

BASIC RESEARCH

Experimental gastric dysrhythmias and its correlation with *in vivo* gastric muscle contractions

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Abstract

AIM: To study the direct correlation between gastric dysrhythmias and *in vivo* gastric muscle tone.

METHODS: Five healthy dogs were implanted with 4 pairs of electrodes along the greater curvature, with a strain gauge (SG) being sutured parallel to the distal electrodes (2 cm above the pylorus). Intravenous vasopressin was given to induce gastric dysrhythmia. The percentage of regular slow waves and SG energy were calculated.

RESULTS: (1) the regularity of gastric myoelectric activity (GMA) was reduced during and after infusion of vasopressin; (2) SG energy was significantly decreased during the infusion of vasopressin; (3) the decrease in SG energy was well correlated with the reduction in GMA regularity; (4) SG energy was negatively correlated with bradygastria and tachygastria.

CONCLUSION: Vasopressin inhibits gastric contractions and impairs gastric slow waves; gastric dysrhythmias are associated with the reduced antral muscle contractions, and are indicative of antral hypomotility.

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Key words: Gastric myoelectric activity; Bradygastria; Tachygastria; Gastric muscle contraction; Canine study

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INTRODUCTION

Gastric motility is under the control of gastric myoelectric activity (GMA). Normally GMA originates from the junctional area of the fundus and proximal stomach, propagates aborally, controlling the frequency and direction of gastric contractions. Disturbances in GMA, so-called gastric dysrhythmias, have been frequently observed in patients with various diseases including gastroparesis^[1], dyspepsia^[2], anorexia nervosa^[3], gastroesophageal reflux diseases (GERD)^[4], motion sickness^[5,6] and pregnancy^[7], etc. Gastric dysrhythmias have also been linked to gastrointestinal symptoms such as nausea and vomiting^[5,6], and improvement of gastric dysrhythmias seems to be associated with relief of such symptoms^[5,8].

Gastric dysrhythmias are classified into tachygastria (frequency higher than normal), bradygastria (frequency lower than normal) and arrhythmia (no rhythmic activity), based on the dominant frequency of GMA. Although it is believed that gastric motility could be affected inevitably, the relationship between gastric dysrhythmias and gastric motility has not well been established. Various techniques including electrogastrography (EGG), gastric emptying and simultaneous recording of EGG and intraluminal pressure have been applied, and available evidence suggests that tachygastria may be associated with hypogastric motility^[9-12], while the effect of bradygastria on gastric motility remains controversial^[13,14]. The association between specific gastric dysrhythmia and gastric muscle contractions has never been carefully investigated.

To address these issues, in this study we specifically evaluated: (1) the effect of intravenous vasopressin on GMA; (2) the correlation between gastric muscle tone and GMA; (3) the effect of experimentally-induced gastric dysrhythmias on gastric muscle contractions, in a canine model.

MATERIALS AND METHODS

Animal preparations

Seven healthy female hound dogs (15-22 kg) were anesthetized with intravenous infusion of thiopental sodium (20 mg/kg) and maintained with inhalation of isoflurane (1%-2%). A midline laparotomy was performed, and four pairs of temporary cardiac pacing wires (A&E Medical, Farmingdale, NJ) were implanted under the serosal surface along the greater curvature of the stomach. The most distal pair was 2 cm above the pylorus, and the distance between adjacent pairs of electrodes was

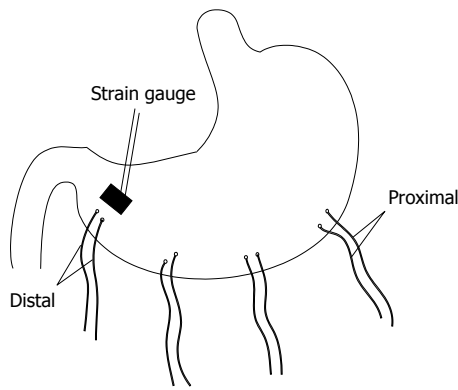


Figure 1 Surgical preparations of stomach. Four pairs of temporary electrodes were implanted along the greater curvature at a distance of 4 cm. The most distal pair was 2 cm above the pylorus. A strain gauge was sutured on the serosal wall parallel to the most distal pair of electrodes.

4 cm (Figure 1). The electrodes in each pair were 1 cm apart. The electrodes were affixed to the gastric serosa by un-absorbable suture in the seromuscular layer of the stomach. The wires were brought out through the anterior abdominal wall, channeled subcutaneously along the right side of the trunk, and placed outside the skin for the attachment for recording gastric myoelectric activity. A strain gauge was sutured on the serosal wall parallel to the most distal pair of electrodes (Figure 1).

The study was initiated about 10 d after the surgery. The protocol was approved by the Animal Use and Care Committee of the University of Texas Medical Branch at Galveston, Texas, USA.

Experimental protocols

Experiments were performed in overnight fasted, fully recovered animals. One study session was performed in each animal. Vasopressin was administered intravenously to induce gastric dysrhythmias in each session. In brief, after a 20 min baseline recording, Vasopressin (0.5 U/kg) was infused continuously for 20 min, followed by another two 20 min recovery periods. GMA and strain gauge signal were recorded simultaneously throughout the session.

Recording of gastric myoelectrical activity and gastric muscle tone

Gastric electrical activity and gastric muscle tone were recorded using a multi-channel recorder system (Acknowledge, Biopac System, Santa Barbara, CA). All signals were displayed on a computer monitor and saved on a hard disk by an HP Pentium IV PC. For GMA, the low and high cutoff frequencies of the amplifier were 0.05 Hz and 35 Hz, and the sampling frequency was 20 Hz. The recordings were then low-pass filtered with a frequency of 1 Hz and re-sampled at a frequency of 24 Hz to reduce the volume of data and potential artifacts before the final review.

Analysis of gastric myoelectrical activity and gastric muscle tone

Regularity of GMA: The percentage of 4- to 6-cpm slow waves is a quantitative assessment of the regularity

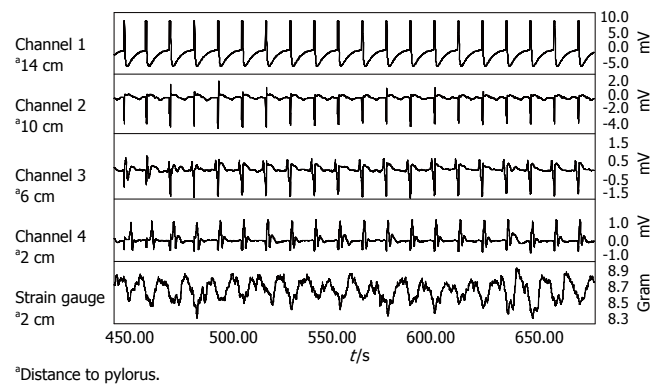


Figure 2 Sample tracings of gastric myoelectric activity recorded from implanted temporary electrodes and gastric contractions from the attached gastric strain gauge. Top 4 channels are gastric myoelectrical activities recorded using serosal electrodes implanted along the greater curvature. The bottom tracing shows gastric contractions measured from a strain gauge implanted in the distal stomach parallel to Channel 4. It was noticed that slow waves propagate from proximal to distal stomach and each gastric slow wave was coupled with a gastric contractile event.

of the GMA measured from the EGG. It was defined as the percentage of time during which normal 4- to 6-cpm gastric slow waves were observed in a specific EGG recording. The percentage of normal 4- to 6-cpm slow waves was computed from the running power spectra of the GMA using an adaptive spectral analysis method^[15]. One power spectrum was generated for every 2 min of EGG data, and the spectral peaks in each spectrum were examined visually. A spectrum was defined as normal if it had a clear peak in the 4- to 6-cpm range. The percentage of regular 4- to 6-cpm slow waves was determined by computing the ratio between the numbers of normal and total spectra.

Gastric dysrhythmias: The percentage of time of gastric dysrhythmias was also computed using running spectral analysis. Bradygastria was defined as a frequency of 0.5-4.0 cpm. Tachygastria was defined as a frequency of 6.0-15.0 cpm. Arrhythmia was defined as any irregular rhythm.

Gastric muscle tone: Reflected by the energy of the strain gauge signals, which were derived by calculating the area under the curve.

Statistical analysis

GMA regularity within each session was compared with One-Way ANOVA. The correlation between gastric muscle tone and gastric dysrhythmias was analyzed with Pearson's correlation test. All data were presented as mean \pm SE. $P < 0.05$ was taken as significance.

RESULTS

Gastric myoelectrical activity and gastric muscle tone at baseline

Intrinsic, distally propagating gastric slow waves were observed in all animals at a mean frequency of 5.6 ± 0.3 cpm, with a range of 4.8-6.2 cpm. Rhythmic variations in gastric muscle tone were consistently present, and coupled with each gastric slow waves at an identical frequency (Figure 2).

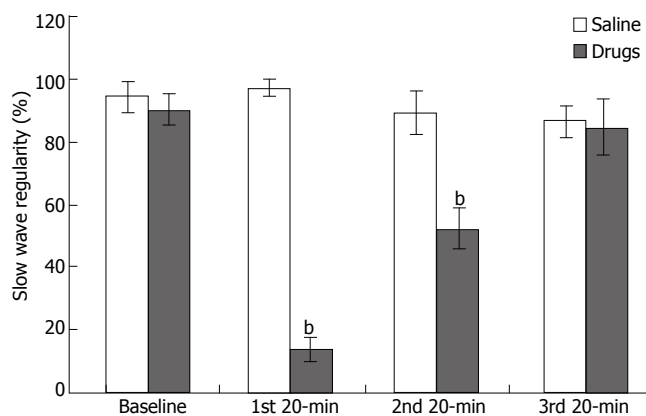


Figure 3 The regularity of gastric myoelectric activity was reduced with and after infusion of vasopressin, and returned somewhat during the last 20-min recovery period. ANOVA, ^b $P < 0.001$ baseline vs 2nd or 3rd 20-min period ($n = 7$).

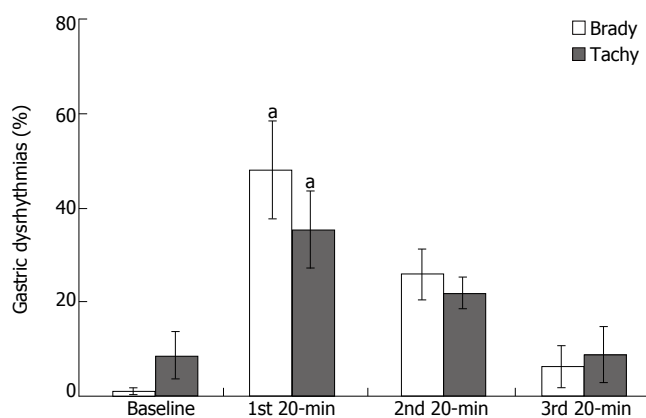


Figure 4 Distribution of gastric dysrhythmias induced by intravenous infusion of vasopressin. Similar percentage of tachygastria and bradycastria was observed, which contributed to the irregularity of gastric myoelectric activity. ANOVA, ^a $P < 0.05$ vs baseline and 3rd 20-min period ($n = 7$).

Effect of vasopressin on GMA

The regularity of GMA was $90.1\% \pm 5.0\%$ at baseline. It was reduced to $13.9\% \pm 3.9\%$ and $52.2\% \pm 6.4\%$ during the following two 20-min periods with and after infusion of vasopressin ($P < 0.01$), and returned to $84.7\% \pm 9.2\%$ during the last 20-min period (Figure 3).

The decrease in the regularity of GMA was attributed to an increase in both bradycastria ($48.0\% \pm 10.4\%$ vs $1.1\% \pm 0.8\%$) and tachycastria ($35.4\% \pm 8.2\%$ vs $8.7\% \pm 5.0\%$) (Figure 4).

Effect of vasopressin on strain gauge energy

The strain gauge energy was significantly decreased during the 20-min infusion of vasopressin. The total energy was 48.4 ± 1.3 dB at baseline, and decreased to 36.6 ± 4.5 dB, 44.5 ± 4.3 dB, 45.8 ± 1.9 dB during the following consecutive three 20-min periods during and after the infusion (vs baseline, $P < 0.05$).

Correlation of GMA and strain gauge energy

The decrease in strain gauge energy was positively correlated with the decrease of GMA regularity ($r = 0.96$, $P < 0.05$). Gastric muscle contractions were reduced or disappeared during periods of bradycastria or tachycastria

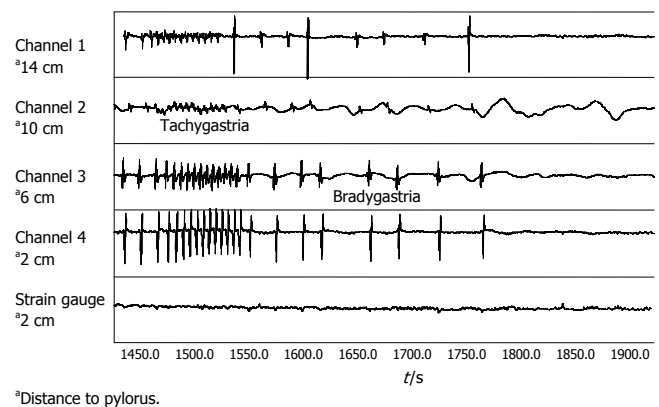


Figure 5 Illustration of gastric dysrhythmias induced by intravenous Vasopressin. Top 4 channels are gastric myoelectrical activities recorded using serosal electrodes implanted along the greater curvature. The bottom tracing shows gastric contractions measured from a strain gauge implanted in the distal stomach parallel to Channel 4. It was noticed that gastric muscle contractions were absent during periods of tachygastria and bradycastria.

(Figure 5). Strain gauge energy was negatively correlated with bradycastria ($r = -0.96$, $P < 0.05$) and tachycastria ($r = -0.95$, $P > 0.05$).

DISCUSSION

Through this study we have found: (1) Intravenous vasopressin at proper doses consistently produced gastric dysrhythmias; (2) strain gauge implanted on gastric wall could reliably detect gastric muscle contractions; gastric muscle contractions were coupled with intrinsic gastric slow waves, and each gastric slow wave was capable of inducing a change in gastric muscle tone; (3) gastric muscle contractions was positively correlated with the regularity of the gastric slow waves; and (4) the reduction or disappearance in gastric muscle contractions was associated with both bradycastria and tachycastria.

Considering the fact that all muscle contractions are coupled and superimposed with myoelectric activities, it is strongly believed that gastric dysrhythmias could cause disturbances in gastric motor functions^[16]. A larger number of clinical studies have been performed recently, and indeed, gastric dysrhythmias have been observed in various motility disorders like gastroparesis, functional dyspepsia and motion sickness, *etc*^[1-7]. In patients with gastroparesis, gastric emptying scintigraphy and EGG were performed concurrently, and it was found that postprandial gastric dysrhythmias correlated with delays in solid phase gastric emptying^[17]. Gastric dysrhythmias may also be in association with the occurrence of GI symptoms^[5,6]. We have noted on many occasions that degeneration of GMA preceded the occurrence of nausea and vomiting in our previous canine studies. Other investigators also reported similar findings^[5,6,18], and all these indicate a possible causative role for the dysrhythmia in the production of nausea and vomiting.

Only few studies have been conducted to qualitatively assess the association between gastric dysrhythmias and gastric motility and there have been no quantitative studies. The first report on tachycastria in humans was published by Telander *et al*^[11]. They reported a 5 month-

old male infant suffering from severe gastric retention and symptoms of intractable nausea, vomiting and weight loss. The symptoms were attributed to impaired motor function of the stomach. The patient underwent resection of the distal $\frac{3}{4}$ of the stomach. The excised tissues were studied *in vitro* by means of intracellular electrodes, and abnormally fast slow waves (> 5 cpm) were detected^[11]. The association between gastric dysrhythmia and gastric motor disorders was further substantiated by other studies. You *et al*^[12] observed tachygastria in a 26-year-old woman with persistent nausea, vomiting and abdominal pain who was found to have severe impairment of antral motor functions. In contrast to tachygastria, data on bradygastria are inconsistent. Van Der Schee *et al* observed bradygastria in dogs and found that it was correlated with strong antral contractions^[13], while Abell *et al*^[14] induced bradygastria in humans with glucagon, and found that it was associated with absence of antral contraction.

In our present study, we attached a strain gauge to the site of recording electrodes, which allows us to study the direct correlation of gastric contractions and GMA. In comparison to the traditional intraluminal manometry, strain gauge is more sensitive and more direct in the detection of subtle contractions of gastric wall than any other methods. In addition, gastric dysrhythmias in this study were assessed quantitatively rather than qualitatively in the previous studies. Our results indicate that both tachygastria and bradygastria are negatively associated with gastric muscle contractile activity, which is consistent with most of the previous report^[11-14].

The exact mechanisms causing gastric dysrhythmias have not been fully understood, though it is well known that GMA is generated by interstitial cells of Cajal, and is influenced by central and autonomic nerve systems, certain hormones and peptides. Tachygastria and bradygastria are different. Tachygastria usually originates from distal stomach, and can be considered as an ectopic rhythm, while transient or persistent bradygastria usually originates from the region of the normal pacemaker at a reduced frequency. Gastric dysrhythmia can be induced experimentally with various agents including vasopressin^[14,19-21]. Vasopressin is a peptide released into the peripheral circulation from the pituitary during experimental motion sickness, and has been confirmed to induce gastric and intestinal dysrhythmia and symptoms of nausea and vomiting^[20-22]. An elevated plasma level of vasopressin was reported in postsurgical patients with impaired gastrointestinal motility^[23]. Vasopressin was known to reduce mesenteric blood flow, which was frequently used in patients with esophageal variceal bleeding^[24], and this vasoconstrictive effect might be the mechanism involved in the induction of GI dysrhythmias.

Gastric myoelectric activity can be recorded in several ways including intraluminal electrodes, surface electrogastrogram and serosal electrodes. Placement of intraluminal electrodes is invasive and uncomfortable. It is very difficult to direct the electrodes to specific sites of stomach and electrodes can be easily dislodged. EGG is a convenient summation technique with various disadvantages: (1) derived data on frequency and power are not strictly region specific; (2) susceptible to a variety

of artifacts; (3) cannot discriminate chaotic rhythm from uncoupling; and (4) susceptible to myoelectric interference from other gut organs, and cutaneously acquired signals may not be gastric in origin^[25]. Serosal recordings, on the other hand, do not have these potential technical problems, and is the most reliable *in vivo* GMA recording except for the need for surgery. This method has allowed us to put the recording electrodes and strain gauge at the same location, thus providing an excellent tool for the study of the direct correlation of dysrhythmias and gastric contraction, which has never been done before.

Another interesting finding from this study is that under control conditions each gastric slow wave seems capable of inducing a certain level of mechanical contractions in gastric muscle. These contractions are so subtle that they could only be reflected by strain gauge, other than other techniques, including intraluminal manometry. This observation is inconsistent with traditional view that only gastric slow waves superimposed with spike activities are capable of inducing gastric contractions.

In summary, in this study we have observed that intravenous administration of vasopressin could impair gastric slow waves and gastric muscle contractions. Experimentally induced tachygastria and bradygastria are negatively correlated with the gastric muscle contractions, suggesting that gastric dysrhythmias, either tachygastria or bradygastria, may be indicative of gastric hypomotility.

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