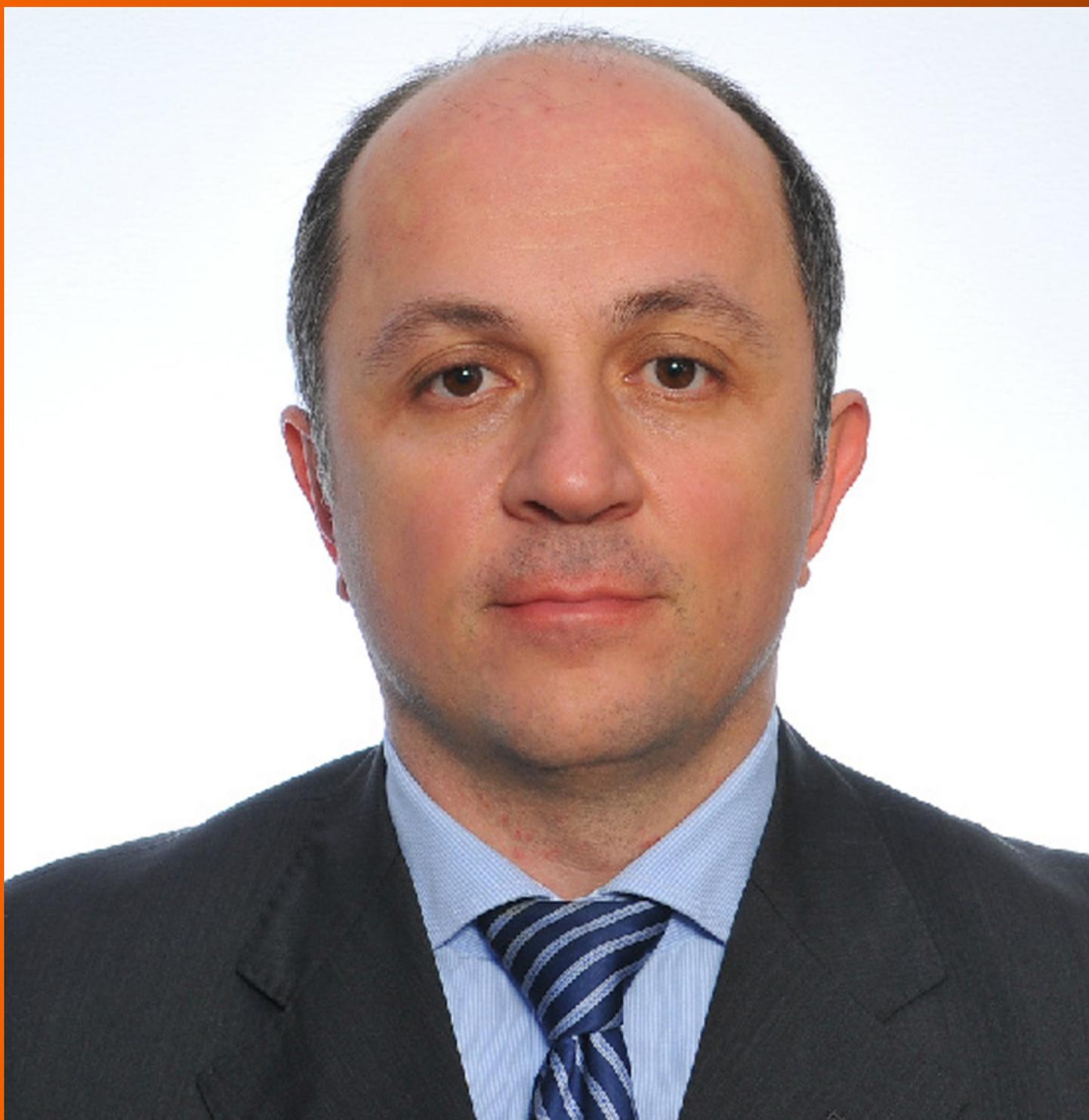


# World Journal of *Clinical Cases*

*World J Clin Cases* 2019 November 26; 7(22): 3683-3914



## Contents

Semimonthly Volume 7 Number 22 November 26, 2019

**REVIEW**

- 3683 Colorectal cancer: The epigenetic role of microbiome  
*Sabit H, Cevik E, Tombuloglu H*

**ORIGINAL ARTICLE****Case Control Study**

- 3698 Human podocyte injury in the early course of hypertensive renal injury  
*Sun D, Wang JJ, Wang W, Wang J, Wang LN, Yao L, Sun YH, Li ZL*

**Retrospective Cohort Study**

- 3711 Relationship between acute hypercarbia and hyperkalaemia during surgery  
*Weinberg L, Russell A, Mackley L, Dunnachie C, Meyerov J, Tan C, Li M, Hu R, Karalapillai D*

**Retrospective Study**

- 3718 Surgical treatment of patients with severe non-flail chest rib fractures  
*Zhang JP, Sun L, Li WQ, Wang YY, Li XZ, Liu Y*

- 3728 Super-selective arterial embolization in the control of acute lower gastrointestinal hemorrhage  
*Lv LS, Gu JT*

- 3734 End-stage liver disease score and future liver remnant volume predict post-hepatectomy liver failure in hepatocellular carcinoma  
*Kong FH, Miao XY, Zou H, Xiong L, Wen Y, Chen B, Liu X, Zhou JJ*

**Observational Study**

- 3742 Treatment of hemorrhoids: A survey of surgical practice in Australia and New Zealand  
*Fowler GE, Siddiqui J, Zahid A, Young CJ*

- 3751 Relationship between homocysteine level and prognosis of elderly patients with acute ischemic stroke treated by thrombolysis with recombinant tissue plasminogen activator  
*Li J, Zhou F, Wu FX*

**CASE REPORT**

- 3757 Cystic fibrosis transmembrane conductance regulator functional evaluations in a G542X+/- IVS8Tn:T7/9 patient with acute recurrent pancreatitis  
*Caldrer S, Bergamini G, Sandri A, Vercellone S, Rodella L, Cerofolini A, Tomba F, Catalano F, Frulloni L, Buffelli M, Tridello G, de Jonge H, Assael BM, Sorio C, Melotti P*

- 3765** Ulcerated intussuscepted jejunal lipoma-uncommon cause of obscure gastrointestinal bleeding: A case report  
*Cuciureanu T, Huiban L, Chiriac S, Singeap AM, Danciu M, Mihai F, Stanciu C, Trifan A, Vlad N*
- 3772** Ultrasonographic evaluation of the effect of extracorporeal shock wave therapy on calcific tendinopathy of the rectus femoris tendon: A case report  
*Lee CH, Oh MK, Yoo JI*
- 3778** Contrast-enhanced computed tomography findings of a huge perianal epidermoid cyst: A case report  
*Sun PM, Yang HM, Zhao Y, Yang JW, Yan HF, Liu JX, Sun HW, Cui Y*
- 3784** Iatrogenic crystalline lens injury during intravitreal injection of triamcinolone acetonide: A report of two cases  
*Su J, Zheng LJ, Liu XQ*
- 3792** Sagliker syndrome: A case report of a rare manifestation of uncontrolled secondary hyperparathyroidism in chronic renal failure  
*Yu Y, Zhu CF, Fu X, Xu H*
- 3800** Pre-eclampsia with new-onset systemic lupus erythematosus during pregnancy: A case report  
*Huang PZ, Du PY, Han C, Xia J, Wang C, Li J, Xue FX*
- 3807** Unilateral congenital scrotal agenesis with ipsilateral cryptorchidism: A case report  
*Fang Y, Lin J, Wang WW, Qiu J, Xie Y, Sang LP, Mo JC, Luo JH, Wei JH*
- 3812** Metastatic infection caused by hypervirulent *Klebsiella pneumonia* and co-infection with *Cryptococcus meningitis*: A case report  
*Shi YF, Wang YK, Wang YH, Liu H, Shi XH, Li XJ, Wu BQ*
- 3821** Allergic fungal rhinosinusitis accompanied by allergic bronchopulmonary aspergillosis: A case report and literature review  
*Cheng KJ, Zhou ML, Liu YC, Zhou SH*
- 3832** Invasive aspergillosis presenting as hilar masses with stenosis of bronchus: A case report  
*Su SS, Zhou Y, Xu HY, Zhou LP, Chen CS, Li YP*
- 3838** Retropharyngeal abscess presenting as acute airway obstruction in a 66-year-old woman: A case report  
*Lin J, Wu XM, Feng JX, Chen MF*
- 3844** Thoracoscopic segmentectomy assisted by three-dimensional computed tomography bronchography and angiography for lung cancer in a patient living with situs inversus totalis: A case report  
*Wu YJ, Bao Y, Wang YL*
- 3851** Single-lung transplantation for pulmonary alveolar microlithiasis: A case report  
*Ren XY, Fang XM, Chen JY, Ding H, Wang Y, Lu Q, Ming JL, Zhou LJ, Chen HW*

- 3859** Respiratory failure and macrophage activation syndrome as an onset of systemic lupus erythematosus: A case report  
*Sun J, Wang JW, Wang R, Zhang H, Sun J*
- 3866** Diagnosis of gastric duplication cyst by positron emission tomography/computed tomography: A case report  
*Hu YB, Gui HW*
- 3872** Peritoneal cancer after bilateral mastectomy, hysterectomy, and bilateral salpingo-oophorectomy with a poor prognosis: A case report and review of the literature  
*Ma YN, Bu HL, Jin CJ, Wang X, Zhang YZ, Zhang H*
- 3881** Apatinib for treatment of a pseudomyxoma peritonei patient after surgical treatment and hyperthermic intraperitoneal chemotherapy: A case report  
*Huang R, Shi XL, Wang YF, Yang F, Wang TT, Peng CX*
- 3887** Novel frameshift mutation causes early termination of the thyroxine-binding globulin protein and complete thyroxine-binding globulin deficiency in a Chinese family: A case report  
*Dang PP, Xiao WW, Shan ZY, Xi Y, Wang RR, Yu XH, Teng WP, Teng XC*
- 3895** Diffuse large B-cell lymphoma arising from follicular lymphoma with warthin's tumor of the parotid gland - immunophenotypic and genetic features: A case report  
*Wang CS, Chu X, Yang D, Ren L, Meng NL, Lv XX, Yun T, Cao YS*
- 3904** Exogenous endophthalmitis caused by *Enterococcus casseliflavus*: A case report  
*Bao QD, Liu TX, Xie M, Tian X*

**LETTER TO THE EDITOR**

- 3912** Microbial transglutaminase should be considered as an environmental inducer of celiac disease  
*Lerner A, Matthias T*

**ABOUT COVER**

Editorial Board Member of *World Journal of Clinical Cases*, Ridvan Hamid Alimehmeti, MD, PhD, Associate Professor, Lecturer, Surgeon, Department of Neuroscience, University of Medicine, Tirana 1000, Albania

**AIMS AND SCOPE**

The primary aim of *World Journal of Clinical Cases (WJCC, World J Clin Cases)* is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

*WJCC* mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

**INDEXING/ABSTRACTING**

The *WJCC* is now indexed in PubMed, PubMed Central, Science Citation Index Expanded (also known as SciSearch®), and Journal Citation Reports/Science Edition. The 2019 Edition of Journal Citation Reports cites the 2018 impact factor for *WJCC* as 1.153 (5-year impact factor: N/A), ranking *WJCC* as 99 among 160 journals in Medicine, General and Internal (quartile in category Q3).

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Responsible Electronic Editor: *Ji-Hong Liu*  
 Proofing Production Department Director: *Yun-Xiaojuan Wu*

<b>NAME OF JOURNAL</b> <i>World Journal of Clinical Cases</i>
<b>ISSN</b> ISSN 2307-8960 (online)
<b>LAUNCH DATE</b> April 16, 2013
<b>FREQUENCY</b> Semimonthly
<b>EDITORS-IN-CHIEF</b> Dennis A Bloomfield, Bao-Gan Peng, Sandro Vento
<b>EDITORIAL BOARD MEMBERS</b> <a href="https://www.wjnet.com/2307-8960/editorialboard.htm">https://www.wjnet.com/2307-8960/editorialboard.htm</a>
<b>EDITORIAL OFFICE</b> Jin-Lei Wang, Director
<b>PUBLICATION DATE</b> November 26, 2019

<b>COPYRIGHT</b> © 2019 Baishideng Publishing Group Inc
<b>INSTRUCTIONS TO AUTHORS</b> <a href="https://www.wjnet.com/bpg/gerinfo/204">https://www.wjnet.com/bpg/gerinfo/204</a>
<b>GUIDELINES FOR ETHICS DOCUMENTS</b> <a href="https://www.wjnet.com/bpg/GerInfo/287">https://www.wjnet.com/bpg/GerInfo/287</a>
<b>GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH</b> <a href="https://www.wjnet.com/bpg/gerinfo/240">https://www.wjnet.com/bpg/gerinfo/240</a>
<b>PUBLICATION MISCONDUCT</b> <a href="https://www.wjnet.com/bpg/gerinfo/208">https://www.wjnet.com/bpg/gerinfo/208</a>
<b>ARTICLE PROCESSING CHARGE</b> <a href="https://www.wjnet.com/bpg/gerinfo/242">https://www.wjnet.com/bpg/gerinfo/242</a>
<b>STEPS FOR SUBMITTING MANUSCRIPTS</b> <a href="https://www.wjnet.com/bpg/GerInfo/239">https://www.wjnet.com/bpg/GerInfo/239</a>
<b>ONLINE SUBMISSION</b> <a href="https://www.f6publishing.com">https://www.f6publishing.com</a>

## Case Control Study

## Human podocyte injury in the early course of hypertensive renal injury

Da Sun, Jiao-Jiao Wang, Wei Wang, Juan Wang, Li-Ning Wang, Li Yao, Ying-Hui Sun, Zi-Long Li

**ORCID number:** Da Sun (0000-0002-5822-5195); Jiao-Jiao Wang (0000-0003-2827-1622); Wei Wang (0000-0003-4743-3308); Juan Wang (0000-0001-7326-9922); Li-Ning Wang (0000-0001-6173-3361); Li Yao (0000-0002-2460-7283); Ying-Hui Sun (0000-0003-1751-3227); Zi-Long Li (0000-0002-2187-2874).

**Author contributions:** Sun YH and Li ZL conceived the study; Sun D, Wang JJ, Wang W and Wang J collected the clinical data and biological specimens and performed the research; Wang LN and Yao L analyzed the data; Sun D and Li ZL drafted the manuscript; all authors approved the final version of the article.

**Supported by** the Natural Science Foundation of Liaoning Provincial Department of Science and Technology, No. 2017225020.

**Institutional review board**

**statement:** The study was approved by Clinical Institutional Review Board for Human Research (Ethics Committee) of China Medical University.

**Informed consent statement:**

Written informed consent was obtained from all participants prior to enrollment in the study.

**Data sharing statement:**

Technical appendix, statistical code, and dataset available from the corresponding author at [yinghui\\_sun@163.com](mailto:yinghui_sun@163.com). Participants gave informed consent for data sharing.

**STROBE statement:** The guidelines of the STROBE Statement have

**Da Sun, Jiao-Jiao Wang, Wei Wang, Juan Wang, Li-Ning Wang, Li Yao, Zi-Long Li,** Department of Nephrology, The First Hospital of China Medical University, Shenyang 110001, Liaoning Province, China

**Ying-Hui Sun,** Department of Experimental Medicine, General Hospital of Northern Theater Command, Shenyang 110016, Liaoning Province, China

**Corresponding author:** Ying-Hui Sun, MD, Associate Chief Physician, Department of Experimental Medicine, General Hospital of Northern Theater Command, No. 83 Wenhua Road, Shenhe District, Shenyang 110016, Liaoning Province, China. [yinghui\\_sun@163.com](mailto:yinghui_sun@163.com)  
**Telephone:** +86-24-28851712

**Abstract****BACKGROUND**

Hypertension is prevalent in the general population and is regarded as the second leading cause of renal damage and dysfunction, outnumbered only by diabetes. However, the mechanisms remain unclear.

**AIM**

To investigate podocyte injury induced by hypertension in the early course without massive proteinuria or renal dysfunction.

**METHODS**

The hypertension group comprised 18 patients with hypertension accompanied by microalbuminuria, diagnosed with hypertensive renal injury according to biopsy results. For a comparison of pathological changes in renal tissue, control group 1 comprised 10 healthy volunteers, and control group 2 comprised 16 patients who underwent surgery for renal trauma.

**RESULTS**

The hypertension group had significantly higher blood pressure ( $P = 0.000$ ) and microalbuminuria ( $P = 0.000$ ) compared with control group 1. In the hypertension group, urinary podocytes were detected following positive staining of podocyte-specific nephrin and/or CD2-associated protein (CD2AP) in urine sediment. Podocyte foot process fusion and a significant decrease in nephrin and/or CD2AP expression in glomeruli were observed in the hypertension group compared with control group 2. This indicated that hypertension caused podocyte injury and detachment from the glomerular basement membrane, which was consistent with urinary detection of podocytes.

**CONCLUSION**

been adopted.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Unsolicited manuscript

**Received:** September 8, 2019

**Peer-review started:** September 8, 2019

**First decision:** October 24, 2019

**Revised:** November 3, 2019

**Accepted:** November 15, 2019

**Article in press:** November 15, 2019

**Published online:** November 26, 2019

**P-Reviewer:** Asselah T, Salami A

**S-Editor:** Wang JL

**L-Editor:** Webster JR

**E-Editor:** Qi LL



Our results suggest that podocyturia appears early in the course of hypertensive renal injury, and may be a sensitive marker for early prediction of hypertensive renal injury.

**Key words:** Podocyte; Hypertension; Hypertensive renal injury; Microalbuminuria; Nephryn; CD2-associated protein

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Podocytes probably play a crucial role in the pathogenesis of hypertensive renal injury, but human data are limited. In this study, we detected podocytes from urine sediments of patients with chronic hypertension and trace proteinuria, and obtained direct evidence that podocyte loss is not only associated with severe renal damage in chronic hypertension, but also occurs in the early course of hypertensive renal injury. We found that intrarenal expression of nephryn and CD2-associated protein was downregulated and consistent with podocyte shedding.

**Citation:** Sun D, Wang JJ, Wang W, Wang J, Wang LN, Yao L, Sun YH, Li ZL. Human podocyte injury in the early course of hypertensive renal injury. *World J Clin Cases* 2019; 7(22): 3698-3710

**URL:** <https://www.wjgnet.com/2307-8960/full/v7/i22/3698.htm>

**DOI:** <https://dx.doi.org/10.12998/wjcc.v7.i22.3698>

## INTRODUCTION

It is generally accepted that renal morphology and function are dependent on the maintenance of normal blood pressure. Long-standing uncontrolled hypertension can lead to hypertensive renal injury, which is a major cause of chronic kidney disease and accounts for significant mortality<sup>[1,2]</sup>.

Glomerular podocytes are highly differentiated cells that cover the outermost layer of the glomerular basement membrane. Podocytes and their foot processes form tight interdigitating networks that contribute to the hydraulic permeability of the glomeruli and play a crucial role as a filter for macromolecules<sup>[3]</sup>.

Due to the biological and morphological characteristics of podocytes, podocyte injury is recognized as a central feature in the development of many types of renal disease<sup>[4,5]</sup>. In fact, podocyte injury and loss has been proved to be causally related to glomerulosclerosis in a variety of kidney diseases such as diabetic nephropathy and minimal change disease<sup>[6-9]</sup>.

In the last 25 years, progressive observations in animal models have shown that podocyte injury is probably another key event in the pathogenesis of hypertensive nephrosclerosis<sup>[10-14]</sup>, subsequent to the lesions of glomerular ischemia resulting from narrowing and hyalinization of interlobular and afferent arterioles. A possible explanation is that elevated systemic blood pressure can give rise to Bowman's capsule hypertension, which may cause podocyte injury in a mechanical-stretch-dependent manner. However, it remains unclear whether podocyte-associated proteins are involved in the regulation and control of podocyte injury and loss. It is also unknown whether podocyturia can replace proteinuria for early prediction of hypertensive renal injury.

To address these problems, we detected urinary podocytes and obtained human data on podocyte injury and loss under hypertensive conditions. We also observed alterations of podocyte-specific proteins, nephryn and CD2-associated protein (CD2AP) in glomeruli in an attempt to understand the molecular mechanism of podocyte injury during chronic hypertension.

## MATERIALS AND METHODS

### *Patients, urine samples, and renal specimens*

This study was approved by the Clinical Institutional Review Board for Human Research (Ethics Committee) of China Medical University and conducted in the Department of Nephrology and Urological Surgery. Written informed consent was

obtained from all participants prior to enrollment in the study.

The hypertension group comprised 18 patients with chronic hypertension and microalbuminuria (9 male and 9 female; mean age, 63.2 years; range, 44–81 years). No patient had any signs of infection. Smokers and patients with a history of primary glomerular disease, cardiac insufficiency, diabetes, tumors, or rheumatoid diseases were excluded. Control group 1 comprised 10 healthy volunteers (5 male and 5 female; mean age, 60.1 years; range, 57–79 years) recruited from friends and family of laboratory staff. Control group 2 comprised 16 patients who underwent partial nephrectomy due to renal trauma (with no previous hypertension or proteinuria) in the Department of Urological Surgery during the study period.

Urine samples voided in the morning were collected in sterile containers and processed within 1 h. Freshly obtained urine specimens (100 mL) were transferred to tubes and centrifuged at 1500 r/min for 10 min. Urinary supernatant was carefully aspirated and the sediment pellet was mixed and cytospun onto slides. Subsequently, the slides were air-dried at room temperature, baked at 60°C for 2 h, and fixed with acetone at 4°C for 10 min.

Renal biopsy was performed in the hypertension group. The normal parts of nephrectomized kidney in control group 2 were used for comparison of pathological changes in kidney tissue. The renal specimens were subjected to routine fixation procedures. The pieces of tissue were immersed in 4% paraformaldehyde at room temperature for 24 h, followed by gradual dehydration in a series of alcohols and then transferred into xylene. Finally, the samples were embedded in paraffin wax. The remaining kidney tissue from both groups was maintained at -80°C for biochemical examination.

#### **Albuminuria, glomerular filtration rate and renal injury**

Albuminuria was evaluated by measuring the urinary albumin-to-creatinine ratio (ACR) from a single morning urine specimen. Microalbuminuria was defined as ACR 30–300 mg/g. Glomerular filtration rate (GFR) was estimated (eGFR) using the Chronic Kidney Disease Epidemiology Collaboration equation. Renal injury was defined as either microalbuminuria or eGFR < 60 mL/min/1.73 m<sup>2</sup>.

#### **Antibodies**

The primary antibodies used were: Rabbit polyclonal anti-nephrin (Boster, Wuhan, China); mouse monoclonal anti-CD2AP (Santa Cruz Biotechnology, Santa Cruz, CA, United States). The secondary antibodies used were: biotinylated sheep anti-rabbit IgG and biotinylated rabbit anti-mouse IgG (Sigma, St. Louis, MO, United States), tetramethylrhodamine (TRITC)-labeled anti-rabbit IgG and fluorescein isothiocyanate (FITC)-labeled anti-mouse IgG (Abcam, Cambridge, MA, United States).

#### **Immunostaining of deparaffinized sections**

The paraffin-embedded tissues were cut into 3- $\mu$ m thick sections and were deparaffinized with xylene and a graded series of alcohols. Some of the sections were stained with hematoxylin and eosin (HE) to observe morphology. Other sections were incubated with 3% hydrogen peroxide to block the nonspecific reactivity of endogenous peroxidase. After washing with phosphate-buffered saline (PBS), the samples were retrieved with sodium citrate buffer at high pressure and were incubated with blocking buffer (PBS containing 5% bovine serum albumin; Boster) at 37°C for 1 h. The sections were subsequently incubated with the primary antibodies (anti-nephrin or anti-CD2AP) at a dilution of 1:100 overnight at 4°C.

On the second day, the specimens were recovered at room temperature for 30 min and divided into two groups: (1) Some of the sections were used for immunohistochemistry staining. After washing in PBS, the corresponding secondary antibody was added at 37°C for 30 min. The sections were then incubated with horseradish peroxidase-conjugated avidin-biotin complex at 37°C for 20 min and visualized with metal-enhanced 3'-diaminobenzidine (DAB) for 5 min. To obtain a clearer structure, the sections were counterstained with hematoxylin for 50 s and then dehydrated in a graded series of hydrochloric acid, alcohols and xylene. Finally, the sections were sealed with neutral balsam, air-dried and viewed with a light microscope (Eclipse 80i, Nikon, Japan). The integral optical density (IOD) of each positive stain was analyzed using Image Pro Plus 6.0, and the result was determined as the sum of the glomeruli. IOD was defined as the sum of the ODs of all the positive pixels in the image, which represented the quality of the targeted protein; and (2) Other sections were used for immunofluorescent staining. The sections were incubated with TRITC-labeled donkey anti-rabbit IgG and FITC-labeled goat anti-mouse IgG at a dilution of 1: 200 at 37°C for 1 h in a moist chamber, followed by washing in PBS. Finally, the specimens were sealed with 50% glycerin and viewed with a confocal laser scanning microscope (FV 10-ASW2.1 Viewer).

### **Immunoelectron microscopy**

To detect ultrastructural changes in the glomeruli and immunolocalization of specific podocyte proteins under various hemodynamic conditions, the extracted renal tissues were fixed in 1% osmium tetroxide for 20 min at 4°C after DAB staining, dehydrated through an acetone gradient, and finally inversion-embedded with Epon 812 and polymerized at 60°C for 48 h. Ultrathin sections (70 nm) were cut with an ultramicrotome and stained with 4% uranyl acetate and 1% lead citrate, and then ultrastructural images were obtained with a transmission electron microscope (HT7650; Hitachi, Tokyo, Japan).

### **Immunostaining for urinary podocytes**

The acetone-fixed slides were stained in a similar manner to the renal tissue described above, and viewed under a light or confocal laser scanning microscope. Nucleated, nephrin-positive or CD2AP-positive cells were considered to be podocytes. Two to four slides were examined per patient sample.

### **Statistical analysis**

The quantitative data are presented as the mean  $\pm$  SD. Statistical analysis was performed using SPSS version 24.0 statistical software. For mean comparisons between the two groups, the Mann-Whitney test was used. For qualitative data comparison, the  $\chi^2$  test was used.  $P < 0.05$  was considered statistically significant.

## **RESULTS**

### **Clinical data**

Clinical characteristics including age, gender, body mass index (BMI), blood pressure, urinary ACR, serum creatinine, and eGFR were obtained by chart review (Table 1). There were no significant differences in age, BMI, serum creatinine and eGFR between the hypertension group and control group 1. The levels of systolic and diastolic blood pressure (including blood pressure under medical treatment) were significantly higher in the hypertension group than in control group 1 ( $P = 0.000$ ). ACR in the hypertension group fulfilled the criteria for microalbuminuria, and was significantly higher than in control group 1 ( $P = 0.000$ ). Of note, the absolute values of urinary albumin in 11 of 18 patients enrolled in our study were  $< 10.6$  mg/L (4.6–33 mg/L), and undetectable in routine urine tests.

### **Podocytes in urinary sediments**

To determine whether podocytes are lost from urine, we examined urine sediments for podocyte-specific proteins (nephrin and CD2AP) under different hemodynamic conditions. Nucleated cells expressing nephrin or CD2AP were assumed to be podocytes (Figure 1). In the hypertension group, 55.6% of urine samples had nephrin-positive cells present on cytopins, and 33.3% of urine samples had CD2AP-positive cells. Approximately 50% of samples positive for nephrin also stained for CD2AP. Double immunofluorescence staining showed partial colocalization of nephrin and CD2AP (Figure 2). However, nephrin and CD2AP were barely detected in control group 1 (Table 2).

### **Histological analysis of glomeruli**

To evaluate glomerular lesions in patients with chronic hypertension, we performed histological examination after HE staining. General morphology of the glomeruli from nephrectomized kidneys was normal (Figure 3). The capillary loops were smooth and plump under normotensive conditions (Figure 3A). However, the glomeruli revealed severe atrophy under chronic hypertension. The capillary loops became severely wizened or obliterated, while Bowman's capsule and the luminal space of tubules became more dilated than in normal glomeruli (Figure 3B).

### **Expression and distribution of nephrin and CD2AP**

Nephrin localized to the slit diaphragm (SD) was strongly detected and stained continually and evenly along the extra-glomerular capillary loops in control group 2 (Figure 4A), but was stained weakly and intermittently in the hypertensive group (Figure 4B). CD2AP is a multifunctional adaptor protein localized in the cytoplasm and membrane ruffles in podocytes. In the glomeruli of control group 2, the appearance of CD2AP was interrupted in a linear-like pattern and distributed diffusely (Figure 4C), but they became more scattered and decreased concomitantly with attenuation of nephrin in the hypertension group (Figure 4D). Figures 4E and F show the IODs of each protein in various groups.

Double immunofluorescent staining showed CD2AP (green) expression in the

**Table 1 Clinical characteristics**

Characteristics	Hypertension group (n = 18)	Control group 1 (n = 10)	P value
Gender			Not analyzed
Male	9	5	
Female	9	5	
Age (yr)	63.2 ± 10	60.1 ± 10.7	0.454
BMI (kg/m <sup>2</sup> )	23.9 ± 2.3	24 ± 1.6	0.969
Blood pressure (mmHg)			
Systolic	184 ± 17 (142 ± 18) <sup>1</sup>	122 ± 8	0.000 (0.003) <sup>2</sup>
Diastolic	106 ± 10 (82 ± 13) <sup>1</sup>	64 ± 11	0.000 (0.001) <sup>2</sup>
Microalbuminuria	13.01 ± 8.9	3.0 ± 1.1	0.000
ACR (mg/g)	107 ± 53	28 ± 3	0.000
Serum creatinine (μmol/L)	66 ± 11	61 ± 8	0.248
eGFR (mL/min/1.73 m <sup>2</sup> )	92.8 ± 11.2	98.9 ± 12.3	0.192

<sup>1</sup>Blood pressure under antihypertensive treatment;

<sup>2</sup>Blood pressure under treatment versus normal control group. Values are mean ± SD. *P* < 0.05 was considered statistically significant. BMI: Body mass index; ACR: Albumin-to-creatinine ratio.

glomeruli. CD2AP was partially co-localized with nephrin (red) resulting in a yellow overlap (Figure 5). The changes in nephrin and CD2AP were confirmed by confocal microscopy (Figure 5). Under hypertensive conditions, the changes in nephrin and CD2AP were similar to those observed by immunohistochemistry.

### Transmission electron microscopy

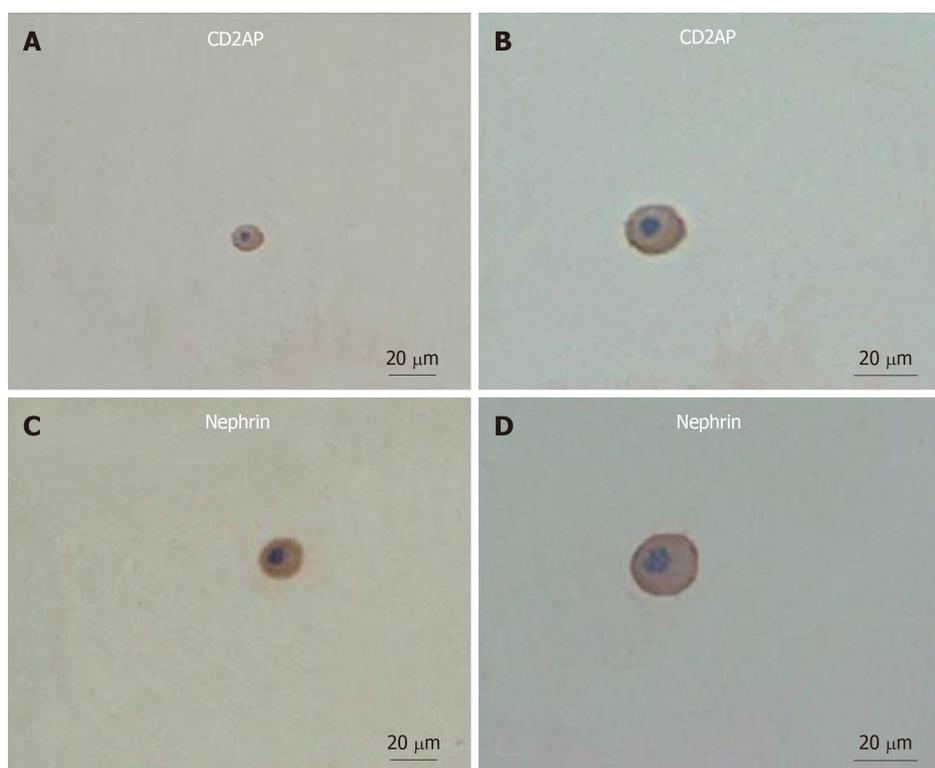
To visualize the distribution of nephrin and CD2AP under different hemodynamic conditions, we detected them using transmission electron microscopy. In control group 2, the foot processes had an intact arrangement and were displayed as interdigitating (Figures 6A and B). In contrast, the foot processes showed different degrees of fusion and partial foot process effacement in the hypertensive group (Figures 6C and D). Nephrin was distributed on the cytoplasmic face of foot processes in many filtration slits (Figure 6A), and CD2AP had a similar distribution (Figure 6B). In the hypertension group, staining of nephrin and CD2AP was scattered and uneven, and accumulated in some fusion foot processes, but the overall level decreased (Figures 6C and D).

## DISCUSSION

Podocyte loss due to severe podocyte injury (including change in expression of podocyte-associated proteins and ultrastructure) has been observed in the early course of nephropathy in rat models of hypertension<sup>[10,13,15]</sup>. However, human data on podocyte injury and loss in hypertensive renal injury are limited.

In the present study, the detection of urinary podocytes was consistent with podocyte loss from the glomeruli and indicated that podocyte injury might occur in the early course of hypertensive renal injury. This result was similar to those in previous studies of human and experimental hypertensive nephrosis<sup>[13,16,17]</sup>. Several studies have also detected podocytes in the urine of healthy subjects, but the number of podocytes shed by healthy controls is significantly lower than in patients with active glomerular diseases<sup>[17,18]</sup>. The possible explanation for these findings may be natural cell senescence and apoptosis<sup>[17,19]</sup>, which will need to be confirmed in a greater number of healthy participants.

Another study has suggested that urinary podocyte loss occurs prior to the onset of proteinuria in the 5/6 nephrectomy model<sup>[20]</sup>, characterized by systemic hypertension and continuing glomerular hyperfiltration. More importantly, podocyturia does not subside and mirrors proteinuria throughout the disease course. Although the level of ACR fulfilled the diagnostic criteria for microalbuminuria, we found that the absolute values of urinary albumin in most patients were in the normal range (< 10.6 mg/L), and could not be detected in routine urine tests. In contrast, podocytes were detected in the urine sediments in the same patients. These findings revealed that podocyturia is a sensitive marker, reflecting active ongoing glomerular damage, and is better than proteinuria. Detection of urinary podocytes may be used for predicting hypertensive



**Figure 1 Immunocytochemical staining of urinary podocytes in the hypertension group.** A: Podocyte expression of CD2-associated protein (CD2AP) (original magnification,  $\times 20$ ); B: Podocyte expression of CD2AP (original magnification,  $\times 40$ ); C: Podocyte expression of nephrin (original magnification,  $\times 20$ ); D: Podocyte expression of nephrin (original magnification,  $\times 40$ ). All cells were nucleated. Scale bars = 20  $\mu\text{m}$ . CD2AP: CD2-associated protein.

renal injury earlier.

Elevated systemic blood pressure can give rise to Bowman's capsule hypertension and glomerular hyperfiltration, which has been suggested to cause podocyte injuries in a mechanical-stretch-dependent manner<sup>[16]</sup>. However, the detailed changes in podocytes morphology in humans have not been clearly illustrated before. In the present study, we observed alterations of podocyte-specific proteins nephrin and CD2AP in glomeruli in an attempt to determine the injury sites of podocytes during chronic hypertension. The results demonstrated that expression of nephrin and CD2AP decreased under chronic hypertension conditions. In addition, the transposition and redistribution of nephrin and CD2AP occurred concomitantly with staining intensity attenuation.

Nephrin is a transmembrane protein expressed by glomerular podocytes and is predominantly localized at the SD<sup>[21]</sup>. Immunoelectron microscopy has revealed that nephrin bridges the filtration slit to form a 35-nm-long globular cross-strand that contributes to a porous SD scaffold<sup>[22]</sup>, which plays a crucial role in controlling glomerular permeability of plasma proteins<sup>[23,24]</sup>.

CD2AP is primarily expressed in podocytes at the cytoplasmic face of the SD, and serves as a linker protein between cell-membrane receptors and actin-modifying proteins<sup>[25,26]</sup>. Via CD2AP, nephrin is anchored to the SD on the actin cytoskeleton to adjust the arrangement of the cytoskeleton<sup>[27,28]</sup>.

Nephrin and CD2AP are critical to the integrity of the glomerular filtration barrier, and our findings indicate that downregulation and distribution of nephrin and CD2AP are causally related to podocyte injury. Consistent results were obtained in experimental hypertensive nephrosis, such as salt-loaded Dahl rats and malignant stroke-prone spontaneously hypertensive rats, which showed decreased expression of nephrin with podocyte injury<sup>[29]</sup>. Furthermore, reduced nephrin expression was also found in renal biopsy specimens from women with pre-eclampsia, a pregnancy-specific disorder characterized by hypertension and proteinuria<sup>[30]</sup>. However, direct evidence of the expression of CD2AP in glomeruli under chronic hypertension is limited. In previous studies, reduced expression of CD2AP has been demonstrated in humans with minimal change nephrotic syndrome and IgA nephropathy<sup>[31]</sup>. Surprisingly, the staining intensity of CD2AP was undiminished in passive Heymann nephritis, a rat model of human membranous nephropathy, although the staining appeared to be more condensed and uneven<sup>[32]</sup>.

One concern raised is that nephrin and CD2AP may also play a crucial role in

**Table 2** Expression of nephrin and CD2-associated protein in urinary sediments, *n* (%)

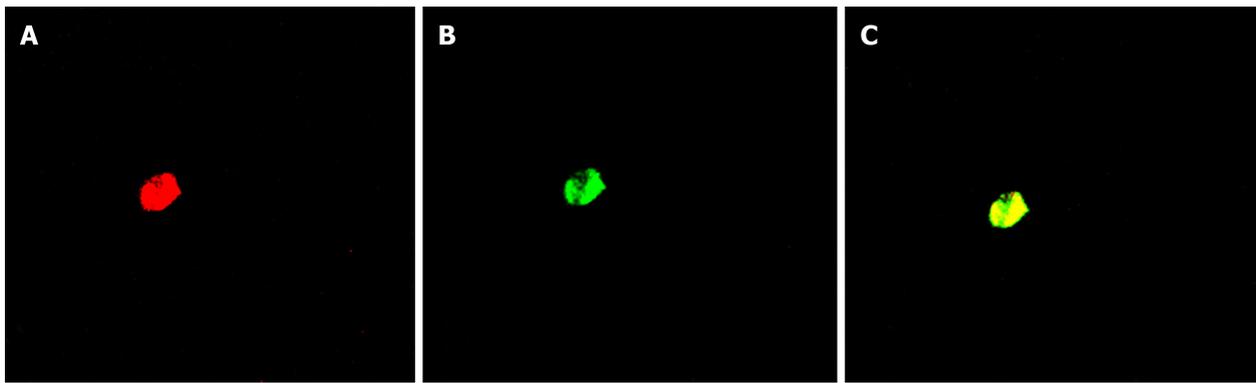
	Hypertension group ( <i>n</i> = 18)	Control group 1 ( <i>n</i> = 10)	<i>P</i> value
Nephrin	10 (55.6)	0 (0)	0.001
CD2AP	6 (33.3)	0 (0)	0.013
Nephrin and CD2AP	5 (27.8)	0 (0)	0.025

Values are positive rate. *P* < 0.05 was considered statistically significant. CD2AP: CD2-associated protein.

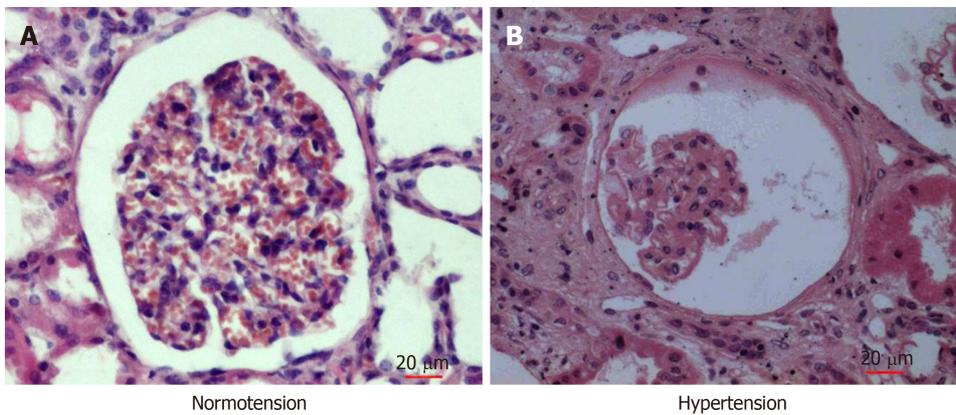
exacerbating podocyte injury. Recent studies have suggested that podocytes exposed to high concentrations of protein actively endocytose extracellular albumin in a time-dependent manner. When the endocytosed albumin is accumulated and out of the podocytes' handling capacity, it causes endoplasmic reticulum (ER) stress and triggers apoptosis of podocytes. Of interest, CD2AP can inhibit the apoptosis of podocytes induced by ER stress<sup>[33]</sup>. Besides, nephrin and CD2AP also bind to a subunit of phosphoinositide 3-kinase (PI3K), and together they stimulate PI3K-dependent AKT signaling and prevent Bad-mediated apoptosis of podocytes in culture<sup>[34,35]</sup>. Thus, we speculate that reduced and altered expression and distribution of nephrin and CD2AP are not only the results of podocyte injury, but also promoting factors in the progression of hypertensive podocyte damage.

All these findings improve our understanding of the mechanism of podocyte injury during chronic hypertension. However, there were several limitations in our study. First, this was a pure observational study and does not suggest a cause-and-effect relationship between urinary podocyte loss and progression of hypertensive renal injury. Second, the possibility that some of the podocyte loss may have resulted from apoptosis could not be excluded. Finally, our study population was complicated with microalbuminuria, so the earlier changes in podocytes before the onset of proteinuria were not observed.

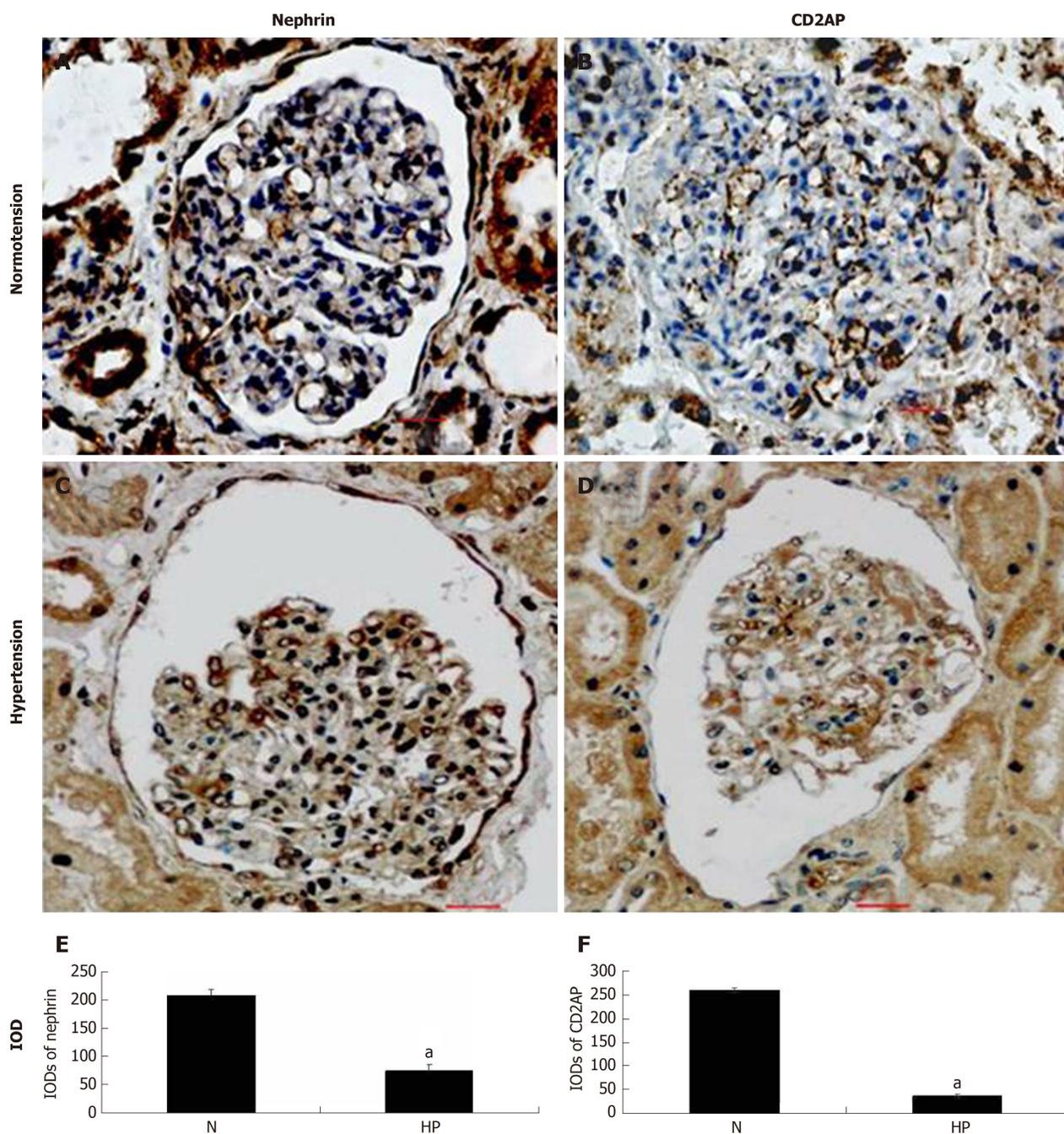
In summary, urinary podocytes were detected in patients with chronic hypertension and microalbuminuria, with immediate evidence of loss of podocytes. This supports podocyturia as an important contributor to podocyte injury and a potentially valuable diagnostic tool; in particular, given its potential to predict hypertensive renal injury earlier than proteinuria can. In addition, we visualized changes in podocyte-specific proteins, nephrin and CD2AP, which located the sites of podocyte injury during hypertensive renal injury. Our next step is to investigate the associated molecules in the nephrin and CD2AP signaling pathway, which should provide new insights into the regulatory mechanisms during hypertensive renal injury.



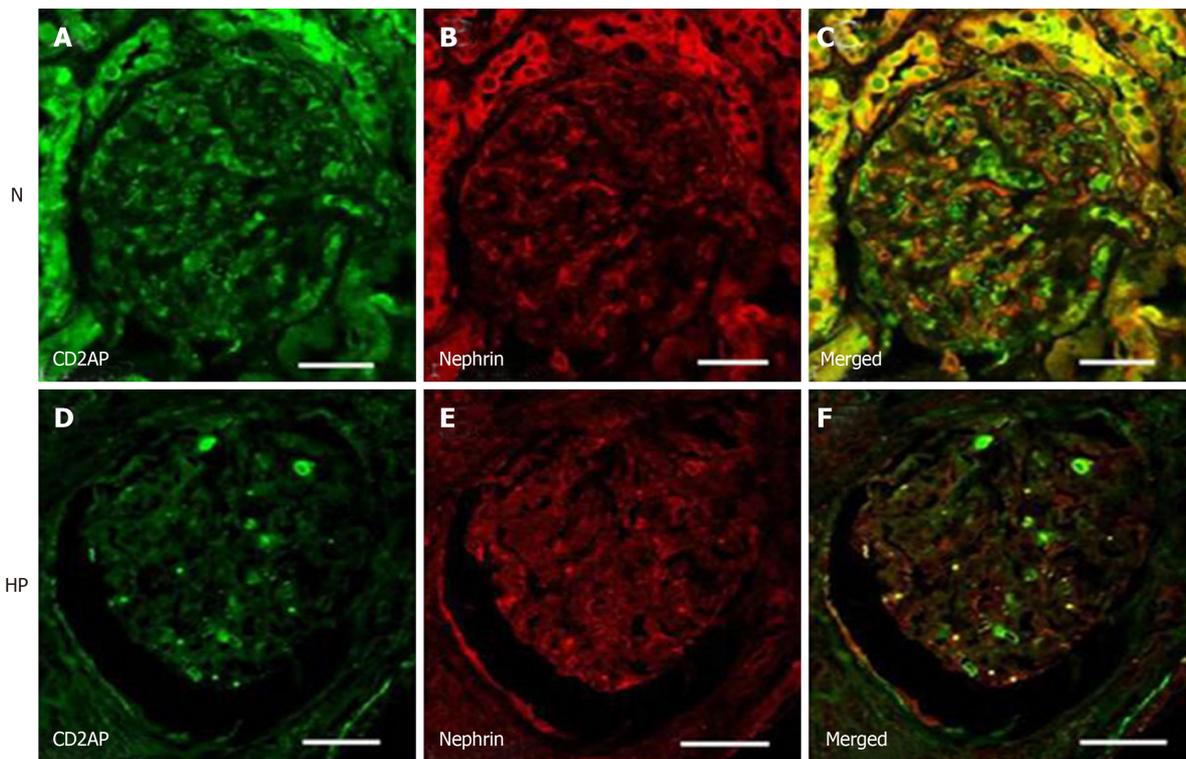
**Figure 2 Immunofluorescence staining of urinary podocytes in the hypertension group.** A: Podocyte expression of nephrin (red, original magnification,  $\times 40$ ); B: Podocyte expression of CD2-associated protein (CD2AP) (green, original magnification,  $\times 40$ ); C: Co-expression of nephrin and CD2AP (yellow, original magnification,  $\times 40$ ).



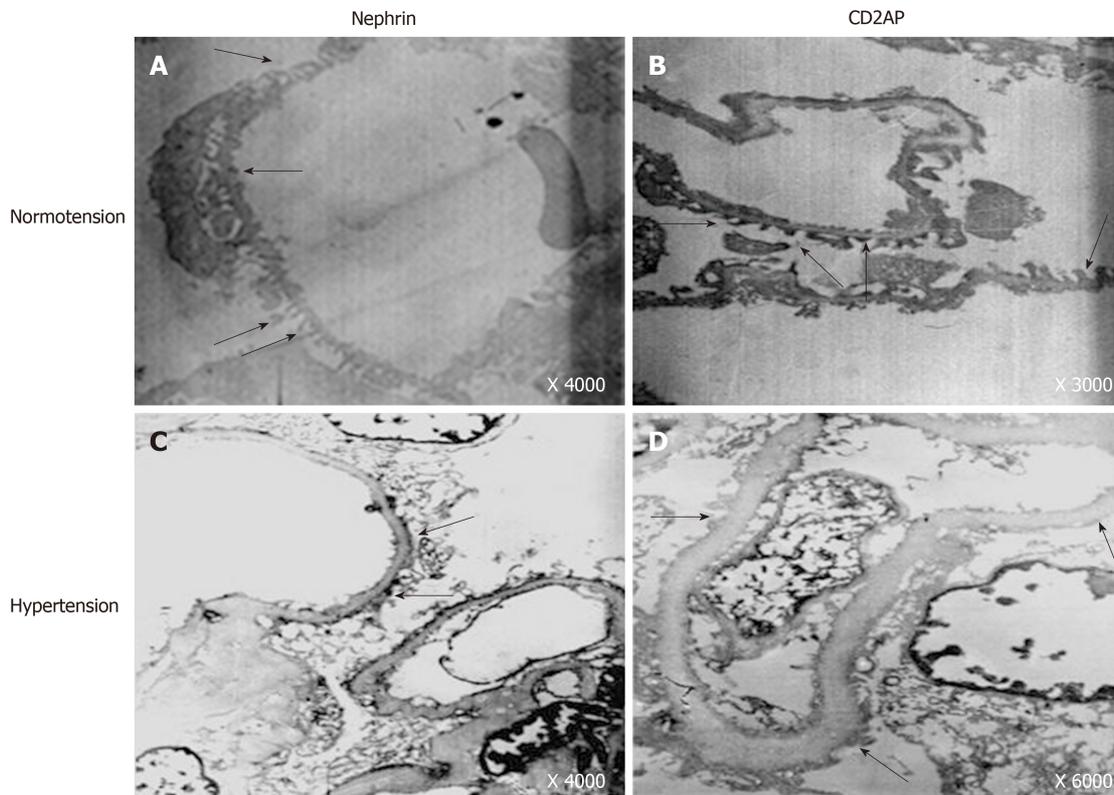
**Figure 3 Light micrographs of human renal cortical tissues stained with hematoxylin and eosin.** Renal tissues from patients in control group 2 who underwent partial nephrectomy due to renal trauma (A) and patients in the hypertension group (B). General morphology of glomeruli from nephrectomized kidney was normal, and the capillary loops were smooth and plump (A). Under hypertensive conditions, the glomeruli showed severe atrophy, the capillary loops became severely widened or obliterated, and Bowman's capsule and the luminal space of tubules were wide open (B). Scale bars = 20  $\mu\text{m}$ .



**Figure 4** Expression of nephrin and CD2-associated protein in human renal tissue by immunohistochemical staining. Tissues were treated with immersion-fixation methods under various hemodynamic conditions. A, C: The localization of nephrin; B, D: The localization of CD2-associated protein; E, F: The integral optical densities of each protein under various hemodynamic conditions. Immunohistochemical localization of both proteins in paraffin-embedded tissue is shown under normotensive (A, B) and hypertensive (C, D) conditions. Scale bar = 20  $\mu$ m. Histogram shows integral optical density of each protein. <sup>a</sup> $P < 0.05$ , Normotension vs Hypertension. N: Normotension; HP: Hypertension; IODs: Integral optical densities; CD2AP: CD2-associated protein.



**Figure 5** Confocal laser scanning micrographs show double immunofluorescence of nephrin-CD2-associated protein under normotensive and hypertensive conditions. Under normotensive conditions, double immunofluorescence staining showed that CD2-associated protein (CD2AP) (A) and nephrin (B) partially colocalized along the basement membrane of glomeruli (C). Under hypertensive conditions, the immunoreactivity of CD2AP and nephrin decreased and stained intermittently (D-F). Scale bar = 20  $\mu$ m. CD2AP: CD2-associated protein; N: Normotension; HP: Hypertension.



**Figure 6** Immunoelectron micrographs of nephrin and CD2-associated protein in glomeruli under normotensive and hypertensive conditions. Under normotensive conditions, the foot processes were intact and displayed as interdigitating (A, B, arrows). In contrast, the foot processes showed different degrees of fusion and partial foot process effacement under hypertensive conditions (C, D). Nephrin was distributed on the cytoplasmic face of foot processes in many filtration slits (A, arrows), and CD2-associated protein (CD2AP) had similar distribution characteristics (B, arrows). Under hypertensive conditions, the staining of nephrin and CD2AP was scattered and uneven, and accumulated in some fusion foot processes, but the overall level decreased (C, D, arrows). CD2AP: CD2-associated protein.

## ARTICLE HIGHLIGHTS

### Research background

Emerging evidence has shown that podocyte injury contributes to the development of renal damage associated with hypertensive kidney injury; however, the mechanism remains unclear and human data are limited.

### Research motivation

This study aimed to investigate human podocyte injury induced by hypertension in the early course without massive proteinuria or decreased renal function. Based on the information obtained in this study, it is expected to provide novel diagnostic methods and therapeutic strategies for hypertensive renal injury in the future.

### Research objectives

In this study, we summarized the method for detecting urinary podocytes, excluded the interference of physiological shedding, and verified the evaluation significance of urinary podocytes for hypertensive renal injury in the clinic. In addition, we preliminarily investigated the molecular mechanism of podocyte injury in the early course of hypertensive renal injury.

### Research methods

Human urine specimens and kidney tissue were used in this study. Urinary podocytes were observed under light microscopy and confocal microscopy. The intrarenal expression of podocyte-specific proteins, nephrin and CD2-associated protein (CD2AP) were detected by immunohistochemical techniques under different hemodynamic conditions and quantified by integral optical density values. The ultrastructure of human glomeruli was examined using transmission electron microscopy.

### Research results

Nucleated cells expressing nephrin or CD2AP were defined to be podocytes, and were detected in urine sediment in the hypertension group. By contrast, few podocytes were observed in control group 1. Moreover, podocyte foot process fusion and significantly decreased nephrin and/or CD2AP expression in glomeruli were observed in the hypertension group compared with control group 2, indicating podocyte injury and detachment from glomerular basement membrane, which was consistent with urinary detection of podocytes.

### Research conclusions

Decreased expression and the abnormal redistribution of nephrin and CD2AP are causally related to the disruption of podocyte morphology, and together with urinary podocytes lost from the glomerulus, confirm that podocyte injury is a key event in the progression of hypertensive kidney injury. Furthermore, podocyturia appears early in the course of hypertensive renal injury, and may be a sensitive marker for early prediction of hypertensive renal injury.

### Research perspectives

Calcium channels, associated signaling pathways and downstream cytokines are closely related to the regulatory mechanisms of podocytes, which is the research focus of hypertensive renal injury in future studies.

## ACKNOWLEDGEMENTS

We thank the Department of Immunodermatology, the First Hospital of China Medical University for providing us with confocal laser microscopy. Thanks also to the Cytobiology College, China Medical University for providing us with transmission electron microscopy.

## REFERENCES

- 1 **Dasgupta I**, Porter C, Innes A, Burden R. "Benign" hypertensive nephrosclerosis. *QJM* 2007; **100**: 113-119 [PMID: 17244670 DOI: 10.1093/qjmed/hcl139]
  - 2 **Yuan X**, Wang W, Wang J, Yin X, Zhai X, Wang L, Li K, Li Z. Down-regulation of integrin  $\beta$ 1 and focal adhesion kinase in renal glomeruli under various hemodynamic conditions. *PLoS One* 2014; **9**: e94212 [PMID: 24705394 DOI: 10.1371/journal.pone.0094212]
  - 3 **Mundel P**, Kriz W. Structure and function of podocytes: an update. *Anat Embryol (Berl)* 1995; **192**: 385-397 [PMID: 8546330]
  - 4 **Somlo S**, Mundel P. Getting a foothold in nephrotic syndrome. *Nat Genet* 2000; **24**: 333-335 [PMID: 10742089 DOI: 10.1038/74139]
  - 5 **Kerjaschki D**. Caught flat-footed: podocyte damage and the molecular bases of focal glomerulosclerosis. *J Clin Invest* 2001; **108**: 1583-1587 [PMID: 11733553 DOI: 10.1172/JCI14629]
  - 6 **Gassler N**, Elger M, Kränzlin B, Kriz W, Gretz N, Hähnel B, Hossler H, Hartmann I. Podocyte injury underlies the progression of focal segmental glomerulosclerosis in the fa/fa Zucker rat. *Kidney Int* 2001; **60**: 106-116 [PMID: 11422742 DOI: 10.1046/j.1523-1755.2001.00777.x]
- Szeto CC, Lai KB, Chow KM, Szeto CY, Yip TW, Woo KS, Li PK, Lai FM. Messenger RNA expression

- 7 of glomerular podocyte markers in the urinary sediment of acquired proteinuric diseases. *Clin Chim Acta* 2005; **361**: 182-190 [PMID: 15996647 DOI: 10.1016/j.cccn.2005.05.016]
- 8 **Kimura M**, Toyoda M, Saito N, Kaneyama N, Miyatake H, Tanaka E, Komaba H, Hara M, Fukagawa M. A Liquid-Based Cytology System, without the Use of Cyto centrifugation, for Detection of Podocytes in Urine Samples of Patients with Diabetic Nephropathy. *J Diabetes Res* 2019; **2019**: 9475637 [PMID: 30911554 DOI: 10.1155/2019/9475637]
- 9 **Raij L**, Tian R, Wong JS, He JC, Campbell KN. Podocyte injury: the role of proteinuria, urinary plasminogen, and oxidative stress. *Am J Physiol Renal Physiol* 2016; **311**: F1308-F1317 [PMID: 27335373 DOI: 10.1152/ajprenal.00162.2016]
- 10 **Kretzler M**, Koepfen-Hagemann I, Kriz W. Podocyte damage is a critical step in the development of glomerulosclerosis in the uninephrectomized-desoxycorticosterone hypertensive rat. *Virchows Arch* 1994; **425**: 181-193 [PMID: 7952502]
- 11 **Kriz W**. Podocyte is the major culprit accounting for the progression of chronic renal disease. *Microsc Res Tech* 2002; **57**: 189-195 [PMID: 12012382 DOI: 10.1002/jemt.10072]
- 12 **D'Agati VD**. The spectrum of focal segmental glomerulosclerosis: new insights. *Curr Opin Nephrol Hypertens* 2008; **17**: 271-281 [PMID: 18408478 DOI: 10.1097/MNH.0b013e3282f94a96]
- 13 **Kato T**, Mizuguchi N, Ito A. Characteristics of podocyte injury in malignant hypertensive nephropathy of rats (MSHRSP/Kpo strain). *Biomed Res* 2015; **36**: 313-321 [PMID: 26522148 DOI: 10.2220/biomedres.36.313]
- 14 **Li N**, Zhang J, Yan X, Zhang C, Liu H, Shan X, Li J, Yang Y, Huang C, Zhang P, Zhang Y, Bu P. SIRT3-KLF15 signaling ameliorates kidney injury induced by hypertension. *Oncotarget* 2017; **8**: 39592-39604 [PMID: 28465484 DOI: 10.18632/oncotarget.17165]
- 15 **Nagase M**, Shibata S, Yoshida S, Nagase T, Gotoda T, Fujita T. Podocyte injury underlies the glomerulopathy of Dahl salt-hypertensive rats and is reversed by aldosterone blocker. *Hypertension* 2006; **47**: 1084-1093 [PMID: 16636193 DOI: 10.1161/01.hyp.0000222003.28517.99]
- 16 **Wang G**, Lai FM, Kwan BC, Lai KB, Chow KM, Li PK, Szeto CC. Podocyte loss in human hypertensive nephrosclerosis. *Am J Hypertens* 2009; **22**: 300-306 [PMID: 19131934 DOI: 10.1038/ajh.2008.360]
- 17 **Vogelmann SU**, Nelson WJ, Myers BD, Lemley KV. Urinary excretion of viable podocytes in health and renal disease. *Am J Physiol Renal Physiol* 2003; **285**: F40-F48 [PMID: 12631553 DOI: 10.1152/ajprenal.00404.2002]
- 18 **Petermann A**, Floege J. Podocyte damage resulting in podocyturia: a potential diagnostic marker to assess glomerular disease activity. *Nephron Clin Pract* 2007; **106**: c61-c66 [PMID: 17570931 DOI: 10.1159/000101799]
- 19 **Ying WZ**, Wang PX, Sanders PW. Induction of apoptosis during development of hypertensive nephrosclerosis. *Kidney Int* 2000; **58**: 2007-2017 [PMID: 11044221 DOI: 10.1111/j.1523-1755.2000.00373.x]
- 20 **Yu D**, Petermann A, Kunter U, Rong S, Shankland SJ, Floege J. Urinary podocyte loss is a more specific marker of ongoing glomerular damage than proteinuria. *J Am Soc Nephrol* 2005; **16**: 1733-1741 [PMID: 15829708 DOI: 10.1681/asn.2005020159]
- 21 **Wang Y**, Zhao S, Loyd S, Groome LJ. Increased urinary excretion of nephrin, podocalyxin, and  $\beta$ g-h3 in women with preeclampsia. *Am J Physiol Renal Physiol* 2012; **302**: F1084-F1089 [PMID: 22301621 DOI: 10.1152/ajprenal.00597.2011]
- 22 **Li X**, Chuang PY, D'Agati VD, Dai Y, Yacoub R, Fu J, Xu J, Taku O, Premririt PK, Holzman LB, He JC. Nephrin Preserves Podocyte Viability and Glomerular Structure and Function in Adult Kidneys. *J Am Soc Nephrol* 2015; **26**: 2361-2377 [PMID: 25644109 DOI: 10.1681/ASN.2014040405]
- 23 **Mundel P**, Shankland SJ. Podocyte biology and response to injury. *J Am Soc Nephrol* 2002; **13**: 3005-3015 [PMID: 12444221 DOI: 10.1097/01.asn.0000039661.06947.fd]
- 24 **Reiser J**, Kriz W, Kretzler M, Mundel P. The glomerular slit diaphragm is a modified adherens junction. *J Am Soc Nephrol* 2000; **11**: 1-8 [PMID: 10616834]
- 25 **Wolf G**, Stahl RA. CD2-associated protein and glomerular disease. *Lancet* 2003; **362**: 1746-1748 [PMID: 14643126 DOI: 10.1016/S0140-6736(03)14856-8]
- 26 **Shih NY**, Li J, Cotran R, Mundel P, Miner JH, Shaw AS. CD2AP localizes to the slit diaphragm and binds to nephrin via a novel C-terminal domain. *Am J Pathol* 2001; **159**: 2303-2308 [PMID: 11733379 DOI: 10.1016/S0002-9440(10)63080-5]
- 27 **Barletta GM**, Kovari IA, Verma RK, Kerjaschki D, Holzman LB. Nephrin and Nep1 co-localize at the podocyte foot process intercellular junction and form cis hetero-oligomers. *J Biol Chem* 2003; **278**: 19266-19271 [PMID: 12646566 DOI: 10.1074/jbc.M301279200]
- 28 **Gerke P**, Huber TB, Sellin L, Benzing T, Walz G. Homodimerization and heterodimerization of the glomerular podocyte proteins nephrin and NEPH1. *J Am Soc Nephrol* 2003; **14**: 918-926 [PMID: 12660326 DOI: 10.1097/01.asn.0000057853.05686.89]
- 29 **Furness PN**, Hall LL, Shaw JA, Pringle JH. Glomerular expression of nephrin is decreased in acquired human nephrotic syndrome. *Nephrol Dial Transplant* 1999; **14**: 1234-1237 [PMID: 10344367 DOI: 10.1093/ndt/14.5.1234]
- 30 **Garovic VD**, Wagner SJ, Petrovic LM, Gray CE, Hall P, Sugimoto H, Kalluri R, Grande JP. Glomerular expression of nephrin and synaptopodin, but not podocin, is decreased in kidney sections from women with preeclampsia. *Nephrol Dial Transplant* 2007; **22**: 1136-1143 [PMID: 17255128 DOI: 10.1093/ndt/gfl711]
- 31 **Mao J**, Zhang Y, Du L, Dai Y, Yang C, Liang L. Expression profile of nephrin, podocin, and CD2AP in Chinese children with MCNS and IgA nephropathy. *Pediatr Nephrol* 2006; **21**: 1666-1675 [PMID: 16941146 DOI: 10.1007/s00467-006-0218-z]
- 32 **Yuan H**, Takeuchi E, Taylor GA, McLaughlin M, Brown D, Salant DJ. Nephrin dissociates from actin, and its expression is reduced in early experimental membranous nephropathy. *J Am Soc Nephrol* 2002; **13**: 946-956 [PMID: 11912254]
- 33 **He F**, Chen S, Wang H, Shao N, Tian X, Jiang H, Liu J, Zhu Z, Meng X, Zhang C. Regulation of CD2-associated protein influences podocyte endoplasmic reticulum stress-mediated apoptosis induced by albumin overload. *Gene* 2011; **484**: 18-25 [PMID: 21679752 DOI: 10.1016/j.gene.2011.05.025]
- 34 **Huber TB**, Hartleben B, Kim J, Schmidts M, Schermer B, Keil A, Egger L, Lecha RL, Borner C, Pavenstädt H, Shaw AS, Walz G, Benzing T. Nephrin and CD2AP associate with phosphoinositide 3-OH kinase and stimulate AKT-dependent signaling. *Mol Cell Biol* 2003; **23**: 4917-4928 [PMID: 12832477 DOI: 10.1128/mcb.23.14.4917-4928.2003]
- 35 **Verdoort A**, Honore PM, Jacobs R, De Waele E, Van Gorp V, De Regt J, Spapen HD. Do Statins Induce





Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA  
Telephone: +1-925-2238242  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>

