley MA, Enriori PJ. Estradiol prevents fat accumulation and overcomes leptin resistance in female high-fat diet mice. *Endocrinology* 2014; **155**: 4447-4460 [PMID: 25147981 DOI: 10.1210/en.2014-1342]
- 280      **MacDonald TL**, MacPherson R, Castellani L, Cervone D, Anderson E, Wright DC, Dyck DJ. Estradiol does not directly regulate adipose lipolysis. *Adipocyte* 2017; **6**: 76-86 [PMID: 28425842 DOI: 10.1080/21623945.2017.1287638]
- 281      **Lundholm L**, Zang H, Hirschberg AL, Gustafsson JA, Arner P, Dahlman-Wright K. Key lipogenic gene expression can be decreased by estrogen in human adipose tissue. *Fertil Steril* 2008; **90**: 44-48 [PMID: 18222430 DOI: 10.1016/j.fertnstert.2007.06.011]
- 282      **Ahluwalia A**, Hoa N, Ge L, Blumberg B, Levin ER. Mechanisms by which membrane and nuclear ER alpha inhibit adipogenesis in cells isolated from female mice. *Endocrinology* 2020; **161** [PMID: 32976570 DOI: 10.1210/endocr/bqaa175]
- 283      **Oliva L**, Aranda T, Alemany M, Fernández-López JA, Remesar X. Unconnected body accrual of dietary lipid and protein in rats fed diets with different lipid and protein content. *Mol Nutr Food Res* 2020; **64**: e2000265 [PMID: 32521082 DOI: 10.1002/mnfr.202000265]
- 284      **Oliva L**, Alemany M, Fernández-López JA, Remesar X. Estradiol determine liver lipid deposition in rats fed standard diets unbalanced with excess lipid or protein. *Chemrxiv* 2020 [DOI: 10.26434/chemrxiv.13072100]
- 285      **Contreras JL**, Smyth CA, Bilbao G, Young CJ, Thompson JA, Eckhoff DE. 17 $\beta$ -estradiol protects isolated human pancreatic islets against proinflammatory cytokine-induced cell death: Molecular mechanisms and islet functionality. *Transplantation* 2002; **74**: 1252-1259 [DOI: 10.1002/tx.10044]

- 286 **de Cleyn K**, Buytaert P, Coppens M. Carbohydrate metabolism during hormonal substitution therapy. *Maturitas* 1989; **11**: 235-242 [PMID: 2687646 DOI: 10.1016/0378-5122(89)90216-8]
- 287 **Stirone C**, Duckles SP, Krause DN, Procaccio V. Estrogen increases mitochondrial efficiency and reduces oxidative stress in cerebral blood vessels. *Mol Pharmacol* 2005; **68**: 959-965 [PMID: 15994367 DOI: 10.1124/mol.105.014662]
- 288 **Viña J**, Sastre J, Pallardó FV, Gambini J, Borrás C. Role of mitochondrial oxidative stress to explain the different longevity between genders: protective effect of estrogens. *Free Radic Res* 2006; **40**: 1359-1365 [PMID: 17090425 DOI: 10.1080/10715760600952851]
- 289 **Klinge CM**. Estrogenic control of mitochondrial function and biogenesis. *J Cell Biochem* 2008; **105**: 1342-1351 [PMID: 18846505 DOI: 10.1002/jcb.21936]
- 290 **Abelenda M**, Puerta M. Dual control of cytochrome-C oxidase activity by female sex steroids. *Eur J Endocrinol* 1999; **141**: 630-636 [PMID: 10601967 DOI: 10.1530/eje.0.1410630]
- 291 **Moreno AJM**, Moreira PI, Custódio JBA, Santos MS. Mechanism of inhibition of mitochondrial ATP synthase by 17 $\beta$ -Estradiol. *J Bioenerg Biomembr* 2013; **45**: 261-270 [DOI: 10.1007/s10863-012-9497-1]
- 292 **Mamounis KJ**, Hernandez MR, Margolies N, Yasrebi A, Roepke TA. Interaction of 17 $\beta$ -estradiol and dietary fatty acids on energy and glucose homeostasis in female mice. *Nutr Neurosci* 2018; **21**: 715-728 [PMID: 28686546 DOI: 10.1080/1028415X.2017.1347374]
- 293 **Martínez de Morentin PB**, González-García I, Martins L, Lage R, Fernández-Mallo D, Martínez-Sánchez N, Ruiz-Pino F, Liu J, Morgan DA, Pinilla L, Gallego R, Saha AK, Kalsbeek A, Fliers E, Bisschop PH, Diéguez C, Nogueiras R, Rahmouni K, Tena-Sempere M, López M. Estradiol regulates brown adipose tissue thermogenesis via hypothalamic AMPK. *Cell Metab* 2014; **20**: 41-53 [PMID: 24856932 DOI: 10.1016/j.cmet.2014.03.031]
- 294 **Sievers W**, Rathner JA, Kettle C, Zacharias A, Irving HR, Green RA. The capacity for oestrogen to influence obesity through brown adipose tissue thermogenesis in animal models: A systematic review and meta-analysis. *Obes Sci Pract* 2019; **5**: 592-602 [PMID: 31890250 DOI: 10.1002/osp4.368]
- 295 **Wieland OH**. The mammalian pyruvate dehydrogenase complex: structure and regulation. *Rev Physiol Biochem Pharmacol* 1983; **96**: 123-170 [PMID: 6338572 DOI: 10.1007/BFb0031008]
- 296 **Connaughton S**, Chowdhury F, Attia RR, Song S, Zhang Y, Elam MB, Cook GA, Park EA. Regulation of pyruvate dehydrogenase kinase isoform 4 (PDK4) gene expression by glucocorticoids and insulin. *Mol Cell Endocrinol* 2010; **315**: 159-167 [PMID: 19703515 DOI: 10.1016/j.mce.2009.08.011]
- 297 **Wu P**, Sato J, Zhao Y, Jaskiewicz J, Popov KM, Harris RA. Starvation and diabetes increase the amount of pyruvate dehydrogenase kinase isoenzyme 4 in rat heart. *Biochem J* 1998; **329**: 197-201 [PMID: 9405294 DOI: 10.1042/bj3290197]
- 298 **Liang H**, Ward WF. PGC-1alpha: a key regulator of energy metabolism. *Adv Physiol Educ* 2006; **30**: 145-151 [PMID: 17108241 DOI: 10.1152/advan.00052.2006]
- 299 **Fernández-Marcos P**, Auwerx J. Regulation of PGC-1, a nodal regulator of mitochondrial biogenesis. *An J Clin Nutr* 2011; **93**: 884S-890S [DOI: 10.3945/ajcn.110.001917]
- 300 **Tcherepanova I**, Puigserver P, Norris JD, Spiegelman BM, McDonnell DP. Modulation of estrogen receptor-alpha transcriptional activity by the coactivator PGC-1. *J Biol Chem* 2000; **275**: 16302-16308 [PMID: 10748020 DOI: 10.1074/jbc.M001364200]
- 301 **Rhee J**, Inoue Y, Yoon JC, Puigserver P, Fan ML, Gonzalez FJ, Spiegelman BM. Regulation of hepatic fasting response by PPAR $\gamma$  coactivator-1 $\alpha$  (PGC-1): Requirement for hepatocyte nuclear factor 4 $\alpha$  in gluconeogenesis. *Proc Natl Acad Sci U S A* 2003; **100**: 4012-4017 [DOI: 10.1073/pnas.0730870100]
- 302 **Zhu LL**, Liu Y, Cui AF, Shao D, Liang JC, Liu XJ, Chen Y, Gupta N, Fang FD, Chang YS. PGC-1 $\alpha$  coactivates estrogen-related receptor- $\alpha$  to induce the expression of glucokinase. *Am J Physiol-cell Ph* 2010; **298**: E1210-E1218 [DOI: 10.1152/ajpendo.00633.2009]
- 303 **Zhang Y**, Ma K, Sadana P, Chowdhury F, Gaillard S, Wang F, McDonnell DP, Unterman TG, Elam MB, Park EA. Estrogen-related receptors stimulate pyruvate dehydrogenase kinase isoform 4 gene expression. *J Biol Chem* 2006; **281**: 39897-39906 [PMID: 17079227 DOI: 10.1074/jbc.M608657200]
- 304 **Heard DJ**, Norby PL, Holloway J, Vissing H. Human ERR $\gamma$ , a third member of the estrogen receptor-related receptor (ERR) subfamily of orphan nuclear receptors: tissue-specific isoforms are expressed during development and in the adult. *Mol Endocrinol* 2000; **14**: 382-392 [PMID: 10707956 DOI: 10.1210/mend.14.3.0431]
- 305 **Horard B**, Vanacker JM. Estrogen receptor-related receptors: orphan receptors desperately seeking a ligand. *J Mol Endocrinol* 2003; **31**: 349-357 [PMID: 14664699 DOI: 10.1677/jme.0.0310349]
- 306 **Hubbard WJ**, Bland KI, Chaudry IH. The ERRor of our ways: Estrogen-related receptors are about energy, not hormones, and are potential new targets for trauma and shock. *Shock* 2015; **44**: 3-15 [DOI: 10.1097/SHK.0000000000000364]
- 307 **Nie Y**, Wong C. Suppressing the activity of ERR  $\alpha$  in 3T3-L1 adipocytes reduces mitochondrial biogenesis but enhances glycolysis and basal glucose uptake. *J Cell Mol Med* 2009; **13**: 3051-3060 [PMID: 18544047 DOI: 10.1111/j.1582-4934.2008.00382.x]
- 308 **Wiik A**, Hellsten Y, Berthelson P, Lundholm L, Fischer H, Jansson E. Activation of estrogen response elements is mediated both via estrogen and muscle contractions in rat skeletal muscle

- myotubes. *Am J Physiol Cell Physiol* 2009; **296**: C215-C220 [PMID: 19020053 DOI: 10.1152/ajpcell.00148.2008]
- 309 **Wright LE**, Brandon AE, Hoy AJ, Forsberg GB, Lelliott CJ, Reznick J, Löfgren L, Oscarsson J, Strömstedt M, Cooney GJ, Turner N. Amelioration of lipid-induced insulin resistance in rat skeletal muscle by overexpression of Pgc-1 $\beta$  involves reductions in long-chain acyl-CoA levels and oxidative stress. *Diabetologia* 2011; **54**: 1417-1426 [PMID: 21331471 DOI: 10.1007/s00125-011-2068-x]
- 310 **Bryzgalova G**, Lundholm L, Portwood N, Gustafsson JA, Khan A, Efendic S, Dahlman-Wright K. Mechanisms of antidiabetogenic and body weight-lowering effects of estrogen in high-fat diet-fed mice. *Am J Physiol Endocrinol Metab* 2008; **295**: E904-E912 [PMID: 18697913 DOI: 10.1152/ajpendo.90248.2008]
- 311 **Kemper MF**, Stirone C, Krause DN, Duckles SP, Procaccio V. Genomic and non-genomic regulation of PGC1 isoforms by estrogen to increase cerebral vascular mitochondrial biogenesis and reactive oxygen species protection. *Eur J Pharmacol* 2014; **723**: 322-329 [PMID: 24275351 DOI: 10.1016/j.ejphar.2013.11.009]
- 312 **Gonzalez-Granillo M**, Savva C, Li X, Fitch M, Pedrelli M, Hellerstein M, Parini P, Korach-Andre M, Gustafsson J-A. ER $\beta$  activation in obesity improves whole body metabolism via adipose tissue function and enhanced mitochondria biogenesis. *Mol Cell Endocrinol* 2019; **479**: 147-158 [DOI: 10.1016/j.mce.2018.10.007]
- 313 **Esteve M**, Rafecas I, Remesar X, Alemany M. Nitrogen balances of lean and obese Zucker rats subjected to a cafeteria diet. *Int J Obes Relat Metab Disord* 1992; **16**: 237-244 [PMID: 1318277]
- 314 **Mauvais-Jarvis F**. Epidemiology of gender differences in diabetes and obesity. *Adv Exp Med Biol* 2017; **1043**: 3-8 [PMID: 29224087 DOI: 10.1007/978-3-319-70178-3\_1]
- 315 **Luciano AA**, Miller BE, Schoenenfeld MJ, Schaser RJ. Effects of estrone sulfate alone or with medroxyprogesterone acetate on serum lipoprotein levels in postmenopausal women. *Obstet Gynecol* 2001; **97**: 101-108 [DOI: 10.1016/S0029-7844(00)01081-4]
- 316 **Esteve M**, Savall P, Blay MT, Fernández-López JA, Remesar X, Alemany M. Intestinal handling of an oral oleoyl-estrone gavage by the rat. *Life Sci* 2001; **69**: 763-777 [PMID: 11487089 DOI: 10.1016/s0024-3205(01)01160-2]
- 317 **Romero MM**, Esteve M, Alemany M. Combined effects of oral oleoyl-estrone and limited food intake on body composition of young overweight male rats. *Int J Obes (Lond)* 2006; **30**: 1149-1156 [PMID: 16418752 DOI: 10.1038/sj.ijo.0803224]
- 318 **Romero MM**, Fernández-López JA, Alemany M, Esteve M. Gene expression modulation of liver energy metabolism by oleoyl-oestrone in overweight rats. *Biosci Rep* 2009; **30**: 81-89 [PMID: 19275765 DOI: 10.1042/BSR20080182]
- 319 **Adán C**, Cabot C, Vilà R, Grasa MM, Masanés RM, Esteve M, Estruch J, Fernández-López JA, Remesar X, Alemany M. Oleoyl-estrone treatment affects the ponderostat setting differently in lean and obese Zucker rats. *Int J Obes Relat Metab Disord* 1999; **23**: 366-373 [PMID: 10340814 DOI: 10.1038/sj.ijo.0800828]
- 320 **Blay M**, Peinado-Onsurbe J, Grasa MM, Díaz-Silva M, Fernandez-López JA, Remesar X, Alemany M. Effect of oral oleoyl-estrone treatment on plasma lipoproteins and tissue lipase activities of Zucker lean and obese female rats. *Int J Obes Relat Metab Disord* 2002; **26**: 618-626 [PMID: 12032744 DOI: 10.1038/sj.ijo.0801985]
- 321 **Romero MM**, Fernández-López JA, Esteve M, Alemany M. Oleoyl-oestrone inhibits lipogenic, but maintains thermogenic, gene expression of brown adipose tissue in overweight rats. *Biosci Rep* 2009; **29**: 237-243 [PMID: 18828761 DOI: 10.1042/BSR20080089]
- 322 **Díaz M**, Grasa MM, Fernández-López JA, Remesar X, Alemany M. Short-term effects of oleoyl-estrone on insulin sensitivity and glucose disposal in the rat. *Int J Obes Relat Metab Disord* 2002; **26**: S204
- 323 **Moos WH**, Dykens JA, Nohynek D, Rubinckik E, Howell N. Review of the effects of 17 $\alpha$  estradiol in humans: A less feminizing estrogen with neuroprotective potential. *Drug Develop Res* 2009; **70**: 1-21 [DOI: 10.1002/ddr.20284]
- 324 **Ferrer-Lorente R**, García-Peláez B, Fernández-López JA, Remesar X, Alemany M. Tamoxifen does not prevent the mobilization of body lipids elicited by oleoyl-estrone. *Steroids* 2004; **69**: 661-665 [DOI: 10.1016/j.steroids.2004.06.001]
- 325 **Mooradian AD**. Antioxidant properties of steroids. *J Steroid Biochem Mol Biol* 1993; **45**: 509-511 [PMID: 8518206 DOI: 10.1016/0960-0760(93)90166-t]
- 326 **Shwaery GT**, Vita JA, Keaney JF Jr. Antioxidant protection of LDL by physiologic concentrations of estrogens is specific for 17-beta-estradiol. *Atherosclerosis* 1998; **138**: 255-262 [PMID: 9690908 DOI: 10.1016/s0021-9150(98)00020-3]
- 327 **Badeau M**, Adlercreutz H, Kaihovaara P, Tikkkanen MJ. Estrogen A-ring structure and antioxidative effect on lipoproteins. *J Steroid Biochem Mol Biol* 2005; **96**: 271-278 [PMID: 15993048 DOI: 10.1016/j.jsbmb.2005.04.034]
- 328 **Larner JM**, Pahuja SL, Shackleton CH, McMurray WJ, Giordano G, Hochberg RB. The isolation and characterization of estradiol-fatty acid esters in human ovarian follicular fluid. Identification of an endogenous long-lived and potent family of estrogens. *J Biol Chem* 1993; **268**: 13893-13899 [PMID: 8314757]
- 329 **Carlson SE**, Carver JD, House SG. High fat diets varying in ratios of polyunsaturated to saturated fatty acid and linoleic to linolenic acid: a comparison of rat neural and red cell membrane

- phospholipids. *J Nutr* 1986; **116**: 718-725 [PMID: 2871142 DOI: 10.1093/jn/116.5.718]
- 330 **Raleigh JA**, Kremers W, Gaboury B. Dose-rate and oxygen effects in models of lipid membranes: linoleic acid. *Int J Radiat Biol Relat Stud Phys Chem Med* 1977; **31**: 203-213 [PMID: 300724 DOI: 10.1080/09553007714550251]
- 331 **Kadoma Y**, Fujisawa S. Radical-scavenging activity of estrogen and estrogen-like compounds using the induction period method. *Int J Mol Sci* 2007; **8**: 295-303
- 332 **Klinge CM**. Estrogens regulate life and death in mitochondria. *J Bioenerg Biomembr* 2017; **49**: 307-324 [PMID: 28401437 DOI: 10.1007/s10863-017-9704-1]
- 333 **Sawada H**, Shimohama S. Neuroprotective effects of estradiol in mesencephalic dopaminergic neurons. *Neurosci Biobehav Rev* 2000; **24**: 143-147 [PMID: 10654671 DOI: 10.1016/s0149-7634(99)00059-7]
- 334 **Moosmann B**, Behl C. The antioxidant neuroprotective effects of estrogens and phenolic compounds are independent from their estrogenic properties. *Proc Natl Acad Sci U S A* 1999; **96**: 8867-8872 [PMID: 10430862 DOI: 10.1073/pnas.96.16.8867]
- 335 **Höckerstedt A**, Jauhainen M, Tikkalanen MJ. Lecithin/cholesterol acyltransferase induces estradiol esterification in high-density lipoprotein, increasing its antioxidant potential. *J Clin Endocrinol Metab* 2004; **89**: 5088-5093 [PMID: 15472210 DOI: 10.1210/jc.2004-0141]
- 336 **Hutson DD**, Gurrala R, Ogola BO, Zimmerman MA, Mostany R, Satou R, Lindsey SH. Estrogen receptor profiles across tissues from male and female *Rattus norvegicus*. *Biol Sex Differ* 2019; **10**: 4 [PMID: 30635056 DOI: 10.1186/s13293-019-0219-9]
- 337 **Dieudonné MN**, Leneveu MC, Giudicelli Y, Pecquery R. Evidence for functional estrogen receptors alpha and beta in human adipose cells: regional specificities and regulation by estrogens. *Am J Physiol Cell Physiol* 2004; **286**: C655-C661 [PMID: 14761887 DOI: 10.1152/ajpcell.00321.2003]
- 338 **Bryzgalova G**, Gao H, Ahren B, Zierath JR, Galuska D, Steiler TL, hlman-Wright K, Nilsson S, Gustafsson JÅ, Efendic S, Khan A. Evidence that oestrogen receptor- $\alpha$  plays an important role in the regulation of glucose homeostasis in mice: insulin sensitivity in the liver. *Diabetologia* 2006; **49**: 588-597 [DOI: 10.1007/s00125-005-0105-3]
- 339 **Ikeda K**, Horie-Inoue K, Inoue S. Functions of estrogen and estrogen receptor signaling on skeletal muscle. *J Steroid Biochem Mol Biol* 2019; **191**: 105375 [PMID: 31067490 DOI: 10.1016/j.jsbmb.2019.105375]
- 340 **Morissette M**, Le Saux M, d'Astous M, Jourdain S, Al Sweidi S, Morin N, Estrada-Camarena E, Méndez P, García-Segura LM, di Paolo T. Contribution of estrogen receptors alpha and beta to the effects of estradiol in the brain. *J Steroid Biochem Mol Biol* 2008; **108**: 327-338 [PMID: 17936613 DOI: 10.1016/j.jsbmb.2007.09.011]
- 341 **Ali MH**, Napit PR, Mahmood ASMH, Bheemanapally K, Alhamami HN, Uddin MM, Mandal SK, Ibrahim MMH, Briski KP. Hindbrain estrogen receptor regulation of ventromedial hypothalamic glycogen metabolism and glucoregulatory transmitter expression in the hypoglycemic male rat. *Neuroscience* 2019; **409**: 253-260 [PMID: 30954669 DOI: 10.1016/j.neuroscience.2019.03.053]
- 342 **Boldarine VT**, Pedroso AP, Brandão-Teles C, loTurco EG, Nascimento CMO, Oyama LM, Bueno AA, Martins-de-Souza D, Ribeiro EB. Ovariectomy modifies lipid metabolism of retroperitoneal white fat in rats: a proteomic approach. *Am J Physiol Endocrinol Metab* 2020; **319**: E427-E437 [PMID: 32663100 DOI: 10.1152/ajpendo.00094.2020]
- 343 **Brown LM**, Clegg DJ. Central effects of estradiol in the regulation of food intake, body weight, and adiposity. *J Steroid Biochem Mol Biol* 2010; **122**: 65-73 [PMID: 20035866 DOI: 10.1016/j.jsbmb.2009.12.005]
- 344 **Liu J**, Bisschop PH, Eggels L, Foppens E, Ackermans MT, Zhou JN, Fliers E, Kalsbeek A. Intrahypothalamic estradiol regulates glucose metabolism via the sympathetic nervous system in female rats. *Diabetes* 2013; **62**: 435-443 [PMID: 23139356 DOI: 10.2337/db12-0488]
- 345 **Shen L**, Wang DQH, Xu M, Woods SC, Liu M. BDNF/TrkB signaling mediates the anorectic action of estradiol in the nucleus tractus solitarius. *Oncotarget* 2017; **8**: 84028-84038 [PMID: 29137402 DOI: 10.18632/oncotarget.21062]
- 346 **Kallen CB**. Estrogen targets fat mass and glucose metabolism by acting in the brain. *Am J Physiol Endocrinol Metab* 2012; **303**: E443-E444 [PMID: 22669245 DOI: 10.1152/ajpendo.00277.2012]
- 347 **Comitato R**, Saba A, Turrini A, Arganini C, Virgili F. Sex hormones and macronutrient metabolism. *Crit Rev Food Sci Nutr* 2015; **55**: 227-241 [PMID: 24915409 DOI: 10.1080/10408398.2011.651177]
- 348 **López M**, Tena-Sempere M. Estrogens and the control of energy homeostasis: a brain perspective. *Trends Endocrinol Metab* 2015; **26**: 411-421 [PMID: 26126705 DOI: 10.1016/j.tem.2015.06.003]
- 349 **Rettberg JR**, Yao J, Brinton RD. Estrogen: a master regulator of bioenergetic systems in the brain and body. *Front Neuroendocrinol* 2014; **35**: 8-30 [PMID: 23994581 DOI: 10.1016/j.yfrne.2013.08.001]
- 350 **van Pelt RE**, Gavin KM, Kohrt WM. Regulation of body composition and bioenergetics by estrogens. *Endocrinol Metab Clin North Am* 2015; **44**: 663-676 [PMID: 26316249 DOI: 10.1016/j.ecl.2015.05.011]
- 351 **Adamski J**, Jakob FJ. A guide to 17 $\beta$ -hydroxysteroid dehydrogenases. *Mol Cell Endocrinol* 2001; **171**: 1-4 [DOI: 10.1016/S0303-7207(00)00383-X]
- 352 **Bonetti A**, Tirelli F, Catapano A, d'Azzi D, dei Cas A, Solito F, Ceda G, Reverberi C, Monica C, Pipitone S, Elia G, Spattini M, Magnati G. Side effects of anabolic androgenic steroids abuse. *Int J Sports Med* 2008; **29**: 679-687 [PMID: 18004690 DOI: 10.1055/s-2007-965808]

- 353 **Hartgens F**, Kuipers H. Effects of androgenic-anabolic steroids in athletes. *Sports Med* 2004; **34**: 513-554 [PMID: 15248788 DOI: 10.2165/00007256-200434080-00003]
- 354 **Wilson C**, Contreras-Ferrat A, Venegas N, Osorio-Fuentealba C, Pávez M, Montoya K, Durán J, Maass R, Lavandero S, Estrada M. Testosterone increases GLUT4-dependent glucose uptake in cardiomyocytes. *J Cell Physiol* 2013; **228**: 2399-2407 [PMID: 23757167 DOI: 10.1002/jcp.24413]
- 355 **Kato Y**, Shigehara K, Nakashima K, Iijima M, Kawagushi S, Nohara T, Izumi K, Kadono Y, Konaka H, Namiki M, Mizokami A. The five-year effects of testosterone replacement therapy on lipid profile and glucose tolerance among hypogonadal men in Japan: a case control study. *Aging Male* 2020; **23**: 23-28 [PMID: 30651019 DOI: 10.1080/13685538.2018.1550060]
- 356 **Quang LM**, Kalhan A. Cardiovascular benefits and risks of testosterone replacement therapy in hypogonadal men with type 2 diabetes mellitus and/or the metabolic syndrome: a systematic review. *Brit J Nutr* 2018; **18**: 141-146 [DOI: 10.15277/bjdn.2018.192]
- 357 **Wise P**. Clearing estrogen's bad name. *The Scientist* 2008; **22**: 40-44
- 358 **Jayachandran M**, Lahr BD, Bailey KR, Miller VM, Kantarci K. Menopausal hormone therapy, blood thrombogenicity, and development of white matter hyperintensities in women of the Kronos Early Estrogen Prevention Study. *Menopause* 2020; **27**: 305-310 [PMID: 31934946 DOI: 10.1097/GME.0000000000001465]
- 359 **Styer AK**. The impact of estrogen alone hormone therapy on breast cancer risk and health outcomes: reassurance for the treatment of climacteric symptoms in black women? *Menopause* 2017; **24**: 124-125 [PMID: 28072609 DOI: 10.1097/GME.0000000000000821]
- 360 **Miller EM**. Hormone replacement therapy affects iron status more than endometrial bleeding in older US women: A role for estrogen in iron homeostasis? *Maturitas* 2016; **88**: 46-51 [PMID: 27105697 DOI: 10.1016/j.maturitas.2016.03.014]
- 361 **Mauvais-Jarvis F**. Is estradiol a biomarker of type 2 diabetes risk in postmenopausal women? *Diabetes* 2017; **66**: 568-570 [PMID: 28223340 DOI: 10.2337/db16-0063]
- 362 **Vermeulen A**, Kaufman JM. Ageing of the hypothalamo-pituitary-testicular axis in men. *Horm Res* 1995; **43**: 25-28 [PMID: 7721258 DOI: 10.1159/000184233]
- 363 **Rahnema CD**, Lipshultz LI, Crosnoe LE, Kovac JR, Kim ED. Anabolic steroid-induced hypogonadism: diagnosis and treatment. *Fertil Steril* 2014; **101**: 1271-1279 [PMID: 24636400 DOI: 10.1016/j.fertnstert.2014.02.002]
- 364 **Gangestad SW**, Thornhill R. Human oestrus. *Proc Biol Sci* 2008; **275**: 991-1000 [PMID: 18252670 DOI: 10.1098/rspb.2007.1425]
- 365 **Anantharaman-Barr HG**, Decombez J. The effect of wheel running and the estrous cycle on energy expenditure in female rats. *Physiol Behav* 1989; **46**: 259-263 [PMID: 2602468 DOI: 10.1016/0031-9384(89)90265-5]
- 366 **Smith RJ**. Allometric scaling in comparative biology: problems of concept and method. *Am J Physiol* 1984; **246**: R152-R160 [PMID: 6696141 DOI: 10.1152/ajpregu.1984.246.2.R152]
- 367 **Calder WA 3rd**. Body size, mortality, and longevity. *J Theor Biol* 1983; **102**: 135-144 [PMID: 6876838 DOI: 10.1016/0022-5193(83)90266-7]
- 368 **Shutt DA**. The effects of plant oestrogens on animal reproduction. *Endeavour* 1976; **35**: 110-113 [PMID: 62660 DOI: 10.1016/0160-9327(76)90004-1]
- 369 **Zang K**, Kurisu F, Kasuga I, Furumai H, Yagi O. Analysis of the phylogenetic diversity of estrone-degrading bacteria in activated sewage sludge using microautoradiography-fluorescence *in situ* hybridization. *Syst Appl Microbiol* 2008; **31**: 206-214 [PMID: 18513907 DOI: 10.1016/j.syapm.2008.03.005]
- 370 **Sadílek J**, Spálovská P, Vrana B, Vávrová M, Maršílek B, Šimek Z. Comparison of extraction techniques for isolation of steroid oestrogens in environmentally relevant concentrations from sediment. *Int J Environ Anal Chem* 2016; **96**: 1022-1037 [DOI: 10.1080/03067319.2016.1232718]
- 371 **Peck M**, Gibson RW, Kortenkamp A, Hill EM. Sediments are major sinks of steroidal estrogens in two United Kingdom rivers. *Environ Toxicol Chem* 2004; **23**: 945-952 [PMID: 15095890 DOI: 10.1897/03-41]
- 372 **Braga O**, Smythe GA, Schäfer AI, Feitz AJ. Steroid estrogens in ocean sediments. *Chemosphere* 2005; **61**: 827-833 [PMID: 15967481 DOI: 10.1016/j.chemosphere.2005.04.053]
- 373 **Servos MR**, Bennie DT, Burnison BK, Jurkovic A, McInnis R, Neheli T, Schnell A, Seto P, Smyth SA, Ternes TA. Distribution of estrogens, 17beta-estradiol and estrone, in Canadian municipal wastewater treatment plants. *Sci Total Environ* 2005; **336**: 155-170 [PMID: 15589256 DOI: 10.1016/j.scitotenv.2004.05.025]
- 374 **Encarnação T**, Pais AA, Campos MG, Burrows HD. Endocrine disrupting chemicals: Impact on human health, wildlife and the environment. *Sci Prog* 2019; **102**: 3-42 [PMID: 31829784 DOI: 10.1177/0036850419826802]
- 375 **Kuhl H**. Pharmacology of estrogens and progestogens: influence of different routes of administration. *Climacteric* 2005; **8** Suppl 1: 3-63 [PMID: 16112947 DOI: 10.1080/13697130500148875]
- 376 **van den Heuvel MW**, van Bragt AJ, Alnabawy AK, Kaptein MC. Comparison of ethinylestradiol pharmacokinetics in three hormonal contraceptive formulations: the vaginal ring, the transdermal patch and an oral contraceptive. *Contraception* 2005; **72**: 168-174 [PMID: 16102549 DOI: 10.1016/j.contraception.2005.03.005]
- 377 **RR**. Effects of developmental exposure to diethylstilbestrol (DES) in rodents: clues for other environmental estrogens. *APMIS* 2001; **109**: S261-S271 [DOI:

- 10.1111/j.1600-0463.2001.tb05775.x]
- 378 **Luine VN.** Estradiol and cognitive function: past, present and future. *Horm Behav* 2014; **66**: 602-618 [PMID: 25205317 DOI: 10.1016/j.ybeh.2014.08.011]
- 379 **McEwen BS.** Invited review: Estrogens effects on the brain: multiple sites and molecular mechanisms. *J Appl Physiol* (1985) 2001; **91**: 2785-2801 [PMID: 11717247 DOI: 10.1152/jappl.2001.91.6.2785]
- 380 **Cutolo M.** Capellino S, Sulli A, Serioli B, Secchi ME, Villaggio B, Straub RH. Estrogens and autoimmune diseases. *Ann N Y Acad Sci* 2006; **1089**: 538-547 [PMID: 17261796 DOI: 10.1196/annals.1386.043]
- 381 **Khan D.** Ansar Ahmed S. The immune system is a natural target for estrogen action: Opposing effects of estrogen in two prototypical autoimmune diseases. *Front Immunol* 2015; **6**: 635 [PMID: 26779182 DOI: 10.3389/fimmu.2015.00635]
- 382 **Trémolieres F.** Contraception orale estro-progestative: quelle différence entre éthinylestradiol et estradiol? *Gynécologie Obstétrique et Fertilité* 2012; **40**: 109-115 [DOI: 10.1016/j.gyobfe.2011.10.009]
- 383 **Coelingh Bennink HJ.** Are all estrogens the same? *Maturitas* 2004; **47**: 269-275 [PMID: 15063479 DOI: 10.1016/j.maturitas.2003.11.009]
- 384 **Shoham Z,** Kopernik G. Tools for making correct decisions regarding hormone therapy. Part I: Background and drugs. *Fertil Steril* 2004; **81**: 1447-1457 [PMID: 15193460 DOI: 10.1016/j.fertnstert.2003.10.052]
- 385 **Patel AS,** Leong JY, Ramos L, Ramasamy R. Testosterone is a contraceptive and should not be used in men who desire fertility. *World J Mens Health* 2019; **37**: 45-54 [PMID: 30350483 DOI: 10.5534/wjmh.180036]
- 386 **Liu JD,** Wu YQ. Anabolic-androgenic steroids and cardiovascular risk. *Chin Med J (Engl)* 2019; **132**: 2229-2236 [PMID: 31478927 DOI: 10.1097/CM9.0000000000000407]
- 387 **O'Connell K,** Davis AR, Kerns J. Oral contraceptives: side effects and depression in adolescent girls. *Contraception* 2007; **75**: 299-304 [PMID: 17362710 DOI: 10.1016/j.contraception.2006.09.008]
- 388 **Skovlund CW,** Mørch LS, Kessing LV, Lidegaard Ø. Association of hormonal contraception with depression. *JAMA Psychiatry* 2016; **73**: 1154-1162 [PMID: 27680324 DOI: 10.1001/jamapsychiatry.2016.2387]



Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

**Telephone:** +1-925-3991568

**E-mail:** [bpgoftice@wjnet.com](mailto:bpgoftice@wjnet.com)

**Help Desk:** <https://www.f6publishing.com/helpdesk>

<https://www.wjnet.com>

