World Journal of *Clinical Cases*

World J Clin Cases 2020 September 26; 8(18): 3920-4279





Published by Baishideng Publishing Group Inc

W J C C World Journal of Clinical Cases

Contents

Semimonthly Volume 8 Number 18 September 26, 2020

OPINION REVIEW

3920 Special features of SARS-CoV-2 in daily practice Charitos IA, Ballini A, Bottalico L, Cantore S, Passarelli PC, Inchingolo F, D'Addona A, Santacroce L

EVIDENCE REVIEW

3934 Gastrointestinal insights during the COVID-19 epidemic

Nie K, Yang YY, Deng MZ, Wang XY

REVIEW

- 3942 From infections to autoimmunity: Diagnostic challenges in common variable immunodeficiency Więsik-Szewczyk E, Jahnz-Różyk K
- 3956 One disease, many faces-typical and atypical presentations of SARS-CoV-2 infection-related COVID-19 disease

Philips CA, Mohan N, Ahamed R, Kumbar S, Rajesh S, George T, Mohanan M, Augustine P

MINIREVIEWS

3971 Application of artificial neural networks in detection and diagnosis of gastrointestinal and liver tumors Mao WB, Lyu JY, Vaishnani DK, Lyu YM, Gong W, Xue XL, Shentu YP, Ma J

3978 Hepatic epithelioid hemangioendothelioma: Update on diagnosis and therapy Kou K, Chen YG, Zhou JP, Sun XD, Sun DW, Li SX, Lv GY

ORIGINAL ARTICLE

Clinical and Translational Research

3988 Streptococcus agalactiae: Identification methods, antimicrobial susceptibility, and resistance genes in pregnant women

Santana FAF, de Oliveira TVL, Filho MBDS, da Silva LSC, de Brito BB, de Melo FF, Souza CL, Marques LM, Oliveira MV

3999 Twelve-month evaluation of the atraumatic restorative treatment approach for class III restorations: An interventional study

Shivanna MM, Ganesh S, Khanagar SB, Naik S, Divakar DD, Al-Kheraif AA, Jhugroo C

Case Control Study

4010 Effects of different doses of metformin on bone mineral density and bone metabolism in elderly male patients with type 2 diabetes mellitus

Wang LX, Wang GY, Su N, Ma J, Li YK



World Journal of Clinical Case	
Conter	nts Semimonthly Volume 8 Number 18 September 26, 2020
4017	Relationship between granulomatous lobular mastitis and methylene tetrahydrofolate reductase gene polymorphism
	Lei QR, Yang X, Miao CM, Wang JC, Yang Y
	Retrospective Cohort Study
4022	First-line chemotherapy in very elderly patients with metastatic pancreatic cancer: Gemcitabine monotherapy vs combination chemotherapy
	Han SY, Kim DU, Seol YM, Kim S, Lee NK, Hong SB, Seo HI
	Retrospective Study
4034	Pre- and intraoperative predictors of acute kidney injury after liver transplantation
	Mrzljak A, Franusic L, Pavicic-Saric J, Kelava T, Jurekovic Z, Kocman B, Mikulic D, Budimir-Bekan I, Knotek M
4043	Clinical value of needleless sling in treatment of female stress urinary incontinence
	Chen YG, Zhang YG, Zhang W, Li X, Wang X
4051	Intratympanic dexamethasone injection for sudden sensorineural hearing loss in pregnancy
	Lyu YL, Zeng FQ, Zhou Z, Yan M, Zhang W, Liu M, Ke ZY
4059	Research on the effect of health care integration on patients' negative emotions and satisfaction with lung cancer nursing activities
	Long FJ, Chen H, Wang YF, He LM, Chen L, Liang ZB, Chen YN, Gong XH
4067	Comparison between computed tomography and magnetic resonance imaging in clinical diagnosis and treatment of tibial platform fractures
	Liu XD, Wang HB, Zhang TC, Wan Y, Zhang CZ
	SYSTEMATIC REVIEWS
4075	Primary sclerosing cholangitis and autoimmune hepatitis overlap syndrome associated with inflammatory bowel disease: A case report and systematic review
	Ballotin VR, Bigarella LG, Riva F, Onzi G, Balbinot RA, Balbinot SS, Soldera J
	CASE REPORT
4094	Epidermolytic acanthoma: A case report
	Ginsberg AS, Rajagopalan A, Terlizzi JP
4100	Management of pembrolizumab-induced steroid refractory mucositis with infliximab: A case report
	Dang H, Sun J, Wang G, Renner G, Layfield L, Hilli J
4109	Small bowel obstruction caused by a bezoar following an adult simultaneous liver-kidney transplantation: A case report
	Pan G, Kim RD, Campsen J, Rofaiel G
4114	Laparoscopic resection of primary retroperitoneal schwannoma: A case report
	Ribeiro Jr MAF, Elias YGB, Augusto SDS, Néder PR, Costa CT, Maurício AD, Sampaio AP, Fonseca AZ

	World Journal of Clinical Cases	
Conter	semimonthly Volume 8 Number 18 September 26, 2020	
4122	Sweet syndrome as a paraneoplastic manifestation of cholangiocarcinoma: A case report	
	Lemaire CC, Portilho ALC, Pinheiro LV, Vivas RA, Britto M, Montenegro M, Rodrigues LFDF, Arruda S, Lyra AC, Cavalcante LN	
4128	Multidisciplinary approach to suspected sudden unexpected infant death caused by milk-aspiration: A case report	
	Maiese A, La Russa R, Arcangeli M, Volonnino G, De Matteis A, Frati P, Fineschi V	
4135	Stress fractures in uncommon location: Six case reports and review of the literature	
	Ficek K, Cyganik P, Rajca J, Racut A, Kiełtyka A, Grzywocz J, Hajduk G	
4151	Celiac disease and Sjögren's syndrome: A case report and review of literature	
	Balaban DV, Mihai A, Dima A, Popp A, Jinga M, Jurcut C	
4162	Nonasthmatic eosinophilic bronchitis in an ulcerative colitis patient – a putative adverse reaction to mesalazine: A case report and review of literature	
	Cernomaz AT, Bordeianu G, Terinte C, Gavrilescu CM	
4169	Insulinoma presenting with postprandial hypoglycemia and a low body mass index: A case report	
	Prídavková D, Samoš M, Kyčina R, Adamicová K, Kalman M, Belicová M, Mokáň M	
4177	Neoadjuvant chemoradiotherapy for locally advanced gastric cancer with bulky lymph node metastasis: Five case reports	
	Nomura E, Kayano H, Machida T, Izumi H, Yamamoto S, Sugawara A, Mukai M, Hasebe T	
4186	Unilateral pleuroparenchymal fibroelastosis as a rare form of idiopathic interstitial pneumonia: A case report	
	Lee JH, Jang HJ, Park JH, Kim HK, Lee S, Kim JY, Kim SH	
4193	Superior mesenteric vein thrombosis induced by influenza infection: A case report	
	Oh GM, Jung K, Kim JH, Kim SE, Moon W, Park MI, Park SJ	
4200	Mucinous adenocarcinoma of the buttock associated with hidradenitis: A case report	
	Kim SJ, Kim TG, Gu MJ, Kim S	
4207	<i>TFE3</i> -expressing malignant perivascular epithelioid cell tumor of the mesentery: A case report and review of literature	
	Kim NI, Lee JS, Choi YD, Ju UC, Nam JH	
4215	Robotic surgery in giant multilocular cystadenoma of the prostate: A rare case report	
	Fan LW, Chang YH, Shao IH, Wu KF, Pang ST	
4223	Multiple recurrent neurofibromas in the abdominal wall: A case report	
-	Zhao XF, Shen YM, Chen J	
4228	Mine disaster survivor's pelvic floor hernia treated with laparoscopic surgery and a perineal approach: A case report	
	Chen K, Lan YZ, Li J, Xiang YY, Zeng DZ	

Conter	World Journal of Clinical Cases	
Conter	Semimonthly Volume 8 Number 18 September 26, 2020	
4234	Successful treatment of encrusted cystitis: A case report and review of literature	
	Fu JG, Xie KJ	
4245	Massive pulmonary haemorrhage due to severe trauma treated with repeated alveolar lavage combined with extracorporeal membrane oxygenation: A case report	
	Zhang BY, Chen XC, You Y, Chen M, Yu WK	
4252	Gitelman syndrome caused by a rare homozygous mutation in the SLC12A3 gene: A case report	
	Yu RZ, Chen MS	
4259	Arterial embolism caused by a peripherally inserted central catheter in a very premature infant: A case report and literature review	
	Huang YF, Hu YL, Wan XL, Cheng H, Wu YH, Yang XY, Shi J	
4266	Left bundle branch pacing with optimization of cardiac resynchronization treatment: A case report	
	Zhang DH, Lang MJ, Tang G, Chen XX, Li HF	
4272	Lymphoplasmacyte-rich meningioma with atypical cystic-solid feature: A case report	
	Gu KC, Wan Y, Xiang L, Wang LS, Yao WJ	



Contents

Semimonthly Volume 8 Number 18 September 26, 2020

ABOUT COVER

Editorial board member of World Journal of Clinical Cases, Dr. Li is a Professor at the Nanjing University Medical School in Nanjing, China. Having received his Bachelor's degree from Xuzhou Medical College in 1997, Dr. Li undertook his postgraduate training first at Nanjing Medical University, receiving his Master's degree in 2004, and then at Fudan University, receiving his PhD in 2007. He advanced to Chief Physician in the Department of Anesthesiology at The Affiliated Hospital of Nanjing University Medical School in 2017 and has held the position since. His ongoing research interests involve ultrasound (transthoracic echo and transesophageal echo) in clinical anesthesia and ultrasound-guided limb and trunk nerve block in postoperative pain management. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of World Journal of Clinical Cases (WJCC, World J Clin Cases) is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The WJCC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, PubMed, and PubMed Central. The 2020 Edition of Journal Citation Reports® cites the 2019 impact factor (IF) for WJCC as 1.013; IF without journal self cites: 0.991; Ranking: 120 among 165 journals in medicine, general and internal; and Quartile category: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ji-Hong Liu; Production Department Director: Xiang Li; Editorial Office Director: Jin-Lei Wang.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Clinical Cases	https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 2307-8960 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
April 16, 2013	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Semimonthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Dennis A Bloomfield, Sandro Vento, Bao-Gan Peng	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/2307-8960/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
September 26, 2020	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2020 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2020 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



W J C C World Journal of Clinical Cases

World Journal of

Submit a Manuscript: https://www.f6publishing.com

World J Clin Cases 2020 September 26; 8(18): 3942-3955

DOI: 10.12998/wjcc.v8.i18.3942

ISSN 2307-8960 (online)

REVIEW

From infections to autoimmunity: Diagnostic challenges in common variable immunodeficiency

Ewa Więsik-Szewczyk, Karina Jahnz-Różyk

ORCID number: Ewa Więsik-Szewczyk 0000-0001-8509-4453; Karina Jahnz-Rózyk 0000-0002-3505-1858.

Author contributions: Więsik-Szewczyk E wrote the manuscript; Jahnz-Różyk K supervised the work

Conflict-of-interest statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/licenses /by-nc/4.0/

Manuscript source: Invited manuscript

Received: May 6, 2020

Ewa Więsik-Szewczyk, Karina Jahnz-Różyk, Department of Internal Medicine, Pulmonology, Allergy and Clinical Immunology, Central Clinical Hospital of the Ministry of National Defense, Military Institute of Medicine, Warsaw 04-141, Poland

Corresponding author: Ewa Więsik-Szewczyk, MD, PhD, Doctor, Department of Internal Medicine, Pulmonology, Allergy and Clinical Immunology, Central Clinical Hospital of the Ministry of National Defense, Military Institute of Medicine, Szaserów 128, Warsaw 04-141, Poland. ewa.w.szewczyk@gmail.com

Abstract

Common variable immunodeficiency (CVID) is the most common clinically significant primary antibody deficiency diagnosed in adults. The early symptoms are not specific. They include common infections, mainly of the respiratory tract, caused by typical microorganisms, so cases can be missed in primary care. In the majority of patients increased susceptibility to infections coexists with signs or symptoms of autoimmunity, inflammation or polyclonal lymphoproliferation, which can divert diagnosis from immune deficiency. The overall incidence of malignancy is increased in CVID and certain cancers are significantly more common. Lymphomas and gastric carcinoma are the most frequently reported malignancies in CVID, so a high index of suspicion is recommended. Diagnostic delay in CVID is seen worldwide. The main goal of this paper is to increase the awareness about CVID among health care professionals. We aim to present features which can be helpful in CVID diagnosis in order to shorten the "latency" of proper management of CVID patients. We review clinical symptoms, complications and laboratory abnormalities of CVID. Immunoglobulin replacement therapy is regarded as the cornerstone of pharmacological intervention. New modes of Ig application, mainly subcutaneously and via the hyaluronidase-facilitated subcutaneous route, help to adjust therapy to patients' needs and preferences. Still there remain unmet needs. It remains to be seen whether CVID complications can be avoided by earlier diagnosis, treatment and thorough monitoring in the context of increased risk of malignancy. Development of patient tailored protocols depending on the clinical phenotype and risk factors might be more appropriate. The most important consideration is to diagnose suspected cases and stratify patients in a precise and timely way. Work is needed to define features predictive of unfavorable prognosis.

Key Words: Primary antibody deficiency; Recurrent respiratory tract infections;



Balabidena® WJCC | https://www.wjgnet.com

Peer-review started: May 6, 2020 First decision: May 15, 2020 Revised: May 29, 2020 Accepted: August 26, 2020 Article in press: August 26, 2020 Published online: September 26, 2020

P-Reviewer: Carbone I S-Editor: Ma YI L-Editor: A P-Editor: Liu JH



Complications; Immunoglobulin replacement; Adults; Early diagnosis

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Common variable immunodeficiency has to be considered in adults with recurrent respiratory tract infections, bronchiectasis, cytopenia, generalized lymphadenopathy, sarcoid-like symptoms, colitis in whom other conditions were excluded. Low calculated globulin, which is the difference between total protein and albumin levels, suggests hypogammaglobulinemia. Patient with common variable immunodeficiency are referred for commencing replacement immunoglobulin therapy, which should be continued regularly throughout life. It can be administered intravenously, subcutaneously and via the hyaluronidase-facilitated subcutaneous route. One must bear in mind that in patients with hypogammaglo-bulinemia and those treated with polyclonal immunoglobulin G products, serological tests based on determination of antibodies are unreliable.

Citation: Więsik-Szewczyk E, Jahnz-Różyk K. From infections to autoimmunity: Diagnostic challenges in common variable immunodeficiency. World J Clin Cases 2020; 8(18): 3942-3955 URL: https://www.wjgnet.com/2307-8960/full/v8/i18/3942.htm DOI: https://dx.doi.org/10.12998/wjcc.v8.i18.3942

INTRODUCTION

Primary immunodeficiencies (PID) are rare diseases. More than half of PID cases are associated with a defect in antibody production or function^[1]. In this group, the most common clinically significant deficiency is common variable immunodeficiency (CVID). Although CVID is innate condition, the peak of disease onset occurs at the age of 20-40, and the condition can be diagnosed even in elderly patients^[2]. Early symptoms are not specific. They include common infections, mainly of the respiratory tract, caused by typical microorganisms, so cases can be missed in primary care. In a significant percentage of patients increased susceptibility to infections coexists with signs or symptoms of autoimmunity or polyclonal lymphoproliferation, which seems to be a paradox. Patients with CVID are consulted by doctors specializing in various areas - hematology, rheumatology, gastroenterology for specific symptoms while the common denominator, immunodeficiency and hypogammaglobulinemia, is overlooked. Indeed, diagnostic delay is seen worldwide^[3]. The delay in diagnosis can lead to organ damage, in particular bronchiectases. In long term follow-up malignancy is also more common among patients with CVID in comparison with healthy population. Unrecognized CVID generates high economical cost^[4].

The main goal of this paper is to increase the awareness about CVID among health care professionals. We aim to present features which can be helpful in CVID diagnosis in order to shorten the "latency" of proper management of CVID patients.

EPIDEMIOLOGY AND DEMOGRAPHY

CVID prevalence ranges from 2/100000 to the highest reported prevalence for Finland - 6.9/100000^[5]. The median age at which the first symptoms occurred in a case series of Polish patients is 24 years^[6], in Italy and Denmark 32 and 29 years^[7], respectively, while in the 2018 European Society of Immune Deficiencies (ESID) registry the figure is 18 years^[3]. A recent study from the United States reported 24 and 28 years as the median age of onset for females and males, respectively^[8]. Some studies report bimodal age distribution^[6,9] where others, reported linear distribution, with decline after the age of 80^[10]. In Germany, the highest percentage of patients experience the first symptoms of CVID between 1 and 5 years, followed by a markedly lower peak incidence at the age of 16-20 years^[11]. Current data indicate that the majority of patients are diagnosed in adulthood (age \geq 18) comprising between 65% and nearly 90% in reported cohorts. The significant variations in epidemiology and demographic data are related to differences between cohorts surveyed, especially with regard to the proportion of adult to pediatric patients, as well as diagnostic criteria used, which are



discussed below. From 2012, when new PID ESID criteria were proposed, professionals tend to identify more cases of late-onset combined immune deficiencies in adults^[12].

The literature informs of a long delay in diagnosing CVID. In a recent report, the overall delay was 5 years in Europe and remained constant over the last 20 years^[3]. A Danish cohort had a mean diagnostic delay of 7 years^[7] and in an Italian one, the figure was 8.9 years^[13]. The longest diagnostic delay of 10.13 years was reported for adult patients with CVID in Poland^[14]. However it was significantly shorter in patients diagnosed before age 18: 5.22 years compared to 11.63 years for patients diagnosed in adulthood (age \geq 18), despite similar clinical phenotypes in children and adults^[14]. At present, in Poland, CVID diagnosis is more rapid than that before 2000^[6]. However, there remains a low percentage of patients whose diagnosis was established within a year of occurrence of the first symptoms^[6]. According to the 2014 ESID registry, a significant shortening of the median delay was achieved only in Spain (9.0 vs 4.6 years)^[2]. There is a need for further research on the relationship between delayed diagnosis and occurrence of complications. It is difficult to compare data coming from different countries and studies, due to different methodology applied for analysis. In our experience, a significant increase in number of diagnosed CVID cases occurred when reimbursement of subcutaneous immunoglobulin treatment for adult patients with primary immunodeficiency was provided and specialized immunology centers established^[6,14].

Taken together, these data suggest that the level of awareness and suspicion of CVID as a differential diagnosis remains poor.

CLINICAL FEATURES

Infections

The hallmark of CVID is susceptibility to infections which affect 99 to 70 per cent of patients. Infections are the leading cause of mortality among patients with primary antibody deficiencies in the United States of America^[15]. The reported etiologies of recurrent respiratory tract infections include common bacteria: Streptococcus pneumoniae, Hemophilus influenzae, Moraxella catarrhalis, Mycoplasma pneumoniae, Chlamydophila pneumoniae and Staphylococcus aureus^[16]. Common sites of infection include paranasal sinuses, middle ear, bronchi and lungs. Gastritis and enterocolitis are less common. The typical causative agents in the alimentary tract are: Giardia lamblia, Campylobacter jejuni, Salmonella enteritidis but some atypical pathogens can be identified, for example: Campylobacter pylori and Aeromonas hydrophila. The differential work-up of enterocolitis must also include viral etiology, including norovirus, which is spontaneously eliminated by immunocompetent persons^[17]. Cytomegalovirus (CMV) has been found in gut biopsies of CVID patients^[18]. CMV enteritis in CVID can lead to severe malabsorption and irreversible chronic damage of the gastro-intestinal tract, which can require parenteral nutritional support^[19].

In contrast to high risk of common bacterial infections patients with CVID do not present an increased susceptibility to influenza infection. Moreover most of them have preserved antigen specific T cell responses after influenza vaccination^[20]. Data about the clinical course of coronavirus disease 2019 (COVID-19) in patients with primary immunodeficiency remains limited. The first, published case series included 5 patients with CVID and 2 patients with agammaglobulinemia^[21]. Patients with CVID presented with a severe form of COVID-19 infection. They required multiple drug treatment, including antiretroviral agents, IL-6 blocking drugs, and mechanical ventilation. In contrast two patients with agammaglobulinemia and absent B cells, had mild symptoms. Authors speculate that strikingly different clinical course of COVID 19 might be explained by a possible role of B lymphocytes in the severe acute respiratory syndrome coronavirus 2 induced inflammation, assuming that patients with agammaglobulinemia lack B lymphocytes whereas patients with CVID have dysfunctional B lymphocytes^[21].

If patients remain untreated, they suffer an increase of frequency and severity of infections with increasing duration of CVID. Patients often require hospitalization and intravenous antibiotics. Bronchiectases are a long-term sequel of recurrent lung infections in CVID^[22]. Still a significant percentage of patients are diagnosed when bronchiectases are already present^[23].

Sepsis is a relatively rare complication of CVID^[14]. Opportunistic infections are more characteristic for combined B and T cell immunodeficiencies than for CVID^[12].



Cumulative incidence of opportunistic infections in CVID is reported between 5.4% and 9.2%, and Pneumocystis jirovecii, varicella zoster virus, *Mycobacterium species* (spp.), *Aspergillus* spp., *Toxoplasma gondii*, and *Candida* spp. are the most common pathogens^[24]. In our CVID cohort we found 1 opportunistic infection and 1 infection caused by an unusual pathogen^[14]. The former case was a severe pneumonia caused by Aspergillus spp. and the latter, a sepsis due to an atypical pathogen: Achromobacter denitrificans. Both patients initially were missed as CVID cases and underwent immunosuppressive treatment for their presenting conditions. The first patient received biological and conventional diseases modifying anti-rheumatic drugs due to psoriatic arthritis^[25]. The second patient was treated with mycophenolate mofetil due to retinitis with granulomas^[26]. It has been found that the risk of opportunistic infections in CVID patients increases with secondary comorbidities and their management^[24].

Microbiological assessment is a crucial element of the diagnostic work-up of patients with CVID. It should be emphasized that in patients with hypogammaglobulinemia and those treated with polyclonal immunoglobulin G (IgG) products, serological tests based on determination of antibodies in IgG class are unreliable. It is therefore obligatory to collect appropriate cultures or perform polymerase chain reaction assays in search of the infectious agent.

Non-infectious complications

The clinical picture of CVID is typically composed of multiple non-infectious pathologies. A non-complicated phenotype, limited to infections, is reported in only up to 30% of patients^[8,14]. Non-infectious pathologies involve autoimmune complications, both organ-specific and general, lymphoid hyperplasia, granulomas and malignancy. In long term follow-up, 68% of patients had at least 2 complications^[8]. These non-infectious complications can precede recurrent infections and be the first presentation of immune dysregulation in CVID^[27,28]. They can also appear simultaneously with infections or later, during follow up. According to published studies, disease complications had a negative impact on patient survival^[10,29,30]. Resnick *et al*^[29] estimated the risk of death to be 11 times higher in complicated *vs* non-complicated CVID.

Autoimmunity: According to reports, autoimmune diseases occur in up to 30% of CVID cohorts. In a recent study focused on noninfectious CVID complications, among 632 patients followed since 1974, autoimmune thrombocytopenic purpura (AITP) was the most common (16.2%), followed by autoimmune hemolytic anemia (AIH 7.7%). Other associated autoimmune conditions include rheumatoid arthritis (2.7%) and uveitis (1%). Rarer autoimmune complications are psoriasis, psoriatic arthritis, vitiligo, alopecia, autoimmune thyroiditis, antiphospholipid syndrome, Sjogren syndrome, vasculitis, type 1 diabetes, myasthenia gravis, autoimmune pancreatitis, severe oral ulcers^[8,10,14]. In one, although small, series of CVID patients the median age of AITP diagnosis was 23 years, while of CVID was 27 years. CVID was diagnosed before AITP in only 19% of patients^[28]. It is recommended to assess IgG level in patients with autoimmune cytopenia.

Gastrointestinal involvement: Gastrointestinal (GI) pathologies in CVID may involve any part of the gastrointestinal tract. They affect from 5% to 20% of patients in published cohort studies^[8,14]. Gastrointestinal symptoms are difficult to differentiate from signs of inflammation due to infectious or non-infectious etiology (colitis-like or celiac-like disease), all the more so since both conditions can coexist.

Non-infectious background of GI symptoms should be searched for after infectious etiology is excluded. The main symptoms are bloating, pain, chronic diarrhea (lasting more than 2 wk) and malabsorption. Comprehensive biopsies revealed lymphocytosis, villous atrophy/blunting, nodular lymphoid hyperplasia, granulomas and non-specific inflammation. If CVID presents with gastrointestinal symptoms, the case is commonly misdiagnosed as celiac disease. The features, which support differential CVID diagnosis, are absence of plasma cells in the histological assessment, negative celiac genetic testing and lack of response to gluten-free diet^[31]. Another diagnostic pitfall is inflammatory bowel disease. Current data highlights the relevance of the endoscopic and histological assessment in CVID patients presenting with chronic diarrhea. The vigilance towards CVID in patients with enterocolitis and recurrent infections is recommended^[32] and immunology centers should agree on a protocol with the local gastroenterology team to guide observations and indicate the pathological samples to be taken at endoscopy in adults with CVID^[33].

Zaishideng® WJCC | https://www.wjgnet.com

Polyclonal lymphoid infiltration: Polyclonal lymphadenopathy occurred in 20% to 40% of patients^[8]. Clinically, this often presents as generalized lymphadenopathy and splenomegaly. The challenge is to differentiate it from lymphoma. Diagnostic excision of a whole lymph node is recommended. Lymph nodes show atypical lymphoid hyperplasia, reactive lymphoid hyperplasia or granulomatous inflammation^[16]. In our experience, we have to repeat histologic evaluation serially in one patient.

Granulomas are less common, but can be located in any organ and may lead to irreversible damage. Granulomas in lungs, liver, skin or eyes can be diagnosed as sarcoidosis^[34]. This highlights the need to include serum main immunoglobulin assessment in a work-up of patients with sarcoidosis. In a subset of patients, lung granulomas can also be accompanied by an intense lymphocytic infiltration; a condition described as granulomatous lymphocytic interstitial lung disease (GLILD)^[35,36]. According to the definition proposed by British Lung Foundation/ United Kingdom Primary Immunodeficiency Network, GLILD is a distinct clinicoradio-pathological interstitial lung disease (ILD) occurring in patients with CVID, associated with lymphocytic infiltration and/or granuloma of the lung, and in whom other conditions had been considered and, where possible, excluded^[37].

Clinically, patients with GLILD experience new or worsening, often productive cough, exertional dyspnea and fatigue. On physical examination, inspiratory crackles are present in 50% of patients^[34]. There is universal agreement for full lung function assessment (spirometry, lung volumes, gas exchange)[37]. In one study including 20 patients with confirmed histopathological GLILD, a restrictive ventilatory defect (total lung capacity < 80% predicted) was found in 33.3% patients and an obstructive defect (forced expiratory volume in 1 s/vital capacity < 70%) in 10 % of patients while an impairment of carbon monoxide diffusing capacity (DLCO <80%) was observed in 70% patients^[34].

Chest computerized tomography (CT) is an essential diagnostic step, where discovery of new abnormalities, which closely correlate with clinical signs and symptoms, is very suggestive. If conventional CT images are stable, but the patient's condition deteriorates, positron emission tomography (PET) can add value^[38]. The most common CT finding in GLILD was nodules, often multiple with air bronchograms and smooth margins possibly surrounded by ground-glass opacities (halo sign). In one study, micronodules were present in 60% of patients, had a random distribution in 64% cases and were perilymphatic in 36% cases. Neither the hallmarks nor their constellation seems to be sufficiently specific to establish diagnosis^[38-40].

A recommended approach is to confirm the diagnosis of GLILD at surgical lung biopsy^[41], but this is not always available^[23]. Diagnosis has to be established by a histopathologist experienced in PID in cooperation with a clinician. The presence of granulomatous inflammation, peribronchiolar lymphoid proliferation, interstitial lymphoid proliferation, and CD4-cell predominance are described, but this is not a universal phenomenon^[41]. Flexible bronchoscopy helps to exclude infection^[37]. The extent of microbiological work-up differs among centers. Recommended essential tests are microscopy and culture, Mycobacterial culture, fungal culture. There is some debate about PCR for Mycobacteria, atypical bacteria, respiratory viruses, Pneumocystis jirovecii and these should be assessed in selected cases^[37]. Some studies indicate that bronchoalveolar lavage can be helpful in assessing GLILD activity and selecting possible treatment. There is some evidence that lymphocytes exceeding 20% of cell counts in BAL material correlate with GLILD diagnosis. There is also growing evidence that GLILD, lymphadenopathy, splenomegaly, hepatomegaly and chronic diarrhea are seen together and that they share common pathology^[42,43].

Up to now the cause and exact significance of this interstitial lung disease is poorly understood and its management is a challenge in clinical practice. Center experiences vary and even the natural history of GLILD is not well described. Patients are often put on corticosteroids with other immunosuppressive therapies and rituximab as a second line treatment^[35,44]. The STILPAD study is a multi-center observational study looking at the effect of various treatments on interstitial lung disease (ILD) in patients with CVID and ILD that had been diagnosed by a chest CT scan^[45]. In selected patients, there may be a role for FDG PET-CT imaging in view of the sensitivity of combined anatomical and functional information obtained, but more data are needed to endorse this modality.

Malignancy: The overall incidence of malignancy is increased in CVID and certain cancers are significantly more common. Lymphomas are the most frequently reported malignancies in CVID^[46,47]. The majority of lymphomas are of the B cell non-Hodgkin's type (90%)^[8]. Childhood onset and previous polyclonal lymphoproliferation are



predictors identified for lymphomas^[10]. Gastric carcinoma is common as well. In a Scandinavian study, the observed to expected ratio for stomach cancer was 10:3^[48]. In an Australian study a standardized incidence ratio for gastric carcinoma was 7.3^[49]. Standardized mortality ratio indicated a 10.1-fold excess mortality among CVID patients with gastric cancer^[50]. A gastric screening protocol was proposed^[51]. This is to include a gastroscopy in individuals with CVID with any of the following risk factors: Age greater than 50, CVID diagnosis since more than 10 years, significant smoking history, family history of gastric carcinoma, persistent diarrhea or upper gastrointestinal symptoms, previous gastroscopy with inadequate biopsies, positive Helicobacter pylori breath test, iron deficiency, and low serum vitamin B12. As CVID patients also have an increased risk of gastrointestinal lymphoma, it seems reasonable to perform multiple biopsies from the gastric antrum and body and additionally from the second part of the duodenum. It is reported that fewer biopsies decreased the probability of detecting early premalignant lesions^[51]. Dhalla et al^[52] suggested to perform upper endoscopy with an interval between the subsequent endoscopic assessment based on histological findings: Every 1-3 years in patients with metaplasia, every 3 years in patients with atrophic gastritis, and every 6-12 mo in those with dysplasia. However, some CVID patients developed a high-grade gastric cancer 12-14 mo after an endoscopy that had shown no histologic signs of dysplasia. Based on this data a yearly evaluation for stomach cancer in all CVID patients might be beneficial^[50,53]. Development of further guideline/consensus on screening and monitoring for gastric cancer and lymphomas is required.

Other malignancies occurred in patients diagnosed in adulthood and according to published data are more likely to develop in the fifth and sixth decade^[54]. Examples are pituitary gland adenoma, colon tubular adenocarcinoma, squamous cell carcinoma (lungs), basal cell carcinoma, intestinal tubular adenoma (low grade), breast adenocarcinoma, metaplasia of the cervix^[14].

CLINICAL PHENOTYPES

The literature defines five clinical phenotypic categories: Autoimmunity (including organ-specific autoimmune conditions and cytopenia); polyclonal lymphocytic infiltration (including unexplained granuloma, unexplained hepatomegaly, persistent lymphadenopathy, and lymphoid interstitial pneumonia); lymphoid malignancy (proven and treated); unexplained enteropathy (biopsy-proven and gluteninsensitive); and no disease-related complications^[10].

LABORATORY MANIFESTATIONS

Despite the heterogeneity of CVID, low IgG levels are found in most patients. At diagnosis, only a minority of patients had $IgG \ge 500 \text{ mg/dL}$, a significant percentage had less than 100 mg/dL and up to 20% had undetectable IgG, IgA and IgM^[14,55]. As normal IgG and IgA levels exclude CVID, quantification of serum Ig levels is a useful initial laboratory test^[14,55]. Protein electrophoresis can draw attention to a low gamma fraction.

Immunoglobulin levels are not always available for primary care. Calculated globulin (CG) can be an alternative option. CG is the difference between total protein and albumin levels. If a CG result is ≤ 17 g/L, the probability of hypogammaglobulinemia increases^[56-58] and the patient is suitable for further immunologic evaluation.

Specific antibody production and isohemaglutinins

Specific antibody production reveals a function of adaptive immunology. It is impaired in the majority of CVID patients.

Currently, the measurement of specific antibody responses varies across countries. It should include evaluation of both T-dependent responses (to protein or glycoprotein antigens) and T-independent responses (to polysaccharide antigens). The procedure involves two steps. First, an initial protective antibody assay is done. If the titer is low, immunization is performed, and a second measurement of protective antibody titer made after 3-6 wk (optimal at 4 wk). Diphtheria and tetanus toxoid vaccines are the most commonly used vaccines to evaluate T-cell dependent responses. Specific antibody T independent responses are mostly measured after vaccination of PPV23 (referred to as test immunization). The vaccination response in adults is satisfactory if



at least 70% of the measured serotype-specific antibody titers are above 1.3 µg/mL or a four-fold increase of the pre-vaccination titers for more than 17 of 23 serotypes is observed at week 4 after immunization^[59]. An alternative to PPV is the measurement of response after immunization against Salmonella typhi^[60,61]. Isohemagglutinins are naturally occurring antibodies of IgM and IgG isotypes to polysaccharide blood group antigens. Isohemagglutinins may be an alternative, clinically relevant, and easily accessible analysis to assess T-independent antibody function in patients with a group other than AB^[55].

Lymphocyte subsets

CVID is a primary antibody defect, so naturally attention is focused on B lymphocyte analysis. Immunophenotyping of the B cell compartment is a routine evaluation of patients with CVID. Low Ig switched memory B-cell (smB) (CD19+CD27+IgM-IgD-) is an abnormality present in 80% of patients, however it is not specific for CVID^[57]. Other B lymphocyte subsets are highly variable. Findings can be classified according several systems, of which two are the most popular: Freiburg^[62] and EUROclass^[63]. Both strategies apply characterization of CD19+ B cell subsets. Low Ig sm B-cell (smB) proportions and abnormally high proportions of B cells with reduced CD21 expression (CD21^{low} B cells) help to distinguish subsets. In addition to these two cell subsets, the EUROclass classification also uses abnormal expansion of transitional B cells (CD19+CD27-CD38+) for further subgrouping.

Multiple studies have demonstrated disturbances in T and natural killer (NK) cell homeostasis. Decreased circulating NK cell numbers in CVID patients were found to be associated with severe bacterial infections and granulomas^[64,65]. T cells play a central role in B cell activation and differentiation into memory and IgG producing B cells. Various reports have associated CVID with other findings, such as CD4 T-cell lymphopenia with reduced counts of naive CD4 T cells^[66] and naive CD8mT-cell^[67]. Regulatory T cells (Treg), Th17, and follicular T helper (Tfh17) cells were specifically reduced in patients with complicated CVID phenotypes^[68]. T cells in CVID have lower proliferative capacities^[69] and abnormal cytokine production^[70]. Low (naïve) CD4 T cells are associated with complications and a poor prognosis in CVID^[71]. Recent publications show an involvement of follicular T cells in CVID pathogenesis^[72]. An increase of circulating memory CXCR5+ CD4 T cells in CVID patients with noninfectious complications has been reported^[73]. Follicular T helper cells could participate in autoimmune manifestations through their role as B cell inducers. As a consequence patients with CVID and autoimmune phenomena exhibit hyperplastic however insufficient germinal center (GC) response. An enrichment of T cells in lymph nodes of patients with CVID with lymphadenopathy and preserved ability to form GCs has been found^[74]. Taken together these data suggest that some distinct pathophysiological alterations are present in a defined subgroup of CVID patients. Work has also been done to better understand correlations between B cell and T cell abnormalities with unique clinical profiles and to use them as a prognostic marker^[68,75].

Real life data revealed that full diagnostic work-up is not always performed. Gaps in laboratory testing were identified. A minority of CVID patients (20%) had been assessed for response to vaccination[7,14]. Problems that need to be solved include test standardization, lack of polysaccharide pneumococcal vaccine on the market, and lack of reimbursement of vaccines for adult patients, challenges in the interpretation of results. Of all reported patients in a Danish cohort 21% had isohemagglutinin titers measured and only 17.9% were characterized by flow cytometric B cell phenotyping^[7].

Genetics

CVID is a sporadic disease, polygenic in nature. A genome-wide association study identified multiple loci with potential susceptibility for CVID, however their significance in clinical practice is unclear. Currently molecular genetic analysis of patients is not a requirement for the diagnosis^[55]. However, pathogenic monogenic variants are found in 2%-30% of screened CVID populations, depending of applied clinical criteria, frequency of parents consanguinity, proportion of familial cases and genetics methods used. Whole exome sequencing (WES) is an effective tool for the discovery of genetic defects in patients with primary immunodeficiencies (PIDs). In 2016 WES was performed on 50 subjects diagnosed with CVID^[76]. Subjects selected for analysis had at least one of the following: Early-onset of manifestations (under age 10) (36%), autoimmune/inflammatory manifestations (76%), low B lymphocyte counts (58%), and/or familial history of hypogammaglobulinemia (16%). With this approach 17 probable disease-causing mutations in 15 patients (30%) were identified. There were mutations in NFKB1, STAT3, CTLA4, PIK3CD, and IKZF1, and biallelic mutations in LRBA and STXBP2. This suggests that WES may be useful in selected CVID



phenotypes. It remains to be seen whether this approach generates change is management and will be beneficial in daily practice^[77].

DIAGNOSIS

There is an ongoing debate about criteria for diagnosis of CVID^[78]. First were proposed by European Society of Immune Deficiencies (ESID)/ Pan-American Group for Immune Deficiency in 1999^[79]. Ameratunga in 2013^[80], ESID in 2014^[12] and International Consensus Document in 2016^[55] proposed new criteria (Table 1). They include features and laboratory parameters which illustrates progress of knowledge and understanding in the pathogenesis of CVID. All criteria may be applied only if secondary causes of hypogammaglobulinemia have been excluded^[81]. Particular domains, features or parameters of diagnostic criteria must be applied sequentially as none is specific individually. Each definition has limitations which are analyzed in a review paper by Ameratunga et al^[78]. In our center we use ESID criteria^[12]. We believe that they are helpful for early diagnosis. In our practice they have a good discriminative value for exclusion of subjects with undifferentiated hypogammaglobulinemia. However ESID criteria are not recommended as an unique criterion for decision of initiation an Ig replacement therapy^[12,80,82].

CONCLUSION

CVID is heterogenous disease. An early and proper diagnosis of CVID is a challenge in everyday practice. It is recognized that early correct diagnosis avoids unnecessary diagnostic procedures and wrong treatments. In the last decade data from published registries and national cohorts of CVID patients increased our knowledge about CVID's natural history and the complexity of its clinical picture, beyond increased susceptibility to infections. Majority of patients have at least one non-infectious manifestation, such as autoimmunization, granulomatous lesions, unexplained polyclonal lymphoproliferation, enteropathy, or malignancy. Immunoglobulin replacement therapy is regarded as the cornerstone of pharmacological intervention. New modes of Ig application, mainly subcutaneously and via the hyaluronidasefacilitated subcutaneous route, help to adjust therapy to patients' need and preferences^[83,84]. Still there remain unmet needs. It remains to be seen whether CVID complications can be avoided by earlier diagnosis, treatment and thorough monitoring in context of increased risk of malignancy. Development of patient tailored protocols depending on the clinical phenotype and risk factors might be more appropriate. In our opinion, from a practical point of view, the most important consideration is to diagnose and stratify patients as precisely as possible from the moment of diagnosis. Predictive features which help identifying a subgroup of patients with unfavorable prognosis are needed.



Ameratunga <i>et al</i> ^[80] 20	
Must meet all major cri	eria:
Hypogammaglobuliner	nia IgG < 5 g/L
No other cause identifie	d for immune defect
Age > 4 yr	
Sequelae directly attrib	table to immune system failure (ISF) (1 or more):
Recurrent, severe or un	usual infections
Poor response to antibio	tics
Breakthrough infection	in spite of prophylactic antibiotics
Infections in spite of ap	propriate vaccination, e.g., human papilloma virus disease
Bronchiectasis and/or o	hronic sinus disease
Inflammatory disorders	or autoimmunity
Supportive laboratory e	vidence (3 or more criteria):
Concomitant reduction	or deficiency of IgA (< 0.8 g/L) and/or IgM (0.4 g/L)
Presence of B cells but r	educed memory B cell subsets and/or
Increased CD21 low sul	sets by flow cytometry
IgG3 deficiency (< 0.2 g	/L)
Impaired vaccine respo	nses compared with age-matched controls
Transient vaccine respo	nses compared with age-matched controls
Absent isohemagglutin	ns (if not blood group AB)
Serological evidence of	significant autoimmunity, e.g., Coombes test
Sequence variations of	enes predisposing to CVID, e.g., TNFRSF13B/TACI, TNFRSF13C/BAFFR and MSH5
Presence of relatively sp category A and B criter	ecific histological markers of CVID (not required for diagnosis but presence increases diagnostic certainty, in the context of a):
Lymphoid interstitial p	neumonitis
Granulomatous disorde	r
Nodular regenerative h	perplasia of the liver
Nodular lymphoid hyp	erplasia of the gut
Absence of plasma cells	on gut biopsy
ESID 2014	
At least one of the follo	ving:
Increased susceptibility	to infection
Autoimmune manifesta	tions
Granulomatous disease	
Unexplained polyclona	proliferation
Affected family membe	with antibody deficiency
AND	
Marked decrease of IgC	and marked decrease of IgA with or without low IgM levels (measured at least twice; < 2SD of the normal levels for their age)
AND	
At least one of the follo	ving:
Poor antibody response	to vaccines (and/or absent isohemaglutinins); <i>i.e.</i> , absence of protective levels despite vaccination were defined

Baisbideng® WJCC | https://www.wjgnet.com

September 26, 2020 Volume 8 Issue 18

Low switched memory B cells (< 70% of age-related normal value)

AND

Secondary causes of hypogammaglobulinemia has been excluded:

AND

Diagnosis is established after the fourth of year of life (but symptoms can be present earlier)

AND

No evidence of profound T-cell deficiency, defined as 2 of the following (y: Year of life):

CD4 numbers/microliter:

2-6 yr < 300, 6-12 yr < 250, 12 yr < 200

% naive CD4:

2-6 yr < 25%, 6-12 yr < 20%, 12 yr < 10%

T cell proliferation absent

ICON 2016

At least 1 of the characteristic clinical manifestations: Infection, autoimmunity, lymphoproliferation

OR

Asymptomatic individuals, especially in familial cases who fulfill criteria 2 to 5, hypogammaglobulinemia should be defined according to the age adjusted reference range for the laboratory in which the measurement is performed

The IgG level must be repeatedly low in at least 2 measurements more than 3 wk apart in all patients. may be omitted if the level is very low < 100-300 mg/dL depending on age, and other characteristic features are present, IgA or IgM level must also be low

There must be a demonstrable impairment of response to at least 1 type of antigen (TD or TI), at the discretion of the practitioner, specific antibody measurement may be dispensed with if all other criteria are satisfied and if the delay incurred by prevaccination and postvaccination antibody measurement is thought to be deleterious to the patient's health

Other causes of hypogammaglobulinemia must be excluded

Genetic studies to investigate monogenic forms of CVID or for disease-modifying polymorphisms are not generally required for diagnosis and management in most of the patients, especially those who present with infections only without immune dysregulation, autoimmunity, malignancy, or other complications

CVID: Common variable immunodeficiency; ESID: European Society of Immune Deficiencies; ICON: International consensus document.

REFERENCES

- Modell V, Orange JS, Quinn J, Modell F. Global report on primary immunodeficiencies: 2018 update from 1 the Jeffrey Modell Centers Network on disease classification, regional trends, treatment modalities, and physician reported outcomes. Immunol Res 2018; 66: 367-380 [PMID: 29744770 DOI: 10.1007/s12026-018-8996-5
- Gathmann B, Mahlaoui N; CEREDIH, Gérard L, Oksenhendler E, Warnatz K, Schulze I, Kindle G, 2 Kuijpers TW, Dutch WID, van Beem RT, Guzman D, Workman S, Soler-Palacín P, De Gracia J, Witte T, Schmidt RE, Litzman J, Hlavackova E, Thon V, Borte M, Borte S, Kumararatne D, Feighery C, Longhurst H, Helbert M, Szaflarska A, Sediva A, Belohradsky BH, Jones A, Baumann U, Meyts I, Kutukculer N, Wågström P, Galal NM, Roesler J, Farmaki E, Zinovieva N, Ciznar P, Papadopoulou-Alataki E, Bienemann K, Velbri S, Panahloo Z, Grimbacher B, European Society for Immunodeficiencies Registry Working Party. Clinical picture and treatment of 2212 patients with common variable immunodeficiency. J Allergy Clin Immunol 2014; 134: 116-126 [PMID: 24582312 DOI: 10.1016/j.jaci.2013.12.1077]
- Odnoletkova I, Kindle G, Quinti I, Grimbacher B, Knerr V, Gathmann B, Ehl S, Mahlaoui N, Van Wilder P, 3 Bogaerts K, de Vries E; Plasma Protein Therapeutics Association (PPTA) Taskforce. The burden of common variable immunodeficiency disorders: a retrospective analysis of the European Society for Immunodeficiency (ESID) registry data. Orphanet J Rare Dis 2018; 13: 201 [PMID: 30419968 DOI: 10.1186/s13023-018-0941-0]
- 4 Sadeghi B, Abolhassani H, Naseri A, Rezaei N, Aghamohammadi A. Economic burden of common variable immunodeficiency: annual cost of disease. Expert Rev Clin Immunol 2015; 11: 681-688 [PMID: 25804338 DOI: 10.1586/1744666X.2015.1029457]
- 5 Selenius JS, Martelius T, Pikkarainen S, Siitonen S, Mattila E, Pietikäinen R, Suomalainen P, Aalto AH, Saarela J, Einarsdottir E, Järvinen A, Färkkilä M, Kere J, Seppänen M. Unexpectedly High Prevalence of Common Variable Immunodeficiency in Finland. Front Immunol 2017; 8: 1190 [PMID: 29033928 DOI: 10.3389/fimmu.2017.01190]
- Ziętkiewicz M, Więsik-Szewczyk E, Matyja-Bednarczyk A, Napiórkowska-Baran K, Zdrojewski Z, Jahnz-6 Różyk K. Shorter Diagnostic Delay in Polish Adult Patients With Common Variable Immunodeficiency and Symptom Onset After 1999. Front Immunol 2020; 11: 982 [PMID: 32655544 DOI: 10.3389/fimmu.2020.00982]
- Westh L, Mogensen TH, Dalgaard LS, Bernth Jensen JM, Katzenstein T, Hansen AE, Larsen OD, Terpling 7



S, Nielsen TL, Larsen CS. Identification and Characterization of a Nationwide Danish Adult Common Variable Immunodeficiency Cohort. Scand J Immunol 2017; 85: 450-461 [PMID: 28370285 DOI: 10.1111/sii.125511

- Ho HE, Cunningham-Rundles C. Non-infectious Complications of Common Variable Immunodeficiency: 8 Updated Clinical Spectrum, Sequelae, and Insights to Pathogenesis. Front Immunol 2020; 11: 149 [PMID: 32117289 DOI: 10.3389/fimmu.2020.00149]
- Sanchez LA, Maggadottir SM, Pantell MS, Lugar P, Rundles CC, Sullivan KE; USIDNET Consortium. Two 0 Sides of the Same Coin: Pediatric-Onset and Adult-Onset Common Variable Immune Deficiency. J Clin Immunol 2017; 37: 592-602 [PMID: 28755066 DOI: 10.1007/s10875-017-0415-5]
- Chapel H, Lucas M, Lee M, Bjorkander J, Webster D, Grimbacher B, Fieschi C, Thon V, Abedi MR, 10 Hammarstrom L. Common variable immunodeficiency disorders: division into distinct clinical phenotypes. Blood 2008; 112: 277-286 [PMID: 18319398 DOI: 10.1182/blood-2007-11-124545]
- 11 El-Helou SM, Biegner AK, Bode S, Ehl SR, Heeg M, Maccari ME, Ritterbusch H, Speckmann C, Rusch S, Scheible R. Warnatz K. Atschekzei F. Beider R. Ernst D. Gerschmann S. Jablonka A. Mielke G. Schmidt RE, Schürmann G, Sogkas G, Baumann UH, Klemann C, Viemann D, von Bernuth H, Krüger R, Hanitsch LG, Scheibenbogen CM, Wittke K, Albert MH, Eichinger A, Hauck F, Klein C, Rack-Hoch A, Sollinger FM, Avila A, Borte M, Borte S, Fasshauer M, Hauenherm A, Kellner N, Müller AH, Ülzen A, Bader P, Bakhtiar S. Lee JY. Heß U. Schubert R. Wölke S. Zielen S. Ghosh S. Laws HJ. Neubert J. Oommen PT. Hönig M. Schulz A, Steinmann S, Schwarz K, Dückers G, Lamers B, Langemeyer V, Niehues T, Shai S, Graf D, Müglich C, Schmalzing MT, Schwaneck EC, Tony HP, Dirks J, Haase G, Liese JG, Morbach H, Foell D, Hellige A, Wittkowski H, Masjosthusmann K, Mohr M, Geberzahn L, Hedrich CM, Müller C, Rösen-Wolff A, Roesler J, Zimmermann A, Behrends U, Rieber N, Schauer U, Handgretinger R, Holzer U, Henes J, Kanz L, Boesecke C, Rockstroh JK, Schwarze-Zander C, Wasmuth JC, Dilloo D, Hülsmann B, Schönberger S, Schreiber S, Zeuner R, Ankermann T, von Bismarck P, Huppertz HI, Kaiser-Labusch P, Greil J, Jakoby D, Kulozik AE, Metzler M, Naumann-Bartsch N, Sobik B, Graf N, Heine S, Kobbe R, Lehmberg K, Müller I, Herrmann F, Horneff G, Klein A, Peitz J, Schmidt N, Bielack S, Groß-Wieltsch U, Classen CF, Klasen J, Deutz P, Kamitz D, Lassay L, Tenbrock K, Wagner N, Bernbeck B, Brummel B, Lara-Villacanas E, Münstermann E, Schneider DT, Tietsch N, Westkemper M, Weiß M, Kramm C, Kühnle I, Kullmann S, Girschick H, Specker C, Vinnemeier-Laubenthal E, Haenicke H, Schulz C, Schweigerer L, Müller TG, Stiefel M, Belohradsky BH, Soetedjo V, Kindle G, Grimbacher B. The German National Registry of Primary Immunodeficiencies (2012-2017). Front Immunol 2019; 10: 1272 [PMID: 31379802 DOI: 10.3389/fimmu.2019.01272]
- ESID Registry Working definitions for clinical diagnosis of PID. [Accessed March, 28, 2020] Available 12 from: https://esid.org/Working-Parties/Registry-Working-Party/Diagnosis-criteria
- 13 Graziano V, Pecoraro A, Mormile I, Quaremba G, Genovese A, Buccelli C, Paternoster M, Spadaro G. Delay in diagnosis affects the clinical outcome in a cohort of cvid patients with marked reduction of iga serum levels. Clin Immunol 2017; 180: 1-4 [PMID: 28347823 DOI: 10.1016/j.clim.2017.03.011]
- 14 Więsik-Szewczyk E, Ziętkiewicz M, Matyja-Bednarczyk A, Napiórkowska-Baran K, Suchanek H, Jahnz-Różyk K. The first Polish cohort of adult patients with common variable immunodeficiency from 4 specialized centers: do we provide standards of care? Pol Arch Intern Med 2018; 128: 563-566 [PMID: 30113020 DOI: 10.20452/pamw.4315]
- Fernández Pérez ER, Hunter M, Katial RK. United States trends in mortality rates for primary 15 immunodeficiency diseases. J Allergy Clin Immunol Pract 2019; 7: 1045-1048 [PMID: 30312803 DOI: 10.1016/j.jaip.2018.09.030]
- 16 Yong PF, Thaventhiran JE, Grimbacher B. "A rose is a rose is a rose," but CVID is Not CVID common variable immune deficiency (CVID), what do we know in 2011? Adv Immunol 2011; 111: 47-107 [PMID: 21970952 DOI: 10.1016/B978-0-12-385991-4.00002-7]
- Woodward J. Gkrania-Klotsas E. Kumararatne D. Chronic norovirus infection and common variable 17 immunodeficiency. Clin Exp Immunol 2017; 188: 363-370 [PMID: 27753065 DOI: 10.1111/cei.12884]
- Daniels JA, Lederman HM, Maitra A, Montgomery EA. Gastrointestinal tract pathology in patients with 18 common variable immunodeficiency (CVID): a clinicopathologic study and review. Am J Surg Pathol 2007; 31: 1800-1812 [PMID: 18043034 DOI: 10.1097/PAS.0b013e3180cab60c]
- 19 Koźlik P, Kosałka J, Gałązka K, Paciorek K, Sydor W, Musiał J. Cytomegalovirus gastroenteritis with pyloric stenosis and persistent malabsorption in a patient with common variable immunodeficiency. Pol Arch Intern Med 2018; 128: 783-784 [PMID: 30403390 DOI: 10.20452/pamw.4367]
- Friedmann D, Goldacker S, Peter HH, Warnatz K. Preserved Cellular Immunity Upon Influenza 20 Vaccination in Most Patients with Common Variable Immunodeficiency. J Allergy Clin Immunol Pract 2020; 8: 2332-2340.e5 [PMID: 32330665 DOI: 10.1016/j.jaip.2020.04.019]
- Ouinti I. Lougaris V. Milito C. Cinetto F. Pecoraro A. Mezzaroma I. Mastroianni CM. Turriziani O. 21 Bondioni MP, Filippini M, Soresina A, Spadaro G, Agostini C, Carsetti R, Plebani A. A possible role for B cells in COVID-19? Lesson from patients with agammaglobulinemia. J Allergy Clin Immunol 2020; 146: 211-213.e4 [PMID: 32333914 DOI: 10.1016/j.jaci.2020.04.013]
- Patrawala M, Cui Y, Peng L, Fuleihan RL, Garabedian EK, Patel K, Guglani L. Pulmonary Disease Burden 22 in Primary Immune Deficiency Disorders: Data from USIDNET Registry. J Clin Immunol 2020; 40: 340-349 [PMID: 31919711 DOI: 10.1007/s10875-019-00738-w]
- Verma N, Grimbacher B, Hurst JR. Lung disease in primary antibody deficiency. Lancet Respir Med 2015; 23 3: 651-660 [PMID: 26188881 DOI: 10.1016/S2213-2600(15)00202-7]
- 24 Kralickova P. Mala E. Vokurkova D. Krcmova I. Pliskova L. Stepanova V. Bartos V. Koblizek V. Tacheci I, Bures J, Brozik J, Litzman J. Cytomegalovirus disease in patients with common variable immunodeficiency: three case reports. Int Arch Allergy Immunol 2014; 163: 69-74 [PMID: 24247002 DOI: 10.1159/000355957
- 25 Wiesik-Szewczyk E, Kucharczyk A, Świerkocka K, Rutkowska E, Jahnz-Różyk K. Recurrent infections in a patient with psoriatic arthritis and hypogammaglobulinemia, treated with conventional and biologic diseasemodifying anti-rheumatic drugs-a primary or secondary entity? Clin Rheumatol 2017; 36: 1677-1681 [PMID:



28488125 DOI: 10.1007/s10067-017-3670-11

- Więsik-Szewczyk E, Jahnz-Różyk K, Rutkowska E. [Common variable immunodeficiency -26
- interdisciplinary problem case report. Polish]. Alergologia i Immunologia Współczesna 2016; 37: 29-33 27 Hanitsch LG, Wittke K, Stittrich AB, Volk HD, Scheibenbogen C. Interstitial Lung Disease Frequently Precedes CVID Diagnosis. J Clin Immunol 2019; 39: 849-851 [PMID: 31637570 DOI:
- 10.1007/s10875-019-00708-21 Michel M, Chanet V, Galicier L, Ruivard M, Levy Y, Hermine O, Oksenhendler E, Schaeffer A, Bierling P, 28 Godeau B. Autoimmune thrombocytopenic purpura and common variable immunodeficiency: analysis of 21
- cases and review of the literature. Medicine (Baltimore) 2004; 83: 254-263 [PMID: 15232313 DOI: 10.1097/01.md.0000133624.65946.40]
- Resnick ES, Moshier EL, Godbold JH, Cunningham-Rundles C. Morbidity and mortality in common 29 variable immune deficiency over 4 decades. Blood 2012; 119: 1650-1657 [PMID: 22180439 DOI: 10.1182/blood-2011-09-377945
- Fischer A, Provot J, Jais JP, Alcais A, Mahlaoui N; members of the CEREDIH French PID study group. 30 Autoimmune and inflammatory manifestations occur frequently in patients with primary immunodeficiencies. J Allergy Clin Immunol 2017; 140: 1388-1393.e8 [PMID: 28192146 DOI: 10.1016/j.jaci.2016.12.978]
- Jørgensen SF, Reims HM, Frydenlund D, Holm K, Paulsen V, Michelsen AE, Jørgensen KK, Osnes LT, 31 Bratlie J, Eide TJ, Dahl CP, Holter E, Tronstad RR, Hanevik K, Brattbakk HR, Kaveh F, Fiskerstrand T, Kran AB, Ueland T, Karlsen TH, Aukrust P, Lundin KE, Fevang B. A Cross-Sectional Study of the Prevalence of Gastrointestinal Symptoms and Pathology in Patients With Common Variable Immunodeficiency. Am J Gastroenterol 2016; 111: 1467-1475 [PMID: 27527747 DOI: 10.1038/ajg.2016.329]
- Pecoraro A, Nappi L, Crescenzi L, D'Armiento FP, Genovese A, Spadaro G. Chronic Diarrhea in Common 32 Variable Immunodeficiency: a Case Series and Review of the Literature. J Clin Immunol 2018; 38: 67-76 [PMID: 29138951 DOI: 10.1007/s10875-017-0461-z]
- 33 Bethune C, Egner W, Garcez T, Huissoon A, Jolles S, Karim Y, Jain R, Savic S, Kelley K, Grosse-Kreul D, Grigoriadou S. British Society for Immunology/United Kingdom Primary Immunodeficiency Network consensus statement on managing non-infectious complications of common variable immunodeficiency disorders. Clin Exp Immunol 2019; 196: 328-335 [PMID: 30724343 DOI: 10.1111/cei.13272]
- Bouvry D, Mouthon L, Brillet PY, Kambouchner M, Ducroix JP, Cottin V, Haroche J, Viallard JF, Lazor R, 34 Lebargy F, Tazi A, Wallaert B, Smail A, Pellegrin JL, Nunes H, Amoura Z, Cordier JF, Valeyre D, Naccache JM: Groupe Sarcoïdose Francophone. Granulomatosis-associated common variable immunodeficiency disorder: a case-control study versus sarcoidosis. Eur Respir J 2013; 41: 115-122 [PMID: 22903958 DOI: 10.1183/09031936.00189011
- Mannina A, Chung JH, Swigris JJ, Solomon JJ, Huie TJ, Yunt ZX, Truong TQ, Brown KK, Achcar RD, 35 Olson AL, Cox CW, Kligerman SJ, Curran-Everett D, Fernández Pérez ER. Clinical Predictors of a Diagnosis of Common Variable Immunodeficiency-related Granulomatous-Lymphocytic Interstitial Lung Disease. Ann Am Thorac Soc 2016; 13: 1042-1049 [PMID: 27064856 DOI: 10.1513/AnnalsATS.201511-728OC
- 36 Bates CA, Ellison MC, Lynch DA, Cool CD, Brown KK, Routes JM. Granulomatous-lymphocytic lung disease shortens survival in common variable immunodeficiency. J Allergy Clin Immunol 2004; 114: 415-421 [PMID: 15316526 DOI: 10.1016/j.jaci.2004.05.057]
- 37 Hurst JR, Verma N, Lowe D, Baxendale HE, Jolles S, Kelleher P, Longhurst HJ, Patel SY, Renzoni EA, Sander CR, Avery GR, Babar JL, Buckland MS, Burns S, Egner W, Gompels MM, Gordins P, Haddock JA, Hart SP, Hayman GR, Herriot R, Hoyles RK, Huissoon AP, Jacob J, Nicholson AG, Rassl DM, Sargur RB, Savic S, Seneviratne SL, Sheaff M, Vaitla PM, Walters GI, Whitehouse JL, Wright PA, Condliffe AM. British Lung Foundation/United Kingdom Primary Immunodeficiency Network Consensus Statement on the Definition, Diagnosis, and Management of Granulomatous-Lymphocytic Interstitial Lung Disease in Common Variable Immunodeficiency Disorders. J Allergy Clin Immunol Pract 2017; 5: 938-945 [PMID: 28351785 DOI: 10.1016/j.jaip.2017.01.021]
- 38 Jolles S, Carne E, Brouns M, El-Shanawany T, Williams P, Marshall C, Fielding P. FDG PET-CT imaging of therapeutic response in granulomatous lymphocytic interstitial lung disease (GLILD) in common variable immunodeficiency (CVID). Clin Exp Immunol 2017; 187: 138-145 [PMID: 27896807 DOI: 10 1111/cei 128561
- Patel S, Anzilotti C, Lucas M, Moore N, Chapel H. Interstitial lung disease in patients with common variable 39 immunodeficiency disorders: several different pathologies? Clin Exp Immunol 2019; 198: 212-223 [PMID: 31216049 DOI: 10.1111/cei.13343]
- Cereser L, De Carli M, d'Angelo P, Zanelli E, Zuiani C, Girometti R. High-resolution computed tomography 40 findings in humoral primary immunodeficiencies and correlation with pulmonary function tests. World J Radiol 2018; 10: 172-183 [PMID: 30568751 DOI: 10.4329/wjr.v10.i11.172]
- Rao N, Mackinnon AC, Routes JM. Granulomatous and lymphocytic interstitial lung disease: a spectrum of 41 pulmonary histopathologic lesions in common variable immunodeficiency--histologic and immunohistochemical analyses of 16 cases. Hum Pathol 2015; 46: 1306-1314 [PMID: 26138782 DOI: 10.1016/j.humpath.2015.05.011]
- 42 Kellner ES, Fuleihan R, Cunningham-Rundles C; USIDNET Consortium, Wechsler JB. Cellular Defects in CVID Patients with Chronic Lung Disease in the USIDNET Registry. J Clin Immunol 2019; 39: 569-576 [PMID: 31250334 DOI: 10.1007/s10875-019-00657-w]
- Hartono S, Motosue MS, Khan S, Rodriguez V, Iyer VN, Divekar R, Joshi AY. Predictors of granulomatous 43 lymphocytic interstitial lung disease in common variable immunodeficiency. Ann Allergy Asthma Immunol 2017; 118: 614-620 [PMID: 28254202 DOI: 10.1016/j.anai.2017.01.004]
- Ng J, Wright K, Alvarez M, Hunninghake GM, Wesemann DR. Rituximab Monotherapy for Common 44 Variable Immune Deficiency-Associated Granulomatous-Lymphocytic Interstitial Lung Disease. Chest 2019; 155: e117-e121 [PMID: 31060706 DOI: 10.1016/j.chest.2019.01.034]
- Centrum für Chronische Immundefizienz CCI. [Accessed on 30 April 2020] Available from:



https://www.uniklinik-freiburg.de/cci/studien/stilpad.html

- 46 Abolhassani H, Aghamohammadi A, Imanzadeh A, Mohammadinejad P, Sadeghi B, Rezaei N. Malignancy phenotype in common variable immunodeficiency. J Investig Allergol Clin Immunol 2012; 22: 133-134 [PMID: 22533236 DOI: 10.1159/000330645]
- Cunningham-Rundles C, Siegal FP, Cunningham-Rundles S, Lieberman P. Incidence of cancer in 98 47 patients with common varied immunodeficiency. J Clin Immunol 1987; 7: 294-299 [PMID: 3611296 DOI: 10 1007/BF009155501
- Mellemkjaer L, Hammarstrom L, Andersen V, Yuen J, Heilmann C, Barington T, Bjorkander J, Olsen JH. 48 Cancer risk among patients with IgA deficiency or common variable immunodeficiency and their relatives: a combined Danish and Swedish study. Clin Exp Immunol 2002; 130: 495-500 [PMID: 12452841 DOI: 10.1046/j.1365-2249.2002.02004.x
- 49 Vajdic CM, Mao L, van Leeuwen MT, Kirkpatrick P, Grulich AE, Riminton S. Are antibody deficiency disorders associated with a narrower range of cancers than other forms of immunodeficiency? Blood 2010; 116: 1228-1234 [PMID: 20466855 DOI: 10.1182/blood-2010-03-272351]
- 50 Pulvirenti F, Pecoraro A, Cinetto F, Milito C, Valente M, Santangeli E, Crescenzi L, Rizzo F, Tabolli S, Spadaro G, Agostini C, Quinti I. Gastric Cancer Is the Leading Cause of Death in Italian Adult Patients With Common Variable Immunodeficiency. Front Immunol 2018; 9: 2546 [PMID: 30455695 DOI: 10.3389/fimmu.2018.02546
- van der Poorten DK, McLeod D, Ahlenstiel G, Read S, Kwok A, Santhakumar C, Bassan M, Culican S, 51 Campbell D, Wong SWJ, Evans L, Jideh B, Kane A, Katelaris CH, Keat K, Ko Y, Lee JA, Limaye S, Lin MW, Murad A, Rafferty M, Suan D, Swaminathan S, Riminton SD, Toong C, Berglund LJ. Gastric Cancer Screening in Common Variable Immunodeficiency. J Clin Immunol 2018; 38: 768-777 [PMID: 30219982 DOI: 10.1007/s10875-018-0546-31
- 52 Dhalla F, da Silva SP, Lucas M, Travis S, Chapel H. Review of gastric cancer risk factors in patients with common variable immunodeficiency disorders, resulting in a proposal for a surveillance programme. Clin Exp Immunol 2011; 165: 1-7 [PMID: 21470209 DOI: 10.1111/j.1365-2249.2011.04384.x]
- 53 Rugge M, Meggio A, Pravadelli C, Barbareschi M, Fassan M, Gentilini M, Zorzi M, Pretis G, Graham DY, Genta RM. Gastritis staging in the endoscopic follow-up for the secondary prevention of gastric cancer: a 5year prospective study of 1755 patients. Gut 2019; 68: 11-17 [PMID: 29306868 DOI: 10.1136/gutjnl-2017-314600]
- 54 Cunningham-Rundles C. Clinical and immunologic analyses of 103 patients with common variable immunodeficiency. J Clin Immunol 1989; 9: 22-33 [PMID: 2784795 DOI: 10.1007/bf00917124]
- Bonilla FA, Barlan I, Chapel H, Costa-Carvalho BT, Cunningham-Rundles C, de la Morena MT, Espinosa-55 Rosales FJ, Hammarström L, Nonoyama S, Quinti I, Routes JM, Tang ML, Warnatz K. International Consensus Document (ICON): Common Variable Immunodeficiency Disorders. J Allergy Clin Immunol Pract 2016; 4: 38-59 [PMID: 26563668 DOI: 10.1016/j.jaip.2015.07.025]
- 56 Holding S, Jolles S. Current screening approaches for antibody deficiency. Curr Opin Allergy Clin Immunol 2015: 15: 547-555 [PMID: 26513714 DOI: 10.1097/ACI.00000000000222]
- Jolles S, Borrell R, Zouwail S, Heaps A, Sharp H, Moody M, Selwood C, Williams P, Phillips C, Hood K, 57 Holding S, El Shanawany T. Calculated globulin (CG) as a screening test for antibody deficiency. Clin Exp Immunol 2014; 177: 671-678 [PMID: 24784320 DOI: 10.1111/cei.12369]
- 58 Pecoraro A, Jolles S, Crescenzi L, Varricchi G, Marone G, Savoia M, Genovese A, Spadaro G. Validation of Calculated Globulin (CG) as a Screening Test for Antibody Deficiency in an Italian University Hospital. Curr Pharm Biotechnol 2018; 19: 728-733 [PMID: 30091407 DOI: 10.2174/1389201019666180808163311]
- 59 Orange JS. Ballow M, Stiehm ER, Ballas ZK, Chinen J, De La Morena M, Kumararatne D, Harville TO. Hesterberg P, Koleilat M, McGhee S, Perez EE, Raasch J, Scherzer R, Schroeder H, Seroogy C, Huissoon A, Sorensen RU, Katial R. Use and interpretation of diagnostic vaccination in primary immunodeficiency: a working group report of the Basic and Clinical Immunology Interest Section of the American Academy of Allergy, Asthma & Immunology. J Allergy Clin Immunol 2012; 130: S1-24 [PMID: 22935624 DOI: 10.1016/j.jaci.2012.07.002]
- Parker AR, Bradley C, Harding S, Sánchez-Ramón S, Jolles S, Kiani-Alikhan S. Measurement and 60 interpretation of Salmonella typhi Vi IgG antibodies for the assessment of adaptive immunity. J Immunol Methods 2018; 459: 1-10 [PMID: 29800575 DOI: 10.1016/j.jim.2018.05.013]
- Evans C, Bateman E, Steven R, Ponsford M, Cullinane A, Shenton C, Duthie G, Conlon C, Jolles S, 61 Huissoon AP, Longhurst HJ, Rahman T, Scott C, Wallis G, Harding S, Parker AR, Ferry BL. Measurement of Typhi Vi antibodies can be used to assess adaptive immunity in patients with immunodeficiency. Clin Exp Immunol 2018; 192: 292-301 [PMID: 29377063 DOI: 10.1111/cei.13105]
- Warnatz K, Denz A, Dräger R, Braun M, Groth C, Wolff-Vorbeck G, Eibel H, Schlesier M, Peter HH. 62 Severe deficiency of switched memory B cells (CD27(+)IgM(-)IgD(-)) in subgroups of patients with common variable immunodeficiency: a new approach to classify a heterogeneous disease. Blood 2002; 99: 1544-1551 [PMID: 11861266 DOI: 10.1182/blood.v99.5.1544]
- 63 Wehr C. Kivioja T, Schmitt C, Ferry B, Witte T, Eren E, Vlkova M, Hernandez M, Detkova D, Bos PR, Poerksen G, von Bernuth H, Baumann U, Goldacker S, Gutenberger S, Schlesier M, Bergeron-van der Cruyssen F, Le Garff M, Debré P, Jacobs R, Jones J, Bateman E, Litzman J, van Hagen PM, Plebani A, Schmidt RE, Thon V, Quinti I, Espanol T, Webster AD, Chapel H, Vihinen M, Oksenhendler E, Peter HH, Warnatz K. The EUROclass trial: defining subgroups in common variable immunodeficiency. Blood 2008; 111: 77-85 [PMID: 17898316 DOI: 10.1182/blood-2007-06-091744]
- 64 Ebbo M, Gérard L, Carpentier S, Vély F, Cypowyj S, Farnarier C, Vince N, Malphettes M, Fieschi C, Oksenhendler E, Schleinitz N, Vivier E; DEFI Study Group. Low Circulating Natural Killer Cell Counts are Associated With Severe Disease in Patients With Common Variable Immunodeficiency. EBioMedicine 2016; 6: 222-230 [PMID: 27211564 DOI: 10.1016/j.ebiom.2016.02.025]
- Aspalter RM, Sewell WA, Dolman K, Farrant J, Webster AD. Deficiency in circulating natural killer (NK) 65 cell subsets in common variable immunodeficiency and X-linked agammaglobulinaemia. Clin Exp Immunol 2000; 121: 506-514 [PMID: 10971518 DOI: 10.1046/j.1365-2249.2000.01317.x]
- Giovannetti A, Pierdominici M, Mazzetta F, Marziali M, Renzi C, Mileo AM, De Felice M, Mora B,



Esposito A, Carello R, Pizzuti A, Paggi MG, Paganelli R, Malorni W, Aiuti F. Unravelling the complexity of T cell abnormalities in common variable immunodeficiency. *J Immunol* 2007; **178**: 3932-3943 [PMID: 17339494 DOI: 10.4049/jimmunol.178.6.3932]

- 67 Bateman EA, Ayers L, Sadler R, Lucas M, Roberts C, Woods A, Packwood K, Burden J, Harrison D, Kaenzig N, Lee M, Chapel HM, Ferry BL. T cell phenotypes in patients with common variable immunodeficiency disorders: associations with clinical phenotypes in comparison with other groups with recurrent infections. *Clin Exp Immunol* 2012; **170**: 202-211 [PMID: 23039891 DOI: 10.1111/j.1365-2249.2012.04643.x]
- 68 Edwards ESJ, Bosco JJ, Aui PM, Stirling RG, Cameron PU, Chatelier J, Hore-Lacy F, O'Hehir RE, van Zelm MC. Predominantly Antibody-Deficient Patients With Non-infectious Complications Have Reduced Naive B, Treg, Th17, and Tfh17 Cells. *Front Immunol* 2019; 10: 2593 [PMID: 31803177 DOI: 10.3389/fimmu.2019.02593]
- 69 Azizi G, Rezaei N, Kiaee F, Tavakolinia N, Yazdani R, Mirshafiey A, Aghamohammadi A. T-Cell Abnormalities in Common Variable Immunodeficiency. *J Investig Allergol Clin Immunol* 2016; 26: 233-243 [PMID: 27374799 DOI: 10.18176/jiaci.0069]
- 70 Unger S, Seidl M, van Schouwenburg P, Rakhmanov M, Bulashevska A, Frede N, Grimbacher B, Pfeiffer J, Schrenk K, Munoz L, Hanitsch L, Stumpf I, Kaiser F, Hausmann O, Kollert F, Goldacker S, van der Burg M, Keller B, Warnatz K. The T_H1 phenotype of follicular helper T cells indicates an IFN-γ-associated immune dysregulation in patients with CD21low common variable immunodeficiency. *J Allergy Clin Immunol* 2018; 141: 730-740 [PMID: 28554560 DOI: 10.1016/j.jaci.2017.04.041]
- 71 von Spee-Mayer C, Koemm V, Wehr C, Goldacker S, Kindle G, Bulashevska A, Proietti M, Grimbacher B, Ehl S, Warnatz K. Evaluating laboratory criteria for combined immunodeficiency in adult patients diagnosed with common variable immunodeficiency. *Clin Immunol* 2019; **203**: 59-62 [PMID: 31004792 DOI: 10.1016/j.clim.2019.04.001]
- 72 Le Saos-Patrinos C, Loizon S, Blanco P, Viallard JF, Duluc D. Functions of Tfh Cells in Common Variable Immunodeficiency. *Front Immunol* 2020; 11: 6 [PMID: 32082308 DOI: 10.3389/fimmu.2020.00006]
- 73 Turpin D, Furudoi A, Parrens M, Blanco P, Viallard JF, Duluc D. Increase of follicular helper T cells skewed toward a Th1 profile in CVID patients with non-infectious clinical complications. *Clin Immunol* 2018; 197: 130-138 [PMID: 30219667 DOI: 10.1016/j.clim.2018.09.006]
- 74 Klocperk A, Unger S, Friedmann D, Seidl M, Zoldan K, Pfeiffer J, Hausmann O, Benes V, Andrieux G, Boettler T, Sediva A, Bengsch B, Warnatz K. Exhausted phenotype of follicular CD8 T cells in CVID. J Allergy Clin Immunol 2020 [PMID: 32169377 DOI: 10.1016/j.jaci.2020.02.025]
- 75 Blanco E, Pérez-Andrés M, Arriba-Méndez S, Serrano C, Criado I, Del Pino-Molina L, Silva S, Madruga I, Bakardjieva M, Martins C, Serra-Caetano A, Romero A, Contreras-Sanfeliciano T, Bonroy C, Sala F, Martín A, Bastida JM, Lorente F, Prieto C, Dávila I, Marcos M, Kalina T, Vlkova M, Chovancova Z, Cordeiro AI, Philippé J, Haerynck F, López-Granados E, Sousa AE, van der Burg M, van Dongen JJM, Orfao A; EuroFlow PID group. Defects in memory B-cell and plasma cell subsets expressing different immunoglobulin-subclasses in patients with CVID and immunoglobulin subclass deficiencies. J Allergy Clin Immunol 2019; 144: 809-824 [PMID; 30826363 DOI: 10.1016/j.jaci.2019.02.017]
- Maffucci P, Filion CA, Boisson B, Itan Y, Shang L, Casanova JL, Cunningham-Rundles C. Genetic Diagnosis Using Whole Exome Sequencing in Common Variable Immunodeficiency. *Front Immunol* 2016; 7: 220 [PMID: 27379089 DOI: 10.3389/fimmu.2016.00220]
- 77 Yska HAF, Elsink K, Kuijpers TW, Frederix GWJ, van Gijn ME, van Montfrans JM. Diagnostic Yield of Next Generation Sequencing in Genetically Undiagnosed Patients with Primary Immunodeficiencies: a Systematic Review, J Clin Immunol 2019; 39: 577-591 [PMID: 31250335 DOI: 10.1007/s10875-019-00656-x]
- 78 Ameratunga R, Lehnert K, Woon ST, Gillis D, Bryant VL, Slade CA, Steele R. Review: Diagnosing Common Variable Immunodeficiency Disorder in the Era of Genome Sequencing. *Clin Rev Allergy Immunol* 2018; 54: 261-268 [PMID: 29030829 DOI: 10.1007/s12016-017-8645-0]
- 79 Conley ME, Notarangelo LD, Etzioni A. Diagnostic criteria for primary immunodeficiencies. Representing PAGID (Pan-American Group for Immunodeficiency) and ESID (European Society for Immunodeficiencies). *Clin Immunol* 1999; **93**: 190-197 [PMID: 10600329 DOI: 10.1006/clim.1999.4799]
- Ameratunga R, Woon ST, Gillis D, Koopmans W, Steele R. New diagnostic criteria for common variable immune deficiency (CVID), which may assist with decisions to treat with intravenous or subcutaneous immunoglobulin. *Clin Exp Immunol* 2013; **174**: 203-211 [PMID: 23859429 DOI: 10.1111/cei.12178]
- 81 Patel SY, Carbone J, Jolles S. The Expanding Field of Secondary Antibody Deficiency: Causes, Diagnosis, and Management. *Front Immunol* 2019; 10: 33 [PMID: 30800120 DOI: 10.3389/fimmu.2019.00033]
- 82 Jolles S, Chapel H, Litzman J. When to initiate immunoglobulin replacement therapy (IGRT) in antibody deficiency: a practical approach. *Clin Exp Immunol* 2017; 188: 333-341 [PMID: 28000208 DOI: 10.1111/cei.12915]
- 83 Misbah S, Sturzenegger MH, Borte M, Shapiro RS, Wasserman RL, Berger M, Ochs HD. Subcutaneous immunoglobulin: opportunities and outlook. *Clin Exp Immunol* 2009; **158** Suppl 1: 51-59 [PMID: 19883424 DOI: 10.1111/j.1365-2249.2009.04027.x]
- 84 Wiesik-Szewczyk E, Sołdacki D, Paczek L, Jahnz-Różyk K. Facilitated Subcutaneous Immunoglobulin Replacement Therapy in Clinical Practice: A Two Center, Long-Term Retrospective Observation in Adults With Primary Immunodeficiencies. *Front Immunol* 2020; 11: 981 [PMID: 32670265 DOI: 10.3389/fimmu.2020.00981]



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

