



TOPIC HIGHLIGHT

Natalia A Osna, MD, PhD, Series Editor

Is the iron regulatory hormone hepcidin a risk factor for alcoholic liver disease?

Duygu Dee Harrison-Findik

Duygu Dee Harrison-Findik, Division of Gastroenterology and Hepatology, Department of Internal Medicine, University of Nebraska Medical Center, Omaha, NE, 68198-5820, United States

Author contributions: Harrison-Findik DD wrote this review. Supported by Grants from the Alcoholic Beverage Medical Research Foundation, Redox Biology Center, University of Nebraska-Lincoln, 2P20RR017675 and NIH grant, R01AA017738-01 to DHF

Correspondence to: Duygu Dee Harrison-Findik, Division of Gastroenterology and Hepatology, Department of Internal Medicine, University of Nebraska Medical Center, Omaha, NE, 68198-5820, United States. dharrisonfindik@unmc.edu

Telephone: +1-402-5596355 Fax: +1-402-5596494

Received: December 17, 2008 Revised: February 11, 2009

Accepted: February 18, 2009

Published online: March 14, 2009

Abstract

Despite heavy consumption over a long period of time, only a small number of alcoholics develop alcoholic liver disease. This alludes to the possibility that other factors, besides alcohol, may be involved in the progression of the disease. Over the years, many such factors have indeed been identified, including iron. Despite being crucial for various important biological processes, iron can also be harmful due to its ability to catalyze Fenton chemistry. Alcohol and iron have been shown to interact synergistically to cause liver injury. Iron-mediated cell signaling has been reported to be involved in the pathogenesis of experimental alcoholic liver disease. Hepcidin is an iron-regulatory hormone synthesized by the liver, which plays a pivotal role in iron homeostasis. Both acute and chronic alcohol exposure suppress hepcidin expression in the liver. The sera of patients with alcoholic liver disease, particularly those exhibiting higher serum iron indices, have also been reported to display reduced prohepcidin levels. Alcohol-mediated oxidative stress is involved in the inhibition of hepcidin promoter activity and transcription in the liver. This in turn leads to an increase in intestinal iron transport and liver iron storage. Hepcidin is expressed primarily in hepatocytes. It is noteworthy that both hepatocytes and Kupffer cells are involved in the progression of alcoholic liver disease. However, the activation of Kupffer cells and TNF- α signaling has been reported not to be involved in the down-regulation of hepcidin expression by alcohol

in the liver. Alcohol acts within the parenchymal cells of the liver to suppress the synthesis of hepcidin. Due to its crucial role in the regulation of body iron stores, hepcidin may act as a secondary risk factor in the progression of alcoholic liver disease. The clarification of the mechanisms by which alcohol disrupts iron homeostasis will allow for further understanding of the pathogenesis of alcoholic liver disease.

© 2009 The WJG Press and Baishideng. All rights reserved.

Key words: Alcohol; Hepatocyte; Kupffer cells; Oxidative stress; Second hit

Peer reviewer: Shivendra Shukla, Professor, Department of Medical Pharmacology and Physiology, University of Missouri School of Medicine, 1 Hospital Drive, M530 Medical Sciences Bldg., Columbia MO 65212, United States

Harrison-Findik DD. Is the iron regulatory hormone hepcidin a risk factor for alcoholic liver disease? *World J Gastroenterol* 2009; 15(10): 1186-1193 Available from: URL: <http://www.wjgnet.com/1007-9327/15/1186.asp> DOI: <http://dx.doi.org/10.3748/wjg.15.1186>

INTRODUCTION

Many eukaryotes require iron for critical biological processes^[1,2]. However, excess iron is toxic, causing lipid peroxidation and oxidative stress^[3]. Due to it being a “double edged sword”, iron uptake and transport must therefore be tightly regulated^[4-7]. Hepcidin, a circulatory peptide synthesized by the liver, performs this important task by regulating iron transport in different parts of the body including duodenum, spleen and bone marrow^[8,9]. Alcohol has been shown to suppress hepcidin expression in the liver leading to an increase in intestinal iron transport^[10-15]. Both iron and alcohol act synergistically to cause liver injury. Disruption of hepcidin synthesis by alcohol may therefore be one of the “second hit” mechanisms leading to the progression of alcoholic liver disease.

ALCOHOL-INDUCED DISTURBANCES IN IRON METABOLISM

Alcohol consumption has long been associated with

changes in iron homeostasis. The reported changes range from anemia to iron overload^[16-22]. Blood loss is common in patients with alcoholic cirrhosis^[23]. Anemia and iron deficiency can be due to gastrointestinal blood loss arising from the complications of alcohol abuse. Nutritional deficiencies, such as folate deficiency, can also be a common cause of anemia^[24-27]. Megaloblastic and sideroblastic anemias, macrocytosis of alcoholism, elevated mean corpuscular volume, and iron deficiency have been reported to be common among hospitalized chronic alcoholics^[23,27-30]. However, it has been demonstrated that anemia and megaloblastic and sideroblastic changes do not occur if an adequate diet is provided during chronic alcohol administration to human volunteers^[25,31]. This has also been shown to be the case for relatively well-nourished alcoholics^[25,26,32,33]. These findings therefore suggest that alcohol by itself does not induce iron deficiency or anemia.

In fact, wine has a high iron content and increases iron absorption^[34]. Decreasing the alcohol content of red wine has been reported to significantly reduce the absorption of non-heme iron in human subjects^[35]. Beer consumption has been suggested to have a more significant effect on serum ferritin levels than wine and spirits, both in males and females^[18]. Alcohol consumption (up to 2 alcoholic drinks per day) has also been shown to exert a protective effect by reducing the risk for iron deficiency anemia in adult participants of the Third National Health and Nutrition Examination Survey^[36]. On the other hand, heavy alcohol consumption (more than two alcoholic drinks per day) elevates the risk of iron overload^[36]. In both male and female adolescents (aged 16-19 years), who participated in the First National Health and Nutrition Examination Survey, serum iron concentration was significantly related to drinking frequency^[17]. Alcohol consumption was associated with elevated serum iron concentration in both male and female adolescents. Interestingly, male adolescents also exhibited increased transferrin saturation and hemoglobin concentration^[17]. Drinking frequency was unrelated to dietary iron intake, poverty index or race^[17]. A Danish population survey also reported a correlation between alcohol intake and elevated serum ferritin in healthy adult males and females (aged 30-60 years)^[37]. African Americans who consume alcohol (4 drinks per day) exhibit a higher prevalence of ferroportin Q248H allele, which is implicated in iron accumulation, and alcohol has been suggested to contribute to higher body iron stores in this population^[38]. Alcohol elevates iron absorption and patients with alcoholic cirrhosis often exhibit elevated liver iron content^[39,40]. *In vivo* whole-body retention studies have demonstrated a two-fold increase in intestinal iron absorption in chronic alcoholics^[41]. Alcoholic cirrhosis patients with higher liver iron content display increased mortality rates^[42]. Experimental animal models of prolonged alcohol exposure also display increased liver iron deposition and hepatocellular injury^[19,43]. In mild cases of alcoholic liver disease (ALD), iron has been reported to accumulate in hepatocytes^[44]. However, in patients with advanced ALD, both parenchymal cells and Kupffer

cells exhibit iron staining^[20].

In some alcoholics, the elevated serum iron indices have been reported to significantly decrease within 2-6 wk of abstinence, suggesting a direct role for alcohol consumption^[45]. However, the mechanisms by which alcohol disrupts iron metabolism are unclear. Recent studies indicate a role for hepcidin in this process.

HEPCIDIN

The discovery of the circulating peptide hormone hepcidin has revolutionized our understanding of iron metabolism. Hepcidin sequence is highly conserved between species, from fish to mammals. Hepcidin was first isolated from human urine and serum as short cysteine-rich peptide forms^[46,47]. Hepcidin is homologous to the members of the defensin family of antimicrobial peptides and has been reported to exhibit antimicrobial activity *in vitro*^[46]. It is the studies with transgenic mice that have highlighted the importance of this protein in the regulation of iron metabolism. Namely, hepcidin knockout mice, deficient in the expression of hepcidin, develop severe iron overload, whereas mice overexpressing hepcidin display iron deficiency and anemia^[48,49]. Moreover, the synthesis of hepcidin has also been shown to be responsive to the body iron levels. Iron overload state induces hepcidin synthesis whereas iron deficiency and anemia have the opposite effect^[50,51].

Liver is the main site of hepcidin synthesis in the body^[50]. It is synthesized as an 84 amino acid precursor protein and is subsequently cleaved into the biologically active 25 amino acid peptide form^[8,9,50,52]. In the liver, hepcidin is primarily expressed in the hepatocytes (Figure 1). Although to a much lesser extent, other cells of the liver and other organs also display hepcidin expression. The exciting discovery of the interaction between hepcidin and the iron exporter protein ferroportin was pivotal to our understanding of how hepcidin regulates iron metabolism^[53,54]. The binding of hepcidin to ferroportin induces the internalization and degradation of the ferroportin protein, which in turn inhibits iron from being transported into the circulation. This is how hepcidin accomplishes the regulation of iron mobilization between distant locations (i.e. duodenum, bone marrow, reticuloendothelial macrophages) in the body (Figure 1)^[8].

The synthesis of hepcidin in the liver is modulated by numerous upstream regulators^[1,2]. Transferrin receptor 2, the genetic hemochromatosis gene product, Hfe, and the juvenile hemochromatosis gene product, HJV, are positive regulators of hepcidin expression^[55-62]. Mutations in these genes result in an increase in body iron stores (i.e. hemochromatosis) due to the down-regulation of hepcidin expression. Recently, a novel candidate TMPRSS6, a transmembrane serine protease also known as matriptase 2, has been identified. Interestingly, TMPRSS6 is the first identified negative regulator of liver hepcidin expression. In other words, TMPRSS6 keeps hepcidin expression under control in the liver^[63,64]. Accordingly, patients expressing TMPRSS6 mutations exhibit iron-refractory iron deficiency anemia due to elevated hepcidin produc-

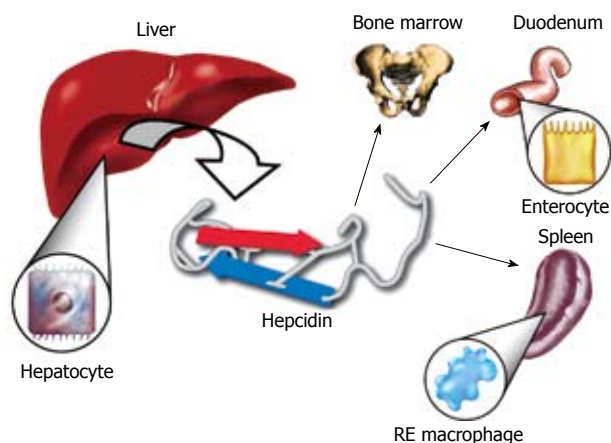


Figure 1 The Iron regulatory hormone hepcidin. Hepcidin is synthesized in the hepatocytes of the liver as an 84 amino acid precursor protein. It is subsequently cleaved into the 25 amino acid biologically active peptide form and is released into the circulation. Hepcidin plays a central role in the regulation of iron metabolism by inhibiting the release of iron from the enterocytes of the duodenum and from reticuloendothelial macrophages. Hepcidin blocks the export of iron from these cells by binding to the iron exporter protein, ferroportin, which induces the internalization and degradation of ferroportin protein. As a soluble mediator, hepcidin establishes the cross-talk between distant organs in the body in order to maintain iron homeostasis.

tion^[63]. Moreover, the splicing defects in the TMPRSS6 gene lead to the mask phenotype, a recessive, chemically induced mutant mouse phenotype^[64]. This phenotype results from the reduced absorption of dietary iron due to high levels of hepcidin expression^[64]. However, the signaling mechanisms involved in these regulatory processes (negative or positive) are as yet unknown.

KUPFFER CELLS, ALCOHOL AND HEPcidIN

Liver macrophages (Kupffer cells) also play a pivotal role in the progression of ALD. Alcoholics have elevated levels of lipopolysaccharide (LPS) in their circulation, which is believed to prime the macrophages leading to the release of proinflammatory cytokines, particularly TNF- α , and reactive oxygen species^[65-67]. The importance of Kupffer cells in ALD has been demonstrated in studies where their depletion or inactivation blocked alcohol-induced symptoms, including inflammation, fatty liver and necrosis, in experimental animal models of chronic alcohol exposure^[68,69].

In advanced forms of ALD and in experimental animal models of ALD, besides parenchymal cells, Kupffer cells also exhibit iron accumulation^[20,70,71]. Reticuloendothelial (RE) macrophages play a major role in iron trafficking and recycling to meet changes in the body's iron demand^[72]. Like the parenchymal cells of the liver, the RE system also stores large amounts of iron bound to the storage protein, ferritin. RE macrophages acquire iron mainly by phagocytosing senescent red blood cells. However, macrophages also express the divalent metal transporter1 (DMT1), natural resistance associated macrophage protein1 (Nramp1), hemoglobin scavenger receptor (CD163) and transferrin receptor1 (TrfR1), all

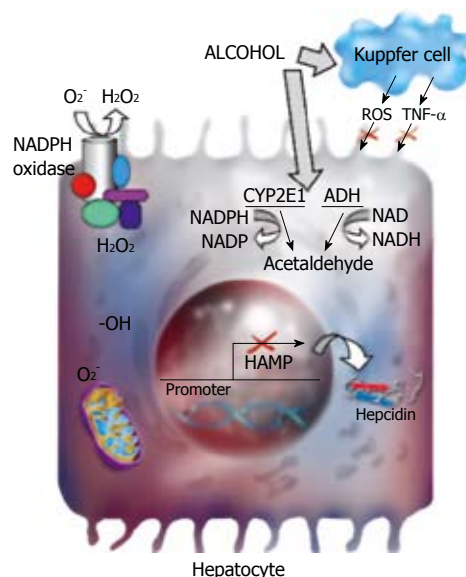


Figure 2 Hepcidin and alcohol. Alcohol is metabolized by alcohol dehydrogenase (ADH) and cytochrome P4502E1 (CYP2E1) in the liver. Alcohol-induced oxidative stress leads to the suppression of hepcidin promoter activity and hepcidin transcription in the liver. The parenchymal, but not the non-parenchymal cells of the liver are involved in the regulation of hepcidin transcription by alcohol-induced oxidative stress. The activation of CYP2E1 or NADPH oxidase and changes in mitochondrial functions are involved in alcohol-induced oxidative stress in hepatocytes. The role of these pathways in the regulation of hepcidin transcription by alcohol requires further investigation.

of which are involved in iron uptake and transport^[72-75]. The livers from experimental animal models of ALD and Japanese patients with ALD have been reported to exhibit significantly more TrfR1 mRNA or protein expression, respectively, compared to control animals or healthy human subjects^[76,77]. Macrophages can take up transferrin-bound iron *in vitro*. However, a significant uptake of transferrin-bound iron by macrophages has not been observed *in vivo* in humans^[72,78]. Although an increase in TrfR1 mRNA expression was observed in Kupffer cells of experimental animals, the increase in TrfR1 protein expression in Japanese patients with ALD was observed mainly in the hepatocytes^[20,76,77]. It should also be noted that mice with disrupted TrfR1 gene, deficient in the global expression of TrfR1 receptor, display abnormalities only in erythropoiesis and neurologic development^[79]. The Kupffer cells of rats exposed to chronic alcohol have also been shown to display an increase in mRNA levels of Hfe, the gene for genetic hemochromatosis^[77]. Hfe gene product does not bind iron and is not directly involved in iron uptake. However, it is involved in the regulation of iron metabolism as shown by the fact that mice deficient in Hfe expression develop iron overload^[80,81]. Hfe is believed to achieve this by modulating the expression and iron responsiveness of hepcidin in the liver^[57,58,82]. On the other hand, the iron release from macrophages is regulated by the iron exporter protein ferroportin^[83,84]. The multicopper ferroxidase, ceruloplasmin may also play a role in iron efflux in macrophages^[85-87]. The Kupffer cells of animals subjected to alcohol exposure have been shown to display an increase in both ferroportin mRNA and protein,

indicative of increased iron export^[77]. Of note, hepcidin binds to ferroportin and induces its internalization and degradation^[53,54]. The increase in ferroportin expression in the Kupffer cells of these animals may be due to suppressed hepcidin expression. Alcohol has been shown to down-regulate hepcidin expression both in animal models of ALD and in patients with ALD^[11-13]. However, Xiong *et al*^[77] have reported no change in plasma pro-hepcidin levels and a decrease in hepcidin mRNA levels in Kupffer cells in their experimental model.

Iron has been reported to prime Kupffer cells for alcoholic liver injury in rats exposed to chronic intragastric alcohol infusion^[70]. The proinflammatory cytokine TNF- α is a key player in alcohol-induced liver injury^[88-90]. Iron has been shown to activate the transcription factor, NF- κ B in hepatic macrophages and induce the synthesis and release of TNF- α ^[70,91]. Following erythrophagocytosis, Kupffer cells have been shown to display elevated levels of LPS-induced NF- κ B activation^[70]. Treatment of cultured Kupffer cells *in vitro* and *ex vivo* with iron chelators blocked both NF- κ B activation and TNF- α synthesis^[70,71]. Proinflammatory cytokines (IL-1, IL-6, TNF- α) have been suggested to increase iron uptake into the monocytes of patients with rheumatoid arthritis^[92]. It is therefore possible that alcohol and/or iron-induced release of TNF- α may serve as a feedback loop for further iron uptake into the Kupffer cells. On the other hand, Olynyk *et al*^[93] have reported that deposition of iron in Kupffer cells impairs LPS-induced proinflammatory cytokine production in these cells. Xiong *et al*^[77] have reported that the intracellular labile non-heme iron pool is involved in peroxynitrite or LPS-mediated activation of NF- κ B and TNF- α release in Kupffer cells isolated from experimental animal models of chronic alcohol exposure. The intracellular labile iron pool is a transitory, free (non-ferritin-bound) pool of iron, which is chelatable by commonly used iron chelators. This pool of iron contributes to the generation of reactive oxygen species but its biological relevance is unclear^[94]. Transient changes in the dynamics of the labile iron pool is not only affected by increased iron uptake but also by prooxidant chemicals, which induce the reductive release of iron from intracellular stores^[95,96]. Xiong *et al*^[77] have demonstrated that iron dextran treatment of cultured Kupffer cells *in vitro* does not affect the basal TNF- α release. Interestingly, Kupffer cells isolated from mice two weeks after iron dextran injection, exhibited significant increases in basal TNF- α release (independent of peroxynitrite or LPS stimulation). These findings suggest that either other cells in the liver (besides Kupffer cells) or yet unknown signals contribute to this process *in vivo*. It is also possible that the discrepancy between *in vitro* and *in vivo* iron loading regarding basal TNF- α release from Kupffer cells may not be due to iron (iron loading) *per se* but rather due to the differences in the level of oxidative stress.

The synthesis of hepcidin in the liver is mediated by several stimuli including iron and inflammation. Kupffer cells have been shown not to play a role in the regulation of hepcidin expression by iron *in vivo*^[97,98]. The data in

the literature regarding the involvement of Kupffer cells in the regulation of liver hepcidin expression by inflammation is contradictory^[97-99]. Montosi *et al*^[97] have reported a role for Kupffer cells in the regulation of hepcidin expression by inflammation in mice. On the other hand, Lou *et al*^[98] and Theurl *et al*^[99] have demonstrated that the depletion of Kupffer cells in mice does not abrogate the up-regulation of liver hepcidin expression by LPS.

The role of Kupffer cell activation and TNF- α signaling in the regulation of hepcidin expression by alcohol *in vivo* has also been reported^[100]. The inactivation of Kupffer cells by gadolinium chloride in rats paired with alcohol Lieber DeCarli diets for 6 wk or mice fed with ethanol in the drinking water for 1 wk did not reverse the alcohol-induced suppression of hepcidin expression in the liver^[100]. Moreover, similar results were obtained when Kupffer cells were depleted by liposomes containing clodronate^[100]. When phagocytosed by the Kupffer cells, clodronate released from the liposomes induces apoptosis and thereby depletes the Kupffer cell^[101]. When co-cultured, Kupffer cells have been suggested to exert a negative effect on hepcidin synthesis in hepatocytes^[99]. However, the depletion or inactivation of the Kupffer cells has been reported to not induce any significant changes in basal hepcidin expression levels in the livers of both control and alcohol-treated animals *in vivo*^[100]. Interestingly, one week of ethanol treatment (in the drinking water) was sufficient to induce NF- κ B activation and TNF- α and IL-6 release in mice, compared to control mice fed with plain water^[100]. The neutralization of TNF- α inhibited the activation of NF- κ B^[100]. However, neither the neutralization of TNF- α nor the absence of TNF- α receptor I and II expression altered the effect of alcohol on hepcidin expression (i.e. down-regulation)^[100]. These findings therefore strongly suggest that the activation of Kupffer cells and TNF- α signaling are not involved in the regulation of hepcidin expression by alcohol *in vivo*. These findings are also in agreement with Lou *et al*^[98], who demonstrated that Kupffer cells are not required for the regulation of liver hepcidin expression by LPS-induced inflammation.

The suppression of hepcidin synthesis in the liver by alcohol occurs very early (within days) following alcohol exposure and involves hepatocytes, but not Kupffer cells (Figure 2)^[12,100]. It is noteworthy that hepatocytes are the main site of hepcidin synthesis in the liver^[50]. Alcohol-induced oxidative stress in hepatocytes is one of the main mechanisms by which hepcidin expression in the liver is down-regulated by alcohol^[12]. The decrease in liver hepcidin synthesis leads to an increase in intestinal iron transport and liver iron content^[10-12]. Hence, alcohol dysregulates iron homeostasis by suppressing hepcidin expression in the liver. Although Kupffer cells are not involved in the initial stages of this process induced by alcohol, the increase in iron stores (due to low levels of hepcidin) will further activate Kupffer cells, and thereby lead to the release of proinflammatory cytokines. It is therefore possible that unlike LPS, the priming of Kupffer cells by iron^[70] occurs in later stages of ALD and exacerbates the inflammatory processes by facilitating further

release of TNF- α , and thereby contributing to liver injury.

ALCOHOL-INDUCED OXIDATIVE STRESS AND HEPCIDIN

Alcohol metabolism generates reactive oxygen species and lipid peroxidation products (malondialdehyde, 4-hydroxynonenal) during the oxidation of ethanol by alcohol dehydrogenase and cytochrome P4502E1 to form acetaldehyde. Serum thioredoxin levels are also an indicator of oxidative stress and they have been reported to be significantly higher in patients with ALD compared to healthy subjects^[102]. Due to its capacity to take part in the Fenton reaction as a transition metal, iron itself induces the generation of reactive oxygen species (superoxide, hydroxyl iron), which subsequently damage cellular membranes *via* lipid peroxidation^[3,103]. Iron and alcohol act as a deadly cocktail to exacerbate liver injury. Orally effective iron chelators have been reported to attenuate alcohol-induced hepatic lipid peroxidation in rats^[104].

Alcohol-induced oxidative stress is involved in the suppression of hepcidin promoter activity and hepcidin transcription in the liver *in vivo* by inhibiting the DNA-binding activity of the transcription factor, C/EBP α ^[12]. Both Kupffer cells and hepatocytes are involved in alcohol-induced oxidative stress. However, a role for Kupffer cells in the regulation of hepcidin expression by alcohol in the liver has been excluded (Figure 2)^[100]. This strongly suggests the involvement of hepatocytes in the regulation of hepcidin expression, and thereby iron metabolism by alcohol. Multiple pathways are involved in alcohol-induced oxidative stress in hepatocytes; redox state changes, the activation of CYP2E1 or NADPH oxidase and changes in mitochondrial functions^[105-109]. Alcohol metabolism, which is accompanied by oxidation and reduction reactions, causes an imbalance in the redox state of the cell by generating excess NADH (reduced nicotinamide adenine dinucleotide). CYP2E1 enzyme, present in liver microsomes, has a high redox potential and is also metabolically active in the absence of alcohol^[107]. It produces reactive oxygen species such as superoxide and hydrogen peroxide, which is increased by alcohol exposure leading to lipid peroxidation, oxidative stress and tissue injury. NADPH oxidase catalyzes the reduction of molecular oxygen to generate superoxide^[110,111]. The liver expresses both phagocytic and non-phagocytic isoforms of NADPH oxidase^[112,113]. Alcohol-induced liver pathology was not observed in mice with deficient NADPH oxidase activity suggesting a role for this enzyme in the progression of alcoholic liver disease^[109]. Mitochondria play a key role both in iron biogenesis and alcohol metabolism^[3,108,114]. Alcohol induces lesions in the proteins of the mitochondrial electron transport chain and decreases the rate of hepatic ATP synthesis^[114-116]. This induces the transfer of unpaired electrons to molecular oxygen and elevates the production of reactive oxygen species. The involvement of

these changes in the regulation of hepcidin expression in hepatocytes by alcohol needs further investigation (Figure 2).

Several factors including iron, viral infection, endotoxin and reactive oxygen species have been implicated to act as second hit factors in the progression of alcoholic liver disease. Of note, hepcidin expression in the liver is regulated by iron, alcohol, hepatitis C viral proteins, inflammation, hypoxia and oxidative stress^[10-12,48,100,117-120]. Nishina *et al*^[119] have recently shown that hepatitis C viral protein-mediated oxidative stress suppresses hepcidin transcription by altering the activity of the transcription factor, C/EBP α . These findings are in agreement with our previously published results demonstrating a role for alcohol-mediated oxidative stress in the inhibition of C/EBP α activity and hepcidin transcription^[11,12]. Hepcidin may therefore act as a potential second hit factor in the progression of alcoholic liver disease.

CONCLUSION

Patients with alcoholic liver disease frequently display elevated iron stores. Recent reports by several groups strongly suggest that this may be a regulated mechanism rather than iron simply leaking out of injured intestinal cells. Hepcidin, an iron regulatory hormone synthesized in hepatocytes, plays a key role in the regulation of iron metabolism. Alcohol-mediated oxidative stress suppresses hepcidin expression in the liver leading to increased intestinal iron uptake and liver iron storage. Reduced hepcidin expression may be one of the mechanisms leading to elevated iron stores following alcohol exposure. Hepcidin may therefore act as a second hit in the progression of ALD. Both hepatocytes and Kupffer cells play a role in ALD. However, Kupffer cells have been reported to not be involved in the regulation of liver hepcidin expression by both acute and chronic alcohol exposure. Hepatocytes are the main site of alcohol metabolism in the liver leading to the generation of reactive oxygen species and the depletion of antioxidants. Alcohol-induced oxidative stress in hepatocytes is therefore one of the mechanisms leading to the suppression of hepcidin synthesis and thereby to iron overload, which in turn acts as a secondary risk factor in ALD. Accordingly, iron has been shown to prime Kupffer cells leading to the release of TNF- α in experimental animal models of ALD. The disruption of hepcidin synthesis and thereby iron metabolism by alcohol may have clinical relevance. Hepcidin therefore holds promise as an early diagnostic marker and as a candidate for therapeutic strategies for ALD.

REFERENCES

- 1 Andrews NC. Forging a field: the golden age of iron biology. *Blood* 2008; **112**: 219-230
- 2 De Domenico I, McVey Ward D, Kaplan J. Regulation of iron acquisition and storage: consequences for iron-linked disorders. *Nat Rev Mol Cell Biol* 2008; **9**: 72-81
- 3 Aisen P, Enns C, Wessling-Resnick M. Chemistry and biology of eukaryotic iron metabolism. *Int J Biochem Cell Biol* 2001; **33**: 940-959

- 4 **Frazer DM**, Anderson GJ. The orchestration of body iron intake: how and where do enterocytes receive their cues? *Blood Cells Mol Dis* 2003; **30**: 288-297
- 5 **Gunshin H**, Fujiwara Y, Custodio AO, Drenzo C, Robine S, Andrews NC. Slc11a2 is required for intestinal iron absorption and erythropoiesis but dispensable in placenta and liver. *J Clin Invest* 2005; **115**: 1258-1266
- 6 **Donovan A**, Brownlie A, Zhou Y, Shepard J, Pratt SJ, Moynihan J, Paw BH, Drejer A, Barut B, Zapata A, Law TC, Brugnara C, Lux SE, Pinkus GS, Pinkus JL, Kingsley PD, Palis J, Fleming MD, Andrews NC, Zon LI. Positional cloning of zebrafish ferroportin1 identifies a conserved vertebrate iron exporter. *Nature* 2000; **403**: 776-781
- 7 **Donovan A**, Roy CN, Andrews NC. The ins and outs of iron homeostasis. *Physiology* (Bethesda) 2006; **21**: 115-123
- 8 **Nemeth E**, Ganz T. Regulation of iron metabolism by hepcidin. *Annu Rev Nutr* 2006; **26**: 323-342
- 9 **Nicolas G**, Viatte L, Bennoun M, Beaumont C, Kahn A, Vaulont S. Hepcidin, a new iron regulatory peptide. *Blood Cells Mol Dis* 2002; **29**: 327-335
- 10 **Harrison-Findik DD**. Role of alcohol in the regulation of iron metabolism. *World J Gastroenterol* 2007; **13**: 4925-4930
- 11 **Harrison-Findik DD**, Klein E, Crist C, Evans J, Timchenko N, Gollan J. Iron-mediated regulation of liver hepcidin expression in rats and mice is abolished by alcohol. *Hepatology* 2007; **46**: 1979-1985
- 12 **Harrison-Findik DD**, Schafer D, Klein E, Timchenko NA, Kulaksiz H, Clemens D, Fein E, Andriopoulos B, Pantopoulos K, Gollan J. Alcohol metabolism-mediated oxidative stress down-regulates hepcidin transcription and leads to increased duodenal iron transporter expression. *J Biol Chem* 2006; **281**: 22974-22982
- 13 **Bridle K**, Cheung TK, Murphy T, Walters M, Anderson G, Crawford DG, Fletcher LM. Hepcidin is down-regulated in alcoholic liver injury: implications for the pathogenesis of alcoholic liver disease. *Alcohol Clin Exp Res* 2006; **30**: 106-112
- 14 **Flanagan JM**, Peng H, Beutler E. Effects of alcohol consumption on iron metabolism in mice with hemochromatosis mutations. *Alcohol Clin Exp Res* 2007; **31**: 138-143
- 15 **Ohtake T**, Saito H, Hosoki Y, Inoue M, Miyoshi S, Suzuki Y, Fujimoto Y, Kohgo Y. Hepcidin is down-regulated in alcohol loading. *Alcohol Clin Exp Res* 2007; **31**: S2-S8
- 16 **Conrad ME**, Barton JC. Anemia and iron kinetics in alcoholism. *Semin Hematol* 1980; **17**: 149-163
- 17 **Friedman IM**, Kraemer HC, Mendoza FS, Hammer LD. Elevated serum iron concentration in adolescent alcohol users. *Am J Dis Child* 1988; **142**: 156-159
- 18 **Whitfield JB**, Zhu G, Heath AC, Powell LW, Martin NG. Effects of alcohol consumption on indices of iron stores and of iron stores on alcohol intake markers. *Alcohol Clin Exp Res* 2001; **25**: 1037-1045
- 19 **Valerio LG Jr**, Parks T, Petersen DR. Alcohol mediates increases in hepatic and serum nonheme iron stores in a rat model for alcohol-induced liver injury. *Alcohol Clin Exp Res* 1996; **20**: 1352-1361
- 20 **Kohgo Y**, Ohtake T, Ikuta K, Suzuki Y, Hosoki Y, Saito H, Kato J. Iron accumulation in alcoholic liver diseases. *Alcohol Clin Exp Res* 2005; **29**: 189S-193S
- 21 **Tavill AS**, Qadri AM. Alcohol and iron. *Semin Liver Dis* 2004; **24**: 317-325
- 22 **Eichner ER**. The hematologic disorders of alcoholism. *Am J Med* 1973; **54**: 621-630
- 23 **Kimber C**, Deller DJ, Ibbotson RN, Lander H. The mechanism of anaemia in chronic liver disease. *Q J Med* 1965; **34**: 33-64
- 24 **Paine CJ**, Eichner ER, Dickson V. Concordance of radioassay and microbiological assay in the study of the ethanol-induced fall in serum folate level. *Am J Med Sci* 1973; **266**: 134-138
- 25 **Lindenbaum J**, Lieber CS. Hematologic effects of alcohol in man in the absence of nutritional deficiency. *N Engl J Med* 1969; **281**: 333-338
- 26 **Lindenbaum J**, Roman MJ. Nutritional anemia in alcoholism. *Am J Clin Nutr* 1980; **33**: 2727-2735
- 27 **Lindenbaum J**. Folate and vitamin B12 deficiencies in alcoholism. *Semin Hematol* 1980; **17**: 119-129
- 28 **Eichner ER**, Buchanan B, Smith JW, Hillman RS. Variations in the hematologic and medical status of alcoholics. *Am J Med Sci* 1972; **263**: 35-42
- 29 **Hines JD**, Cowan DH. Studies on the pathogenesis of alcohol-induced sideroblastic bone-marrow abnormalities. *N Engl J Med* 1970; **283**: 441-446
- 30 **Sheehy TW**, Berman A. The anemia of cirrhosis. *J Lab Clin Med* 1960; **56**: 72-82
- 31 **Eichner ER**, Hillman RS. The evolution of anemia in alcoholic patients. *Am J Med* 1971; **50**: 218-232
- 32 **Waters AH**, Morley AA, Rankin JG. Effect of alcohol on haemopoiesis. *Br Med J* 1966; **2**: 1565-1568
- 33 **Pierce HI**, McGuffin RG, Hillman RS. Clinical studies in alcoholic sideroblastosis. *Arch Intern Med* 1976; **136**: 283-289
- 34 **Hallberg L**, Rossander L. Effect of different drinks on the absorption of non-heme iron from composite meals. *Hum Nutr Appl Nutr* 1982; **36**: 116-123
- 35 **Cook JD**, Reddy MB, Hurrell RF. The effect of red and white wines on nonheme-iron absorption in humans. *Am J Clin Nutr* 1995; **61**: 800-804
- 36 **Ioannou GN**, Dominitz JA, Weiss NS, Heagerty PJ, Kowdley KV. The effect of alcohol consumption on the prevalence of iron overload, iron deficiency, and iron deficiency anemia. *Gastroenterology* 2004; **126**: 1293-1301
- 37 **Milman N**, Kirchhoff M. Relationship between serum ferritin, alcohol intake, and social status in 2235 Danish men and women. *Ann Hematol* 1996; **72**: 145-151
- 38 **Gordeuk VR**, Diaz SF, Onojobi GO, Kasvosve I, Debebe Z, Edossa A, Pantin JM, Xiong S, Nekhai S, Nouraie M, Tsukamoto H, Taylor RE. Ferroportin Q248h, dietary iron, and serum ferritin in community African-Americans with low to high alcohol consumption. *Alcohol Clin Exp Res* 2008; **32**: 1947-1953
- 39 **Chapman RW**, Morgan MY, Laulicht M, Hoffbrand AV, Sherlock S. Hepatic iron stores and markers of iron overload in alcoholics and patients with idiopathic hemochromatosis. *Dig Dis Sci* 1982; **27**: 909-916
- 40 **Irving MG**, Halliday JW, Powell LW. Association between alcoholism and increased hepatic iron stores. *Alcohol Clin Exp Res* 1988; **12**: 7-13
- 41 **Duane P**, Raja KB, Simpson RJ, Peters TJ. Intestinal iron absorption in chronic alcoholics. *Alcohol Alcohol* 1992; **27**: 539-544
- 42 **Ganne-Carrie N**, Christidis C, Chastang C, Ziol M, Chapel F, Imbert-Bismut F, Trinchet JC, Guettier C, Beaugrand M. Liver iron is predictive of death in alcoholic cirrhosis: a multivariate study of 229 consecutive patients with alcoholic and/or hepatitis C virus cirrhosis: a prospective follow up study. *Gut* 2000; **46**: 277-282
- 43 **Sanchez J**, Casas M, Rama R. Effect of chronic ethanol administration on iron metabolism in the rat. *Eur J Haematol* 1988; **41**: 321-325
- 44 **Takada A**, Takase S, Tsutsumi M. Characteristic features of alcoholic liver disease in Japan: a review. *Gastroenterol Jpn* 1993; **28**: 137-148
- 45 **Bell H**, Skinningsrud A, Raknerud N, Try K. Serum ferritin and transferrin saturation in patients with chronic alcoholic and non-alcoholic liver diseases. *J Intern Med* 1994; **236**: 315-322
- 46 **Park CH**, Valore EV, Waring AJ, Ganz T. Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem* 2001; **276**: 7806-7810
- 47 **Krause A**, Neitz S, Magert HJ, Schulz A, Forssmann WG, Schulz-Knappe P, Adermann K. LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Lett* 2000; **480**: 147-150
- 48 **Nicolas G**, Bennoun M, Devaux I, Beaumont C, Grandchamp B, Kahn A, Vaulont S. Lack of hepcidin gene

- expression and severe tissue iron overload in upstream stimulatory factor 2 (USF2) knockout mice. *Proc Natl Acad Sci USA* 2001; **98**: 8780-8785
- 49 **Nicolas G**, Bennoun M, Porteu A, Mativet S, Beaumont C, Grandchamp B, Sirito M, Sawadogo M, Kahn A, Vaulont S. Severe iron deficiency anemia in transgenic mice expressing liver hepcidin. *Proc Natl Acad Sci USA* 2002; **99**: 4596-4601
 - 50 **Pigeon C**, Ilyin G, Courselaud B, Leroyer P, Turlin B, Brissot P, Loreal O. A new mouse liver-specific gene, encoding a protein homologous to human antimicrobial peptide hepcidin, is overexpressed during iron overload. *J Biol Chem* 2001; **276**: 7811-7819
 - 51 **Detivaud L**, Nemeth E, Boudjema K, Turlin B, Troade MB, Leroyer P, Ropert M, Jacquelinet S, Courselaud B, Ganz T, Brissot P, Loreal O. Hepcidin levels in humans are correlated with hepatic iron stores, hemoglobin levels, and hepatic function. *Blood* 2005; **106**: 746-748
 - 52 **Ganz T**. Hepcidin in iron metabolism. *Curr Opin Hematol* 2004; **11**: 251-254
 - 53 **Nemeth E**, Preza GC, Jung CL, Kaplan J, Waring AJ, Ganz T. The N-terminus of hepcidin is essential for its interaction with ferroportin: structure-function study. *Blood* 2006; **107**: 328-333
 - 54 **Nemeth E**, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, Ganz T, Kaplan J. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* 2004; **306**: 2090-2093
 - 55 **Nemeth E**, Roetto A, Garozzo G, Ganz T, Camaschella C. Hepcidin is decreased in TFR2 hemochromatosis. *Blood* 2005; **105**: 1803-1806
 - 56 **Wallace DF**, Summerville L, Subramaniam VN. Targeted disruption of the hepatic transferrin receptor 2 gene in mice leads to iron overload. *Gastroenterology* 2007; **132**: 301-310
 - 57 **Muckenthaler M**, Roy CN, Custodio AO, Minana B, deGraaf J, Montross LK, Andrews NC, Hentze MW. Regulatory defects in liver and intestine implicate abnormal hepcidin and Cybrd1 expression in mouse hemochromatosis. *Nat Genet* 2003; **34**: 102-107
 - 58 **Bridle KR**, Frazer DM, Wilkins SJ, Dixon JL, Purdie DM, Crawford DH, Subramaniam VN, Powell LW, Anderson GJ, Ramm GA. Disrupted hepcidin regulation in HFE-associated haemochromatosis and the liver as a regulator of body iron homeostasis. *Lancet* 2003; **361**: 669-673
 - 59 **Papanikolaou G**, Samuels ME, Ludwig EH, MacDonald ML, Franchini PL, Dube MP, Andres L, MacFarlane J, Sakellaropoulos N, Politou M, Nemeth E, Thompson J, Risler JK, Zaborowska C, Babakaiff R, Radomski CC, Pape TD, Davidas O, Christakis J, Brissot P, Lockitch G, Ganz T, Hayden MR, Goldberg YP. Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat Genet* 2004; **36**: 77-82
 - 60 **Babitt JL**, Huang FW, Wrighting DM, Xia Y, Sidis Y, Samad TA, Campagna JA, Chung RT, Schneyer AL, Woolf CJ, Andrews NC, Lin HY. Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat Genet* 2006; **38**: 531-539
 - 61 **Wang RH**, Li C, Xu X, Zheng Y, Xiao C, Zervas P, Cooperman S, Eckhaus M, Rouault T, Mishra L, Deng CX. A role of SMAD4 in iron metabolism through the positive regulation of hepcidin expression. *Cell Metab* 2005; **2**: 399-409
 - 62 **Fleming RE**, Ahmann JR, Migas MC, Waheed A, Koeffler HP, Kawabata H, Britton RS, Bacon BR, Sly WS. Targeted mutagenesis of the murine transferrin receptor-2 gene produces hemochromatosis. *Proc Natl Acad Sci USA* 2002; **99**: 10653-10658
 - 63 **Finberg KE**, Heeney MM, Campagna DR, Aydinok Y, Pearson HA, Hartman KR, Mayo MM, Samuel SM, Strouse JJ, Markianos K, Andrews NC, Fleming MD. Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA). *Nat Genet* 2008; **40**: 569-571
 - 64 **Du X**, She E, Gelbart T, Truksa J, Lee P, Xia Y, Khovananth K, Mudd S, Mann N, Moresco EM, Beutler E, Beutler B. The serine protease TMPRSS6 is required to sense iron deficiency. *Science* 2008; **320**: 1088-1092
 - 65 **Takei Y**, Arteel GE, Bergheim I, Lambert JC, McMullen MR, Nagy LE, Enomoto N, Sato N. Roles of Kupffer cells in alcoholic liver disease. *Alcohol Clin Exp Res* 2005; **29**: 1116-1120
 - 66 **Tsukamoto H**, Takei Y, McClain CJ, Joshi-Barve S, Hill D, Schmidt J, Deaciuc I, Barve S, Colell A, Garcia-Ruiz C, Kaplowitz N, Fernandez-Checa JC, Yokoyama H, Okamura Y, Nakamura Y, Ishii H, Chawla RK, Barve S, Joshi-Barve S, Watson W, Nelson W, Lin M, Ohata M, Motomura K, Enomoto N, Ikejima K, Kitamura T, Oide H, Hirose M, Bradford BU, Rivera CA, Kono H, Peter S, Yamashina S, Konno A, Ishikawa M, Shimizu H, Sato N, Thurman R. How is the liver primed or sensitized for alcoholic liver disease? *Alcohol Clin Exp Res* 2001; **25**: 171S-181S
 - 67 **Wheeler MD**. Endotoxin and Kupffer cell activation in alcoholic liver disease. *Alcohol Res Health* 2003; **27**: 300-306
 - 68 **Adachi Y**, Bradford BU, Gao W, Bojes HK, Thurman RG. Inactivation of Kupffer cells prevents early alcohol-induced liver injury. *Hepatology* 1994; **20**: 453-460
 - 69 **Koop DR**, Klopfenstein B, Iimuro Y, Thurman RG. Gadolinium chloride blocks alcohol-dependent liver toxicity in rats treated chronically with intragastric alcohol despite the induction of CYP2E1. *Mol Pharmacol* 1997; **51**: 944-950
 - 70 **Tsukamoto H**, Lin M, Ohata M, Giulivi C, French SW, Brittenham G. Iron primes hepatic macrophages for NF-kappaB activation in alcoholic liver injury. *Am J Physiol* 1999; **277**: G1240-G1250
 - 71 **Xiong S**, She H, Tsukamoto H. Signaling role of iron in NF-kappa B activation in hepatic macrophages. *Comp Hepatol* 2004; **3** Suppl 1: S36
 - 72 **Knutson M**, Wessling-Resnick M. Iron metabolism in the reticuloendothelial system. *Crit Rev Biochem Mol Biol* 2003; **38**: 61-88
 - 73 **Gunshin H**, Mackenzie B, Berger UV, Gunshin Y, Romero MF, Boron WF, Nussberger S, Gollan JL, Hediger MA. Cloning and characterization of a mammalian proton-coupled metal-ion transporter. *Nature* 1997; **388**: 482-488
 - 74 **Vidal SM**, Pinner E, Lepage P, Gauthier S, Gros P. Natural resistance to intracellular infections: Nramp1 encodes a membrane phosphoglycoprotein absent in macrophages from susceptible (Nramp1 D169) mouse strains. *J Immunol* 1996; **157**: 3559-3568
 - 75 **Kristiansen M**, Graversen JH, Jacobsen C, Sonne O, Hoffman HJ, Law SK, Moestrup SK. Identification of the haemoglobin scavenger receptor. *Nature* 2001; **409**: 198-201
 - 76 **Suzuki Y**, Saito H, Suzuki M, Hosoki Y, Sakurai S, Fujimoto Y, Kohgo Y. Up-regulation of transferrin receptor expression in hepatocytes by habitual alcohol drinking is implicated in hepatic iron overload in alcoholic liver disease. *Alcohol Clin Exp Res* 2002; **26**: 26S-31S
 - 77 **Xiong S**, She H, Zhang AS, Wang J, Mkrtchyan H, Dynnyk A, Gordeuk VR, French SW, Enns CA, Tsukamoto H. Hepatic macrophage iron aggravates experimental alcoholic steatohepatitis. *Am J Physiol Gastrointest Liver Physiol* 2008; **295**: G512-G521
 - 78 **Finch CA**, Deubelbeiss K, Cook JD, Eschbach JW, Harker LA, Funk DD, Marsaglia G, Hillman RS, Slichter S, Adamson JW, Ganzoni A, Biblett ER. Ferrokinetics in man. *Medicine* (Baltimore) 1970; **49**: 17-53
 - 79 **Levy JE**, Jin O, Fujiwara Y, Kuo F, Andrews NC. Transferrin receptor is necessary for development of erythrocytes and the nervous system. *Nat Genet* 1999; **21**: 396-399
 - 80 **Levy JE**, Montross LK, Cohen DE, Fleming MD, Andrews NC. The C282Y mutation causing hereditary hemochromatosis does not produce a null allele. *Blood* 1999; **94**: 9-11
 - 81 **Zhou XY**, Tomatsu S, Fleming RE, Parkkila S, Waheed A, Jiang J, Fei Y, Brunt EM, Ruddy DA, Prass CE, Schatzman RC, O'Neill R, Britton RS, Bacon BR, Sly WS. HFE gene knockout produces mouse model of hereditary

- hemochromatosis. *Proc Natl Acad Sci USA* 1998; **95**: 2492-2497
- 82 **Piperno A**, Girelli D, Nemeth E, Trombini P, Bozzini C, Poggiali E, Phung Y, Ganz T, Camaschella C. Blunted hepcidin response to oral iron challenge in HFE-related hemochromatosis. *Blood* 2007; **110**: 4096-4100
 - 83 **Knutson MD**, Vafa MR, Haile DJ, Wessling-Resnick M. Iron loading and erythrophagocytosis increase ferroportin 1 (FPN1) expression in J774 macrophages. *Blood* 2003; **102**: 4191-4197
 - 84 **Andriopoulos B**, Pantopoulos K. Hepcidin generated by hepatoma cells inhibits iron export from co-cultured THP1 monocytes. *J Hepatol* 2006; **44**: 1125-1131
 - 85 **Harris ZL**, Durley AP, Man TK, Gitlin JD. Targeted gene disruption reveals an essential role for ceruloplasmin in cellular iron efflux. *Proc Natl Acad Sci USA* 1999; **96**: 10812-10817
 - 86 **Ragan HA**, Nacht S, Lee GR, Bishop CR, Cartwright GE. Effect of ceruloplasmin on plasma iron in copper-deficient swine. *Am J Physiol* 1969; **217**: 1320-1323
 - 87 **Sarkar J**, Seshadri V, Tripoulas NA, Ketterer ME, Fox PL. Role of ceruloplasmin in macrophage iron efflux during hypoxia. *J Biol Chem* 2003; **278**: 44018-44024
 - 88 **Bird GL**, Sheron N, Goka AK, Alexander GJ, Williams RS. Increased plasma tumor necrosis factor in severe alcoholic hepatitis. *Ann Intern Med* 1990; **112**: 917-920
 - 89 **Iimuro Y**, Gallucci RM, Luster MI, Kono H, Thurman RG. Antibodies to tumor necrosis factor alpha attenuate hepatic necrosis and inflammation caused by chronic exposure to ethanol in the rat. *Hepatology* 1997; **26**: 1530-1537
 - 90 **Yin M**, Wheeler MD, Kono H, Bradford BU, Gallucci RM, Luster MI, Thurman RG. Essential role of tumor necrosis factor alpha in alcohol-induced liver injury in mice. *Gastroenterology* 1999; **117**: 942-952
 - 91 **She H**, Xiong S, Lin M, Zandi E, Giulivi C, Tsukamoto H. Iron activates NF-kappaB in Kupffer cells. *Am J Physiol Gastrointest Liver Physiol* 2002; **283**: G719-G726
 - 92 **Telfer JF**, Brock JH. Proinflammatory cytokines increase iron uptake into human monocytes and synovial fibroblasts from patients with rheumatoid arthritis. *Med Sci Monit* 2004; **10**: BR91-BR95
 - 93 **Olynyk JK**, Clarke SL. Iron overload impairs pro-inflammatory cytokine responses by Kupffer cells. *J Gastroenterol Hepatol* 2001; **16**: 438-444
 - 94 **Kakhlon O**, Cabantchik ZI. The labile iron pool: characterization, measurement, and participation in cellular processes(1). *Free Radic Biol Med* 2002; **33**: 1037-1046
 - 95 **Funk F**, Lenders JP, Crichton RR, Schneider W. Reductive mobilisation of ferritin iron. *Eur J Biochem* 1985; **152**: 167-172
 - 96 **Thomas CE**, Aust SD. Release of iron from ferritin by cardiotoxic anthracycline antibiotics. *Arch Biochem Biophys* 1986; **248**: 684-689
 - 97 **Montosi G**, Corradini E, Garuti C, Barelli S, Recalcatti S, Cairo G, Valli L, Pignatti E, Vecchi C, Ferrara F, Pietrangelo A. Kupffer cells and macrophages are not required for hepatic hepcidin activation during iron overload. *Hepatology* 2005; **41**: 545-552
 - 98 **Lou DQ**, Lesbordes JC, Nicolas G, Viatte L, Bennoun M, Van Rooijen N, Kahn A, Renia L, Vaulont S. Iron- and inflammation-induced hepcidin gene expression in mice is not mediated by Kupffer cells in vivo. *Hepatology* 2005; **41**: 1056-1064
 - 99 **Theurl M**, Theurl I, Hochegger K, Obrist P, Subramaniam N, van Rooijen N, Schuemann K, Weiss G. Kupffer cells modulate iron homeostasis in mice via regulation of hepcidin expression. *J Mol Med* 2008; **86**: 825-835
 - 100 **Harrison-Findik DD**, Klein E, Evans J, Gollan J. Regulation of liver hepcidin expression by alcohol in vivo does not involve Kupffer cell activation or TNF-alpha signaling. *Am J Physiol Gastrointest Liver Physiol* 2009; **296**: G112-G118
 - 101 **Van Rooijen N**, Sanders A. Liposome mediated depletion of macrophages: mechanism of action, preparation of liposomes and applications. *J Immunol Methods* 1994; **174**: 83-93
 - 102 **Sumida Y**, Nakashima T, Yoh T, Kakisaka Y, Nakajima Y, Ishikawa H, Mitsuyoshi H, Okanoue T, Nakamura H, Yodoi J. Serum thioredoxin elucidates the significance of serum ferritin as a marker of oxidative stress in chronic liver diseases. *Liver* 2001; **21**: 295-299
 - 103 **Winterbourn CC**. Toxicity of iron and hydrogen peroxide: the Fenton reaction. *Toxicol Lett* 1995; **82-83**: 969-974
 - 104 **Sadrzadeh SM**, Nanji AA, Price PL. The oral iron chelator, 1,2-dimethyl-3-hydroxypyrid-4-one reduces hepatic-free iron, lipid peroxidation and fat accumulation in chronically ethanol-fed rats. *J Pharmacol Exp Ther* 1994; **269**: 632-636
 - 105 **Arteel GE**. Oxidants and antioxidants in alcohol-induced liver disease. *Gastroenterology* 2003; **124**: 778-790
 - 106 **Molina PE**, Hoek JB, Nelson S, Guidot DM, Lang CH, Wands JR, Crawford JM. Mechanisms of alcohol-induced tissue injury. *Alcohol Clin Exp Res* 2003; **27**: 563-575
 - 107 **Cederbaum AI**. Iron and CYP2E1-dependent oxidative stress and toxicity. *Alcohol* 2003; **30**: 115-120
 - 108 **Hoek JB**, Cahill A, Pastorino JG. Alcohol and mitochondria: a dysfunctional relationship. *Gastroenterology* 2002; **122**: 2049-2063
 - 109 **Kono H**, Rusyn I, Yin M, Gabele E, Yamashina S, Dikalova A, Kadiiska MB, Connor HD, Mason RP, Segal BH, Bradford BU, Holland SM, Thurman RG. NADPH oxidase-derived free radicals are key oxidants in alcohol-induced liver disease. *J Clin Invest* 2000; **106**: 867-872
 - 110 **Rotrosen D**, Yeung CL, Leto TL, Malech HL, Kwong CH. Cytochrome b558: the flavin-binding component of the phagocyte NADPH oxidase. *Science* 1992; **256**: 1459-1462
 - 111 **Chanock SJ**, el Benna J, Smith RM, Babior BM. The respiratory burst oxidase. *J Biol Chem* 1994; **269**: 24519-24522
 - 112 **Reinehr R**, Becker S, Eberle A, Grether-Beck S, Haussinger D. Involvement of NADPH oxidase isoforms and Src family kinases in CD95-dependent hepatocyte apoptosis. *J Biol Chem* 2005; **280**: 27179-27194
 - 113 **Cheng G**, Cao Z, Xu X, van Meir EG, Lambeth JD. Homologs of gp91phox: cloning and tissue expression of Nox3, Nox4, and Nox5. *Gene* 2001; **269**: 131-140
 - 114 **Bailey SM**, Cunningham CC. Contribution of mitochondria to oxidative stress associated with alcoholic liver disease. *Free Radic Biol Med* 2002; **32**: 11-16
 - 115 **Cahill A**, Cunningham CC, Adachi M, Ishii H, Bailey SM, Fromenty B, Davies A. Effects of alcohol and oxidative stress on liver pathology: the role of the mitochondrion. *Alcohol Clin Exp Res* 2002; **26**: 907-915
 - 116 **Cunningham CC**, Bailey SM. Ethanol consumption and liver mitochondria function. *Biol Signals Recept* 2001; **10**: 271-282
 - 117 **Nemeth E**, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, Ganz T. IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest* 2004; **113**: 1271-1276
 - 118 **Lee P**, Peng H, Gelbart T, Wang L, Beutler E. Regulation of hepcidin transcription by interleukin-1 and interleukin-6. *Proc Natl Acad Sci USA* 2005; **102**: 1906-1910
 - 119 **Nishina S**, Hino K, Korenaga M, Vecchi C, Pietrangelo A, Mizukami Y, Furutani T, Sakai A, Okuda M, Hidaka I, Okita K, Sakaida I. Hepatitis C virus-induced reactive oxygen species raise hepatic iron level in mice by reducing hepcidin transcription. *Gastroenterology* 2008; **134**: 226-238
 - 120 **Nicolas G**, Chauvet C, Viatte L, Danan JL, Bigard X, Devaux I, Beaumont C, Kahn A, Vaulont S. The gene encoding the iron regulatory peptide hepcidin is regulated by anemia, hypoxia, and inflammation. *J Clin Invest* 2002; **110**: 1037-1044