

# World Journal of *Hepatology*

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## Alkaline sphingomyelinase (NPP7) in hepatobiliary diseases: A field that needs to be closely studied

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### Abstract

Alkaline sphingomyelinase cleaves phosphocholine from sphingomyelin, platelet-activating factor, lysophosphatidylcholine, and less effectively phosphatidyl-

choline. The enzyme shares no structure similarities with acid or neutral sphingomyelinase but belongs to ecto-nucleotide pyrophosphatase/phosphodiesterase (NPP) family and therefore is also called NPP7 nowadays. The enzyme is expressed in the intestinal mucosa in many species and additionally in human liver. The enzyme in the intestinal tract has been extensively studied but not that in human liver. Studies on intestinal alkaline sphingomyelinase show that it inhibits colonic tumorigenesis and inflammation, hydrolyses dietary sphingomyelin, and stimulates cholesterol absorption. The review aims to summarize the current knowledge on liver alkaline sphingomyelinase in human and strengthen the necessity for close study on this unique human enzyme in hepatobiliary diseases.

**Key words:** Sphingomyelin; Alkaline sphingomyelinase; Nucleotide pyrophosphatase/phosphodiesterase 7; Autotaxin; Platelet-activating factor; Cholangiocarcinoma; Liver diseases; Gallstone

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**Core tip:** Alkaline sphingomyelinase is an enzyme expressing in the intestinal tract and additionally human liver. It hydrolyzes sphingomyelin, platelet activating factor and lysophospholipase. In the intestinal tract, it digests dietary sphingomyelin, stimulates cholesterol absorption, and inhibits development of colon cancer. Less is known about the implications of the enzyme in liver diseases. The review summarizes the current knowledge of its roles in hepatobiliary disease and raised special topics for future investigations.

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## ALK-SMASE IN NPP FAMILY

An enzyme that catalyzes sphingomyelin (SM) hydrolysis to ceramide at optimal alkaline pH was first identified in the intestinal tract by Nilsson in 1969<sup>[1]</sup>. The enzyme was thereafter named as alkaline sphingomyelinase (alk-SMase)<sup>[2]</sup> in line with acid and neutral SMases. However, after purification, characterization, and gene cloning<sup>[3-6]</sup>, it was found that alk-SMase actually had no structural similarities with either acid or neutral SMase, but shared about 30% amino acid sequence similarities with enzymes in ecto nucleotide pyrophosphatase/phosphodiesterase (NPP) family. As a novel member in NPP family, alk-SMase is nowadays also called NPP7.

Comparing with other six NPP members, alk-SMase is distinctive in several aspects. Not like most NPP members, alk-SMase has no nucleotidase activity but a phospholipase C activity. It cleaves phosphocholine from phospholipids including SM, platelet activating factor (PAF)<sup>[7]</sup>, lysophosphatidylcholine (lyso-PC), and phosphatidylcholine (PC) less effectively<sup>[5]</sup>. In NPP family, NPP2 (autotaxin) can also hydrolyze lyso-PC, but with a phospholipase D activity<sup>[8]</sup>. Another NPP member NPP6 can cleave phosphocholine from lysophospholipids mainly lyso-PC and lyso-PAF but not from SM<sup>[9]</sup>. Alk-SMase is so far the only one that has decent activities against SM and PAF in this family. In addition, while the activities of other NPP members could be identified in many organs and tissues, expression of alk-SMase is only restricted to intestinal tract in most species<sup>[10]</sup>. Western blot of rat tissues only shows positive band in intestinal mucosa and content but not in other organs including brain, heart, lung, liver, spleen, kidney and pancreas<sup>[4]</sup>. Interestingly, additional high activity was found in the bile of human<sup>[11]</sup>, but not in bile of other species including rat, mouse, pig, cow, sheep, dog, guinea pig, and baboon<sup>[10]</sup> (and unpublished data). Furthermore, most NPP members are functioning as a proliferative and inflammatory factors that are important for cell survival<sup>[12]</sup>, alk-SMase displays inhibitory effects on cell proliferation and inflammation<sup>[13,14]</sup>. Cell culture studies show that alk-SMase can inhibit cell proliferation by about 50%<sup>[15]</sup> and its activity *in vivo* is positively correlated to the activity of caspase 3, the key enzyme that triggers apoptosis<sup>[16-18]</sup>. Rectal administration of alk-SMase in rats suppresses colitis induced by dextran sulfate sodium (DSS)<sup>[19]</sup>. Recently the studies with alk-SMase knockout mice clearly showed that both initiation and malignant transformation of colon cancer induced by azoxymethane and DSS was enhanced by about 5 times in the knockout mice comparing with the wild type mice<sup>[20]</sup>. In agreement with the animal studies, clinical studies also found reduced alk-SMase activity in patients with inflammatory bowel diseases (IBD) and colon cancer, and the reduction is progressive from 25% in IBD to 75% in colonic carcinoma<sup>[21-23]</sup>.

The anticancer effects of alk-SMase are thought to be achieved with a three armed mechanism<sup>[13]</sup>. First,

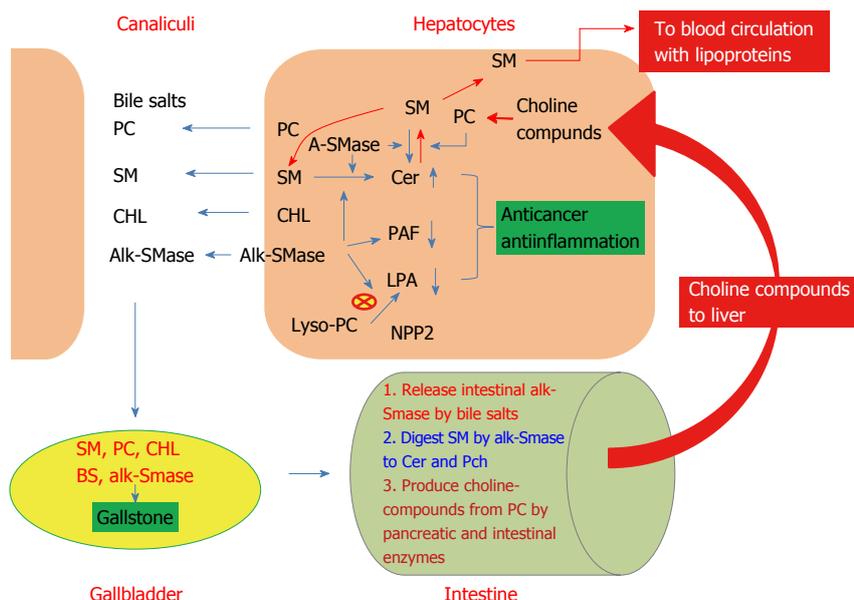
it hydrolyzes SM to ceramide, which is a well-known antiproliferative and apoptotic molecule<sup>[18,24]</sup>. Second, it cleaves phosphocholine moiety from PAF and inactivates PAF<sup>[7]</sup>, which is widely expressed in many inflammatory tissues promoting inflammation and tumorigenesis<sup>[25]</sup>. And finally NPP7 converts lyso-PC to monoacylglycerol<sup>[5]</sup>, thus reducing the production of lysophosphatidic acid (LPA), which otherwise can be formed by NPP2<sup>[8]</sup>. LPA has emerged as an important messenger with potent inflammatory and carcinogenic effects mediated *via* several signaling transduction pathways after binding to G protein coupled receptors<sup>[26,27]</sup>. In supporting this three arm hypothesis, decreased ceramide and increased PAF<sup>[20]</sup> and LPA (Zhang P *et al* Abstract presented in AACR symposium, Shanghai, China, 2016) have been found in NPP7 knockout mice.

Similar to other NPP members, alk-SMase is anchored on the surface of the cell membrane with a short hydrophobic domain. The remaining part of the enzyme including the catalytic domain is exposed extracellularly<sup>[13]</sup>. The enzyme can be released by bile salt<sup>[28]</sup>, and also by pancreatic trypsin, as there is a tryptic site just above the hydrophobic domain embedded inside the membrane<sup>[29]</sup>.

## ALK-SMASE IN HUMAN BILE

Alk-SMase in human bile was discovered by Nyberg *et al*<sup>[11]</sup> in bile collected from patients in our hospital. The activity in human bile is not derived from bacteria since it is similarly present in the samples with and without bacterial infection. Although gallbladder bile has higher activity than the hepatic bile, no activity was found in the homogenates of gallbladder mucosa, confirming that it is liver not gallbladder that expresses the enzyme. Because PCR experiment identifies alk-SMase mRNA in human HepG2 cells<sup>[30]</sup>, the enzyme is believed to be expressed by hepatocytes, transported to the surface of the microvilli that extend into the bile canaliculi and released by bile salt into the lumen.

Alk-SMase in human bile shares similar characteristics as the one in the intestinal mucosa<sup>[31,32]</sup>. The enzyme becomes active at the pH around 7 and the maximal activity occurs at pH 9.0. Its activity requires the presence of bile salts<sup>[4,6]</sup>. The bile salt dependency is type specific, which differs from other lipases such as bile salt stimulated lipase<sup>[33,34]</sup>. Although different bile salts more or less increase alk-SMase activity with the maximal effects at their critical micelle concentrations, taurocholate (TC) and taurochenodeoxycholate (TCDC) are much more effective than other bile salts. On the other hand, the nonionic detergent Triton X100 and zwitterionic non-denaturing detergent CHAPS with similar structure as TCDC and TC have no stimulatory effect but inhibit alk-SMase activity in the presence of other bile salts<sup>[4,31,35]</sup>. The finding indicates that the bile salt induced activation is not a simple detergent effect on the physical state of the substrate SM in mixed micelles. Additional interaction between the enzyme



**Figure 1 Metabolism of sphingomyelin in the liver and potential implications of alk-SMase in liver diseases.** Liver alk-SMase is localized on the hepatocyte canaliculi membrane. It hydrolyzes SM, PAF, and lyso-PC, resulting in increased ceramide (Cer) and decreased PAF and LPA, thus having anticancer and anti-inflammatory effects. Together with PC, cholesterol, and bile salts, SM is released in canaliculi and transported to gallbladder, where the interactions of these compounds affect gallstone formation. When the bile is delivered into the intestinal tract, bile salt will release additional alk-SMase from intestinal mucosa, and digest intestinal SM to ceramide and phosphocholine (Pch). Meanwhile, PC will be hydrolyzed by enzymes from pancreas and intestinal mucosa to choline compounds such as free choline, lyso-PC, and Pch. These choline compounds will be transported to liver where to be used for synthesis of PC. Pch moiety in PC can be transferred to ceramide to form SM by SM synthases. Part of the SM formed will be released into blood together with lipoproteins, part to bile, and part to be degraded by alk-SMase and acid SMase (ASMase) in the liver.

protein and the bile salt is likely involved. Supporting this hypothesis, recent studies on crystal structure of human alk-SMase by Gorelik *et al*<sup>[36]</sup> showed that the enzyme forms a hydrophobic loop and a positively charged surface which can interact with bile salts. This needs to be proved in further investigations.

Similar also to intestinal alk-SMase, human bile alk-SMase is inhibited by PC, the most abundant phospholipid in the bile<sup>[31,37]</sup>. This might be related to a competition between PC and SM for the substrate binding site of the enzyme. As shown by both computer homology modelling studies<sup>[38]</sup> and crystal structural studies<sup>[36]</sup>, alk-SMase forms a specific pocket and a long narrow groove that fits the phosphocholine head group, and the tails of these substrates, respectively. The binding affinity is stronger for SM than for PC<sup>[39,40]</sup>.

Presence of alk-SMase in human bile enhances SM digestion in humans. Intestinal SM is derived from diet, shedding mucosal cells and bile. The enzyme responsible for digesting SM in the gut is alk-SMase and in alk-SMase knockout mice, about 90% of ingested SM cannot be digested but accumulated in the colon<sup>[41]</sup>. In many species except human, alk-SMase activity is absent in duodenum, increasing in the jejunum and declining in the colon<sup>[2]</sup>. SM digestion in the gut normally starts in the middle of the jejunum where alk-SMase is high and PC, the major inhibitor of alk-SMase, has been decreased due to the absorption<sup>[42]</sup>. The process of SM digestion is slow and incomplete in most species, resulting in about 40% ceramide and SM being identified in feces<sup>[43,44]</sup>. However, due to the presence

of additional alk-SMase in the bile of human, human duodenum has considerable alk-SMase activity. Ohlsson *et al*<sup>[45]</sup> found that digestion of SM in human is more efficient than other species and about 81% of ingested SM can be digested. The more effective digestion of SM could be important for human health, as dietary SM stimulates the development of the gut of the new born and inhibits colonic tumorigenesis<sup>[46-48]</sup>.

## ROLE OF LIVER IN SPHINGOLIPID METABOLISM

It is well known that liver is an important organ for lipid metabolism such as fatty acid beta-oxidation, ketone body generation, cholesterol metabolism, lipoprotein synthesis, and phospholipid metabolism. For the phospholipid metabolism in liver, most previous studies focused on PC not SM, but the interest in SM was increasing in the latest decades. Liver is an organ with relatively high levels of SM. Comparing with subcutaneous and intra-abdominal adipose tissues, SM in human liver is 7-8 fold higher than in these adipose tissues<sup>[49]</sup>.

The high levels of PC and SM in the liver are attributable to the fact that liver efficiently takes up choline containing compounds such as choline, lyso-PC and phosphocholine derived from digestion of phospholipids in the intestinal tract and uses them for synthesis of PC and SM<sup>[42,50]</sup> (Figure 1). As shown in animal studies, after feeding choline labeled SM, up to 30% of the

labeled choline is accumulated in the liver and more than 95% of them is utilized for PC synthesis<sup>[42]</sup>. This is important for SM synthesis, as at the last step of SM synthesis, phosphocholine is transferred from PC to ceramide, catalyzed by SM synthase, which is highly expressed in the liver<sup>[51,52]</sup>. For the hydrolysis of SM, liver has high acid SMase activity than most other organs in many species<sup>[41]</sup>. Acid SMase is an enzyme with two isoforms. One is the lysosome enzyme that breaks down internalized SM in lysosome and the other is a secretory form that can be secreted to the plasma membrane and hydrolyzes membrane bound SM<sup>[53]</sup>. That is why in Niemann Pick diseases with acid SMase deficiency, liver is one of the most affected organs with SM accumulation<sup>[54]</sup>. There are many other factors that influence SM levels in the liver, such as high fat diet<sup>[55]</sup>, endotoxin infection<sup>[56]</sup>, hepatitis B virus infection<sup>[57]</sup> and liver cancer<sup>[58]</sup>.

SM synthesized in the liver can be released into plasma and bile (Figure 1). Comparing with other species, human plasma has at least two fold higher levels of SM than other species<sup>[59]</sup>. The plasma SM from liver is mainly transported with lipoproteins mainly VLDL and less with LDL and HDL<sup>[43]</sup>. SM in plasma is also derived from intestinal mucosa and other tissue cells, being about 1-1.5 g in total<sup>[43]</sup>. Most SM secreted from intestine is in chylomicron. SM in chylomicron is mainly not from the dietary products but from the membrane of enterocytes, because most sphingosine, the final digestion product of SM by alk-SMase and neutral ceramidase in the gut is not utilized to resynthesize SM after absorption in mucosal cells, but to convert to fatty acid and then to chyle triglyceride<sup>[44,60]</sup>.

Secretion of phospholipids from hepatocyte canalicular membrane is by a mechanism related to the interactions of bile salt and ABCB4<sup>[61]</sup>. Under physiological conditions, more than 95% of the phospholipids in bile is PC, and SM is accounted for about 3%<sup>[62]</sup>. The levels of SM in human bile is relatively low comparing with other species which don't have alk-SMase in the bile such as sheep<sup>[63]</sup>. But under pathological conditions, the level of SM in bile is subject to change. Barnwell *et al*<sup>[64]</sup> showed that *in vivo* perfusion of bile salts in rat significantly reduced PC content and meanwhile induced about 10 time increase of SM in the bile without obvious damage to the liver.

The implications of SM in plasma and bile are getting increasing interest. It has been known that high concentration of plasma SM is a risk factor for atherosclerosis<sup>[65]</sup>. Recent studies also found that plasma SM could be a biomarker for various liver diseases such as hepatitis, primary sclerosing cholangitis (PSC), and steatosis<sup>[57,66,67]</sup>, indicating SM metabolism in liver significantly contributes to plasma SM levels. SM in the bile may have important implications, as it has stronger van der Waal interactions with cholesterol than PC<sup>[40]</sup> and its physical properties of SM is affected by bile salt which may affect gallstone formation in gallbladder<sup>[68]</sup>.

## ALK-SMASE IN HEPATOBILIARY DISEASES

Comparing the extensive studies on intestinal alk-SMase, which showed its important roles in SM digestion, colon cancer prevention, and cholesterol absorption<sup>[13,14,20]</sup>, the progress of the research on human bile alk-SMase obviously lags behind. The main obstacle is lack of an animal model that expresses alk-SMase in the liver, and lack of a cell line that highly expresses alk-SMase, as the enzyme has already been downregulated in tumorigenesis. In a pilot study, we measured alk-SMase activity in 30 human liver biopsies and the results indicated a reduction of the enzyme activity in steatosis and PSC<sup>[30]</sup>. Recently we determined alk-SMase activity in 59 bile samples taken under ERCP and found significant reduction of alk-SMase activity in bile of patients with PSC and tumorigenic diseases, with the most remarkable reduction in cholangiocarcinoma<sup>[69]</sup>.

Besides the reduction of activity, an abnormal transcript of alk-SMase was identified in both liver cancer HepG2 cells and colon cancer HT29 cells, which is caused by a shift of RNA splice site at transcriptional level, resulting in exon 4 deletion<sup>[30,70]</sup>. The enzyme translated from this transcript is totally inactive, as 73 amino acids coded by exon 4 were absent, of which a histidine is critical for formation of the substrate binding site<sup>[70]</sup>. According to the size of their mRNA, the wild type and the mutant isoform have been called 1.4 kb and 1.2 kb form, respectively. These two forms were found in the bile of many of the 59 patients with different hepatic diseases<sup>[69]</sup>, but the activity of alk-SMase is positively correlated with the ratio of 1.4/1.2 kb form. Decrease in the 1.4 kb product and increase in the 1.2 kb product are likely associated with the development of cholangiocarcinoma. In the bile of one PSC and one cholangiocarcinoma patient, no alk-SMase activity and no 1.4 kb product but only high levels of 1.2 kb form were identified<sup>[69]</sup>.

On the other hand, hepatobiliary diseases may also affect the levels of alk-SMase and SM digestion in the intestine because bile diversion strongly reduced alk-SMase activity in the small intestinal content by 85% and in the feces by 68% in rat<sup>[71]</sup>. The changes are believed to be related to bile salts, which release the alk-SMase from the intestinal mucosa to gut lumen<sup>[28]</sup>.

## HUMAN HEPATIC ALK-SMASE IN PERSPECTIVES

Considering the results from previous studies on alk-SMase, it is predictable that human liver alk-SMase may also have important implications for hepatobiliary diseases. The following questions are worth close investigation: (1) Can the remarkable reduction of alk-SMase activity and the increase in the aberrant isoform

in the bile be warning signals for carcinogenesis in the liver, particularly cholangiocarcinoma? It is well known that cholangiocarcinoma is a disease lacking an early biomarker, and most patients are not curable at the time when the disease is diagnosed<sup>[72]</sup>. Making the situation worsen is the fact that the incidence of cholangiocarcinoma is increasing<sup>[73]</sup>. To find an early biomarker for cholangiocarcinoma is therefore a challenge for clinical doctors and medical researchers. Alk-SMase activity in bile is significantly decreased in cholangiocarcinoma to an extent greater than in other hepatic diseases associated with increased expression of the 1.2 kb isoform<sup>[69]</sup>. The reduction of alk-SMase activity seems already occurring in both bile and liver biopsies in PSC patients<sup>[31]</sup>. PSC is a major risk factor for cholangiocarcinoma and about 15% of PSC patients may finally develop this type of cancer<sup>[74]</sup>. Future studies in a relatively large scale are necessary to evaluate whether the changed activity and 1.2 kb isoform expression in PSC patients can be biomarker for cholangiocarcinoma. To follow these changes might be helpful for identifying the early carcinogenesis. In addition, it is worthwhile to point out that about 70% of PSC patients may have IBD, particularly ulcerative colitis<sup>[75,76]</sup>. Reduction of alk-SMase activity in chronic ulcerative colitis has been reported<sup>[23]</sup>;

(2) Are there a cross communication between NPP7 (alk-SMase) and NPP2 (autotaxin) in hepatobiliary diseases? NPP2 hydrolyzes lyso-PC with a phospholipase D activity and generating LPA, a potent inflammatory and proliferative factor<sup>[77]</sup>. Increased levels of NPP2 are a feature of many important hepatobiliary diseases such as PBC, PSC<sup>[78]</sup>, steatosis<sup>[79]</sup>, liver fibrosis<sup>[80]</sup>, hepatitis C<sup>[81]</sup> and liver cancer<sup>[82]</sup>. Alk-SMase shares the same substrate lyso-PC with NPP2 but it cleaves phosphocholine instead of choline and thus generates monoacylglycerol not LPA<sup>[5]</sup>. Alk-SMase therefore may counteract NPP2 and thus reduce the formation of LPA. Recently we did find that in alk-SMase knockout mice treated with DSS, the levels of LPA in the colonic mucosa is higher in the knockout mice than in the wild type mice (Zhang P *et al*, Abstract presented in AACR symposium, Shanghai, China, 2016). Interestingly, a recent cohort study showed that primary biliary cirrhosis (PBC) patients who did not respond to ursodeoxycholic acid (UDCA) treatment display higher NPP2 levels than the responders<sup>[78]</sup>. Changes of alk-SMase may be implicated in the results, as alk-SMase activity can be increased by UDCA in liver cells<sup>[17]</sup>. No response to UDCA in these PBC patients may indicate a failure to upregulate alk-SMase by UDCA in these patients, leading to increased levels of NPP2; (3) Can alk-SMase protect liver against noxious effects of PAF? PAF is a type of bioactive lipid and can be synthesized rapidly in various inflammatory tissues. After binding to its G protein coupled receptors, PAF triggers several signal transduction pathways leading to activation of various phospholipases including C, D and A2 and to calcium mobilization, MAP kinase activation, and neutrophil mobilization<sup>[83,84]</sup>. In the liver PAF induces vasoconstriction<sup>[85]</sup> and may play

a key role in several hepatic diseases such as CCl<sub>4</sub> induced cirrhosis, ethanol and acetaminophen induced liver injury, viral induced hepatitis, and hepatocellular carcinoma<sup>[86-89]</sup>. All these diseases are associated with increased formation of PAF levels and PAF receptor expression. To inhibit the effects of PAF, previous studies were focused on PAF acetyl hydrolase and PAF receptor antagonists<sup>[90]</sup>. PAF is a substrate for alk-SMase which inactivates PAF by degrading it to phosphocholine and alkyl acetyl glycerol<sup>[7]</sup>. The activity is bile salt dependent with optimal pH at 7.5, which well fits the niche of the hepatobiliary system<sup>[7]</sup>. In alk-SMase knockout mice, PAF has been found to be significantly increased in the intestinal lumen<sup>[20]</sup>. It is therefore worthwhile examining the impact of alk-SMase on PAF action in these hepatic diseases and whether upregulation of alk-SMase may counteract the effects of PAF and benefit the patients; (4) What is the role of human bile alk-SMase in regulating SM levels in bile? Alk-SMase affects SM levels in the cell membrane. Overexpression of alk-SMase in COS7 cells<sup>[5]</sup> and incubation of the cells with purified alk-SMase result in reduced SM in the cell membrane<sup>[13]</sup>. Alk-SMase in human bile most likely can do the same things and thus affecting SM levels both in the hepatocyte canaliculi and bile. Phospholipids particularly PC and SM affect crystallization of cholesterol which is a key event involved in gallstone formation<sup>[68]</sup>. SM levels in the bile can be increased in the presence of high concentrations of bile salt<sup>[64]</sup>, and be decreased in the presence of high levels of alk-SMase released from canaliculi. Considering the influence of the membrane SM on cholesterol translocation and synthesis<sup>[40]</sup>, and the more appreciable interaction of SM than PC with cholesterol<sup>[39,40]</sup>, the impact of bile alk-SMase on gallstone formation through regulating SM levels both in the canalicular membrane and bile might be also worthwhile for close investigation.

## CONCLUSION

Additional expression of alk-SMase in liver is unique for humans. As shown in the figure, by hydrolyzing its substrate SM, PAF, and Lyso-PC, alk-SMase generates anticancer and apoptotic molecule ceramide, reduces levels of PAF and LPA, which have been shown to be involved in a series of liver diseases including viral infection, steatosis, fibrosis, sclerosis, and tumorigenesis. Alk-SMase thus may play important roles in protecting the organ from these diseases. In addition, the enzyme is released into bile together with SM, PC, bile salt, and cholesterol and may interfere SM and PC levels and the physical-chemical interactions of these molecules in bile, thus affecting gallstone formation. Liver is an active organ for SM metabolism and for regulating plasma SM levels. Changed alk-SMase activity in bile and SM levels in plasma have been found in several hepatobiliary diseases, and such changes may have diagnostic and prognostic values. The contributions of alk-SMase, a unique human liver enzyme, for these changes need close investigation.

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