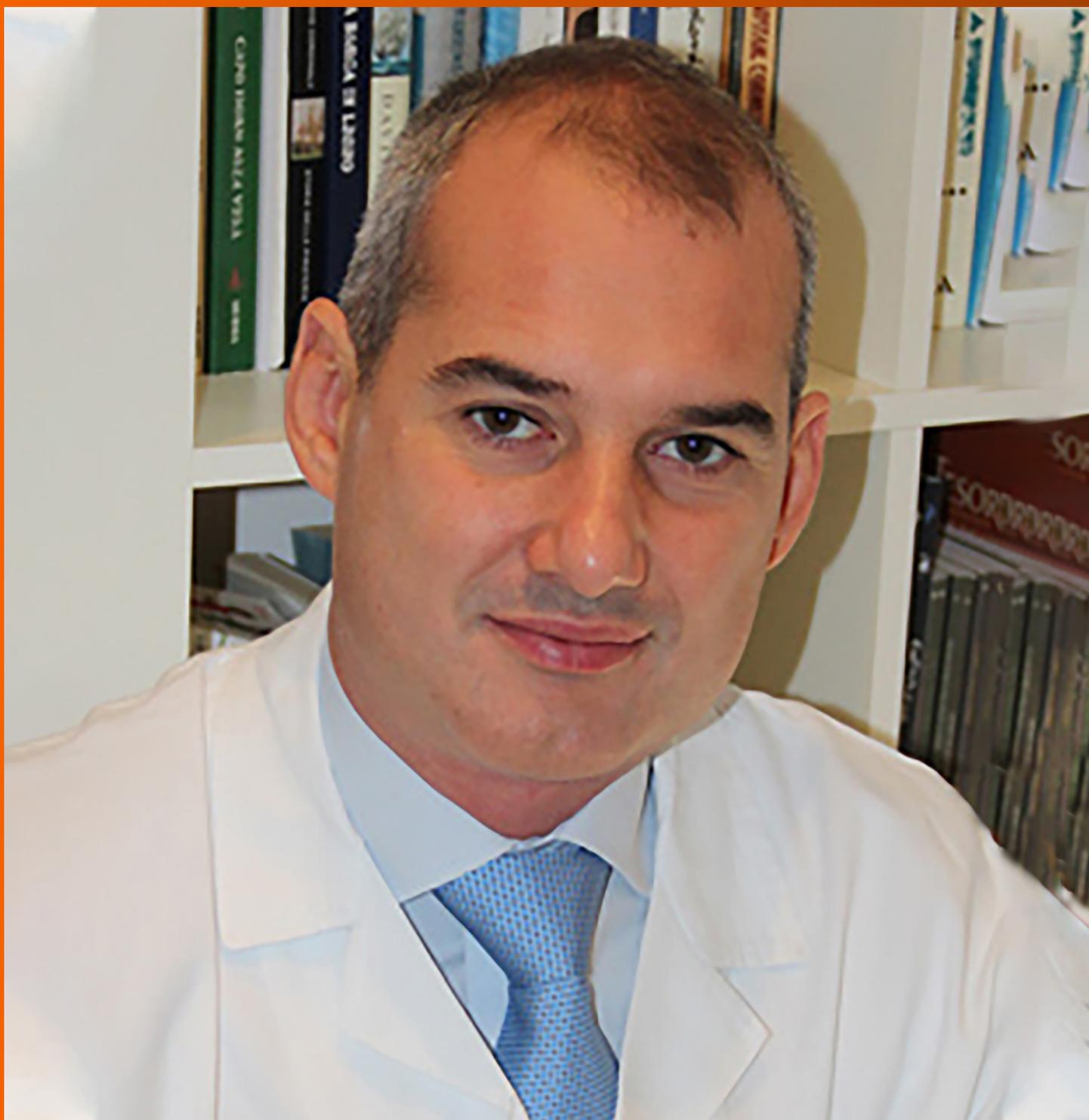


World Journal of *Clinical Cases*

World J Clin Cases 2021 August 16; 9(23): 6582-6963



OPINION REVIEW

- 6582 COVID-19 pandemic, as experienced in the surgical service of a district hospital in Spain
Pérez Lara FJ, Jimenez Martinez MB, Pozo Muñoz F, Fontalba Navas A, Garcia Cisneros R, Garcia Larrosa MJ, Garcia Delgado I, Callejon Gil MDM

REVIEW

- 6591 Beta-carotene and its protective effect on gastric cancer
Chen QH, Wu BK, Pan D, Sang LX, Chang B
- 6608 Liver transplantation during global COVID-19 pandemic
Alfishawy M, Nso N, Nassar M, Ariyaratnam J, Bhuiyan S, Siddiqui RS, Li M, Chung H, Al Balakosy A, Alqassieh A, Fülöp T, Rizzo V, Daoud A, Soliman KM
- 6624 Nonalcoholic fatty pancreas disease: An emerging clinical challenge
Zhang CL, Wang JJ, Li JN, Yang Y

MINIREVIEWS

- 6639 Novel mechanism of hepatobiliary system damage and immunoglobulin G4 elevation caused by *Clonorchis sinensis* infection
Zhang XH, Huang D, Li YL, Chang B
- 6654 Intestinal microbiota participates in nonalcoholic fatty liver disease progression by affecting intestinal homeostasis
Zhang Y, Li JX, Zhang Y, Wang YL
- 6663 Theory and reality of antivirals against SARS-CoV-2
Zhao B, Yang TF, Zheng R
- 6674 Acute acalculous cholecystitis due to infectious causes
Markaki I, Konsoula A, Markaki L, Spornovasilis N, Papadakis M

ORIGINAL ARTICLE**Case Control Study**

- 6686 Innate immunity – the hallmark of *Helicobacter pylori* infection in pediatric chronic gastritis
Meliş LE, Mărginean CO, Săsăran MO, Mocan S, Ghiga DV, Bogliş A, Duicu C

Retrospective Study

- 6698 Effects on newborns of applying bupivacaine combined with different doses of fentanyl for cesarean section
Wang Y, Liu WX, Zhou XH, Yang M, Liu X, Zhang Y, Hai KR, Ye QS

- 6705** Awake fiberoptic intubation and use of bronchial blockers in ankylosing spondylitis patients
Yang SZ, Huang SS, Yi WB, Lv WW, Li L, Qi F
- 6717** Efficacy of different antibiotics in treatment of children with respiratory mycoplasma infection
Zhang MY, Zhao Y, Liu JF, Liu GP, Zhang RY, Wang LM
- 6725** Expression of caspase-3 and hypoxia inducible factor 1 α in hepatocellular carcinoma complicated by hemorrhage and necrosis
Liang H, Wu JG, Wang F, Chen BX, Zou ST, Wang C, Luo SW
- 6734** Increased morbidity and mortality of hepatocellular carcinoma patients in lower cost of living areas
Sempokuya T, Patel KP, Azawi M, Ma J, Wong LL

SYSTEMATIC REVIEWS

- 6747** Safety of pancreatic surgery with special reference to antithrombotic therapy: A systematic review of the literature
Fujikawa T, Naito S
- 6759** What paradigm shifts occurred in the management of acute diverticulitis during the COVID-19 pandemic? A scoping review
Gallo G, Ortenzi M, Grossi U, Di Tanna GL, Pata F, Guerrieri M, Sammarco G, Di Saverio S

CASE REPORT

- 6768** Pylephlebitis – a rare complication of a fish bone migration mimicking metastatic pancreatic cancer: A case report
Bezerra S, França NJ, Mineiro F, Capela G, Duarte C, Mendes AR
- 6775** Solitary seminal vesicle metastasis from ileal adenocarcinoma presenting with hematospermia: A case report
Cheng XB, Lu ZQ, Lam W, Yiu MK, Li JS
- 6781** Hepatic abscess caused by esophageal foreign body misdiagnosed as cystadenocarcinoma by magnetic resonance imaging: A case report
Pan W, Lin LJ, Meng ZW, Cai XR, Chen YL
- 6789** 2+0 CYP21A2 deletion carrier – a limitation of the genetic testing and counseling: A case report
Xi N, Song X, Wang XY, Qin SF, He GN, Sun LL, Chen XM
- 6798** Psoriasis treatment using minimally manipulated umbilical cord-derived mesenchymal stem cells: A case report
Ahn H, Lee SY, Jung WJ, Pi J, Lee KH
- 6804** Double intussusception in a teenage child with Peutz-Jeghers syndrome: A case report
Chiew J, Sambanthan ST, Mahendran HA

- 6810** Nedaplatin-induced syndrome of inappropriate secretion of antidiuretic hormone: A case report and review of the literature
Tian L, He LY, Zhang HZ
- 6816** Nasal metastases from neuroblastoma-a rare entity: Two case reports
Zhang Y, Guan WB, Wang RF, Yu WW, Jiang RQ, Liu Y, Wang LF, Wang J
- 6824** Nocardiosis with diffuse involvement of the pleura: A case report
Wang P, Yi ML, Zhang CZ
- 6832** Prenatal diagnosis of triphalangeal thumb-polysyndactyly syndrome by ultrasonography combined with genetic testing: A case report
Zhang SJ, Lin HB, Jiang QX, He SZ, Lyu GR
- 6839** Blue LED as a new treatment to vaginal stenosis due pelvic radiotherapy: Two case reports
Barros D, Alvares C, Alencar T, Baqueiro P, Marianno A, Alves R, Lenzi J, Rezende LF, Lordelo P
- 6846** Diverse microbiota in palatal radicular groove analyzed by Illumina sequencing: Four case reports
Tan XL, Chen X, Fu YJ, Ye L, Zhang L, Huang DM
- 6858** Autism with dysphasia accompanied by mental retardation caused by *FOXP1* exon deletion: A case report
Lin SZ, Zhou XY, Wang WQ, Jiang K
- 6867** *FGFR2-TSC22D1*, a novel *FGFR2* fusion gene identified in a patient with colorectal cancer: A case report
Kao XM, Zhu X, Zhang JL, Chen SQ, Fan CG
- 6872** Trismus originating from rare fungal myositis in pterygoid muscles: A case report
Bi L, Wei D, Wang B, He JF, Zhu HY, Wang HM
- 6879** Retroperitoneal laparoscopic partial nephrectomy for unilateral synchronous multifocal renal carcinoma with different pathological types: A case report
Xiao YM, Yang SK, Wang Y, Mao D, Duan FL, Zhou SK
- 6886** Diffuse large B cell lymphoma originating from the maxillary sinus with skin metastases: A case report and review of literature
Usuda D, Izumida T, Terada N, Sangen R, Higashikawa T, Sekiguchi S, Tanaka R, Suzuki M, Hotchi Y, Shimosawa S, Tokunaga S, Osugi I, Katou R, Ito S, Asako S, Takagi Y, Mishima K, Kondo A, Mizuno K, Takami H, Komatsu T, Oba J, Nomura T, Sugita M, Kasamaki Y
- 6900** Manifestation of acute peritonitis and pneumonedema in scrub typhus without eschar: A case report
Zhou XL, Ye QL, Chen JQ, Li W, Dong HJ
- 6907** Uterine tumor resembling an ovarian sex cord tumor: A case report and review of literature
Zhou FF, He YT, Li Y, Zhang M, Chen FH
- 6916** Dopamine agonist responsive burning mouth syndrome: Report of eight cases
Du QC, Ge YY, Xiao WL, Wang WF

- 6922** Complete withdrawal of glucocorticoids after dupilumab therapy in allergic bronchopulmonary aspergillosis: A case report
Nishimura T, Okano T, Naito M, Tsuji C, Iwanaka S, Sakakura Y, Yasuma T, Fujimoto H, D'Alessandro-Gabazza CN, Oomoto Y, Kobayashi T, Gabazza EC, Iбата H
- 6929** Sirolimus treatment for neonate with blue rubber bleb nevus syndrome: A case report
Yang SS, Yang M, Yue XJ, Tou JF
- 6935** Combined thoracoscopic and laparoscopic approach to remove a large retroperitoneal compound paraganglioma: A case report
Liu C, Wen J, Li HZ, Ji ZG
- 6943** Menetrier's disease and differential diagnosis: A case report
Wang HH, Zhao CC, Wang XL, Cheng ZN, Xie ZY
- 6950** Post-salpingectomy interstitial heterotopic pregnancy after *in vitro* fertilization and embryo transfer: A case report
Wang Q, Pan XL, Qi XR
- 6956** Ulnar nerve injury associated with displaced distal radius fracture: Two case reports
Yang JJ, Qu W, Wu YX, Jiang HJ

ABOUT COVER

Editorial Board Member of *World Journal of Clinical Cases*, Luigi Valentino Berra, MD, Assistant Professor, Neurosurgeon, Department of Neurosurgery, Policlinico Umberto I - Sapienza Università di Roma, Roma 00161, Italy. luigivbe@tin.it

AIMS AND SCOPE

The primary aim of *World Journal of Clinical Cases (WJCC, World J Clin Cases)* is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The *WJCC* is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Scopus, PubMed, and PubMed Central. The 2021 Edition of Journal Citation Reports® cites the 2020 impact factor (IF) for *WJCC* as 1.337; IF without journal self cites: 1.301; 5-year IF: 1.742; Journal Citation Indicator: 0.33; Ranking: 119 among 169 journals in medicine, general and internal; and Quartile category: Q3. The *WJCC*'s CiteScore for 2020 is 0.8 and Scopus CiteScore rank 2020: General Medicine is 493/793.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Jia-Hui Li*; Production Department Director: *Xiang Li*; Editorial Office Director: *Jin-Lei Wang*.

NAME OF JOURNAL

World Journal of Clinical Cases

ISSN

ISSN 2307-8960 (online)

LAUNCH DATE

April 16, 2013

FREQUENCY

Thrice Monthly

EDITORS-IN-CHIEF

Dennis A Bloomfield, Sandro Vento, Bao-Gan Peng

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2307-8960/editorialboard.htm>

PUBLICATION DATE

August 16, 2021

COPYRIGHT

© 2021 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Intestinal microbiota participates in nonalcoholic fatty liver disease progression by affecting intestinal homeostasis

Yang Zhang, Jun-Xiang Li, Yan Zhang, Yun-Liang Wang

ORCID number: Yang Zhang 0000-0002-2704-9846; Jun-Xiang Li 0000-0001-7590-9444; Yan Zhang 0000-0001-9543-6795; Yun-Liang Wang 0000-0002-0806-1399.

Author contributions: Zhang Y and Zhang Y drafted the manuscript; Li JX and Wang YL revised the manuscript for important intellectual content.

Supported by National Natural Science Foundation of China, No. 81503407; and Fundamental Research Funds for the Central Universities (Scientific Research Innovation Team), No. 2019-JYB-TD004.

Conflict-of-interest statement: The authors declare no conflicts of interest for this manuscript.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/License>

Yang Zhang, Jun-Xiang Li, Yan Zhang, Yun-Liang Wang, Department of Gastroenterology, Dong Fang Hospital, Beijing University of Chinese Medicine, Beijing 100078, China

Corresponding author: Yun-Liang Wang, MD, PhD, Associate Chief Physician, Doctor, Department of Gastroenterology, Dong Fang Hospital, Beijing University of Chinese Medicine, No. 6 District 1 Fangxingyuan, Beijing 100078, China. yunliang_wang@sina.com

Abstract

Nonalcoholic fatty liver disease (NAFLD) is a chronic liver disease with a pathogenesis that has not been fully elucidated. With the development of the theory of the gut-liver axis and the deepening of related research, the role of the intestinal tract in the pathogenesis of NAFLD has been investigated more. Intestinal microbiota, intestinal metabolites, and intestinal epithelial and immune-based barriers constitute the intestinal environment, which uses crosstalk to maintain the homeostasis of the intestinal environment. This paper reviews the progress in the study of intestinal microbiota, intestinal environment, and NAFLD and suggests that repair of intestinal functional balance may be a new idea for early prevention and intervention of NAFLD.

Key Words: Nonalcoholic fatty liver disease; Gut-liver axis; Intestinal microbiota; Metabolites; Intestinal homeostasis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Intestinal microbiota, intestinal metabolites, the intestinal epithelial barrier, and the immune barrier constitute the intestinal environment. The importance of intestinal homeostasis in the occurrence and development of nonalcoholic fatty liver disease (NAFLD) has been shown. Intestinal microbiota plays a leading role in maintaining the balance of the intestinal environment, but the complex interaction mechanism has not been elucidated fully. This article reviews the current research from the perspective of intestinal homeostasis and proposes that the repair of intestinal functional balance may be a new idea for early prevention and intervention of NAFLD.

Citation: Zhang Y, Li JX, Zhang Y, Wang YL. Intestinal microbiota participates in nonalcoholic fatty liver disease progression by affecting intestinal homeostasis. *World J Clin Cases* 2021;

[s/by-nc/4.0/](#)**Manuscript source:** Invited manuscript**Specialty type:** Medicine, research and experimental**Country/Territory of origin:** China**Peer-review report's scientific quality classification**Grade A (Excellent): 0
Grade B (Very good): 0
Grade C (Good): C, C
Grade D (Fair): 0
Grade E (Poor): 0**Received:** March 12, 2021**Peer-review started:** March 12, 2021**First decision:** April 17, 2021**Revised:** April 25, 2021**Accepted:** June 22, 2021**Article in press:** June 22, 2021**Published online:** August 16, 2021**P-Reviewer:** Jamali R, Zhang L**S-Editor:** Fan JR**L-Editor:** Wang TQ**P-Editor:** Li JH

9(23): 6654-6662

URL: <https://www.wjgnet.com/2307-8960/full/v9/i23/6654.htm>**DOI:** <https://dx.doi.org/10.12998/wjcc.v9.i23.6654>

INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is one of the most common liver diseases[1], characterized by metabolic stress causing liver damage. Risk factors include insulin resistance and genetic susceptibility[2]. The primary pathological change is diffuse bullae steatosis in hepatocytes, which is similar to alcoholic liver disease but without a history of heavy drinking. NAFLD has a multifactorial and complex pathogenesis and diverse clinical manifestations[3]. The classical “two-hit” theory is not enough to explain the complex pathogenesis of NAFLD[4]. The “gut-liver axis” theory, which hypothesizes a bidirectional relationship between the intestinal tract and the liver, has become a prominent theory to explain the key role of metabolic disorders in the pathogenesis of many liver diseases[5-7]. The relationship between intestinal homeostasis and the occurrence and development of NAFLD has also been gradually revealed due to this theory[8,9].

INTESTINAL ENVIRONMENT

The intestinal tract is an immune, metabolic, and nervous system organ, and its functional impairment plays an important role in the occurrence of many diseases, which can lead to death[10]. The intestinal tract is exposed to a variety of food components, antigens, symbiotic microorganisms, and pathogens. It is responsible for the digestion and absorption of nutrients and prevents harmful substances, such as pathogenic bacteria and toxins, from entering the circulation system through the intestinal tract[11]. Intestinal homeostasis constitutes a congenital protective barrier [12]. According to its composition, the intestinal barrier can be divided into an immune barrier, a mechanical barrier, a chemical barrier, and a biological barrier[13]. The immune barrier includes secretory immunoglobulin, lymphoid tissue, and mesenteric lymph node[14]. The mechanical barrier is composed of the mucus layer, intestinal epithelial cells, tight junctions between cells, and peristalsis function of the intestine. The chemical barrier is composed of gastric acid, bile, digestive enzymes, mucopolysaccharides, lysozyme, antimicrobial peptides, *etc.* The biological barrier refers to the microecosystem composed of the normal flora in the intestine. There are more than 4 billion microorganisms in the intestinal tract, and their metabolites can directly affect human health[15].

The composite barrier formed with multiple barriers is the key to keeping the dynamic balance in the intestinal environment[16]. The multiple barriers effectively resist intestinal bacterial translocation and endotoxin release and maintain normal bowel function, immune regulation, and body health[17]. Intestinal microbiota, metabolites, intestinal epithelium, and host immunity are important components of the intestinal environment. Under physiological conditions, all factors maintain a dynamic balance to ensure that the body is protected from harmful factors. Under pathological conditions, the imbalance of the intestinal environmental homeostasis leads to the destruction of the protective barrier, which causes the translocation of pathogenic microorganisms and toxic metabolites (an important cause of many diseases)[18].

INTESTINAL MICROBIOTA AND NAFLD

There are a variety of microbial communities in the human intestinal tract[19]. They function in a symbiotic relationship by promoting metabolism, digestion, and absorption[20] and by playing a role as a biological barrier. In both human and animal studies, the potential pathogenicity of intestinal microbial disorders in the development of NAFLD/nonalcoholic steatohepatitis (NASH) has been found[21-25]. However, due to the influence of multiple factors, such as age, region, diet, and obesity, no consistent results have been obtained on the characteristics of intestinal microbiota in patients with NAFLD/NASH[26].

A study of adult patients with NAFLD has shown that the occurrence of steatohepatitis is associated with an increase in the proportion of *Bacillus globulis* and a decrease in the proportion of *Bacteroides*[27]. In combination with 14C-D-xylose and lactulose breath test, researchers found bacterial overgrowth in the small intestine in approximately 50% of NASH patients, which was significantly higher than that in healthy controls[28]. Other studies found that gut microbiome samples from patients with NAFLD or NASH had a lower proportion of *Ruminococcaceae* than healthy subjects, and *Escherichia coli* was significantly disproportionate between obese children and children with NASH[29]. The proportion of *Clostridium globulae* in biopsy-confirmed adult NASH patients was significantly higher than that in NAFLD patients[27]. Obesity and insulin resistance are risk factors for NAFLD[30]. It has been confirmed by experiments that sterile mice can avoid obesity caused by a high-fat diet, indicating that the obesity caused by a high-fat diet, to some extent, requires the involvement of intestinal microorganisms[31].

The mechanism of action between intestinal microbiota and hosts is complex. The disorder of intestinal microbiota affects the levels of metabolites, such as ethanol, choline, and endotoxin, in the NAFLD inflammation progress[32-34] and damages the function of the intestinal barrier by increasing the permeability. This leads to exposure of the liver to endotoxin, which further promotes fatty liver inflammation and enhances fibrosis in NASH development[35]. Intestinal microbiota participates in the process of NAFLD, which is closely related to its influence on the intestinal environment.

INTESTINAL METABOLITES

Dietary substances enter the intestinal tract under the action of intestinal microbiota after digestion to produce a large number of metabolites, such as secondary bile acids, short-chain fatty acids, ethanol, choline, and endotoxin[36]. The metabolites that cannot be reabsorbed can be excreted through feces. If the production of metabolites exceeds the load capacity that the body is adapted to, it will cause metabolic disorders [37]. At present, the effects of bile acids and fatty acids have the most data in relation to NAFLD[38].

Bile acids

Bile acids are an important component of bile, which regulate lipid digestion and absorption and cholesterol metabolism. They also play a key role in insulin sensitivity and metabolic homeostasis[39]. Primary bile acids synthesized in the liver are transformed into secondary bile acids under the action of intestinal flora after entering the intestine, and their level is affected by the composition, abundance, and proportion of intestinal flora[40]. Bile acids can also change the structure of the intestinal flora [41]. By promoting the activation of the farnesoid X receptor, intestinal microorganisms can further induce the expression of fibroblast growth factor 15, inhibit CYP7YPA1, and affect bile acid synthesis. In addition, the intestinal flora interferes with secondary bile acid reabsorption by affecting the transport of normal tissue complication probability and automated blood sampling telemetry[42].

Studies have found that serum bile acid levels in patients with NAFLD are significantly increased[43], and the severity of steatosis is positively correlated with the level of bile acid synthesis[44]. Increased levels of key bile acids, such as taurocholic acid and glycocholic acid, were associated with steatosis, lobular and portal vein inflammation, and hepatocyte ballooning. In addition, the risk of NAFLD progression to fibrosis was positively correlated with the ratio of secondary bile acids to primary bile acids and the concentration of bound bile acids[45]. Bile acid may also damage bacterial membranes by binding phospholipids on the bacterial membrane, thus playing an antibacterial role in adhesion and neutralizing of endotoxins[46-48]. Mice fed food rich in saturated fatty acids promoted changes in the composition of bile acids, which showed significant dysregulation of intestinal microorganisms[49]. These results suggest that bile acid metabolism in the intestine is involved in the process of NAFLD, and there is an interaction between bile acids and intestinal microorganisms. The complex mechanism behind this interaction remains to be further elucidated.

Fatty acid metabolism

Abnormal lipid metabolism is a key factor in the occurrence and development of NAFLD[50]. Fatty acid deposition in liver cells and the resulting insulin resistance, lipotoxic injury, oxidative stress, apoptosis, and inflammation are important

pathological mechanisms of the progression of NAFLD[51]. Fatty acids can be derived from glucose synthesis absorbed by hepatocytes, from peripheral adipose tissue release, and from intestinal metabolism into blood[52]. Under the action of intestinal microorganisms, fat is hydrolyzed and emulsified in the upper part of the small intestine to produce fatty acids. Long-chain fatty acids are absorbed by intestinal epithelial mucosal cells, transformed into triglycerides, and combined with apolipoproteins to form chylous particles that are released into circulation. Medium-chain and short-chain fatty acids enter circulation directly through the portal system. The free fatty acid levels in blood increase, and excessive intake of the fatty acids by hepatocytes leads to NAFLD[53].

The incidence of NAFLD is correlated with short-chain fatty acid levels. Short-chain fatty acids are produced by gut microbes through the glycolysis of carbohydrates. The most prevalent short-chain fatty acids are acetic acid, propionic acid, and butyric acid [54]. Acetic acid participates in body energy metabolism through acetyl-coA, propionic acid inhibits cholesterol synthesis in the liver, and butyric acid can protect intestinal mucosa barrier permeability[55,56].

Fatty acid metabolism in the intestine is affected by intestinal microbiota and regulates the intestinal barrier, which plays a complex role in the progression of NAFLD[57,58]. When the liver is overloaded with free fatty acids, a large amount of reactive oxygen species will be produced, causing oxidative stress and mitochondrial damage. A clinical study[59] related to short-chain fatty acids found that the concentration of propionate/acetate in feces of patients with NAFLD was positively correlated with the ratio of Th17/Treg cells in peripheral blood, but negatively correlated with Tregs in peripheral blood. Th17/Treg dynamic balance is an important factor in maintaining immune homeostasis, Th17 plays an inflammatory regulation role, while Tregs plays an immune tolerance role[60], suggesting that short-chain fatty acids may participate in the progression of NAFLD by affecting T cell immune function.

INTESTINAL MICROBIOTA AND INTESTINAL BARRIER INJURY IN NAFLD

The intestinal tract, as an important organ for nutrient absorption, forms an effective barrier network to avoid harmful substance intake. Normal intestinal barrier function is the key to maintaining homeostasis of the intestinal environment. The composition of the different barriers varies but they are closely related to each other. Intestinal biological barriers and chemical barriers have been described. Intestinal microorganisms stimulate intestinal epithelial cells to secrete a variety of immune mediators, including cytokines and chemokines, that play an important role in maintaining the integrity of the epithelial barrier, shaping the mucosal immune system, and regulating the host immune response[61]. In order to avoid an abnormal excessive immune response, intestinal epithelial cells isolate intestinal microorganisms from immune cells through chemical and mechanical barriers, thereby establishing host-symbiotic relationships. Intestinal immune cells are also involved in maintaining a healthy intestinal microbial community and enhancing epithelial barrier function[15]. Intestinal barrier damage has been confirmed in NAFLD, and the intestinal flora is a participant in that process[62,63].

Intestinal epithelial barrier

The intestinal epithelial barrier is the main component of the mechanical barrier. It includes columnar epithelial cells, Pan's cells, goblet cells, intestinal endocrine cells, cluster cells, and other cell populations, which play a crucial role in the stability of the intestinal environment[64]. The integrity and regenerative capacity of the intestinal epithelium are the structural basis. Intestinal bacterial metabolites, such as short-chain fatty acids and secondary bile acids, play an important role in maintaining intestinal epithelial integrity.

Short-chain fatty acids are an energy source for the host and regulate the physiological function of intestinal epithelial cells. Short-chain fatty acids can regulate intestinal adaptability and promote proliferation through activation of G-protein-coupled receptors (GPR41, GPR43, and GPR109A)[65]. Other studies have shown that short-chain fatty acids can activate the nuclear factor- κ B signaling pathway through Toll-like receptors (TLRs), which regulates the integrity of intestinal epithelial cells [66]. Short-chain fatty acids can also activate the inflammatory body NOD-like receptor family pyrin domain containing 3 (NLRP3), upregulate interleukin (IL)-18, and maintain intestinal dynamic balance[67].

Primary bile acids secreted by the liver enter the small intestine and promote the digestion and absorption of fat in the small intestine. Under the action of intestinal microorganisms, most of the transformed secondary bile acids are reabsorbed by intestinal epithelial cells and return to the liver through the portal vein. Studies have shown that secondary bile acids also regulate the proliferation of intestinal epithelial cells. For example, cholic acid induces the proliferation of intestinal epithelial cells by activating Src, epidermal growth factor receptor, and extracellular signal-regulated kinase, while deoxycholic acid inhibits the proliferation of intestinal epithelial cells by activating the farnesoid X receptor[68].

The junction of the intestinal epithelium depends on tight junction proteins, including occludin, the claudin family proteins, and zona occludens 1. The imbalance of intestinal flora in patients with NASH and excessive growth and reproduction of intestinal Gram-negative bacilli, such as *Enterobacteriaceae*, causes tight junction proteins to contract and move to the cytoplasm under the pathological conditions of hypoxia and inflammatory stimulation. This increases intestinal mucosal permeability, epithelial barrier destruction, and intestinal bacterial translocation. In addition, the metabolite lipopolysaccharide is released into the blood and liver through the portal system, promoting the release of inflammatory factors and resulting in NAFLD[69]. Studies have found that the application of intestinal microbiota regulator probiotics or synbiotics can reduce hepatic steatosis and inflammation in NAFLD[70], which may be related to the improvement of intestinal permeability and regulation of intestinal microecological balance[71].

Intestinal immune barrier

The intestinal immune barrier plays an important role in the pathogenesis of NAFLD. The intestinal mucosal immune barrier is composed of immune cells, intestinal mucosal lymphoid tissues, and immunoglobulin A (secreted by the immune cells)[72], which plays an important role in protecting the body from pathogenic bacteria and pathogens. Immune cells include T cells, B cells, dendritic cells, macrophages, eosinophils, and mast cells that are distributed in different tissue layers of the intestinal mucosa and have an inherent immune effect. These cells play a key role in maintaining immune homeostasis by inhibiting responses to harmless antigens and enhancing the integrity of the intestinal mucosal barrier function[73].

The intestinal immune function acts as a sentinel to prevent harmful substances from entering the portal system. Then, the liver can monitor enteric metabolites and pathogens and induce an immune response from harmful substances entering the liver *via* the portal vein. The function of the intestinal immune barrier is related to pattern recognition receptors, such as TLRs and nucleotide binding oligomerization domain-like receptors[74]. The intestinal epithelium can express TLRs, recognize intestinal metabolites, and further participate in the inflammatory progression of NAFLD by activating MYD88. Mice with intestinal epithelial cell-specific *MYD88* gene deletion fed a high-fat diet showed improved oral glucose tolerance and associated hepatic steatosis and triglyceride content, in contrast to wild-type mice[75]. In addition, lipopolysaccharide produced by the flora can activate NLRP3 inflammatory bodies through TLR4 and TLR9, activate caspase-1, and cleave IL-1 β and IL-18 precursors into activated IL-1 β and IL-18, which promotes the progression of NAFLD inflammation and fibrosis[76].

Intestinal regulatory T cells play a key role in inhibiting the immune response induced by symbiotic microorganisms and metabolites. Intestinal short-chain fatty acids can induce epithelial transforming growth factor- β production and promote regulatory T cell production to participate in the immune process[77]. Therefore, the intestinal immune system response is a key link in the occurrence and development of NAFLD.

CONCLUSION

Intestinal microbiota, intestinal metabolites, and intestinal epithelial and immune-based barriers constitute the intestinal environment. These components are all dependent on the others, requiring crosstalk to maintain the homeostasis of the intestinal environment. Current studies have shown that intestinal flora play a leading role in maintaining the balance of the intestinal environment. The balance of intestinal environmental factors is the key to maintaining a healthy state, and the importance of homeostasis imbalance in the occurrence and development of NAFLD has been shown. Although some progress has been made in all aspects of research, the complex

interaction mechanisms have not been fully elucidated. Most of the previous studies highlight a single factor. The research on the relationship between the factors must be expanded. The balance of multiple factors likely involves a variety of complex mechanisms, which also presents difficulties in the research efforts.

At present, there is no specific therapeutic drug for NAFLD. The discovery of the “gut-liver axis” and the importance of the intestinal microbiota in the pathogenesis of NAFLD suggest that focusing on the repair of intestinal functional balance may be a new drug target for prevention and intervention of NAFLD. In the future, with the further development of more studies, major mechanisms will be revealed, which will bring new treatment options for NAFLD.

REFERENCES

- 1 **Sheka AC**, Adeyi O, Thompson J, Hameed B, Crawford PA, Ikramuddin S. Nonalcoholic Steatohepatitis: A Review. *JAMA* 2020; **323**: 1175-1183 [PMID: [32207804](#) DOI: [10.1001/jama.2020.2298](#)]
- 2 **Wu Y**, Zheng Q, Zou B, Yeo YH, Li X, Li J, Xie X, Feng Y, Stave CD, Zhu Q, Cheung R, Nguyen MH. The epidemiology of NAFLD in Mainland China with analysis by adjusted gross regional domestic product: a meta-analysis. *Hepatol Int* 2020; **14**: 259-269 [PMID: [32130675](#) DOI: [10.1007/s12072-020-10023-3](#)]
- 3 **Lonardo A**, Nascimbeni F, Maurantonio M, Marrazzo A, Rinaldi L, Adinolfi LE. Nonalcoholic fatty liver disease: Evolving paradigms. *World J Gastroenterol* 2017; **23**: 6571-6592 [PMID: [29085206](#) DOI: [10.3748/wjg.v23.i36.6571](#)]
- 4 **Bessone F**, Razori MV, Roma MG. Molecular pathways of nonalcoholic fatty liver disease development and progression. *Cell Mol Life Sci* 2019; **76**: 99-128 [PMID: [30343320](#) DOI: [10.1007/s00018-018-2947-0](#)]
- 5 **Szabo G**. Gut-liver axis in alcoholic liver disease. *Gastroenterology* 2015; **148**: 30-36 [PMID: [25447847](#) DOI: [10.1053/j.gastro.2014.10.042](#)]
- 6 **Albillos A**, de Gottardi A, Rescigno M. The gut-liver axis in liver disease: Pathophysiological basis for therapy. *J Hepatol* 2020; **72**: 558-577 [PMID: [31622696](#) DOI: [10.1016/j.jhep.2019.10.003](#)]
- 7 **Wiest R**, Albillos A, Trauner M, Bajaj JS, Jalan R. Targeting the gut-liver axis in liver disease. *J Hepatol* 2017; **67**: 1084-1103 [PMID: [28526488](#) DOI: [10.1016/j.jhep.2017.05.007](#)]
- 8 **Tripathi A**, Debelius J, Brenner DA, Karin M, Loomba R, Schnabl B, Knight R. The gut-liver axis and the intersection with the microbiome. *Nat Rev Gastroenterol Hepatol* 2018; **15**: 397-411 [PMID: [29748586](#) DOI: [10.1038/s41575-018-0011-z](#)]
- 9 **Kolodziejczyk AA**, Zheng D, Shibolet O, Elinav E. The role of the microbiome in NAFLD and NASH. *EMBO Mol Med* 2019; **11**: e9302 [PMID: [30591521](#) DOI: [10.15252/emmm.201809302](#)]
- 10 **Druml W**. [Intestinal cross-talk : The gut as motor of multiple organ failure]. *Med Klin Intensivmed Notfmed* 2018; **113**: 470-477 [PMID: [30120495](#) DOI: [10.1007/s00063-018-0475-1](#)]
- 11 **Fabbiano S**, Suárez-Zamorano N, Chevalier C, Lazarević V, Kieser S, Rigo D, Leo S, Veyrat-Durebex C, Gaia N, Maresca M, Merkler D, Gomez de Agüero M, Macpherson A, Schrenzel J, Trajkovski M. Functional Gut Microbiota Remodeling Contributes to the Caloric Restriction-Induced Metabolic Improvements. *Cell Metab* 2018; **28**: 907-921.e7 [PMID: [30174308](#) DOI: [10.1016/j.cmet.2018.08.005](#)]
- 12 **Chopyk DM**, Grakoui A. Contribution of the Intestinal Microbiome and Gut Barrier to Hepatic Disorders. *Gastroenterology* 2020; **159**: 849-863 [PMID: [32569766](#) DOI: [10.1053/j.gastro.2020.04.077](#)]
- 13 **Vancamelbeke M**, Vermeire S. The intestinal barrier: a fundamental role in health and disease. *Expert Rev Gastroenterol Hepatol* 2017; **11**: 821-834 [PMID: [28650209](#) DOI: [10.1080/17474124.2017.1343143](#)]
- 14 **Weström B**, Arévalo Sureda E, Pierzynowska K, Pierzynowski SG, Pérez-Cano FJ. The Immature Gut Barrier and Its Importance in Establishing Immunity in Newborn Mammals. *Front Immunol* 2020; **11**: 1153 [PMID: [32582216](#) DOI: [10.3389/fimmu.2020.01153](#)]
- 15 **Kayama H**, Okumura R, Takeda K. Interaction Between the Microbiota, Epithelia, and Immune Cells in the Intestine. *Annu Rev Immunol* 2020; **38**: 23-48 [PMID: [32340570](#) DOI: [10.1146/annurev-immunol-070119-115104](#)]
- 16 **Wells JM**, Brummer RJ, Derrien M, MacDonald TT, Troost F, Cani PD, Theodorou V, Dekker J, Méheust A, de Vos WM, Mercenier A, Nauta A, Garcia-Rodenas CL. Homeostasis of the gut barrier and potential biomarkers. *Am J Physiol Gastrointest Liver Physiol* 2017; **312**: G171-G193 [PMID: [27908847](#) DOI: [10.1152/ajpgi.00048.2015](#)]
- 17 **Julio-Pieper M**, Bravo JA. Intestinal Barrier and Behavior. *Int Rev Neurobiol* 2016; **131**: 127-141 [PMID: [27793215](#) DOI: [10.1016/bs.irm.2016.08.006](#)]
- 18 **Salvo Romero E**, Alonso Cotoner C, Pardo Camacho C, Casado Bedmar M, Vicario M. The intestinal barrier function and its involvement in digestive disease. *Rev Esp Enferm Dig* 2015; **107**: 686-696 [PMID: [26541659](#) DOI: [10.17235/reed.2015.3846/2015](#)]
- 19 **Thursby E**, Juge N. Introduction to the human gut microbiota. *Biochem J* 2017; **474**: 1823-1836 [PMID: [28512250](#) DOI: [10.1042/BCJ20160510](#)]

- 20 **Zmora N**, Suez J, Elinav E. You are what you eat: diet, health and the gut microbiota. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 35-56 [PMID: [30262901](#) DOI: [10.1038/s41575-018-0061-2](#)]
- 21 **Le Roy T**, Llopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, Martin P, Philippe C, Walker F, Bado A, Perlemuter G, Cassard-Doulier AM, Gérard P. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. *Gut* 2013; **62**: 1787-1794 [PMID: [23197411](#) DOI: [10.1136/gutjnl-2012-303816](#)]
- 22 **Loomba R**, Wolfson T, Ang B, Hooker J, Behling C, Peterson M, Valasek M, Lin G, Brenner D, Gamst A, Ehman R, Sirlin C. Magnetic resonance elastography predicts advanced fibrosis in patients with nonalcoholic fatty liver disease: a prospective study. *Hepatology* 2014; **60**: 1920-1928 [PMID: [25103310](#) DOI: [10.1002/hep.27362](#)]
- 23 **Martínez-Castillo M**, Rosique-Oramas D, Medina-Avila Z, Pérez-Hernández JL, Higuera-De la Tijera F, Santana-Vargas D, Montalvo-Jave EE, Sanchez-Avila F, Torre A, Kershenovich D, Gutierrez-Reyes G. Differential production of insulin-like growth factor-binding proteins in liver fibrosis progression. *Mol Cell Biochem* 2020; **469**: 65-75 [PMID: [32301061](#) DOI: [10.1007/s11010-020-03728-4](#)]
- 24 **Ji Y**, Yin Y, Li Z, Zhang W. Gut Microbiota-Derived Components and Metabolites in the Progression of Non-Alcoholic Fatty Liver Disease (NAFLD). *Nutrients* 2019; **11**: 1712 [PMID: [31349604](#) DOI: [10.3390/nu11081712](#)]
- 25 **Abdou RM**, Zhu L, Baker RD, Baker SS. Gut Microbiota of Nonalcoholic Fatty Liver Disease. *Dig Dis Sci* 2016; **61**: 1268-1281 [PMID: [26898658](#) DOI: [10.1007/s10620-016-4045-1](#)]
- 26 **Aron-Wisnewsky J**, Vigiotti C, Witjes J, Le P, Holleboom AG, Verheij J, Nieuwdorp M, Clément K. Gut microbiota and human NAFLD: disentangling microbial signatures from metabolic disorders. *Nat Rev Gastroenterol Hepatol* 2020; **17**: 279-297 [PMID: [32152478](#) DOI: [10.1038/s41575-020-0269-9](#)]
- 27 **Mouzaki M**, Comelli EM, Arendt BM, Bonengel J, Fung SK, Fischer SE, McGilvray ID, Allard JP. Intestinal microbiota in patients with nonalcoholic fatty liver disease. *Hepatology* 2013; **58**: 120-127 [PMID: [23401313](#) DOI: [10.1002/hep.26319](#)]
- 28 **Wigg AJ**, Roberts-Thomson IC, Dymock RB, McCarthy PJ, Grose RH, Cummins AG. The role of small intestinal bacterial overgrowth, intestinal permeability, endotoxaemia, and tumour necrosis factor alpha in the pathogenesis of non-alcoholic steatohepatitis. *Gut* 2001; **48**: 206-211 [PMID: [11156641](#) DOI: [10.1136/gut.48.2.206](#)]
- 29 **Zhu L**, Baker SS, Gill C, Liu W, Alkhoury R, Baker RD, Gill SR. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH. *Hepatology* 2013; **57**: 601-609 [PMID: [23055155](#) DOI: [10.1002/hep.26093](#)]
- 30 **Turnbaugh PJ**, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; **444**: 1027-1031 [PMID: [17183312](#) DOI: [10.1038/nature05414](#)]
- 31 **Bäckhed F**, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci USA* 2007; **104**: 979-984 [PMID: [17210919](#) DOI: [10.1073/pnas.0605374104](#)]
- 32 **Cope K**, Risby T, Diehl AM. Increased gastrointestinal ethanol production in obese mice: implications for fatty liver disease pathogenesis. *Gastroenterology* 2000; **119**: 1340-1347 [PMID: [11054393](#) DOI: [10.1053/gast.2000.19267](#)]
- 33 **Blumberg H**, McCollum EV. The prevention by choline of liver cirrhosis in rats on high fat, low protein diets. *Science* 1941; **93**: 598-599 [PMID: [17795968](#) DOI: [10.1126/science.93.2425.598](#)]
- 34 **Alisi A**, Manco M, Devito R, Piemonte F, Nobili V. Endotoxin and plasminogen activator inhibitor-1 serum levels associated with nonalcoholic steatohepatitis in children. *J Pediatr Gastroenterol Nutr* 2010; **50**: 645-649 [PMID: [20400911](#) DOI: [10.1097/MPG.0b013e3181c7bdf1](#)]
- 35 **Jadhav K**, Cohen TS. Can You Trust Your Gut? *Front Endocrinol (Lausanne)* 2020; **11**: 592157 [PMID: [33193105](#) DOI: [10.3389/fendo.2020.592157](#)]
- 36 **Venetsanaki V**, Karabouta Z, Polyzos SA. Farnesoid X nuclear receptor agonists for the treatment of nonalcoholic steatohepatitis. *Eur J Pharmacol* 2019; **863**: 172661 [PMID: [31536725](#) DOI: [10.1016/j.ejphar.2019.172661](#)]
- 37 **Miyachi S**, Hirasawa A, Ichimura A, Hara T, Tsujimoto G. New frontiers in gut nutrient sensor research: free fatty acid sensing in the gastrointestinal tract. *J Pharmacol Sci* 2010; **112**: 19-24 [PMID: [20093784](#) DOI: [10.1254/jphs.09r09fm](#)]
- 38 **Wieland A**, Frank DN, Harnke B, Bambha K. Systematic review: microbial dysbiosis and nonalcoholic fatty liver disease. *Aliment Pharmacol Ther* 2015; **42**: 1051-1063 [PMID: [26304302](#) DOI: [10.1111/apt.13376](#)]
- 39 **Dawson PA**, Karpen SJ. Intestinal transport and metabolism of bile acids. *J Lipid Res* 2015; **56**: 1085-1099 [PMID: [25210150](#) DOI: [10.1194/jlr.R054114](#)]
- 40 **Gonzalez FJ**, Jiang C, Patterson AD. An Intestinal Microbiota-Farnesoid X Receptor Axis Modulates Metabolic Disease. *Gastroenterology* 2016; **151**: 845-859 [PMID: [27639801](#) DOI: [10.1053/j.gastro.2016.08.057](#)]
- 41 **Wahlström A**, Kovatcheva-Datchary P, Ståhlman M, Bäckhed F, Marschall HU. Crosstalk between Bile Acids and Gut Microbiota and Its Impact on Farnesoid X Receptor Signalling. *Dig Dis* 2017; **35**: 246-250 [PMID: [28249261](#) DOI: [10.1159/000450982](#)]
- 42 **Wahlström A**, Sayin SI, Marschall HU, Bäckhed F. Intestinal Crosstalk between Bile Acids and Microbiota and Its Impact on Host Metabolism. *Cell Metab* 2016; **24**: 41-50 [PMID: [27320064](#) DOI: [10.1016/j.cmet.2016.05.002](#)]

- 10.1016/j.cmet.2016.05.005]
- 43 **Yang ZX**, Shen W, Sun H. Effects of nuclear receptor FXR on the regulation of liver lipid metabolism in patients with non-alcoholic fatty liver disease. *Hepatol Int* 2010; **4**: 741-748 [PMID: 21286345 DOI: 10.1007/s12072-010-9202-6]
 - 44 **Bechmann LP**, Kocabayoglu P, Sowa JP, Sydor S, Best J, Schlattjan M, Beilfuss A, Schmitt J, Hannivoort RA, Kilicarslan A, Rust C, Berr F, Tschopp O, Gerken G, Friedman SL, Geier A, Canbay A. Free fatty acids repress small heterodimer partner (SHP) activation and adiponectin counteracts bile acid-induced liver injury in superobese patients with nonalcoholic steatohepatitis. *Hepatology* 2013; **57**: 1394-1406 [PMID: 23299969 DOI: 10.1002/hep.26225]
 - 45 **Puri P**, Daita K, Joyce A, Mirshahi F, Santhekadur PK, Cazanave S, Luketic VA, Siddiqui MS, Boyett S, Min HK, Kumar DP, Kohli R, Zhou H, Hylemon PB, Contos MJ, Idowu M, Sanyal AJ. The presence and severity of nonalcoholic steatohepatitis is associated with specific changes in circulating bile acids. *Hepatology* 2018; **67**: 534-548 [PMID: 28696585 DOI: 10.1002/hep.29359]
 - 46 **Ridlon JM**, Kang DJ, Hylemon PB, Bajaj JS. Bile acids and the gut microbiome. *Curr Opin Gastroenterol* 2014; **30**: 332-338 [PMID: 24625896 DOI: 10.1097/MOG.0000000000000057]
 - 47 **Jüngst C**, Berg T, Cheng J, Green RM, Jia J, Mason AL, Lammert F. Intrahepatic cholestasis in common chronic liver diseases. *Eur J Clin Invest* 2013; **43**: 1069-1083 [PMID: 23927644 DOI: 10.1111/eci.12128]
 - 48 **Fouts DE**, Torralba M, Nelson KE, Brenner DA, Schnabl B. Bacterial translocation and changes in the intestinal microbiome in mouse models of liver disease. *J Hepatol* 2012; **56**: 1283-1292 [PMID: 22326468 DOI: 10.1016/j.jhep.2012.01.019]
 - 49 **Devkota S**, Wang Y, Musch MW, Leone V, Fehlner-Peach H, Nadimpalli A, Antonopoulos DA, Jabri B, Chang EB. Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in IL10^{-/-} mice. *Nature* 2012; **487**: 104-108 [PMID: 22722865 DOI: 10.1038/nature11225]
 - 50 **Dongiovanni P**, Anstee QM, Valenti L. Genetic predisposition in NAFLD and NASH: impact on severity of liver disease and response to treatment. *Curr Pharm Des* 2013; **19**: 5219-5238 [PMID: 23394097 DOI: 10.2174/13816128113199990381]
 - 51 **Legrand-Poels S**, Esser N, L'homme L, Scheen A, Paquot N, Piette J. Free fatty acids as modulators of the NLRP3 inflammasome in obesity/type 2 diabetes. *Biochem Pharmacol* 2014; **92**: 131-141 [PMID: 25175736 DOI: 10.1016/j.bcp.2014.08.013]
 - 52 **Perry RJ**, Samuel VT, Petersen KF, Shulman GL. The role of hepatic lipids in hepatic insulin resistance and type 2 diabetes. *Nature* 2014; **510**: 84-91 [PMID: 24899308 DOI: 10.1038/nature13478]
 - 53 **Neuschwander-Tetri BA**. Hepatic lipotoxicity and the pathogenesis of nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. *Hepatology* 2010; **52**: 774-788 [PMID: 20683968 DOI: 10.1002/hep.23719]
 - 54 **Ríos-Covián D**, Ruas-Madiedo P, Margolles A, Gueimonde M, de Los Reyes-Gavilán CG, Salazar N. Intestinal Short Chain Fatty Acids and their Link with Diet and Human Health. *Front Microbiol* 2016; **7**: 185 [PMID: 26925050 DOI: 10.3389/fmicb.2016.00185]
 - 55 **Markowiak-Kopeć P**, Śliżewska K. The Effect of Probiotics on the Production of Short-Chain Fatty Acids by Human Intestinal Microbiome. *Nutrients* 2020; **12**: 1107 [PMID: 32316181 DOI: 10.3390/nu12041107]
 - 56 **Nicholson JK**, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, Pettersson S. Host-gut microbiota metabolic interactions. *Science* 2012; **336**: 1262-1267 [PMID: 22674330 DOI: 10.1126/science.1223813]
 - 57 **Walker AW**, Ince J, Duncan SH, Webster LM, Holtrop G, Ze X, Brown D, Stares MD, Scott P, Bergerat A, Louis P, McIntosh F, Johnstone AM, Lobley GE, Parkhill J, Flint HJ. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *ISME J* 2011; **5**: 220-230 [PMID: 20686513 DOI: 10.1038/ismej.2010.118]
 - 58 **Woods CP**, Hazlehurst JM, Tomlinson JW. Glucocorticoids and non-alcoholic fatty liver disease. *J Steroid Biochem Mol Biol* 2015; **154**: 94-103 [PMID: 26241028 DOI: 10.1016/j.jsbmb.2015.07.020]
 - 59 **Rau M**, Rehman A, Dittrich M, Groen AK, Hermanns HM, Seyfried F, Beyersdorf N, Dandekar T, Rosenstiel P, Geier A. Fecal SCFAs and SCFA-producing bacteria in gut microbiome of human NAFLD as a putative link to systemic T-cell activation and advanced disease. *United European Gastroenterol J* 2018; **6**: 1496-1507 [PMID: 30574320 DOI: 10.1177/2050640618804444]
 - 60 **Newton R**, Priyadarshini B, Turka LA. Immunometabolism of regulatory T cells. *Nat Immunol* 2016; **17**: 618-625 [PMID: 27196520 DOI: 10.1038/ni.3466]
 - 61 **Okumura R**, Takeda K. Roles of intestinal epithelial cells in the maintenance of gut homeostasis. *Exp Mol Med* 2017; **49**: e338 [PMID: 28546564 DOI: 10.1038/emmm.2017.20]
 - 62 **Milosevic I**, Vujovic A, Barac A, Djelic M, Korac M, Radovanovic Spurnic A, Gmizic I, Stevanovic O, Djordjevic V, Lekic N, Russo E, Amedei A. Gut-Liver Axis, Gut Microbiota, and Its Modulation in the Management of Liver Diseases: A Review of the Literature. *Int J Mol Sci* 2019; **20**: 395 [PMID: 30658519 DOI: 10.3390/ijms20020395]
 - 63 **Acharya C**, Sahingur SE, Bajaj JS. Microbiota, cirrhosis, and the emerging oral-gut-liver axis. *JCI Insight* 2017; **2**: e94416 [PMID: 28978799 DOI: 10.1172/jci.insight.94416]
 - 64 **Kurashima Y**, Kiyono H. Mucosal Ecological Network of Epithelium and Immune Cells for Gut Homeostasis and Tissue Healing. *Annu Rev Immunol* 2017; **35**: 119-147 [PMID: 28125357 DOI: 10.1146/annurev-immunol-051116-052424]
 - 65 **Marchix J**, Goddard G, Helmuth MA. Host-Gut Microbiota Cross-Talk in Intestinal Adaptation.

- Cellu Molecul Gastroenterol* 2018; 1-13 [DOI: [10.1016/j.jcmgh.2018.01.024](https://doi.org/10.1016/j.jcmgh.2018.01.024)]
- 66 **Sun M**, Wu W, Liu Z, Cong Y. Microbiota metabolite short chain fatty acids, GPCR, and inflammatory bowel diseases. *J Gastroenterol* 2017; **52**: 1-8 [PMID: [27448578](https://pubmed.ncbi.nlm.nih.gov/27448578/) DOI: [10.1007/s00535-016-1242-9](https://doi.org/10.1007/s00535-016-1242-9)]
- 67 **Macia L**, Tan J, Vieira AT, Leach K, Stanley D, Luong S, Maruya M, Ian McKenzie C, Hijikata A, Wong C, Binge L, Thorburn AN, Chevalier N, Ang C, Marino E, Robert R, Offermanns S, Teixeira MM, Moore RJ, Flavell RA, Fagarasan S, Mackay CR. Metabolite-sensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nat Commun* 2015; **6**: 6734 [PMID: [25828455](https://pubmed.ncbi.nlm.nih.gov/25828455/) DOI: [10.1038/ncomms7734](https://doi.org/10.1038/ncomms7734)]
- 68 **Mroz MS**, Lajczak NK, Goggins BJ, Keely S, Keely SJ. The bile acids, deoxycholic acid and ursodeoxycholic acid, regulate colonic epithelial wound healing. *Am J Physiol Gastrointest Liver Physiol* 2018; **314**: G378-G387 [PMID: [29351391](https://pubmed.ncbi.nlm.nih.gov/29351391/) DOI: [10.1152/ajpgi.00435.2016](https://doi.org/10.1152/ajpgi.00435.2016)]
- 69 **Cui Y**, Wang Q, Chang R, Zhou X, Xu C. Intestinal Barrier Function-Non-alcoholic Fatty Liver Disease Interactions and Possible Role of Gut Microbiota. *J Agric Food Chem* 2019; **67**: 2754-2762 [PMID: [30798598](https://pubmed.ncbi.nlm.nih.gov/30798598/) DOI: [10.1021/acs.jafc.9b00080](https://doi.org/10.1021/acs.jafc.9b00080)]
- 70 **Kobyliak N**, Abenavoli L, Falalyeyeva T, Mykhalchyshyn G, Boccuto L, Kononenko L, Kyriienko D, Komisarenko I, Dynnyk O. Beneficial effects of probiotic combination with omega-3 fatty acids in NAFLD: a randomized clinical study. *Minerva Med* 2018; **109**: 418-428 [PMID: [30221912](https://pubmed.ncbi.nlm.nih.gov/30221912/) DOI: [10.23736/S0026-4806.18.05845-7](https://doi.org/10.23736/S0026-4806.18.05845-7)]
- 71 **de Roos NM**, van Hemert S, Rovers JMP, Smits MG, Witterman BJM. The effects of a multispecies probiotic on migraine and markers of intestinal permeability-results of a randomized placebo-controlled study. *Eur J Clin Nutr* 2017; **71**: 1455-1462 [PMID: [28537581](https://pubmed.ncbi.nlm.nih.gov/28537581/) DOI: [10.1038/ejcn.2017.57](https://doi.org/10.1038/ejcn.2017.57)]
- 72 **Bieghs V**, Trautwein C. Innate immune signaling and gut-liver interactions in non-alcoholic fatty liver disease. *Hepatobiliary Surg Nutr* 2014; **3**: 377-385 [PMID: [25568861](https://pubmed.ncbi.nlm.nih.gov/25568861/) DOI: [10.3978/j.issn.2304-3881.2014.12.04](https://doi.org/10.3978/j.issn.2304-3881.2014.12.04)]
- 73 **Honda K**, Littman DR. The microbiota in adaptive immune homeostasis and disease. *Nature* 2016; **535**: 75-84 [PMID: [27383982](https://pubmed.ncbi.nlm.nih.gov/27383982/) DOI: [10.1038/nature18848](https://doi.org/10.1038/nature18848)]
- 74 **Huang Y**, Chen Z. Inflammatory bowel disease related innate immunity and adaptive immunity. *Am J Transl Res* 2016; **8**: 2490-2497 [PMID: [27398134](https://pubmed.ncbi.nlm.nih.gov/27398134/)]
- 75 **Everard A**, Geurts L, Caesar R, Van Hul M, Matamoros S, Duparc T, Denis RG, Cochez P, Pierard F, Castel J, Bindels LB, Plovier H, Robine S, Muccioli GG, Renaud JC, Dumoutier L, Delzenne NM, Luquet S, Bäckhed F, Cani PD. Intestinal epithelial MyD88 is a sensor switching host metabolism towards obesity according to nutritional status. *Nat Commun* 2014; **5**: 5648 [PMID: [25476696](https://pubmed.ncbi.nlm.nih.gov/25476696/) DOI: [10.1038/ncomms6648](https://doi.org/10.1038/ncomms6648)]
- 76 **Wree A**, McGeough MD, Peña CA, Schlattjan M, Li H, Inzaugarat ME, Messer K, Canbay A, Hoffman HM, Feldstein AE. NLRP3 inflammasome activation is required for fibrosis development in NAFLD. *J Mol Med (Berl)* 2014; **92**: 1069-1082 [PMID: [24861026](https://pubmed.ncbi.nlm.nih.gov/24861026/) DOI: [10.1007/s00109-014-1170-1](https://doi.org/10.1007/s00109-014-1170-1)]
- 77 **Tanoue T**, Atarashi K, Honda K. Development and maintenance of intestinal regulatory T cells. *Nat Rev Immunol* 2016; **16**: 295-309 [PMID: [27087661](https://pubmed.ncbi.nlm.nih.gov/27087661/) DOI: [10.1038/nri.2016.36](https://doi.org/10.1038/nri.2016.36)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-3991568
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>

