World Journal of *Clinical Cases*

World J Clin Cases 2021 January 16; 9(2): 291-520





Published by Baishideng Publishing Group Inc

W T C C World Journal of Clinical Cases

Contents

Thrice Monthly Volume 9 Number 2 January 16, 2021

OPINION REVIEW

Continuity of cancer care in the era of COVID-19 pandemic: Role of social media in low- and middle-291 income countries

Yadav SK, Yadav N

REVIEW

- Effect of a fever in viral infections the 'Goldilocks' phenomenon? 296 Belon L, Skidmore P, Mehra R, Walter E
- 308 Overview of bile acid signaling in the cardiovascular system Zhang R, Ma WQ, Fu MJ, Li J, Hu CH, Chen Y, Zhou MM, Gao ZJ, He YL

MINIREVIEWS

321 Gut microbiota and inflammatory bowel disease: The current status and perspectives Zheng L, Wen XL

ORIGINAL ARTICLE

Retrospective Cohort Study

334 Effective immune-inflammation index for ulcerative colitis and activity assessments

Zhang MH, Wang H, Wang HG, Wen X, Yang XZ

Retrospective Study

344 Risk factors associated with acute respiratory distress syndrome in COVID-19 patients outside Wuhan: A double-center retrospective cohort study of 197 cases in Hunan, China

Hu XS, Hu CH, Zhong P, Wen YJ, Chen XY

META-ANALYSIS

357 Limb length discrepancy after total knee arthroplasty: A systematic review and meta-analysis

Tripathy SK, Pradhan SS, Varghese P, Purudappa PP, Velagada S, Goyal T, Panda BB, Vanyambadi J

CASE REPORT

Lateral position intubation followed by endoscopic ultrasound-guided angiotherapy in acute esophageal 372 variceal rupture: A case report

Wen TT, Liu ZL, Zeng M, Zhang Y, Cheng BL, Fang XM

379 Perioperative mortality of metastatic spinal disease with unknown primary: A case report and review of literature

Li XM. Jin LB



- .	World Journal of Clinical Cases
Conter	Thrice Monthly Volume 9 Number 2 January 16, 2021
389	Massive gastric bleeding - perforation of pancreatic pseudocyst into the stomach: A case report and review of literature
	Jin Z, Xiang YW, Liao QS, Yang XX, Wu HC, Tuo BG, Xie R
396	Natural history of inferior mesenteric arteriovenous malformation that led to ischemic colitis: A case report
	Kimura Y, Hara T, Nagao R, Nakanishi T, Kawaguchi J, Tagami A, Ikeda T, Araki H, Tsurumi H
403	Coil embolization of arterioportal fistula complicated by gastrointestinal bleeding after Caesarian section: A case report
	Stepanyan SA, Poghosyan T, Manukyan K, Hakobyan G, Hovhannisyan H, Safaryan H, Baghdasaryan E, Gemilyan M
410	Cholecystoduodenal fistula presenting with upper gastrointestinal bleeding: A case report
	Park JM, Kang CD, Kim JH, Lee SH, Nam SJ, Park SC, Lee SJ, Lee S
416	Rare case of fecal impaction caused by a fecalith originating in a large colonic diverticulum: A case report
	Tanabe H, Tanaka K, Goto M, Sato T, Sato K, Fujiya M, Okumura T
422	Intravitreal dexamethasone implant – a new treatment for idiopathic posterior scleritis: A case report
	Zhao YJ, Zou YL, Lu Y, Tu MJ, You ZP
429	Inflammatory myofibroblastic tumor successfully treated with metformin: A case report and review of literature
	Liang Y, Gao HX, Tian RC, Wang J, Shan YH, Zhang L, Xie CJ, Li JJ, Xu M, Gu S
436	Neonatal isovaleric acidemia in China: A case report and review of literature
	Wu F, Fan SJ, Zhou XH
445	Malignant solitary fibrous tumor of the greater omentum: A case report and review of literature
	Guo YC, Yao LY, Tian ZS, Shi B, Liu Y, Wang YY
457	Paratesticular liposarcoma: Two case reports
	Zheng QG, Sun ZH, Chen JJ, Li JC, Huang XJ
463	Sinistral portal hypertension associated with pancreatic pseudocysts - ultrasonography findings: A case report
	Chen BB, Mu PY, Lu JT, Wang G, Zhang R, Huang DD, Shen DH, Jiang TT
469	Epstein-Barr virus-associated monomorphic post-transplant lymphoproliferative disorder after pediatric kidney transplantation: A case report
	Wang Z, Xu Y, Zhao J, Fu YX
476	Postoperative complications of concomitant fat embolism syndrome, pulmonary embolism and tympanic membrane perforation after tibiofibular fracture: A case report
	Shao J, Kong DC, Zheng XH, Chen TN, Yang TY
482	Double-hit lymphoma (rearrangements of MYC, BCL-2) during pregnancy: A case report
	Xie F, Zhang LH, Yue YQ, Gu LL, Wu F



Conton	World Journal of Clinical Cases
Conten	Thrice Monthly Volume 9 Number 2 January 16, 2021
489	Is sinusoidal obstructive syndrome a recurrent disease after liver transplantation? A case report
	Liu Y, Sun LY, Zhu ZJ, Wei L, Qu W, Zeng ZG
496	Portal hypertension exacerbates intrahepatic portosystemic venous shunt and further induces refractory hepatic encephalopathy: A case report
	Chang YH, Zhou XL, Jing D, Ni Z, Tang SH
502	Repair of a severe palm injury with anterolateral thigh and ilioinguinal flaps: A case report
	Gong HY, Sun XG, Lu LJ, Liu PC, Yu X
509	Indirect inguinal hernia containing portosystemic shunt vessel: A case report
	Yura M, Yo K, Hara A, Hayashi K, Tajima Y, Kaneko Y, Fujisaki H, Hirata A, Takano K, Hongo K, Yoneyama K, Nakagawa M
516	Recurrent inverted papilloma coexisted with skull base lymphoma: A case report
	Hsu HJ, Huang CC, Chuang MT, Tien CH, Lee JS, Lee PH



Contents

Thrice Monthly Volume 9 Number 2 January 16, 2021

ABOUT COVER

Editorial Board Member of World Journal of Clinical Cases, Dr. Mukul Vij is Senior Consultant Pathologist and Lab Director at Dr Rela Institute and Medical Center in Chennai, India (since 2018). Having received his MBBS degree from King George Medical College in 2004, Dr. Vij undertook postgraduate training at Sanjay Gandhi Postgraduate Institute of Medical Sciences, receiving his Master's degree in Pathology in 2008 and his PDCC certificate in Renal Pathology in 2009. After 2 years as senior resident, he became Assistant Professor in the Department of Pathology at Christian Medical College, Vellore (2011), moving on to Global Health City as Consultant Pathologist and then Head of the Pathology Department (2013). (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of World Journal of Clinical Cases (WJCC, World J Clin Cases) is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The WJCC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, PubMed, and PubMed Central. The 2020 Edition of Journal Citation Reports® cites the 2019 impact factor (IF) for WJCC as 1.013; IF without journal self cites: 0.991; Ranking: 120 among 165 journals in medicine, general and internal; and Quartile category: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Jia-Hui Li; Production Department Director: Yu-Jie Ma; Editorial Office Director: Jin-Lei Wang,

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS		
World Journal of Clinical Cases	https://www.wjgnet.com/bpg/gerinfo/204		
ISSN	GUIDELINES FOR ETHICS DOCUMENTS		
ISSN 2307-8960 (online)	https://www.wjgnet.com/bpg/GerInfo/287		
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH		
April 16, 2013	https://www.wjgnet.com/bpg/gerinfo/240		
FREQUENCY	PUBLICATION ETHICS		
Thrice Monthly	https://www.wjgnet.com/bpg/GerInfo/288		
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT		
Dennis A Bloomfield, Sandro Vento, Bao-gan Peng	https://www.wignet.com/bpg/gerinfo/208		
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE		
https://www.wjgnet.com/2307-8960/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242		
PUBLICATION DATE January 16, 2021	STEPS FOR SUBMITTING MANUSCRIPTS https://www.wignet.com/bpg/GerInfo/239		
COPYRIGHT	ONLINE SUBMISSION		
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com		

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



W J C C World Journal of Clinical Cases

World Journal of

Submit a Manuscript: https://www.f6publishing.com

World J Clin Cases 2021 January 16; 9(2): 308-320

DOI: 10.12998/wjcc.v9.i2.308

ISSN 2307-8960 (online)

REVIEW

Overview of bile acid signaling in the cardiovascular system

Rou Zhang, Wen-Qi Ma, Meng-Jun Fu, Juan Li, Chun-Hua Hu, Yi Chen, Mi-Mi Zhou, Zhi-Jie Gao, Ying-Li He

ORCID number: Rou Zhang 0000-0002-7620-3424; Wen-Qi Ma 0000-0003-4570-9200; Meng-Jun Fu 0000-0002-7381-4916; Juan Li 0000-0002-8246-9798; Chun-Hua Hu 0000-0001-7292-6222; Yi Chen 0000-0001-6409-7479; Mi-Mi Zhou 0000-0001-8249-0207; Zhi-Jie Gao 0000-0002-1644-260X; Ying-Li He 0000-0001-9444-3678.

Author contributions: Zhang R performed the majority of the writing; Ma WQ prepared the figures and tables; Li J, Fu MJ, Hu CH, and Chen Y acquired the data and wrote the paper; Zhou MM and Gao ZJ revised the paper; He YL designed the outline and coordinated the writing of the paper; All authors have read and approve the final manuscript.

Supported by National Natural Science Foundation of China, No. 82070641.

Conflict-of-interest statement: All authors declare that: (1) No support, financial or otherwise, has been received from any organization that may have an interest in the submitted work; and (2) There are no other relationships or activities that could appear to have influenced the submitted work.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external

Rou Zhang, Wen-Qi Ma, Meng-Jun Fu, Juan Li, Chun-Hua Hu, Yi Chen, Mi-Mi Zhou, Zhi-Jie Gao, Ying-Li He, Department of Infectious Diseases, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Corresponding author: Ying-Li He, MD, PhD, Associate Chief Physician, Department of Infectious Diseases, The First Affiliated Hospital of Xi'an Jiaotong University, No. 277 Yanta Road (w), Xi'an 710061, Shaanxi Province, China. heyingli2000@xjtu.edu.cn

Abstract

Bile acids (BAs) are classically known to play a vital role in the metabolism of lipids and in absorption. It is now well established that BAs act as signaling molecules, activating different receptors (such as farnesoid X receptor, vitamin D receptor, Takeda G-protein-coupled receptor 5, sphingosine-1-phosphate, muscarinic receptors, and big potassium channels) and participating in the regulation of energy homeostasis and lipid and glucose metabolism. In addition, increased BAs can impair cardiovascular function in liver cirrhosis. Approximately 50% of patients with cirrhosis develop cirrhotic cardiomyopathy. Exposure to high concentrations of hydrophobic BAs has been shown to be related to adverse effects with respect to vascular tension, endothelial function, arrhythmias, coronary atherosclerotic heart disease, and heart failure. The BAs in the serum BA pool have relevant through their hydrophobicity, and the lipophilic BAs are more harmful to the heart. Interestingly, ursodeoxycholic acid is a hydrophilic BA, and it is used as a therapeutic drug to reverse and protect the harmful cardiac effects caused by hydrophobic elevated BAs. In order to elucidate the mechanism of BAs and cardiovascular function, abundant experiments have been conducted in vitro and in vivo. The aim of this review was to explore the mechanism of BAs in the cardiovascular system.

Key Words: Bile acids; Cardiovascular; Arteries; Receptors; Signaling; Cirrhosis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: In the literature, there are some reviews on the relationship between bile acids (BAs) and the cardiovascular system. However, this is the first review to use molecular and cellular mechanisms of related pathways to explore the possible mechanism of BAs in the pathogenesis of cardiovascular disease and to classify the role of BAs in



reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Unsolicited manuscript

Specialty type: Medicine, research and experimental

Country/Territory of origin: China

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: July 6, 2020 Peer-review started: July 6, 2020 First decision: September 12, 2020 Revised: September 28, 2020 Accepted: October 20, 2020 Article in press: October 20, 2020 Published online: January 16, 2021

P-Reviewer: Enosawa S, Park YM S-Editor: Huang P L-Editor: Filipodia P-Editor: Liu JH



heart and other organs using a tabular form. The goal was to provide readers a more comprehensive, deeper, and clearer understanding of the function of BAs.

Citation: Zhang R, Ma WQ, Fu MJ, Li J, Hu CH, Chen Y, Zhou MM, Gao ZJ, He YL. Overview of bile acid signaling in the cardiovascular system. World J Clin Cases 2021; 9(2): 308-320

URL: https://www.wjgnet.com/2307-8960/full/v9/i2/308.htm DOI: https://dx.doi.org/10.12998/wjcc.v9.i2.308

INTRODUCTION

Bile acids (BAs) comprise the primary catabolic pathway of cholesterol metabolism in the body, consisting of steroid cores and side-chains that can bind with either taurine or glycine groups^[1]. BAs are also the primary lipid component of bile, because their side chains can combine with carboxylic acid or sulfonic acid group, granting these molecules water-soluble and lipid-soluble amphiphilic properties. BAs are divided into the free and combined types according to their structure. Cholic acid (CA), chenodeoxycholic acid (CDCA), deoxycholic acid (DCA), lithocholic acid (LCA), and ursodeoxycholic acid (UDCA) constitute the main free BAs. The 24-carboxyl groups of the above free BAs combine with glycine or taurine to form combined BAs, which increase water solubility^[2]. The hydrophobicity of BAs depends on the location and number of hydroxyl groups in the structure of the ring, which is related to cytotoxicity that can be reduced by hydroxylation of the BAs^[3-5].

There are several mechanisms for the cytotoxicity of hydrophobic BAs. For example, BAs facilitate the production of reactive oxygen species (ROS) that oxidize and modify lipids, protein, and nucleic acids, eventually leading to apoptosis of the hepatocyte^[6]. In addition, hydrophobic BAs can activate liver Kupffer cells to produce ROS, which may further insult liver cells^[7]. In addition, mitochondria also play a role in the toxicity of BAs^[8]. The order of BA hydrophobicity is LCA > DCA > CDCA > CA > UDCA^[9] (Table 1). Hydrophilic BAs antagonize the cytotoxicity of hydrophobic BAs, and this antagonism correlates with their hydrophilicity^[10]. BAs are also divided into primary and secondary BAs according to their sources. Primary BAs are directly synthesized from cholesterol in liver cells and are stored in the gallbladder. When stimulated by food digestion, primary BAs are secreted into the intestine. Secondary BAs are produced from primary BAs by intestinal bacteria, which are then reabsorbed by the brush border cells of the small intestine and transported back to the liver through the hepatic portal vein circulation. In normal physiological conditions, approximately 95% of BAs are reabsorbed.

BA flow occurs continuously between the intestines and the liver and is called "enterohepatic circulation", which is a critical regulatory mechanism of the rate of BA metabolism, maintaining the balance of BAs and cholesterol in the body and preventing the formation of cholesterol stones^[11].

BAs AS SIGNALING MOLECULES

The farnesoid X receptor (FXR) was the first identified receptor of BA signaling, discovered in 1999^[12]. The G-protein-coupled receptor specific for BAs, the Takeda Gprotein-coupled receptor 5 (TGR5), was subsequently identified^[13]. Moreover, there are other types of receptors reportedly involved in regulating BA signaling, such as the muscarinic (M) receptors, sphingosine-1-phosphate (S1P), and large conductance voltage-and Ca2+-activated potassium (K+) [big potassium (BK)] channels. This section primary summarizes research on BAs as signaling molecules (Table 2)[14-28].

Nuclear receptor signaling

The FXR was identified as a BA receptor in 1999 and is highly expressed in the liver, kidney, and gastrointestinal tract^[29]. Since then, many studies on BA receptors have been performed, and CDCA is the most potent endogenous ligand of FXR. CDCA binding to FXR causes a conformational change of FXR, facilitating formation with the



Table 1 Hydrophobicity of bile acid increases with the decrease of OH groups						
Hydrophobicity	Types of bile acids					
Low	UDCA					
\downarrow	СА					
	CDCA					
	DCA					
High	LCA					

CA: Cholic acid; CDCA: Chenodeoxycholic acid; DCA: Deoxycholic acid; LCA: Lithocholic acid; UDCA: Ursodeoxycholic acid.

retinoic acid X receptor in the cytoplasm. The latter enters the nucleus to recognize target gene promoter regions and regulates the transcription of target genes by FXR response elements^[30]. For the LCA-induced cholestasis model, it was reported that CDCA activity down-regulated the bile salt export pump expression. This resulted in increased BA concentration and decreased liver bile secretion, which precipitated liver injury^[31]. CDCA can also activate the expression of FXR in the intestinal tract, thereby activating intestinal acid-binding protein expression to mediate cholesterol secretion^[32]. FXR is also expressed in the cardiovascular system, including in the coronary arteries, aorta, atherosclerotic arteries, and cardiomyocytes^[33]. It was reported that suppression of the proliferator-activated receptor- γ co-activator 1 α gene, which is a key regulator of fatty acid metabolism, caused development of cardiac dysfunction in FXR and SHP double-knockout mice (a model of cirrhosis)^[34]. This suggested that BAs may reduce plasma triglyceride levels and prevent signs of atherosclerosis^[35]. FXR ligands also inhibit the inflammatory response of rat aortic smooth muscle cells induced by interleukin-1β. The putative mechanism of this effect includes activating the ligand resistance of FXR binding nuclear factor-kappa B (NF-KB) to resist this proinflammatory pathway. This suggested that FXR agonists have anti-atherosclerosis potential^[36,37]. Liu et al^[38] and Yang et al^[39] reported that increased tumor necrosis factor alpha (TNFa) and NF-KB and decreased cardiac function were observed in bile duct ligation (BDL) animal models, and anti-TNFa antibody therapy significantly improved this cardiac dysfunction. These studies suggested that there are relationships among bacterial translocation, increased activity of the endocannabinoid TNFα, NF-κB, and cardiac dysfunction. BAs (primarily DCA) have antibacterial properties because they can destroy the integrity of bacteria, which can affect the composition of the intestinal microbiota^[40]. Pu et al^[41] found in cultured cardiomyocytes that activation of FXR through mitochondrial death signal transduction causes significant apoptosis, which was verified in a myocardial ischemia/reperfusion injury mouse model, demonstrating that FXR signaling might be involved in the growth and apoptosis of cardiomyocytes. These studies suggest that BAs play diverse roles by activating distinct receptors in different tissues, including different BAs that activate FXR receptors in the heart tissue to exert anti-atherosclerosis or pro-atherosclerosis functions.

After FXR was identified as a nuclear BA receptor, two other receptors, the pregnane X receptor (PXR) and the vitamin D receptor (VDR), were found^[42]. FXR, PXR, and VDR play essential roles in eliminating BA-induced toxicity by downregulating cholesterol 7a hydroxylase (CYP7A1) expression^[43], which is a rate-limiting enzyme for BA synthesis. In an animal study, activation of PXR was shown to regulate energy and lipid metabolism, thereby preventing obesity and insulin resistance caused by a high-fat diet. Thus, this study demonstrated the essential role of PXR in maintaining lipid metabolism^[44]. VDR is expressed in almost every tissue in the human body and is classified as an endocrine nuclear receptor^[45]. The VDR receptor can be activated by LCA, its natural ligands, and 1a, 25-dihydroxy-vitamin D3 [1a, 25 (OH)₂-D3]. Two ligands were found to activate the VDR signaling pathway through extracellular signal-regulated kinase 1/2, leading to phosphorylation of VDR and translocation into the nucleus. VDR can inhibit CYP7A1 transcription, thereby protecting hepatocyte cells from further damage due to cholestatic liver injury^[46]. It localizes the cardiomyocyte t-tubules. According to previous studies, the t-tube is the ideal location for regulating intracellular calcium flow and contractile forces. The inflow rate of calcium through calcium channels primarily determines the speed and pressure of myocardial contraction^[47]. Loss of VDR selectivity in cardiomyocytes leads to enlargement, hypertrophy, and systolic and diastolic dysfunction of cardiomy-



Table 2 Central receptor involved in bile acid signaling

Receptor	Tissue	Cardiovascular tissue	Ligand	Regulatory mechanism	Direct action	Ref.
FXR	Liver, kidney, gastrointestinal	Arteries, cardiomyocytes	CDCA, LCA, DCA	Regulates BSEP expression and participates in the process of liver injury, activates I-BABP expression to mediate cholesterol secretion, inhibits vascular inflammation, decreases lipogenesis synthesis, increases lipoprotein clearance to prevent atherosclerosis, ameliorates post-MI cardiac dysfunction and remodeling	No	[14-16]
PXR,VDR	Liver	t-Tubulescardiomyocytes	LCA	Down-regulates CYP7A1 expression to eliminate BA toxicity, maintains lipid metabolism, regulates intracellular calcium flow and contractile forces, maintains the normal operation of cardiomyocytes	No	[17-19]
М	Nervous, intestinal, gastrointestinal	Aorta, cardiomyocytes	TC, LCT, TCA	Stimulates acetylcholine-induced inositol phosphorylation and MAP kinase phosphorylation, inhibits cAMP, amplitude, reduces CM contraction	Yes	[20,21]
S1P	Liver, nervous, immune	Endothelial smooth muscle, cardiomyocytes	TCA, UDCA	Inhibits formation of cAMP and antagonizes adrenergic receptor-mediated contractile force, protects heart from ischemia/reperfusion injury, involved in remodeling and differentiation of cardiac fibroblasts	No	[22,23]
TGR5	Liver, glands, fat, muscle, enteric, immune	Aortic endothelial	CA, DCA, CDCA, TLCA, TCDCA	Inhibits the secretion of TNF induced by LPS, downregulates CSK3 and upregulates PKB associated with cardiac hypertrophy, metabolic transformation of energy in cardiomyocytes	Yes	[24,25]
BK channel	Liver, brain	Endothelial, cardiomyocytes	LCA	Cause vasodilation in liver and cerebral artery, plays role in diabetes, through mitochondrial BK channels confer cardioprotection	Yes	[26-28]

BA: Bile acid; BK channel: Large conductance voltage- and Ca²⁺-activated potassium (K⁺) (BK) channels; BSEP: Bile salt export pump; CA: Cholic acid; CAMP: Cyclic AMP; CDCA: Chenodeoxycholic acid; CM: Myocardial cell; CYP7A1: Cholesterol 7α hydroxylase; DCA: Deoxycholic acid; FXR: Farnesoid X receptor; GSK3: Glycogen synthase kinase-3; I-BABP: Encoding intestinal bile acid binding protein; LCA: Lithocholic acid; LCT: Lithocholyltaurine; LPS: Lipopolysaccharide; M: Muscarinic; MAP: Mitogen-activated; MI: Myocardial infarction; PKB: Protein kinase B; PXR: Pregnane X receptor; RXR: Retinoic acid X receptor; S1P: Sphingosine-1-phosphate; TCA: Taurocholate; TCA: Taurocholic acid; TOPCA: Taurocholic acid; VDR: Vitamin D receptor.

> ocytes^[48]. Vitamin D supplementation improves left ventricular structure and function in heart failure (HF) patients^[49], illustrating that expression of VDR is vital for maintaining the normal function of cardiomyocytes. Nuclear receptors are transcription factors and cannot explain the immediate changes in myocardial function caused by BA stimulation. However, as the duration of action of BAs varies, their possible effects on the heart vary.

Muscarinic receptor

BAs also interact with membrane receptors to activate a cascade of intracellular effectors. BAs interact with three membrane G-protein-coupled receptors, the M receptors, TGR5, and S1P receptor. M receptors are widely expressed in the gastrointestinal tract, intestinal smooth muscle, and central nervous system and can be divided into M1, M2, M3, M4, and M5 receptors. Among these, there are two classes based on conjugation with different G proteins that stimulate phosphoinositide hydrolysis (M1, M3, and M5) or inhibit adenylate cyclase (M2 and M4)^[50]. The ligand

for M3 receptors is the BA lithocholyltaurine^[51], which can stimulate acetylcholineinduced inositol phosphorylation and mitogen-activated kinase phosphorylation^[52]. Taurocholate binds to the M2 receptor, inhibiting cyclic AMP (cAMP), affecting transient calcium amplitude, and reducing myocardial cell contraction^[53]. The role of the remaining muscarinic receptors in the heart is unclear due to the lack of identified corresponding ligands.

S1P receptor

S1P is the most effective substrate for sphingolipids and is produced by phosphorylation of sphingolipids catalyzed by sphingolipid kinase. SIP determines cell fate through pro-apoptotic or survival signals. There are five subtypes of S1P receptor, which are S1P1, S1P2, S1P3, S1P4, and S1P5. Highly expressed S1P1 and S1P2 are detected in hepatocytes and activate extracellular signal-regulated kinase 1/2 and protein kinase B (PKB). S1P1, S1P2, and S1P3 receptors are primarily located in the heart, whereas S1P4 and S1P5 are limited to the nervous and immune systems^[54]. Taurocholic acid induces S1P2 receptor expression and promotes cholangiocarcinoma growth^[55]. In cardiomyocytes, the S1P1 receptor is the foremost expressed subtype, and its activation inhibits the formation of cAMP and antagonizes adrenergic receptormediated contractile force. Low levels of the S1P3 receptor mediate the bradycardia effect of S1P agonists. Studies have shown that S1P2 and S1P3 receptors play essential roles in heart protection from *in vivo*-mediated ischemia/reperfusion injury in mice using knockout mice. S1P receptors are also involved in proliferation, remodeling, and cardiac fibroblasts' differentiation. Furthermore, S1P receptors are found in smooth muscle cells and endothelial cells, which could mediate peripheral vascular tension and responses of the endothelium. Despite these findings, the role of the regulatory system in the cardiovascular system remains unclear^[56].

TGR5

TGR5 expression is detected in different cell types, such as fat cells, endocrine glands, muscle, immune cells, and the enteric nervous system. TGR5 has been reported to inhibit the response of rabbit alveolar macrophages to BAs (DCA, CDCA, and LCA), subsequently inhibiting the secretion of TNFa induced by lipopolysaccharide (LPS)^[57]. TGR5 also protects the liver by inhibiting expression of cytokines induced by LPS in Kupffer cells^[58]. LPS-induced inhibition of mitophagy increases oxidative stress and promotes inflammation in hepatic stellate cells during the process of acute liver failure^[59]. In recent years, TGR5 mRNA has been identified in human, rabbit, cow, and mouse heart tissues^[60]. The effects of mouse cardiac-specific TGR5 activated by taurodeoxycholic acid include LCA down-regulation of glycogen synthase kinase-3 and up-regulation of PKB, which are known to be associated with cardiac hypertrophy^[61]. TGR5 is also expressed in aortic endothelial cells that play an antiatherosclerotic role through producing nitric oxide (NO) in a dose-dependent manner, inhibiting NF-KB activity, and regulating monocyte adhesion and the inflammatory response^[62]. BA activation of TGR5 is also involved in the metabolic transformation of energy in cardiomyocytes. However, the mechanism of TGR5's action on cardiomyocytes remains to be clarified.

Large conductance voltage-and Ca2+-activated potassium (K+) (BK) channels

Studies have shown that in addition to known BA receptors (FXR, LXR, VDR, PXR, TGR5, M, and S1P), BAs can also activate nonclassical receptor reactions, such as largeconductance voltage-and Ca²⁺-activated potassium (K⁺) (BK) channels^[63]. It has been speculated that systemic vasodilation in hepatobiliary disease partly causes vascular smooth muscle cells' relaxation through the activation of BK_{Ca}. LCA induced a 30% increase in cerebral artery vasodilation in an endothelium-independent manner. This effect was eliminated in a BK β-1 subunit knockout mouse model, demonstrating that the role of this potassium channel subunit in diabetes is essential^[64]. In another study, BAs were shown to activate the BK pathway in cirrhosis patients and to increase the risk of developing cirrhotic cardiomyopathy. Meanwhile, taurine conjugated hydrophobic BAs activate BK channels, which can expand outward potassium currents, reduce the duration of action potentials, and exert negative inotropic effects^[65]. Since this receptor primarily mediates ion changes, it is speculated to play a primary role in the cardiac conduction related functions.

Zaishideng® WJCC | https://www.wjgnet.com

BAs AND CARDIOVASCULAR FUNCTION

BAs and vascular function

Studies have shown that BAs can regulate vascular tension. Increased BAs in the liver portal vein of rats with BDL were observed to decrease norepinephrine-induced vasoconstriction. These findings show that BAs are vasodilators. According to previous studies, the primary driver of cardiovascular disease is endothelial dysfunction, which leads to an imbalance in the synthesis and release of harmful and protective mediators, among which NO is the most important^[66,67]. FXR is identified in vascular smooth muscle cells and endothelium, and, as a transcription factor, it can regulate vascular relaxation and contraction by altering the term of vasoactive molecules or other receptors. Studies have shown that activated FXR induces vasodilation in endothelial cells by down-regulating endothelin-1 (IL-1), up-regulating endothelial NO synthase (eNOS), modulating angiotensin II receptor expression, and inhibiting inflammation and migration in vascular smooth muscle cells^[68]. It was found that CDCA activation leads to decreased IL-1 mRNA expression in a concentrationdependent manner. As is known, IL-1 is the most effective vasoconstrictor, and its BAinhibited expression may be an essential factor in systemic vasodilation in cirrhotic patients^[69]. The same team proposed the presence of FXR response elements in the promoter region of eNOS. Their activation led to up-regulation of eNOS and subsequent increases in the production of vasodilated NO^[70]. S1P receptor 2 (S1PR2) is another BA-sensitive receptor found in vascular smooth muscle cells that is involved in NO signaling. Nevertheless, it works through inhibiting the synthase of inducible NO, thus reducing a part of NO levels in vascular injury^[71-73].

In contrast, long-term stimulation of FXR weakens NO-dependent vasodilation due to increased cGMP passivation in smooth muscle cells. These observations suggest that short-term and long-term FXR stimulation has differential effects on NO production and sensitivity^[74]. Thus, time should be taken into account in the study of BA receptor-related effects. Pak *et al*^[75] found that BA increases can cause vasodilation, and speculated that this might occur by inhibiting calcium from passing through membrane channels. This effect has no relationship with blockers or endothelial stripping. However, it is strongly affected by the type of BAs, and hydrophobic and lipophilic BAs are more likely to induce vasodilation. The authors speculate on the mechanism by which BAs achieve this effect and conclude that they must directly interact with cell membrane components, emphasizing the role of BA components rather than merely the concept that increasing concentration is essential in the cardiovascular function.

BAs and coronary atherosclerotic heart disease

Recent clinical studies have found that fasting total bile acid (TBA) serum levels are closely related to the severity of coronary heart disease (CHD), serving as an indicator of the seriousness of the severity CHD^[76]. According to research, the fasting levels of BA concentration inhibit atherosclerosis^[77]. Animal models are resistant to developing atherosclerosis because they can excrete excess cholesterol by secreting large amounts of BAs into the intestines^[78]. We hypothesize that patients with coronary atherosclerotic disease might have impaired BA secretion and excretion, resulting in high serum cholesterol levels that promote progression of atherosclerotic lesions. Clinical studies have shown that fecal BA content in CHD patients is indeed significantly lower than that of non-CHD control groups^[79]. BAs and their synthetic derivatives exert antiatherogenic effects by activating the FXR receptor in some animal models. Oral administration of CDCA derivatives to apolipoprotein E-deficient mice reduced aortic plaque formation by 95% and reduced aortic expression of inflammatory factors, including IL-6, IL-1, etc.^[80]. In mammalian models, oral administration of BAs or their synthetic derivatives reduce serum triglycerides and total cholesterol levels, and inhibit the formation of atherosclerosis in a dose-dependent manner. These findings suggest that oral administration of BAs or their synthetic derivatives may represent a method for treating atherosclerotic lesions^[81].

Interestingly, Fxr^{-/-} mice showed pro-atherogenic lipid characteristics even when fed a high-fat/high-cholesterol diet but did not show enhanced atherosclerosis. Studies have demonstrated that CDCA or FXR ligand activated by FXR can reduce the activity of cardiomyocytes by triggering apoptosis of cardiomyocytes, promoting myocardial ischemia/reperfusion injury. These conflicting observations suggest that further *in vivo* studies are needed to determine the effect of the BA-FXR interaction on atherosclerosis.

Zaisbideng® WJCC | https://www.wjgnet.com

BAs and cardiac cell arrhythmia

Through previous studies, we know that the influence of BAs on cardiac function can be divided into indirect and direct effects. Direct effects require interaction between BAs and cardiomyocytes to affect the conduction and contraction of the myocardium. These effects may be receptor-dependent or independent. The cardiotoxicity of BAs was observed when high doses of BAs intravenously injected into animal specimens caused severe bradycardia. Further studies confirmed the dose-dependent negative time-varying effect of BAs on cardiomyocytes^[82]. Binash et al^[65] reported that in vitro sodium taurocholate slightly increased the outward potassium current, reduced the calcium current, and slowed the inward sodium current, ultimately reducing the duration of the action potential. Voltage clamp experiments in mice demonstrated that BAs decrease slow inward Na⁺ and Ca²⁺ currents and increase outward K⁺ currents. Of note, BAs can alter the function of heart muscle cells as pacemakers. In a partial *in vivo* study, investigators found the plasma nonursodeoxycholic BA ratio was significantly increased in the atrial fibrillation group. Data analysis showed that the serum ursodeoxycholic BA concentration and nonursodeoxycholic BA ratio were independent predictors of atrial fibrillation^[83]. BAs can affect the exchange of sodium and calcium on the myocardial cell membrane as polar amphiphilic molecules, leading to depolarization of the resting potential and inducing posterior depolarization of cells. Subsequent depolarization and triggering are one of the initiating mechanisms of HF.

BAs and HF

As the relationship between BAs and the heart continues to evolve, BAs have been shown to also play a role in HF^[84]. Vascular endothelial dysfunction is one of the critical manifestations of chronic HF. Injury to endothelial function will activate endothelium-dependent injury pathways, eventually leading to decreased exercise tolerance and affecting the quality of life in HF patients. One of the most important factors is TNF-a^[85]. LPS, which exists in the cell walls of Gram-negative bacteria in the gut, can enter the circulation through swollen intestinal mucosa due to decreased intestinal mucosal barrier function during the development of chronic HF^[86]. Secondary BAs are produced from primary BAs under the action of intestinal microorganisms, indicating that the intestinal flora is related to the severity of HF^[87]. In a recently published cross-sectional study, serum primary BA levels in patients with chronic HF decrease revealed specific secondary BA level increases and an increased ratio of secondary BAs to primary BAs. Therefore, the relationship between BAs and cardiomyocytes involves a complex regulatory system of multiple factors and multiple systems (organ, tissue, cellular, molecular, and endocrine) that interact with each other (as shown in Figure 1).

DISEASE THAT CAUSES BA ABNORMALITIES IS ASSOCIATED WITH THE HEART

Cirrhosis is often accompanied by cardiac dysfunction, which has aroused interest in the study of the relationship between abnormal BA metabolism and cardiac pathology. The relative risk of cardiovascular disease after primary sclerosing cholangitis onset was 3.34 and after primary biliary cholangitis (PBC) was 2.2^[88,99]. Significant prolongation of the corrected QT (QTc) interval was found in PBC patients, which may lead to ventricular arrhythmia and increase the risk of sudden death^[90,91]. Furthermore, studies have shown that patients with PBC have significantly reduced heart rate variability and stress-response sensitivity^[92].

Intrahepatic cholestasis of pregnancy (ICP) is a condition in which the mother's TBA concentration is higher than the normal range. ICP can cause accumulation of BAs in fetal serum, inducing fetal heart problems^[93]. Studies in patients with mild and severe ICP have shown that TBA concentration is associated with ventricular arrhythmia^[94].

Relationship between BAs, liver disease, and cardiomyopathy due to cirrhosis

Various causes of cholestatic disease eventually lead to cirrhosis^[95]. It is estimated that about half of all cirrhosis cases result in cirrhotic cardiomyopathy (CC), which is characterized by systolic or diastolic dysfunction and can lead to morphological changes in the heart^[96]. A prolonged QT interval is the most common feature of CC. The mechanism is unclear, but most studies have shown that it is determined by a combination of factors^[97]. Studies also found that the severity of cirrhosis is positively





Figure 1 Bile acids affect heart function in a number of ways and affect each other. BAs: Bile acids; eNOS: Endothelial nitric oxide synthase; IL-1: Interleukin-1; IL-6: Interleukin-6; LPS: Lipopolysaccharide; TNF-α: Tumor necrosis factor-alpha.

correlated with prolonged QTc septum, and it is more common in alcoholic cirrhosis^[98]. The mouse BDL model revealed that elevated BAs increase the production of NO by mediating intracellular Ca²⁺ signaling and induce apoptosis of cardiomyocytes, leading to CC. It showed that NO can also promote apoptosis and inhibit autophagy in hepatocellular carcinoma. Ma *et al*^[99] showed that reduced flow of the myocardial membrane in BDL rats, resulting in adrenergic dysfunction and the inability to produce cAMP, ultimately resulted in decreased contractility of the myocardium. However, most of the data obtained from the modeling of BDL has involved rodents, so the mechanism of application in the human body remains to be further confirmed.

UDCA has a protective effect on heart disease

UDCA is a highly hydrophilic BA that was initially used to treat chronic liver disease because it can dissolve cholesterol and reduce cholesterol absorption^[100]. In animal experiments, increased BA concentrations often lead to arrhythmia and decreased cardiac function, while UDCA does not. UDCA has both apoptotic and anti-apoptotic effects, suggesting that it plays distinct roles depending on the cell type^[101]. It protects the myocardium by counteracting more hydrophobic BAs, which is thought to be mediated by protein kinase C and intracellular calcium (Ca²⁺)^[102]. Lee et al^[103] found that UDCA protects myocardial damage by enhancing the recovery of systolic cardiac function during ischemia-reperfusion and reducing the release of lactate dehydrogenase during ischemia-reperfusion. In addition, the protective effect of UDCA on myocardial reperfusion injury in rats is thought to occur by inhibiting the mitochondrial permeability transition pore, which is dependent on the phosphoinositide 3 kinase/PKB pathway^[104]. In patients with HF, UDCA therapy has been shown to improve endothelial-dependent and nondependent vasodilation, thereby maintaining impaired NO production of arterial blood flow^[105]. In another clinical study, patients with chronic HF received 4 wk UDCA 500 mg twice daily, resulting in improved ischemic blood flow. Furthermore, levels of gammaglutamyltransferase, aspartate transaminase, and tumor necrosis factor receptor 1 (TNFR1) were reduced and liver function improved after treatment compared to before treatment^[106]. UDCA also protects fetal arrhythmia through BA-induced function^[107]. Although numerous studies on UDCA have shown that its use can improve myocardial injury, the mechanisms are not well understood. To further confirm the importance of UDCA in humans, more patients are needed for more comprehensive studies.

Zaishidena® WJCC | https://www.wjgnet.com

CONCLUSION

Based on the above discussion, we know that BA signaling plays a vital role through receptor-dependent (FXR, VDR, TGR5, S1P, M) and channel-mediated (BK channel) mechanisms in different cell types. Particularly, for the cardiac system, most studies have shown that BA signaling affects heart function, and cardiac dysfunction in liver diseases, such as cirrhosis and ICP, is common. Interestingly, UDCA was found to play a protective role in dysrhythmia. Future work should be devoted to deciphering the complex interactions between BAs and their receptors to provide a pharmacological basis for the clinical treatment of related diseases.

ACKNOWLEDGEMENTS

The authors thank the Xi'an Jiaotong University School of Medicine for the financial support and facilities provided.

REFERENCES

- Zhuang S, Li Q, Cai L, Wang C, Lei X. Chemoproteomic Profiling of Bile Acid Interacting 1 Proteins. ACS Cent Sci 2017; 3: 501-509 [PMID: 28573213 DOI: 10.1021/acscentsci.7b00134]
- Di Ciaula A, Garruti G, Lunardi Baccetto R, Molina-Molina E, Bonfrate L, Wang DQ, Portincasa P. Bile Acid Physiology. Ann Hepatol 2017; 16: s4-s14 [PMID: 29080336 DOI: 10.5604/01.3001.0010.5493]
- 3 Chen J, Zhao KN, Chen C. The role of CYP3A4 in the biotransformation of bile acids and therapeutic implication for cholestasis. Ann Transl Med 2014; 2: 7 [PMID: 25332983 DOI: 10.3978/j.issn.2305-5839.2013.03.02
- 4 Hofmann AF. Detoxification of lithocholic acid, a toxic bile acid: relevance to drug hepatotoxicity. Drug Metab Rev 2004; 36: 703-722 [PMID: 15554243 DOI: 10.1081/dmr-200033475]
- 5 Chen J, Raymond K. Nuclear receptors, bile-acid detoxification, and cholestasis. Lancet 2006; 367: 454-456 [PMID: 16473109 DOI: 10.1016/s0140-6736(06)68156-7]
- 6 Sokol RJ, Straka MS, Dahl R, Devereaux MW, Yerushalmi B, Gumpricht E, Elkins N, Everson G. Role of oxidant stress in the permeability transition induced in rat hepatic mitochondria by hydrophobic bile acids. Pediatr Res 2001; 49: 519-531 [PMID: 11264436 DOI: 10.1203/00006450-200104000-00014]
- Gong Z, Zhou J, Zhao S, Tian C, Wang P, Xu C, Chen Y, Cai W, Wu J. Chenodeoxycholic acid 7 activates NLRP3 inflammasome and contributes to cholestatic liver fibrosis. Oncotarget 2016; 7: 83951-83963 [PMID: 27924062 DOI: 10.18632/oncotarget.13796]
- Palmeira CM, Rolo AP. Mitochondrially-mediated toxicity of bile acids. Toxicology 2004; 203: 1-8 15 [PMID: 15363577 DOI: 10.1016/j.tox.2004.06.001]
- Perez MJ, Briz O. Bile-acid-induced cell injury and protection. World J Gastroenterol 2009; 15: 1677-1689 [PMID: 19360911 DOI: 10.3748/wjg.15.1677]
- Danchenko E, Petermann H, Chirkin A, Dargel R. Effect of bile acids on the proliferative activity 10 and apoptosis of rat hepatocytes. Exp Toxicol Pathol 2001; 53: 227-233 [PMID: 11484843 DOI: 10.1078/0940-2993-00178]
- Russell DW. The enzymes, regulation, and genetics of bile acid synthesis. Annu Rev Biochem 2003; 72: 137-174 [PMID: 12543708 DOI: 10.1146/annurev.biochem.72.121801.161712]
- 12 Ding L, Yang L, Wang Z, Huang W. Bile acid nuclear receptor FXR and digestive system diseases. Acta Pharm Sin B 2015; 5: 135-144 [PMID: 26579439 DOI: 10.1016/j.apsb.2015.01.004]
- Duboc H, Taché Y, Hofmann AF. The bile acid TGR5 membrane receptor: from basic research to 13 clinical application. Dig Liver Dis 2014; 46: 302-312 [PMID: 24411485 DOI: 10.1016/j.dld.2013.10.021]
- 14 Shaik FB, Prasad DV, Narala VR. Role of farnesoid X receptor in inflammation and resolution. Inflamm Res 2015; 64: 9-20 [PMID: 25376338 DOI: 10.1007/s00011-014-0780-y]
- 15 Mencarelli A, Fiorucci S. FXR an emerging therapeutic target for the treatment of atherosclerosis. J *Cell Mol Med* 2010; **14**: 79-92 [PMID: 20041971 DOI: 10.1111/j.1582-4934.2009.00997.x]
- Xia Y, Zhang F, Zhao S, Li Y, Chen X, Gao E, Xu X, Xiong Z, Zhang X, Zhang J, Zhao H, Wang 16 W, Wang H, Guo Y, Liu Y, Li C, Wang S, Zhang L, Yan W, Tao L. Adiponectin determines farnesoid X receptor agonism-mediated cardioprotection against post-infarction remodelling and dysfunction. Cardiovasc Res 2018; 114: 1335-1349 [PMID: 29668847 DOI: 10.1093/cvr/cvy093]
- Pike JW, Meyer MB, Lee SM, Onal M, Benkusky NA. The vitamin D receptor: contemporary 17 genomic approaches reveal new basic and translational insights. J Clin Invest 2017; 127: 1146-1154 [PMID: 28240603 DOI: 10.1172/JCI88887]
- 18 Zhang J, Huang W, Qatanani M, Evans RM, Moore DD. The constitutive androstane receptor and pregnane X receptor function coordinately to prevent bile acid-induced hepatotoxicity. J Biol Chem 2004; 279: 49517-49522 [PMID: 15358766 DOI: 10.1074/jbc.M409041200]



- 19 Glenn DJ, Cardema MC, Gardner DG. Amplification of lipotoxic cardiomyopathy in the VDR gene knockout mouse. J Steroid Biochem Mol Biol 2016; 164: 292-298 [PMID: 26429397 DOI: 10.1016/j.jsbmb.2015.09.034]
- 20 Ibrahim E, Diakonov I, Arunthavarajah D, Swift T, Goodwin M, McIlvride S, Nikolova V, Williamson C, Gorelik J. Bile acids and their respective conjugates elicit different responses in neonatal cardiomyocytes: role of Gi protein, muscarinic receptors and TGR5. Sci Rep 2018; 8: 7110 [PMID: 29740092 DOI: 10.1038/s41598-018-25569-4]
- Takagi N, Miyake-Takagi K, Takagi K, Tamura H, Takeo S. Altered extracellular signal-regulated 21 kinase signal transduction by the muscarinic acetylcholine and metabotropic glutamate receptors after cerebral ischemia. J Biol Chem 2002; 277: 6382-6390 [PMID: 11714707 DOI: 10.1074/jbc.M108081200]
- Donati C, Meacci E, Nuti F, Becciolini L, Farnararo M, Bruni P. Sphingosine 1-phosphate regulates 22 myogenic differentiation: a major role for S1P2 receptor. FASEB J 2005; 19: 449-451 [PMID: 15625079 DOI: 10.1096/fj.04-1780fje]
- 23 Takuwa Y, Okamoto Y, Yoshioka K, Takuwa N. Sphingosine-1-phosphate signaling and biological activities in the cardiovascular system. Biochim Biophys Acta 2008; 1781: 483-488 [PMID: 18472021 DOI: 10.1016/j.bbalip.2008.04.003]
- 24 Keitel V, Häussinger D. Role of TGR5 (GPBAR1) in Liver Disease. Semin Liver Dis 2018; 38: 333-339 [PMID: 30357770 DOI: 10.1055/s-0038-1669940]
- Eblimit Z, Thevananther S, Karpen SJ, Taegtmeyer H, Moore DD, Adorini L, Penny DJ, Desai MS. 25 TGR5 activation induces cytoprotective changes in the heart and improves myocardial adaptability to physiologic, inotropic, and pressure-induced stress in mice. Cardiovasc Ther 2018; 36: e12462 [PMID: 30070769 DOI: 10.1111/1755-5922.12462]
- 26 Bukiya AN, McMillan JE, Fedinec AL, Patil SA, Miller DD, Leffler CW, Parrill AL, Dopico AM. Cerebrovascular dilation via selective targeting of the cholane steroid-recognition site in the BK channel β1-subunit by a novel nonsteroidal agent. Mol Pharmacol 2013; 83: 1030-1044 [PMID: 23455312 DOI: 10.1124/mol.112.083519]
- Zhu Y, Ye P, Chen SL, Zhang DM. Functional regulation of large conductance Ca²⁺-activated K⁺ 27 channels in vascular diseases. Metabolism 2018; 83: 75-80 [PMID: 29373813 DOI: 10.1016/j.metabol.2018.01.008
- Bentzen BH, Olesen SP, Rønn LC, Grunnet M. BK channel activators and their therapeutic 28 perspectives. Front Physiol 2014; 5: 389 [PMID: 25346695 DOI: 10.3389/fphys.2014.00389]
- 29 Parks DJ, Blanchard SG, Bledsoe RK, Chandra G, Consler TG, Kliewer SA, Stimmel JB, Willson TM, Zavacki AM, Moore DD, Lehmann JM. Bile acids: natural ligands for an orphan nuclear receptor. Science 1999; 284: 1365-1368 [PMID: 10334993 DOI: 10.1126/science.284.5418.1365]
- 30 Laffitte BA, Kast HR, Nguyen CM, Zavacki AM, Moore DD, Edwards PA. Identification of the DNA binding specificity and potential target genes for the farnesoid X-activated receptor. J Biol Chem 2000; 275: 10638-10647 [PMID: 10744760 DOI: 10.1074/jbc.275.14.10638]
- 31 Yu J, Lo JL, Huang L, Zhao A, Metzger E, Adams A, Meinke PT, Wright SD, Cui J. Lithocholic acid decreases expression of bile salt export pump through farnesoid X receptor antagonist activity. J Biol Chem 2002; 277: 31441-31447 [PMID: 12052824 DOI: 10.1074/jbc.M200474200]
- 32 Tu H, Okamoto AY, Shan B. FXR, a bile acid receptor and biological sensor. Trends Cardiovasc Med 2000; 10: 30-35 [PMID: 11150726 DOI: 10.1016/s1050-1738(00)00043-8]
- 33 **Bishop-Bailey D**, Walsh DT, Warner TD. Expression and activation of the farnesoid X receptor in the vasculature. Proc Natl Acad Sci US A 2004; 101: 3668-3673 [PMID: 14990788 DOI: 10.1073/pnas.0400046101]
- 34 Desai MS, Mathur B, Eblimit Z, Vasquez H, Taegtmeyer H, Karpen SJ, Penny DJ, Moore DD, Anakk S. Bile acid excess induces cardiomyopathy and metabolic dysfunctions in the heart. Hepatology 2017; 65: 189-201 [PMID: 27774647 DOI: 10.1002/hep.28890]
- Hageman J, Herrema H, Groen AK, Kuipers F. A role of the bile salt receptor FXR in 35 atherosclerosis. Arterioscler Thromb Vasc Biol 2010; 30: 1519-1528 [PMID: 20631352 DOI: 10.1161/ATVBAHA.109.197897]
- Li YT, Swales KE, Thomas GJ, Warner TD, Bishop-Bailey D. Farnesoid x receptor ligands inhibit 36 vascular smooth muscle cell inflammation and migration. Arterioscler Thromb Vasc Biol 2007; 27: 2606-2611 [PMID: 18029909 DOI: 10.1161/ATVBAHA.107.152694]
- 37 Ghosh S, Dass JFP. Study of pathway cross-talk interactions with NF-κB leading to its activation via ubiquitination or phosphorylation: A brief review. Gene 2016; 584: 97-109 [PMID: 26968890 DOI: 10.1016/j.gene.2016.03.008
- Liu H, Lee SS. Nuclear factor-kappaB inhibition improves myocardial contractility in rats with 38 cirrhotic cardiomyopathy. Liver Int 2008; 28: 640-648 [PMID: 18346133 DOI: 10.1111/j.1478-3231.2008.01692.x]
- 39 Yang YY, Liu H, Nam SW, Kunos G, Lee SS. Mechanisms of TNFalpha-induced cardiac dysfunction in cholestatic bile duct-ligated mice: interaction between TNFalpha and endocannabinoids. J Hepatol 2010; 53: 298-306 [PMID: 20626112 DOI: 10.1016/j.jhep.2010.03.011]
- De Fabiani E, Mitro N, Gilardi F, Galmozzi A, Caruso D, Crestani M. When food meets man: the 40 contribution of epigenetics to health. Nutrients 2010; 2: 551-571 [PMID: 22254041 DOI: 10.3390/nu2050551]
- Pu J, Yuan A, Shan P, Gao E, Wang X, Wang Y, Lau WB, Koch W, Ma XL, He B. Cardiomyocyte-41



expressed farnesoid-X-receptor is a novel apoptosis mediator and contributes to myocardial ischaemia/reperfusion injury. Eur Heart J 2013; 34: 1834-1845 [PMID: 22307460 DOI: 10.1093/eurheartj/ehs011]

- 42 Adachi R, Shulman AI, Yamamoto K, Shimomura I, Yamada S, Mangelsdorf DJ, Makishima M. Structural determinants for vitamin D receptor response to endocrine and xenobiotic signals. Mol Endocrinol 2004; 18: 43-52 [PMID: 14525957 DOI: 10.1210/me.2003-0244]
- 43 Jung D, Mangelsdorf DJ, Meyer UA. Pregnane X receptor is a target of farnesoid X receptor. J Biol Chem 2006; 281: 19081-19091 [PMID: 16682417 DOI: 10.1074/jbc.M600116200]
- 44 Ma Y, Liu D. Activation of pregnane X receptor by pregnenolone 16 α-carbonitrile prevents high-fat diet-induced obesity in AKR/J mice. PLoS One 2012; 7: e38734 [PMID: 22723881 DOI: 10.1371/journal.pone.0038734]
- Bouillon R, Carmeliet G, Verlinden L, van Etten E, Verstuyf A, Luderer HF, Lieben L, Mathieu C, 45 Demay M. Vitamin D and human health: lessons from vitamin D receptor null mice. Endocr Rev 2008; 29: 726-776 [PMID: 18694980 DOI: 10.1210/er.2008-0004]
- 46 Han S, Li T, Ellis E, Strom S, Chiang JY. A novel bile acid-activated vitamin D receptor signaling in human hepatocytes. Mol Endocrinol 2010; 24: 1151-1164 [PMID: 20371703 DOI: 10.1210/me.2009-0482]
- Tishkoff DX, Nibbelink KA, Holmberg KH, Dandu L, Simpson RU. Functional vitamin D receptor (VDR) in the t-tubules of cardiac myocytes: VDR knockout cardiomyocyte contractility. Endocrinology 2008; 149: 558-564 [PMID: 17974622 DOI: 10.1210/en.2007-0805]
- 48 Chen S, Law CS, Grigsby CL, Olsen K, Hong TT, Zhang Y, Yeghiazarians Y, Gardner DG. Cardiomyocyte-specific deletion of the vitamin D receptor gene results in cardiac hypertrophy. Circulation 2011; 124: 1838-1847 [PMID: 21947295 DOI: 10.1161/CIRCULATIONAHA.111.032680]
- Rodriguez AJ, Mousa A, Ebeling PR, Scott D, de Courten B. Effects of vitamin D supplementation 49 on inflammatory markers in heart failure: a systematic review and meta-analysis of randomized controlled trials. Sci Rep 2018; 8: 1169 [PMID: 29348609 DOI: 10.1038/s41598-018-19708-0]
- 50 Wess J, Eglen RM, Gautam D. Muscarinic acetylcholine receptors: mutant mice provide new insights for drug development. Nat Rev Drug Discov 2007; 6: 721-733 [PMID: 17762886 DOI: 10.1038/nrd2379]
- Cheng K, Khurana S, Chen Y, Kennedy RH, Zimniak P, Raufman JP. Lithocholylcholine, a bile 51 acid/acetylcholine hybrid, is a muscarinic receptor antagonist. J Pharmacol Exp Ther 2002; 303: 29-35 [PMID: 12235229 DOI: 10.1124/jpet.102.036376]
- 52 Raufman JP, Chen Y, Cheng K, Compadre C, Compadre L, Zimniak P. Selective interaction of bile acids with muscarinic receptors: a case of molecular mimicry. Eur J Pharmacol 2002; 457: 77-84 [PMID: 12464352 DOI: 10.1016/s0014-2999(02)02690-0]
- Sheikh Abdul Kadir SH, Miragoli M, Abu-Hayyeh S, Moshkov AV, Xie Q, Keitel V, Nikolaev 53 VO, Williamson C, Gorelik J. Bile acid-induced arrhythmia is mediated by muscarinic M2 receptors in neonatal rat cardiomyocytes. PLoS One 2010; 5: e9689 [PMID: 20300620 DOI: 10.1371/journal.pone.0009689
- 54 Serriere-Lanneau V, Teixeira-Clerc F, Li L, Schippers M, de Wries W, Julien B, Tran-Van-Nhieu J, Manin S, Poelstra K, Chun J, Carpentier S, Levade T, Mallat A, Lotersztajn S. The sphingosine 1phosphate receptor S1P2 triggers hepatic wound healing. FASEB J 2007; 21: 2005-2013 [PMID: 17341687 DOI: 10.1096/fj.06-6889com]
- 55 Liu R, Li X, Qiang X, Luo L, Hylemon PB, Jiang Z, Zhang L, Zhou H. Taurocholate Induces Cyclooxygenase-2 Expression via the Sphingosine 1-phosphate Receptor 2 in a Human Cholangiocarcinoma Cell Line. J Biol Chem 2015; 290: 30988-31002 [PMID: 26518876 DOI: 10.1074/jbc.M115.668277]
- Means CK, Brown JH. Sphingosine-1-phosphate receptor signalling in the heart. Cardiovasc Res 56 2009; 82: 193-200 [PMID: 19282351 DOI: 10.1093/cvr/cvp086]
- Kawamata Y, Fujii R, Hosoya M, Harada M, Yoshida H, Miwa M, Fukusumi S, Habata Y, Itoh T, 57 Shintani Y, Hinuma S, Fujisawa Y, Fujino M. A G protein-coupled receptor responsive to bile acids. J Biol Chem 2003; 278: 9435-9440 [PMID: 12524422 DOI: 10.1074/jbc.M209706200]
- 58 Keitel V, Donner M, Winandy S, Kubitz R, Häussinger D. Expression and function of the bile acid receptor TGR5 in Kupffer cells. Biochem Biophys Res Commun 2008; 372: 78-84 [PMID: 18468513 DOI: 10.1016/j.bbrc.2008.04.171]
- Zhang X, Jin L, Tian Z, Wang J, Yang Y, Liu J, Chen Y, Hu C, Chen T, Zhao Y, He Y. Nitric oxide 59 inhibits autophagy and promotes apoptosis in hepatocellular carcinoma. Cancer Sci 2019; 110: 1054-1063 [PMID: 30657629 DOI: 10.1111/cas.13945]
- Watanabe M, Houten SM, Mataki C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney 60 JW, Ezaki O, Kodama T, Schoonjans K, Bianco AC, Auwerx J. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. Nature 2006; 439: 484-489 [PMID: 16400329 DOI: 10.1038/nature04330]
- 61 Desai MS, Shabier Z, Taylor M, Lam F, Thevananther S, Kosters A, Karpen SJ. Hypertrophic cardiomyopathy and dysregulation of cardiac energetics in a mouse model of biliary fibrosis. Hepatology 2010; 51: 2097-2107 [PMID: 20512997 DOI: 10.1002/hep.23585]
- 62 Kida T, Tsubosaka Y, Hori M, Ozaki H, Murata T. Bile acid receptor TGR5 agonism induces NO production and reduces monocyte adhesion in vascular endothelial cells. Arterioscler Thromb Vasc Biol 2013; 33: 1663-1669 [PMID: 23619297 DOI: 10.1161/ATVBAHA.113.301565]



- 63 Dopico AM, Walsh JV Jr, Singer JJ. Natural bile acids and synthetic analogues modulate large conductance Ca2+-activated K+ (BKCa) channel activity in smooth muscle cells. J Gen Physiol 2002; 119: 251-273 [PMID: 11865021 DOI: 10.1085/jgp.20028537]
- 64 Bukiya AN, Liu J, Toro L, Dopico AM. Beta1 (KCNMB1) subunits mediate lithocholate activation of large-conductance Ca2+-activated K+ channels and dilation in small, resistance-size arteries. Mol Pharmacol 2007; 72: 359-369 [PMID: 17468198 DOI: 10.1124/mol.107.034330]
- 65 Binah O, Rubinstein I, Bomzon A, Better OS. Effects of bile acids on ventricular muscle contraction and electrophysiological properties: studies in rat papillary muscle and isolated ventricular myocytes. Naunyn Schmiedebergs Arch Pharmacol 1987; 335: 160-165 [PMID: 3561530 DOI: 10.1007/BF00177718]
- 66 Fiorucci S, Zampella A, Cirino G, Bucci M, Distrutti E. Decoding the vasoregulatory activities of bile acid-activated receptors in systemic and portal circulation: role of gaseous mediators. Am J Physiol Heart Circ Physiol 2017; 312: H21-H32 [PMID: 27765751 DOI: 10.1152/ajpheart.00577.2016]
- 67 Guizoni DM, Vettorazzi JF, Carneiro EM, Davel AP. Modulation of endothelium-derived nitric oxide production and activity by taurine and taurine-conjugated bile acids. Nitric Oxide 2020; 94: 48-53 [PMID: 31669041 DOI: 10.1016/j.niox.2019.10.008]
- Zhang Q, He F, Kuruba R, Gao X, Wilson A, Li J, Billiar TR, Pitt BR, Xie W, Li S. FXR-mediated 68 regulation of angiotensin type 2 receptor expression in vascular smooth muscle cells. Cardiovasc Res 2008; 77: 560-569 [PMID: 18006431 DOI: 10.1093/cvr/cvm068]
- He F, Li J, Mu Y, Kuruba R, Ma Z, Wilson A, Alber S, Jiang Y, Stevens T, Watkins S, Pitt B, Xie W, Li S. Downregulation of endothelin-1 by farnesoid X receptor in vascular endothelial cells. Circ Res 2006; 98: 192-199 [PMID: 16357303 DOI: 10.1161/01.RES.0000200400.55539.85]
- 70 Li J, Wilson A, Kuruba R, Zhang Q, Gao X, He F, Zhang LM, Pitt BR, Xie W, Li S. FXR-mediated regulation of eNOS expression in vascular endothelial cells. Cardiovasc Res 2008; 77: 169-177 [PMID: 18006476 DOI: 10.1093/cvr/cvm016]
- 71 Nakajima T, Okuda Y, Chisaki K, Shin WS, Iwasawa K, Morita T, Matsumoto A, Suzuki JI, Suzuki S, Yamada N, Toyo-Oka T, Nagai R, Omata M. Bile acids increase intracellular Ca(2+) concentration and nitric oxide production in vascular endothelial cells. Br J Pharmacol 2000; 130: 1457-1467 [PMID: 10928945 DOI: 10.1038/sj.bjp.0703471]
- 72 Khurana S, Raina H, Pappas V, Raufman JP, Pallone TL. Effects of deoxycholylglycine, a conjugated secondary bile acid, on myogenic tone and agonist-induced contraction in rat resistance arteries. PLoS One 2012; 7: e32006 [PMID: 22359652 DOI: 10.1371/journal.pone.0032006]
- 73 Machida T, Matamura R, Iizuka K, Hirafuji M. Cellular function and signaling pathways of vascular smooth muscle cells modulated by sphingosine 1-phosphate. J Pharmacol Sci 2016; 132: 211-217 [PMID: 27581589 DOI: 10.1016/j.jphs.2016.05.010]
- 74 Kida T, Murata T, Hori M, Ozaki H. Chronic stimulation of farnesoid X receptor impairs nitric oxide sensitivity of vascular smooth muscle. Am J Physiol Heart Circ Physiol 2009; 296: H195-H201 [PMID: 19011043 DOI: 10.1152/ajpheart.00679.2008]
- 75 Pak JM, Adeagbo AS, Triggle CR, Shaffer EA, Lee SS. Mechanism of bile salt vasoactivity: dependence on calcium channels in vascular smooth muscle. Br J Pharmacol 1994; 112: 1209-1215 [PMID: 7952883 DOI: 10.1111/j.1476-5381.1994.tb13212.x]
- 76 Li W, Shu S, Cheng L, Hao X, Wang L, Wu Y, Yuan Z, Zhou J. Fasting serum total bile acid level is associated with coronary artery disease, myocardial infarction and severity of coronary lesions. Atherosclerosis 2020; 292: 193-200 [PMID: 31811964 DOI: 10.1016/j.atherosclerosis.2019.11.026]
- 77 Pols TW. TGR5 in inflammation and cardiovascular disease. Biochem Soc Trans 2014; 42: 244-249 [PMID: 24646225 DOI: 10.1042/BST20130279]
- Charach G, Rabinovich A, Argov O, Weintraub M, Rabinovich P. The role of bile Acid excretion in 78 atherosclerotic coronary artery disease. Int J Vasc Med 2012; 2012: 949672 [PMID: 21918722 DOI: 10.1155/2012/949672]
- 79 Gylling H, Hallikainen M, Rajaratnam RA, Simonen P, Pihlajamäki J, Laakso M, Miettinen TA. The metabolism of plant sterols is disturbed in postmenopausal women with coronary artery disease. Metabolism 2009; 58: 401-407 [PMID: 19217458 DOI: 10.1016/j.metabol.2008.10.015]
- 80 Hanniman EA, Lambert G, McCarthy TC, Sinal CJ. Loss of functional farnesoid X receptor increases atherosclerotic lesions in apolipoprotein E-deficient mice. J Lipid Res 2005; 46: 2595-2604 [PMID: 16186601 DOI: 10.1194/jlr.M500390-JLR200]
- Ali AH, Carey EJ, Lindor KD. Recent advances in the development of farnesoid X receptor agonists. 81 Ann Transl Med 2015; 3: 5 [PMID: 25705637 DOI: 10.3978/j.issn.2305-5839.2014.12.06]
- 82 Joubert P. An in vivo investigation of the negative chronotropic effect of cholic acid in the rat. Clin *Exp Pharmacol Physiol* 1978; **5**: 1-8 [PMID: 639354 DOI: 10.1111/j.1440-1681.1978.tb00645.x]
- Rainer PP, Primessnig U, Harenkamp S, Doleschal B, Wallner M, Fauler G, Stojakovic T, Wachter 83 R, Yates A, Groschner K, Trauner M, Pieske BM, von Lewinski D. Bile acids induce arrhythmias in human atrial myocardium--implications for altered serum bile acid composition in patients with atrial fibrillation. Heart 2013; 99: 1685-1692 [PMID: 23894089 DOI: 10.1136/heartjnl-2013-304163]
- Mayerhofer CCK, Ueland T, Broch K, Vincent RP, Cross GF, Dahl CP, Aukrust P, Gullestad L, Hov JR, Trøseid M. Increased Secondary/Primary Bile Acid Ratio in Chronic Heart Failure. J Card Fail 2017; 23: 666-671 [PMID: 28688889 DOI: 10.1016/j.cardfail.2017.06.007]
- Schumacher SM, Naga Prasad SV. Tumor Necrosis Factor-α in Heart Failure: an Updated Review. 85



Curr Cardiol Rep 2018; 20: 117 [PMID: 30259192 DOI: 10.1007/s11886-018-1067-7]

- 86 Pasini E, Aquilani R, Testa C, Baiardi P, Angioletti S, Boschi F, Verri M, Dioguardi F. Pathogenic Gut Flora in Patients With Chronic Heart Failure. JACC Heart Fail 2016; 4: 220-227 [PMID: 26682791 DOI: 10.1016/j.jchf.2015.10.009]
- 87 Kitai T, Tang WHW. Gut microbiota in cardiovascular disease and heart failure. Clin Sci (Lond) 2018; 132: 85-91 [PMID: 29326279 DOI: 10.1042/CS20171090]
- 88 Ludvigsson JF, Bergquist A, Montgomery SM, Bahmanyar S. Risk of diabetes and cardiovascular disease in patients with primary sclerosing cholangitis. J Hepatol 2014; 60: 802-808 [PMID: 24291242 DOI: 10.1016/j.jhep.2013.11.017]
- Czul F, Peyton A, Levy C. Primary biliary cirrhosis: therapeutic advances. Clin Liver Dis 2013; 17: 89 229-242 [PMID: 23540499 DOI: 10.1016/j.cld.2012.12.003]
- Kempler P, Váradi A, Kádar E, Szalay F. Autonomic and peripheral neuropathy in primary biliary 90 cirrhosis: evidence of small sensory fibre damage and prolongation of the QT interval. J Hepatol 1994; 21: 1150-1151 [PMID: 7699249 DOI: 10.1016/s0168-8278(05)80640-3]
- 91 Bogaard K, van der Steen MS, Tan HL, Tukkie R. Short-coupled variant of torsade de pointes. Neth Heart J 2008; 16: 246-249 [PMID: 18711611 DOI: 10.1007/BF03086155]
- 92 Newton JL, Elliott C, Frith J, Ghazala C, Pairman J, Jones DE. Functional capacity is significantly impaired in primary biliary cirrhosis and is related to orthostatic symptoms. Eur J Gastroenterol Hepatol 2011; 23: 566-572 [PMID: 21593676 DOI: 10.1097/MEG.0b013e3283470256]
- 93 Geenes V, Lövgren-Sandblom A, Benthin L, Lawrance D, Chambers J, Gurung V, Thornton J, Chappell L, Khan E, Dixon P, Marschall HU, Williamson C. The reversed feto-maternal bile acid gradient in intrahepatic cholestasis of pregnancy is corrected by ursodeoxycholic acid. PLoS One 2014; 9: e83828 [PMID: 24421907 DOI: 10.1371/journal.pone.0083828]
- 94 Kirbas O, Biberoglu EH, Kirbas A, Daglar K, Kurmus O, Danisman N, Biberoglu K. Evaluation of ventricular repolarization in pregnant women with intrahepatic cholestasis. Int J Cardiol 2015; 189: 25-29 [PMID: 25885869 DOI: 10.1016/j.ijcard.2015.04.001]
- 95 Woolbright BL. Inflammation: Cause or consequence of chronic cholestatic liver injury. Food Chem Toxicol 2020; 137: 111133 [PMID: 31972189 DOI: 10.1016/j.fct.2020.111133]
- 96 Ruiz-del-Arbol L, Serradilla R. Cirrhotic cardiomyopathy. World J Gastroenterol 2015; 21: 11502-11521 [PMID: 26556983 DOI: 10.3748/wjg.v21.i41.11502]
- 97 Zambruni A, Trevisani F, Caraceni P, Bernardi M. Cardiac electrophysiological abnormalities in patients with cirrhosis. J Hepatol 2006; 44: 994-1002 [PMID: 16510203 DOI: 10.1016/j.jhep.2005.10.034]
- 98 Mozos I, Costea C, Serban C, Susan L. Factors associated with a prolonged QT interval in liver cirrhosis patients. J Electrocardiol 2011; 44: 105-108 [PMID: 21146831 DOI: 10.1016/j.jelectrocard.2010.10.034]
- 99 Ma Z, Lee SS, Meddings JB. Effects of altered cardiac membrane fluidity on beta-adrenergic receptor signalling in rats with cirrhotic cardiomyopathy. J Hepatol 1997; 26: 904-912 [PMID: 9126806 DOI: 10.1016/s0168-8278(97)80259-0]
- 100 Ambros-Rudolph CM, Glatz M, Trauner M, Kerl H, Müllegger RR. The importance of serum bile acid level analysis and treatment with ursodeoxycholic acid in intrahepatic cholestasis of pregnancy: a case series from central Europe. Arch Dermatol 2007; 143: 757-762 [PMID: 17576942 DOI: 10.1001/archderm.143.6.757
- 101 Rodrigues CM, Fan G, Ma X, Kren BT, Steer CJ. A novel role for ursodeoxycholic acid in inhibiting apoptosis by modulating mitochondrial membrane perturbation. J Clin Invest 1998; 101: 2790-2799 [PMID: 9637713 DOI: 10.1172/JCI1325]
- 102 Murakami M, Une N, Nishizawa M, Suzuki S, Ito H, Horiuchi T. Incretin secretion stimulated by ursodeoxycholic acid in healthy subjects. Springerplus 2013; 2: 20 [PMID: 23450079 DOI: 10.1186/2193-1801-2-20
- 103 Lee WY, Han SH, Cho TS, Yoo YH, Lee SM. Effect of ursodeoxycholic acid on ischemia/reperfusion injury in isolated rat heart. Arch Pharm Res 1999; 22: 479-484 [PMID: 10549575 DOI: 10.1007/BF02979156]
- 104 Rajesh KG, Suzuki R, Maeda H, Yamamoto M, Yutong X, Sasaguri S. Hydrophilic bile salt ursodeoxycholic acid protects myocardium against reperfusion injury in a PI3K/Akt dependent pathway. J Mol Cell Cardiol 2005; 39: 766-776 [PMID: 16171810 DOI: 10.1016/j.yjmcc.2005.07.014]
- Sinisalo J, Vanhanen H, Pajunen P, Vapaatalo H, Nieminen MS. Ursodeoxycholic acid and 105 endothelial-dependent, nitric oxide-independent vasodilatation of forearm resistance arteries in patients with coronary heart disease. Br J Clin Pharmacol 1999; 47: 661-665 [PMID: 10383544 DOI: 10.1046/j.1365-2125.1999.00940.x]
- 106 von Haehling S, Schefold JC, Jankowska EA, Springer J, Vazir A, Kalra PR, Sandek A, Fauler G, Stojakovic T, Trauner M, Ponikowski P, Volk HD, Doehner W, Coats AJ, Poole-Wilson PA, Anker SD. Ursodeoxycholic acid in patients with chronic heart failure: a double-blind, randomized, placebo-controlled, crossover trial. J Am Coll Cardiol 2012; 59: 585-592 [PMID: 22300693 DOI: 10.1016/j.jacc.2011.10.880]
- 107 Miragoli M, Kadir SH, Sheppard MN, Salvarani N, Virta M, Wells S, Lab MJ, Nikolaev VO, Moshkov A, Hague WM, Rohr S, Williamson C, Gorelik J. A protective antiarrhythmic role of ursodeoxycholic acid in an in vitro rat model of the cholestatic fetal heart. Hepatology 2011; 54: 1282-1292 [PMID: 21809354 DOI: 10.1002/hep.24492]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

