

World Journal of *Clinical Cases*

World J Clin Cases 2023 March 16; 11(8): 1669-1887



Contents

Thrice Monthly Volume 11 Number 8 March 16, 2023

REVIEW

- 1669** Understanding the multifaceted etiopathogenesis of foot complications in individuals with diabetes
Matijević T, Talapko J, Meštrović T, Matijević M, Erić S, Erić I, Škrlec I

MINIREVIEWS

- 1684** Diabetic foot ulcer: A comprehensive review of pathophysiology and management modalities
Raja JM, Maturana MA, Kayali S, Khouzam A, Efevbokhan N
- 1694** Isoperistaltic *vs* antiperistaltic anastomosis after right hemicolectomy: A comprehensive review
Symeonidis D, Karakantas KS, Kissa L, Samara AA, Bompou E, Tepetes K, Tzovaras G
- 1702** Evolving paradigm of thrombolysis in pulmonary embolism: Comprehensive review of clinical manifestations, indications, recent advances and guideline
Ochani RK, Aibani R, Jatoi HN, Anwar M, Khan SA, Ratnani I, Surani S
- 1712** Corneal endothelial cells and acoustic cavitation in phacoemulsification
Chen K, Xu WY, Sun SS, Zhou HW
- 1719** Modern blepharoplasty: From bench to bedside
Miotti G, Zeppieri M, Pederzani G, Salati C, Parodi PC
- 1730** Pregnancy and medications for inflammatory bowel disease: An updated narrative review
Akiyama S, Steinberg JM, Kobayashi M, Suzuki H, Tsuchiya K
- 1741** Pathogenesis, clinical manifestations, diagnosis, and treatment progress of achalasia of cardia
Li MY, Wang QH, Chen RP, Su XF, Wang DY

ORIGINAL ARTICLE

Retrospective Study

- 1753** Patients with hepatocellular carcinoma that die during the first year of liver transplantation have high blood sFasL concentrations
Lorente L, Rodriguez ST, Sanz P, González-Rivero AF, Pérez-Cejas A, Padilla J, Díaz D, González A, Martín MM, Jiménez A, Cerro P, Portero J, Barrera MA

Prospective Study

- 1761** Epidemiological and clinical characteristics of COVID-19 in a Brazilian public hospital
Pinheiro FD, Lopes LW, Dórea RSDM, Araújo GRL, Silva FAFD, de Brito BB, Cordeiro Santos ML, Júnior GMS, de Lorenzo Barcia MTA, Marques RA, Botelho AB, Dantas ACS, Costa DT, Teixeira AF, Souza CL, Marques LM, Campos GB, Oliveira MV, de Magalhães Queiroz DM, Freire de Melo F

CASE REPORT

- 1771** Pediatric acute heart failure caused by endocardial fibroelastosis mimicking dilated cardiomyopathy: A case report
Xie YY, Li QL, Li XL, Yang F
- 1782** Extensively infarcted giant solitary hamartomatous polyp treated with endoscopic full-thickness resection: A case report
Ye L, Zhong JH, Liu YP, Chen DD, Ni SY, Peng FQ, Zhang S
- 1788** Combined hamartoma of the retina and retinal pigment epithelium: A case report
Ren Q, Han N, Zhang R, Chen RF, Yu P
- 1794** Testicular pain originating from lumbar disc degeneration: A case report
Yan XJ, Wu B, He X, Tian ZK, Peng BG
- 1799** Glucocorticoid-induced thrombotic microangiopathy in paroxysmal nocturnal hemoglobinuria: A case report and review of literature
Yang XD, Ju B, Xu J, Xiu NN, Sun XY, Zhao XC
- 1808** Giant juvenile fibroadenoma in a 14-year old Chinese female: A case report
Wang J, Zhang DD, Cheng JM, Chen HY, Yang RJ
- 1814** A complementary comment on primary hepatic angiosarcoma: A case report
Gulmez AO, Aydin S, Kantarci M
- 1823** Primary membranous nephrotic syndrome with chylothorax as first presentation: A case report and literature review
Feng LL, Du J, Wang C, Wang SL
- 1830** Continuous positive airway pressure for treating hypoxemia due to pulmonary vein injury: A case report
Zhou C, Song S, Fu JF, Zhao XL, Liu HQ, Pei HS, Guo HB
- 1837** False positive detection of serum cryptococcal antigens due to insufficient sample dilution: A case series
Chen WY, Zhong C, Zhou JY, Zhou H
- 1847** Lactation breast abscess treated with Gualou Xiaoyong decoction and painless lactation manipulation: A case report and review of literature
Jin LH, Zheng HL, Lin YX, Yang Y, Liu JL, Li RL, Ye HJ
- 1857** Treatment of a large area perioral viral herpes infection following noninvasive ventilation: A case report
Tang AM, Xu JY, Wang R, Li YM
- 1862** Gastroparesis after video-assisted thoracic surgery: A case report
An H, Liu YC
- 1869** Hyperlactemia associated with secondary hepatocellular carcinoma resection in relation to circulation stability and quality of recovery: A case report
Meng Y, Pei HS, Yu JJ

- 1878** Sclerosing odontogenic carcinoma of maxilla: A case report

Soh HY, Zhang WB, Yu Y, Zhang R, Chen Y, Gao Y, Peng X

ABOUT COVER

Editorial Board Member of *World Journal of Clinical Cases*, Zaza Demetrashvili, FACS, FICS, MD, PhD, Professor, Department of Surgery, Tbilisi State Medical University, Tbilisi 0177, Georgia. zdemetr@yahoo.com

AIMS AND SCOPE

The primary aim of *World Journal of Clinical Cases* (WJCC, *World J Clin Cases*) is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The WJCC is now abstracted and indexed in Science Citation Index Expanded (SCIE, also known as SciSearch®), Journal Citation Reports/Science Edition, Current Contents®/Clinical Medicine, PubMed, PubMed Central, Scopus, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 Edition of Journal Citation Reports® cites the 2021 impact factor (IF) for WJCC as 1.534; IF without journal self cites: 1.491; 5-year IF: 1.599; Journal Citation Indicator: 0.28; Ranking: 135 among 172 journals in medicine, general and internal; and Quartile category: Q4. The WJCC's CiteScore for 2021 is 1.2 and Scopus CiteScore rank 2021: General Medicine is 443/826.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Hua-Ge Yin*; Production Department Director: *Xu Guo*; Editorial Office Director: *Jin-Lei Wang*.

NAME OF JOURNAL

World Journal of Clinical Cases

ISSN

ISSN 2307-8960 (online)

LAUNCH DATE

April 16, 2013

FREQUENCY

Thrice Monthly

EDITORS-IN-CHIEF

Bao-Gan Peng, Jerzy Tadeusz Chudek, George Kontogeorgos, Maurizio Serati, Ja Hyeon Ku

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2307-8960/editorialboard.htm>

PUBLICATION DATE

March 16, 2023

COPYRIGHT

© 2023 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Corneal endothelial cells and acoustic cavitation in phacoemulsification

Kai Chen, Wen-Ya Xu, Si-Si Sun, Hong-Wei Zhou

Specialty type: Ophthalmology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): B
Grade C (Good): C
Grade D (Fair): D
Grade E (Poor): 0

P-Reviewer: Cheraqpour K, Iran;
Dragonieri S, Italy

Received: December 14, 2022

Peer-review started: December 14, 2022

First decision: January 3, 2023

Revised: January 14, 2023

Accepted: February 22, 2023

Article in press: February 22, 2023

Published online: March 16, 2023



Kai Chen, Wen-Ya Xu, Si-Si Sun, Department of Ophthalmology, Lianshui County People's Hospital, Huai'an 223400, Jiangsu Province, China

Hong-Wei Zhou, Department of Ophthalmology, Huai'an 82 Hospital, Huai'an 223001, Jiangsu Province, China

Hong-Wei Zhou, Department of Industrial Engineering, Tsinghua University, Beijing 100084, China

Corresponding author: Hong-Wei Zhou, MD, PhD, Chief Physician, Director, Doctor, Department of Ophthalmology, Huai'an 82 Hospital, No. 100 Jiankang Road, Qingjiang District, Huai'an 223001, Jiangsu Province, China. leave0307@163.com

Abstract

Postoperative complications of phacoemulsification, such as corneal edema caused by human corneal endothelial cell (CEC) injury, are still a matter of concern. Although several factors are known to cause CEC damage, the influence of ultrasound on the formation of free radicals during surgery should be considered. Ultrasound in aqueous humor induces cavitation and promotes the formation of hydroxyl radicals or reactive oxygen species (ROS). ROS-induced apoptosis and autophagy in phacoemulsification have been suggested to significantly promote CEC injury. CEC cannot regenerate after injury, and measures must be taken to prevent the loss of CEC after phacoemulsification or other CEC injuries. Antioxidants can reduce the oxidative stress injury of CEC during phacoemulsification. Evidence from rabbit eye studies shows that ascorbic acid infusion during operation or local application of ascorbic acid during phacoemulsification has a protective effect by scavenging free radicals or reducing oxidative stress. Both in experiments and clinical practice, hydrogen dissolved in the irrigating solution can also prevent CEC damage during phacoemulsification surgery. Astaxanthin (AST) can inhibit oxidative damage, thereby protecting different cells from most pathological conditions, such as myocardial cells, luteinized granulosa cells of the ovary, umbilical vascular endothelial cells, and human retina pigment epithelium cell line (ARPE-19). However, existing research has not focused on the application of AST to prevent oxidative stress during phacoemulsification, and the related mechanisms need to be studied. The Rho related helical coil kinase inhibitor Y-27632 can inhibit CEC apoptosis after phacoemulsification. Rigorous experiments are required to confirm whether its effect is realized through improving the ROS clearance ability of CEC.

Key Words: Cataract; Phacoemulsification; Corneal endothelial cells; Ultrasound; Acoustic cavitation; Oxidative stress; Antioxidant

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Ultrasound in aqueous solution induces cavitation and promotes the formation of hydroxyl radicals or reactive oxygen species (ROS). ROS-induced apoptosis and autophagy in phacoemulsification have been suggested to significantly promote corneal endothelial cell (CEC) injury. Antioxidants can reduce the oxidative stress-induced injury to CECs during phacoemulsification.

Citation: Chen K, Xu WY, Sun SS, Zhou HW. Corneal endothelial cells and acoustic cavitation in phacoemulsification. *World J Clin Cases* 2023; 11(8): 1712-1718

URL: <https://www.wjgnet.com/2307-8960/full/v11/i8/1712.htm>

DOI: <https://dx.doi.org/10.12998/wjcc.v11.i8.1712>

INTRODUCTION

Phacoemulsification has recently become an increasingly popular surgical procedure for cataracts, with improvements in devices and surgical techniques. However, postoperative complications are still a problem worthy of concern, such as infective endophthalmitis, retinal detachment, and corneal edema due to damage to human corneal endothelial cells (CECs). Although several factors are known to cause CEC damage, the effects of free radical formation by ultrasound waves during the procedure should be considered. Ultrasound in aqueous humor induces acoustic cavitation[1] and promotes the formation of reactive oxygen species (ROS)[2]. ROS-induced corneal endothelial damage has been demonstrated to significantly promote apoptosis and autophagy during phacoemulsification[3]. CECs cannot regenerate after injuries[4], and strategies must be taken to prevent CEC loss after phacoemulsification or other endothelial injuries. This paper discusses corneal endothelial injury caused by oxidative stress secondary to acoustic cavitation during phacoemulsification and also related protective measures and implications for related fields.

THE IMPORTANCE OF PREVENTING CECS LOSS AND INJURY

Phacoemulsification surgery is the most commonly used surgery to treat cataracts. The complications of phacoemulsification, which affect the prognosis of visual acuity, are still a matter of concern. These complications include macular cystoid edema, infectious endophthalmitis, retinal detachment, and corneal edema due to loss of CEC. Corneal edema caused by decompensation of CEC is one of the most serious complications of phacoemulsification and can lead to severe visual loss and pain. CECs play a crucial role in regulating the constant dehydration of the corneal stroma and transparency[5]. CECs play a role mainly through active fluid pump and barrier function[6]. Both clinical and experimental studies have shown that the proliferation ability of human CECs is limited[7]. CEC density (ECD) usually starts from 4000 cells/mm² at birth and gradually decreases with age. The average value for adults is approximately 2500 cells/mm², while the average value for elderly individuals is less than 2000 cells/mm². Corneas with an ECD less than 1000 cells/mm² may not be able to tolerate intraocular surgery. When ECD is less than 500 cells/mm², corneal edema and compensatory imbalance usually occur[8]. In previous studies, uneventful cataract surgery has been proven to induce CEC loss ranging from 12% to 20%[9,10]. At present, the only effective option to treat corneal endothelial dysfunction is corneal transplantation (*e.g.*, full thickness penetrating keratoplasty or lamellar endothelial keratoplasty)[11]. In phacoemulsification, CEC damage is caused by ultrasonic mechanical damage, such as turbulence of the anterior chamber fluid and mechanical collision of bubbles and crystal fragments[12]. The application of viscoelastic materials in ophthalmology can limit the CEC damage caused by this aspect. Studies have confirmed that endogenous damage caused by oxidative stress plays a major role in CEC damage caused by phacoemulsification[3,13]. Therefore, how to maximize the prevention of CEC loss and injury during and after phacoemulsification and how to find the best effective safety strategy to protect CECs is a research priority.

ACOUSTIC CAVITATION AND ITS EFFECTS ON CECS

Ultrasonic energy is used in various surgical operations and is generated by the conversion of electrical energy by piezoelectric crystals located in the casing head. The probe is a hollow or solid metal conduit that directs ultrasonic vibration energy from the transducer to the target tissue (Figure 1). The low-frequency and high-energy ultrasonic radiation from the transducer can lead to the periodic compression and expansion of bubbles suspended in the liquid medium. Acoustic cavitation (that is, bubble expansion and rapid adiabatic collapse in solution) can generate enough energy to generate free radicals, sonoluminescence, high pressure and temperature rise in the bubble gas phase, bubble core and liquid interface[14]. In a water medium, water molecules decompose to form H^+ and OH^- radicals as the main primary radicals. The generation of acoustic cavitation in a water medium is a random, nonlinear, complex and multifactor-related phenomenon.

Bond[14] proposed a mechanical model to describe the interaction between phacoemulsification or other ultrasonic operations and various tissues. In the process of phacoemulsification, the direct mechanical vibration effect between the probe and the tissue will produce a large amount of heat accumulation, so continuous synchronous irrigation can be used to reduce the heat effect. The application of irrigating fluid in ophthalmic surgery can reduce friction and thermal effects; however, the gas nuclei in these aqueous solutions may produce acoustic cavitation. Acoustic cavitation may interact with tissues through chemical, electromagnetic radiation, thermal and mechanical effects and reportedly cause cell and intracellular damage. The combined effects of acoustic cavitation on the survival of damaged cells (that is, the generation of radiation, ultraviolet radiation, high temperature and high pressure leading to acoustic perforation) may lead to ultrasonic energy damage and may explain immediate cell damage, apoptosis and death[15,16].

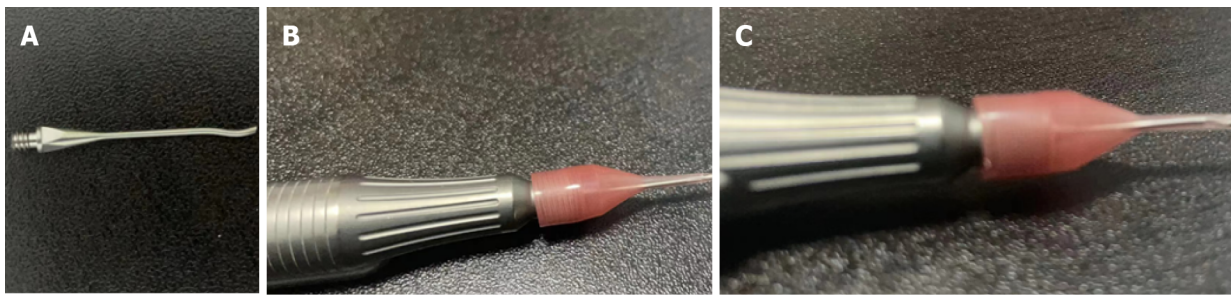
In phacoemulsification, because monolayer endothelial cells cannot regenerate, the damage and destruction of CECS should be minimized or eliminated. Aqueous humor contains natural antioxidants. In phacoemulsification, the irrigating solution circulates through the anterior chamber of the eye at a rate of approximately 20-30 mL/min to prevent heat accumulation. This effectively irrigates all aqueous humor within a few seconds, thus removing all natural antioxidants that usually protect the anterior chamber from oxidative damage. Therefore, ultrasonic emulsification in the irrigating solution environment can produce acoustic cavitation, but it depletes the natural antioxidants of aqueous humor.

The main expected effect of phacoemulsification is to chop, emulsify and empty the cataractous lens through the small clear corneal incision. To reduce heat accumulation and help lens fragmentation and emulsification, as well as the removal of debris, synchronous irrigation must be carried out continuously. However, for the generation of free radicals and sonoluminescence, cavitation may be considered an undesirable byproduct of phacoemulsification. Free radicals have been shown to cause toxic ocular oxidative damage, such as in the retina, lens and cornea[17]. The cytotoxicity of ROS to photosensitizer-treated and light-treated bovine CEC has been proven to be mediated by a parallel pathway leading to apoptosis and necrosis[18].

INHIBITING OXIDATIVE STRESS INDUCED BY ACOUSTIC CAVITATION PROTECTS CECS

Studies have confirmed that endogenous damage caused by oxidative stress plays an important role in CEC damage caused by phacoemulsification[3,16]. Therefore, some antioxidants can reduce the damage to CECS in phacoemulsification.

Holst *et al*[18] determined the formation of free radicals by adding luminol to the buffer solution and measuring the chemiluminescence *in vitro* and in rabbit eyes during phacoemulsification. They found that free radicals were formed during the process of phacoemulsification, and the number of free radicals was related to the power of ultrasound. In addition, the free radical scavenger superoxide dismutase (SOD) can inhibit the formation of free radicals, indicating that adding SOD to the irrigating solution during phacoemulsification can reduce the damage to CECS. Rubowitz *et al*[19] performed long-term phacoemulsification in the anterior chamber on 17 rabbit eyes. In 9 eyes, balanced salt eye solution was used in the phacoemulsification, and in 8 eyes, 0.001 M ascorbic acid was added to the solution. All other parameters were the same between the two groups. They found that there was no significant difference between the two groups in the number of CECS before the operation, but the ECD of the ascorbic acid treated group was higher compared to the other group after the operation ($P = 0.011$). They believed that adding ascorbic acid in the irrigating solution could significantly reduce the loss of CEC during phacoemulsification by approximately 70% and that it was due to the free radical scavenging properties of ascorbic acid. M Padua *et al*[12] showed that during phacoemulsification surgery in dogs, the application of ascorbic acid at a final concentration of 0.001 M in the irrigating solution significantly reduced the loss of CEC, the loss of hexagonal cells, and the coefficient of variation of CEC. Lee *et al*[20] reported the clinical application of ascorbic acid to prevent CEC damage related to phacoemulsification surgery in 2 patients during the perioperative period. In addition to cataracts, 2 patients suffered from Fuchs corneal endothelial dystrophy and corneal endotheliitis. Nakamura *et al*



DOI: 10.12998/wjcc.v11.i8.1712 Copyright ©The Author(s) 2023.

Figure 1 Photograph of the probe and sleeve. A: Probe; B: Front of probe with sleeve; C: Side of probe with sleeve.

[21] emphasized the protective effect of the antioxidant glutathione on CEC caused by ROS and found that oxidized glutathione has a better effect. Both oxidized glutathione and reduced glutathione can protect CECs.

The most relevant studies on acoustic cavitation and its effects on CECs are summarized in Table 1. A figure describing acoustic cavitation and its effects on CECs is included (Figure 2).

IMPLICATIONS FOR RELEVANT FIELDS

Endogenous injury of CECs induced by oxidative stress is the main cause of CEC injury induced by phacoemulsification. Reducing the damage to CECs during cataract surgery has been a hot topic in the field of ophthalmology.

Astaxanthin (AST) is an orange red carotenoid pigment that is the strongest antioxidant in nature. It has a variety of biological activities, including anticancer, anti-inflammatory, antiaging, antidiabetic, immune regulation and neuroprotective activities. However, it is unclear whether AST can protect CECs from endogenous damage caused by oxidative stress during phacoemulsification.

AST does not exist in human eyes and aqueous humor. At present, there are some studies on AST in ophthalmology. Otsuka *et al* [22] found that AST can prevent retinal ischemic damage through its antioxidant effect in a retinal ischemia-reperfusion model. Dong *et al* [23] found that in the diabetes db/db mouse model, AST can reduce the apoptosis of retinal ganglion cells, reduce the oxidative stress pressure of retinal tissue in db/db mice, reduce superoxide anion, reduce malondialdehyde, reduce 8-hydroxy-2-deoxyguanosine (8-OHdG) and increase manganese superoxide dismutase (Mn SOD) activity. Hashimoto *et al* [24] conducted a clinical experiment to evaluate the antioxidant effect of AST through changes in superoxide scavenging activity, hydrogen peroxide level and total organic peroxide in human aqueous humor. The subjects were 35 patients who underwent bilateral cataract surgery before and after taking AST (6 mg/day for 2 wk), and aqueous humor was taken during the surgery. After AST intake, the superoxide scavenging activity increased significantly, and the total organic peroxide level decreased significantly. The superoxide scavenging activity was significantly, negatively correlated with the total organic peroxide level ($R = -0.485$, $P < 0.01$), indicating that AST intake obviously enhanced the superoxide scavenging activity of human aqueous humor and inhibited the production of total organic peroxide in aqueous humor.

Many scholars have conducted in-depth research on the mechanism of AST in various cells. In a recent study, an ultraviolet (UV) B-induced oxidative stress model was used to evaluate the antioxidant effect of AST on a human retina pigment epithelium cell line (ARPE-19). The results showed that 20 μ M and 40 μ M AST can increase cell viability and have synergistic effects with ascorbic acid [25,26]. Increasing evidence shows that ROS production stimulates Phosphatidylinositol-3-kinase-serine/threonine kinase Akt (PI3K-Akt) mediated autophagy, while AST improves cell survival under oxidative inflammation conditions by activating the Akt signaling pathway [27]. Li Z and colleagues have proven that AST plays a protective role in H_2O_2 -induced oxidative damage by activating the PI3K-Akt pathway, and then PI3K-Akt further activates several downstream signal transduction media, such as mammalian target of rapamycin (mTOR) and the Nuclear factor erythroid 2-related factor 2-antioxidant response elements (Nrf2-ARE) pathway [28].

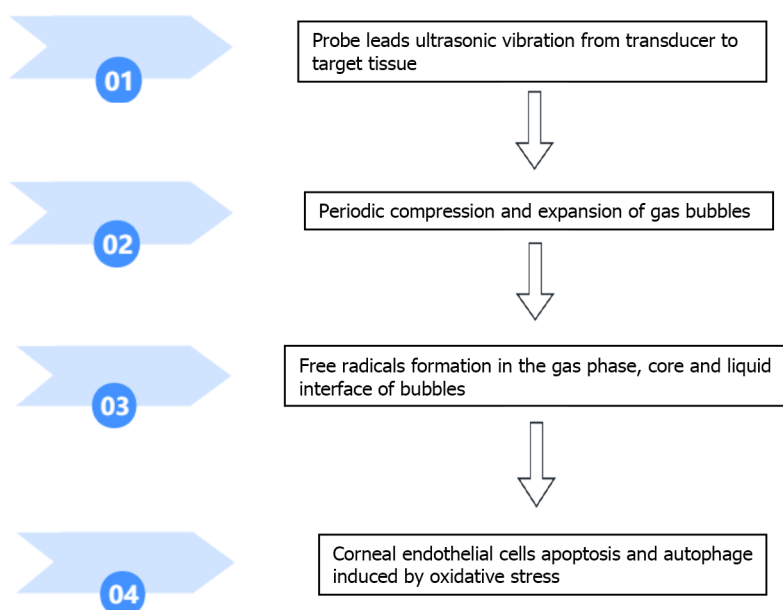
However, no previous studies have focused on the role of AST in the oxidative stress injury of CEC during phacoemulsification and its related mechanisms. In fact, in addition to phacoemulsification, there are many risk factors for the increase in CEC, such as solar ultraviolet radiation, aging and malnutrition. Therefore, local application of AST may contribute to the protection of patients with fragile CEC.

Previous papers have suggested that Rho related helical coil kinase (ROCK) plays important roles in cell cycle control because ROCK inhibits the premature separation of two centrioles in G1 period and is indispensable for the contraction of the cleavage groove (a necessary step for the completion of

Table 1 The most relevant studies about acoustic cavitation effects and its effects on corneal endothelial cells

Ref.	Journal	Key points
Hsueh <i>et al</i> [3]	<i>Cells</i>	ROS-induced CECs damage has been demonstrated to significantly promote apoptosis and autophagy during phacoemulsification
M Padua <i>et al</i> [12]	<i>Vet Ophthalmol</i>	Oxidative stress plays a major role in CECs damage and ascorbic acid significantly protected CECs during phacoemulsification
Ashush <i>et al</i> [13]	<i>Cancer Res</i>	Acoustic cavitation can generate free radicals, sonoluminescence, high pressure and temperature rise
Holst <i>et al</i> [18]	<i>Curr Eye Res</i>	SOD can inhibit the formation of free radicals during phacoemulsification
Rubowitz <i>et al</i> [19]	<i>Invest Ophthalmol Vis Sci</i>	The number of postoperative CECs in the study group treated with ascorbic acid was significantly more
Lee <i>et al</i> [20]	<i>World J Clin Cases</i>	Clinical application of ascorbic acid can prevent CECs damage related to phacoemulsification

CECs: Corneal endothelial cells; SOD: Superoxide dismutase; ROS: Reactive oxygen species.



DOI: 10.12998/wjcc.v11.i8.1712 Copyright ©The Author(s) 2023.

Figure 2 Acoustic cavitation and its effects on corneal endothelial cells.

cytokinesis)[29]. The ROCK inhibitor Y-27632 has been proven to increase proliferation and even immortalize primary keratinocytes in the presence of feeder cells[30]. Achiron *et al*[30] divided the corneal ring of a human donor into fragments, stored them in commercial storage medium with or without 10 mmol/L ROCK inhibitor, and then exposed them to phacoemulsification. The sample was separated into single cells by trypsin digestion. CEC was labeled with anti-CD166 antibodies to evaluate the early and late apoptosis rate of survival of CEC by flow cytometry analysis of annexin V and propidium iodide (PI) double staining. Six corneal and scleral rings from 4 donors were studied. After phacoemulsification, compared with the control group, the CEC exposed to Y-27632 showed that the early apoptosis rate decreased by 37.06%, and the late apoptosis rate decreased by 45.27%. The authors believe that ROCK inhibitors can be used before cataract surgery, especially in high-risk patients. This may be a promising new method to prevent pseudophakic bullous keratopathy. However, the author has not clarified the mechanism behind the effect. Zhou *et al*[31] found that the primary limbal epithelial cells of rabbits treated with Y-27632 also showed improved colony formation efficiency by enhancing the expansion of stem/progenitor cells. They proved that Y-27632 improved the cloning efficiency of rabbit limbal stem/progenitor cells by improving their adhesion and ROS clearance ability. Therefore, rigorous experiments are required to confirm whether the effect of Y-27632 on inhibiting the apoptosis of CECs after phacoemulsification is realized through the mechanism of improving the ROS clearance ability of CECs.

CONCLUSION

Ultrasound in aqueous humor can induce cavitation and promote the formation of ROS. ROS-induced apoptosis and autophagy have been suggested to significantly promote CEC injury during phacoemulsification. AST can inhibit oxidative damage, thus protecting different cells from most pathological conditions. However, existing research has not focused on the application of AST to prevent oxidative stress during phacoemulsification. Y-27632 inhibited CEC apoptosis after phacoemulsification. Further experiments are required to confirm whether the effect is realized by improving the ROS clearance ability of CECs.

FOOTNOTES

Author contributions: Chen K and Xu WY searched the references, designed and discussed the manuscript; Zhou HW and Sun SS searched the references, designed, composed, revised and submitted the manuscript.

Conflict-of-interest statement: Chen K has received research funding from Scientific Research Project of Lianshui County Health Committee in 2022; Xu WY, Sun SS, and Zhou HW have nothing to disclose.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: China

ORCID number: Kai Chen 0000-0003-4707-8246; Wen-Ya Xu 0000-0003-1955-6642; Si-Si Sun 0000-0001-9640-1455; Hong-Wei Zhou 0000-0002-5320-3886.

S-Editor: Yan JP

L-Editor: A

P-Editor: Yan JP

REFERENCES

- 1 **Topaz M**, Motiei M, Assia E, Meyerstein D, Meyerstein N, Gedanken A. Acoustic cavitation in phacoemulsification: chemical effects, modes of action and cavitation index. *Ultrasound Med Biol* 2002; **28**: 775-784 [PMID: 12113790 DOI: 10.1016/s0301-5629(02)00514-8]
- 2 **Augustin AJ**, Dick HB. Oxidative tissue damage after phacoemulsification: influence of ophthalmic viscosurgical devices. *J Cataract Refract Surg* 2004; **30**: 424-427 [PMID: 15030835 DOI: 10.1016/S0886-3350(03)00577-7]
- 3 **Hsueh YJ**, Meir YJ, Yeh LK, Wang TK, Huang CC, Lu TT, Cheng CM, Wu WC, Chen HC. Topical Ascorbic Acid Ameliorates Oxidative Stress-Induced Corneal Endothelial Damage via Suppression of Apoptosis and Autophagic Flux Blockage. *Cells* 2020; **9** [PMID: 32290365 DOI: 10.3390/cells9040943]
- 4 **Català P**, Thuret G, Skottman H, Mehta JS, Parekh M, Ni Dhubbghaill S, Collin RWJ, Nuijts RMMA, Ferrari S, LaPointe VLS, Dickman MM. Approaches for corneal endothelium regenerative medicine. *Prog Retin Eye Res* 2022; **87**: 100987 [PMID: 34237411 DOI: 10.1016/j.preteyeres.2021.100987]
- 5 **Sahu PK**, Das GK, Agrawal S, Kumar S. Comparative Evaluation of Corneal Endothelium in Patients with Diabetes Undergoing Phacoemulsification. *Middle East Afr J Ophthalmol* 2017; **24**: 74-80 [PMID: 28936050 DOI: 10.4103/meajo.MEAJO_242_15]
- 6 **Hsueh YJ**, Chen HC, Wu SE, Wang TK, Chen JK, Ma DH. Lysophosphatidic acid induces YAP-promoted proliferation of human corneal endothelial cells via PI3K and ROCK pathways. *Mol Ther Methods Clin Dev* 2015; **2**: 15014 [PMID: 26029725 DOI: 10.1038/mtm.2015.14]
- 7 **Rouhbakhshzaeri M**, Rabiee B, Azar N, Ghahari E, Putra I, Eslani M, Djalilian AR. New ex vivo model of corneal endothelial phacoemulsification injury and rescue therapy with mesenchymal stromal cell secretome. *J Cataract Refract Surg* 2019; **45**: 361-366 [PMID: 30527441 DOI: 10.1016/j.jcrs.2018.09.030]
- 8 **Choi JY**, Han YK. Long-term (≥10 years) results of corneal endothelial cell loss after cataract surgery. *Can J Ophthalmol* 2019; **54**: 438-444 [PMID: 31358141 DOI: 10.1016/j.jcjo.2018.08.005]
- 9 **Al-Mohtaseb Z**, He X, Yesilirmak N, Waren D, Donaldson KE. Comparison of Corneal Endothelial Cell Loss Between Two Femtosecond Laser Platforms and Standard Phacoemulsification. *J Refract Surg* 2017; **33**: 708-712 [PMID: 28991340 DOI: 10.3928/1081597X-20170731-01]
- 10 **Shen L**, Sun P, Zhang C, Yang L, Du L, Wu X. Therapy of corneal endothelial dysfunction with corneal endothelial cell-like cells derived from skin-derived precursors. *Sci Rep* 2017; **7**: 13400 [PMID: 29042661 DOI: 10.1038/s41598-017-13787-1]
- 11 **Wilkinson SW**, Park SSE, Ungricht EL, Trapnell M, Nydegger J, Cardenas IA, Brintz BJ, Mamalis N, Olson RJ, Werner

- L. Effect of simulated lenticular debris on corneal endothelial cells: experimental study in rabbit eyes. *J Cataract Refract Surg* 2022; **48**: 1325-1330 [PMID: [36026701](#) DOI: [10.1097/j.jcrs.0000000000001032](#)]
- 12 **M Padua IR**, P Valdetaro G, B Lima T, K Kobashigawa K, E S Silva P, Aldrovani M, M Padua PP, Laus JL. Effects of intracameral ascorbic acid on the corneal endothelium of dogs undergoing phacoemulsification. *Vet Ophthalmol* 2018; **21**: 151-159 [PMID: [28714237](#) DOI: [10.1111/vop.12490](#)]
- 13 **Ashush H**, Rozenszajn LA, Blass M, Barda-Saad M, Azimov D, Radnay J, Zipori D, Rosenschein U. Apoptosis induction of human myeloid leukemic cells by ultrasound exposure. *Cancer Res* 2000; **60**: 1014-1020 [PMID: [10706118](#)]
- 14 **Bond LJ**, Cimino WW. Physics of ultrasonic surgery using tissue fragmentation. *Ultrasonics* 1996; **34**: 579-585 [PMID: [8701547](#) DOI: [10.1016/0041-624x\(96\)00039-x](#)]
- 15 **Murano N**, Ishizaki M, Sato S, Fukuda Y, Takahashi H. Corneal endothelial cell damage by free radicals associated with ultrasound oscillation. *Arch Ophthalmol* 2008; **126**: 816-821 [PMID: [18541846](#) DOI: [10.1001/archophth.126.6.816](#)]
- 16 **Giblin FJ**, McCready JP. The effect of inhibition of glutathione reductase on the detoxification of H₂O₂ by rabbit lens. *Invest Ophthalmol Vis Sci* 1983; **24**: 113-118 [PMID: [6826306](#)]
- 17 **Cho KS**, Lee EH, Choi JS, Joo CK. Reactive oxygen species-induced apoptosis and necrosis in bovine corneal endothelial cells. *Invest Ophthalmol Vis Sci* 1999; **40**: 911-919 [PMID: [10102288](#)]
- 18 **Holst A**, Rolfsen W, Svensson B, Ollinger K, Lundgren B. Formation of free radicals during phacoemulsification. *Curr Eye Res* 1993; **12**: 359-365 [PMID: [8391415](#) DOI: [10.3109/02713689308999460](#)]
- 19 **Rubowitz A**, Assia EI, Rosner M, Topaz M. Antioxidant protection against corneal damage by free radicals during phacoemulsification. *Invest Ophthalmol Vis Sci* 2003; **44**: 1866-1870 [PMID: [12714617](#) DOI: [10.1167/iops.02-0892](#)]
- 20 **Lee CY**, Chen HT, Hsueh YJ, Chen HC, Huang CC, Meir YJ, Cheng CM, Wu WC. Perioperative topical ascorbic acid for the prevention of phacoemulsification-related corneal endothelial damage: Two case reports and review of literature. *World J Clin Cases* 2019; **7**: 642-649 [PMID: [30863764](#) DOI: [10.12998/wjcc.v7.i5.642](#)]
- 21 **Nakamura M**, Nakano T, Hikida M. Effects of oxidized glutathione and reduced glutathione on the barrier function of the corneal endothelium. *Cornea* 1994; **13**: 493-495 [PMID: [7842706](#)]
- 22 **Otsuka T**, Shimazawa M, Inoue Y, Nakano Y, Ojino K, Izawa H, Tsuruma K, Ishibashi T, Hara H. Astaxanthin Protects Against Retinal Damage: Evidence from In Vivo and In Vitro Retinal Ischemia and Reperfusion Models. *Curr Eye Res* 2016; **41**: 1465-1472 [PMID: [27158842](#) DOI: [10.3109/02713683.2015.1127392](#)]
- 23 **Dong LY**, Jin J, Lu G, Kang XL. Astaxanthin attenuates the apoptosis of retinal ganglion cells in db/db mice by inhibition of oxidative stress. *Mar Drugs* 2013; **11**: 960-974 [PMID: [23519150](#) DOI: [10.3390/md11030960](#)]
- 24 **Hashimoto H**, Arai K, Hayashi S, Okamoto H, Takahashi J, Chikuda M, Obara Y. Effects of astaxanthin on antioxidation in human aqueous humor. *J Clin Biochem Nutr* 2013; **53**: 1-7 [PMID: [23874063](#) DOI: [10.3164/jcbrn.13-6](#)]
- 25 **Oh S**, Kim YJ, Lee EK, Park SW, Yu HG. Antioxidative Effects of Ascorbic Acid and Astaxanthin on ARPE-19 Cells in an Oxidative Stress Model. *Antioxidants (Basel)* 2020; **9** [PMID: [32899990](#) DOI: [10.3390/antiox9090833](#)]
- 26 **Zhang J**, Ding C, Zhang S, Xu Y. Neuroprotective effects of astaxanthin against oxygen and glucose deprivation damage via the PI3K/Akt/GSK3 β /Nrf2 signalling pathway in vitro. *J Cell Mol Med* 2020; **24**: 8977-8985 [PMID: [32567157](#) DOI: [10.1111/jcmm.15531](#)]
- 27 **Li Z**, Dong X, Liu H, Chen X, Shi H, Fan Y, Hou D, Zhang X. Astaxanthin protects ARPE-19 cells from oxidative stress via upregulation of Nrf2-regulated phase II enzymes through activation of PI3K/Akt. *Mol Vis* 2013; **19**: 1656-1666 [PMID: [23901249](#)]
- 28 **Riento K**, Ridley AJ. ROCKs: multifunctional kinases in cell behaviour. *Nat Rev Mol Cell Biol* 2003; **4**: 446-456 [PMID: [12778124](#) DOI: [10.1038/nrm1128](#)]
- 29 **Liu X**, Ory V, Chapman S, Yuan H, Albanese C, Kallakury B, Timofeeva OA, Nealon C, Dakic A, Simic V, Haddad BR, Rhim JS, Dritschilo A, Riegel A, McBride A, Schlegel R. ROCK inhibitor and feeder cells induce the conditional reprogramming of epithelial cells. *Am J Pathol* 2012; **180**: 599-607 [PMID: [22189618](#) DOI: [10.1016/j.ajpath.2011.10.036](#)]
- 30 **Achiron A**, Feldman A, Karmona L, Avizemer H, Barequet IS, Rosner M, Knyazer B, Bartov E, Burgansky Z, Vishnevskia-Dai V. Prophylactic exposure of human corneal endothelial cells to Rho-associated kinase inhibitor reduced apoptosis rate after phacoemulsification: Ex vivo study. *J Cataract Refract Surg* 2018; **44**: 1261-1266 [PMID: [30172567](#) DOI: [10.1016/j.jcrs.2018.04.044](#)]
- 31 **Zhou Q**, Duan H, Wang Y, Qu M, Yang L, Xie L. ROCK inhibitor Y-27632 increases the cloning efficiency of limbal stem/progenitor cells by improving their adherence and ROS-scavenging capacity. *Tissue Eng Part C Methods* 2013; **19**: 531-537 [PMID: [23151007](#) DOI: [10.1089/ten.TEC.2012.0429](#)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

