



## EDITORIAL

# Molecular basis of the potential of mesalazine to prevent colorectal cancer

Carmine Stolfi, Roberto Pellegrini, Eleonora Franzè, Francesco Pallone, Giovanni Monteleone

Carmine Stolfi, Roberto Pellegrini, Eleonora Franzè, Francesco Pallone, Giovanni Monteleone, Department of Internal Medicine, University Tor Vergata of Rome, Rome 00133, Italy

Author contributions: Stolfi C, Pellegrini R, and Franzè E performed *in vitro* and *in vivo* experimental research and contributed to the draft manuscript; Pallone F and Monteleone G supervised the work and wrote the manuscript.

Correspondence to: Giovanni Monteleone, Cattedra di Gastroenterologia, Dipartimento di Medicina Interna, Università Tor Vergata, Via Montpellier 1, Rome 00133, Italy. [Gi.Monteleone@Med.uniroma2.it](mailto:Gi.Monteleone@Med.uniroma2.it)

Telephone: +39-6-72596150 Fax: +39-6-72596391

Received: March 26, 2008 Revised: April 24, 2008

Accepted: April 30, 2008

Published online: July 28, 2008

[com/1007-9327/14/4434.asp](http://com/1007-9327/14/4434.asp) DOI: <http://dx.doi.org/10.3748/wjg.14.4434>

## INTRODUCTION

Patients with ulcerative colitis (UC) and Crohn's disease (CD), the major forms of inflammatory bowel disease (IBD) in humans, are at increased risk of developing colorectal cancer (CRC)<sup>[1,2]</sup>. Chronic inflammation is believed to be the driving force for neoplastic transformation, and clinical factors that increase the risk include disease duration > 10 years, extensive disease, severity of colitis, a positive family history of sporadic CRC, and the concomitant presence of primary sclerosing cholangitis<sup>[3-5]</sup>.

While in sporadic CRC, the dysplastic precursor is usually the adenomatous polyp, dysplasia in IBD patients can be both polypoid and flat, localized or diffuse. Detection of dysplasia during programmed screening and surveillance colonoscopy is the goal of the current strategy for CRC prevention in IBD patients. When dysplasia or CRC is identified, proctocolectomy is performed to remove the at-risk organ<sup>[6]</sup>. However, no evidence exists that screening for colonic dysplasia and cancer with surveillance colonoscopy prolongs survival in IBD patients. Indirect evidence also suggests that this is a cost-effective approach<sup>[7,8]</sup>. Therefore, gastroenterologists have recently shifted their attention towards alternative strategies of chemoprevention, and particular interest has been given to the possibility of reducing the risk of IBD-related CRC by using mesalazine or 5-aminosalicylic acid (5-ASA). Mesalazine is widely used in the maintenance of remission and in the treatment of mild flare-ups of IBD, and recent epidemiological studies have suggested that this drug is chemopreventive for CRC development in UC patients<sup>[9-11]</sup>, even though some studies have documented no benefit<sup>[12]</sup>. The mechanisms behind the antineoplastic effect of mesalazine are incompletely understood, but it is likely that they are mostly dependent on the ability of the drug to attenuate ongoing mucosal inflammation. Indeed, it is well known that mesalazine can modulate various inflammatory pathways (e.g. production of inflammatory cytokines, activity of inducible nitric oxide synthase, activation of nuclear factor- $\kappa$ B) that are

## Abstract

Patients with ulcerative colitis (UC) and Crohn's disease (CD) are at increased risk for developing colorectal cancer (CRC), and this is believed to be a result of chronic inflammation. Although conclusive evidence is still missing, both epidemiological and experimental observations suggest that certain drugs used to treat inflammation, such as mesalazine, can reduce the incidence of colitis-associated CRC. Therefore, in recent years, several studies have been conducted to dissect the mechanisms by which mesalazine interferes with CRC cell growth and survival. This review summarizes the current information on the molecular mechanisms that underlie the antineoplastic action of mesalazine.

© 2008 The WJG Press. All rights reserved.

**Key words:** Chemoprevention; Colorectal cancer; Cyclooxygenase-2; Epidermal growth factor receptor; Inflammatory bowel disease; Mesalazine; Wnt/ $\beta$ -catenin

**Peer reviewer:** Tamara Cacev, MSc, Division of Molecular Medicine, Rudjer Boskovic Institute, Bijenicka c. 54, Zagreb 10000, Croatia

Stolfi C, Pellegrini R, Franzè E, Pallone F, Monteleone G. Molecular basis of the potential of mesalazine to prevent colorectal cancer. *World J Gastroenterol* 2008; 14(28): 4434-4439 Available from: URL: <http://www.wjgnet.com>

relevant to CRC initiation and progression<sup>[13-16]</sup>. There is also evidence that mesalazine inhibits the formation of reactive oxygen species (ROS) from polymorphonuclear leukocytes<sup>[17]</sup>, which leads to a decrease or complete inhibition of DNA damage, a phenomenon that has been involved in colon carcinogenesis. More recently, it has also been shown that mesalazine can directly target CRC cells and interfere with biological pathways that control their growth and survival<sup>[18]</sup>. The object of this article is to summarize recent data that elucidate the basic mechanisms of the antineoplastic effect of mesalazine.

## MESALAZINE HAS DIRECT INHIBITORY EFFECTS ON CRC CELL GROWTH AND SURVIVAL

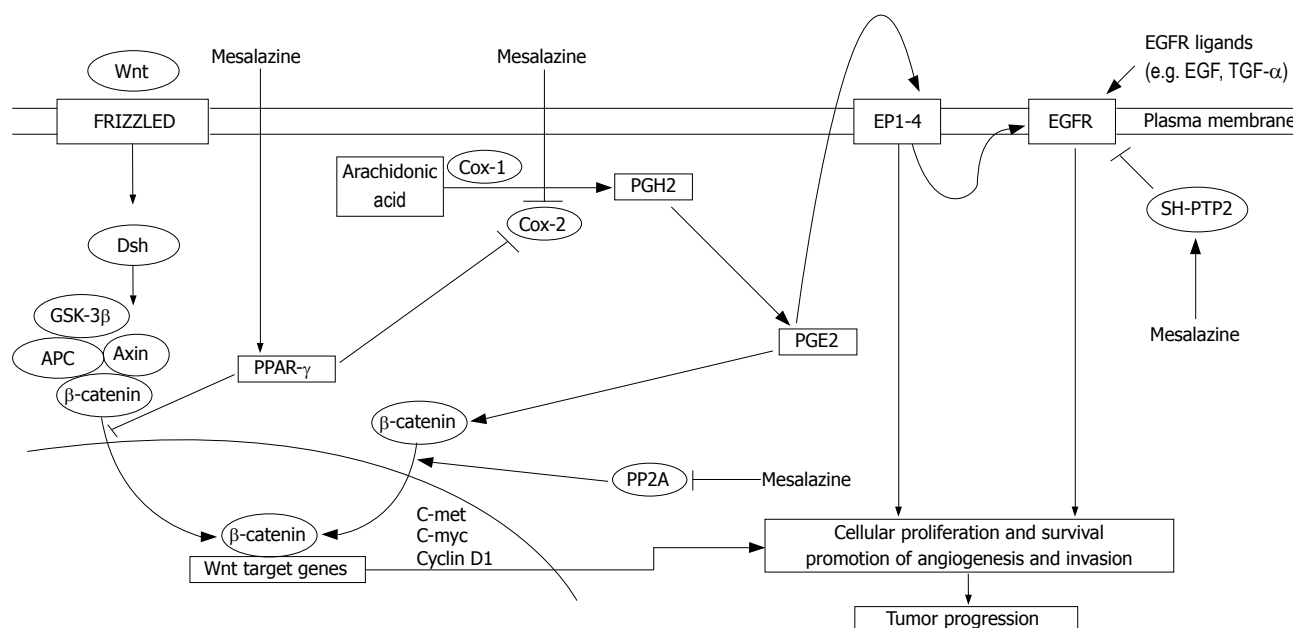
Carcinogenesis is a complex and multistage process that involves interactions between genes and environmental insults which ultimately affect cell proliferation and apoptosis. Apoptosis progressively decreases and proliferation increases in the sequential stages from normal colonic mucosa to dysplastic and CRC tissue. Therefore, strategies that inhibit cell proliferation and/or restore cell susceptibility to apoptosis have been shown to be effective in interfering with CRC initiation and/or progression.

Accumulating evidence indicates that mesalazine can block growth and promote apoptosis of CRC cells. This was initially suggested by *ex vivo* studies in patients with colonic adenoma. In particular, Reinacker-Schick *et al* have analyzed the effect of orally administered mesalazine on apoptosis and proliferation of colorectal mucosa in 21 patients with sporadic polyps. An increase in the apoptotic rate and decrease in cell proliferation were seen 1 and 3 d, respectively, after the initiation of treatment with mesalazine<sup>[19]</sup>. Bus *et al* have demonstrated that 2 wk treatment with 4 g/d mesalazine enema in patient with sporadic CRC resulted in enhanced apoptosis of tumor cells, while no change was seen in the normal mucosa that surrounded the tumor lesion. Moreover, the cellular proliferation rate as assessed by means of Ki-67 expression was unchanged in both the tumor and normal tissue<sup>[20]</sup>. Studies in rodent models of CRC showed that mesalazine inhibits tumor growth and reduces the number of aberrant cryptic foci<sup>[21,22]</sup>. Moreover, in a mouse model of colitis-associated CRC, Ikeda *et al* have shown that mesalazine, given in the remission stage of colitis, markedly suppresses the number and size of neoplasms. Notably, mesalazine treatment reduces the rate of proliferation of tumor cells, which leaves the proliferation of normal epithelial cells unaltered<sup>[23]</sup>. These observations have been reinforced by *in vitro* studies that show that mesalazine inhibits the growth and enhances apoptosis of several cultured CRC cell lines, in a time- and dose-dependent manner<sup>[18,24]</sup>. Altogether, these later findings

indicate that mesalazine has direct effects on CRC cells. This novel information has boosted new research aimed at dissecting the molecular mechanisms by which mesalazine interferes with CRC development/growth.

## EFFECTS OF MESALAZINE ON REPLICATION FIDELITY

Many of the molecular alterations that are believed to play a major role in the development of sporadic CRC are also seen in IBD-associated CRC tissue. For instance, both these types of CRC are characterized by a very similar frequency of the two main types of genomic instability, namely chromosomal instability (CIN, 85%) and microsatellite instability (MSI, 15%)<sup>[25]</sup>. CIN results in abnormal segregation of chromosomes and abnormal DNA content (aneuploidy). As a result, loss of chromosomal material often occurs, which contributes to the loss of function of important tumor suppressor genes [e.g. adenomatous polyposis coli (*APC*) and *p53*]. The MSI pathway involves the primary loss of function of genes that usually repair DNA base-pair mismatches that occur during the normal process of DNA replication in dividing cells. During this process, frameshift mutations, called microsatellites, tend to accumulate. Since microsatellites are mainly located in intronic DNA sequences, microsatellite mutations generally result in no gene function alteration. However, if microsatellites are located in exonic gene regions, the mutations can lead to a shift in the codon reading frame and changes in the amino acid sequence during mRNA translation. The introduction of an early stop codon can eventually cause protein truncation, and this is generally associated with a loss of protein function<sup>[26]</sup>. Recently, Gasche and colleagues, using an assay based on a stable transfection of a plasmid carrying a microsatellite sequence into HCT-116 cells, have shown that mesalazine improves replication fidelity in cultured CRC cell lines by reducing frameshift mutations at microsatellites. Since the effect of mesalazine is seen in mismatch repair-deficient CRC cells, it is highly likely that mesalazine acts on replication fidelity independently of post-replicative mismatch repair. The molecular mechanism by which mesalazine inhibits the generation of frameshift mutations has not yet been characterized. However, studies by the same group have shown that mesalazine can interact with cellular machinery involved in cell-cycle progression<sup>[28]</sup>. In particular, it has been shown that mesalazine can slow down DNA replication and cell division, thus allowing cells to either repair DNA damage or undergo apoptosis. Analysis of cell-cycle phases has revealed that CRC cells are arrested in S-phase when treated with mesalazine for 24-48 h. Such results are, however, somewhat different from those published by Reinacker-Schick *et al* that have shown that CRC cells are arrested in G2-M phase when exposed to mesalazine<sup>[18]</sup>. The



**Figure 1** Some putative molecular mechanisms that underlie the antineoplastic activity of mesalazine. Mesalazine inhibits COX-2, thereby blocking synthesis of PGH<sub>2</sub>, an intermediate that is metabolized into various prostaglandins, including PGE<sub>2</sub>. PGE<sub>2</sub> binds to its cell surface cognate receptors (EP1-4) and sustains various functions of tumor cells, including proliferation, survival, angiogenesis and invasion. These are mediated through the transactivation of EGFR or activation of other intracellular pathways. The antineoplastic effects of mesalazine rely also on its ability to inhibit Wnt/β-catenin, through inactivation of PP2A (and downstream down-regulation of β-catenin), and EGFR pathways. Moreover, mesalazine activates PPAR-γ, thus leading to inactivation of the Wnt/β-catenin pathway and down-regulation of COX-2.

reasons for these discrepancies remain unclear, but it is likely that they are due to differences in the culture systems used by these investigators.

which suggests that the effect of this drug on CRC cell growth is partially independent of inhibition of the COX-2/PGE<sub>2</sub> axis.

## THE ANTI-MITOTIC EFFECT OF MESALAZINE IS NOT STRICTLY DEPENDENT ON CYCLOOXYGENASE (COX)-2 INHIBITION

COX-2 is a major target of CRC chemopreventive programs, as it is highly expressed in both sporadic and familial CRC, and activation of COX-2 is known to trigger and/or amplify biological pathways that sustain CRC growth<sup>[29-32]</sup>. COX-2 is also over-expressed in IBD-related CRC tissue<sup>[33]</sup>. Since mesalazine inhibits COX-2 in inflammatory cells, it is hypothesized that the antineoplastic effect of this drug is strictly dependent on the inhibition of COX-2 in CRC cells. In this context, we have recently shown that mesalazine inhibits the growth of HT-115, a CRC cell line that expresses a functionally active COX-2, and that the anti-mitogenic effect of mesalazine is associated with a marked down-regulation of COX-2 at the protein and mRNA level<sup>[34]</sup>. Consistently, treatment of HT-115 cells with mesalazine causes a significant reduction in secretion of prostaglandin (PG) E<sub>2</sub>, a product of COX-2 activity that positively regulates CRC cell growth (Figure 1). However, exogenously added PGE<sub>2</sub> does not abrogate the inhibitory effect of mesalazine on HT-115 cell growth. Moreover, mesalazine blocks the growth of DLD-1, a CRC cell line that does not express COX-2,

## MESALAZINE INHIBITS BOTH THE WNT/β-CATENIN AND EPIDERMAL GROWTH FACTOR RECEPTOR (EGFR) SIGNALING PATHWAYS IN CRC CELLS

An intriguing hypothesis that has emerged from the available experimental data is that mesalazine may act on one or more pathways that are both early and common in colorectal carcinogenesis. A potential candidate is the wntless and integration site growth factor (Wnt)/β-catenin pathway, since it is constitutively activated in the majority of CRC<sup>[35-37]</sup>. In this pathway, Wnt binds to the transmembrane Frizzled receptor, which leads to activation of the cytoplasmic dishevelled (Dsh) protein. Dsh forms a complex with the β-catenin degradation complex, which consists of the APC gene product, glycogen synthase kinase-3β (GSK-3β), axin and β-catenin. The current model for the Wnt signaling pathway proposes that, in the absence of Wnt, GSK-3β phosphorylates β-catenin, thereby promoting ubiquitination and degradation of β-catenin<sup>[38]</sup>. In response to Wnt signals, β-catenin is no longer targeted for degradation and accumulates to high levels in the cytoplasm<sup>[39]</sup>. The accumulated β-catenin translocates to the nucleus, associates with the transcriptional enhancers of the lymphoid enhancer-binding factor/Tcf family, and stimulates the expression of genes, such as *Myc*, that

play important roles in tumor progression<sup>[40,41]</sup>.

Elegant studies by Bos *et al* have shown that mesalazine inhibits the Wnt/ $\beta$ -catenin pathway in APC-mutated CRC cells with intact  $\beta$ -catenin (Figure 1)<sup>[42]</sup>. Consistent with this, mesalazine treatment reduces expression of nuclear  $\beta$ -catenin and Wnt/ $\beta$ -catenin target genes (e.g. *cyclin D1*, *c-met* and *c-Myc*), and increased  $\beta$ -catenin phosphorylation. Mesalazine fails to inhibit the expression of Wnt/ $\beta$ -catenin target genes in  $\beta$ -catenin mutant CRC cell lines, in which the Wnt/ $\beta$ -catenin pathway is not regulated by  $\beta$ -catenin phosphorylation. These observations suggest that inhibition of the Wnt/ $\beta$ -catenin pathway by mesalazine is dependent on increased phosphorylation of  $\beta$ -catenin. In line with this, pre-incubation of CRC cells with okadaic acid, a specific phospho-serine/phospho-threonine phosphatase inhibitor, prevents mesalazine-induced  $\beta$ -catenin phosphorylation. The precise mechanism by which mesalazine enhances  $\beta$ -catenin phosphorylation remains to be ascertained, even though Bos *et al* have demonstrated that mesalazine reduces the activity of protein phosphatase 2A (PP2A), a known regulator of the  $\beta$ -catenin phosphorylation status and Wnt/ $\beta$ -catenin pathway in CRC cells.

Another target of mesalazine is EGFR (Figure 1). This receptor is highly activated in CRC cells, in which it is supposed to trigger mitogenic and pro-survival signals<sup>[43,44]</sup>. Consistent, EGFR inhibitors, such as monoclonal antibodies or small molecules that inhibit tyrosine kinase activity, have been shown to be effective in patients with advanced CRC<sup>[45]</sup>. A very high percentage of IBD-associated CRC displays immunohistochemical positivity for EGFR. Expression of EGFR is frequent in IBD-associated intestinal cancer<sup>[46]</sup>. Expression occurs at an early stage (i.e. premalignant lesions), as described in sporadic CRC<sup>[47]</sup>, which suggests that blocking EGFR activity is useful for reducing the occurrence of IBD-related CRC<sup>[48,49]</sup>. We have shown that exposure of CRC cell lines to mesalazine results in a marked suppression of EGFR phosphorylation/activation, and that mesalazine-induced EGFR dephosphorylation is dependent on neither CRC cell death induction nor shedding of the receptor<sup>[50]</sup>. Moreover, mesalazine suppresses EGFR activation induced by exogenous EGF or TGF- $\alpha$ , thus excluding the possibility that dephosphorylation of EGFR in mesalazine-treated cells is secondary to inhibition of EGFR ligand synthesis. EGFR phosphorylation is a tightly controlled phenomenon, which is the net result of the action of tyrosine kinase and phosphatases (PTPs). Therefore, we have examined whether mesalazine-mediated inhibition of EGFR activation reflects changes in the expression/activity of PTPs, which have been reported to control the extent of EGFR activation<sup>[51]</sup>. Notably, treatment of CRC cells with mesalazine causes a significant increase in the activity, but not expression, of phosphorylated-EGFR-targeting PTPs, and pre-incubation of cells with PTP inhibitors largely reduces the inhibitory effect of mesalazine on EGFR activation. Among these PTPs, both SH-PTP1 and SH-PTP2 interact with EGFR upon

mesalazine treatment. However, targeted silencing of SH-PTP2, but not SH-PTP1 prevents mesalazine-induced EGFR dephosphorylation. Consistent with these data, mesalazine also attenuates EGFR phosphorylation in *ex vivo* organ cultures of human sporadic CRC explants.

## MESALAZINE ACTIVATES PEROXISOME PROLIFERATOR ACTIVATED RECEPTOR- $\gamma$ (PPAR- $\gamma$ ) IN CRC CELLS

PPAR- $\gamma$  is a nuclear receptor that is highly expressed in the colon, and plays a key role in bacteria-induced inflammation. Many factors can modulate the activity of PPAR- $\gamma$ , but the most important activating factor in colon epithelial cells appears to be the luminal flora<sup>[52]</sup>. Activation of PPAR- $\gamma$  also has anti-tumorigenic effects that are manifested as both anti-proliferative and pro-apoptotic activities<sup>[53, 54]</sup>, inhibition of the formation of aberrant cryptic foci<sup>[55]</sup>, and inhibition of CRC development<sup>[56]</sup>. It has also been shown that PPAR- $\gamma$  suppresses tumor formation by interfering with the Wnt/ $\beta$ -catenin signaling pathway<sup>[57,58]</sup>. Recent *in vitro* and *in vivo* studies have shown that mesalazine can activate PPAR- $\gamma$  (Figure 1). In particular, using HT-29 CRC cells, Rousseaux *et al* have shown that mesalazine enhances PPAR- $\gamma$  expression, stimulates translocation of PPAR- $\gamma$  to the nucleus, induces conformational changes in the PPAR- $\gamma$  molecule, and increases the interaction between PPAR- $\gamma$  and vitamin D3 receptor-interacting protein 205<sup>[59]</sup>. In competitive binding studies, mesalazine displaces rosiglitazone and the selective PPAR- $\gamma$  ligand GW1929 from their binding sites on the PPAR- $\gamma$  molecule<sup>[59]</sup>. In line with these *in vitro* findings, it has been shown that the antineoplastic effects of mesalazine are mediated by PPAR- $\gamma$  in a model of CRC, in which SCID mice were engrafted with human CRC cells. In particular, in this model, locally administered mesalazine significantly reduced the growth of xenografts, and this effect was blocked by the selective PPAR- $\gamma$  antagonist GW9662<sup>[60]</sup>.

## CONCLUSION

In recent years, there has been great interest in the possibility of chemoprevention of IBD-related CRC by mesalazine. Given the difficulty of performing double-blind, placebo-controlled, randomized clinical trials in patients, investigators have turned to experimental models of cancer, and indeed, the existing data suggest that mesalazine can reduce the risk of CRC by directly interfering with CRC cell biology, other than by simply controlling inflammation. However, definitive conclusions from experimental findings are not always completely correct, and therefore, future studies will be necessary to ascertain whether data generated from studies with cultured cells or animal models of CRC can be generalized to IBD-associated dysplasia or CRC.

Another important issue that needs further investigation regards the dosage/concentration of



mesalazine required to interfere with CRC cell growth and survival. *In vitro* studies have indicated that the antineoplastic effect of mesalazine is seen with relative high drug dosages (e.g. 10-50 mmol/L), which are not always reached within the colonic tissue under standard oral treatment. In this context, it is also relevant to take into consideration mesalazine metabolism, for instance, mesalazine oxidation and acetylation, which could differ considerably between *in vitro* and *in vivo* circumstances, and limit the amount of biologically active compound.

## REFERENCES

- Eaden JA, Abrams KR, Mayberry JF. The risk of colorectal cancer in ulcerative colitis: a meta-analysis. *Gut* 2001; **48**: 526-535
- Jess T, Gamborg M, Matzen P, Munkholm P, Sorensen TI. Increased risk of intestinal cancer in Crohn's disease: a meta-analysis of population-based cohort studies. *Am J Gastroenterol* 2005; **100**: 2724-2729
- Munkholm P. Review article: the incidence and prevalence of colorectal cancer in inflammatory bowel disease. *Aliment Pharmacol Ther* 2003; **18** Suppl 2: 1-5
- Rutter M, Saunders B, Wilkinson K, Rumbles S, Schofield G, Kamm M, Williams C, Price A, Talbot I, Forbes A. Severity of inflammation is a risk factor for colorectal neoplasia in ulcerative colitis. *Gastroenterology* 2004; **126**: 451-459
- Gupta RB, Harpaz N, Itzkowitz S, Hossain S, Matula S, Kornbluth A, Bodian C, Ullman T. Histologic inflammation is a risk factor for progression to colorectal neoplasia in ulcerative colitis: a cohort study. *Gastroenterology* 2007; **133**: 1099-1105; quiz 1340-1341
- Eaden J. Review article: colorectal carcinoma and inflammatory bowel disease. *Aliment Pharmacol Ther* 2004; **20** Suppl 4: 24-30
- Obrador A, Ginard D, Barranco L. Review article: colorectal cancer surveillance in ulcerative colitis - what should we be doing? *Aliment Pharmacol Ther* 2006; **24** Suppl 3: 56-63
- Jess T, Loftus EV Jr, Velayos FS, Harmsen WS, Zinsmeister AR, Smyrk TC, Schleck CD, Tremaine WJ, Melton LJ 3rd, Munkholm P, Sandborn WJ. Risk of intestinal cancer in inflammatory bowel disease: a population-based study from olmsted county, Minnesota. *Gastroenterology* 2006; **130**: 1039-1046
- Velayos FS, Terdiman JP, Walsh JM. Effect of 5-aminosalicylate use on colorectal cancer and dysplasia risk: a systematic review and metaanalysis of observational studies. *Am J Gastroenterol* 2005; **100**: 1345-1353
- Van Staa TP, Card T, Logan RF, Leufkens HG. 5-Aminosalicylate use and colorectal cancer risk in inflammatory bowel disease: a large epidemiological study. *Gut* 2005; **54**: 1573-1578
- Eaden J, Abrams K, Ekbohm A, Jackson E, Mayberry J. Colorectal cancer prevention in ulcerative colitis: a case-control study. *Aliment Pharmacol Ther* 2000; **14**: 145-153
- Bernstein CN, Blanchard JF, Metge C, Yogendran M. Does the use of 5-aminosalicylates in inflammatory bowel disease prevent the development of colorectal cancer? *Am J Gastroenterol* 2003; **98**: 2784-2788
- Bantel H, Berg C, Vieth M, Stolte M, Kruis W, Schulze-Osthoff K. Mesalazine inhibits activation of transcription factor NF-kappaB in inflamed mucosa of patients with ulcerative colitis. *Am J Gastroenterol* 2000; **95**: 3452-3457
- Kaiser GC, Yan F, Polk DB. Mesalazine blocks tumor necrosis factor growth inhibition and nuclear factor kappaB activation in mouse colonocytes. *Gastroenterology* 1999; **116**: 602-609
- Mahida YR, Lamming CE, Gallagher A, Hawthorne AB, Hawkey CJ. 5-Aminosalicylic acid is a potent inhibitor of interleukin 1 beta production in organ culture of colonic biopsy specimens from patients with inflammatory bowel disease. *Gut* 1991; **32**: 50-54
- Kennedy M, Wilson L, Szabo C, Salzman AL. 5-aminosalicylic acid inhibits iNOS transcription in human intestinal epithelial cells. *Int J Mol Med* 1999; **4**: 437-443
- Miyachi Y, Yoshioka A, Imamura S, Niwa Y. Effect of sulphasalazine and its metabolites on the generation of reactive oxygen species. *Gut* 1987; **28**: 190-195
- Reinacher-Schick A, Schoeneck A, Graeven U, Schwarte-Waldhoff I, Schmiegeler W. Mesalazine causes a mitotic arrest and induces caspase-dependent apoptosis in colon carcinoma cells. *Carcinogenesis* 2003; **24**: 443-451
- Reinacher-Schick A, Seidensticker F, Petrasch S, Reiser M, Philippou S, Theegarten D, Freitag G, Schmiegeler W. Mesalazine changes apoptosis and proliferation in normal mucosa of patients with sporadic polyps of the large bowel. *Endoscopy* 2000; **32**: 245-254
- Bus PJ, Nagtegaal ID, Verspaget HW, Lamers CB, Geldof H, Van Krieken JH, Griffioen G. Mesalazine-induced apoptosis of colorectal cancer: on the verge of a new chemopreventive era? *Aliment Pharmacol Ther* 1999; **13**: 1397-1402
- Brown WA, Farmer KC, Skinner SA, Malcontenti-Wilson C, Misajon A, O'Brien PE. 5-aminosalicylic acid and olsalazine inhibit tumor growth in a rodent model of colorectal cancer. *Dig Dis Sci* 2000; **45**: 1578-1584
- MacGregor DJ, Kim YS, Sleisenger MH, Johnson LK. Chemoprevention of colon cancer carcinogenesis by balsalazide: inhibition of azoxymethane-induced aberrant crypt formation in the rat colon and intestinal tumor formation in the B6-Min/+ mouse. *Int J Oncol* 2000; **17**: 173-179
- Ikeda I, Tomimoto A, Wada K, Fujisawa T, Fujita K, Yonemitsu K, Nozaki Y, Endo H, Takahashi H, Yoneda M, Inamori M, Kubota K, Saito S, Nagashima Y, Nakagama H, Nakajima A. 5-aminosalicylic acid given in the remission stage of colitis suppresses colitis-associated cancer in a mouse colitis model. *Clin Cancer Res* 2007; **13**: 6527-6531
- Fina D, Franchi L, Caruso R, Peluso I, Naccari GC, Bellinva S, Testi R, Pallone F, Monteleone G. 5-aminosalicylic acid enhances anchorage-independent colorectal cancer cell death. *Eur J Cancer* 2006; **42**: 2609-2616
- Lengauer C, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. *Nature* 1998; **396**: 643-649
- Ionov Y, Peinado MA, Malkhosyan S, Shibata D, Perucho M. Ubiquitous somatic mutations in simple repeated sequences reveal a new mechanism for colonic carcinogenesis. *Nature* 1993; **363**: 558-561
- Gasche C, Goel A, Natarajan L, Boland CR. Mesalazine improves replication fidelity in cultured colorectal cells. *Cancer Res* 2005; **65**: 3993-3997
- Luciani MG, Campregher C, Fortune JM, Kunkel TA, Gasche C. 5-ASA affects cell cycle progression in colorectal cells by reversibly activating a replication checkpoint. *Gastroenterology* 2007; **132**: 221-235
- Huls G, Koornstra JJ, Kleibeuker JH. Non-steroidal anti-inflammatory drugs and molecular carcinogenesis of colorectal carcinomas. *Lancet* 2003; **362**: 230-232
- Koehne CH, Dubois RN. COX-2 inhibition and colorectal cancer. *Semin Oncol* 2004; **31**: 12-21
- Dubois RN, Abramson SB, Crofford L, Gupta RA, Simon LS, Van De Putte LB, Lipsky PE. Cyclooxygenase in biology and disease. *FASEB J* 1998; **12**: 1063-1073
- Tsujii M, Kawano S, DuBois RN. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. *Proc Natl Acad Sci USA* 1997; **94**: 3336-3340
- Eaden J. Review article: the data supporting a role for aminosalicylates in the chemoprevention of colorectal cancer in patients with inflammatory bowel disease. *Aliment Pharmacol Ther* 2003; **18** Suppl 2: 15-21
- Stolfi C, Fina D, Caruso R, Caprioli F, Sarra M, Fantini MC, Rizzo A, Pallone F, Monteleone G. Cyclooxygenase-

- 2-dependent and -independent inhibition of proliferation of colon cancer cells by 5-aminosalicylic acid. *Biochem Pharmacol* 2008; **75**: 668-676
- 35 **Polakis P.** Wnt signaling and cancer. *Genes Dev* 2000; **14**: 1837-1851
- 36 **Behrens J, Lustig B.** The Wnt connection to tumorigenesis. *Int J Dev Biol* 2004; **48**: 477-487
- 37 **Oving IM, Clevers HC.** Molecular causes of colon cancer. *Eur J Clin Invest* 2002; **32**: 448-457
- 38 **Orford K, Crockett C, Jensen JP, Weissman AM, Byers SW.** Serine phosphorylation-regulated ubiquitination and degradation of beta-catenin. *J Biol Chem* 1997; **272**: 24735-24738
- 39 **Miller JR, Hocking AM, Brown JD, Moon RT.** Mechanism and function of signal transduction by the Wnt/beta-catenin and Wnt/Ca<sup>2+</sup> pathways. *Oncogene* 1999; **18**: 7860-7872
- 40 **Ponzielli R, Katz S, Barsyte-Lovejoy D, Penn LZ.** Cancer therapeutics: targeting the dark side of Myc. *Eur J Cancer* 2005; **41**: 2485-2501
- 41 **Mann B, Gelos M, Siedow A, Hanski ML, Gratchev A, Ilyas M, Bodmer WF, Moyer MP, Riecken EO, Buhr HJ, Hanski C.** Target genes of beta-catenin-T cell-factor/lymphoid-enhancer-factor signaling in human colorectal carcinomas. *Proc Natl Acad Sci USA* 1999; **96**: 1603-1608
- 42 **Bos CL, Diks SH, Hardwick JC, Walburg KV, Peppelenbosch MP, Richel DJ.** Protein phosphatase 2A is required for mesalazine-dependent inhibition of Wnt/beta-catenin pathway activity. *Carcinogenesis* 2006; **27**: 2371-2382
- 43 **Jorissen RN, Walker F, Pouliot N, Garrett TP, Ward CW, Burgess AW.** Epidermal growth factor receptor: mechanisms of activation and signalling. *Exp Cell Res* 2003; **284**: 31-53
- 44 **Bradley SJ, Garfinkle G, Walker E, Salem R, Chen LB, Steele G Jr.** Increased expression of the epidermal growth factor receptor on human colon carcinoma cells. *Arch Surg* 1986; **121**: 1242-1247
- 45 **Janmaat ML, Giaccone G.** The epidermal growth factor receptor pathway and its inhibition as anticancer therapy. *Drugs Today (Barc)* 2003; **39** Suppl C: 61-80
- 46 **Svrcek M, Cosnes J, Tiret E, Bennis M, Parc Y, Flejou JF.** Expression of epidermal growth factor receptor (EGFR) is frequent in inflammatory bowel disease (IBD)-associated intestinal cancer. *Virchows Arch* 2007; **450**: 243-244
- 47 **Martin ES, Tonon G, Sinha R, Xiao Y, Feng B, Kimmelman AC, Protopopov A, Ivanova E, Brennan C, Montgomery K, Kucherlapati R, Bailey G, Redston M, Chin L, DePinho RA.** Common and distinct genomic events in sporadic colorectal cancer and diverse cancer types. *Cancer Res* 2007; **67**: 10736-10743
- 48 **Mendelsohn J.** The epidermal growth factor receptor as a target for cancer therapy. *Endocr Relat Cancer* 2001; **8**: 3-9
- 49 **Mendelsohn J, Baselga J.** The EGF receptor family as targets for cancer therapy. *Oncogene* 2000; **19**: 6550-6565
- 50 **Monteleone G, Franchi L, Fina D, Caruso R, Vavassori P, Monteleone I, Calabrese E, Naccari GC, Bellinva S, Testi R, Pallone F.** Silencing of SH-PTP2 defines a crucial role in the inactivation of epidermal growth factor receptor by 5-aminosalicylic acid in colon cancer cells. *Cell Death Differ* 2006; **13**: 202-211
- 51 **Moghal N, Sternberg PW.** Multiple positive and negative regulators of signaling by the EGF-receptor. *Curr Opin Cell Biol* 1999; **11**: 190-196
- 52 **Dubuquoy L, Rousseaux C, Thuru X, Peyrin-Biroulet L, Romano O, Chavatte P, Chamailard M, Desreumaux P.** PPARgamma as a new therapeutic target in inflammatory bowel diseases. *Gut* 2006; **55**: 1341-1349
- 53 **Matthiessen MW, Pedersen G, Albrechtsen T, Adamsen S, Fleckner J, Brynskov J.** Peroxisome proliferator-activated receptor expression and activation in normal human colonic epithelial cells and tubular adenomas. *Scand J Gastroenterol* 2005; **40**: 198-205
- 54 **Shimada T, Kojima K, Yoshiura K, Hiraishi H, Terano A.** Characteristics of the peroxisome proliferator activated receptor gamma (PPARgamma) ligand induced apoptosis in colon cancer cells. *Gut* 2002; **50**: 658-664
- 55 **Tanaka T, Kohno H, Yoshitani S, Takashima S, Okumura A, Murakami A, Hosokawa M.** Ligands for peroxisome proliferator-activated receptors alpha and gamma inhibit chemically induced colitis and formation of aberrant crypt foci in rats. *Cancer Res* 2001; **61**: 2424-2428
- 56 **Osawa E, Nakajima A, Wada K, Ishimine S, Fujisawa N, Kawamori T, Matsuhashi N, Kadowaki T, Ochiai M, Sekihara H, Nakagama H.** Peroxisome proliferator-activated receptor gamma ligands suppress colon carcinogenesis induced by azoxymethane in mice. *Gastroenterology* 2003; **124**: 361-367
- 57 **Jansson EA, Are A, Greicius G, Kuo IC, Kelly D, Arulampalam V, Pettersson S.** The Wnt/beta-catenin signaling pathway targets PPARgamma activity in colon cancer cells. *Proc Natl Acad Sci USA* 2005; **102**: 1460-1465
- 58 **Lu D, Cottam HB, Corr M, Carson DA.** Repression of beta-catenin function in malignant cells by nonsteroidal antiinflammatory drugs. *Proc Natl Acad Sci USA* 2005; **102**: 18567-18571
- 59 **Rousseaux C, Lefebvre B, Dubuquoy L, Lefebvre P, Romano O, Auwerx J, Metzger D, Wahli W, Desvergne B, Naccari GC, Chavatte P, Farce A, Bulois P, Cortot A, Colombel JF, Desreumaux P.** Intestinal antiinflammatory effect of 5-aminosalicylic acid is dependent on peroxisome proliferator-activated receptor-gamma. *J Exp Med* 2005; **201**: 1205-1215
- 60 **Desreumaux P, Ghosh S.** Review article: mode of action and delivery of 5-aminosalicylic acid - new evidence. *Aliment Pharmacol Ther* 2006; **24** Suppl 1: 2-9

S- Editor Li DL L- Editor Kerr C E- Editor Zhang WB