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*WJGP* mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal pathophysiology and covering a wide range of topics including disorders of the esophagus, stomach and duodenum, small intestines, pancreas, biliary system, and liver.

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## Electrical neuromodulation therapy for inflammatory bowel disease

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### Abstract

Inflammatory bowel disease (IBD) is an inflammatory disease of the gastrointestinal (GI) tract. It has financial and quality of life impact on patients. Although there has been a significant advancement in treatments, a considerable number of patients do not respond to it or have severe side effects. Therapeutic approaches such as electrical neuromodulation are being investigated to provide alternate options. Although bioelectric neuromodulation technology has evolved significantly in the last decade, sacral nerve stimulation (SNS) for fecal incontinence remains the only neuromodulation protocol commonly utilized use for GI disease. For IBD treatment, several electrical neuromodulation techniques have been studied, such as vagus NS, SNS, and tibial NS. Several animal and clinical experiments were conducted to study the effectiveness, with encouraging results. The precise underlying mechanisms of action for electrical neuromodulation are unclear, but this modality appears to be promising. Randomized control trials are required to investigate the efficacy of intrinsic processes. In this review, we will discuss the electrical modulation therapy for the IBD and the data pertaining to it.

**Key Words:** Inflammatory bowel disease; Sacral nerve stimulation; Vagus nerve stimulation; Tibial nerve stimulation; Electrical neuromodulation; Crohn's disease;

Ulcerative colitis; Neuromodulation

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**Core Tip:** Inflammatory bowel disease (IBD) is an inflammatory disease of the gastrointestinal tract with no known available treatment. Electrical neuromodulation is the use of electric stimulation of nerves or brain regions as a therapeutic technique. Electrical neuromodulation therapy has been studied as a possible treatment regimen for IBD. There are several forms of neuromodulation that use various types of nerves, such as sacral nerve stimulation, vagal NS (VNS), and tibial NS. As indicated by many clinical investigations, VNS as a potential therapy for IBD has a lot of promise. More research is needed to assess the possibility of VNS as a viable cure for IBD.

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## INTRODUCTION

Inflammatory bowel disease (IBD) comprises ulcerative colitis (UC) and Crohn's disease (CD). In these conditions, neutrophils and macrophages produce cytokines, proteolytic enzymes, and free radicals, leading to inflammation and ulceration of the intestinal lining. Both UC and CD share similar manifestations, including abdominal pain, diarrhea, weight loss, and hematochezia. Malnutrition, anemia, fatigue, fever, mouth ulcers, joint pain, and skin lesions, including erythema nodosum or pyoderma gangrenosum, are the common findings[1].

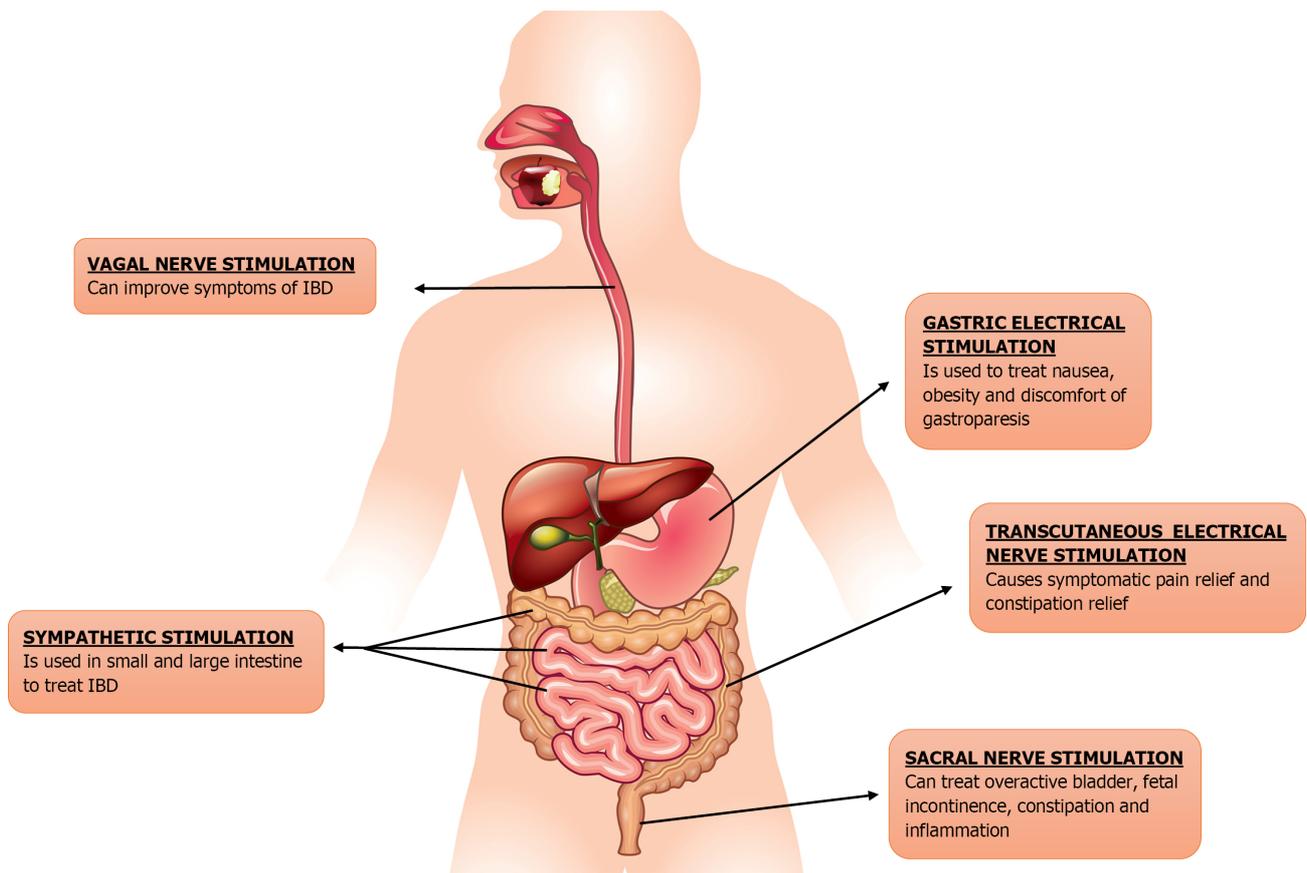
The exact etiology of IBD is unknown, but the altered immune system is suggested as a possible explanation. Risk factors include race, family history, ethnicity, cigarette smoking, and non-steroidal drugs. Colon cancer, skin infection, eye and joint infection, pharmaceutical side effects, and blood clots are all common complications of CD and UC[2].

Diagnostic procedures for IBD include blood work (for anemia and infection), endoscopic procedures (colonoscopy, flexible sigmoidoscopy, upper endoscopy, capsule endoscopy, and balloon aided enteroscopy), and imaging treatments (X-ray, computerized tomography scan, magnetic resonance imaging)[2].

The common medical treatment consists of antibiotics, corticosteroids, immune regulators, aminosalicylates, Janus kinase inhibitor (JAK), anti-tumor necrosis factor-alpha (Anti-TNF- $\alpha$ ), anti-integrin, and anti-interleukin (IL) 12/IL23. Adverse reactions include itching, erythema, and delayed allergic reactions can be seen in patients due to these medication use[3]. Therefore, more effective, and safer therapeutic choices are needed. Nerves or brain structures electrical stimulation is being studied as an intervention in a growing number of conditions, including Parkinson's disease, arthritis, and depressive disorders. The idea that bioelectrical neuromodulation can be used to treat gastrointestinal (GI) disorders has piqued the interest of the medical community[4].

## ELECTROMODULATION THERAPY FOR IBDS

The usage of electric stimulation of nerves or brain centers as a therapeutic tool is being tested in a wide variety of conditions as Parkinson's disease and schizophrenia. This approach is called neuromodulation or bioelectric neuromodulation, or electroceuticals[5]. GI tract is connected to the central nervous system *via* vagus and sacral nerve, providing disease-modifying bioelectric neuromodulation therapy opportunities[4]. Electrical neuromodulation (ENM) has been used effectively to treat variety of gastrointestinal disorders including GERD, dyspepsia, gastroparesis, fecal incontinence and constipation as shown in **Figure 1**. Neuromodulation may be central, as in thalamic stimulation or trans-magnetic stimulation; spinally, as in spinal cord stimulation for ache and movement in spinal cord damage; vagal as regional, as in auricular stimulation for seizures; sacral, as in stimulation for genitourinary (GU)/GI dysfunction; and peripherally, as in electrified stimulation for GU/GI dysfunction peripherally, as in electroacupuncture; and enteric, as in gastric/GI electrical stimulation (GES)[6]. Sacral nerve stimulation (SNS) is the most effective neuromodulation protocol for GI disease that is currently in use[7]. Because of the dysregulation of brain-gut interactions in IBD, ENM can be



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**Figure 1 Sites of bioelectric neuromodulation to improve gastro-intestinal symptoms.** In animal research and experimental clinical settings, neuromodulation has been used to treat a ramification of gastrointestinal (GI) illnesses at numerous sites on neurons innervating the gastrointestinal tract. Some of the neuromodulation techniques such as transcutaneous electrical nerve stimulation, sympathetic stimulation, vagal nerve stimulation, and gastric electrical stimulation are mentioned in the figure above that relieve the symptoms related to inflammatory bowel disease. IBD: Inflammatory bowel disease.

considered as a treatment option[8]. Numerous electrical neuromodulation techniques for treating IBD, *i.e.*, we will be discussing vagus NS (VNS), SNS, and tibial NS (TNS), in this review.

## INTERPLAY BETWEEN BRAIN-GUT AXIS/EXTRINSIC GI INNERVATION

The GI tract (GIT) has intrinsic (enteric nervous system) as well as extrinsic innervation (gut-brain axis). The gut-brain axis is bidirectional in nature, mediated through hormonal, neural, metabolic, and immunological responses. It carries different sensations such as GIT pressure changes, ischemia, poisons, bacterial infection, gastric acidity, and inflammation of the brain through afferent fibers, as demonstrated in Figure 2[9]. These fibers then carry information to the brain, which sends efferent signals to the gut and associated organs, causing toxic substances to be removed, decreasing acid production, increasing satiety, and nausea, to name a few. Recently, the gut microbiota is also included in the gut-brain axis[10], which links intestinal microbiota and the brain[11].

Accurate extrinsic innervation is crucial for the proper functioning of the gut as well as for the balanced emotional and psychological responses through dual connections between brain and gut[12]. Various researches have listed the effects of the brain on the gut or vice versa, signaling, *e.g.*, how depression and impaired brain functioning can increase an individual's vulnerability to IBD. Whereas other experimentations have shown the prevalence of psychic and anxiety-related disorders in IBD patients, these researches show a close interplay between the gut and the brain[8].

The complex pathway connects the central nervous system (CNS), sympathetic ganglia, enteric nervous system, and gastrointestinal effector tissues. The nucleus tractus solitarius receives the communications *via* the vagal afferents, while the thoracolumbar spinal cord receives the input *via* the spinal afferents. Cervical afferents also link the esophagus to the cervical spinal cord. Intestinal-fugal neurons that amplify from the intestine to the CNS are involved in certain afferent routes. To accurately understand the specifics of the extrinsic innervation in the form of the gut-brain axis, the various pathways through which the dual interaction between the gut and the brain takes place are described in Figure 2.

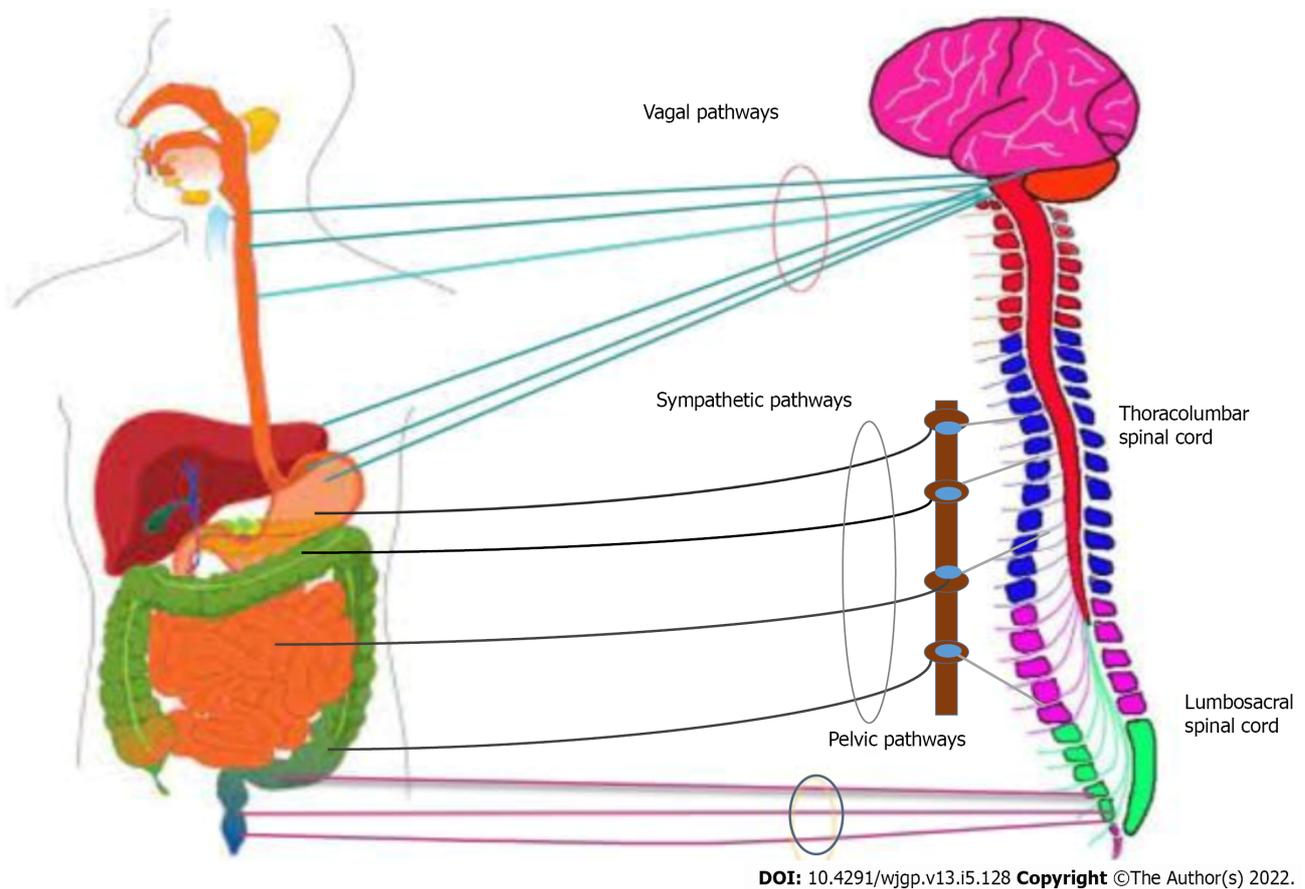


Figure 2 Gastrointestinal tract's extrinsic innervation.

## NEURAL CONTROL OF GUT INFLAMMATION

### *Influence of vagal pathway on gut inflammation*

Preganglionic neurons of cranial nerve corticofugal fibers protrude from the medulla' dorsal motor nucleus of the vagus nerve and innervate the muscular and tissue layer layers of the gut, each within the lamina propria and also the muscularis externa of the viscus wall[13]. Food, antigens, potential pathogens, and symbiotic intestinal microbiota are always present in the gastrointestinal system, and some of them may present as risk factors for intestinal inflammation[14]. TNF- $\alpha$ , a cytokine, is released by activated macrophages, nerve fiber cells, and different tissue layer cells in response to the infective toxin and other harmful stimuli to cause inflammation[15,16]. Counter-regulatory mechanisms consist of capable immune cells and anti-inflammatory cytokines that inhibit inflammatory mediators' transfer into the circulation. As an anti-inflammatory mechanism, there is a fine relationship between neurological and immune system processes. The dorsal vagus complicated (DVC), which has the sensory nuclei of the solitary tract (NTS), the area postrema (AP), and also the dorsal motor core of the cranial nerve (DMN), responds to higher current levels of TNF- $\alpha$  by increasing motor levels activity within the vagus nerve[17]. Two studies have shown that electrical cranial nerve stimulation will suppress inflammation in models of inflammation[18,19]. Furthermore, due to the lack of control on immunological mediating cells, the sub-diaphragmatic vagotomy increases inflammation in the gut. The brain can monitor immunological states and detect peripheral inflammation through two mechanisms.

### *Neural pathway*

Stimulation of the vagus nerve is triggered directly or indirectly by cytokines discharged by nerve fiber cells, macrophages, and different vagus-associated immune cells and indirectly by chemoreceptors[20]. Visceral afferent vagus fibers within the neural structure nodosum principally end in the DVC of the medulla oblongata. DVC includes NTS, the dorsal motor nucleus of the vagus (DMV), and also the post-mammillary region (AP)[21]. DMN is a critical region for the formation of preganglionic vagus efferent fibers. The majority of sensory vagal input is received by the NTS[22]. The paraventricular nucleus (PVN) of hypothalamus, receives signals from the NTS. PVN causes the production and release of corticotropin-releasing hormones (CRH), which is an important chemical on the hypothalamus-pituitary-adrenocortical (HPA) axis (described below)[23].

**Humoral pathway**

Circulating cytokines in the humoral route interact directly with areas of the brain involved in anti-inflammatory response. Circulatory IL-1 and TNF can move across the blood-brain barrier through a saturated transport mechanism to get into the CSF and interstitial space of the brain and spinal cord, where they can directly stimulate the brain to produce an anti-inflammatory reaction[24]. Circumventricular organs that lack regular blood-brain barrier protection uses cytokine-to-brain transmission. Postrema is the most well-known circumventricular organ[23].

Followings are a few pathways that are included in the neural control of gut inflammation.

**HPA axis pathway**

The HPA axis is composed of three major components (the hypothalamus, the anterior and posterior pituitary gland, and the adrenal cortex). Steinlein[25] demonstrated the role of vagal afferents in the neuro-immune axis in the control of the HPA axis. According to L E Goehler and co-workers, peripheral administration of lipopolysaccharides (LPS), a pro-inflammatory cytokine that stimulates vagal afferents *via* IL-1 receptors, induces the production of IL-1, a pro-inflammatory cytokine[26]. The vagal nerve is susceptible to peripheral pro-inflammatory cytokines generated by macrophages and other immune cells, such as IL-1, IL-6, and TNF- $\alpha$ [27]. Vagal afferent receptors (IL-1 beta) convey information to the parvo-cellular zone of the paraventricular nucleus of the hypothalamus (PVH) around corticotrophin-releasing-factor (CRF)-containing neurons. These CRF neurons subsequently drive the hypophysis to release the adreno-corticotrophin hormone, which stimulates the adrenal glands to release glucocorticoids, reducing peripheral inflammation[27]. Glucocorticoids affect the inflammatory response by suppressing immune cell release of pro-inflammatory cytokines, as well as inhibiting vasodilation and vascular permeability caused by inflammation. The brain can influence the activity of functional intestinal effector cells such as immune cells, smooth muscle cells, epithelial cells, interstitial cells of Cajal, enteric neurons, and enterochromaffin cells through neuronal and hormonal communication lines [24]. These cells, on the other hand, are influenced by the gut microbiome. The internal organ microbiota encompasses a vital influence on the intestinal axis, not solely through native interaction with intestinal cells conjointly with the enteric systema nervosum, but also through direct effects on the system and metabolic processes[28]. Emerging evidence supports the function of gut bacteria in anxiety and depressive-like behavior[29].

**Cholinergic anti-inflammatory pathway (ach axis)**

Acetylcholine is a crucial neurochemical and neuromodulator within the brain, mediates neuronal transmission in sympathetic and parasympathetic neurons, and acts as a primary neurotransmitter in parasympathetic/pneumogastric neural structure corticoefferent neurons[23]. This neurotransmitter acts through two varieties of receptors: muscarinic (metabotropic) and nicotinic (ionotropic)[30,31]. The seven component of the nicotinic acetylcholine receptor is displayed on phagocytes[32]. TNF production from human macrophages used by endotoxins is considerably reduced by acetylcholine through a post-transcriptional mechanism and is concentration-dependent. The authors demonstrated the connection of a bungarotoxin-insensitive vasoconstrictor receptor in suppressing cytokine production *in vitro* by neurotransmitter mistreatment specific muscarinic and nicotinic agonists and antagonists[23]. Apart from TNF, acetylcholine suppresses alternative endotoxin-inducible pro-inflammatory proteins reminiscent of IL-1, IL-6, and IL-18 through a post-transcriptional mechanism. However, acetylcholine has no effect on the discharge of the anti-inflammatory cytokine IL-10 from endotoxin-stimulated macrophages[32]. Nicotinic acetylcholine receptors are a ligand-gated pentameric ion channel family. The HPA axis (afferent vagal Fibers) activates the cholinergic anti-inflammatory pathway. Proinflammatory cytokines discharged throughout the immunologic response will activate vagal receptive signals, resulting in direct or indirect activation (via the core of the neurons of the solitary tract NTS) of the vagal efferents in the DMN. As a result, the sensory vagal afferents and motor vagus efferents produce an inflammatory reflex that constantly monitors and modifies the inflammatory condition in the periphery[33]. Since the tetravalent guanyl hydrazone CNI1493 induces the activation of the vagus and, by activating the cholinergic anti-inflammatory signal pathway, confers anti-inflammatory effects in each native and general model of inflammation, it's going to be attainable to activate the cholinergic anti-inflammatory pathway (with centrally active substances)[34].

**Vagal sympathetic pathway**

The celiac, superior mesenteric, and inferior mesenteric ganglia contain the cell bodies of the bulk of postganglionic sympathetic neurons that innervate the gastrointestinal tract[35]. In gut noradrenaline (NA) is the primary neurotransmitter released from sympathetic postganglionic nerve terminals; however, ATP and neuropeptide Y (NPY) can also engage in sympathetic neurotransmission within the GI tract[36,37]. The vagal afferent Fibers terminate in the NTS, which ultimately activates the central autonomic network (CAN). The sympathetic outlet is operated by 5 CAN brain regions (the paraventricular nucleus of the neural structure HPV, the noradrenergic cluster A5, the area of the caudal raphe, the rostral ventrolateral medulla, and ventromedial medulla)[38]. By increasing sympathetic outflow, the vagal nerve can generate a non-direct anti-inflammatory reaction. Abe *et al*[39] explained

the role of the C1 adrenergic cluster. They concluded that these neurons are concerned with protecting the result of stress in reperfusion injuries because of nephritic anemia *via* a sympathetic pathway. They conjointly mentioned how activation of vagal afferents in mice twenty-four hours before injury considerably reduced acute excretory organ inflammation and plasma levels of TNF- $\alpha$ [30]. Tyrosine hydroxylase is found in the lamina propria, the submucosa, the ganglia of the nerve plexus, and lymph follicles (Peyer' plaques)[40]. Adrenergic receptors of diverse types are expressed by macrophages. In vitro, beta receptors mediate the anti-inflammatory effects of agonists on macrophages derived from the intestine[41]. Although sympathetic nerves decrease gut inflammation, persistent nerve stimulation should be avoided as it can promote stasis and aggravate bacterial growth in Crohn's illness[4].

### **Vagal splenic pathway**

Through an association between the VN and the splenic nerve, the Vago-splenic pathway works collectively[42]. In general inflammatory conditions, the spleen is a crucial supply of inflammatory cytokines, and excision considerably reduces circulating TNF $\alpha$  levels in mouse endotoxemia[43]. Tracey *et al* identified the vagal splenic route, finding that VNS caused the celiac ganglion to produce acetylcholine (ACh), which subsequently adhered to the c7nAChR of the splenic nerve to release norepinephrine (NE) in the spleen[24]. Following that, it binds to beta two adrenergic receptors of splenic lymphocytes, which produces acetylcholine, which will act on the  $\alpha$  c7nAChR of splenic macrophages limiting release of TNF, resulting in an anti-inflammatory impact[44]. According to Martelli *et al*[45], there is also a non-nervous relationship between the vagus and splenic sympathetic nerves. In another article, Martelli *et al*[46] noted how the sympathetic nerve, not the vagal nerve, is the efferent mediator of the cholinergic anti-inflammatory pathway (splenic nerve).

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## **CAPSAICIN-SENSITIVE AFFERENTS AND INFLAMMATION REGULATION**

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Electrical and physiological stimulation of receptive neurons, particularly afferent nerves of the digestive tract, generates the release of transmitters at their peripheral ends, most often tachykinin and the amide (CGRP) linked to the calcitonin gene[47]. CGRP serves a number of purposes *via* serving as a modulator, transmitter, and hormone. CGRP-containing nerve Fibers are numerous surrounding blood vessels, particularly arterioles, suggesting that they may have a physiologic role in regulating blood flow to the gastric mucosa[48]. Capsaicin-sensitive afferent Fibers conduct protective anti-inflammatory activities in the gastrointestinal tract by releasing peptides from their peripheral ends[49-52]. Sensory inputs innervating the stomach generate CGRP, which reduces mucosal damage and improves mesenteric and mucosal blood flow in stomachic ulcer models in rats and mice[50,51]. Once administered at the time of injury, capsaicin promotes the discharge of neuropeptides and reduces the extent of ethanolin-induced gastric injury in rats[49,52]. This impact is operated by the discharge of CGRP from receptive nerve endings before their degeneration, which happens hours or days after the capsaicin injection. Numerous studies have shown that hCGRP (837), a fraction of human CGRP lacking the cyclic loop at the amino terminus of native CGRP, inhibits the action of exogenous CGRP[53,54].

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## **VNS FOR INTESTINAL BOWEL DISEASE**

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VNS is a unique therapeutic method for chronic TNF-mediated inflammatory illnesses in the framework of bioelectronic medicine, with the objective of employing tiny stimulators to provide electrical nerve signals for therapeutic, rather than pharmaceutical, purposes[55-57]. VNS is already used to treat depression and epilepsy which is resistant to drugs[58]. There is currently no recognized curative medicine for IBD. Current medicines reduce disease activity, and when therapy is stopped, the condition recurs. TNF is one of the most significant cytokines in IBD, and anti-TNF medicines have transformed the therapy of the disease[59]. New compounds are available that target pro-inflammatory cytokines such as IL-12, IL-23, anti-integrin, and anti-JAK therapies[60,61]. In the case of treatment failure or an IBD consequence (perforation, abscess, stenosis), surgery is an option, although the disease reappears after the procedure. While anti-TNF medications are effective in IBD, there is a 20%-30% initial non-response rate, and the yearly chance of anti-TNF reactivity is 13% per patient year for infliximab and 20% per patient year for adalimumab[62-64]. This lack of secondary response is attributable to (i) the formation of autoantibodies, particularly for infliximab but also, to a lesser extent, for adalimumab, or (ii) secondary failure due to insufficient dose[65,66]. As a result of the risk of adverse effects and the requirement for ongoing therapy for these disorders, patients are increasingly hesitant to begin and maintain these treatments once they are in remission. The non-compliance rate is 30%-50% [67,68]. Therefore, targeted therapy for pro-inflammatory cytokines such as TNF-alpha and others using CAP could be extremely helpful with fewer side effects, no compliance issues, and cheaper than biologics (*i.e.*, anti TNF-alpha). In this case, targeting the VN's anti-inflammatory characteristics might be of interest. VNS, particularly as a non-drug therapy, has the potential to be employed as an

alternative to conventional biological therapies. A number of animal and clinical research have been undertaken in recent years to investigate the efficacy of VNS in the treatment of IBD (Supplementary Table 1).

### Animal evidence

Vagotomy has been found in several studies to enhance the disease activity index (DAI), gross and pro-inflammatory cytokine levels in mice[69-71]. To replicate UC, Chen and colleagues employed dextran sodium sulphate (DSS) colitis in mice. They observed that VNS eased cerebral cortical microinfarcts induced by a two-photon laser and reduced DSS colitis. This neuroprotection was linked to decreased blood-brain barrier permeability and inflammatory processes[72].

### Human evidence

Indirect data suggests that a vagal anti-inflammatory action plays a role in IBD. Vagal activity has been demonstrated to be inversely associated with inflammatory markers in healthy and cardiac patients as evaluated by HRV spectral analysis[73]. VNS might be an attractive method for the treatment of IBD based on pre-clinical results in rats with colitis and two recent clinical pilot trials targeting two distinct categories of patients with active CD, either ignorant of anti-TNF on inclusion or resistant to biologics [74].

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## LABORATORY AND CLINICAL STUDIES

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### Animal studies

Miceli and Jacobson[75] published the first data on the anti-inflammatory effects of VN in digestive inflammation. Colitis in rats with 2,4,6 trinitrobenzenesulfonic acid (TNBS) improved with early treatment of anticholinesterase medications such as neostigmine, which does not cross the blood-brain barrier, or physostigmine. This impact was more pronounced with physostigmine, indicating a dominating central mechanism. In mice, vagotomy aggravated experimental colitis, indicating that NV serves a protective function[76]. It was demonstrated that in the non-vagotomized watchful rat, 3 h per day for five consecutive days, low-frequency VNS (5 Hz) led in an improvement in TNBS colitis in rats [31] VNS inhibited weight loss and inflammatory indicators.

An improvement in a multivariate measure of colitis was also observed as an anti-inflammatory impact (which includes body mass, temperature, and motor function, macroscopic area of the lesions, histological and biological parameters such as myeloperoxidase activity, cytokines, and mRNAs related to cytokines)[77]. Sun *et al*[32] showed that chronic VNS increased the clinical activity index, the histological scores, the biological inflammation due to myeloperoxidase activity, the iNOS, TNF, and IL-6 Levels among rats with colitis, and the inflammatory response induced by LPS in cells of the human epithelial colorectal adenocarcinoma (Caco2) by ACh *in vitro*. In 2000, Kevin Tracey's team first described CAP[78,79]. They found that there is an inflammatory reflex in which proinflammatory cytokines stimulate vagus afferents, which activate vagus efferents, causing the production of these cytokines by tissue macrophages, mainly TNF, but also other pro-inflammatory cytokines such as IL-6. IL-1b, but not IL-10, an anti-inflammatory cytokine VN has anti-inflammatory effects because it inhibits pro-inflammatory cytokines.

### Human studies

Decreased vagus activity was observed to be related to systemic inflammatory markers in both IC and CD patients[80,81]. VNS improved several inflammatory markers in rats' small intestines, including fecal quality, inflammatory processes, and leukocyte infiltration. Furthermore, considerable cardiac and respiratory changes happened with supra-threshold cervical VNS, while abdominal VNS caused alterations. Due to the lack of side effects and effectiveness in reducing inflammation, abdominal VNS appears to be a viable alternative to cervical VNS. This evidence supports the application of this novel peripheral nerve network for abdominal VNS as a potential therapy for IBD like CD[82]. A pilot study on VNS was carried out for the first time in patients with moderate to severe celiac disease as an alternative to drug anti-TNF therapy or in untreated patients in a translational approach from the laboratory to the bedside[56]. A VNS device and electrode were implanted in nine patients. At the time of implantation, two patients had failed immunosuppressive drugs (azathioprine), while the other seven received no treatment[56]. ENV was carried out on a continuous basis over a period of one year. In April 2012, the first patient was implanted, and then the last in March 2016.

Due to increasing condition, two patients were removed from the trial after three months of neurostimulation: The first had ileocecal resection but elected to continue neurostimulation until the end of the study due to an early good response and rejection of pharmaceutical therapy. The second patient took infliximab and azathioprine and continued to use an active VNS. Six patients were in remission owing to neurostimulation alone after one year of follow-up, while the seventh was in relapse. In April 2012, the first patient to get the implant was in remission from azathioprine in ileal CD with a history of ileocecal resection[56]. In conclusion, five out of seven patients who received the one-year VNS attained

clinical improvement (CDAI 150), and all gained the CDAI70 response (CDAI decreased 70 points from baseline). Similarly, the Endoscopic CD Severity Score (CDEIS) decreased from 60% to 100% in five patients. Other than complaints caused by the output current/intensity of the device, no adverse events were observed[56]. In patients with UC decrease activity has been linked with autonomic function[83].

### Devices and methods

Currently, the generally used VNS therapeutic equipment is invasive and implantable. The VNS Therapy System consists of an implanted pulse generator, a bipolar VNS electrode, a small handheld device, programming software, a programming stick, and hand magnets. VNS is traditionally used to treat epilepsy and depression, as well as in the two pilot studies in patients with CD.

It is invasive, generally performed by a neurosurgeon who is experienced in the surgery, and lasts 1 h with minimal side effects. Noninvasive (n) VNS may be beneficial in certain patients who are afraid to have surgery in a vasculo-nervous location, such as the vein or the external carotid artery, which are close to the VN. Furthermore, if the device is removed, the electrode wrapped around the VN is normally kept in place, although some writers have removed it without causing significant nerve and artery damage[84]. Anesthesia is necessary for the operation, which requires two small incisions. The bipolar lead is looped around the left cervical VN and the pulse generator is positioned in the top-left chest. Physicians program the stimulator with a small handheld device, programming software, and a programming stick. After implantation, patients are given a wearable magnet to manipulate the stimulation on their own. The left vagus, which is more intimately linked to cardiovascular activities, is considered more suitable than the right cervical vagus. In the treatment of epilepsy, right-sided VNS has been observed in numerous patients[85-87]. Right-sided VNS appears to be as effective as, if not more successful than, left VNS[88]. Gadgets stimulating the VN on the cervical degree or on the auricular degree were produced (Figure 3). Certainly, the cymbal concha of the external ear is innervated by means of a sensory auricular branch of the VN that sends projection inside the NTS in cats and human beings[89-91]. These noninvasive devices have not been associated with any significant major side effects. In comparison to invasive VNS, n-VNS has the disadvantage of low compliance, which is a major concern in the treatment of IBD. Indeed, 30%-40% of IBD patients fail to take their medicine[92]. One can wonder if the same problem arises with these noninvasive devices. Furthermore, in the case of the Gamma core device, the repeatability of the placement of the discs in contact with the VN is unknown. Finally, ta-VNS was less efficient than VNS in decreasing the LPS-induced serum cytokine (TNF, IL-1, and IL-6) response in a septic shock animal[93].

### Mechanism of VNS

An unexpected receptor mechanism underpins the anti-inflammatory effect of the Vagus nerve. In comparison to many "classical" physiological activities, which might be managed with the aid of metabotropic mAChRs, the anti-inflammatory effects of the Vagus nerve are mediated *via* ionotropic nicotinic acetylcholine receptors (nAChRs)[94]. The frequency of stimulation for VN activation is critical to the function of various therapies[95-100]. A couple of studies have indicated that mAChRs, especially the M1 mAChR, play a role in this regulation in endotoxemia, inflammatory bowel disorder (colitis), hemorrhagic shock, and other illnesses[101,102-104].

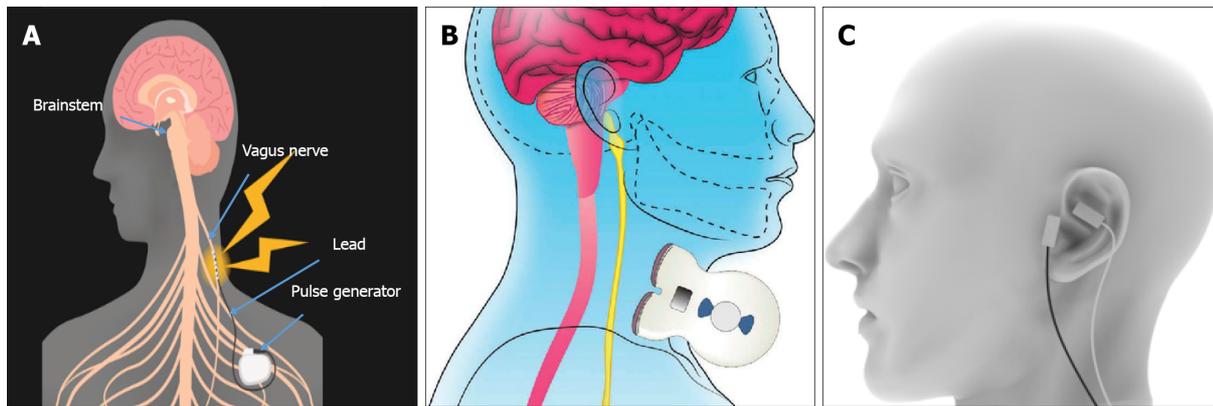
Increased cholinergic transmission in the brain with centrally acting acetylcholinesterase inhibitors, particularly galantamine, leads to inhibition of unusual inflammatory responses generated by vagus nerve impulses in mice models of endotoxemia, colitis, and lupus[105-107]. The most recent work, which used targeted optogenetic stimulation and sophisticated pharmacological methods, discovered that forebrain signal transduction and M1 mAChR play a unique role in the modulation of peripheral inflammatory responses in endotoxemia mice *via* vagus nerve transduction[108]. VNS blocks splenic TNF, which has been identified as a primary contributor to systemic TNF. It is critical to understand how the vagus nerve regulates cytokines in the spleen. The vagus nerve innervates the celiac ganglia and the superior mesenteric ganglion, which have been shown to provide neurons to the splenic nerve [101].

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## SNS

The sacral nerves are divided into five pairs. Each contains an afferent and efferent component, allowing for effective interaction between the lower GIT and the nervous system. The activity of the lower GIT (descending colon, rectum), sexual organs, and urinary bladder is modulated by the parasympathetic component of sacral nerves. The principal somatic nerve of the sacral plexus is the pudendal nerve (S2-S4). It is both sensory and motor. The external anal sphincter, which is under our conscious control, receives sensory and motor innervation from it. It also gives sensation to the external genitalia, the skin around the anal area, the anal canal, the perineum, and motor innervation to the external urethral sphincter.

SNS, also known as "sacral neuromodulation", is a relatively new and promising treatment option. SNS uses an implanted device that stimulates the S3 nerve root and offers a wide range of applications



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**Figure 3 Vagus nerve stimulation.** A: Direct vagus nerve stimulation; B and C: Noninvasive vagus nerve stimulation; transcutaneous cervical vagus nerve stimulation (VNS) (B); transcutaneous auricular VNS (C).

in conditions such as urgency urinary incontinence, pelvic pain, detrusor stimulation with transurethral approach, FI, *etc.* Following are some applications of SNS in relation to IBD[3].

### SACRAL NEUROMODULATION FOR INFLAMMATION AND INTESTINAL BARRIER IN IBD

Experimental and clinical evidence from several studies signifies the potential of SNS as a treatment option for IBD Patients who have received SNS had less severe mucosal lesions than those who have not received SNS. SNS also improves the recovery of enema caused by trinitrobenzene sulfonic acid (TNBS enema). With elevated TNF1 and trypsin levels, SNS also increases the number of mucosal neutrophils. SNS also stopped TNBS-induced inflammatory factors, including IL-4 and IL-1, from rising. All of these variables indicate that sacral neuromodulation is beneficial in restoring the intestinal barrier following mucosa injury[109]. In IBD, SNS has a significant anti-inflammatory impact. SNS enhanced the spinal afferent-vagal efferent pathway and improved autonomic function by increased vagal efferent activity. SNS also causes anti-inflammatory effects due to the SNS-mediated release of Ach[110]. In a study using the TNBS rat model, sacral neuromodulation lowered the level of pro-inflammatory cytokines and improved colonic inflammation[111].

### SACRAL NEUROMODULATION IN FECAL INCONTINENCY

The inability to regulate bowel movements, which can range from modest rectum leaks to total bowel control, is known as FI. Viability of sacral neuromodulations as a treatment option for FI is tremendous. Many studies have demonstrated that FI responds positively to SNS. SNS has proven to be a reliable method for dealing with FI in children[112]. Clinical trials have also shown that SNS can help with FI [113]. Another extensive approach conveys the benefits of SNS in patients with neuropathic FI[114].

### OTHER METHODS OF NEUROMODULATION

Other than SNS and VNS, other neuromodulation methods to treat IBD are TNS and Spinal Cord Stimulation (SCS). The sensory, motor and autonomic fibers in the tibial nerve make it a mixed nerve. It is caused by the L4-S3 nerves, which feed the colorectum, bladder, and pelvic floor. TNS uses electrical impulses to treat bladder and pelvic floor issues. TNS is classified into two types: Percutaneous TNS (PTNS) and transcutaneous TNS (TCTNS) (TTNS). The former makes use of a needle electrode, whilst the latter makes use of a sticky electrode[3]. PTNS is a minimally invasive method that has been demonstrated to be beneficial in treating overactive bladder, FI, and pelvic discomfort. Having few side effects is highly convenient, but it is limited by the necessity that patients visit the clinic weekly to obtain the series of treatments[115].

The actual mechanism of TNS is uncertain however it appears to involve excitation of afferent pathways to the sacral spinal cord as well as regulation of efferent nerves[116]. Retrospective research looked at 183 individuals with refractory overactive bladder (OAB) who had 30-min PTNS sessions for 12 wk during nine years. There was a significant improvement in micturition frequency, nocturia, and urge incontinence episodes in the PTNS group, with the impact obvious by week 10 of therapy. With a

wide range of PTNS times, 61.5 percent of subjects self-proclaimed > 50% improvement in signs and symptoms, raising the subjective accomplishment percentages[117]. For a 12-wk treatment period, a recent randomized research of forty women with nocturia of weekly TTNS periods compared pelvic floor muscle training and behavioral therapy. Both medicines improved sleep quality by reducing the number of times people awoke to pee (45 percent reduced by 1 in both groups)[118]. A spinal cord stimulator (SCS) is surgically implanted under the skin and delivers a weak electrical current to the spinal cord. Current from a pulse generator is carried to the spinal cords' nerve fibers by thin wires. When the SCS is activated, it stimulates the nerves in the area where a person is feeling pain. The pain signal is altered and masked by electrical impulses, prohibiting it from going to the brain[119].

For more than a half-century, spinal cord stimulation (SCS) has been used to treat chronic pain. Several studies have demonstrated that SCS can help with stomach discomfort[120]. Randomized trial has shown that SCS can lessen diarrhea and pain in persons with irritable bowel syndrome[121]. Although it has been quite successful, some people might experience device-related challenges such as pain at the implantation site or subsequent infections. But it doesn't cause any serious complications like paralysis or hemorrhage in the epidural space[122,123].

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## CONCLUSION

The digestive system's broad and approachable interaction with the CNS, the predominance of IBD, and the lack of effective treatment options make it an appealing target for bioelectrical neuromodulation therapy for digestive system innervation. A wide range of gastrointestinal problems has been treated with various degrees of success. This approach has been tried with different degrees of effectiveness in a range of gastrointestinal diseases. SNS for faecal incontinence has become a popular bio-electric therapy for gastrointestinal disorders. The development of bioelectrical digestive system neuromodulation medicines requires investigation. The advancement of our understanding of the multiple roles of the mixed nerve components, such as vagus nerves and sympathetic routes to the intestines, should allow us to take IBS treatment to a new level.

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## FOOTNOTES

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## REFERENCES

- 1 Szigethy E, McLafferty L, Goyal A. Inflammatory bowel disease. *Child Adolesc Psychiatr Clin N Am* 2010; **19**: 301-318, ix [PMID: 20478501 DOI: 10.1016/j.chc.2010.01.007]
- 2 Mayo Clinic gastroenterology and hepatology board review 95 ed. Oxford Medicine Online. Inflammatory bowel disease (IBD)-Symptoms and causes-Mayo Clinic. [cited 6 July 2022] [DOI: 10.1093/med/9780199373338.003.0019]

- 3 **Cheng J**, Shen H, Chowdhury R, Abdi T, Selaru F, Chen JDZ. Potential of Electrical Neuromodulation for Inflammatory Bowel Disease. *Inflamm Bowel Dis* 2020; **26**: 1119-1130 [PMID: 31782957 DOI: 10.1093/ibd/izz289]
- 4 **Payne SC**, Furness JB, Stebbing MJ. Bioelectric neuromodulation for gastrointestinal disorders: effectiveness and mechanisms. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 89-105 [PMID: 30390018 DOI: 10.1038/s41575-018-0078-6]
- 5 Erratum for the Research Article "Identification of Integrator-PP2A complex (INTAC), an RNA polymerase II phosphatase" by H. Zheng, Y. Qi, S. Hu, X. Cao, C. Xu, Z. Yin, X. Chen, Y. Li, W. Liu, J. Li, J. Wang, G. Wei, K. Liang, F. X. Chen, Y. Xu. *Science* 2021; **371** [PMID: 33446562 DOI: 10.1126/science.abg4122]
- 6 **Adams D**, Stocker A, Lancaster W, Abell T. The Surgeon's Role in Gastric Electrical Stimulation Therapy for Gastroparesis. *J Gastrointest Surg* 2021; **25**: 1053-1064 [PMID: 33236324 DOI: 10.1007/s11605-020-04850-w]
- 7 **Engel MA**, Neurath MF. New pathophysiological insights and modern treatment of IBD. *J Gastroenterol* 2010; **45**: 571-583 [PMID: 20213337 DOI: 10.1007/s00535-010-0219-3]
- 8 **Smart NA**, Fisher S, Pearson MJ. On "Physical Therapist Clinical Practice Guideline for the Management of Individuals With Heart Failure." Shoemaker MJ, Dias KJ, Lefebvre KM, Heick JD, Collins SM. *Phys Ther*. 2020;100:14-43. *Phys Ther* 2020; **100**: 1882 [PMID: 32737486 DOI: 10.1093/ptj/pzaa130]
- 9 **Furness JB**, Rivera LR, Cho HJ, Bravo DM, Callaghan B. The gut as a sensory organ. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 729-740 [PMID: 24061204 DOI: 10.1038/nrgastro.2013.180]
- 10 **Collins SM**, Surette M, Bercik P. The interplay between the intestinal microbiota and the brain. *Nat Rev Microbiol* 2012; **10**: 735-742 [PMID: 23000955 DOI: 10.1038/nrmicro2876]
- 11 **Goehler LE**, Gaykema RP, Opitz N, Reddaway R, Badr N, Lyte M. Activation in vagal afferents and central autonomic pathways: early responses to intestinal infection with *Campylobacter jejuni*. *Brain Behav Immun* 2005; **19**: 334-344 [PMID: 15944073 DOI: 10.1016/j.bbi.2004.09.002]
- 12 **Nishida A**, Inoue R, Inatomi O, Bamba S, Naito Y, Andoh A. Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin J Gastroenterol* 2018; **11**: 1-10 [PMID: 29285689 DOI: 10.1007/s12328-017-0813-5]
- 13 **Berthoud HR**, Neuhuber WL. Functional and chemical anatomy of the afferent vagal system. *Auton Neurosci* 2000; **85**: 1-17 [PMID: 11189015 DOI: 10.1016/s1566-0702(00)00215-0]
- 14 **Becker C**, Neurath MF, Wirtz S. The Intestinal Microbiota in Inflammatory Bowel Disease. *ILAR J* 2015; **56**: 192-204 [PMID: 26323629 DOI: 10.1093/ilar/ilv030]
- 15 **Bonaz B**, Sinniger V, Pellissier S. Vagus nerve stimulation: a new promising therapeutic tool in inflammatory bowel disease. *J Intern Med* 2017; **282**: 46-63 [PMID: 28421634 DOI: 10.1111/joim.12611]
- 16 **Tracey KJ**. The inflammatory reflex. *Nature* 2002; **420**: 853-859 [PMID: 12490958 DOI: 10.1038/nature01321]
- 17 **Goehler LE**, Gaykema RP, Hansen MK, Anderson K, Maier SF, Watkins LR. Vagal immune-to-brain communication: a visceral chemosensory pathway. *Auton Neurosci* 2000; **85**: 49-59 [PMID: 11189026 DOI: 10.1016/s1566-0702(00)00219-8]
- 18 **HARRIS GW**. The hypothalamus and endocrine glands. *Br Med Bull* 1950; **6**: 345-350 [PMID: 15420401 DOI: 10.1093/oxfordjournals.bmb.a073628]
- 19 **Goehler LE**, Relton JK, Dripps D, Kiechle R, Tartaglia N, Maier SF, Watkins LR. Vagal paraganglia bind biotinylated interleukin-1 receptor antagonist: a possible mechanism for immune-to-brain communication. *Brain Res Bull* 1997; **43**: 357-364 [PMID: 9227848 DOI: 10.1016/s0361-9230(97)00020-8]
- 20 **Bonaz B**, Sinniger V, Pellissier S. Anti-inflammatory properties of the vagus nerve: potential therapeutic implications of vagus nerve stimulation. *J Physiol* 2016; **594**: 5781-5790 [PMID: 27059884 DOI: 10.1113/JP271539]
- 21 **Pavlov VA**, Wang H, Czura CJ, Friedman SG, Tracey KJ. The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation. *Mol Med* 2003; **9**: 125-134 [PMID: 14571320]
- 22 **Caulfield MP**, Birdsall NJM; International union of pharmacology. XVII. Classification of muscarinic acetylcholine receptors. In: *Pharmacol*, 1998
- 23 **Lindstrom J**. Nicotinic acetylcholine receptors in health and disease. *Mol Neurobiol* 1997; **15**: 193-222 [PMID: 9396010 DOI: 10.1007/BF02740634]
- 24 **Borovikova LV**, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, Wang H, Abumrad N, Eaton JW, Tracey KJ. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000; **405**: 458-462 [PMID: 10839541 DOI: 10.1038/35013070]
- 25 **Steinlein O**. New functions for nicotinic acetylcholine receptors? *Behav Brain Res* 1998; **95**: 31-35 [PMID: 9754874 DOI: 10.1016/s0166-4328(97)00207-6]
- 26 **Leonard S**, Bertrand D. Neuronal nicotinic receptors: from structure to function. *Nicotine Tob Res* 2001; **3**: 203-223 [PMID: 11506765 DOI: 10.1080/14622200110050213]
- 27 **Tracey KJ**. Hacking the inflammatory reflex. *The Lancet Rheumatology* 2021; **3**: e237 e239 [DOI: 10.1016/s2665-9913(20)30448-3]
- 28 **Mayer EA**, Savidge T, Shulman RJ. Brain-gut microbiome interactions and functional bowel disorders. *Gastroenterology* 2014; **146**: 1500-1512 [PMID: 24583088 DOI: 10.1053/j.gastro.2014.02.037]
- 29 **Gacias M**, Gaspari S, Santos PM, Tamburini S, Andrade M, Zhang F, Shen N, Tolstikov V, Kiebish MA, Dupree JL, Zachariou V, Clemente JC, Casaccia P. Microbiota-driven transcriptional changes in prefrontal cortex override genetic differences in social behavior. *Elife* 2016; **5** [PMID: 27097105 DOI: 10.7554/eLife.13442]
- 30 **Foster JA**, McVey Neufeld KA. Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci* 2013; **36**: 305-312 [PMID: 23384445 DOI: 10.1016/j.tins.2013.01.005]
- 31 **Meregani J**, Clarençon D, Vivier M, Peinnequin A, Mouret C, Sinniger V, Picq C, Job A, Canini F, Jacquier-Sarlin M, Bonaz B. Anti-inflammatory effect of vagus nerve stimulation in a rat model of inflammatory bowel disease. *Auton Neurosci* 2011; **160**: 82-89 [PMID: 21071287 DOI: 10.1016/j.autneu.2010.10.007]
- 32 **Sun P**, Zhou K, Wang S, Li P, Chen S, Lin G, Zhao Y, Wang T. Involvement of MAPK/NF- $\kappa$ B signaling in the activation of the cholinergic anti-inflammatory pathway in experimental colitis by chronic vagus nerve stimulation. *PLoS One* 2013; **8**: e69424 [PMID: 23936328 DOI: 10.1371/journal.pone.0069424]

- 33 **Banks WA**, Kastin AJ, Broadwell RD. Passage of cytokines across the blood-brain barrier. *Neuroimmunomodulation* 1995; **2**: 241-248 [PMID: 8963753 DOI: 10.1159/000097202]
- 34 **Buller KM**. Role of circumventricular organs in pro-inflammatory cytokine-induced activation of the hypothalamic-pituitary-adrenal axis. *Clin Exp Pharmacol Physiol* 2001; **28**: 581-589 [PMID: 11458886 DOI: 10.1046/j.1440-1681.2001.03490.x]
- 35 **Lomax AE**, Sharkey KA, Furness JB. The participation of the sympathetic innervation of the gastrointestinal tract in disease states. *Neurogastroenterol Motil* 2010; **22**: 7-18 [PMID: 19686308 DOI: 10.1111/j.1365-2982.2009.01381.x]
- 36 **Evans RJ**, Surprenant A. Vasoconstriction of guinea-pig submucosal arterioles following sympathetic nerve stimulation is mediated by the release of ATP. *Br J Pharmacol* 1992; **106**: 242-249 [PMID: 1356556 DOI: 10.1111/j.1476-5381.1992.tb14323.x]
- 37 **Lundberg JM**, Rudehill A, Sollevi A. Pharmacological characterization of neuropeptide Y and noradrenaline mechanisms in sympathetic control of pig spleen. *Eur J Pharmacol* 1989; **163**: 103-113 [PMID: 2744085 DOI: 10.1016/0014-2999(89)90401-9]
- 38 **Strack AM**, Sawyer WB, Platt KB, Loewy AD. CNS cell groups regulating the sympathetic outflow to adrenal gland as revealed by transneuronal cell body labeling with pseudorabies virus. *Brain Res* 1989; **491**: 274-296 [PMID: 2548665 DOI: 10.1016/0006-8993(89)90063-2]
- 39 **Abe C**, Inoue T, Inglis MA, Viar KE, Huang L, Ye H, Rosin DL, Stornetta RL, Okusa MD, Guyenet PG. C1 neurons mediate a stress-induced anti-inflammatory reflex in mice. *Nat Neurosci* 2017; **20**: 700-707 [PMID: 28288124 DOI: 10.1038/nn.4526]
- 40 **Cervi AL**, Lukewich MK, Lomax AE. Neural regulation of gastrointestinal inflammation: role of the sympathetic nervous system. *Auton Neurosci* 2014; **182**: 83-88 [PMID: 24412637 DOI: 10.1016/j.autneu.2013.12.003]
- 41 **Gabanyi I**, Muller PA, Feighery L, Oliveira TY, Costa-Pinto FA, Mucida D. Neuro-immune Interactions Drive Tissue Programming in Intestinal Macrophages. *Cell* 2016; **164**: 378-391 [PMID: 26777404 DOI: 10.1016/j.cell.2015.12.023]
- 42 **Bellinger DL**, Felten SY, Lorton D, Felten DL. Origin of noradrenergic innervation of the spleen in rats. *Brain Behav Immun* 1989; **3**: 291-311 [PMID: 2575409 DOI: 10.1016/0889-1591(89)90029-9]
- 43 **Huston JM**, Ochani M, Rosas-Ballina M, Liao H, Ochani K, Pavlov VA, Gallowitsch-Puerta M, Ashok M, Czura CJ, Foxwell B, Tracey KJ, Ulloa L. Splenectomy inactivates the cholinergic antiinflammatory pathway during lethal endotoxemia and polymicrobial sepsis. *J Exp Med* 2006; **203**: 1623-1628 [PMID: 16785311 DOI: 10.1084/jem.20052362]
- 44 **Rosas-Ballina M**, Olofsson PS, Ochani M, Valdés-Ferrer SI, Levine YA, Reardon C, Tusche MW, Pavlov VA, Andersson U, Chavan S, Mak TW, Tracey KJ. Acetylcholine-synthesizing T cells relay neural signals in a vagus nerve circuit. *Science* 2011; **334**: 98-101 [PMID: 21921156 DOI: 10.1126/science.1209985]
- 45 **Martelli D**, McKinley MJ, McAllen RM. The cholinergic anti-inflammatory pathway: a critical review. *Auton Neurosci* 2014; **182**: 65-69 [PMID: 24411268 DOI: 10.1016/j.autneu.2013.12.007]
- 46 **Martelli D**, Farmer DG, Yao ST. The splanchnic anti-inflammatory pathway: could it be the efferent arm of the inflammatory reflex? *Exp Physiol* 2016; **101**: 1245-1252 [PMID: 27377300 DOI: 10.1113/EP085559]
- 47 **Holzer P**. Local effector functions of capsaicin-sensitive sensory nerve endings: involvement of tachykinins, calcitonin gene-related peptide and other neuropeptides. *Neuroscience* 1988; **24**: 739-768 [PMID: 3288903 DOI: 10.1016/0306-4522(88)90064-4]
- 48 **Sternini C**, Reeve JR Jr, Brecha N. Distribution and characterization of calcitonin gene-related peptide immunoreactivity in the digestive system of normal and capsaicin-treated rats. *Gastroenterology* 1987; **93**: 852-862 [PMID: 3305137 DOI: 10.1016/0016-5085(87)90450-1]
- 49 **Holzer P**, Lippe IT. Stimulation of afferent nerve endings by intragastric capsaicin protects against ethanol-induced damage of gastric mucosa. *Neuroscience* 1988; **27**: 981-987 [PMID: 3252180 DOI: 10.1016/0306-4522(88)90201-1]
- 50 **Holzer P**, Lippe IT. Role of calcitonin gene-related peptide in gastrointestinal blood flow. *Ann N Y Acad Sci* 1992; **657**: 228-239 [PMID: 1637087 DOI: 10.1111/j.1749-6632.1992.tb22771.x]
- 51 **Jacobson ED**. Vascular mediation of gastric mucosal damage and cytoprotection. *Indian J Physiol Pharmacol* 1990; **34**: 223-234 [PMID: 2100285]
- 52 **Lambrecht N**, Burchert M, Respondek M, Müller KM, Peskar BM. Role of calcitonin gene-related peptide and nitric oxide in the gastroprotective effect of capsaicin in the rat. *Gastroenterology* 1993; **104**: 1371-1380 [PMID: 8387040 DOI: 10.1016/0016-5085(93)90345-d]
- 53 **Han SP**, Naes L, Westfall TC. Inhibition of periarterial nerve stimulation-induced vasodilation of the mesenteric arterial bed by CGRP (8-37) and CGRP receptor desensitization. *Biochem Biophys Res Commun* 1990; **168**: 786-791 [PMID: 2159300 DOI: 10.1016/0006-291x(90)92390-1]
- 54 **Gardiner SM**, Compton AM, Kemp PA, Bennett T, Bose C, Foulkes R, Hughes B. Antagonistic effect of human alpha-CGRP [8-37] on the in vivo regional haemodynamic actions of human alpha-CGRP. *Biochem Biophys Res Commun* 1990; **171**: 938-943 [PMID: 2222455 DOI: 10.1016/0006-291x(90)90774-h]
- 55 **Jo BG**, Kim SH, Namgung U. Vagal afferent fibers contribute to the anti-inflammatory reactions by vagus nerve stimulation in concanavalin A model of hepatitis in rats. *Mol Med* 2020; **26**: 119 [PMID: 33272194 DOI: 10.1186/s10020-020-00247-2]
- 56 **Bonaz B**, Sinniger V, Hoffmann D, Clarençon D, Mathieu N, Dantzer C, Vercueil L, Picq C, Trocmé C, Faure P, Cracowski JL, Pellissier S. Chronic vagus nerve stimulation in Crohn's disease: a 6-month follow-up pilot study. *Neurogastroenterol Motil* 2016; **28**: 948-953 [PMID: 26920654 DOI: 10.1111/nmo.12792]
- 57 **Olofsson PS**, Tracey KJ. Bioelectronic medicine: technology targeting molecular mechanisms for therapy. *J Intern Med* 2017; **282**: 3-4 [PMID: 28621493 DOI: 10.1111/joim.12624]
- 58 **Bonaz B**, Picq C, Sinniger V, Mayol JF, Clarençon D. Vagus nerve stimulation: from epilepsy to the cholinergic anti-inflammatory pathway. *Neurogastroenterol Motil* 2013; **25**: 208-221 [PMID: 23360102 DOI: 10.1111/nmo.12076]
- 59 **Peyrin-Biroulet L**. Anti-TNF therapy in inflammatory bowel diseases: a huge review. *Minerva Gastroenterol Dietol* 2010; **56**: 233-243 [PMID: 20485259]
- 60 **Pagnini C**, Pizarro TT, Cominelli F. Novel Pharmacological Therapy in Inflammatory Bowel Diseases: Beyond Anti-

- Tumor Necrosis Factor. *Front Pharmacol* 2019; **10**: 671 [PMID: 31316377 DOI: 10.3389/fphar.2019.00671]
- 61 **D'Amico F**, Peyrin-Biroulet L, Danese S, Fiorino G. New drugs in the pipeline for the treatment of inflammatory bowel diseases: what is coming? *Curr Opin Pharmacol* 2020; **55**: 141-150 [PMID: 33254032 DOI: 10.1016/j.coph.2020.10.015]
- 62 **Ford AC**, Sandborn WJ, Khan KJ, Hanauer SB, Talley NJ, Moayyedi P. Efficacy of biological therapies in inflammatory bowel disease: systematic review and meta-analysis. *Am J Gastroenterol* 2011; **106**: 644-659, quiz 660 [PMID: 21407183 DOI: 10.1038/ajg.2011.73]
- 63 **Gisbert JP**, Panés J. Loss of response and requirement of infliximab dose intensification in Crohn's disease: a review. *Am J Gastroenterol* 2009; **104**: 760-767 [PMID: 19174781 DOI: 10.1038/ajg.2008.88]
- 64 **Billioud V**, Sandborn WJ, Peyrin-Biroulet L. Loss of response and need for adalimumab dose intensification in Crohn's disease: a systematic review. *Am J Gastroenterol* 2011; **106**: 674-684 [PMID: 21407178 DOI: 10.1038/ajg.2011.60]
- 65 **Ben-Horin S**, Chowers Y. Review article: loss of response to anti-TNF treatments in Crohn's disease. *Aliment Pharmacol Ther* 2011; **33**: 987-995 [PMID: 21366636 DOI: 10.1111/j.1365-2036.2011.04612.x]
- 66 **Chaparro M**, Guerra I, Muñoz-Linares P, Gisbert JP. Systematic review: antibodies and anti-TNF- $\alpha$  levels in inflammatory bowel disease. *Aliment Pharmacol Ther* 2012; **35**: 971-986 [PMID: 22443153 DOI: 10.1111/j.1365-2036.2012.05057.x]
- 67 **Chan W**, Chen A, Tiao D, Selinger C, Leong R. Medication adherence in inflammatory bowel disease. *Intest Res* 2017; **15**: 434-445 [PMID: 29142511 DOI: 10.5217/ir.2017.15.4.434]
- 68 **Torres J**, Ellul P, Langhorst J, Mikocka-Walus A, Barreiro-de Acosta M, Basnayake C, Ding NJS, Gilardi D, Katsanos K, Moser G, Opheim R, Palmela C, Pellino G, Van der Marel S, Vavricka SR. European Crohn's and Colitis Organisation Topical Review on Complementary Medicine and Psychotherapy in Inflammatory Bowel Disease. *J Crohns Colitis* 2019; **13**: 673-685e [PMID: 30820529 DOI: 10.1093/ecco-jcc/ijz051]
- 69 **Ghia JE**, Blennerhassett P, Kumar-Ondiveeran H, Verdu EF, Collins SM. The vagus nerve: a tonic inhibitory influence associated with inflammatory bowel disease in a murine model. *Gastroenterology* 2006; **131**: 1122-1130 [PMID: 17030182 DOI: 10.1053/j.gastro.2006.08.016]
- 70 **Ghia JE**, Blennerhassett P, El-Sharkawy RT, Collins SM. The protective effect of the vagus nerve in a murine model of chronic relapsing colitis. *Am J Physiol Gastrointest Liver Physiol* 2007; **293**: G711-G718 [PMID: 17673544 DOI: 10.1152/ajpgi.00240.2007]
- 71 **Di Giovangiulio M**, Bosmans G, Meroni E, Stakenborg N, Florens M, Farro G, Gomez-Pinilla PJ, Matteoli G, Boeckxstaens GE. Vagotomy affects the development of oral tolerance and increases susceptibility to develop colitis independently of the alpha-7 nicotinic receptor. *Mol Med* 2016; **22**: 464-476 [PMID: 27341335 DOI: 10.2119/molmed.2016.00062]
- 72 **Chen X**, He X, Luo S, Feng Y, Liang F, Shi T, Huang R, Pei Z, Li Z. Vagus Nerve Stimulation Attenuates Cerebral Microinfarct and Colitis-induced Cerebral Microinfarct Aggravation in Mice. *Front Neurol* 2018; **9**: 798 [PMID: 30319530 DOI: 10.3389/fneur.2018.00798]
- 73 **Frasure-Smith N**, Lespérance F, Irwin MR, Talajic M, Pollock BG. The relationships among heart rate variability, inflammatory markers and depression in coronary heart disease patients. *Brain Behav Immun* 2009; **23**: 1140-1147 [PMID: 19635552 DOI: 10.1016/j.bbi.2009.07.005]
- 74 **Bonaz B**. Is-there a place for vagus nerve stimulation in inflammatory bowel diseases? *Bioelectron Med* 2018; **4**: 4 [PMID: 32232080 DOI: 10.1186/s42234-018-0004-9]
- 75 **Miceli PC**, Jacobson K. Cholinergic pathways modulate experimental dinitrobenzene sulfonic acid colitis in rats. *Auton Neurosci* 2003; **105**: 16-24 [PMID: 12742187 DOI: 10.1016/S1566-0702(03)00023-7]
- 76 **Caravaca AS**, Levine YA, Drake A, Eberhardson M, Olofsson PS. Vagus Nerve Stimulation Reduces Indomethacin-Induced Small Bowel Inflammation. *Front Neurosci* 2021; **15**: 730407 [PMID: 35095387 DOI: 10.3389/fnins.2021.730407]
- 77 **Bonaz B**, Sinniger V, Pellissier S. Therapeutic Potential of Vagus Nerve Stimulation for Inflammatory Bowel Diseases. *Front Neurosci* 2021; **15**: 650971 [PMID: 33828455 DOI: 10.3389/fnins.2021.650971]
- 78 **Borovikova LV**, Ivanova S, Nardi D, Zhang M, Yang H, Ombrellino M, Tracey KJ. Role of vagus nerve signaling in CNI-1493-mediated suppression of acute inflammation. *Auton Neurosci* 2000; **85**: 141-147 [PMID: 11189021 DOI: 10.1016/S1566-0702(00)00233-2]
- 79 **Jarczyk J**, Yard BA, Hoeger S. The Cholinergic Anti-Inflammatory Pathway as a Conceptual Framework to Treat Inflammation-Mediated Renal Injury. *Kidney Blood Press Res* 2019; **44**: 435-448 [PMID: 31307039 DOI: 10.1159/000500920]
- 80 **Pellissier S**, Dantzer C, Mondillon L, Trocme C, Gauchez AS, Ducros V, Mathieu N, Toussaint B, Fournier A, Canini F, Bonaz B. Relationship between vagal tone, cortisol, TNF-alpha, epinephrine and negative affects in Crohn's disease and irritable bowel syndrome. *PLoS One* 2014; **9**: e105328 [PMID: 25207649 DOI: 10.1371/journal.pone.0105328]
- 81 **Gunterberg V**, Simrén M, Öhman L, Friberg P, Jones MP, Van Oudenhove L, Strid H. Autonomic nervous system function predicts the inflammatory response over three years in newly diagnosed ulcerative colitis patients. *Neurogastroenterol Motil* 2016; **28**: 1655-1662 [PMID: 27265090 DOI: 10.1111/nmo.12865]
- 82 **Payne SC**, Furness JB, Burns O, Sedo A, Hyakumura T, Shepherd RK, Fallon JB. Anti-inflammatory Effects of Abdominal Vagus Nerve Stimulation on Experimental Intestinal Inflammation. *Front Neurosci* 2019; **13**: 418 [PMID: 31133776 DOI: 10.3389/fnins.2019.00418]
- 83 **Maunder RG**, Greenberg GR, Nolan RP, Lancee WJ, Steinhart AH, Hunter JJ. Autonomic response to standardized stress predicts subsequent disease activity in ulcerative colitis. *Eur J Gastroenterol Hepatol* 2006; **18**: 413-420 [PMID: 16538114 DOI: 10.1097/00042737-200604000-00016]
- 84 **Champeaux C**, Landré E, Chassoux F, Mann MW, Devaux B, Turak B. Vagus Nerve Stimulation Removal or Replacement Involving the Lead and the Electrode: Surgical Technique, Institutional Experience and Outcome. *World Neurosurg* 2017; **99**: 275-281 [PMID: 28012885 DOI: 10.1016/j.wneu.2016.11.103]
- 85 **McGregor A**, Wheless J, Baumgartner J, Bettis D. Right-sided vagus nerve stimulation as a treatment for refractory epilepsy in humans. *Epilepsia* 2005; **46**: 91-96 [PMID: 15660773 DOI: 10.1111/j.0013-9580.2005.16404.x]

- 86 **Spuck S**, Nowak G, Renneberg A, Tronnier V, Sperner J. Right-sided vagus nerve stimulation in humans: an effective therapy? *Epilepsy Res* 2008; **82**: 232-234 [PMID: 18801642 DOI: 10.1016/j.epilepsyres.2008.08.003]
- 87 **Navas M**, Navarrete EG, Pascual JM, Carrasco R, Núñez JA, Shakur SF, Pastor J, Sola RG. Treatment of refractory epilepsy in adult patients with right-sided vagus nerve stimulation. *Epilepsy Res* 2010; **90**: 1-7 [PMID: 20488666 DOI: 10.1016/j.epilepsyres.2010.04.007]
- 88 **Navas M**, Navarrete EG, Pascual JM, Carrasco R, Núñez JA, Shakur SF, Pastor J, Sola RG. Treatment of refractory epilepsy in adult patients with right-sided vagus nerve stimulation. *Epilepsy Res* 2010; **90**: 1-7 [PMID: 20488666 DOI: 10.1016/j.epilepsyres.2010.04.007]
- 89 **Stakenborg N**, Viola MF, Boeckxstaens GE. Intestinal neuro-immune interactions: focus on macrophages, mast cells and innate lymphoid cells. *Curr Opin Neurobiol* 2020; **62**: 68-75 [PMID: 31862627 DOI: 10.1016/j.conb.2019.11.020]
- 90 **Peuker ET**, Filler TJ. The nerve supply of the human auricle. *Clin Anat* 2002; **15**: 35-37 [PMID: 11835542 DOI: 10.1002/ca.1089]
- 91 **Nomura S**, Mizuno N. Central distribution of primary afferent fibers in the Arnold's nerve (the auricular branch of the vagus nerve): a transganglionic HRP study in the cat. *Brain Res* 1984; **292**: 199-205 [PMID: 6692153 DOI: 10.1016/0006-8993(84)90756-x]
- 92 **Frangos E**, Ellrich J, Komisaruk BR. Non-invasive Access to the Vagus Nerve Central Projections via Electrical Stimulation of the External Ear: fMRI Evidence in Humans. *Brain Stimul* 2015; **8**: 624-636 [PMID: 25573069 DOI: 10.1016/j.brs.2014.11.018]
- 93 **Herman ML**, Kane SV. Treatment Nonadherence in Inflammatory Bowel Disease: Identification, Scope, and Management Strategies. *Inflamm Bowel Dis* 2015; **21**: 2979-2984 [PMID: 26355470 DOI: 10.1097/MIB.0000000000000581]
- 94 **Zhao YX**, He W, Jing XH, Liu JL, Rong PJ, Ben H, Liu K, Zhu B. Transcutaneous auricular vagus nerve stimulation protects endotoxemic rat from lipopolysaccharide-induced inflammation. *Evid Based Complement Alternat Med* 2012; **2012**: 627023 [PMID: 23346208 DOI: 10.1155/2012/627023]
- 95 **Wang H**, Yu M, Ochani M, Amella CA, Tanovic M, Susarla S, Li JH, Wang H, Yang H, Ulloa L, Al-Abed Y, Czura CJ, Tracey KJ. Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature* 2003; **421**: 384-388 [PMID: 12508119 DOI: 10.1038/nature01339]
- 96 **Attenello F**, Amar AP, Liu C, Apuzzo ML. Theoretical Basis of Vagus Nerve Stimulation. *Prog Neurol Surg* 2015; **29**: 20-28 [PMID: 26393349 DOI: 10.1159/000434652]
- 97 **Zhao M**, He X, Bi XY, Yu XJ, Gil Wier W, Zang WJ. Vagal stimulation triggers peripheral vascular protection through the cholinergic anti-inflammatory pathway in a rat model of myocardial ischemia/reperfusion. *Basic Res Cardiol* 2013; **108**: 345 [PMID: 23519622 DOI: 10.1007/s00395-013-0345-1]
- 98 **Matteoli G**, Gomez-Pinilla PJ, Nemethova A, Di Giovangiulio M, Cailotto C, van Bree SH, Michel K, Tracey KJ, Schemann M, Boesmans W, Vanden Berghe P, Boeckxstaens GE. A distinct vagal anti-inflammatory pathway modulates intestinal muscularis resident macrophages independent of the spleen. *Gut* 2014; **63**: 938-948 [PMID: 23929694 DOI: 10.1136/gutjnl-2013-304676]
- 99 **Reyt S**, Picq C, Sinniger V, Clarençon D, Bonaz B, David O. Dynamic Causal Modelling and physiological confounds: a functional MRI study of vagus nerve stimulation. *Neuroimage* 2010; **52**: 1456-1464 [PMID: 20472074 DOI: 10.1016/j.neuroimage.2010.05.021]
- 100 **Barbella G**, Cocco I, Freri E, Marotta G, Visani E, Franceschetti S, Casazza M. Transcutaneous vagal nerve stimulation (t-VNS): An adjunctive treatment option for refractory epilepsy. *Seizure* 2018; **60**: 115-119 [PMID: 29940349 DOI: 10.1016/j.seizure.2018.06.016]
- 101 **Kibleur A**, Pellissier S, Sinniger V, Robert J, Gronlier E, Clarençon D, Vercueil L, Hoffmann D, Bonaz B, David O. Electroencephalographic correlates of low-frequency vagus nerve stimulation therapy for Crohn's disease. *Clin Neurophysiol* 2018; **129**: 1041-1046 [PMID: 29573733 DOI: 10.1016/j.clinph.2018.02.127]
- 102 **Pavlov VA**, Tracey KJ. Neural circuitry and immunity. *Immunol Res* 2015; **63**: 38-57 [PMID: 26512000 DOI: 10.1007/s12026-015-8718-1]
- 103 **Pavlov VA**, Ochani M, Gallowitsch-Puerta M, Ochani K, Huston JM, Czura CJ, Al-Abed Y, Tracey KJ. Central muscarinic cholinergic regulation of the systemic inflammatory response during endotoxemia. *Proc Natl Acad Sci U S A* 2006; **103**: 5219-5223 [PMID: 16549778 DOI: 10.1073/pnas.0600506103]
- 104 **Munyaka P**, Rabbi MF, Pavlov VA, Tracey KJ, Khafipour E, Ghia JE. Central muscarinic cholinergic activation alters interaction between splenic dendritic cell and CD4+CD25- T cells in experimental colitis. *PLoS One* 2014; **9**: e109272 [PMID: 25295619 DOI: 10.1371/journal.pone.0109272]
- 105 **Lee ST**, Chu K, Jung KH, Kang KM, Kim JH, Bahn JJ, Jeon D, Kim M, Lee SK, Roh JK. Cholinergic anti-inflammatory pathway in intracerebral hemorrhage. *Brain Res* 2010; **1309**: 164-171 [PMID: 19900419 DOI: 10.1016/j.brainres.2009.10.076]
- 106 **Pavlov VA**, Parrish WR, Rosas-Ballina M, Ochani M, Puerta M, Ochani K, Chavan S, Al-Abed Y, Tracey KJ. Brain acetylcholinesterase activity controls systemic cytokine levels through the cholinergic anti-inflammatory pathway. *Brain Behav Immun* 2009; **23**: 41-45 [PMID: 18639629 DOI: 10.1016/j.bbi.2008.06.011]
- 107 **Ji H**, Rabbi MF, Labis B, Pavlov VA, Tracey KJ, Ghia JE. Central cholinergic activation of a vagus nerve-to-spleen circuit alleviates experimental colitis. *Mucosal Immunol* 2014; **7**: 335-347 [PMID: 23881354 DOI: 10.1038/mi.2013.52]
- 108 **Pham GS**, Wang LA, Mathis KW. Pharmacological potentiation of the efferent vagus nerve attenuates blood pressure and renal injury in a murine model of systemic lupus erythematosus. *Am J Physiol Regul Integr Comp Physiol* 2018; **315**: R1261-R1271 [PMID: 30332305 DOI: 10.1152/ajpregu.00362.2017]
- 109 **Lehner KR**, Silverman HA, Addorisio ME, Roy A, Al-Onaizi MA, Levine Y, Olofsson PS, Chavan SS, Gros R, Nathanson NM, Al-Abed Y, Metz CN, Prado VF, Prado MAM, Tracey KJ, Pavlov VA. Forebrain Cholinergic Signaling Regulates Innate Immune Responses and Inflammation. *Front Immunol* 2019; **10**: 585 [PMID: 31024522 DOI: 10.3389/fimmu.2019.00585]
- 110 **Brégeon J**, Coron E, Da Silva AC, Jaulin J, Aubert P, Chevalier J, Vergnolle N, Meurette G, Neunlist M. Sacral nerve

- stimulation enhances early intestinal mucosal repair following mucosal injury in a pig model. *J Physiol* 2016; **594**: 4309-4323 [PMID: 26939757 DOI: 10.1113/JP271783]
- 111 **Tu L**, Gharibani P, Zhang N, Yin J, Chen JD. Anti-inflammatory effects of sacral nerve stimulation: a novel spinal afferent and vagal efferent pathway. *Am J Physiol Gastrointest Liver Physiol* 2020; **318**: G624-G634 [PMID: 32068444 DOI: 10.1152/ajpgi.00330.2019]
- 112 **Jin H**, Guo J, Liu J, Lyu B, Foreman RD, Yin J, Shi Z, Chen JDZ. Anti-inflammatory effects and mechanisms of vagal nerve stimulation combined with electroacupuncture in a rodent model of TNBS-induced colitis. *Am J Physiol Gastrointest Liver Physiol* 2017; **313**: G192-G202 [PMID: 28546285 DOI: 10.1152/ajpgi.00254.2016]
- 113 **Pauwels N**, Willemsse C, Hellemans S, Komen N, Van den Broeck S, Roenen J, Van Aggelpoel T, De Schepper H. The role of neuromodulation in chronic functional constipation: a systematic review. *Acta Gastroenterol Belg* 2021; **84**: 467-476 [PMID: 34599572 DOI: 10.51821/84.3.012]
- 114 **Thaha MA**, Abukar AA, Thin NN, Ramsanahie A, Knowles CH. Sacral nerve stimulation for faecal incontinence and constipation in adults. *Cochrane Database Syst Rev* 2015; CD004464 [PMID: 26299888 DOI: 10.1002/14651858.cd004464.pub3]
- 115 **Weledji EP**. A historical perspective on sacral nerve stimulation (sns) for bowel dysfunction. 2021. [cited 2021 Sep 19]. Available from: <https://www.preprints.org/manuscript/202108.0404/v1> [DOI: 10.20944/PREPRINTS202108.0404.V1]
- 116 **Gupta P**, Ehlert MJ, Sirls LT, Peters KM. Percutaneous tibial nerve stimulation and sacral neuromodulation: an update. *Curr Urol Rep* 2015; **16**: 4 [PMID: 25630918 DOI: 10.1007/s11934-014-0479-1]
- 117 **Marti L**, Galata C, Beutner U, Hetzer F, Pipitone N, Wolff K, Borovicka J, Brunner W, Sulz MC, Maurus C. Percutaneous tibial nerve stimulation (pTNS): success rate and the role of rectal capacity. *Int J Colorectal Dis* 2017; **32**: 789-796 [PMID: 28391449 DOI: 10.1007/s00384-017-2804-x]
- 118 **Iyer S**, Laus K, Rugino A, Botros C, Lozo S, Botros SM, Goldberg R, Tomezsko J, Gafni-Kane A, Wroblewski K, Sand P. Subjective and objective responses to PTNS and predictors for success: a retrospective cohort study of percutaneous tibial nerve stimulation for overactive bladder. *Int Urogynecol J* 2019; **30**: 1253-1259 [PMID: 30467763 DOI: 10.1007/s00192-018-3822-0]
- 119 **Furtado-Albانهزي D**, Jürgensen SP, Avila MA, Correia GN, Driusso P. Effects of two nonpharmacological treatments on the sleep quality of women with nocturia: a randomized controlled clinical trial. *Int Urogynecol J* 2019; **30**: 279-286 [PMID: 29508046 DOI: 10.1007/s00192-018-3584-8]
- 120 **Spinal Cord Stimulation Cincinnati**. OH Mayfield Brain & Spine. [cited 2021 Sep 19]. Available from: <https://mayfieldclinic.com/pe-stim.htm>
- 121 **Levine AB**, Parrent AG, MacDougall KW. Stimulation of the Spinal Cord and Dorsal Nerve Roots for Chronic Groin, Pelvic, and Abdominal Pain. *Pain Physician* 2016; **19**: 405-412 [PMID: 27454271]
- 122 **Lind G**, Winter J, Linderöth B, Hellström PM. Therapeutic value of spinal cord stimulation in irritable bowel syndrome: a randomized crossover pilot study. *Am J Physiol Regul Integr Comp Physiol* 2015; **308**: R887-R894 [PMID: 25786486 DOI: 10.1152/ajpregu.00022.2015]
- 123 **Eldabe S**, Buchser E, Duarte RV. Complications of Spinal Cord Stimulation and Peripheral Nerve Stimulation Techniques: A Review of the Literature. *Pain Med* 2016; **17**: 325-336 [PMID: 26814260 DOI: 10.1152/ajpregu.00022.2015]



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