



## GASTRIC CANCER

# Anticancer effect of Jinlongshe granules on *in situ*-transplanted human MKN-45 gastric cancer in nude mice and xenografted sarcoma 180 in Kunming mice and its mechanism

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## Abstract

**AIM:** To study the antitumor effect of Chinese compound Jinlongshe (JLS) granules on sarcoma 180 and MKN-45 human gastric cancer cell lines *in vivo* and its mechanism.

**METHODS:** After establishment of S180 sarcoma (S180) and MKN-45 gastric cancer model of nude mice, the tumor-bearing mice were divided into 5 groups at random. Three experimental groups were respectively given the aqueous extract of JLS granules at doses of 120 g, 60 g and 20 g/(kg per 6/wk, *i.g*) for 3 wk in S180 and 6 wk in nude mice model. Positive control was given cyclophosphamide (Cy) at a dose of 50 mg/(kg per 3 /wk, *i.g*) for 3 wk in S180 models and 5- Fluorouracil (5-FU) 20 mg/(kg per 3 /wk, *i.g*) for 3 wk in nude mice model. Negative control was given normal saline (NS) at a dose of 0.18 g/(kg per 6/wk, *i.g*) respectively. After 3 wk in mice bearing S180 tumor and 6 wk in nude mice model, the experimental animals were sacrificed and the masses of tumor were weighed, and the rates of tumor inhibition of each treated group were calculated respectively. To determine the antitumor mechanisms, the morphological changes, cell cycle and apoptosis were observed in MKN-45 nude mice model. Annexin V-FITC/PI double staining FCM assay was used to further determine the live cells, apoptotic cells, necrotic cells and debris.

**RESULTS:** The inhibitory rates of JLS granules at the doses of 20 g/kg, 60 g/kg and 120 g/kg were 50.31%, 55.94% and 68.13% ( $P < 0.01$ ) in nude mice models and 40.90%, 50.32% and 58.46% ( $P < 0.01$ ) in S180 model. The inhibitory rate of Cy was 85.22% in S180

models and the inhibitory rate of 5-FU was 53.43% in nude mice model ( $P < 0.01$ ). Nuclear chromatin and margination were observed under a transmission electron microscope (TEM). The G0/G1 phase was arrested, typical apoptotic peak appeared, the apoptotic rate was 22.81%-38.54% in three JLS granule-treated groups. Annexin V-FITC/PI double staining FCM assay showed that the apoptotic cells were 4.36%, 3.08% and 7.08% in three dosages, most cells were localized in the low right quadrant.

**CONCLUSION:** Jinlongshe granules possess anti-tumor effects on experimental tumor models *in vivo*, and apoptosis induction is one of its anti-tumor mechanisms.

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**Key words:** Jinlongshe granules; Sarcoma 180; Human MKN-45 gastric cancer lines; Traditional Chinese medicine

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## INTRODUCTION

Gastric cancer is one of the most leading causes of cancer-related deaths worldwide<sup>[1]</sup>. Despite its decreasing trend in the Western world during the recent two decades, gastric cancer still accounts for about 23.2% of all cancer deaths in China<sup>[2-4]</sup>. Although surgery, chemotherapy, and irradiation are the mainstream therapeutic methods for gastric cancer, undesirable chemotherapy reaction and poor prognosis in the advanced gastric cancer patients are still the major problem for gastric cancer treatment<sup>[5]</sup>. Consequently, combination of multi-therapeutic methods may effectively improve gastric cancer patient's symptoms and prolong their survival time<sup>[6-7]</sup>.

Jinlongshe (JLS) granules are an oral Chinese medicine compound, consisting of Chinese herbs (*Rhizome Arisaema*, *Rhizoma Pinelliae*, *corium stomachium galli*, *Radix*

*Glycyrrhizae preparata*, etc.), developed by our department based on TCM theories of “phlegm” as the key factor in preventing and treating gastric cancer<sup>[8]</sup>. However, the exact antitumor mechanism of JLS granules is unclear. This study was to observe the anti-tumor effect of JLS granules and to determine its mechanism using sarcoma 180 and human gastric cancer cell line MKN-45 *in vivo*.

## MATERIALS AND METHODS

### Experimental animals

BALB/C nude mice (19–22 g) and Kunming mice (20–22 g) were provided by Experimental Animal Center of Chinese Academy of Sciences, Shanghai, China. All experimental animals were housed in specific-pathogen free (SPF) condition for 1 wk to adapt the surroundings before initiation of the experiment and had free access to food and water throughout the study.

### Cancer cell lines

S180 cell line and MKN-45 human gastric denocarcinoma cell line were provided by Institute of Pharmaceutical Research, Shanghai, Chinese Academy of Sciences.

### Drug samples and reagents

Crude JLS granules were purchased from Leiyunshang Company, Shanghai, China. An aqueous extract of JLS granules at a concentration of 6 g/mL of raw materials was provided by Shanghai Changzheng Hospital. The aqueous extract was diluted with purified water before use. Cyclophosphamide injection (Cy, Lot No. 040909) was produced in Hualian Pharmaceutical Co., Ltd, Shanghai, China. OB Biogel (Lot No.040514) was produced in Baiyun Medical Biogel Co., Ltd, Guangzhou, China. Ketamine hydrochloride injection (Lot No.02072332) was produced in Hengrui Pharmaceutical Co., Ltd, Jiangsu, China.

### Establishment of *in situ*-transplanted human MKN-45 gastric carcinoma nude mouse model

The procedure of *in situ*-transplanted gastric cancer nude mouse model was referred to Xu and Chen Ling *et al.*<sup>[9–10]</sup>. In brief, before transplantation, nude mice bearing gastric denocarcinoma MKN-45 cell lines were killed. Then the tumor was dissected and put into aseptic saline. Pieces of intact, fish meat-like tumor tissue of 1 mm × 1 mm × 1 mm in diameter from margin were prepared after removal of tumor capsule. Mice anesthetized with ketamine (50 mg/kg, ip) were sterilized with caseiodine and a transverse incision was made on the left upper quadrant. After exposure of the greater curvature of stomach, serosa and muscular layer were incised with needle. Then tumor tissues were transplanted under serosa of stomach and one drop of OB biogel was applied to the tumor tissue. After 40 s, stomach was brought into abdominal cavity and abdominal wall was finally closed. The whole operation procedure was completed following the doctrine of aseptic manipulation.

### Establishment of S180 model

This model was by subcutaneous injection as previously described<sup>[11]</sup>. Briefly, the S180 mouse sarcoma cells with

ascites were harvested, diluted with sterilized saline at a ratio of 1:8 (cell concentration was adjusted to  $1.0 \times 10^6$ /mL), and inoculated subcutaneously into the right armpit region of Kunming mice.

### Inhibitory effect of JLS granules on S180 and MKN-45 *in vivo*

Forty-eight hours after establishment of the MKN-45 gastric cancer model and 24 h after establishment of S180 model, the transplanted mice were divided into 5 groups according to the method described in “National Requirements for *in vivo* Screening of Anti-neoplastic Drugs”<sup>[12]</sup>. The dose of JLS granules was decided according to the clinical dose used in humans. The mice bearing S180 were killed by dislocation of cervical vertebra 3 wk after treatment. The nude mice bearing *in situ*-transplanted MKN-45 gastric cancer cell line were sacrificed 6 wk after treatment. The tumor weights were measured and the inhibitory rates were calculated according to the following equation: rate of inhibition (%) = (mean tumor wt of negative control - mean wt of treated group)/mean tumor wt of negative control × 100.

### Morphologic assessment of apoptosis in MKN-45 nude mice

We selected NS group, 5-FU group and 60 g/kg JLS granules group for morphologic assessment of apoptosis in MKN-45 nude mice. Tumor tissue was cut into small pieces, homogenized into 1 mm × 2 mm, prefixed with 4% paraformaldehyde for 4 h and washed with PBS (0.1 mmol/L), post-fixed with 1% osmic acid, dehydrated in graded ethanol-acetonum, embedded in an Epon 812 mixture, and cut into sections on a ultramicrotome. After stained with uranyl acetate and lead citrate, the sections were examined under a Hitachi-800 transmission electron microscope.

### Cell cycle and apoptosis in MKN-45 nude mice

Fresh tumor tissue separated from nude mice was chopped with mechanical method. Single cell-suspended solution was obtained with 200 filter screens. The suspensions were divided into two parts. One part was fixed with 70% ice-cold ethanol for at least 12 h at 4 °C and then washed with PBS again before examination. The fixed cells were treated with RNase A and stained with PI at 37 °C for 30 min. Analysis was performed with a Coulter FCM (EPICS-XL) and data were analyzed with Multicycle DNA content and cell analysis software.

### Annexin V-FITC/PI double staining for FCM-assessed apoptosis

The Annexin V FITC kit (Jingmei Company, Lot No. 20050910) was used for detecting apoptosis according to manufacturer's instructions. The suspensions were washed twice and adjusted to a concentration of  $1 \times 10^6$  cells/mL with ice cold 4 °C PBS. The Falcon tubes (12 mm × 75 mm, polystyrene round-bottom) were used in experiment, 100 µL of suspensions was added to each labeled tube, 10 µL of Annexin V-FITC and 10 µL 20 µg/mL PI were added into the labeled tube, incubated for at least 20 min at room temperature in the dark, then 400 µL of PBS

**Table 1** Anti-tumor effect of JLS granules on mice bearing S180

Group	n	Tumor weight (g)	Inhibition rate (%)
Control	10	4.67 ± 3.27	-
120 g/kg	10	1.94 ± 2.03 <sup>ab</sup>	58.46
60 g/kg	10	2.32 ± 2.83 <sup>ab</sup>	50.32
20 g/kg	10	2.76 ± 2.32 <sup>ab</sup>	40.90
Cy	10	0.69 ± 0.33 <sup>b</sup>	85.22

<sup>a</sup>*P* < 0.05 vs Cy; <sup>b</sup>*P* < 0.01 vs control.**Table 2** Anti-tumor effect of JLS granules on MKN-45 cells in nude mice

Group	n	Tumor weight (g)	Inhibition rate (%)
Control	10	3.20 ± 0.96	-
120 g/kg	10	1.02 ± 0.57 <sup>b</sup>	68.13
60 g/kg	10	1.41 ± 1.39 <sup>b</sup>	55.94
20 g/kg	10	1.59 ± 1.12 <sup>b</sup>	50.31
5-FU	8	1.49 ± 1.37 <sup>b</sup>	53.43

<sup>b</sup>*P* < 0.01 vs control.

binding buffer was added to each tube without washing and analyzed by FCM as soon as possible (within 1 h).

### Statistical analysis

Data were presented as mean ± SD and analyzed with SPSS 11.0 software. The analysis of variance (ANOVA) was used and least significant difference *t* test (LSD-*t*) was used for group comparison. *P* < 0.05 was considered statistically significant.

## RESULTS

### Anti-tumor effect of JLS granules on S180

JLS granule at the doses of 20 g/kg, 60 g/kg and 120 g/kg significantly inhibited the growth of S180 subcutaneous tumor (*P* < 0.01) in a dose-dependent manner, but lower than the inhibitory rates of Cy (*P* < 0.05) (Table 1).

### Anti-tumor effect of JLS granules on in situ-transplanted gastric cancer MKN-45 cell lines

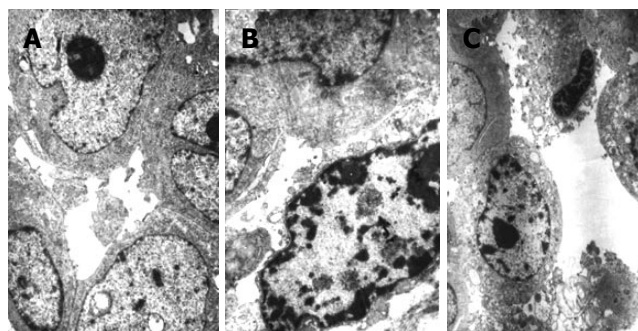
JLS granule at the doses of 20 g/kg, 60 g/kg and 120 g/kg significantly inhibited the growth of orthotopic-transplanted MKN-45 gastric cancer (*P* < 0.01) in a dose-dependent manner, but no difference statistically compared with 5-FU (*P* > 0.05) (Table 2).

### Morphology changes

Condensation of chromatin at margins of nuclei, disintegration of nucleolus, vacuoles in cytoplasm were observed in 60 g/kg group and 5-FU also had the same apoptotic appearance under TEM (Figure 1).

### Assessment of cell cycle and apoptosis of MKN-45 cells in nude mice

The percentage of G0/G1 cells in JLS granules group was higher than that in NS group and 5-FU group in a dose-

**Figure 1** Ultra-structure observed under transmission electron microscopy in control group × 4 000 (A), 60 g/kg JLS group × 6 000 (B), 5-FU group × 3 000 (C).**Table 3** Effect of different treatments on cell cycle and apoptosis of MKN-45 cells (mean ± SD)

Group	n	G <sub>0</sub> /G <sub>1</sub>	G <sub>2</sub> /M	S	Apoptotic rate
Control	10	42.40 ± 11.28	21.28 ± 11.43	36.92 ± 12.21	19.88 ± 14.83 <sup>c</sup>
120 g/kg	10	50.58 ± 8.78	26.45 ± 12.53	14.56 ± 9.35 <sup>a</sup>	38.54 ± 12.65 <sup>a</sup>
60 g/kg	10	47.55 ± 7.02	26.51 ± 15.05	19.36 ± 12.69 <sup>a</sup>	28.27 ± 11.25
20 g/kg	10	46.37 ± 12.09	25.47 ± 14.79	24.04 ± 13.27 <sup>a</sup>	22.81 ± 10.65 <sup>c</sup>
5-FU	8	41.78 ± 4.30	23.59 ± 11.97	19.97 ± 13.31 <sup>a</sup>	24.97 ± 9.93 <sup>c</sup>

<sup>a</sup>*P* < 0.05 vs control, <sup>c</sup>*P* < 0.05 vs 120 g/kg.

dependent manner (*P* > 0.05). The cell percentage of S phase in of JLS group and 5-FU group was lower than that in control group (*P* < 0.05), the apoptotic rate in 120 g/kg JLS group was higher than that in control group, 20 g/kg JLS group and 5-FU group (*P* < 0.05). The typical apoptotic peak was observed (Table 3, Figure 2).

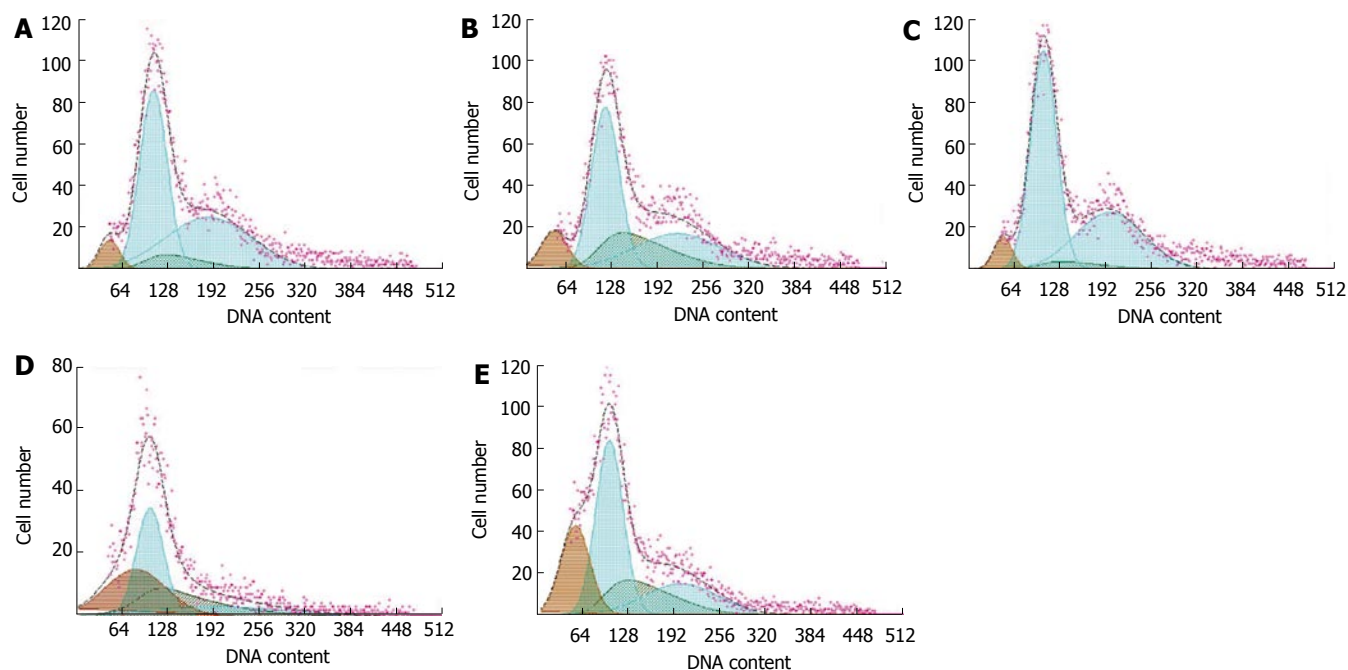
### Assessment of apoptosis in MKN-45 nude mice

Results by Annexin V-FITC/PI double staining for FCM assay showed that the early apoptotic cells were 4.36%, 3.08% and 7.08% in the three JLS granules groups (Figure 3), and most of cell populations in 5-FU treated group were located in the up right quadrant and the cells on the lower right quadrant were not many as compared with JLS granules groups.

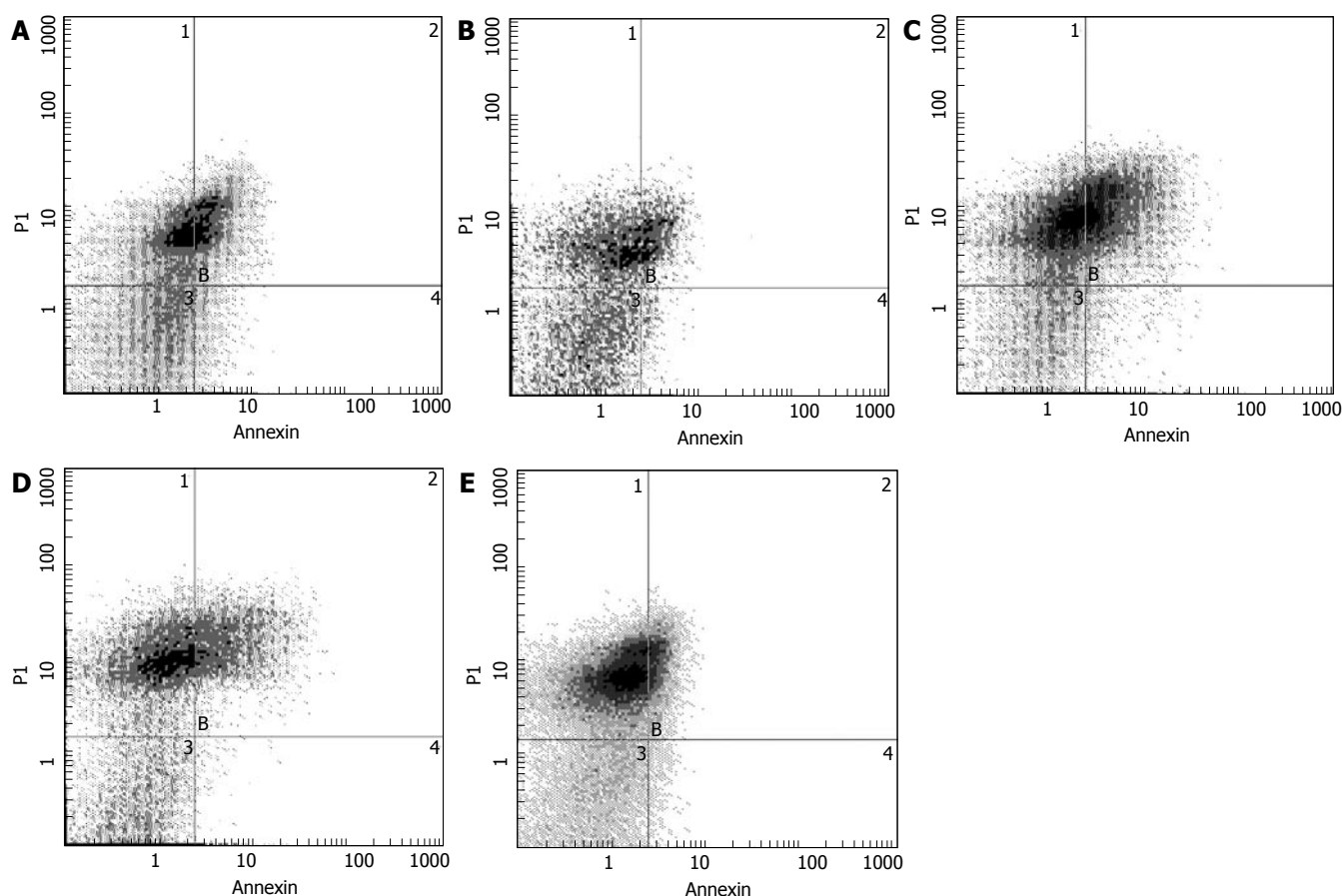
## DISCUSSION

Tumorigenesis, in a sense, is due to the deregulation in cell proliferation and apoptosis<sup>[13-14]</sup>, accordingly tumor treatment not only inhibits cell proliferation, but also induces apoptosis. More and more attention is paid to apoptosis in treatment of tumors and the effectiveness of anti-tumor agents is often evaluated<sup>[15,16]</sup>. Investigators have confirmed that many Chinese herbs have antitumor property and induction of apoptosis is one of the mechanisms<sup>[17-20]</sup>. But most of these studies aimed directly at single herbs, bioactive constituents or leading compounds extracted from herbs, and could not educe the multi-target action of Chinese compounds and traditional Chinese medicine (TCM) theories in disease therapy<sup>[21]</sup>.

In TCM theories, “phlegm” is not only a pathologic metabolite, but also a causative agent. The metabolites such as ectopic hormone, growth factor and tumor



**Figure 2** Histogram observed under FCM in 120 g/kg JLS group (A), 60 g/kg JLS group (B), 20 g/kg JLS group (C), control group (D), and 5-FU group (E).



**Figure 3** Cytogram observed by Annexin V-FITC/PI double staining assay in 120 g/kg JLS group (A), 60 g/kg JLS group (B), 20 g/kg JLS group (C), control group (D), and 5-FU group (E).

markers produced by tumor cells all belong to “phlegm”, which can devitalize physiologic functions and stimulate tumor invasion and metastasis in turn. Therefore,

“dispersing phlegm and eliminating stagnation” (cleaning metabolites produced by tumor) is one of the regimens for tumor treatment in TCM. JLS granules are an anti-

tumor compound prescription developed according to the methods of “dispersing phlegm and eliminating stagnation”. Our previous experiments have shown that “dispersing phlegm and eliminating stagnation” inhibit could VEGF, Ras mRNA and protein expression<sup>[22-24]</sup> and have good antitumor and anti-metastasis potency *in vivo*. However whether it induces apoptosis is unclear. Our studies showed that JLS granules had anti-tumor effects on S180 and MKN-45 human gastric cancer. The ultra-structure of MKN-45 gastric cancer tissue treated with JLS granules showed that there were apoptosis changes. Cell cycle and apoptosis analysis also demonstrated that JLS granules could arrest MKN-45 cells at G0/G1 phase, decrease S phase cell populations. The typical apoptosis peak further confirmed its apoptosis-inducing effect.

In addition, changes in the plasma membrane are one of the earliest morphological features of apoptosis. At this time, the membrane phospholipid phosphatidylserine (PS) is translocated from the inner to the outer leaflet of the plasma membrane, thereby exposing PS to the external cellular environment. Annexin V is a 35-36kD  $\text{Ca}^{2+}$ -dependent phospholipid binding protein that has a high affinity for PS, thus Annexin V staining serves as a sensitive probe for flow cytometric analysis of cells undergoing apoptosis<sup>[25-26]</sup>. Therefore, staining with Annexin V in conjunction with vital dyes such as propidium iodide (PI) can identify early apoptotic cells (Annexin V positive, PI negative). In our study, cells treated with JLS granules were located in the low right quadrant and cells in 5-FU-treated group were located in the upper left quadrant, suggesting that the earlier apoptotic cells in JLS granules-treated groups are higher than those in 5-FU-treated group and that JLS granules induce apoptosis in human gastric cancer MKN-45 cells.

In conclusion, JLS granules have the inhibiting effects on human gastric carcinoma and induction of apoptosis may be one of its mechanisms.

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