

Bile salts inhibit growth and induce apoptosis of human esophageal cancer cell line

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Abstract

AIM: To explore the effect of six bile salts, including glycocholate (GC), glycochenodeoxycholate (GCDC), glycodeoxycholate (GDC), taurocholate (TC), taurochenodeoxycholate (TCDC), taurodeoxycholate (TDC), and two bile acids including cholic acid (CA) and deoxycholic acid (DCA) on esophageal cancer Eca109 cell line.

METHODS: Eca109 cells were exposed to six bile salts, two bile acids and the mixed bile salts at different concentrations for 24-72 h. 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium bromide (MTT) assay was used to detect the cell proliferation. Apoptotic morphology was observed by phase-contrast video microscopy and deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay. Sub-G1 DNA fragmentations and early apoptosis cells were assayed by flow cytometry (FCM) with propidium iodide (PI) staining and annexin V-FITC conjugated with PI staining. Apoptosis DNA ladders on agarose were observed. Activation of caspase-3 was assayed by FCM with FITC-conjugated monoclonal rabbit anti-active caspase-3 antibody and expressions of Bcl-2 and Bax proteins were examined immunocytochemically in 500 $\mu\text{mol/L}$ -TC-induced apoptosis cells.

RESULTS: Five bile salts except for GC, and two bile acids and the mixed bile salts could initiate growth inhibition of Eca109 cells in a dose- and time-dependent manner. TUNEL, FCM, and DNA ladder assays all demonstrated apoptosis induced by bile salts and bile acids at 500 $\mu\text{mol/L}$, except for GC. Early apoptosis cell percentages in Eca109 cells treated with GCDC, GDC, TC, TCDC, TDC, CA at 500 $\mu\text{mol/L}$ for 12 h, DCA at 500 $\mu\text{mol/L}$ for 6 h, and mixed bile salts at 1 000 $\mu\text{mol/L}$ for 12 h were 7.5%, 8.7%, 14.8%, 8.9%, 7.8%, 9.3%, 22.6% and 12.5%, respectively, all were significantly higher than that in

control (1.9%). About 22% of the cell population treated with TC at 500 $\mu\text{mol/L}$ for 24 h had detectable active caspase-3, and were higher than that in the control (1%). Immunocytochemical assay suggested that TC down-regulated Bcl-2 protein level and up-regulated Bax protein level.

CONCLUSION: GCDC, GDC, TC, TCDC, TDC, CA and DCA, except for GC, can inhibit growth and induce apoptosis of esophageal cancer Eca109 cells. Activation of caspase-3, decreased Bcl-2 protein and increased Bax protein are involved in TC-induced apoptosis of Eca109 cells.

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Key words: Bile salts; Esophageal cancer cells; Proliferation; Apoptosis

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INTRODUCTION

Duodeno-gastro-esophageal reflux or bile reflux is involved in the pathogenesis of reflux esophagitis, Barrett's esophagus (BE), and esophageal adenocarcinoma. Recent studies have shown that bile acids and bile salts present frequently in the refluxate of BE patients, and influence the development and persistence of metaplasia^[1-3]. Bile salts can enter mucosal cells in their non-ionized lipophilic form, cause injuries to cell membranes and tight junctions, and cell necrosis^[4]. However, studies have shown that different bile salts induce cell apoptosis^[5-7]. Whether different bile salts induce apoptosis of esophageal cancer cells remains unknown. Therefore, it is necessary to identify the effect of bile salts on esophageal cancer cells.

Glycine-conjugated bile acids such as glycocholic (GC) and glycochenodeoxycholic acids (GCDC) induce hepatocyte apoptosis *in vitro*^[8], whereas taurine-conjugated bile acids such as taurocholic (TC) and taurochenodeoxycholic acids (TCDC) are well tolerated by hepatocytes^[9,10]. Cell apoptosis is specifically induced by hydrophobic bile salts such as deoxycholate (DC) and dihydroxy bile salt, whereas conjugated bile salt glycodeoxycholate (GDC) and trihydroxy bile salt (TDC) and its conjugate glycocholate (GC) do not induce apoptosis in colorectal cancer cell lines^[6]. Caspase cleavage and activation occur as early as 30 min after the

addition of DC to cells^[6]. In a recent report, direct oligomerization of the Fas receptor (CD95/Apo-1) has been suggested as the primary causative mechanism of bile salt-mediated hepatocyte apoptosis^[11-13]. However, CD95 is not involved in DC-induced apoptosis, but bile salt-mediated induction of mitochondrial permeability transition has been seen in colorectal cancer cell lines^[6,14].

In the present study, we observed whether six bile salts and two bile acids could inhibit cell proliferation and induce apoptosis of human esophageal cancer Eca109 cells. We characterized the kinetics of bile salt-induced apoptosis of esophageal cancer cells. The difference in effects of different bile salts and bile acids on esophageal cancer cells was demonstrated. Activation of caspase-3 and expression of Bcl-2 and Bax proteins, as the key components in mitochondrial apoptosis "death-signal transduction" pathway^[15-18], were described in bile salt-mediated apoptotic esophageal cancer cells.

MATERIALS AND METHODS

Materials

DMEM was purchased from Hyclone and trypsin was from Sigma. Fetal bovine serum (FBS) was from Hangzhou Jiangbin Biotechnology Co., Ltd. GC, GCDC, GDC, TC, TCDC, TDC, CA, and DCA were from Sigma. SP HistostainTM-Plus kits and deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) kits were purchased from Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd. Mouse mAbs to Bcl-2 and Bax were from Antibody Diagnostica Inc. Annexin V-FITC apoptosis detection kit I and FITC-conjugated monoclonal active caspase-3 antibody apoptosis kit I were from BD Biosciences.

Cell lines

Eca109 cell lines were obtained from the Fourth Military Medical University. Cells were cultured in DMEM plus 10% FBS. All cell lines were cultured under standard tissue culture conditions.

Methods

Assessment of cell proliferation MTT assay was conducted to determine the cell proliferation^[19]. Eight bile salts at concentrations of 500, 250, and 50 $\mu\text{mol/L}$, and mixed bile salts at concentrations of 1 500, 1 000, 500, and 250 $\mu\text{mol/L}$, including GC, GCDC, GDC, TC, TCDC, and TDC at the ratio of 2:2:1:2:2:1, were observed in MTT assay.

Eca109 cells were seeded in a 96-well plate (4 to 6×10^4 cells/well). After 24 h of post seeding, cells were treated with different bile salts for 3 d, and untreated cells served as a control. Prior to the determination, 10 μL of the 2.5 g/L stock solution of MTT was added to each well. After 4 h of incubation, the culture media were discarded, and then 100 μL of DMSO was added to each well and vibrated for 10 min. The absorbance (A) was measured at 492 nm with a microplate reader. The percentage of viable cells was calculated as follows: (A of experimental group/ A of control group) $\times 100\%$.

TUNEL assay

Cell apoptosis was analyzed using *in situ* cell death detection

kit based on the terminal TUNEL technique. Cells were grown on chamber glass culture slides and treated with bile salts. To avoid disadherence of apoptotic cells, the slides were coated with poly-lysine. In brief, after the cells were treated with or without bile salts for the indicated time, they were fixed overnight in 100 g/L formaldehyde, and then treated with proteinase K and H_2O_2 , labeled with dUTP in a humid box at 37 $^\circ\text{C}$ for 1 h. The cells without addition of TdT enzyme were used as negative control.

Flow cytometry by PI staining

Cells from the medium supernatant and adherent cells treated with bile salts were collected and pelleted at 1 200 U/min. The harvested cells were fixed with 1 mL of 75% cold ethanol at -20 $^\circ\text{C}$ for a night, and then washed with PBS. Cell pellets were incubated with 10 $\mu\text{g/mL}$ RNase and stained with 50 $\mu\text{g/mL}$ propidium iodide (PI) for 30 min in the dark. Samples were analyzed by FACSCalibur FCM at excitation wavelength of 488 nm. In the assay, 10 000 cells were detected by FCM. The resulting histograms were analyzed by the program CELLQuest.

Flow cytometry by annexin V-FITC conjugated with PI staining^[20]

Cells from the medium supernatant and adherent cells treated with bile salts were collected and pelleted at 1 200 U/min. Pellets were washed twice with cold PBS and then resuspended in a binding buffer at a concentration of 1×10^6 cells/mL, and 100 μL of the solution (1×10^5 cells) was transferred to each of two 5-mL culture tubes. Five microliters of annexin V-FITC and 5 μL of PI were added into each 100 μL solution, the cells were gently vortexed and incubated for 15 min at RT in the dark. Four hundred microliters of 1st binding buffer was added to the sample and analyzed by FACSCalibur FCM within 1 h.

DNA fragmentation

A total of 10^6 - 10^7 cells treated with bile salts were collected and low molecular weight DNA was extracted using cell apoptosis DNA ladder extraction kit (Beijing Dingguo Biotechnology Co., Ltd). Standard loading buffer was added, 20 μL DNA samples was run on an 1% agarose gel containing 0.1% ethidium bromide in TAE buffer (Tris 40 mmol/L, sodium acetate 20 mmol/L, and 1 mmol/L EDTA, pH 8.0).

Caspase-3 assay

Caspase-3 was analyzed by FCM using FITC-conjugated monoclonal active caspase-3 antibody apoptosis kit I. Cells were collected and washed with cold PBS, then resuspended in Cytofix/CytopermTM solution at a concentration of 1×10^6 cells/0.5 mL and incubated for 20 min on ice. Samples were washed twice with Perm/washTM buffer, and then FITC-conjugated monoclonal rabbit anti-active caspase-3 antibody was added and incubated for 30 min at room temperature. The marked samples were analyzed by FACSCalibur FCM.

Immunocytochemical assay of Bcl-2 and Bax

Cells were grown on chamber glass culture slides and treated with TC at 500 $\mu\text{mol/L}$. To avoid disadherence of apoptotic cells, the slides were coated with poly-lysine. Slides with cells

were fixed in 65% cold acetone for 20 min. Bcl-2 and Bax were stained with the mAb using standard immunocytochemical techniques. Positive-staining-area percentages were calculated using a cell image analysis system (Qwin550CW). A mean of 20 adjacent fields, at a magnification of $\times 400$, was analyzed for each section^[21].

Statistical analysis

All values were expressed as mean \pm SD. Statistical differences between means were calculated by Student's *t*-test and χ^2 test. $P < 0.05$ was considered statistically significant.

RESULTS

Morphological changes

The influence of different bile salts on the apoptosis of Eca109 cells was established. Morphological criteria of cell apoptosis, such as membrane blebbing, cell shrinkage, nuclear condensation and fragmentation, were assessed by phase-contrast video microscopy. Morphological changes were observed 3 h after the addition of DCA at 500 $\mu\text{mol/L}$, and 24 h after the addition of the other bile salts at 500 $\mu\text{mol/L}$, except for GC.

Inhibition of Eca109 cell proliferation by some bile salts

The experimental Eca109 cells were treated with eight kinds of bile salts and mixed bile salts at different concentrations as described above. The cell viability was determined by MTT assay (Figure 1). Except for GC, the other bile salts or bile acids inhibited the growth of esophageal cancer cells in a dose- and time-dependent manner. Cell growth was suppressed by 85% after 24 h treatment with DCA at 500 $\mu\text{mol/L}$, higher than that after treatment with other bile salts and bile acid at the same concentration ($P < 0.01$). Cell growth was suppressed by 65% and 95% after 24- and 72-h treatment with mixed bile salts at 1 000 $\mu\text{mol/L}$, by 59% and 94% after 24- and 72-h treatment with TC at 500 $\mu\text{mol/L}$, which were higher than those after treatment with the other five kinds of bile salts.

TUNEL assay

TUNEL assay revealed that bile salts, except for GC, induced apoptosis of Eca109 cells. When treated with GC, GCDC, GDC, TC, TCDC, TDC, CA at 500 $\mu\text{mol/L}$ for 24 h, DCA at 500 $\mu\text{mol/L}$ for 12 h, and mixed bile salts at 100 $\mu\text{mol/L}$ for 24 h, the positive-cell percentages were 2%, 11%, 9%, 25%, 11%, 12%, 15%, 43% and 30%, respectively. Compared to 2% in control, the percentages were higher in the cells treated with bile salts ($P < 0.05$) except for GC. Figure 2 shows the image of cells treated with 500 $\mu\text{mol/L}$ TC for 24 h.

FCM by PI staining

Eca109 cells treated with bile salts and bile acids at 500 $\mu\text{mol/L}$ and mixed bile salts at 1 000 $\mu\text{mol/L}$ were stained with PI and analyzed by FCM. Except for GC, the bile salts induced cell apoptosis in a time-dependent manner (Figure 3A). When treated with DCA at 500 $\mu\text{mol/L}$, the apoptotic cell percentage reached 44.9% after 12 h and 80.5% after 24 h (Figure 3B). After 24 and 72 h, the percentages were 27.7% and 72.4%

after being treated with mixed bile salts at 1 000 $\mu\text{mol/L}$, and 19.7% and 75.5% after being treated with TC at 500 $\mu\text{mol/L}$, respectively.

FCM by annexin V-FITC conjugated with PI staining

Early apoptotic cells were annexin V-FITC-positive and PI-negative^[20]. Early apoptotic cells increased in the cells treated with bile salts except for GC. The percentages of early apoptotic cells of Eca109 cells treated with GC, GCDC, GDC, TC, TCDC, TDC, CA at 500 $\mu\text{mol/L}$ for 12 h, DCA at 500 $\mu\text{mol/L}$ for 6 h, and mixed bile salts at 100 $\mu\text{mol/L}$ for 12 h were 1.9%, 7.5%, 8.7%, 14.8%, 8.9%, 7.8%, 9.3%, 22.6%, and 12.5%, respectively. Some of them are shown in Figure 4. Compared to 1.9% in control, the percentages were higher in the cells treated with bile salts ($P < 0.05$) except for GC.

DNA fragmentation

In this study, we observed DNA ladder formation in cells treated with GC, TC, and DCA at 500 $\mu\text{mol/L}$ for 24 h, and mixed bile salts at 1 000 $\mu\text{mol/L}$ for 24 and 48 h (Figure 5).

Caspase-3 assay

We analyzed active caspase-3 in Eca109 cells treated with TC and GC at 500 $\mu\text{mol/L}$ for 24 h by FCM with FITC-conjugated monoclonal active caspase-3 antibody (Figure 6). In control and cells treated with GC at 500 $\mu\text{mol/L}$, 1% was found to be primarily active caspase-3 cells, and there was no difference between the two groups ($P > 0.05$). But about 22% of the cells treated with TC at 500 $\mu\text{mol/L}$ for 24 h had detectable active caspase-3, being higher than that in the control ($P < 0.01$).

Expressions of Bcl-2 and Bax proteins

Figure 7 shows the morphologic images of cells with immunocytochemical staining of Bcl-2 and Bax proteins. Bcl-2 protein staining was weaker than Bax because the positive staining was weaker. Expression of Bcl-2 protein was reduced in 500- $\mu\text{mol/L}$ -TC-treated Eca109 cells, while expression of Bax protein was increased (Table 1), suggesting that TC could down regulate Bcl-2 protein level and up-regulate Bax protein level.

Table 1 TC's effects on the expression of Bcl-2 and Bax in Eca109 cells (mean \pm SD)

Positive-area-percentage (%)	Bcl-2	Bax
Control	45 \pm 3	22 \pm 4
500 $\mu\text{mol/L}$ TC, 24 h	26 \pm 4 ^a	47 \pm 6 ^b
500 $\mu\text{mol/L}$ TC, 48 h	24 \pm 5 ^a	71 \pm 4 ^b

^a $P < 0.05$, ^b $P < 0.01$ vs control.

DISCUSSION

Bile salts and bile acids are likely to influence the development and persistence of esophageal mucosal cell metaplasia^[1-3]. It is necessary to investigate the effect of bile salts on esophageal mucosal cancer cells. In our studies, we have demonstrated

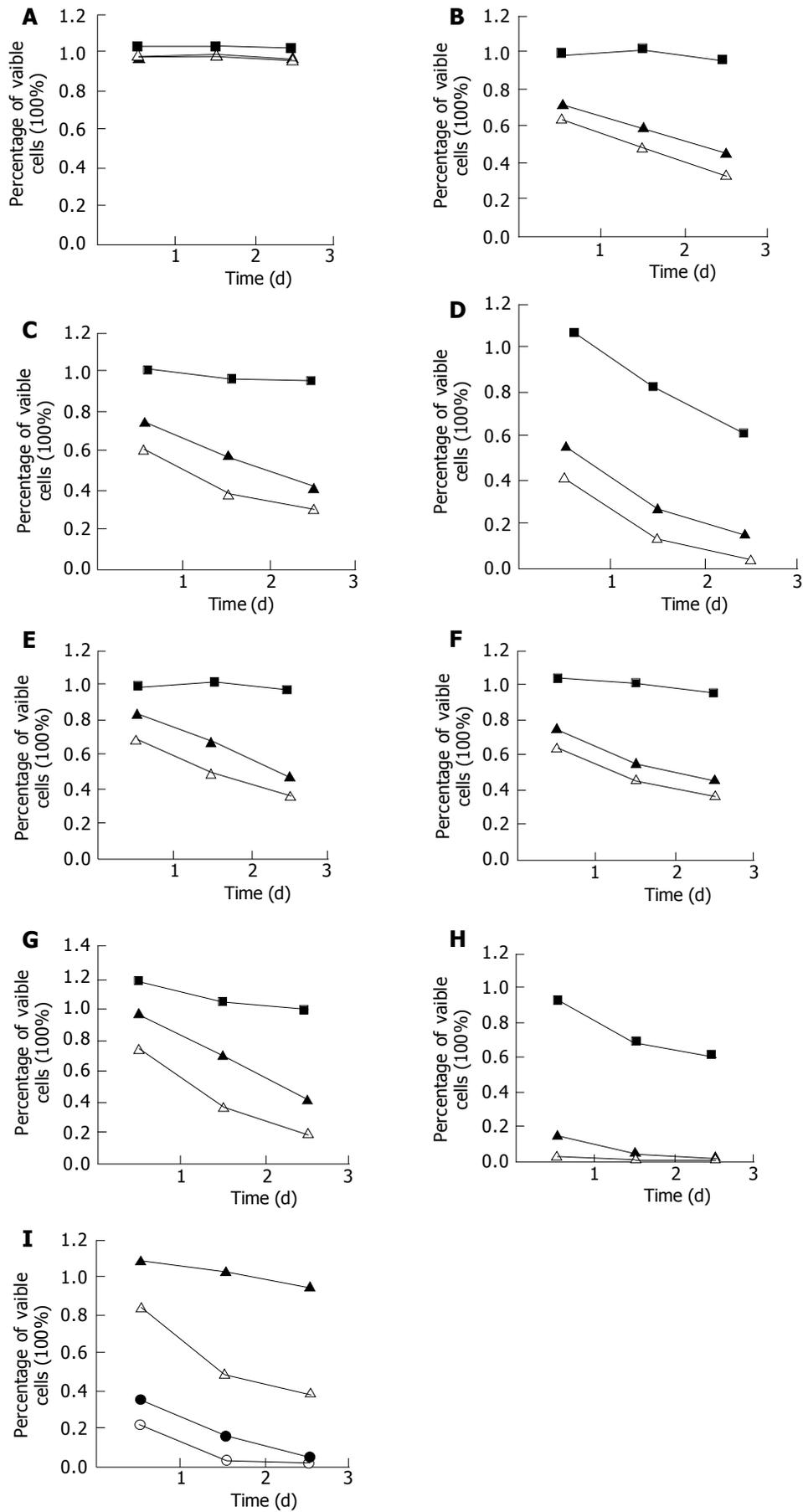


Figure 1 Effect of bile salts on Eca109 cell growth. ■ (at 50 μmol/L), ▲ (at 250 μmol/L), △ (at 500 μmol/L), ● (1 000 μmol/L), ○ (1 500 μmol/L). A: Glycocholate; B: Glycochenodeoxycholate; C: Glycodeoxycholate; D:

Taurocholate; E: Taurochenodeoxychoate; F: Taurodeoxycholate; G: Cholic acid; H: Deoxycholic acid; I: Mixed bile salts.

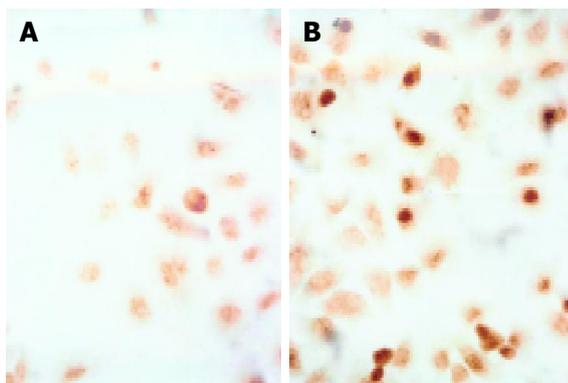


Figure 2 TC-induced apoptosis of control (A) and cells treated with TC (B) ×400.

that specific bile salts and bile acids at 50-500 μmol/L could inhibit growth and induce apoptosis of Eca109 cells.

Bile acids constitute 67% of normal bile, and are

conjugated with glycine or taurine in normal human bile. Bile acids consisted of 40% CA, 40% CDCA, and 20% DCA. Lithocholic acid (LCA) and ursodeoxycholic acid (UDCA) is less than 5% respectively^[22]. We selected GC, GCDC, GDC, TC, TCDC, and TDC, and CA and DCA in this study. The concentrations of bile salts in our study were from 25 to 500 μmol/L and those of mixed bile salts were 50 to 1 500 μmol/L, because the total bile acid concentration in esophagus of patients suffering from bile reflux is 0.89 mg/mL^[23]. Six bile salts at concentration of less than 500 μmol/L are all water-soluble sodium salts in pH 7.1-7.5. Because of its hydrophobicity, the largest portion of DCA is not soluble in water, but a considerable amount of 700 μmol/L DCA is detectable in fecal water^[24,25]. As this portion, at least theoretically, can get into contact with colonic epithelial cells, we selected the concentration at 25-500 μmol/L to investigate the effect of DCA on esophageal cancer cells. Our study demonstrated that less than 500 μmol/L DCA and CA could be dissolved thoroughly

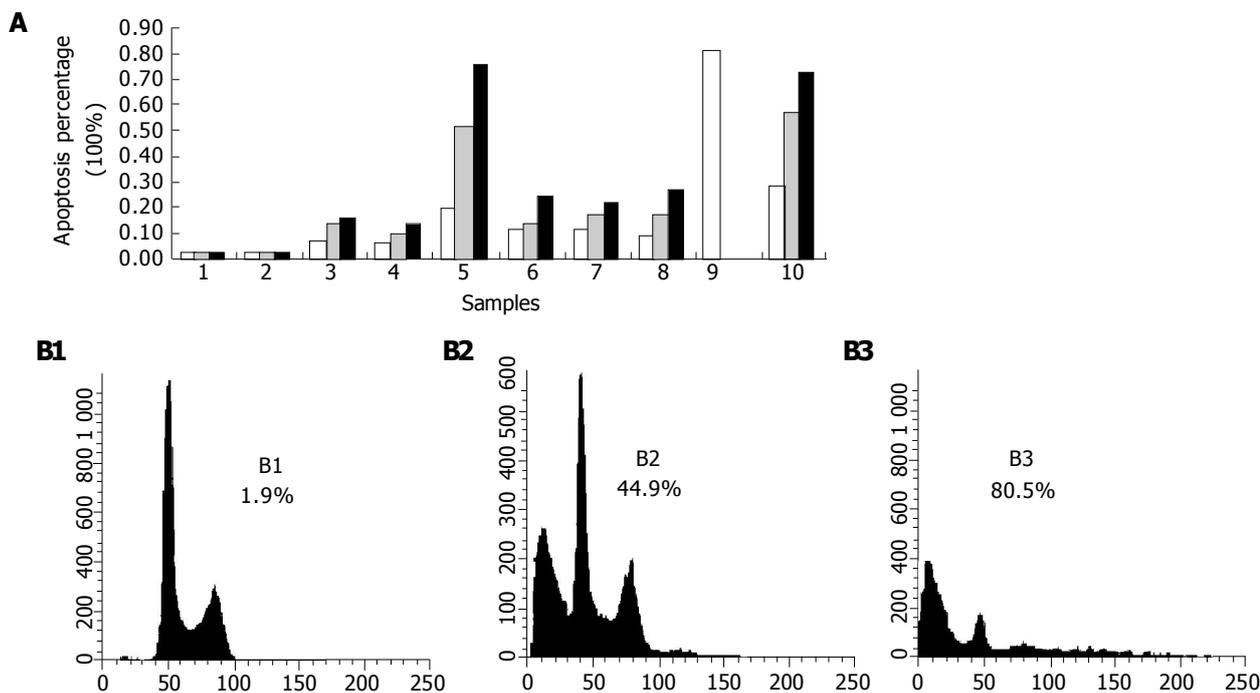


Figure 3 Time-dependent apoptosis of Eca109 cells induced by bile salts or bile acids (A) and DCA (B1-3). 1: Control; 2: GC; 3: GCDC; 4: GDC; 5: TC; 6: TCDC; 7: TDC; 8: CA; 9: DCA; 10: mixed bile salts. ■ (after 24 h), ▒ (after 48 h), □ (after 72 h).

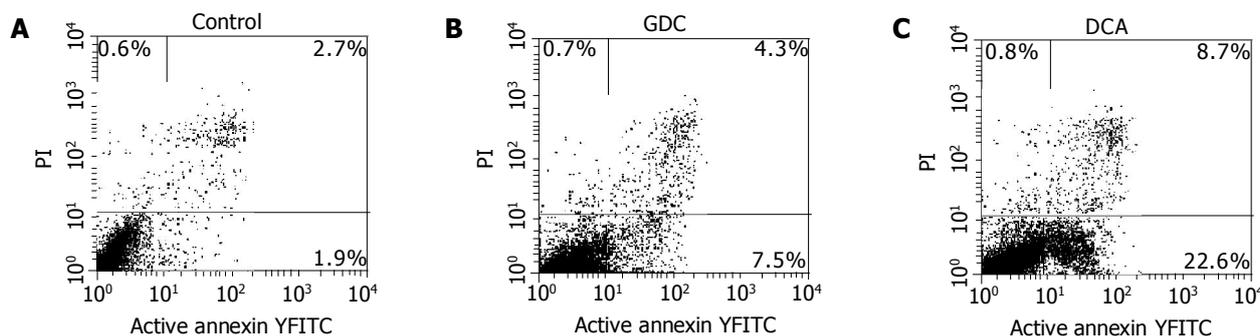


Figure 4 Apoptosis of control (A) and cells treated with GCDC (B) and DCA (C).

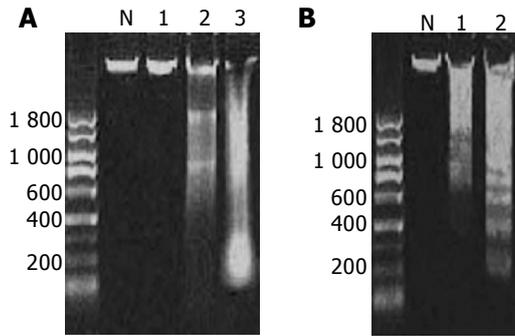


Figure 5 DNA ladder formation in cells treated with TC (A, Lane 2) and DCA (A, Lane 3) and mixed bile salts (B, Lane 1 and 2).

in the media. Based on the components of normal bile, the mixed bile salts in our study comprised GC, GCDC, GDC, TC, TCDC, and TDC at 2:2:1:2:2:1.

The present study demonstrated that five bile salts, two bile acids and the mixed bile salts except for GC, could inhibit the growth of human esophageal cancer Eca109 cells in a dose- and time-dependent manner. Our results also showed that the induced-cell death was a major reason for the cell growth inhibition. Necrosis is the major effect of bile salts on esophageal mucosal cells, by entering mucosal cells and causing injuries to cell membranes and tight junctions^[4]. However, the effects of five bile salts, two bile

acids and the mixed bile salts except for GC, on apoptosis-inducing activity in esophageal cancer cells were examined in our study. They caused typical apoptotic alterations including morphological changes assessed by phase-contrast video microscopy and TUNEL assay, positive annexin V staining by FCM, apoptotic sub-G₁ peak by FCM with PI staining, and DNA fragmentation by gel electrophoresis.

Seven bile salts or acids and mixed bile salts have different effects between Eca109 cells and hepatocytes^[8-10] and colon cancer cell lines^[6]. They could induce apoptosis of esophageal cancer cells in a dose- and time-dependent manner. Conversely, hepatocyte-cytotoxic glycine-conjugated bile acids such as GC induce hepatocyte apoptosis *in vitro*^[8], but not apoptosis of esophageal cancer cells, whereas hepatocyte-tolerated TC and TCDC^[9] induce apoptosis of esophageal cancer cells. Our results also showed that TC was more toxic to esophageal cancer cells than other bile salts, suggesting that apoptosis-induced effect is correlative with the conjugated forms or structures of bile acids in esophageal cancer cells. In hepatocytes, the effect of bile salts is based on the glycine- or taurine-conjugated structure. It was reported that some hydrophobic bile acids such as TC activate PI3K-dependent survival pathways, which prevent their inherent toxicity^[5]. But it seemed that PI3K was not activated by TC in esophageal cancer cells. The different mechanism of different bile salts needs to be defined by further studies.

The apoptosis “death-signal transduction” pathway is currently classified into the Fas/FasL pathway^[26-28], the

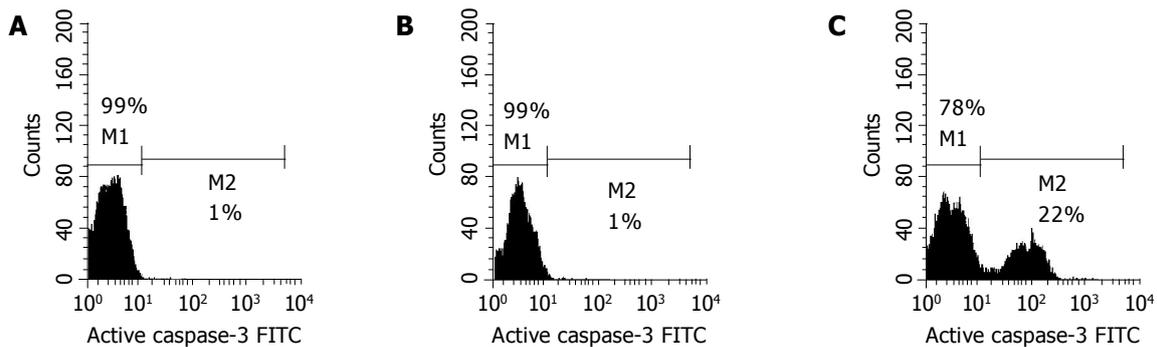


Figure 6 Active caspase-3 in the control (A) and cells treated with GC (B) and TC (C).

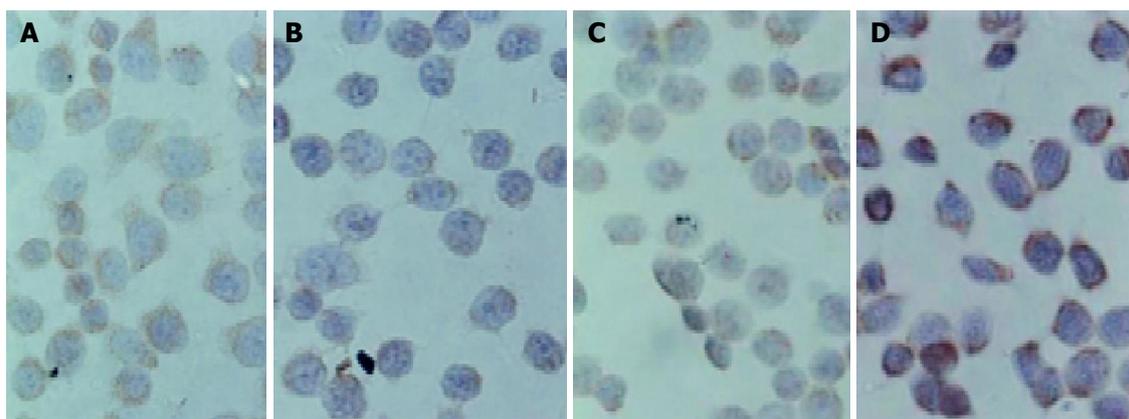


Figure 7 Expression of Bcl-2 (B) and Bax proteins (D) and controls (A: Bcl-2 and C: Bax) ×400.

mitochondrial pathway^[16,29], and the endoplasmic reticulum-stress activated pathway^[30,31]. It was reported that direct oligomerization of the Fas receptor is the primary causative mechanism of bile salt-mediated hepatocyte apoptosis^[11-13]. The signal transduction of bile salt-mediated apoptosis of esophageal cancer cells was also investigated in our study. The data demonstrate that caspase-3 is activated in TC-induced cells. Activation of caspase-3 occurs as a consequence of TC-induced apoptosis. However, caspase-3 plays a pivotal role in the three apoptosis pathways^[32-34]. We further observed the expression of Bcl-2 and Bax protein. Members of the Bcl-2 family, as the key components in the mitochondrial apoptosis "death-signal transduction" pathway, are important regulators in the apoptotic pathway^[15,16]. As an oncogene-derived protein, Bcl-2 confers a negative control in the pathway of cellular suicide, whereas Bax, a Bcl-2-homologous protein, promotes cell death by competing with Bcl-2^[17,18]. Reduced expression of Bcl-2 protein and increased expression of Bax protein in Eca109 cells treated with TC demonstrated in this study, suggest that TC-induced apoptosis is relative with the mitochondrial death-signal pathway in Eca109 cells. As opposed to hepatocyte death mediated by bile acids, CD95 is not involved in deoxycholate-induced apoptosis of colon cancer cell lines^[6]. Because the changes of CD95 were not observed in our study, our results cannot prove that Fas receptor (CD95) is not involved in TC-induced esophageal cancer cell apoptosis. Toxic bile acids may be capable of stimulating multiple pathways to initiate cell death, but further studies are needed.

In conclusion, GCDC, GDC, TC, TCDC, TDC, CA, and DCA, except for GC, inhibit growth and induce apoptosis of Eca109 cells in a dose- and time-dependent manner *in vitro*. Activation of caspase-3, decreased Bcl-2 protein, and increased Bax protein are involved in TC-induced apoptosis of esophageal cancer cells.

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