

Basic Study

Helicobacter pylori inhibits the cleavage of TRAF1 via a CagA-dependent mechanism

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Abstract

AIM

To study the impact on cleavage of tumor necrosis factor receptor-associated factor 1 (TRAF1) regulated by *Helicobacter pylori* (*H. pylori*).

METHODS

Cleavage of TRAF1 was detected by western blotting in the human gastric cancer cell line AGS following treatment with an apoptosis inducer. Cleavage of TRAF1 mediated by caspase was examined *in vitro* using specific caspase inhibitors. The effect of the COOH-terminal TRAF1 fragment on gastric cell apoptosis during *H. pylori* infection was measured using flow cytometry. The impact of *H. pylori* infection on TRAF1 cleavage was detected in the presence of apoptosis inducer. The roles of *H. pylori* virulence factors that may regulate TRAF1 cleavage were analyzed using isogenic *cagA*-, *vacA*- and *cagE*-null mutants.

RESULTS

TRAF1 was found to be cleaved in AGS cells treated with the apoptosis inducer, and caspase-8 was the major caspase involved in the cleavage of TRAF1. The COOH-terminal TRAF1 fragment significantly induced cell apoptosis ($P < 0.05$) as well as promoted *H. pylori*-induced cell apoptosis ($P < 0.05$). *H. pylori* infection was found to significantly inhibit the cleavage of TRAF1 and to inhibit the activation of caspase-8 in the

presence of the apoptosis inducer at specific infection times and different cell/bacteria ratios. We also found that the effects of *cagE*- and *cagA*-null mutants on the inhibition of TRAF1 cleavage and activation of caspase-8 were significantly attenuated, compared with wild-type *H. pylori*, in the presence of the apoptosis inducer, showing that the virulence factor CagA was mainly involved in the inhibition of TRAF1 cleavage.

CONCLUSION

H. pylori infection significantly inhibits the cleavage of TRAF1 *via* a CagA-dependent mechanism, which would increase the relative amounts of full-length TRAF1 and exert an antiapoptotic effect on *H. pylori*-infected cells.

Key words: *Helicobacter pylori*; Tumor necrosis factor receptor-associated factor 1; CagA; Cleavage; Apoptosis

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Core tip: We report our first results from a study of *Helicobacter pylori* (*H. pylori*)-mediated tumor necrosis factor receptor-associated factor 1 (TRAF1) cleavage. The impact of *H. pylori* infection and its virulence factors on TRAF1 cleavage were detected in the presence of an apoptosis inducer. This study demonstrates, for the first time, that the cleavage of TRAF1 and the activation of caspase-8 were significantly inhibited by *H. pylori* infection in the presence of an apoptosis inducer. In addition, the virulence factor CagA was mainly involved in the inhibition of TRAF1 cleavage.

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INTRODUCTION

Gastric cancer is one of the most common malignant tumors and the third leading cause of cancer-related deaths worldwide^[1]. *Helicobacter pylori* (*H. pylori*) is a gram-negative, spiral shaped pathogen that successfully colonizes human gastric mucosa and is a strong risk factor for chronic gastritis, peptic ulcer, mucosa-associated lymphoid tissue lymphoma and gastric cancer^[2-4]. *H. pylori* pathogenesis is determined largely by interactions between bacterial factors and host cells. The best-characterized virulence determinants of *H. pylori* are the cytotoxin-associated gene pathogenicity island (*cag* PAI) and the vacuolating cytotoxin A (*vacA*). The *cag* PAI encodes a type IV secretion system (T4SS), which is responsible for injecting virulence factors, such as CagA protein,

directly into host cells. Once injected into cells, the CagA protein will induce complex cell changes involving various host signaling pathways^[5,6]. Though numerous studies have identified the association between *H. pylori* infection and the development of gastric carcinoma, the mechanisms underlying the carcinogenic potential of *H. pylori* are still not completely understood.

TRAF1 is a member of the TRAF family and is characterized by its diverse biological functions, acting through direct or indirect interactions with multiple tumor necrosis factor receptor (TNFR) family members and intracellular proteins^[7]. Several studies have demonstrated that TRAF1 might exert an antiapoptotic role in lymphoma cells through regulation of the activation of NF- κ B^[8-10]. Wang *et al.*^[11] reported that TRAF1 expression was up-regulated in the human gastric mucosal samples infected with *H. pylori* in a clinical immunohistochemical analysis. Moreover, the up-regulation of TRAF1 was increased as the gastric disease progressed from chronic gastritis to gastric cancer.

Our previous study showed that the expression of TRAF1 is up-regulated by *H. pylori* infection in both gastric epithelial cells and mice. The up-regulation of TRAF1 inhibited cell apoptosis as well as increased the viability of infected cells, which suggested that TRAF1 is an important protein that contributes to the pathogenesis of *H. pylori*-related gastric cancer^[12]. Interestingly, several studies have shown that TRAF1 could be transformed into a pro-apoptotic form after cleavage by caspase-8 in the presence of Fas ligand or TNF- α -induced apoptosis^[13-15]. Caspase-8 cleaves TRAF1 into two fragments and overexpression of the COOH-terminal fragment could enhance Fas or TNF- α mediated apoptosis. However, whether the cleavage of TRAF1 could be influenced by *H. pylori* and the mechanisms are not known.

The aim of our work was to elucidate the effect on TRAF1 cleavage regulated by *H. pylori* and the roles of *H. pylori* virulence factors regulating TRAF1 cleavage. To gain a better understanding of the role of *H. pylori* infection related to TRAF1 cleavage, in the present study we detected the cleavage of TRAF1 in AGS cells co-cultured with *H. pylori* or sterile saline alone in the presence of an apoptosis inducer. We also analyzed the roles of *H. pylori* virulence factors that may regulate TRAF1 cleavage using isogenic *cagA*-, *vacA*- and *cagE*-null mutants.

MATERIALS AND METHODS

Cell line and *H. pylori* strains

The human gastric cancer cell line AGS was obtained from the Cell Bank of the Chinese Academy of Sciences (Shanghai, China). AGS cells were grown in F-12 medium (Invitrogen, United States) supplemented with 10% (vol/vol) fetal bovine serum and were cultured at 37 °C in a humidified 5% CO₂ atmosphere. *H. pylori*

strain NCTC11637, obtained from Chinese Center for Disease Control and Prevention (Beijing, China) was grown on Columbia agar plates with 5% sheep blood. The plates were incubated in a mixed atmosphere of 10% CO₂, 5% O₂, and 85% N₂, at 37 °C for 24 h, after which they were harvested by centrifugation at 1000 × *g* for 5 min at 4 °C, washed 3 times with sterile saline, resuspended in sterile saline and added to gastric cells as indicated. Isogenic *cagA*-, *cagE*- and *vacA*-null mutants of *H. pylori* strain NCTC11637 were constructed by insertional mutagenesis using homologous recombination and selected with 25 µg/mL kanamycin.

Antibodies and reagents

The following primary antibodies were used: mouse anti-HA tag from Zhongshan Golden Bridge Biotechnology (Beijing, China); rabbit anti-GAPDH from Bioworld Technology (United States); rabbit anti-caspase-8 from Beyotime Biotechnology (Jiangsu, China). The following secondary antibodies were used: peroxidase-conjugated goat anti-mouse IgG (H + L) and peroxidase-conjugated goat anti-rabbit IgG (H + L) from Zhongshan Golden Bridge Biotechnology.

The pharmacological inhibitors were as follows: caspase-8 inhibitor Z-IETD-FMK from BioVision (United States); pan caspase inhibitor Z-VAD-FMK and caspase-3 inhibitor Ac-DEVD-CHO from Beyotime Biotechnology. The apoptosis inducer cycloheximide (CHX) and TNF-α were purchased from BioVision and Peprotech (United States) respectively.

Plasmids and transfection

The plasmids pcDNA-TRAF1-HA, pcDNA-TRAF1(D163A)-HA, pcDNA-TRAF1-N-HA, pcDNA-TRAF1-C-HA were constructed by inserting the specific cDNA and HA-tag sequences into pcDNA3.1 (+) (conserved in our laboratory) respectively.

In brief, AGS cells were seeded in a 24-well plate at a density of 1 × 10⁵ cells/well and cultured for 24 h, with a target of 60%-80% confluency at the time of transfection. Cells were transfected with 0.5 µg of plasmids using jetPRIME Transfection Reagent (Polyplus, France) according to the manufacturer's protocol. After 24 h of transfection, cells were co-cultured with *H. pylori* for the indicated experiments.

Western blot analysis

Proteins were extracted from the cultured cells and homogenized in a lysis buffer containing a protease inhibitor cocktail (CWBiotech, Beijing, China) according to the instructions. Then, the proteins were separated on 12% SDS-PAGE gels and transferred to nitrocellulose membranes (GE Healthcare Life Sciences, United States). The membranes were blocked in phosphate-buffered saline plus Tween-20 (PBST) containing 5% skim milk powder for 2 h at 37 °C, and then incubated with the primary antibodies for 1.5 h

at 37 °C. Next, the membranes were incubated with peroxidase-conjugated secondary antibodies for 1 h at 37 °C. The membranes were treated with ECL reagents (Engreen Biosystem, Beijing, China) and visualized using a Minichemi Lane1D imager (Sagecreation, Beijing, China), according to the manufacturer's instructions.

Cell apoptosis analysis

Cell apoptosis was measured according to the instructions of the Annexin V-FITC Apoptosis Detection Kit purchased from Beyotime Biotechnology. Briefly, AGS cells were transfected with the respective plasmids for 24 h and followed by co-culture with *H. pylori* for 24 h. Then, the cells were collected, washed in sterile PBS, and resuspended at a density of 1 × 10⁶ cells/mL. After that, the cells were stained with Annexin V-FITC and propidium iodide (PI) for 10 min prior to the analysis using a FACScan flow cytometer (Bio-Rad, United States).

Statistical analysis

Statistical analyses in our paper were performed using the software GraphPad Prism 5. The results are presented as mean ± SD. Differences among groups were compared using variance analysis, and comparisons between two groups were performed using an unpaired Student's *t*-test. Differences were considered to be significant at *P* < 0.05.

RESULTS

TRAF1 is cleaved via caspase-8 in AGS cells infected with *H. pylori*

To confirm that TRAF1 could be cleaved under the condition of *H. pylori* infection, AGS cells transfected with HA-tagged TRAF1 were treated with the apoptosis inducer (0.3 µg/mL CHX and 80 ng/mL TNF-α) for 6 h, and then analyzed for the cleavage of TRAF1. Our results showed that TRAF1 could be significantly cleaved in AGS cells treated with the apoptosis inducer (Figure 1A). To confirm that the cleavage site of TRAF1 is located at aspartic acid 163 in the ¹⁶⁰LVED¹⁶³ motif, AGS cells were transfected with HA-tagged TRAF1 mutation (D163A) and treated with the apoptosis inducer. The result showed that the TRAF1 mutation (D163A) could not be cleaved by the apoptosis inducer, which suggested the cleavage site of TRAF1 is located at aspartic acid 163 (Figure 1B). To further confirm the cleavage of TRAF1 mediated by caspase-8, TRAF1 cleavage was examined *in vitro* using specific caspase inhibitors in the presence of the apoptosis inducer treatment or *H. pylori* infection. As shown in Figure 1C and 1D, TRAF1 cleavage was blocked completely by the addition of Z-VAD-FMK, a broad specificity inhibitor of caspases, and was mainly blocked by Z-IETD-FMK, a caspase-8-specific inhibitor. However, Z-YVAD-FMK, a caspase-3-specific inhibitor, had no apparent effect on TRAF1 cleavage. The results show caspase-8 is the

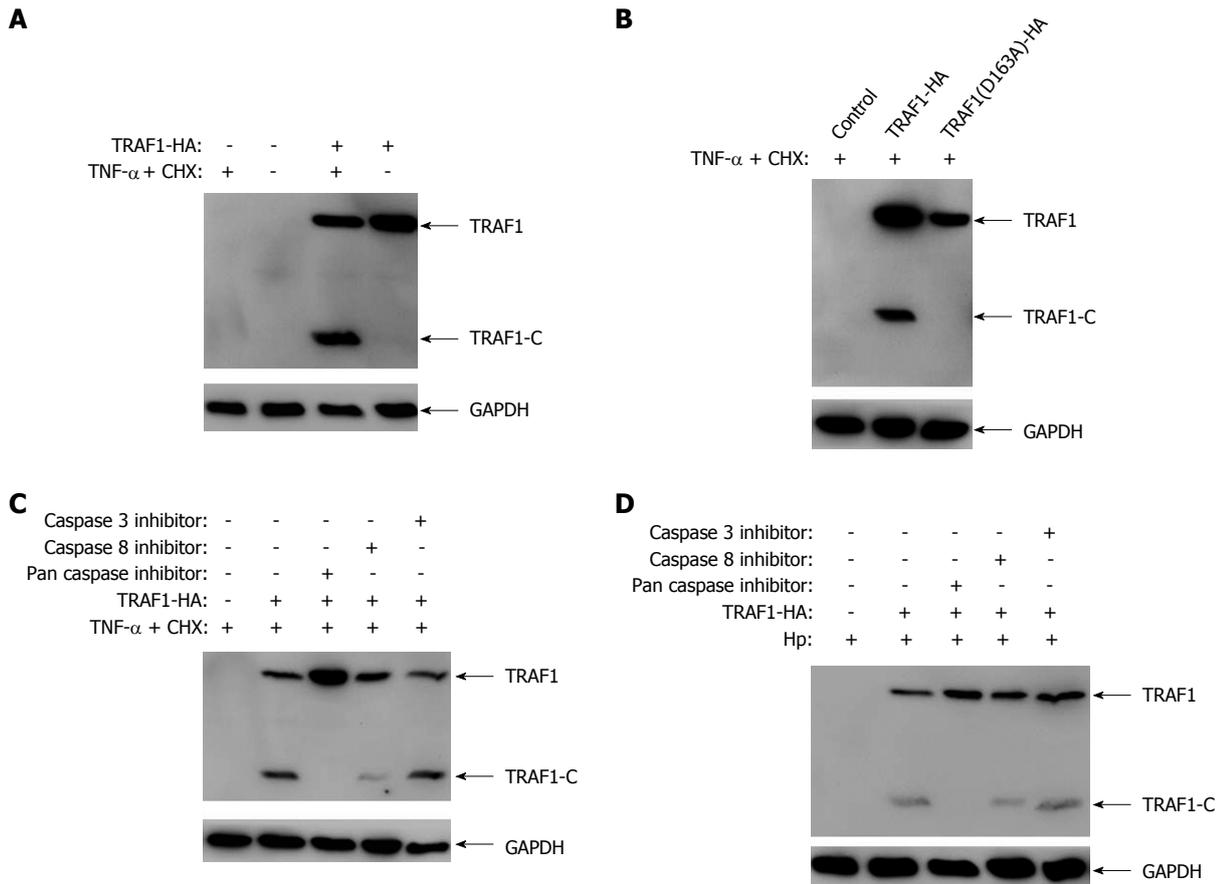


Figure 1 Tumor necrosis factor receptor-associated factor 1 is cleaved via caspase-8 in AGS cells infected with *Helicobacter pylori*. A: AGS cells transfected with either HA-tagged tumor necrosis factor receptor-associated factor 1 (TRAF1) or empty vector were treated with the apoptosis inducer (0.3 μ g/mL CHX and 80 ng/mL TNF- α) for 6 h, and the cleavage of TRAF1 was detected by western blotting; B: AGS cells were transfected with wild-type or D163A mutant of TRAF1 and analyzed for the cleavage of TRAF1 by western blotting; C: AGS cells transfected with the HA-tagged TRAF1 were pretreated with 20 μ mol/L of Z-VAD-FMK, Z-IETD-FMK, or Z-YVAD-FMK for 30 min, followed by treatment with the apoptosis inducer for 6 h and then analyzed for the cleavage of TRAF1 by western blotting; D: AGS cells transfected with the HA-tagged TRAF1 were pretreated with 20 μ mol/L of Z-VAD-FMK, Z-IETD-FMK, or Z-YVAD-FMK for 30 min, followed by infection with *Helicobacter pylori* strain NCTC11637 for 6 h and then analyzed for the cleavage of TRAF1 by western blotting.

major caspase involved in the cleavage of TRAF1 in cells infected with *H. pylori*.

COOH-terminal TRAF1 fragment promoted cell apoptosis in *H. pylori* infected cells

To confirm the pro-apoptotic role of the COOH-terminal TRAF1 fragment, AGS cells were transfected with HA-tagged TRAF1, TRAF1 (D163A), the NH₂-terminal TRAF1 (TRAF1-N), the COOH-terminal TRAF1 (TRAF1-C), or empty vector respectively and cell apoptosis was analyzed by flow cytometry. The results showed transfection with TRAF1, TRAF1 (D163A), or TRAF1-N had no significant effect on cell apoptosis compared with empty vector control, whereas transfection with TRAF1-C significantly promoted cell apoptosis (Figure 2A). In addition, to confirm the pro-apoptotic role of the COOH-terminal TRAF1 fragment in AGS cells infected with *H. pylori*, AGS cells were also transfected with the above five plasmids respectively and infected with *H. pylori* for 24 h and analyzed for cell apoptosis (Figure 2B). The results showed transfection with TRAF1-N has no significant

effect on cell apoptosis. Transfection with TRAF1 or TRAF1 (D163A) significantly inhibited *H. pylori*-induced apoptosis, whereas transfection with TRAF1-C significantly promoted *H. pylori*-induced apoptosis, thereby indicating the pro-apoptotic role of the COOH-terminal TRAF1 fragment in *H. pylori* infected cells.

H. pylori infection significantly inhibits the cleavage of TRAF1

To investigate the effect of *H. pylori* infection on TRAF1 cleavage in cells, the expression of TRAF1 and its cleavage were examined in AGS cells when co-cultured with *H. pylori* or sterile saline alone in the presence of the apoptosis inducer for the specific infection times and cell/bacteria ratios. The cleavage of TRAF1 in AGS cells decreased significantly 4 h after *H. pylori* infection compared with the uninfected cells (Figure 3A). The effect of cell/bacteria ratios (1:10-1:500) on TRAF1 cleavage was then tested. A significantly decreased cleavage was observed at a ratio of 1:50, and the cleavage of TRAF1 gradually decreased as the bacteria amounts increased (Figure 3B). As caspase-8 is the

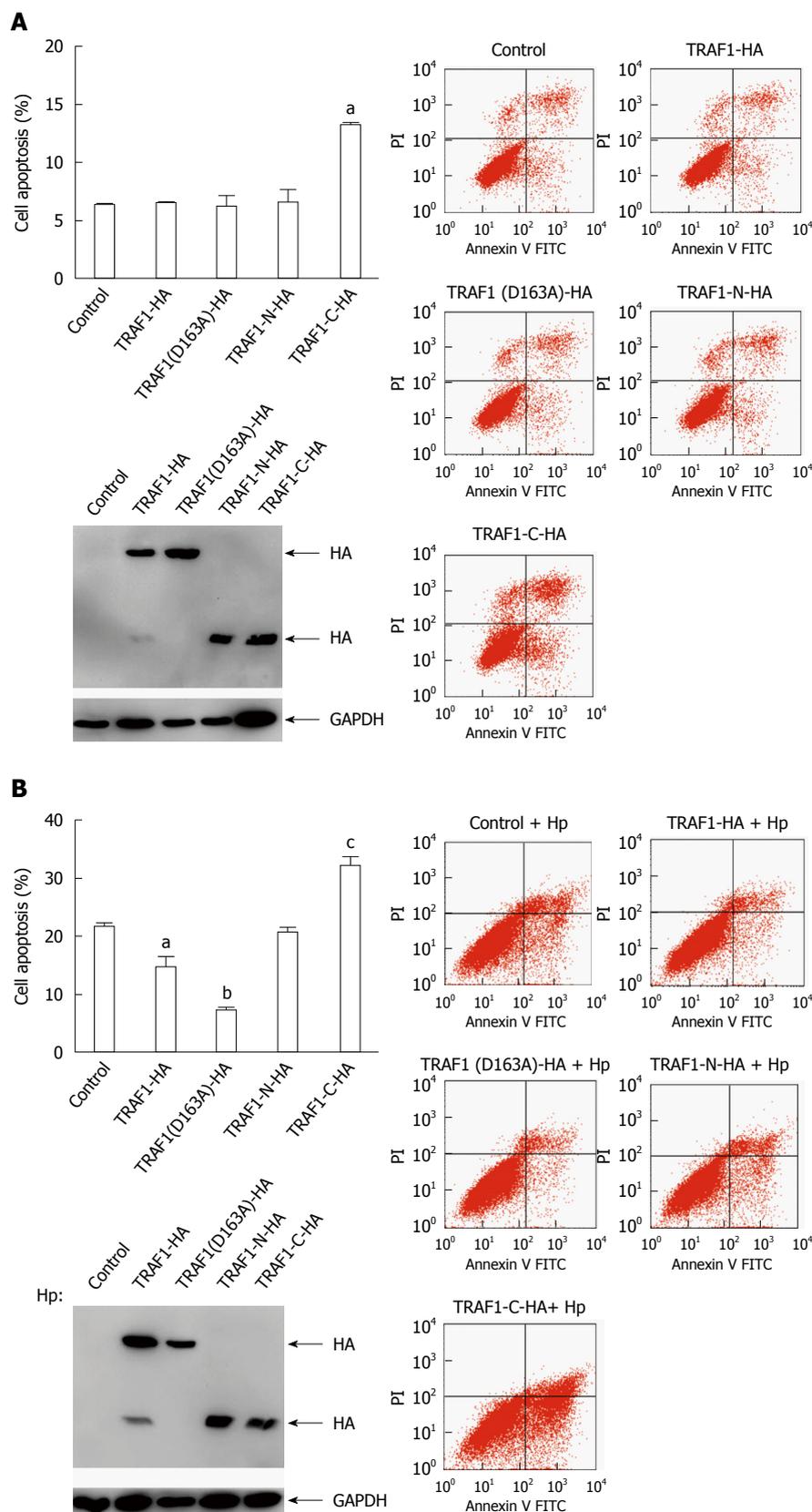


Figure 2 COOH-terminal tumor necrosis factor receptor-associated factor 1 fragment promoted cell apoptosis in *Helicobacter pylori* infected cells. A: AGS cells were transfected with HA-tagged tumor necrosis factor receptor-associated factor 1 (TRAF1), TRAF1(D163A), TRAF1-N, TRAF1-C for 24 h and then analyzed for cell apoptosis by flow cytometry using annexin V-FITC and propidium iodide staining (^a*P* < 0.05 vs control). The expression levels of these proteins in the analyzed cells were shown (Lower); B: AGS cells were transfected with HA-tagged TRAF1, TRAF1(D163A), TRAF1-N, TRAF1-C for 24 h respectively, followed by infection with *Helicobacter pylori* strain NCTC11637 for 24 h and then analyzed for cell apoptosis (^a*P* < 0.05 vs control, ^b*P* < 0.01 vs Control, ^c*P* < 0.05 vs control). The expression levels of these proteins in the analyzed cells are shown (Lower).

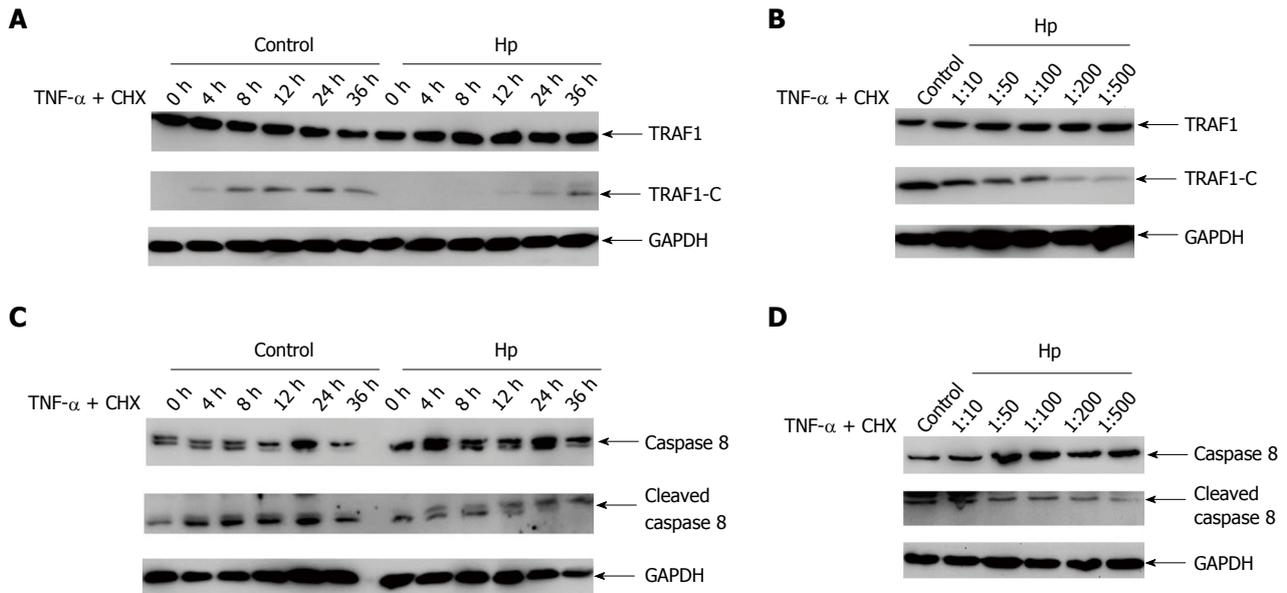


Figure 3 *Helicobacter pylori* inhibits the cleavage of tumor necrosis factor receptor-associated factor 1. A: AGS cells transfected with HA-tagged tumor necrosis factor receptor-associated factor 1 (TRAF1) were treated with the apoptosis inducer (0.3 μ g/mL CHX and 80 ng/mL TNF- α) and co-cultured with *Helicobacter pylori* (*H. pylori*) strain NCTC11637, or left untreated for the infection time indicated, and analyzed for the cleavage of TRAF1 by western blotting; B: AGS cells transfected with HA-tagged TRAF1 were treated with the apoptosis inducer and co-cultured with *H. pylori* strain NCTC11637, or left untreated, at the indicated cell/bacteria ratios and analyzed for the cleavage of TRAF1 by western blotting; C: The same as shown in (A), but analyzed for the uncleaved and cleaved caspase-8 by western blotting; D: The same as shown in (B), but analyzed for the uncleaved and cleaved caspase-8 by western blotting.

major caspase involved in the cleavage of TRAF1, we next analyzed the activation of caspase-8. As shown in Figure 3C and D, the cleaved caspase-8 was significantly decreased when co-cultured with *H. pylori* in the presence of the apoptosis inducer for the specific infection times as well as the indicated cell/bacteria ratios, suggesting that the activity of caspase-8 was inhibited. These results indicate that *H. pylori* infection significantly inhibits the cleavage of TRAF1.

CagA is the major virulence factor involved in inhibiting the cleavage of TRAF1

As CagA is a major virulence factor in *H. pylori*-related gastric pathogenesis, we next investigated whether CagA could affect TRAF1 cleavage. The isogenic *cagA*-, *vacA*- and *cagE*-null mutants were used. *H. pylori* lacking CagE cannot form an effective type IV secretion system and inhibits the translocation of CagA into cells, whereas knocking-out of *cagA* and *vacA* leads to no expression of the VacA and CagA virulence factors, respectively.

AGS cells were infected with the wild-type *H. pylori* or the isogenic *H. pylori* mutants in the presence of the apoptosis inducer, and analyzed for the TRAF1 cleavage. The results showed the TRAF1 cleavage significantly increased in 6 h and 12 h after infection with *cagE*- and *cagA*-null mutants compared with wild-type *H. pylori*, while the *vacA*-null mutant had no significant changes compared with wild-type *H. pylori* (Figure 4A). A significantly increased cleavage was also observed in *cagE*- and *cagA*-null mutants at the indicated cell/bacteria ratios (1:100-1:500) (Figure 4B). This suggested that the effect of *cagE*- and *cagA*-

null mutants on the inhibition of TRAF1 cleavage was significantly attenuated compared with wild-type *H. pylori* in the presence of the apoptosis inducer. We also analyzed the uncleaved and cleaved caspase-8 forms at the indicated infection time and cell/bacteria ratios. As shown in Figure 4C and D, the activity of caspase-8 significantly increased in the *cagE*- and *cagA*-null mutants compared with wild-type *H. pylori*. Thus, these results suggested that the virulence factor CagA plays an important role in the *H. pylori*-mediated inhibition of TRAF1 cleavage.

DISCUSSION

In the present study, we show the previously unidentified role of TRAF1 cleavage regulated by *H. pylori* infection. TRAF1 was found to be cleaved *via* caspase-8 in AGS cells treated with an apoptosis inducer, and the COOH-terminal TRAF1 fragment was found to promote *H. pylori*-induced cell apoptosis. However, *H. pylori* infection was found to inhibit the cleavage of TRAF1 as well as inhibit the activation of caspase-8 in the presence of the apoptosis inducer. We also found that the *H. pylori* virulence factor CagA is mainly involved in the *H. pylori*-mediated inhibition of TRAF1 cleavage.

TRAF1 is a member of the TRAF family and is dysregulated in various diseases, such as atheroma, lymphoma, and solid tumors^[16-19]. Some studies suggested that TRAF1 plays an antiapoptotic role^[8,9] and could suppress TNFR-induced cell apoptosis^[20,21]. Immunohistochemistry of gastric mucosal specimens from *H. pylori*-positive patients with chronic gastritis (CG), intestinal metaplasia (IM), dysplasia, and

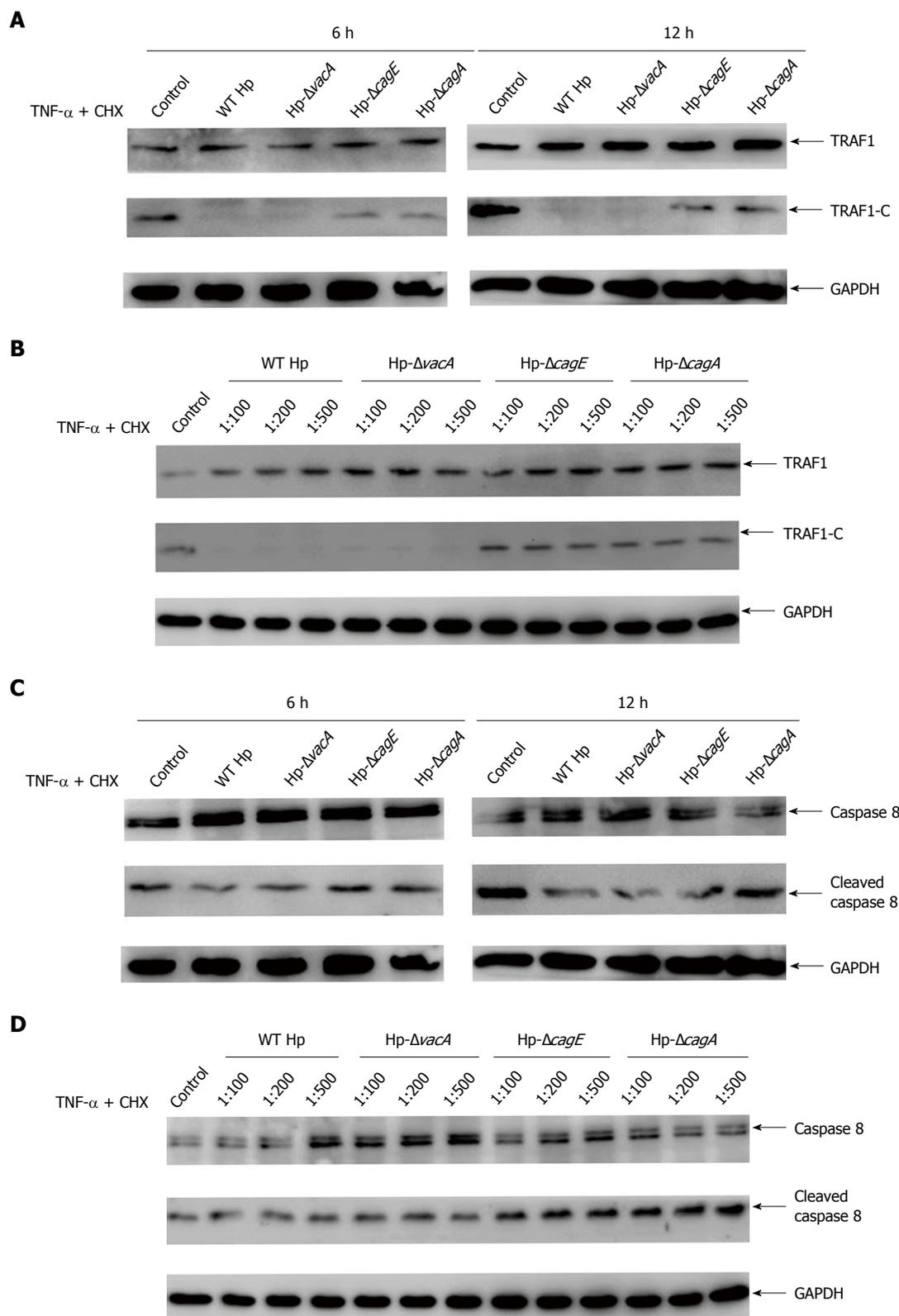


Figure 4 CagA is the major factor involved in inhibiting the cleavage of tumor necrosis factor receptor-associated factor 1. A: AGS cells transfected with tumor necrosis factor receptor-associated factor 1 (TRAF1) were treated with the apoptosis inducer (0.3 μ g/mL CHX and 80 ng/mL TNF- α), and co-cultured with *Helicobacter pylori* (*H. pylori*) strain NCTC11637 or its isogenic *vacA*-, *cagE*- or *cagA*-null mutants, and analyzed for the cleavage of TRAF1 at 6 h and 12 h by western blotting; B: AGS cells transfected with TRAF1 were treated with the apoptosis inducer, and co-cultured with *H. pylori* strain NCTC11637, or its isogenic *vacA*-, *cagE*- or *cagA*-null mutants at the indicated cell/bacteria ratios, and analyzed for the cleavage of TRAF1 by western blotting; C: The same as shown in (A), but analyzed for uncleaved and cleaved caspase-8 by western blotting; D: The same as shown in (B), but analyzed for uncleaved and cleaved caspase-8 by western blotting.

gastric carcinoma (GC) showed that positive TRAF1 expression was detected in 34.8%, 53.3%, 72.7%, and 88.9% specimens of CG, IM, dysplasia, and GC, respectively, suggesting that TRAF1 may be involved in *H. pylori*-induced gastric carcinogenesis^[11]. Our previous study showed that the up-regulation of TRAF1 could be induced by *H. pylori* in both gastric epithelial cells and mice. The up-regulation of TRAF1 inhibited cell apoptosis and increased the viability of gastric epithelial cells infected with *H. pylori*. We also determined that *H. pylori* infection significantly enhanced the formation of the antiapoptotic complex containing TRAF1, TRAF2, cIAP1, and cIAP2, which might play a major role in suppressing the activation of caspase-8 and in inhibiting cell apoptosis^[12].

Though the full-length TRAF1 protein exerts an anti-apoptotic role during *H. pylori* infection, TRAF1 could be converted into a pro-apoptotic protein after cleavage^[13-15]. So, it is important to explore the regulation of TRAF1 cleavage by *H. pylori*. In our study, we found the cleavage of TRAF1 as well as the activation of caspase-8 induced by the apoptosis inducer could be significantly inhibited by *H. pylori* infection. As the COOH-terminal TRAF1 fragment has a pro-apoptotic role, the inhibition of TRAF1 cleavage may further protect *H. pylori*-infected cells from apoptosis. Our results showed that the cleavage of TRAF1 decreased significantly after 4 h following *H. pylori* infection and the cleavage of TRAF1 gradually decreased as the bacteria amounts increased, which suggests the inhibition of TRAF1 cleavage is related to the infection time and the bacterial amounts.

Another important finding is that the inhibition of TRAF1 cleavage was mainly dependent on the *H. pylori* virulence factor CagA. Our results suggested that the effects of *cagE*- and *cagA*-null mutants on the inhibition of TRAF1 cleavage were significantly attenuated compared with wild-type *H. pylori* or the *vacA*-null mutant in the presence of the apoptosis inducer. However, the mechanisms are not completely understood. It is possible that CagA-positive *H. pylori* up-regulates the expression of TRAF1, TRAF2, cIAP1, and cIAP2, which can increase formation of the antiapoptotic complex to suppress caspase-8 activation and then inhibit TRAF1 cleavage. It is also possible that *H. pylori* inhibits caspase-8 activation via other signaling pathways mediated by activation of NF- κ B, thereby inhibiting TRAF1 cleavage. NF- κ B is a major transcription regulator involved in inflammation, cell proliferation, and apoptosis, which are considered to be anti-apoptotic and to have an oncogenic role^[22]. *H. pylori* infection could activate NF- κ B through host-bacterial interactions via multiple signaling pathways^[23,24]. Among these pathways, CagA is the major bacterial component to induce activation of NF- κ B through interaction with multiple signaling regulators, such as receptor tyrosine kinase c-Met, guanine exchange factor α -Pix, and Ras, thereby leading to activation of phosphatidylinositol-3-kinase (PI3K)/Akt kinase, transforming growth factor beta-

activated kinase 1 (TAK1), and MAPK/ERK signaling pathways^[25-28].

In summary, our study suggested that during its evolutionary adaptation to the host gastric environment and to counteract detrimental conditions, *H. pylori* infection induced the up-regulation of TRAF1 as well as inhibited its cleavage. Therefore, the dysregulation and increase of TRAF1 may be advantageous for *H. pylori* by allowing this pathogen long-term habitation in the gastric mucosa without triggering cell apoptosis. However, this change may increase the risk of gastric carcinoma.

ACKNOWLEDGMENTS

We would like to express our great appreciation to Weiwei Liu for his statistical review of our data.

COMMENTS

Background

Our previous study showed that tumor necrosis factor receptor-associated factor 1 (TRAF1) is up-regulated by *Helicobacter pylori* (*H. pylori*) infection in both gastric epithelial cells and mice. The up-regulation of TRAF1 has been shown to inhibit cell apoptosis and to increase the viability of infected cells, suggesting that TRAF1 may be an important molecule during the pathogenesis of *H. pylori*-related gastric cancer. Interestingly, several studies have shown that TRAF1 can be converted into a pro-apoptotic protein after cleavage by caspase-8 in the presence of Fas ligand or TNF- α -induced apoptosis. However, whether the cleavage of TRAF1 could be influenced by *H. pylori* infection and the related mechanism are not known.

Research frontiers

Gastric cancer is the fifth most common malignancy and the third leading cause of cancer-related deaths worldwide. Though numerous studies have identified an association between *H. pylori* infection and the development of gastric carcinoma, the mechanisms underlying the carcinogenic potential of *H. pylori* are still not completely understood.

Innovations and breakthroughs

The authors report our first results from a study of *H. pylori*-mediated TRAF1 cleavage. This study demonstrates for the first time that the cleavage of TRAF1 and the activation of caspase-8 were significantly inhibited by *H. pylori* infection in the presence of an apoptosis inducer. In addition, the virulence factor CagA was found to be mainly involved in the inhibition of TRAF1 cleavage.

Applications

This results suggest that, during its evolutionary adaptation to the host environment and to counteract detrimental conditions, *H. pylori* infection induced the up-regulation of TRAF1 as well as inhibited its cleavage. The up-regulation of TRAF1 inhibited cell apoptosis and increased the viability of infected cells, suggesting that TRAF1 may be a potential target for prevention and diagnosis during the pathogenesis of gastric cancer related to *H. pylori* infection.

Terminology

Cytotoxin-associated gene A (CagA), an important virulence factor and the only bacterial oncoprotein, is delivered into gastric epithelial cells via type IV secretion of *H. pylori*. Upon delivery, CagA perturbs multiple host signaling pathways by interacting with the host signaling molecules, resulting in cytopathic effects and subsequent cell transformation.

Peer-review

The following are comments and questions to authors. I hope to take

consideration for further improvement: (1) the detailed characteristics of AGS cell and the reason why this cell line was selected needs to be explained; (2) the authors revealed TRAF1 was cleaved by an apoptosis inducer and it was attenuated by a caspase-8 inhibitor. Do the authors have data that caspase-8 itself cleaved TRAF1? (3) for easy understanding, a correlation diagram in *H. pylori* and TRAF1 cleavage should be presented.

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