

• BRIEF REPORTS •

# Effects of endostatin on expression of vascular endothelial growth factor and its receptors and neovascularization in colonic carcinoma implanted in nude mice

Yun-He Jia, Xin-Shu Dong, Xi-Shan Wang

**Yun-He Jia, Xin-Shu Dong, Xi-Shan Wang**, Department of Abdominal Surgery, Tumor Hospital of Harbin Medical University, Harbin 150040, Heilongjiang Province, China

**Supported by** the Key Technologies Research and Development Program of Heilongjiang Province During the 9th Five-Year Plan Period, No. G99C19-5

**Correspondence to:** Dr. Yun-He Jia, Department of General Surgery of Nanjing Jinling Hospital, Nanjing 210002, Jiangsu Province, China. jyhcruise@0451.com

**Telephone:** +86-25-80860034

**Received:** 2003-12-23 **Accepted:** 2004-02-01

## Abstract

**AIM:** To investigate the antiangiogenic effects of endostatin on colonic carcinoma cell line implanted in nude mice and its mechanism.

**METHODS:** Nude mice underwent subcutaneous injection with LS-174t colonic carcinoma cell line to generate carcinoma and were randomly separated into two groups. Mice received injection of vehicle or endostatin every day for two weeks. After the tumor was harvested, the tumor volumes were determined, and the expressions of CD34, VEGF and Flk-1 were examined by immunohistochemical method.

**RESULTS:** Tumor volume was significantly inhibited in the endostatin group (84.17%) and tumor weight was significantly inhibited in the endostatin group ( $0.197 \pm 0.049$ ) compared to the control group ( $1.198 \pm 0.105$ ) ( $F = 22.56$ ,  $P = 0.001$ ), microvessel density (MVD) was significantly decreased in the treated group ( $31.857 \pm 3.515$ ) compared to the control group ( $100.143 \pm 4.290$ ) ( $F = 151.62$ ,  $P < 0.001$ ). Furthermore, the expression of Flk-1 was significantly inhibited in the treated group (34.29%) compared to the control group (8.57%) ( $\chi^2 = 13.745$ ,  $P = 0.001$ ). However no significant decrease was observed in the expression of vascular endothelial growth factor (VEGF) between these two groups ( $\chi^2 = 0.119$ ,  $P = 0.730$ ).

**CONCLUSION:** Endostatin can inhibit tumor growth and angiogenesis by blocking Vegf/Flk-1 pathway. This experiment provides the theory basis for developing a new anti-carcinoma drug through studying the properties of anti-angiogenesis inhibitors.

Jia YH, Dong XS, Wang XS. Effects of endostatin on expression of vascular endothelial growth factor and its receptors and neovascularization in colonic carcinoma implanted in nude mice. *World J Gastroenterol* 2004; 10(22): 3361-3364  
<http://www.wjgnet.com/1007-9327/10/3361.asp>

## INTRODUCTION

Angiogenesis is the process of sprouting of capillaries from

preexisting blood vessels. The overall process is a complex one that involves many biological functions and cell types. Previous researches since 1970s have demonstrated that tumor angiogenesis is required for the growth and metastasis of primary solid tumor. Endothelial cell activation, migration, and proliferation are major cellular events in this process. All of these processes are under the tight regulation of factors that either “promote” or “inhibit” angiogenesis. When the balance of these factors is disturbed, angiogenic factors can be released from tumor cells, such as vascular endothelial growth factor (VEGF)<sup>[1]</sup> and basic fibroblast growth factor (bFGF), and migrate to the nearby blood vessel endothelia, signal the activation of the angiogenic response. Tumor cells also produce anti-angiogenic factors including angiostatin, endostatin. Administration of endostatin in tumor-bearing mice has been shown to keep the primary tumor in a dormant state<sup>[2]</sup>. But the mechanisms are still unclear. One mechanism is that endostatin in combination with the endothelial cells to form a compound with tyrosine activity and thereby inducing apoptosis of endothelial cells. The other is that endostatin functions as a ligand for the integrin family of adhesion receptors on the surface of endothelial cells and is associated with MMP family<sup>[3]</sup>. So, in order to explore the mechanism of endostatin, we provided data that endostatin showed a broad antitumor effect, which might be a result of its inhibitory mechanism on tumor angiogenesis.

## MATERIALS AND METHODS

### Materials

Endostatin donated from Hefei Sunny Biology-technology Institute, was a 20 ku C-terminal fragment of collagen X VIII. Human endostatin was cloned and expressed in *Escherichia coli* and purified by gene cloning technology by Gene Med Ltd.

Colon cancer cell line HL-174T was purchased from Shanghai Cellular Research Institute. The cells were cultured in 1640 medium supplemented with 100 mL/L fetal bovine serum at 37 °C in 50 mL/L CO<sub>2</sub> and the culture media were changed twice a week and observed in the invert microscope to assay the reproduction situation. All reagents and media for cell culture were obtained from GIBCO.

### Methods

**Subcutaneous xenograft models** Tumor cells were implanted ( $0.5 \times 10^6$  cells/animal) subcutaneously using a 27-gauge needle in the back region of BALB/C-nu/nu female mice of 6-8 wk old. Tumor volume was measured every 3 d using a slide gauge. Tumor volumes were measured as the product of (length × width) ×  $\pi/6$ . Mice were fed with a standard rodent diet for two weeks, and 14 of 20 mice with a similar tumor volume were selected and divided into two groups randomly. Animals of each group were treated once daily with a 0.1 mL (2 mg/mL) ip bolus injection of endostatin in saline or saline alone for 14 d. Upon termination of the efficacy portion of endostatin (3 d after drug withdrawal), the animals were euthanized, and the tumor was harvested from the animals and the volume was weighted. The tumors were submitted for immunohistochemical test.

**Evaluation of MVD, VEGF and VEGFR/flk-1 expression in tumors** Five- $\mu$ m paraffin embedded sections of the tumors were evaluated by immunohistochemistry. The sections were stained with anti-CD34 antibody (DAKO, 1:2 000) and polyclonal anti-VEGF antibody and polyclonal anti-Flk-1 antibody that could recognize VEGF isoforms and their receptors, following avidin-biotin complex methods.

Microvessel density was assessed by light microscopy in areas of invasive tumors containing the highest number of capillaries and small venules per area (neovascular "hot point"). Areas of the highest neovascularization were found by scanning the tumor sections at a low magnification ( $\times 100$ ) and identified with the greatest number of distinct CD34 stained (brown) microvessels per area. Necrotic areas within tumors, where microvessels were sparse, were not included in the vessel counts. After the area of the highest neovascularization was identified, individual microvessels were counted under  $\times 400$  field. A brown-stained endothelial cell or a endothelial cell cluster clearly separated from adjacent microvessels, tumor cells, and other connective-tissue elements, considered a single, countable microvessel. Vessel lumens were not necessary for a structure to be defined as a microvessel, and red cells were not used to define a vessel lumen. Results were expressed as the number of microvessels identified within 10 fields ( $\times 400$ ).

VEGF was assessed by light microscopy ( $\times 100$ ) in the areas of invasive tumor cells. VEGF expression was determined on adjacent serial visual fields and scored by a pathologic doctor. The number of positively and negatively stained cells was counted in 10 continuous visual fields. If more than 5% cells were brown stained, they were considered as positive, otherwise negative. The results were expressed as mean $\pm$ SD.

Flk-1 was assessed by light microscopy ( $\times 400$ ) in the areas of microvessel endothelial cells. The number of endothelial cells stained positively was counted in 10 continuous visual fields. Over 5% positively stained endothelial cells were determined as positive and else negative. The results were expressed as mean $\pm$ SD.

### Statistics

The analysis of variance test was used to compare the differences in tumor weight and MVD between the treated and control groups. The Chi-squared test was used to compare the differences in the expression of VEGF or VEGFR2/Flk-1 between the treated and control groups. The difference was considered statistically significant if the *P* value was less than 0.05. All the data were analyzed with SAS 6.22 by the statistical office of the Second Military Medical University.

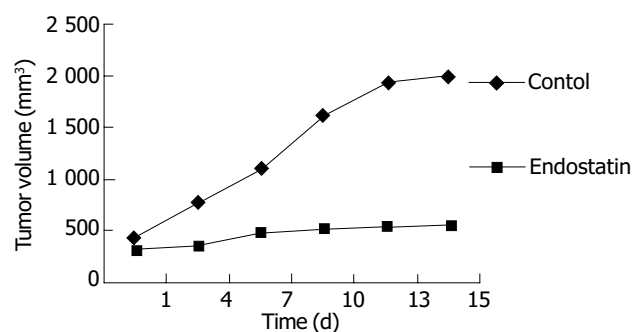
## RESULTS

### Inhibition of tumor growth by endostatin

To determine the efficacy of endostatin on tumor growth, the volume of tumors were measured every three days after endostatin was used and tumors were weighed immediately after they were harvested. The tumor growth in the mice treated with endostatin was inhibited by 84.17% in volume. The growth of the tumors was stopped 3-5 d after using endostatin except two of them. The growth of primary colon tumor was significantly inhibited by endostatin (Figure 1). The tumor weight of the treated group was significantly less than that of the control group ( $F = 22.56$ ;  $P = 0.0005$ ) (Table 1).

### Effects of endostatin on angiogenesis in primary tumors

Immunohistochemical analysis showed a significant inhibition of angiogenesis in the tumors treated with endostatin ( $P < 0.05$ ) (Table 1). The MVD of tumors was significantly different ( $F = 151.62$ ,  $P = 0.001$ ) between the two groups, the treated group had a significant lower MVD.



**Figure 1** Changes in volume of tumors transplanted in dorsal skinfold chambers in nude mice.

**Table 1** Reduction in tumor mass and MVD after endostatin treatment (mean $\pm$ SD)

	Control	Endostatin	<i>F</i>	<i>P</i>
Mass	1.198 $\pm$ 0.105	0.197 $\pm$ 0.049	22.56	<sup>a</sup> <i>P</i> = 0.0005
MVD	100.143 $\pm$ 4.290	31.857 $\pm$ 3.515	151.62	<sup>b</sup> <i>P</i> = 0.0001

<sup>a</sup>*P* = Mass difference between Control group and Endostatin group, <sup>b</sup>*P* = MVD difference between Control group and Endostatin group.

**Table 2** Expression of VEGF and VEGFR2/Flk-1 in tumor after Endostatin treatment

	Positive	Negative		$\chi^2$	<i>P</i>
Control-VEGF (1)	65	5	(1) vs (2)	0.119	0.730
Endostatin-VEGF (2)	66	4			
Control-Flk-1 (3)	64	6	(3) vs (4)	13.745	<sup>a</sup> <i>P</i> = 0.001
Endostatin-Flk-1 (4)	46	24			

<sup>a</sup>*P* = Flk-1 difference between Control group and Endostatin group.

### Selectivity and potency of endostatin on VEGF and Flk-1

Immunohistochemical analysis showed that endostatin had a potent inhibition on Flk-1 expression ( $\chi^2 = 13.745$ ,  $P = 0.001$ ) but no inhibition on VEGF expression in tumor cells (Table 2). There was a significant difference in the tumors of treated mice and control mice after staining of endothelial cells with Flk-1 monoclonal antibody (Table 2) but no difference in the tumor tissue stained with VEGF between the two groups (Table 2).

## DISCUSSION

Angiogenesis is a fundamental process by which new blood vessels are formed. It is essential in reproduction, development, and wound repair. Under these conditions, angiogenesis is highly regulated, turned on for brief periods and then completely inhibited. However some diseases are driven by persistently unregulated angiogenesis. Tumor growth is one of these kinds of diseases. Tumor growth and metastasis are angiogenesis-dependent. A tumor must continuously stimulate the growth of new capillary blood vessels for the tumor itself to grow. MVD was significantly high in colon cancer tissues compared with normal colon tissues<sup>[4]</sup>. Tumor cells get oxygen and nutrition through angiogenesis. The Phenomena were also observed in other cancers, for example, breast carcinoma<sup>[5]</sup> lung cancer<sup>[6]</sup> prostate cancer<sup>[7]</sup> and pancreatic tumor<sup>[8]</sup>, etc.

Endostatin, a 20-kD C-terminal cleavage product of collagen XVIII, was originally identified by O'Reilly *et al.*<sup>[2]</sup> as a tumor-derived, highly active, and endothelial specific angiogenic inhibitor<sup>[9]</sup>. Recombinant endostatin has been shown to inhibit

the growth of a wide variety of tumors in mice, with no toxic side effects observed. Importantly, tumors treated with several cycles of endostatin did not develop drug resistance and become dormant, which persisted even when the endostatin therapy was discontinued<sup>[10]</sup>. In our experiment, we found that the tumor growth slowed down or even stopped three days after usage of endostatin, but when the treatment was stopped the tumor began to grow rapidly. So endostatin must be used continuously to prevent tumor growth. It is difficult for the patients to accept the treatment in their whole lifetime, so further research is necessary for the usage of endostatin. At present, the molecular mechanism of endostatin action remains unknown. There are some theories which are supported by various studies.

Our results showed that endostatin blocked the VEGF-VEGFR/Flk-1 pathway to inhibit the angiogenesis of colon tumor in nude mice. VEGF pathway is the only well-defined signaling pathway known to be required for the normal development of vasculature as well as for the pathologic angiogenesis that accompanies cancer and other disease states<sup>[11,12]</sup>. The VEGF family of growth factors comprises at least five members, i.e., VEGF, placenta growth factor (PGF), VEGF-B, VEGF-C<sup>[13]</sup>, and VEGF-D. The VEGF pathway is initiated when VEGF binds to its receptors in endothelial cells. The three best characterized VEGF receptors are termed VEGF receptor 1 (VEGFR1/Flt-1) and VEGF receptor 2 (VEGFR2/Flk-1/KDR) and VEGF receptor 3 (VEGFR3/Flk-4). VEGFR1 and VEGFR2 are highly related transmembrane tyrosine kinases that use their ectodomains to bind to VEGF<sup>[14,15]</sup>. These binding in turn, activate the intrinsic tyrosine kinase activity of their cytodomains, initiating intracellular signaling. VEGFR3 is expressed almost exclusively by the lymphatic endothelium, thus being considered as a major regulator of lymphangiogenesis<sup>[16]</sup>. The blockade of VEGF by using a soluble VEGFR-3 extracellular domain could inhibit tumor lymphangiogenesis<sup>[17]</sup>. While PlGF could bind selectively to VEGFR1, VEGF-C and VEGF-D could bind to both VEGFR3<sup>[18,19]</sup> and VEGFR2. The corresponding receptor(s) for VEGF-B is VEGFR1. VEGF/VEGFR2 pathway is the most important for tumor angiogenesis, growth and metastasis. In our studies, we found VEGFR2 was obviously down-regulated in the endostatin group compared to the control group, but the expression of VEGF was not changed in both groups. So we think that endostatin can block the VEGF/VEGFR2 pathway by binding to VEGFR2, thus inhibiting the neovascularization.

Endostatin could function as a ligand for the integrin family of adhesion receptors on the surface of endothelial cells<sup>[20]</sup>. Integrins are potential targets for endostatin function and they are important in endothelial cell biology and angiogenesis. The endostatin-integrin interaction is of functional significance *in vitro*, and the immobilized endostatin supports endothelial cell survival and migration in an integrin-dependent manner. Soluble endostatin in turn could inhibit integrin-dependent endothelial cell functions, such as cell migration<sup>[20]</sup>.

Matrix metalloproteinases (MMPs), a family of extracellular and membrane-associated endopeptidases, collectively digest almost all extracellular matrix and basement membrane components, thus playing an important role in tumor progression. Endostatin could prevent the fragmentation of pro-MMP-2 that is associated with reduction of catalytic activity. Endostatin had no effect on MMP-8 as shown by collagenase activity assays<sup>[3]</sup>. Endostatin could block the activation and activities of certain tumor-associated pro-MMPs, such as pro-MMP-2, -9, and -13, which may explain at least in part, the antitumor effect of endostatin<sup>[3,21]</sup>.

Inhibition of angiogenesis has been shown to be an effective strategy in cancer therapy in mice. However, its widespread application has been hampered by difficulties in the large-scale production of antiangiogenic proteins. This limitation may be resolved by *in vivo* delivery and expression of antiangiogenic

genes. A recombinant adenovirus that could express murine endostatin that was biologically active *in vitro* as determined in endothelial cell proliferation assays, and *in vivo* by suppression of angiogenesis induced by VEGF has been constructed<sup>[22]</sup>. Persistent high serum levels of endostatin were achieved after systemic administration of the vector to nude mice, which resulted in a significant reduction of the growth rates and the volumes of metastatic brain tumor<sup>[23]</sup>. If the method can be used safely in clinic, it can effectively resolve the recurrence and overgrowth of tumors during the rest periods. Different targets have been found and effective results were achieved in gene therapy. An adenoviral vector carrying Tie2 gene, an endostatin-specific receptor tyrosine kinase, was constructed and tested in established primary tumor, a murine mammary carcinoma or a murine melanoma, which could significantly inhibit the growth rate of both murine mammary carcinoma and melanoma by 64% and 47%, respectively. So the potential of vector-mediated antiangiogenic gene therapy as a component is a very novel and effective strategy in cancer therapy<sup>[24,25]</sup>.

Synergy between endostatin and chemotherapy<sup>[26]</sup> can be used to eradicate spontaneous tumor and metastases of colon cancer or other angiogenesis dependent diseases. Tumor necrosis was demonstrated only in animals receiving the combination therapy, but not when each agent was applied as monotherapy. The results suggested that these synergistic treatment modalities might provide a novel and effective tool for future therapies of metastatic cancer. Synergy between endostatin and interventional therapy is another way to inhibit the neovascularization. Endostatin can be sent directly to the microvessels around the tumor and destroy the tunica intima of vessels to starve the tumor to death. Other strategies also were used to block the VEGF/VEGFR pathway to inhibit the tumor growth. VEGF-trap is a potent blocker against the angiogenesis of the tumor and could result in stunted and almost completely avascular tumors. VEGF-Trap-mediated blockade may be a hopeful way for tumor treatment<sup>[27]</sup>. More and more attention has been paid to the effectiveness and carrier and medication methods are paid more and more attention throughout the world. Endostatin may provide a novel and effective tool for future therapies of cancer<sup>[28,29]</sup>.

## REFERENCES

- 1 Potti A, Moazzam N, Tendulkar K, Javed NA, Koch M, Kargas S. Immunohistochemical determination of vascular endothelial growth factor (VEGF) overexpression in malignant melanoma. *Anticancer Res* 2003; **23**: 4023-4026
- 2 O'Reilly MS, Boehm T, Shing Y, Fukai N, Vasios G, Lane WS, Flynn E, Birkhead JR, Olsen BR, Folkman J. Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 1997; **24**: 277-285
- 3 Nyberg P, Heikkilä P, Sorsa T, Luostarinen J, Heljasvaara R, Stenman UH, Pihlajaniemi T, Salo T. Endostatin inhibits human tongue carcinoma cell invasion and intravasation and blocks the activation of matrix metalloproteinase-2, -9, and -13. *J Biol Chem* 2003; **278**: 22404-22411
- 4 Dkhissi F, Lu H, Soria C, Opolon P, Griscelli F, Liu H, Khattar P, Mishal Z, Perricaudet M, Li H. Endostatin exhibits a direct antitumor effect in addition to its antiangiogenic activity in colon cancer cells. *Hum Gene Ther* 2003; **14**: 997-1008
- 5 Atiqur Rahman M, Toi M. Anti-angiogenic therapy in breast cancer. *Biomed Pharmacother* 2003; **57**: 463-470
- 6 Sridhar SS, Shepherd FA. Targeting angiogenesis: a review of angiogenesis inhibitors in the treatment of lung cancer. *Lung Cancer* 2003; **42**(2 Suppl): S81-S91
- 7 Uehara H. Angiogenesis of prostate cancer and antiangiogenic therapy. *J Med Invest* 2003; **50**: 146-153
- 8 Schuch G, Kisker O, Atala A, Soker S. Pancreatic tumor growth is regulated by the balance between positive and negative modulators of angiogenesis. *Angiogenesis* 2002; **5**: 181-190
- 9 Sim BK. Angiostatin and endostatin: endothelial cell-specific

- endogenous inhibitors of angiogenesis and tumor growth. *Angiogenesis* 1998; **2**: 37-48
- 10 **Kuroiwa M**, Takeuchi T, Lee JH, Yoshizawa J, Hirato J, Kaneko S, Choi SH, Suzuki N, Ikeda H, Tsuchida Y. Continuous versus intermittent administration of human endostatin in xenografted human neuroblastoma. *J Pediatr Surg* 2003; **38**: 1499-1505
- 11 **Zachary I**. VEGF signalling: integration and multi-tasking in endothelial cell biology. *Biochem Soc Trans* 2003; **31**(Pt 6): 1171-1177
- 12 **Qi L**, Robinson WA, Brady BM, Glode LM. Migration and invasion of human prostate cancer cells is related to expression of VEGF and its receptors. *Anticancer Res* 2003; **23**: 3917-3922
- 13 **Karkkainen MJ**, Haiko P, Sainio K, Partanen J, Taipale J, Petrova TV, Jeltsch M, Jackson DG, Talikka M, Rauvala H, Betsholtz C, Alitalo K. Vascular endothelial growth factor C is required for sprouting of the first lymphatic vessels from embryonic veins. *Nat Immunol* 2004; **5**: 74-80
- 14 **Dales JP**, Garcia S, Bonnier P, Duffaud F, Carpentier S, Djemli A, Ramuz O, Andrac L, Lavaut M, Allasia C, Charpin C. Prognostic significance of VEGF receptors, VEGFR-1 (Flt-1) and VEGFR-2 (KDR/Flk-1) in breast carcinoma. *Ann Pathol* 2003; **23**: 297-305
- 15 **Endo A**, Fukuhara S, Masuda M, Ohmori T, Mochizuki N. Selective inhibition of vascular endothelial growth factor receptor-2 (VEGFR-2) identifies a central role for VEGFR-2 in human aortic endothelial cell responses to VEGF. *J Recept Signal Transduct Res* 2003; **23**: 239-254
- 16 **Pimenta FJ**, Sa AR, Gomez RS. Lymphangiogenesis in human dental pulp. *Int Endod J* 2003; **36**: 853-856
- 17 **Kaipainen A**, Korhonen J, Mustonen T, van Hinsbergh VW, Fang GH, Dumont D, Breitman M, Alitalo K. Expression of the fms-like tyrosine kinase 4 gene becomes restricted to lymphatic endothelium during development PNAS. *Proc Natl Acad Sci U S A* 1995; **92**: 3566-3570
- 18 **Nakamura Y**, Yasuoka H, Tsujimoto M, Yang Q, Imabun S, Nakahara M, Nakao K, Nakamura M, Mori I, Kakudo K. Flt-4-positive vessel density correlates with vascular endothelial growth factor-d expression, nodal status, and prognosis in breast cancer. *Clin Cancer Res* 2003; **9**: 5313-5317
- 19 **Van Trappen PO**, Steele D, Lowe DG, Baithun S, Beasley N, Thiele W, Weich H, Krishnan J, Shepherd JH, Pepper MS, Jackson DG, Sleeman JP, Jacobs IJ. Expression of vascular endothelial growth factor (VEGF)-C and VEGF-D, and their receptor VEGFR-3, during different stages of cervical carcinogenesis. *J Pathol* 2003; **201**: 544-554
- 20 **Rehn M**, Veikkola T, Kukkk-Valdre E, Nakamura H, Ilmonen M, Lombardo CR, Pihlajaniemi T, Alitalo K, Vuori K. Interaction of endostatin with integrins implicated in angiogenesis. *Proc Natl Acad Sci U S A* 2001; **98**: 1024-1029
- 21 **Guan KP**, Ye HY, Yan Z, Wang Y, Hou SK. Serum levels of endostatin and matrix metalloproteinase-9 associated with high stage and grade primary transitional cell carcinoma of the bladder. *Urology* 2003; **61**: 719-723
- 22 **Sauter BV**, Martinet O, Zhang WJ, Mandeli J, Woo SLC. Adenovirus-mediated gene transfer of endostatin *in vivo* results in high level of transgene expression and inhibition of tumor growth and metastases PNAS. *Proc Natl Acad Sci U S A* 2000; **97**: 4802-4807
- 23 **Oga M**, Takenaga K, Sato Y, Nakajima H, Koshikawa N, Osato K, Sakiyama S. Inhibition of metastatic brain tumor growth by intramuscular administration of the endostatin gene. *Int J Oncol* 2003; **23**: 73-79
- 24 **Chen W**, Fu J, Liu Q, Ruan C, Xiao S. Retroviral endostatin gene transfer inhibits human colon cancer cell growth *in vivo*. *Chin Med J* 2003; **116**: 1582-1584
- 25 **Sridhar SS**, Shepherd FA. Targeting angiogenesis: a review of angiogenesis inhibitors in the treatment of lung cancer. *Lung Cancer* 2003; **42**(2 Suppl): S81-S91
- 26 **Plum SM**, Hanson AD, Volker KM, Vu HA, Sim BK, Fogler WE, Fortier AH. Synergistic activity of recombinant human endostatin in combination with adriamycin: analysis of *in vitro* activity on endothelial cells and *in vivo* tumor progression in an orthotopic murine mammary carcinoma model. *Clin Cancer Res* 2003; **9**: 4619-4626
- 27 **Byrne AT**, Ross L, Holash J, Nakanishi M, Hu L, Hofmann JJ, Yancopoulos GD, Jaffe RB. Vascular endothelial growth factor-trap decreases tumor burden, inhibits ascites, and causes dramatic vascular remodeling in an ovarian cancer model. *Clin Cancer Res* 2003; **9**: 5721-5728
- 28 **Shibuya M**. VEGF-receptor inhibitors for anti-angiogenesis. *Nippon Yakurigaku Zasshi* 2003; **122**: 498-503
- 29 **Tee D**, DiStefano J 3rd. Simulation of tumor-induced angiogenesis and its response to anti-angiogenic drug treatment: mode of drug delivery and clearance rate dependencies. *J Cancer Res Clin Oncol* 2004; **130**: 15-24

Edited by Kumar M and Wang XL Proofread by Xu FM