



Generation and characterization of a transgenic mouse model for pancreatic cancer

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Abstract

AIM: To generate a SV40Tag transgenic tumor animal model and to study the mechanism underlying tumorigenesis.

METHODS: A mammary gland expression vector containing SV40Tag DNA was generated. Transgene fragments were microinjected into fertilized eggs of FVB mice. The genetically manipulated embryos were transferred into the oviducts of pseudo-pregnant female mice. PCR and Northern blot analysis were used for genotype analysis of F1 and F2 mice. Transgene expression was detected by RT-PCR and immunohistochemistry.

RESULTS: SV40Tag gene was detected in two lines of transgenic mice. One of them delivered the transgene to F1 and a tumor was found in the pancreas of these mice. RT-PCR and immunohistochemistry showed that SV40Tag gene was expressed in the tumor. Pathological characterization of the transgenic mice demonstrated that the tumor belonged to pancreatic cystic neoplasm.

CONCLUSION: SV40Tag transgenic mouse model can be successfully established. The transgenic mice develop a pancreatic tumor, which can be used for investigation of the molecular mechanism of tumorigenesis *in vivo*.

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Key words: SV40Tag; Transgenic mice; Animal model; Pancreatic neoplasms

INTRODUCTION

Pancreatic cancer is one of the most common malignant diseases threatening human life. The survival rate of pancreatic cancer patients is among the lowest in all kinds of cancer patients. The 1- 3- and 5-year survival rates have been reported to be 16%, 5% and 4%, respectively^[1]. Early diagnosis and effective therapeutic intervention are essential to improve the situation. Animal models of pancreatic cancer should provide important tool for the identification of biomarkers for early detection and molecular mechanism studies of the pathological procession of pancreatic cancer.

Simian virus 40 T antigen (SV40Tag) is a virus oncogene encoded by simian virus 40 (SV40). SV40Tag protein is a multifunctional regulatory protein that binds to and inactivates tumor suppressor genes, including P53 and Rb. It has been shown that interaction between SV40Tag and tumor suppressor proteins stimulates DNA duplication, thus facilitating cell proliferation and inducing cell transformation^[2,3]. A number of tumor transgenic animal models have been established, including pancreatic cancer^[2,3], bone sarcoma^[4], urinary bladder carcinoma^[5] and liver cancer^[6]. In this report, we described the generation and characterization of a SV40Tag transgenic mouse model for pancreatic cancer. The animal model may be useful for molecular mechanism studies on this disease.

MATERIALS AND METHODS

Experimental animals

FVB and CD-1 mice were provided by Center of Comparative Medicine of Yangzhou University. Mice were maintained in standard laboratory conditions in a 12h light/12h dark cycle.

Expression vectors

A 2482-bp SV40Tag DNA fragment was generated by PCR using COS-1 genomic DNA as template. The PCR

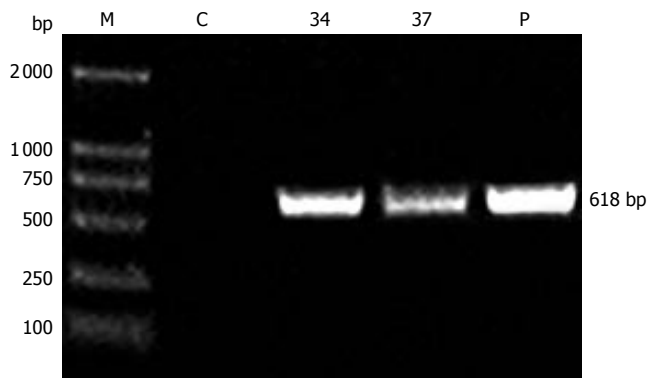


Figure 1 PCR results of transgenic mice. M: DL 2000 marker; C: Non-transgenic mouse control; P: pBC-SV40Tag plasmid; 34 and 37: the number of gene positive mice.

product containing the entire SV40Tag coding region and a 346 bp intron with two Xho I sites at both ends was purified, digested with Xho I and then cloned into pBC-1 vector to generate the transgenic construct pBC-SV40Tag.

Preparation of transgenic mice

An 18.2 kb pBC-SV40Tag fragment was purified from agarose gel (Qiagen, CA, USA) and adjusted to a final concentration of 2 µg/mL in PBS. FVB female mice were hormonally super-ovulated and mated with FVB male mice. The fertilized eggs were collected from the oviduct 24 h later. Transgene pBC-SV40Tag in PBS was microinjected into the pronuclei of fertilized eggs. Finally, the injected fertilized eggs were transplanted into the oviduct of pseudo-pregnant CD-1 mice.

Genotype of transgenic mice

Founder (G0) mice were identified by PCR and Southern blotting analysis. Genomic DNA was isolated from the tails of transgenic mice using a standard method. For PCR, a pair of primers spanning SV40Tag introns was designed (forward: 5'-ACTTTGGAGGCTTCTGGGAT-3' and reverse: 5'-GGTGTAATAGCAAAGCAAGCA-3'), which would produce a 618 bp fragment for genomic DNA template and a 272 bp fragment for cDNA template. PCR using genomic DNA as template was performed under the following condition: 35 cycles at 94 °C for 1 min, at 60 °C 45 s, at 72 °C for 50 s. PCR products were detected by electrophoresis on 1.0 % agarose gels. After PCR screening, transgenic mice were further confirmed by Southern blot analysis. Genomic DNA was digested overnight with Nde I and subjected to electrophoresis on a 0.8 % agarose gels. DNA was transferred onto nylon membrane (Millipore Co., Ltd). The 1 kb fragment of plasmid pBC-SV40Tag digested with Nde I was used as a probe. Southern blot analysis was performed according to ECLTM direct nucleic acid labeling and detection system (Amersham).

Morphology and anatomy examination

Transgenic mice were examined every day. Mice with bulgy belly and showing listlessness were anatomized and photographed. Various organs and tumors were dissected

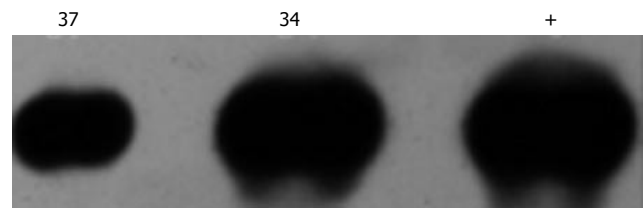


Figure 2 Southern blotting of transgenic mice. +: pBC-SV40Tag plasmid positive control; 34 and 37: the number of gene positive mice.

from the transgenic mice and fixed in 10 % formalin. Sections were obtained from paraffin-embedded tissue samples, stained with hematoxylin-eosin, and examined under a microscope and photographed.

Expression of transgene

Total RNA was isolated from tumor and other tissues with TRIzol reagent (Invitrogen) according to the manufacturer's instructions. First strand cDNA was synthesized using reverse transcriptase (Promega). RT-PCR was performed using the SV40Tag primers described above. PCR was performed for 35 cycles at 94 °C for 1 min, at 60 °C for 45 s, and at 72 °C for 50 s. The PCR products were analyzed by electrophoresis on 1.0 % agarose gels.

Immunohistochemistry analysis

Tissues expressing SV40Tag were collected and fixed in 10 % formalin. Paraffin-embedded tissue samples were cut into 5µm-thick sections. For immunohistochemistry, tissues were de-waxed and treated with newly diluted 3 % H₂O₂ for 15 min to destroy internal peroxidases. Tissues were incubated with normal goat serum at room temperature for 20 min, then incubated with polyclonal antibody of SV40Tag (Santa cruz biotechnology, sc-20800, 1 : 400) at 37 °C for 120 min, and finally with goat anti-rabbit secondary antibodies at 37 °C for 30 min. The samples were stained with DAB using SABC kit (Boster). The nuclei were counterstained with hematoxylin. Slices were observed under microscope and photographed (Olympus).

RESULTS

Establishment of pBC-SV40Tag transgenic mice

pBC-SV40Tag was generated and confirmed by restriction enzyme map and DNA sequence analysis. A total of 39 mice were analyzed for production of transgenic mice. Two mice, one male and one female, were identified as transgenic mice by PCR (Figure 1). The serial numbers of the positive mice were line 34 (female) and line 37 (male). These transgenic mice were confirmed by Southern blotting analysis using Nde I digested SV40Tag DNA fragment as a probe (Figure 2). The line 34 developed a tumor at early stage (about 4 wk). Line 37 produced F1 off-springs by mating with normal FVB female mice.

Tumorigenesis of pBC-SV40Tag transgenic mice

Line 37 mice and positive off-springs developed tumor. At age of 10 wk, the mice showed a palpable abdominal mass and became gradually depressed and moribund (Figure 3A). After the skin of abdomen was dissected,

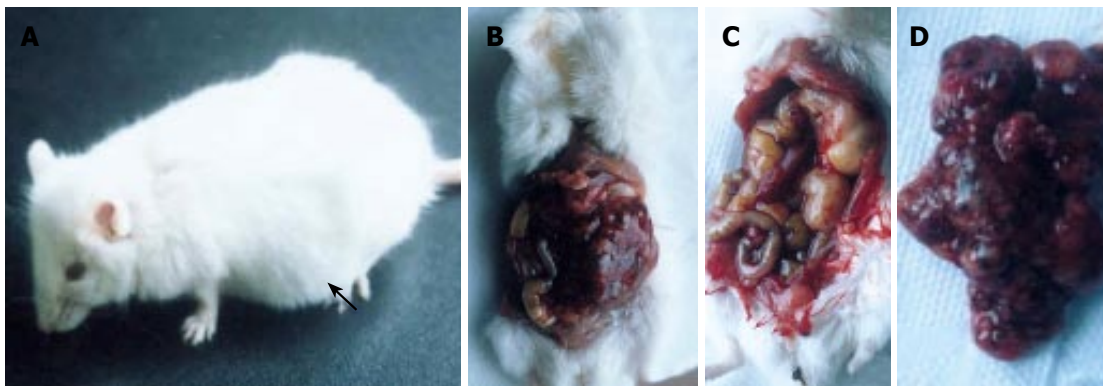


Figure 3 Appearance and anatomy of transgenic mice. **A:** Transgenic mice showing ridgy belly (arrowhead); **B:** Transgenic mice showing a tumor in its abdominal cavity; **C** and **D:** Abdominal cavity and tumor excised from the body.

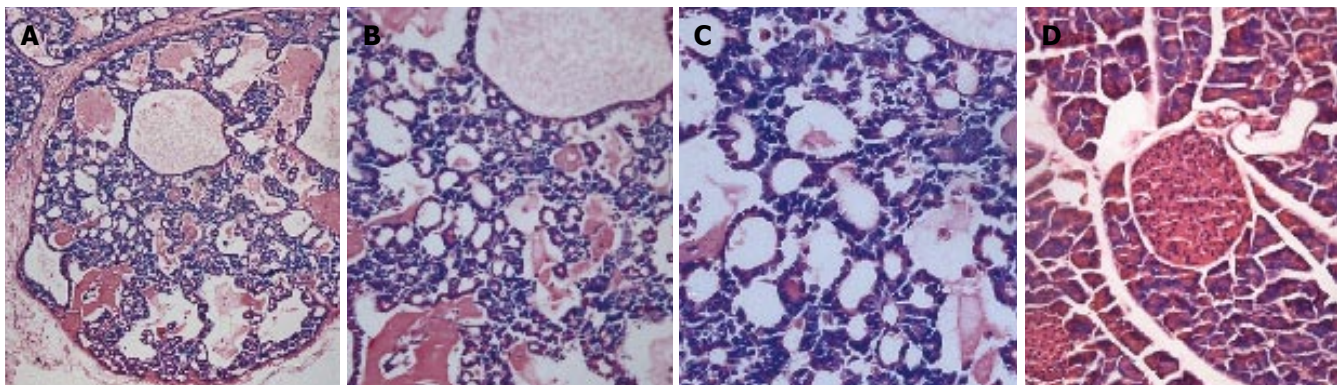


Figure 4 Normal pancreas and tumor tissues-stained with H&E. **A, B** and **C:** Tissue of cystic neoplasms from transgenic mice (**A:** $\times 100$; **B:** $\times 200$; **C:** $\times 400$); **D:** Normal pancreas with islet and acinar cell ($\times 200$).

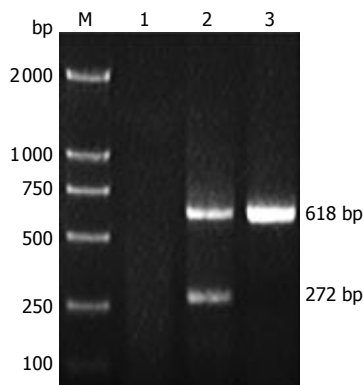


Figure 5 RT-PCR results of transgene expression in tumor. **M:** DL-2000 marker, **1:** blank control (H_2O), **2:** cDNA of tumor tissue, **3:** negative control (non-reverse transcribed RNA).

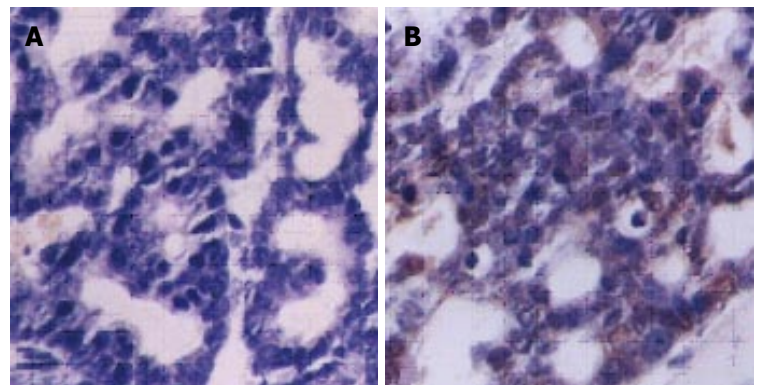


Figure 6 SV40Tag expression in tumor tissue detected by immunohistochemistry without anti-Tag antibody (**A**) and with anti-Tag antibody (**B**).

tumor tissue in the abdominal cavity was observed across peritoneum. In addition, a multilocular cystic tumor was identified in the abdominal cavity (Figures 3B and 3C). The diameter of the cysts ranged from 2 mm to 2 cm. The cystic cavities were filled with wine fluid (Figure 3D). Additionally, the spleen and kidney became swollen, and the kidney showed obvious pathological changes. No other obvious pathological changes were observed in other organs (data not shown). The cystic spaces were lined by multi-layer epithelium cells (Figures 4A-4C).

SV40Tag expression in tumor tissue

To examine whether the transgene was expressed in

multiple tissues and tumor of transgenic mice, we performed RT-PCR for SV40Tag. A 272 bp PCR product was generated from tumor cDNA (Figure 5) by using SV40Tag specific primers, but not in other tissues. Furthermore, immunohistochemical analysis detected SV40Tag protein expression in tumor tissues of transgenic mice (Figure 6).

DISCUSSION

The early region of simian virus 40 (SV40) encodes a tumor antigen (Tag), a multifunctional regulatory protein responsible for the alterations of cellular processes is

required for viral infection^[7]. SV40 Tag forms complexes with and inactivates the key cell cycle regulatory proteins, such as p53 and retinoblastoma (Rb) tumor suppressor gene product^[7-10]. Loss of these functional tumor suppressor gene products is often associated with human cancer. SV40 Tag expression under the control of tissue specific promoters has been used to generate a variety of transgenic models for human cancers^[11-14]. We have used breast-specific β -casein promoter for SV40 Tag transgenic models. Since the β -casein promoter can specifically drive the expression of SV40 Tag in the breast, we expected to generate a breast cancer transgenic mice. Surprisingly, the transgenic mice failed to produce breast cancer but developed pancreatic neoplastic lesions. One possible explanation is that the transgene has been integrated into a particular location of the mouse genome and leads to the expression of SV40 Tag in the pancreas. Another possibility is that the regulated element of the expression vector has been activated in the pancreas. In addition, the transgenic female mice developed a pancreatic tumor and died before lactation. Thus we have no way to know whether SV40 Tag is expressed in the galactophore of the transgenic mice.

Cystic pancreatic neoplasms comprise a heterogeneous group of pathologic entities. Although the incidence rate and lethality of the cystic pancreatic neoplasm are not as high as that of the pancreatic adenocarcinoma, some of the cystic pancreatic neoplasms can develop into metastatic pancreatic adenocarcinoma. The most common cystic pancreatic neoplasms are mucinous cystic neoplasm and serous cystadenoma. These two entities account for more than 75% of the cystic pancreatic neoplasms. Other less common cystic pancreatic neoplasms include papillary cystic tumor (also known as solid and papillary tumor), cystic neuroendocrine tumor, acinar cystadenocarcinoma, cystic teratoma, lymphangioma, hemangioma, and paraganglioma^[15]. Due to the low incidence and recent nosographic arrangement, the natural evolution and the clinical development of these neoplasms are not well known. It is, therefore, difficult to identify all types of these cystic lesions. Pancreatic cancers are rarely observed spontaneously or induced by carcinogen administration in the laboratory mice. However, genetic engineering has been shown to effectively generate mice with germline oncogenic lesions similar to human pancreatic adenocarcinomas^[16]. We have generated a transgenic animal model with classic features of human cystic pancreatic cancer. This model provides a tractable genetic system to dissect the complexities of evolving cancers in a physiological context. We believe that the mouse model is helpful for the identification of early disease markers and effective therapeutic targets for the disease.

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