

BRIEF REPORTS

Relationship between expression and distribution of cyclooxygenase-2 and bcl-2 in human gastric adenocarcinoma

Xiao-Li Chen, Bao-Shan Su, Run-Qin Sun, Jun Zhang, Yi-Li Wang

Xiao-Li Chen, Bao-Shan Su, Run-Qin Sun, Department of Pathology, Second Hospital of Xi'an Jiaotong University, Xi'an 710004, Shaanxi Province, China

Jun Zhang, Section of Gastroenterology, Department of Medicine, Second Hospital of Xi'an, Jiaotong University, Xi'an 710004, Shaanxi

Yi-Li Wang, Institute for Cancer Research, Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Co-correspondents: Yi-Li Wang

Correspondence to: Dr. Xiao-Li Chen, Department of Pathology, Second Hospital of Xi'an Jiaotong University, Xi'an 710004, Shaanxi

Province, China. chenxiaoli64.student@sina.com

Telephone: +86-29-88546322

Gastroenterol 2005; 11(8): 1228-1231

methods for antitumor therapy.

gene (bcl-2); Cyclooxygenase (COX-2)

and bcl-2 in human gastric adenocarcinoma. World J

http://www.wjgnet.com/1007-9327/11/1228.asp

© 2005 The WJG Press and Elsevier Inc. All rights reserved.

Key words: Gastric adenocarcinoma; Apoptosis suppressor

Chen XL, Su BS, Sun RQ, Zhang J, Wang YL. Relationship

between expression and distribution of cyclooxygenase-2

Abstract

AIM: To explore expression and distribution features of COX-2 and bcl-2 in human gastric adenocarcinoma tissues and to study its biological significance.

METHODS: Totally 36 human gastric carcinoma samples were enrolled in this study (cardiac adenocarcinoma 16 cases, distal gastric adenocarcinoma 20 cases). The expressions of COX-2 and bcl-2 in cancerous tissues and corresponding para-cancerous tissues were investigated by immunohistochemistry using COX-2 polyclonal antibody and bcl-2 monoclonal antibody. The normal gastric mucosa tissues were used as control.

RESULTS: The expressions of COX-2 and bcl-2 in gastric carcinoma were significantly higher than that in the paracancerous tissues (77.8% vs 47.2%, P<0.01, 80.56% vs 58.33%, P<0.05). The expression of COX-2 in cardiac adenocarcinoma was remarkably higher than that in the distal gastric carcinoma (93.8% vs 65.0%, P<0.01). The expression of COX-2 was mainly localized in the cytoplasm of tumor cells and partly in the nucleus. There is a transition of the COX-2 cytoplasmic positivity to nucleic in tumor cells with the increase of gastric carcinoma pathological grade. Interstitial macrophages, fibroblasts and vascular endothelial cells also expressed COX-2. The tissues with higher expression of COX-2 also expressed high level of bcl-2 protein.

CONCLUSION: Abnormal expression pattern of COX-2 within the tissues of human gastric cancer is correlated with tumor location and lymph node metastasis. COX-2 may regulate expression of apoptosis suppressor gene (bcl-2) through interaction of tumor cells and stromal cells and play an important role in the generation and development of tumors, which will be of great help in developing new

INTRODUCTION

Cyclooxygenases (COXes), which are the key enzymes in prostaglandin (PG) biosynthesis, are thought to be important in maintaining the normal physiological functions and certain pathological process. Currently, cyclooxygenases have at least 2 isoenzymes in the mammal, namely COX-1 and COX-2. COX-1 is expressed in most tissues and is thought to be important in maintaining the normal physiological functions. COX-2 may be involved in the repairment of peptic ulcer and colitis, development and progression of tumor by promoting proliferation of cells and tumor angiogenesis, and inhibition of apoptosis^[1,2]. Recently, the relationship between COX-2 and human gastric cancer has been emphasized. The increased expression of COX-2 mRNA or protein has been demonstrated in human tumors of digestive system^[3-10], such as esophageal cancer, colorectal cancer, liver cancer and pancreas cancer. The expression of COX-2 protein in human gastric carcinoma has also been reported. However, distribution features of COX-2 protein in human gastric carcinoma and the relationship between COX-2 and bcl-2 are still not clear. To clarify the contribution of COX-2 and bcl-2 in gastric carcinoma, it is essential to examine the expression of COX-2 and bcl-2 within tissues of human gastric carcinoma.

In this study, we used immunohistochemistry method to detect the expression of COX-2 and bcl-2 protein in human gastric adenocarcinoma and their corresponding paracancerous tissues, and to explore the distribution features of COX-2 and bcl-2 in human gastric adenocarcinoma and the biological significance, and to provide rational foundation for designing effective schemes of antitumor therapy.

MATERIALS AND METHODS

A total of 36 human gastric adenocarcinoma (including 16

cardiac adenocarcinomas and 20 distal gastric adenocarcinomas) cases were investigated. All tissues were surgically resected. Cancerous tissue and para-cancerous gastric mucosa were all from the same specimens. Morphologically normal gastric mucosa tissues were used as control. Each specimen was fixed in 10% phosphate-buffered formalin immediately after resection, embedded in paraffin and sectioned into 4 μm for immunohistochemical study and routine histological examination.

The study was approved by the Ethics Committee of our hospital and all patients gave written informed consent prior to enrollment.

Reagents and methods

Polyclonal mouse anti-human COX-2 antibody was purchased from Santa Cruz Biotechnology Co. Mouse anti-human monoclonal antibody bcl-2 was purchased from Danmark Co. The SP staining kit was purchased from Zymed Co. The procedures of immunohistochemical staining were performed according to the manufacturer's instructions. The known positive colorectal cancer tissues were used as positive control. As negative control, PBS was used to replace primary antibody.

Assessment of staining

Under the light microscope, the immunoreactivity was evaluated both qualitatively and quantitatively. Intensity and percentage of positive cells were used to evaluate each tissue section. The immunostaining intensity was scored as 0 (achromatic), 1 (light yellow), 2 (yellow), and 3 (brown).

Immunostaining quantity was scored as follows: 1 = <25%, 2 = 26-50%, 3 = 51-75% and 4 = >75% of tumor cell showing positivity. The combined score, based on both qualitative and quantitative immunostaining, was determined by multiplying the qualitative and quantitative scores. Based on the combined score, the samples were grouped as follows: negative (-) 0-1, weak (+) 2-3, moderate (++) 4-6, strong (+++) >6.

Statistical analysis

With statistical analysis package (SPSS) version 10.0 for Windows, difference between groups was evaluated by using chi-square test and Student's *t* test. The *P* value less than 0.05 was considered statistically significant. All reported *P* values were two-sided.

RESULTS

Expression of COX-2 and bcl-2 in normal gastric mucosa

In the normal gastric mucosa, the expression of COX-2 was mostly negative. Smattering deep glandular epithelia cells showed weak-positive expression of COX-2. The expression of bcl-2 was negative or focus weak-positive reaction in the normal gastric mucosa.

Distribution of COX-2 within tissues of human gastric carcinomas (Table 1)

The expression of COX-2 was mainly localized in the cytoplasm of tumor cells, partly in the nucleus (Figure 1). The expressions of COX-2 were detected in the paracancerous tissues (Figure 2), including gastric mucosal

Table 1 Pathological characteristics of gastric adenocarcinoma and immunohistochemical COX-2 staining

Pathological characteristics	n		Expression	Positive rate (%)	Р		
		-	+	++	+++	Fositive rate (%)	Р
Gastric carcinomas	36	8	12	10	6	77.8	<0.01 ^b
Para-cancerous tissues	36	19	10	7	0	47.2	
Location of tumor							
Cardiac carcinoma	16	1	4	8	3	93.8	<0.01 ^b
Non-cardiac carcinoma	20	7	8	2	3	65.0	
Histological type							
Intestinal carcinoma	17	3	5	5	4	82.3	
Diffuse carcinoma	19	5	7	4	3	74.1	
Metastatic lymph node							
Positive	22	1	10	5	6	95.5	<0.005b
Negative	14	7	2	5	0	50.0	

^bP<0.01 vs the expression of COX-2 was significant between gastric carcinoma and para-cancerous tissues, location of tumor, and metastatic lymph node.

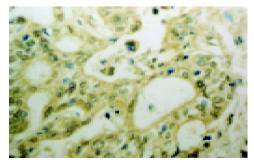


Figure 1 Expression of COX-2 in tissue of human gastric adenocarcinoma. Some stromal cells showed positive stains. Immunohistochemical staining, ×400.



Figure 2 Expression of COX-2 in the para-cancerous tissue. Intestinal metaplasia epithelia and goblet cells showed plasmatic stain. Immunohistochemical staining, ×400.

epithelia, intestinal metaplasia epithelia, goblet cells and a few glandular epithelia, but with lower intensity. The COX-2 expression was also observed on interstitial macrophages, fibroblasts and endothelial cells. The expression of COX-2 in cancerous tissues was significantly higher than that in the para-cancerous tissues (77.8% vs 47.2%). Compared to the distal gastric adenocarcinoma, the rate of COX-2 expression was dramatically higher in cardiac adenocarcinoma (93.8% vs 65%). There was no significant difference in the expression of COX-2 based on the tumor differentiation. However, the cytoplasmic positivity of COX-2 was transformed to nucleic positivity with the increase of pathological grade of gastric carcinoma. Meanwhile, we observed that the expression of COX-2 in metastatic cases was higher than that in non-metastatic one (P<0.005), which suggested that the expression of COX-2 may contribute to infiltrating metastasis potential of cancer cells.

The expression of bcl-2 protein in gastric cancer tissues (see

The expression of bcl-2 protein in gastric cancer tissues was located in cytoplasm of tumor cells and stromal lymphocytes. In para-cancerous tissues, only a few deep glandular epithelium cells and intestinal metaplasia cells expressed bcl-2 protein. The positive rate and intensity of bcl-2 expression in gastric cancer tissues were higher than that in para-cancerous tissues. The expression of bcl-2 protein was not correlated with location and histological type of tumor and lymph node metastasis (data was not shown).

Table 2 Expression of bcl-2 in gastric cancer and para-cancerous tissues

Type of tissue n		Ex	Expression of bcl-2 Positive rate (%)				
Type of tissue n		-	+	++	+++		1
Gastric cancer tissue	36	7	9	12	8	80.56	<0.025 ^a
Para-cancerous tissues		15	14	4	3	58.33	

 $^{^{\}mathrm{a}}P$ <0.05 vs the expression of bcl-2 was significant between gastric carcinoma and para-cancerous tissues.

DISCUSSION

COXes are thought to be important in maintaining the normal physiological and pathological functions. COX-2 may be involved in the development and progress of tumor by promoting proliferation of cells and tumor angiogenesis and inhibiting apoptosis of cells[1,2].

The increased expression of COX-2 mRNA or protein has been demonstrated in human tumors of digestive system^[3-8], such as esophageal cancer, colorectal cancer, liver cancer and pancreas cancer. The expression of COX-2 protein in human gastric cancer tissues has also been reported. However, distribution features of COX-2 protein in human gastric cancer tissues, relationship between COX-2 and bcl-2 are not completely clear. In the present study, we demonstrated that COX-2 protein was overexpressed in human gastric cancer tissues. The location of COX-2 products varied with difference of tumor differentiation.

In intestinal adenocarcinoma, COX-2 protein was present in the cytoplasm of tumor cells with moderate staining intensity. But in diffuse adenocarcinoma, COX-2 protein was localized in nucleus of tumor cells and stained with brown granules. Sometimes, COX-2 protein simultaneously appeared in nucleus and cytoplasm of tumor cells. Interestingly, the positive rate of COX-2 expression and stained intensity in cardiac carcinoma were significantly higher than that in gastric distal carcinomas. These findings differed from other reports[9] in which similar rate of positivity was evident in intestinal-type cardiac carcinoma (52%) when compared to the noncardiac ones (64%). However, there was a trend of lower incidence of strong COX-2 positivity in cardiac than in noncardiac tumors (14% vs 41%). This difference of COX-2 distribution may be associated with specificity and sensitivity related to different COX-2 antibodies and staining protocols or procedures. There may be ethnic difference between different peoples. Recent studies[11] indicated that genesis of cardiac cancer was related to reflux esophagitis, intestinal metaplasia in the junction of stomach and esophagus and atypical hyperplasia. Incidence of cardiac adenocarcinoma was increasing in Chinese, which may be one of the factors that lead to highlevel expression of COX-2 in cardiac cancer.

Our results showed that COX-2 protein was not only expressed in neoplastic epithelial cells, but also in interstitial macrophages and fibroblasts, which might be an incidental phenomenon in the progression of diseases unrelated to carcinogenesis itself or might be a reaction to mucosal lesion. The stromal cells expressing COX-2 can stimulate proliferation of epithelium cells and promote tumorigenesis through paracrine ways^[12]. Williams and his colleagues^[13] found that the growth of a lung carcinoma cell line is markedly attenuated in COX-2 -/- mice or mice treated with a COX-2selective inhibitor. In sporadic colorectal adenomas, COX-2 was localized on superficial interstitial macrophages and dysplastic epithelial cells. The mechanism to induce epithelial carcinogenesis may be COX-2-mediated paracrine signaling between the macrophages and epithelial cells within tumors^[14].

The development and progression of stomach cancer is a pathological process involving many factors, stages and oncogenes, in which imbalance between apoptosis and proliferation of cells plays an important role. Bcl-2 gene is a well-known apoptosis suppressor gene leading to increase of abnormal cells by inhibiting cell apoptosis and prolonging life span of cells, but not promoting cell proliferation, so it will make way for the development of tumors^[15]. Our results showed that the positive rate and intensity of bcl-2 protein expression in tissues of gastric adenocarcinomas was higher than that in para-cancerous tissues, suggesting that epithelial cells might have overexpression of bcl-2 protein during malignancy transformation of gastric mucosa, which makes regulatory mechanism of cells changeable. Therefore, the balance between apoptosis and proliferation of cell was destroyed, which will conduce to development of tumors. The expression of bcl-2 protein was not related to differentiation and location of gastric adenocarcinoma.

Recent studies[16-19] suggest that COX-2 was involved in the development of tumors through following ways: (1) COX-2 can increase products of prostaglandin; (2) COX-2 can transform pre-carcinogen to carcinogen; (3) COX-2 can inhibit apoptosis of cells and promote adhesion of cells; (4) COX-2 can promote angiogenesis of tumor; (5) COX-2 can regulate inflammation and immune functions; (6) COX-2 can enhance invasive ability of tumor cells. We also found in the present experiment that the tissues with a high-level expression of COX-2 expressed simultaneously high-level of bcl-2 protein, indicating the COX-2 can promote expression of bcl-2 and decrease cell apoptosis. Therefore, we suppose COX-2 plays an important regulatory role in bcl-2 expression. Further study of interaction between COX-2 and bcl-2 in the gastric cancer is warranted.

REFERENCES

- 1 Uefuji K, Ichikura T, Mochizuki H. Expression of cyclooxygenase-2 in human gastric adenomas and adenocarcinomas. J Surg Oncol 2001; 76: 26-30
- Wu AW, Gu J, Ji JF, Li ZF, Xu GW. Role of COX-2 in carcinogenesis of colorectal cancer and its relationship with tumor biological characteristics and patients' prognosis. World J Gastroenterol 2003; 9: 1990-1994
- Fournier DB, Gordon GB. COX-2 and colon cancer: potential targets for chemoprevention. J Cell Biochem Suppl 2000; 34: 97-102
- 4 Lim HY, Joo HJ, Choi JH, Yi JW, Yang MS, Cho DY, Kim HS, Nam DK, Lee KB, Kim HC. Increased expression of cyclooxygenase-2 protein in human gastric carcinoma. *Clin Cancer Res* 2000; **6**: 519-525
- 5 Tucker ON, Dannenberg AJ, Yang EK, Zhang F, Teng L, Daly JM, Soslow RA, Masferrer JL, Woerner BM, Koki AT, Fahey TJ. Cyclooxygenase-2 expression is up-regulated in human pancreatic cancer. *Cancer Res* 1999; 59: 987-990
- Yu J, Leung WK, Lee TL, Tse PC, To KF, Sung JJ. Promoter hypermethylation of cyclooxygenase-2 in gastric carcinoma. *Int J Oncol* 2003; 22: 1025-1031
- 7 Shiota G, Okubo M, Noumi T, Noguchi N, Oyama K, Takano Y, Yashima K, Kishimoto Y, Kawasaki H. Cyclooxygenase-2 expression in hepatocellular carcinoma. *Hepatogastroenterology* 1999: 46: 407-412
- 8 Zimmermann KC, Sarbia M, Weber AA, Borchard F, Gabbert

- HE, Schror K. Cyclooxygenase-2 expression in human esophageal carcinoma. *Cancer Res* 1999; **59:** 198-204
- 9 Saukkonen K, Nieminen O, van Rees B, Vilkki S, Harkonen M, Juhola M, Mecklin JP, Sipponen P, Ristimaki A. Expression of cyclooxygenase-2 in dysplasia of the stomach and in intestinal-type gastric adenocarcinoma. *Clin Cancer Res* 2001; 7: 1923-1931
- 10 Zhang J, Liu Y, Cao ZL, Chen XL, Wang KM, Guo XD, Zuo AL, Gong J. Analysis pathological lesions at distal esophagus and esophagogastric junction. Zhonghua Xiaohua Neijing Zazhi 2002; 19: 214-217
- van Rees BP, Saukkonen K, Ristimaki A, Polkowski W, Tytgat GN, Drillenburg P, Offerhaus GJ. Cyclooxygenase-2 expression during carcinogenesis in the human stomach. *J Pathol* 2002: 196: 171-179
- 12 Williams CS, Tsujii M, Reese J, Dey SK, DuBois RN. Host cyclooxygenase-2 modulates carcinoma growth. *J Clin Invest* 2000; **105**: 1589-1594
- 13 Chapple KS, Cartwright EJ, Hawcroft G, Tisbury A, Bonifer C, Scott N, Windsor AC, Guillou PJ, Markham AF, Coletta PL, Hull MA. Localization of cyclooxygenase-2 in human sporadic colorectal adenomas. Am J Pathol 2000; 156: 545-553
- 14 Lauwers GY, Scott GV, Karpeh MS. Immunohistochemical evaluation of bcl-2 protein expression in gastric adenocarcinomas. *Cancer* 1995; 75: 2209-2213
- 15 Sung JJ, Leung WK, Go MY, To KF, Cheng AS, Ng EK, Chan FK. Cyclooxygenase-2 expression in *Helicobacter pylori*-associated premalignant and malignant gastric lesions. *Am J Pathol* 2000; 157: 729-735
- 16 Kelly LM, Hill AD, Kennedy S, Connolly EM, Ramanath R, Teh S, Dijkstra B, Purcell R, McDermott EW, O'Higgins N. Lack of prognostic effect of Cox-2 expression in primary breast cancer on short-term follow-up. Eur J Surg Oncol 2003; 29: 707-710
- 17 Howe LR, Dannenberg AJ. COX-2 inhibitors for the prevention of breast cancer. J Mammary Gland Biol Neoplasia 2003; 8: 31-43
- 18 Zang T, Sun F, Li Y. Expression of COX-2 in prostatic cancer and benign prostatic hyperplasia *Zhonghua WaiKe Zazhi* 2001; 39: 702-703
- Higashi Y, Kanekura T, Kanzaki T. Enhanced expression of cyclooxygenase (COX)-2 in human skin epidermal cancer cells: evidence for growth suppression by inhibiting COX-2 expression. *Int J Cancer* 2000; 86: 667-671