

# World Journal of *Gastroenterology*

*World J Gastroenterol* 2021 December 7; 27(45): 7739-7865



**EDITORIAL**

- 7739 Orosomuroid in liver diseases  
*Elpek GO*

**FRONTIER**

- 7748 Novel frontiers of agents for bowel cleansing for colonoscopy  
*Di Leo M, Iannone A, Arena M, Losurdo G, Palamara MA, Iabichino G, Consolo P, Rendina M, Luigiano C, Di Leo A*
- 7771 Chronic rejection after liver transplantation: Opening the Pandora's box  
*Angelico R, Sensi B, Manzia TM, Tisone G, Grassi G, Signorello A, Milana M, Lenci I, Baiocchi L*

**OPINION REVIEW**

- 7784 Humans have intestinal bacteria that degrade the plant cell walls in herbivores  
*Fujimori S*

**MINIREVIEWS**

- 7792 Gut microbiome in allogeneic hematopoietic stem cell transplantation and specific changes associated with acute graft *vs* host disease  
*Le Bastard Q, Chevallier P, Montassier E*

**ORIGINAL ARTICLE****Clinical and Translational Research**

- 7801 MicroRNAs expression influence in ulcerative colitis and Crohn's disease: A pilot study for the identification of diagnostic biomarkers  
*Quaglio AEV, Santaella FJ, Rodrigues MAM, Sasaki LY, Di Stasi LC*

**Observational Study**

- 7813 Multimodality management of gallbladder cancer can lead to a better outcome: Experience from a tertiary care oncology centre in North India  
*Goel S, Aggarwal A, Iqbal A, Talwar V, Mitra S, Singh S*
- 7831 In-hospital mortality of hepatorenal syndrome in the United States: Nationwide inpatient sample  
*Kaewput W, Thongprayoon C, Dumancas CY, Kanduri SR, Kovvuru K, Kaewput C, Pattharanitima P, Petnak T, Lertjitbanjong P, Boonpheng B, Wijarnpreecha K, Zabala Genovez JL, Vallabhajosyula S, Jadowiec CC, Qureshi F, Cheungpasitporn W*

**CASE REPORT**

- 7844** Clinical presentation of gastric Burkitt lymphoma presenting with paraplegia and acute pancreatitis: A case report

*Lin Y, Pan YH, Li MK, Zong XD, Pan XM, Tan SY, Guo YW*

**LETTER TO THE EDITOR**

- 7855** SARS-CoV-2 infection in people with pre-existing liver disease: Further research is warranted

*Verma HK, Bhaskar L*

- 7859** Therapeutic potentials of fasudil in liver fibrosis

*Xi Y, Xu PF*

- 7862** Diagnostic biomarkers for pancreatic cancer: An update

*Yang M, Zhang CY*

**ABOUT COVER**

Editorial Board Member of *World Journal of Gastroenterology*, Oscar Teramoto-Matsubara, MD, AGAF, FACG, FACP, Associate Specialist, Department of Gastroenterology, ABC Medical Center, Mexico City 11000, Mexico. [teramotomd@prodigy.net.mx](mailto:teramotomd@prodigy.net.mx)

**AIMS AND SCOPE**

The primary aim of *World Journal of Gastroenterology* (WJG, *World J Gastroenterol*) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

**INDEXING/ABSTRACTING**

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: *Ying-Yi Yuan*; Production Department Director: *Xiang Li*; Editorial Office Director: *Ze-Mao Gong*.

**NAME OF JOURNAL**

*World Journal of Gastroenterology*

**ISSN**

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

**LAUNCH DATE**

October 1, 1995

**FREQUENCY**

Weekly

**EDITORS-IN-CHIEF**

Andrzej S Tarnawski, Subrata Ghosh

**EDITORIAL BOARD MEMBERS**

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

**PUBLICATION DATE**

December 7, 2021

**COPYRIGHT**

© 2021 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION ETHICS**

<https://www.wjgnet.com/bpg/GerInfo/288>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Gut microbiome in allogeneic hematopoietic stem cell transplantation and specific changes associated with acute graft vs host disease

Quentin Le Bastard, Patrice Chevallier, Emmanuel Montassier

**ORCID number:** Quentin Le Bastard [0000-0001-6930-0720](https://orcid.org/0000-0001-6930-0720); Patrice Chevallier [0000-0003-3142-5581](https://orcid.org/0000-0003-3142-5581); Emmanuel Montassier [0000-0002-2313-1172](https://orcid.org/0000-0002-2313-1172).

**Author contributions:** Le Bastard Q, Chevallier P, and Montassier E have been involved equally and have read and approved the final manuscript; all authors meet the criteria for authorship established by the International Committee of Medical Journal Editors and verify the validity of the results reported.

**Conflict-of-interest statement:**

There is no conflict of interest associated with any of the senior author or other coauthors who contributed their efforts in this manuscript.

**Country/Territory of origin:** France

**Specialty type:** Gastroenterology and hepatology

**Provenance and peer review:**

Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review report's scientific quality classification**

Grade A (Excellent): 0  
Grade B (Very good): B, B

**Quentin Le Bastard, Emmanuel Montassier**, Department of Emergency Medicine, Nantes University Hospital, Nantes 44093, France

**Patrice Chevallier**, Department of Hematology, Nantes University Hospital, Nantes 44093, France

**Corresponding author:** Emmanuel Montassier, MD, PhD, Professor, Department of Emergency Medicine, Nantes University Hospital, 1 place Alexis Ricordeau, Nantes 44093, France. [emmanuel.montassier@chu-nantes.fr](mailto:emmanuel.montassier@chu-nantes.fr)

### Abstract

Allogeneic hematopoietic stem cell transplantation (aHSCT) is a standard validated therapy for patients suffering from malignant and nonmalignant hematological diseases. However, aHSCT procedures are limited by potentially life-threatening complications, and one of the most serious complications is acute graft-versus-host disease (GVHD). During the last decades, DNA sequencing technologies were used to investigate relationship between composition or function of the gut microbiome and disease states. Even if it remains unclear whether these microbiome alterations are causative or secondary to the presence of the disease, they may be useful for diagnosis, prevention and therapy in aHSCT recipients. Here, we summarized the most recent findings of the association between human gut microbiome changes and acute GVHD in patients receiving aHSCT.

**Key Words:** Gut microbiome; DNA sequencing technologies; Allogeneic hematopoietic stem cell transplantation; Transplants; Acute graft vs host disease; Biomarkers; Composition; Function

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** This review reports the compositional and functional changes in gut microbiome of allogeneic hematopoietic stem cell transplantation recipients associated with acute graft-versus-host disease that could serve a biomarker for diagnosis and

Grade C (Good): 0  
 Grade D (Fair): 0  
 Grade E (Poor): 0

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

**Received:** April 20, 2021

**Peer-review started:** April 20, 2021

**First decision:** July 27, 2021

**Revised:** August 5, 2021

**Accepted:** November 20, 2021

**Article in press:** November 20, 2021

**Published online:** December 7, 2021

**P-Reviewer:** Hegazy MAE, Sameer S

**S-Editor:** Fan JR

**L-Editor:** A

**P-Editor:** Fan JR



prevention in patients receiving allogeneic hematopoietic stem cell transplantation.

**Citation:** Le Bastard Q, Chevallier P, Montassier E. Gut microbiome in allogeneic hematopoietic stem cell transplantation and specific changes associated with acute graft vs host disease. *World J Gastroenterol* 2021; 27(45): 7792-7800

**URL:** <https://www.wjnet.com/1007-9327/full/v27/i45/7792.htm>

**DOI:** <https://dx.doi.org/10.3748/wjg.v27.i45.7792>

## INTRODUCTION

In human health, DNA sequencing technologies, including 16S rRNA gene-based amplicon sequence analysis and whole genome shotgun metagenomic analysis were used to discover how exogenous and intrinsic host factors influence gut microbiome composition[1,2]. In large cohorts, scientists investigated the impact of factors such as lifestyle, dietary information, anthropometrics and drugs on the gut microbiome communities[3,4]. They found that age, gender, dietary factors and intrinsic parameters were highly correlated with composition and function of the gut microbiome. They also observed that several drug categories, such as antibiotics, proton-pump, metformin, statins, and laxatives, had a strong effect on the gut microbiome. On the other hand, gut microbiome can affect the bioavailability of oral drugs[5]. Thus, microorganisms can impact drug absorption and metabolism, that may explain, in part, inter-individual heterogeneity in drug response and disposition[6].

In addition, DNA sequencing technologies were used to investigate associations between composition or function of the gut microbiome and various disease states. Studies performed cross sectional comparisons between subjects with and without disease, and reported changes in the gut microbiome of patients with inflammatory bowel diseases, colorectal cancer, diabetes, obesity and metabolic disease, or autoimmune diseases compared to controls[7]. Even if it remains unclear whether these microbiome alterations are causative or secondary to the presence of the disease, gut microbiome could be viewed as a diagnostic biomarker[8].

Allogeneic hematopoietic stem cell transplantation (aHSCT) is a standard validated therapy, which every year allows an increasing number of patients suffering from malignant and nonmalignant hematological diseases, to benefit from an HSC transplant. The donor can be related (called geno-identical if HLA matched or haploidentical if sharing only one haplotype) or unrelated (called pheno-identical if HLA matched and including cord blood). However, aHSCT procedures are limited by potentially life-threatening complications, and one of the most serious complications is acute graft-versus-host disease (GVHD). GVHD occurs when immune competent T cells in the donated tissue recognize the recipient as foreign[9], that induces tissue damages in target organs of the host[10], including skin, liver and the gut, and typically occurs 3 to 6 wk after transplantation in nearly 60% of related donor transplants and 80% of unrelated donor transplants[11]. Hahn *et al*[12] assessed acute GVHD risk factors in 1960 adults after HLA-identical sibling myeloablative transplant in 226 centers worldwide. They found that cyclophosphamide and total-body irradiation, blood cell *vs* bone marrow grafts in patients age 18 to 39 years, recipient age 40 and older, chronic myeloid leukemia, Karnofsky performance score less than 90, and recipient/donor cytomegalovirus-seronegative were independent factors significantly associated with grade 2 to 4 acute GVHD[12].

Based on preclinical data demonstrating reduced acute GVHD severity in germ-free or antibiotic-treated mice (*i.e.*, gut microbiome with reduced diversity and richness), the gut microbiome was suggested to play a pivotal role in the pathogenesis of acute GVHD following aHSCT[13]. These findings are in contradiction with the common paradigm that loss of microbiome diversity is associated with diseases[14]. Most recent studies, based on microbiome sequencing, however, reported that higher diversity of intestinal microbiota at the time of neutrophil engraftment was associated with lower mortality in patients undergoing allogeneic hematopoietic-cell transplantation[15].

Here, we summarized the most recent findings of the association between human gut microbiome changes and graft *vs* host disease of the intestinal tract in patients receiving aHSCT.

## ALTERATIONS OF THE GUT MICROBIOME DURING aHSCT

Several studies reported the profound alteration of the gut microbiota during the aHSCT procedure, as summarized in [Table 1](#). Following various conditioning regimens, both myeloablative or of reduced-intensity, a recent 16S rRNA gene-based amplicon sequence analysis that profiled 8767 fecal samples from 1362 patients undergoing aHSCT at four different centers observed a significant gut microbiota disruption characterized by loss of diversity and domination by single taxa[15], defined as occupation of at least 30% of the gut microbiota by a single predominating bacterial taxon. They also identified an association between lower intestinal diversity and higher risks of transplantation-related death. Moreover, samples collected prior transplantation also showed evidence of microbiome disruption, and lower diversity before transplantation was associated with poor survival. In another study, mortality outcomes were significantly worse in patients with lower intestinal diversity, with an overall survival at 3 years of 36% in low gut microbiome diversity patients compared to 67% in high diversity groups. Overall, low diversity showed a strong effect on mortality after multivariate adjustment for other clinical predictors. Furthermore, in subjects with lower diversity, the gut microbiota was generally dominated by a single bacterial genus, including *Enterococcus*, *Streptococcus*, Enterobacteriaceae (*Escherichia* and *Kluyvera*), and *Lactobacillus*[16].

Taur *et al*[16] analyzed a total of 439 fecal specimens obtained from 94 patients during their transplant hospitalization and reported that over the course of aHSCT, gut microbiome diversity index decreased and remained low until the end of the observation period (Day 35 post-transplant). Patients also demonstrated significant changes in gut microbiome composition, and in most patients, the microbial composition became dominated by a single bacterial taxon: *Enterococcus* (40% of the patients), *Streptococcus* (37%) and the phylum Proteobacteria (13%). Interestingly, they demonstrated that patients with enterococcal domination in the gut had a 9-fold increased risk of Vancomycin Resistant *Enterococcus* bacteremia, and intestinal domination by Proteobacteria increased the risk of bacteremia with aerobic gram-negative bacilli 5-fold[17]. In a recent study using metagenomic shotgun, able to achieve bacterial identification to species and strain level, Ilett *et al*[10] reported that, in addition to a significant loss of gut microbiota diversity, post-aHSCT samples were enriched in *Staphylococcus*, *Eggerthella*, *Streptococcus*, *Enterococcus* and *Lactobacillus* compared to pre-aHSCT samples, and several samples were dominated by a single micro-organism [*Enterococcus* ( $n = 41$  of 112) or *Streptococcus* ( $n = 10$  of 112)]. At species level, they observed an enrichment of *Enterococcus faecium*, *Lactobacillus delbriekii*, *Staphylococcus epidermidis*, and *Streptococcus thermophilus* in the post-aHSCT samples compared to pre-aHSCT samples[10].

In patients receiving intensive chemotherapy regimen used as myeloablative conditioning treatment to prepare patients for aHSCT, gut microbiota after chemotherapy exhibited significant decrease in Firmicutes and Actinobacteria, and significant increase in Proteobacteria compared to samples collected before chemotherapy. At the genus level, gut microbiota after chemotherapy was significantly depleted in *Ruminococcus*, *Oscillospira*, *Blautia*, *Lachnospira*, *Roseburia*, *Dorea*, *Coprococcus*, *Anaerostipes*, *Clostridium*, *Collinsella*, *Adlercreutzia* and *Bifidobacterium*, and with significant increase in *Citrobacter*, *Klebsiella*, *Enterococcus*, *Megasphaera* and *Parabacteroides* compared with samples collected before chemotherapy. In addition, functional composition assessed using PICRUSt[18] revealed that following conditioning regimen, patients had reduced capacity for nucleotide metabolism, energy metabolism, metabolism of cofactors and vitamins, and increased capacity for glycan metabolism, signal transduction and xenobiotics biodegradation[19].

Thus, overall, aHSCT procedure is associated with a loss of gut microbiota diversity, a decrease in micro-organisms associated with health-promoting effects[20], and with a significant increase or domination by potentially pathobionts ([Figure 1](#)).

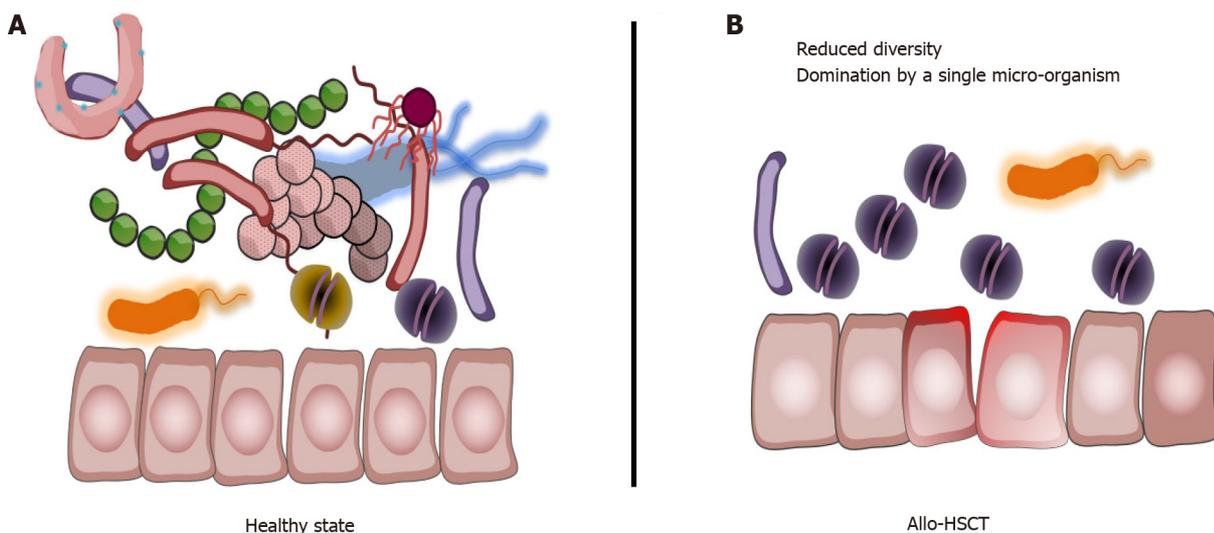
## ALTERATIONS OF THE DIVERSITY OF THE GUT MICROBIOME AND ACUTE GVHD

Studies reported that decreased gut microbiota diversity and richness was associated with the onset of acute intestinal GVHD. Jenq *et al*[21], in 2015, reported in a cohort of 115 patients receiving aHSCT, that increased bacterial diversity was associated with reduced GVHD-related mortality[21]. Moreover, in their recent 16S rRNA gene-based amplicon sequence analysis, Peled *et al*[15] found that higher intestinal diversity was

**Table 1 Gut microbiome diversity, composition and function changes in gut microbiota during allogeneic hematopoietic stem cell transplantation procedure**

Ref.	Sequencing technology	Change in diversity	Changes in composition	Change in functions
Peled <i>et al</i> [15]	16S rRNA gene-based amplicon sequence	Loss of diversity	Gut microbiota dominated by single taxa (occupation of at least 30% of the gut microbiota by a single predominating bacterial taxon), including <i>Enterococcus</i> , <i>Streptococcus</i> , Enterobacteriaceae ( <i>Escherichia</i> and <i>Kluyvera</i> ), and <i>Lactobacillus</i>	
Taur <i>et al</i> [16]	16S rRNA gene-based amplicon sequence	Decrease of the gut microbiome diversity index	Gut microbiota composition frequently dominated by a single bacterial taxon, including <i>Enterococcus</i> , <i>Streptococcus</i> or Proteobacteria	
Ilett <i>et al</i> [10]	Metagenomic shotgun	Loss of gut microbiota diversity	Post-aHST samples enriched in <i>Staphylococcus</i> , <i>Eggerthella</i> , <i>Streptococcus</i> , <i>Enterococcus</i> and <i>Lactobacillus</i> compared to pre-aHST samples, &t species level enrichment in <i>Enterococcus faecium</i> , <i>Lactobacillus delbrueckii</i> , <i>Staphylococcus epidermidis</i> , and <i>Streptococcus thermophilus</i>	
Montassier <i>et al</i> [19]	16S rRNA gene-based amplicon sequence	Loss of gut microbiota diversity	Decreases in abundances of Firmicutes and Actinobacteria, and significant increases in abundances of Proteobacteria	Reduced capacity for nucleotide metabolism, energy metabolism, metabolism of cofactors and vitamins, and increased capacity for glycan metabolism, signal transduction and xenobiotics biodegradation

HST: Hematopoietic stem cell transplantation.



**Figure 1 Gut microbiota changes during the allogeneic hematopoietic stem cell transplantation procedure.** A: Healthy state with diverse and rich gut microbiota; B: Following conditioning regimen, gut microbiome alterations are marked by a drastic loss of diversity, a significant decrease in micro-organisms associated with health-promoting effects, and with a significant increase or domination, defined as occupation of at least 30% of the microbiota by a single predominating bacterial taxon, by potentially pathobionts. Allo-HSCT: Allogeneic hematopoietic stem cell transplantation.

associated with decreased risk of deaths attributable to GVHD (17 GVHD-related deaths among 244 patients in the higher-diversity group *vs* 26 such deaths among 184 patients in the lower-diversity group; hazard ratio, 0.49; 95%CI: 0.26-0.90)[15].

In a cohort of 44 patients, in which 16 (36%) experienced acute intestinal GVHD (median time to diagnosis: 53 d), Galloway-Peña *et al*[22] found that lower Shannon diversity index, popular diversity index in the ecological literature, of fecal samples collected at the time of engraftment was significantly associated with increased incidence of acute GVHD[22]. Golob *et al*[23] also found that diversity was statistically significantly lower in patients with acute GVHD when compared to those with no acute GVHD[23]. In another cohort of 57 patients, Liu *et al*[24] reported that decreased gut microbiota recipients' diversity was not associated with decreased risk of aGVHD. However, they found that high gut microbiota donor diversity was associated with

decreased risk of acute GVHD[24]. In a cohort of 70 patients, Payen *et al*[25] reported that patients with severe aGVHD had reduced gut microbiota diversity at disease onset, whereas patients with mild aGVHD had gut microbiota diversity more similar to those of controls[25].

Ilett *et al*[10] applied shotgun metagenomic sequencing to study a large cohort of adults ( $n = 150$ ) undergoing aHSCT. Among them, 36 developed acute GVHD (median time to development: 34 d (interquartile range, 26-50 d post-aHSCT). They did not find significant association between diversity measures and acute GVHD in samples collected from the pre-aHSCT period. However, in samples collected in the early post-aHSCT period, patients who later developed acute GVHD had a significantly lower gene richness compared with those who did not develop acute GVHD[10].

Altogether, these findings clearly associate decreased diversity and richness of the gut microbiota, known to be associated with enhanced inflammation and impaired immunity[26], to onset of acute GVHD (Table 2).

## COMPOSITION CHANGES IN GUT MICROBIOME AND ACUTE GVHD

Using a 16S rRNA-based sequencing analysis, Jenq *et al*[21] reported that genus *Blautia* were most significantly associated with reduced GVHD-related mortality, whereas genus *Veillonella*, was associated with increased GVHD-related mortality[21].

Holler *et al*[27] reported that post-transplant samples were increased in enterococci, and that the increase was more pronounced in patients developing acute GVHD compared to patients who did not develop aGVHD. They confirmed this trend using enterococcal polymerase chain reaction (PCR) and observed a predominance of *Enterococcus faecium* and *Enterococcus faecalis* in samples collected in patients who developed acute GVHD. Moreover, post-transplant samples collected in patients with acute GVHD were significantly depleted in Clostridia and *Eubacterium rectale*[27]. Stein-Thoeringer *et al*[28] also reported that fecal domination by *Enterococcus* (*i.e.*, relative genus abundance  $\geq 30\%$  in samples collected during early post-transplant period) was associated with increased risk of acute GVHD, and increased GVHD-related mortality [28]. Importantly, in a gnotobiotic model, the authors demonstrated that *Enterococcus* growth is dependent on disaccharide lactose, and that dietary lactose depletion attenuates *Enterococcus* outgrowth and reduces the severity of GVHD[28].

In their cohort of 44 patients, Galloway-Peña *et al*[22] reported that only one taxon at the time of engraftment, *Coriobacteriia*, a class of Gram-positive bacteria within the Actinobacteria phylum, was negatively correlated with the incidence of aGVHD[22]. In another cohort of 107 patients, Doki *et al*[29] reported patients who developed acute GVHD exhibited a significantly higher abundance of phylum Firmicutes than patients who did not develop aGVHD[29].

Golob *et al*[23] found in a cohort of 66 patients, that in samples collected at neutrophil recovery post-HCT, the presence of Actinobacteria and Firmicutes was positively correlated with subsequent acute GVHD, whereas Lachnospiraceae were negatively correlated. In detail, *Butyricoccus*, *Bacteroides luti*, *Bacteroides thetaiotaomicron*, *Bacteroides ovatus*, and *Bacteroides caccae* were negatively correlated with subsequent acute severe GVHD, while *Rothia mucilaginosa*, *Solobacterium moorei*, *Veillonella parvula*, and *Bacteroides dorei* were positively correlated[23]. Payen *et al*[25] reported that *Lachnospiraceae*, *Blautia*, *Sellimonas*, *Anaerostipes*, *Faecalibacterium*, *Flavonifractor*, *Erysipelatoclostridium* and *Lactococcus* were negatively associated with subsequent acute severe GVHD, whereas *Prevotella* and *Stenotrophomonas* were considered positive biomarkers of severe aGVHD. Moreover, using qPCR, they observed a significant depletion of the *Blautia coccoides* group (cluster XIVA) in patients with aGVHD compared with controls and patients with no aGVHD[25].

Based on a shotgun metagenomic sequencing analysis in 150 patients, Ilett *et al*[10] found that no bacteria were associated with acute GVHD in samples collected during the pre-aHSCT period. In samples collected during the early post-aHSCT period, they found that *Blautia*, *Akkermansia*, and *Campylobacter*, as well as the specific species *Akkermansia muciniphila*, *Blautia obeum*, *Blautia hydrogenotropica*, and *Blautia hansenii* were all significantly associated with reduced risk of aGVHD[10]. Still using shotgun metagenomic sequencing analysis, Turner assessed samples collected in nine patients with aGVHD and treated with standard-of-care high-dose steroids. Three of these patients were steroid-refractory, whereas six had a response. They showed that *Dorea longicatena* was associated with response to high-dose steroids treatment whereas *Akkermansia muciniphila* was associated with refractoriness. They also reported that maintenance of a stable *Dorea/Akkermansia* ratio predicted steroid response, whereas a

**Table 2** Gut microbiome diversity, composition and function changes in patients receiving allogeneic hematopoietic stem cell transplantation procedure and developing acute graft versus host disease

Ref.	Sequencing technology	Change in diversity	Changes in composition	Change in functions
Ilett <i>et al</i> [10] (150 patients)	Shotgun metagenomic	Decrease in gene richness during the early post-aHSCT period in patients with aGVHD	Decrease in <i>Akkermansia muciniphila</i> , <i>Blautia obeum</i> , <i>Blautia hydrogenotropa</i> , and <i>Blautia hansenii</i> during the early post-aHSCT period in patients with aGVHD	Increase in toxin named PetZ, that triggers bacterial autolysis in pathological bacteria during the pre-aHSCT and the early post-aHSCT period in patients with aGVHD
Holler <i>et al</i> [27] (31 patients)	16S rRNA V3 sequencing		Increase in enterococci in patients who subsequently developed acute GVHD	
Galloway-Peña <i>et al</i> [22] (44 patients)	16S rRNA V4 sequencing	Lower Shannon diversity index in fecal samples collected at the time of engraftment in patients with subsequent aGVHD	<i>Coriobacteriia</i> negatively correlated with the incidence of acute GVHD	Fecal metabolites (Fecal indole and butyrate levels determined using liquid chromatography tandem mass spectrometry) associated with acute GVHD
Liu <i>et al</i> [24] (57 patients)	16S rRNA V4 sequencing	High gut microbiota donor diversity associated with decreased risk of aGVHD in recipient		
Doki <i>et al</i> [29] (107 patients)	16S rRNA V4 sequencing		Higher abundance of phylum Firmicutes in samples collected before aHSCT in patients with acute GVHD	
Golob <i>et al</i> [23] (66 patients)	16S rRNA V3-V4 sequencing	Diversity significantly lower in patients with GVHD	<i>Butyricoccus</i> , <i>Bacteroides luti</i> , <i>Bacteroides thetaiotaomicron</i> , <i>Bacteroides ovatus</i> , and <i>Bacteroides caccae</i> negatively correlated with subsequent acute GVHD, <i>Rothia mucilaginoso</i> , <i>Solobacterium moorei</i> , <i>Veillonella parvula</i> , and <i>Bacteroides dorei</i> positively correlated with subsequent onset of GVHD	
Michonneau <i>et al</i> [31] (99 patients)	Metabolomics			Significant decrease in tryptophan metabolites, including microbiota-produced compounds, such as 3-indoxyl sulfate, indoleacetate, indoleacetylglutamine, and indolepropionate in patients with aGVHD
Payen <i>et al</i> [25] (70 patients)	16S rRNA V3-V4 sequencing	Decreased diversity in acute GVHD	<i>Lachnoclostridium</i> , <i>Blautia</i> , <i>Sellimonas</i> , <i>Anaerostipes</i> , <i>Faecalibacterium</i> , <i>Flavonifractor</i> , <i>Erysipelatoclostridium</i> and <i>Lactococcus</i> negatively associated with subsequent acute GVHD	

HSCT: Hematopoietic stem cell transplantation; GVHD: Graft-versus-host disease.

decline in this ratio preceded refractory disease. Importantly, Shono *et al*[30] previously demonstrated in a mouse model an increased in *Akkermansia muciniphila*, a commensal bacterium with mucus-degrading capabilities, raising the possibility that mucus degradation may contribute to murine GVHD[30].

Further studies are needed to confirm or not the controversial role of *Akkermansia* in acute GVHD. Moreover, some species of the genus *Blautia* should be investigated as a potential biomarker: High relative abundance at the time of engraftment being protective against GVHD, while low relative abundance could be considered a risk factor for secondary development of GVHD.

## FUNCTION CHANGES IN GUT MICROBIOME AND ACUTE GVHD

The functional alterations of the intestinal microbiome in patients receiving aHSCT are currently poorly described because the majority of studies have used 16S ribosomal RNA gene-based techniques, which boil down to providing bacterial taxonomy only at the genus level, but are ineffective in obtaining functional information. In a cohort of 44 patients, Galloway-Peña *et al*[22] reported that fecal metabolites (fecal indole and butyrate levels) were not associated with aGVHD[22]. In another study, Michonneau *et al*[31] reported that aGVHD was characterized by specific metabolomics changes in

two cohorts of patients ( $n = 99$ ). They found that bile acids, plasmalogens, tryptophan, and arginine metabolites were the main contributors involved, especially the aryl hydrocarbon receptor ligand 3-indoxyl sulfate. In addition to host-derived metabolites, they also identified significant variation in microbiota-derived indole compounds, especially in aryl hydrocarbon receptor ligands. The authors suggested that allogeneic immune response during aGVHD might be influenced by bile acids and by the decreased production of aryl hydrocarbon receptor ligands by gut microbiome that could limit indoleamine 2,3-dioxygenase induction and influence allogeneic T cell reactivity[31]. To assess if altered composition of the gut microbiota may result in an altered metabolome, which potentially disrupts functionalities at the onset of aGVHD, Payen *et al*[25] quantified the Short Chain Fatty Acids (SCFA) content in fecal samples with measurement of total SCFAs and acetate, propionate, and butyrate. They found that the fecal amount of all SCFAs was drastically diminished at aGVHD onset. In detail, they observed that total SCFAs, acetate and butyrate respectively decreased by 80%, 75% and 95% in severe aGVHD patients as compared with controls[25]. Importantly, in a mouse model, Mathewson *et al*[32] demonstrated that butyrate restoration improved intestinal epithelial cells junctional integrity and mitigated aGVHD[32].

Based on a shotgun metagenomic sequencing analysis, Ilett *et al*[10] found that a total of 1267 and 1289 genes were present in significantly different amounts among those who developed aGVHD *vs* those who did not develop aGVHD during pre-HSCT and early post-aHSCT, respectively. Of these genes, 24 overlapped between the 2 time periods, all being significantly higher in abundance among those who did not develop aGVHD. They pointed out that 1 gene (O2.CD1-0-PT\_GL0039283) had a 3-log fold-change in both periods and is known to function as a toxin named Petz, that triggers bacterial autolysis in pathological bacteria[10].

## CONCLUSION

The investigations described in this mini review provide an understanding of the role of the gut microbiome in the pathophysiology of aGVHD in patients receiving aHSCT. Observational studies have shown that a decrease in diversity of the gut microbiome and specific species or metabolic pathways were associated with aGVHD. These specific changes could serve as biomarkers for diagnosis and prevention in patients receiving aHSCT. Moreover, functional alterations of the gut microbiome during aHSCT should be more investigated so that modulations of the gut microbiome could be tested to prevent this potentially life-threatening complication.

## REFERENCES

- 1 **Kurilshikov A**, Medina-Gomez C, Bacigalupe R, Radjabzadeh D, Wang J, Demirkan A, Le Roy CI, Raygoza Garay JA, Finnicum CT, Liu X, Zhernakova DV, Bonder MJ, Hansen TH, Frost F, Rühlemann MC, Turpin W, Moon JY, Kim HN, Lüll K, Barkan E, Shah SA, Fornage M, Szopinska-Tokov J, Wallen ZD, Borisevich D, Agreus L, Andreasson A, Bang C, Bedrani L, Bell JT, Bisgaard H, Boehnke M, Boomsma DI, Burk RD, Claringbould A, Croitoru K, Davies GE, van Duijn CM, Duijts L, Falony G, Fu J, van der Graaf A, Hansen T, Homuth G, Hughes DA, Ijzerman RG, Jackson MA, Jaddoe VWV, Joossens M, Jørgensen T, Keszthelyi D, Knight R, Laakso M, Laudes M, Launer LJ, Lieb W, Lusi AJ, Masclee AAM, Moll HA, Mujagic Z, Qibin Q, Rothschild D, Shin H, Sørensen SJ, Steves CJ, Thorsen J, Timpson NJ, Tito RY, Vieira-Silva S, Völker U, Völzke H, Vösa U, Wade KH, Walter S, Watanabe K, Weiss S, Weiss FU, Weissbrod O, Westra HJ, Willemsen G, Payami H, Jonkers DMAE, Arias Vasquez A, de Geus EJC, Meyer KA, Stokholm J, Segal E, Org E, Wijmenga C, Kim HL, Kaplan RC, Spector TD, Uitterlinden AG, Rivadeneira F, Franke A, Lerch MM, Franke L, Sanna S, D'Amato M, Pedersen O, Paterson AD, Kraaij R, Raes J, Zhernakova A. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat Genet* 2021; **53**: 156-165 [PMID: 33462485 DOI: 10.1038/s41588-020-00763-1]
- 2 **Rothschild D**, Weissbrod O, Barkan E, Kurilshikov A, Korem T, Zeevi D, Costea PI, Godneva A, Kalka IN, Bar N, Shilo S, Lador D, Vila AV, Zmora N, Pevsner-Fischer M, Israeli D, Kosower N, Malka G, Wolf BC, Avnit-Sagi T, Lotan-Pompan M, Weinberger A, Halpern Z, Carmi S, Fu J, Wijmenga C, Zhernakova A, Elinav E, Segal E. Environment dominates over host genetics in shaping human gut microbiota. *Nature* 2018; **555**: 210-215 [PMID: 29489753 DOI: 10.1038/nature25973]
- 3 **Zhernakova A**, Kurilshikov A, Bonder MJ, Tigchelaar EF, Schirmer M, Vatanen T, Mujagic Z, Vila AV, Falony G, Vieira-Silva S, Wang J, Imhann F, Brandsma E, Jankipersadsing SA, Joossens M, Cenit MC, Deelen P, Swertz MA; LifeLines cohort study, Weersma RK, Feskens EJ, Netea MG, Gevers D, Jonkers D, Franke L, Aulchenko YS, Huttenhower C, Raes J, Hofker MH, Xavier RJ,

- Wijmenga C, Fu J. Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. *Science* 2016; **352**: 565-569 [PMID: 27126040 DOI: 10.1126/science.aad3369]
- 4 **Vandeputte D**, Falony G, Vieira-Silva S, Tito RY, Joossens M, Raes J. Stool consistency is strongly associated with gut microbiota richness and composition, enterotypes and bacterial growth rates. *Gut* 2016; **65**: 57-62 [PMID: 26069274 DOI: 10.1136/gutjnl-2015-309618]
  - 5 **Haiser HJ**, Gootenberg DB, Chatman K, Sirasani G, Balskus EP, Turnbaugh PJ. Predicting and manipulating cardiac drug inactivation by the human gut bacterium *eggerthella lenta*. *Science* 2013; **341**: 295-298 [PMID: 23869020 DOI: 10.1126/science.1235872]
  - 6 **EIRakaiby M**, Dutilh BE, Rizkallah MR, Boleij A, Cole JN, Aziz RK. Pharmacomicrobiomics: the impact of human microbiome variations on systems pharmacology and personalized therapeutics. *OMICS* 2014; **18**: 402-414 [PMID: 24785449 DOI: 10.1089/omi.2014.0018]
  - 7 **Young VB**. The role of the microbiome in human health and disease: an introduction for clinicians. *BMJ* 2017; **356**: j831 [PMID: 28298355 DOI: 10.1136/bmj.j831]
  - 8 **Yang J**, McDowell A, Kim EK, Seo H, Lee WH, Moon CM, Kym SM, Lee DH, Park YS, Jee YK, Kim YK. Development of a colorectal cancer diagnostic model and dietary risk assessment through gut microbiome analysis. *Exp Mol Med* 2019; **51**: 1-15 [PMID: 31582724 DOI: 10.1038/s12276-019-0313-4]
  - 9 **Zeiser R**, Blazar BR. Acute Graft-versus-Host Disease - Biologic Process, Prevention, and Therapy. *N Engl J Med* 2017; **377**: 2167-2179 [PMID: 29171820 DOI: 10.1056/NEJMra1609337]
  - 10 **Ilett EE**, Jørgensen M, Noguera-Julian M, Norgaard JC, Daugaard G, Helleberg M, Paredes R, Murray DD, Lundgren J, MacPherson C, Reekie J, Sengeløv H. Associations of the gut microbiome and clinical factors with acute GVHD in allogeneic HSCT recipients. *Blood Adv* 2020; **4**: 5797-5809 [PMID: 33232475 DOI: 10.1182/bloodadvances.2020002677]
  - 11 **Vargas-Díez E**, García-Díez A, Marín A, Fernández-Herrera J. Life-threatening graft-vs-host disease. *Clin Dermatol* 2005; **23**: 285-300 [PMID: 15896544 DOI: 10.1016/j.clinidematol.2004.06.005]
  - 12 **Hahn T**, McCarthy PL Jr, Zhang MJ, Wang D, Arora M, Frangoul H, Gale RP, Hale GA, Horan J, Isola L, Maziarz RT, van Rood JJ, Gupta V, Halter J, Reddy V, Tiberghien P, Litzow M, Anasetti C, Pavletic S, Ringden O. Risk factors for acute graft-versus-host disease after human leukocyte antigen-identical sibling transplants for adults with leukemia. *J Clin Oncol* 2008; **26**: 5728-5734 [PMID: 18981462 DOI: 10.1200/JCO.2008.17.6545]
  - 13 **van Bekkum DW**, Roodenburg J, Heidt PJ, van der Waaij D. Mitigation of secondary disease of allogeneic mouse radiation chimeras by modification of the intestinal microflora. *J Natl Cancer Inst* 1974; **52**: 401-404 [PMID: 4150164 DOI: 10.1093/jnci/52.2.401]
  - 14 **Mosca A**, Leclerc M, Hugot JP. Gut Microbiota Diversity and Human Diseases: Should We Reintroduce Key Predators in Our Ecosystem? *Front Microbiol* 2016; **7**: 455 [PMID: 27065999 DOI: 10.3389/fmicb.2016.00455]
  - 15 **Peled JU**, Gomes ALC, Devlin SM, Littmann ER, Taur Y, Sung AD, Weber D, Hashimoto D, Slingerland AE, Slingerland JB, Maloy M, Clurman AG, Stein-Thoeringer CK, Markey KA, Docampo MD, Burgos da Silva M, Khan N, Gessner A, Messina JA, Romero K, Lew MV, Bush A, Bohannon L, Brereton DG, Fontana E, Amoretti LA, Wright RJ, Armijo GK, Shono Y, Sanchez-Escamilla M, Castillo Flores N, Alarcon Tomas A, Lin RJ, Yáñez San Segundo L, Shah GL, Cho C, Scordo M, Politikos I, Hayasaka K, Hasegawa Y, Gyurkocza B, Ponce DM, Barker JN, Perales MA, Giral SA, Jenq RR, Teshima T, Chao NJ, Holler E, Xavier JB, Pamer EG, van den Brink MRM. Microbiota as Predictor of Mortality in Allogeneic Hematopoietic-Cell Transplantation. *N Engl J Med* 2020; **382**: 822-834 [PMID: 32101664 DOI: 10.1056/NEJMoa1900623]
  - 16 **Taur Y**, Jenq RR, Perales MA, Littmann ER, Morjaria S, Ling L, No D, Gouborne A, Viale A, Dahi PB, Ponce DM, Barker JN, Giral SA, van den Brink M, Pamer EG. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. *Blood* 2014; **124**: 1174-1182 [PMID: 24939656 DOI: 10.1182/blood-2014-02-554725]
  - 17 **Taur Y**, Xavier JB, Lipuma L, Ubeda C, Goldberg J, Gouborne A, Lee YJ, Dubin KA, Socci ND, Viale A, Perales MA, Jenq RR, van den Brink MR, Pamer EG. Intestinal domination and the risk of bacteremia in patients undergoing allogeneic hematopoietic stem cell transplantation. *Clin Infect Dis* 2012; **55**: 905-914 [PMID: 22718773 DOI: 10.1093/cid/cis580]
  - 18 **Langille MG**, Zaneveld J, Caporaso JG, McDonald D, Knights D, Reyes JA, Clemente JC, Burkepile DE, Vega Thurber RL, Knight R, Beiko RG, Huttenhower C. Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat Biotechnol* 2013; **31**: 814-821 [PMID: 23975157 DOI: 10.1038/nbt.2676]
  - 19 **Montassier E**, Gastinne T, Vangay P, Al-Ghalith GA, Bruley des Varannes S, Massart S, Moreau P, Potel G, de La Cochetière MF, Batard E, Knights D. Chemotherapy-driven dysbiosis in the intestinal microbiome. *Aliment Pharmacol Ther* 2015; **42**: 515-528 [PMID: 26147207 DOI: 10.1111/apt.13302]
  - 20 **Eisenstein M**. The hunt for a healthy microbiome. *Nature* 2020; **577**: S6-S8 [PMID: 31996823 DOI: 10.1038/d41586-020-00193-3]
  - 21 **Jenq RR**, Taur Y, Devlin SM, Ponce DM, Goldberg JD, Ahr KF, Littmann ER, Ling L, Gouborne AC, Miller LC, Docampo MD, Peled JU, Arpaia N, Cross JR, Peets TK, Lumish MA, Shono Y, Dudakov JA, Poeck H, Hanash AM, Barker JN, Perales MA, Giral SA, Pamer EG, van den Brink MR. Intestinal *Blautia* Is Associated with Reduced Death from Graft-versus-Host Disease. *Biol Blood Marrow Transplant* 2015; **21**: 1373-1383 [PMID: 25977230 DOI: 10.1016/j.bbmt.2015.04.016]
  - 22 **Galloway-Peña JR**, Peterson CB, Malik F, Sahasrabhojane PV, Shah DP, Brumlow CE, Carlin LG,

- Chemaly RF, Im JS, Rondon G, Felix E, Veillon L, Lorenzi PL, Alousi AM, Jenq RR, Kontoyiannis DP, Shpall EJ, Shelburne SA, Okhuysen PC. Fecal Microbiome, Metabolites, and Stem Cell Transplant Outcomes: A Single-Center Pilot Study. *Open Forum Infect Dis* 2019; **6**: ofz173 [PMID: 31065565 DOI: 10.1093/ofid/ofz173]
- 23 **Golob JL**, Pergam SA, Srinivasan S, Fiedler TL, Liu C, Garcia K, Mielcarek M, Ko D, Aker S, Marquis S, Loeffelholz T, Plantinga A, Wu MC, Celustka K, Morrison A, Woodfield M, Fredricks DN. Stool Microbiota at Neutrophil Recovery Is Predictive for Severe Acute Graft vs Host Disease After Hematopoietic Cell Transplantation. *Clin Infect Dis* 2017; **65**: 1984-1991 [PMID: 29020185 DOI: 10.1093/cid/cix699]
- 24 **Liu C**, Frank DN, Horch M, Chau S, Ir D, Horch EA, Tretina K, van Besien K, Lozupone CA, Nguyen VH. Associations between acute gastrointestinal GvHD and the baseline gut microbiota of allogeneic hematopoietic stem cell transplant recipients and donors. *Bone Marrow Transplant* 2017; **52**: 1643-1650 [PMID: 28967895 DOI: 10.1038/bmt.2017.200]
- 25 **Payen M**, Nicolis I, Robin M, Michonneau D, Delannoye J, Mayeur C, Kapel N, Berçot B, Butel MJ, Le Goff J, Socié G, Rousseau C. Functional and phylogenetic alterations in gut microbiome are linked to graft-versus-host disease severity. *Blood Adv* 2020; **4**: 1824-1832 [PMID: 32353108 DOI: 10.1182/bloodadvances.2020001531]
- 26 **Simms-Waldrup TR**, Sunkersett G, Coughlin LA, Savani MR, Arana C, Kim J, Kim M, Zhan X, Greenberg DE, Xie Y, Davies SM, Koh AY. Antibiotic-Induced Depletion of Anti-inflammatory Clostridia Is Associated with the Development of Graft-versus-Host Disease in Pediatric Stem Cell Transplantation Patients. *Biol Blood Marrow Transplant* 2017; **23**: 820-829 [PMID: 28192251 DOI: 10.1016/j.bbmt.2017.02.004]
- 27 **Holler E**, Butzhammer P, Schmid K, Hundsruker C, Koestler J, Peter K, Zhu W, Sporrer D, Hehlhans T, Kreutz M, Holler B, Wolff D, Edinger M, Andreesen R, Levine JE, Ferrara JL, Gessner A, Spang R, Oefner PJ. Metagenomic analysis of the stool microbiome in patients receiving allogeneic stem cell transplantation: loss of diversity is associated with use of systemic antibiotics and more pronounced in gastrointestinal graft-versus-host disease. *Biol Blood Marrow Transplant* 2014; **20**: 640-645 [PMID: 24492144 DOI: 10.1016/j.bbmt.2014.01.030]
- 28 **Stein-Thoeringer CK**, Nichols KB, Lazrak A, Docampo MD, Slingerland AE, Slingerland JB, Clurman AG, Armijo G, Gomes ALC, Shono Y, Staffas A, Burgos da Silva M, Devlin SM, Markey KA, Bajic D, Pinedo R, Tsakmaklis A, Littmann ER, Pastore A, Taur Y, Monette S, Arcila ME, Pickard AJ, Maloy M, Wright RJ, Amoretti LA, Fontana E, Pham D, Jamal MA, Weber D, Sung AD, Hashimoto D, Scheid C, Xavier JB, Messina JA, Romero K, Lew M, Bush A, Bohannon L, Hayasaka K, Hasegawa Y, Vehreschild MJGT, Cross JR, Ponce DM, Perales MA, Giral SA, Jenq RR, Teshima T, Holler E, Chao NJ, Pamer EG, Peled JU, van den Brink MRM. Lactose drives *Enterococcus* expansion to promote graft-versus-host disease. *Science* 2019; **366**: 1143-1149 [PMID: 31780560 DOI: 10.1126/science.aax3760]
- 29 **Doki N**, Suyama M, Sasajima S, Ota J, Igarashi A, Mimura I, Morita H, Fujioka Y, Sugiyama D, Nishikawa H, Shimazu Y, Suda W, Takeshita K, Atarashi K, Hattori M, Sato E, Watakabe-Inamoto K, Yoshioka K, Najima Y, Kobayashi T, Kakihana K, Takahashi N, Sakamaki H, Honda K, Ohashi K. Clinical impact of pre-transplant gut microbial diversity on outcomes of allogeneic hematopoietic stem cell transplantation. *Ann Hematol* 2017; **96**: 1517-1523 [PMID: 28733895 DOI: 10.1007/s00277-017-3069-8]
- 30 **Shono Y**, Docampo MD, Peled JU, Perobelli SM, Velardi E, Tsai JJ, Slingerland AE, Smith OM, Young LF, Gupta J, Lieberman SR, Jay HV, Ahr KF, Porosnicu Rodriguez KA, Xu K, Calarfiore M, Poeck H, Caballero S, Devlin SM, Rapaport F, Dudakov JA, Hanash AM, Gyurkocza B, Murphy GF, Gomes C, Liu C, Moss EL, Falconer SB, Bhatt AS, Taur Y, Pamer EG, van den Brink MRM, Jenq RR. Increased GVHD-related mortality with broad-spectrum antibiotic use after allogeneic hematopoietic stem cell transplantation in human patients and mice. *Sci Transl Med* 2016; **8**: 339ra71 [PMID: 27194729 DOI: 10.1126/scitranslmed.aaf2311]
- 31 **Michonneau D**, Latis E, Curis E, Dubouchet L, Ramamoorthy S, Ingram B, de Latour RP, Robin M, de Fontbrune FS, Chevret S, Rogge L, Socié G. Metabolomics analysis of human acute graft-versus-host disease reveals changes in host and microbiota-derived metabolites. *Nat Commun* 2019; **10**: 5695 [PMID: 31836702 DOI: 10.1038/s41467-019-13498-3]
- 32 **Mathewson ND**, Jenq R, Mathew AV, Koenigsknecht M, Hanash A, Toubai T, Oravec-Wilson K, Wu SR, Sun Y, Rossi C, Fujiwara H, Byun J, Shono Y, Lindemans C, Calarfiore M, Schmidt TM, Honda K, Young VB, Pennathur S, van den Brink M, Reddy P. Gut microbiome-derived metabolites modulate intestinal epithelial cell damage and mitigate graft-versus-host disease. *Nat Immunol* 2016; **17**: 505-513 [PMID: 26998764 DOI: 10.1038/ni.3400]



Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA  
**Telephone:** +1-925-3991568  
**E-mail:** [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
**Help Desk:** <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>

