

RAPID COMMUNICATION

Xeroderma pigmentosum group D 751 polymorphism as a predictive factor in resected gastric cancer treated with chemo-radiotherapy

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Abstract

AIM: To evaluate the potential association of xeroderma pigmentosum group D (XPD) codon 751 variant with outcome after chemo-radiotherapy in patients with resected gastric cancer.

METHODS: We used PCR-RFLP to evaluate the genetic XPD *Lys751Gln* polymorphisms in 44 patients with stage III (48%) and IV (20%) gastric cancer treated with surgery following radiation therapy plus 5-fluorouracil/leucovorin based chemotherapy.

RESULTS: Statistical analysis showed that 75% (12 of 16) of relapse patients showed *Lys/Lys* genotype more frequently ($P = 0.042$). The *Lys* polymorphism was an independent predictor of high-risk relapse-free survival from Cox analysis (HR: 3.07, 95% CI: 1.07-8.78, $P = 0.036$) and Kaplan-Meier test ($P = 0.027$, log-rank test).

CONCLUSION: XPD *Lys751Gln* polymorphism may be an important marker in the prediction of clinical outcome to chemo-radiotherapy in resected gastric cancer patients.

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Key words: Xeroderma pigmentosum group D gene; Polymorphism; Gastric cancer; Radiotherapy

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INTRODUCTION

The xeroderma pigmentosum group D (XPD) gene encodes a protein required for nucleotide excision repair (NER). This product recognizes and repairs a wide range of structurally unrelated lesions such as bulky adducts caused by UV light, environmental agents, cross links and oxidative damage^[1,2]. Moreover, XPD gene is one of the components of basal transcription factor IIIH (TFIIH), participating also in transcription initiation. Since XPD is involved in both transcription and NER, it may be able to repair other types of damage, such as radiation therapy-induced damage. Because XPD interacts with many different proteins as part of TFIIH transcription factor, amino acid variants in different domains of XPD, such as 683 and 751, may affect different protein interactions, and result in expression of different phenotypes^[3,4].

Intrinsic and acquired resistance to cancer chemotherapeutic agents results from polymorphisms in genes encoding DNA repair enzymes. Differences in responsiveness of cancer cells to anticancer agents can be differently affected by changes in repair efficiency. Clinical outcomes after chemotherapy may be influenced by pharmacogenetic polymorphisms in DNA repair enzymes. In this sense, three single nucleotide polymorphisms (SNP) in the XPD gene (in codons 156, 312, and 751) are related with different DNA repair capacity^[5-7]. Thus, variants generated by amino acids can change *Asp312Asn* (exon 10, G > A substitution) and *Lys751Gln* (exon 23 A > C substitution) located in conserved regions of the XPD protein, and are associated with lower efficiency of damage repair and significantly higher background frequency of apoptotic cells in irradiated lymphocytes^[8].

Although gastrectomy is the only potentially curative treatment in gastric-cancer patients, the overall survival results remain unsatisfactory. The main factor accounting for high mortality is relapse after surgical resection. During the past few decades, the principle of combined treatment modality has been developed and applied in various solid tumors including gastric cancer. In order to prevent recurrence and increase the cure rate of gastric-cancer after surgery, multiple studies using variable treatment modalities

ties have been undertaken. One of the landmark studies reported that chemo-radiotherapy can significantly improve survival after resection of stage IB to stage IV gastric cancers^[9]. Chemo-radiotherapy has been increasingly recognized as a standard of care since then^[10,11]. However, whether adjuvant chemo-radiotherapy can prolong the survival of patients with extensive lymph node dissection remains debatable. In this paper, we evaluated the potential role of XPD codon 751 variant in the outcome of 44 patients with resected gastric cancer after chemo-radiotherapy.

MATERIALS AND METHODS

Patients

Characteristics of the patients are listed in Table 1. From October 1992 to January 1999, 44 patients consisting of 32 men (73%) and 12 women (27%) with a median age of 60 years (range: 33-77 years) with diagnosis of gastric cancer after having undergone gastrectomy were treated with radiation plus 5-fluorouracil (5-FU) and leucovorin. Thirty-two percent of the patients had stage I - II and 68% stage III-IV gastric cancer at the time of diagnosis (Table 1). The median follow-up time was 46.9 mo (range: 5.33 to 124.06 mo). The median time to progression was 37.5 mo (range: 2.3 to 95.4 mo). This study was conducted in Navarra Hospital Center, and informed consent was obtained from all the patients for using their tissues. The clinicopathological information of each subject was obtained from the tumor registry at Navarra Hospital.

Treatment schedule

The patients received postoperative treatment with 5-FU plus leucovorin and local radiation 20-40 d after gastrectomy. This chemotherapy regimen developed by the North Central Cancer Treatment Group^[12] was used before and after radiation. Chemotherapy (fluorouracil, 425 mg/m² per day, and leucovorin, 20 mg/m² per day) was initiated on d 1 followed by chemo-radiotherapy on d 28 after the initial cycle of chemotherapy. Chemo-radiotherapy consisted of 4500 cGy of radiation at 180 cGy/d, 5 d/wk for 5 wk, with 5-FU (400 mg/m² per day) and leucovorin (20 mg/m² per day) on the first four and the last three days of radiotherapy. One month after radiotherapy, two 5-d cycles of 5-FU (425 mg/m² per day) plus leucovorin (20 mg/m² per day) were given every other month.

The 4500 cGy of radiation was delivered in 25 fractions (5 d/wk) to the tumor bed, regional nodes and 2 cm beyond the proximal and distal margins of resection. The adjuvant treatment was performed as previously described^[9].

Samples and DNA extraction

Surgical specimens (paraffin blocks) were cut into 5 µm-thick sections for molecular analysis.

Three tissue sections were transferred into a micro centrifuge tube and 1.2 mL of xylene was added. After centrifugation at 14 000 r/min for 5 min at room temperature (RT), the supernatant was removed. Subsequently, the tissue samples were washed in 1.2 mL of 96 mL/L

Table 1 Characteristics of the patients

Baseline factors	n (%)
Sex	
Male	32 (73)
Female	12 (27)
Age (yr)	
≤ 50	16 (36)
51-64	13 (30)
≥ 65	15 (34)
Median (Range)	60 (33-77)
Stage at diagnostic	
I - II	14 (32)
III A + III B	21 (48)
IV	9 (20)
Histotype	
Intestinal	9 (20)
Diffuse	35 (80)
Grading	
I	3 (7)
II	8 (18)
III	33 (75)
Tumor location	
Cardia	9 (21)
Fundus	9 (21)
Body	10 (24)
Antrum	14 (33)
Gastrectomy	
R0	35 (80)
R1	8 (18)
R2	1 (2)
XPD 751	
Lys/Lys	19 (51)
Lys/Gln	14 (38)
Gln/Gln	4 (11)
Unknown	7

ethanol. After centrifugation at 14 000 r/min for 5 min at RT, the supernatant was discarded. The washing procedure was repeated another time. The samples were dried for 3-5 min, in a vacuum pump (any letter between were and dried).

DNA was extracted using the QIAamp DNA Mini Kit (Qiagen-IZASA- Barcelona Spain). The deparaffinized samples were re-suspended in 180 µL of ATL buffer plus 20 µL of proteinase K and incubated overnight at 56°C. The samples were processed according to the provided protocol. The purified DNA was finally eluted in a total volume of 150 µL. DNA yield was quantified by NanoDrop 3.0.0 (Nucliber, Wilmington, Delaware USA).

About 7-509 mg/L of DNA was extracted and 45-200 ng of genomic DNA was used as template. PCR/RFLP-based assays were performed as described previously^[5] with slight modifications. The PCR product (20 µL) was digested with 25 units of Pst I enzyme (New England BioLabs, -IZASA, S.A, Barcelona-, Spain) in 50 µL reaction mixture for 1 h. The digestion product was visualized in 3% agarose gel (Pronadisa, Madrid, Spain). The wild-type homozygote was defined by 104- and 220-bp banding patterns; the heterozygote by 63-, 104-, 157-, and 220-bp fragments; and variant homozygote

Table 2 Toxicity evaluated

Toxicity grade III-IV	n (%)
No	28 (64)
Yes	16 (36)
Toxicity type	
Hematological	10 (62)
Digestive	3 (19)
Others	3 (19)

Table 3 Relapse site

Relapse	n (%)
No	24 (54.5)
Yes	20 (45.5)
Relapse sites	
Local	5 (25)
Distant	10 (50)
Local + Distant	5 (25)

by 63-, 104-, and 157-bp fragments. Some samples were noted as “unknown” because they could not be amplified due to a relatively frequent occurrence of PCR inhibitory substances in samples prepared with this DNA extraction method (Table 1). Furthermore, three DNA samples corresponding to each genotype selected by direct sequencing were used. The results were in concordance with RFLP genotyping.

Statistical analysis

This study was designed to analyze the role of XPD polymorphism in the prediction of relapse. Relapse free-survival (RFS) was defined as the time from the start of chemotherapy to the first evidence of disease progression. RFS was calculated using the Kaplan-Meier method. Contingency tables and chi-square test (χ^2) were used to summarize the association of relapse with XPD polymorphism. All *P* values were two-sided. Cox proportional hazards models were also used to evaluate the different variables considered. All statistical tests were conducted by SPSS software 11.0 version for Windows (SPSS, Inc. Chicago). *P* ≤ 0.05 was considered statistically significant.

RESULTS

Thirty-seven patients were evaluated for the *Lys751Gln* polymorphism. Seven patients were not assessed due to poor sample extraction quality. Fifty-one percent (19 of 37) of the patients were homozygous for the *Lys/Lys* genotype, 38% (14/37) heterozygous for *Lys/Gln*, and 11% (4 of 37) homozygous for the glutamine variant (Table 1). The *751Gln* allele frequency was 0.33, similar to that observed in other studies^[5,7].

Sixteen patients (36%) developed grade III-IV toxicity. Hematological toxicity was found in 62% (10/16) patients compared with gastrointestinal and other toxicity in 19% (3/16) patients (Table 2). However, there was no

Table 4 Results of chi-square test for disease relapse

Relapse	XPD			Stage	
	<i>Lys</i>	<i>Lys/Gln</i>	<i>Gln/Gln</i>	I + II	III A + III B + IV
No	7	11	3	11	13
Yes	12	3	1	3	17
<i>P</i>	0.042			0.050	

Table 5 Unadjusted XPD genotype and relapse-free survival (RFS)

XPD	RFS (median)	Hazard Ratio	95% CI
<i>Lys/Lys</i>	11.79	1.0	Reference
<i>Lys/Gln</i>	14.33	0.35	0.11 to 1.09
<i>Gln/Gln</i>	23.33	0.25	0.03 to 1.96

Table 6 Adjusted Cox multivariate analysis for RFS

XPD	RFS (median)	Hazard ratio	95% CI	<i>P</i>
<i>Lys/Lys</i>	11.79	3.07	1.07 to 8.78	0.036
<i>Lys/Gln</i> + <i>Gln/Gln</i>	20.26	1.0	Reference	

association between XPD genotype and toxicity (data not shown).

Gastrectomy with D1 lymphadenectomy was performed in all patients. Of the 44 patients, 20 (45.5%) had relapse and 17 (85%) of them died. The first relapse site was local in 5, distant in 10, and both local and distant in 5 (Table 3).

Statistical analysis showed that 75% (12 of 16) of relapse patients showed *Lys/Lys* genotype more frequently (*P* = 0.042, Table 4).

Other clinico-pathological features such as histology, grading and localization were not significant (data not shown).

The relative risk of progression (with *Lys/Lys* genotype used as the reference) was 0.35 (95% CI: 0.11-1.09) for patients carrying the *Lys/Gln* genotype and 0.25 (95% CI: 0.03-1.96) for the *Gln/Gln* group (Table 5).

Only XPD genotype and stage to diagnosis showed a significant relation with cancer relapse and were evaluated as potential predictors of RFS by Cox test. We used the combined genotypes *LysGln/GlnGln* as reference in these analyses and found that the *Lys/Lys* genotype was more strongly associated with disease progression (*P* = 0.036) than the combined *LysGln/GlnGln* genotype (adjusted HR: 3.07, 95% CI: 1.07-8.78) (Table 6, Figure 1).

DISCUSSION

Most cases of gastric cancer are diagnosed at an advanced stage with poor prognosis. Surgery remains the only potentially curative treatment, but it is associated with a high rate of local recurrence and distant metastases. When irradiation is combined with surgical resection for all or a

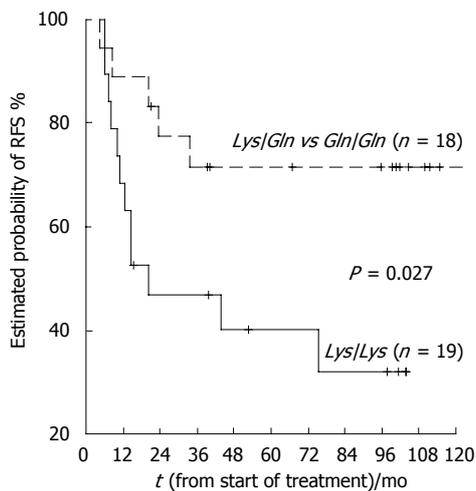


Figure 1 XPD751 polymorphism and RFS under radio-5Fu/Lv treatment.

majority of tumors, both survival and local control appear to be better than those for the unresected patients^[13]. Epidemiological studies have shown that *Gln* variant at XPD-751 polymorphism is partially able to repair DNA in lung cancer^[7,14]. In this sense, cancer treatment is to cause DNA damage and tumor cell apoptosis. *Gln* variant seems to show a lower ability to repair DNA damage and cancer patients are more sensitive to chemo-radiotherapy. In contrast, the presence of allele *Lys* represents a positive effect in different DNA repair pathways and a possible poor response to cancer treatment. In this study, we found a significant relationship between clinical response to chemo-radiotherapy and the XPD *Lys751Gln* polymorphism. Patients with the *Lys/Lys* genotype were more likely to have relapse compared to those with the combined *Lys/Gln* and *Gln/Gln* genotype.

Scientific publications are available but with diverging results in the XPD751 polymorphism and cancer risk or treatment efficiency^[3,5-7]. Thus, decreased DNA repair has been associated with *Lys* allele^[6]. Epidemiological study has reported association of this allele with a higher risk of basal cell carcinoma^[5] even though no significant relationship between this polymorphic gene and DNA repair proficiency has been reported^[15,16]. Spitz *et al*^[7] and other research group^[17] reported a suboptimal DNA repair capacity (DRC) particularly in subjects who were homozygous for 751*Gln* alleles. Moreover, a recently study has found a tendency toward a higher background frequency of apoptotic cells in irradiated lymphocytes carrying variant homozygote *Gln* at XPD codon 751^[8]. Our results are in agreement with these studies, suggesting that heterozygote and homozygote *Gln* alleles may therefore increase their clinical response to suboptimal DNA damage repair after treatment.

Repair of DNA damage is a complex process^[1,18,19]. Functional protein of XPD could be affected by a structural change (*Lys* → *Gln*) from a basic to a polar amino acid change located at about 50-base upstream from the poly (A) signal^[5]. Moreover, close proximity in the genome between XPD and other polymorphic DNA repair genes such as X-ray repair cross complementing (XRCC1) and excision repair cross complementing (ERCC1) may influ-

ence the interaction between them leading to different or synergic DNA repair capability. A recent molecular epidemiology study has genotyped 44 SNPs in 20 genes involved in four DNA damage repair pathways, showing that homozygous *Gln751Gln* and other gene variants genotyped are highly associated with lung cancer risk^[20]. In the same way, variant *Gln* allele and some nucleotide and base excision repair (NER and BER, respectively) variant gene members are associated with increased level of polycyclic aromatic hydrocarbon-DNA adducts^[15,21].

In conclusion, XPD *Lys751Gln* polymorphism may be an important marker of genotoxicity that can predict the clinical outcome of resected gastric cancer treated with chemo-radiotherapy.

REFERENCES

- 1 Sancar A. DNA repair in humans. *Annu Rev Genet* 1995; **29**: 69-105
- 2 Flejter WL, McDaniel LD, Johns D, Friedberg EC, Schultz RA. Correction of xeroderma pigmentosum complementation group D mutant cell phenotypes by chromosome and gene transfer: involvement of the human ERCC2 DNA repair gene. *Proc Natl Acad Sci USA* 1992; **89**: 261-265
- 3 Stoehlmacher J, Park DJ, Zhang W, Yang D, Groshen S, Zahedy S, Lenz HJ. A multivariate analysis of genomic polymorphisms: prediction of clinical outcome to 5-FU/oxaliplatin combination chemotherapy in refractory colorectal cancer. *Br J Cancer* 2004; **91**: 344-354
- 4 Takayama K, Salazar EP, Broughton BC, Lehmann AR, Sarasin A, Thompson LH, Weber CA. Defects in the DNA repair and transcription gene ERCC2(XPD) in trichothiodystrophy. *Am J Hum Genet* 1996; **58**: 263-270
- 5 Dybdahl M, Vogel U, Frentz G, Wallin H, Nexø BA. Polymorphisms in the DNA repair gene XPD: correlations with risk and age at onset of basal cell carcinoma. *Cancer Epidemiol Biomarkers Prev* 1999; **8**: 77-81
- 6 Lunn RM, Helzlsouer KJ, Parshad R, Umbach DM, Harris EL, Sanford KK, Bell DA. XPD polymorphisms: effects on DNA repair proficiency. *Carcinogenesis* 2000; **21**: 551-555
- 7 Spitz MR, Wu X, Wang Y, Wang LE, Shete S, Guo Z, Lei L, Mohrenweiser H, Wei Q. Modulation of nucleotide excision repair capacity by XPD polymorphisms in lung cancer patients. *Cancer Res* 2001; **61**: 1354-1357
- 8 Rzeszowska-Wolny J, Polanska J, Pietrowska M, Palyvoda O, Jaworska J, Butkiewicz D, Hancock R. Influence of polymorphisms in DNA repair genes XPD, XRCC1 and MGMT on DNA damage induced by gamma radiation and its repair in lymphocytes in vitro. *Radiat Res* 2005; **164**: 132-140
- 9 Macdonald JS, Smalley SR, Benedetti J, Hundahl SA, Estes NC, Stemmermann GN, Haller DG, Ajani JA, Gunderson LL, Jessup JM, Martenson JA. Chemoradiotherapy after surgery compared with surgery alone for adenocarcinoma of the stomach or gastroesophageal junction. *N Engl J Med* 2001; **345**: 725-730
- 10 Macdonald JS. Adjuvant therapy for gastric cancer. *Semin Oncol* 2003; **30**: 19-25
- 11 Macdonald JS. Clinical overview: adjuvant therapy of gastrointestinal cancer. *Cancer Chemother Pharmacol* 2004; **54** Suppl 1: S4-S11
- 12 Poon MA, O'Connell MJ, Moertel CG, Wieand HS, Cullinan SA, Everson LK, Krook JE, Mailliard JA, Laurie JA, Tschetter LK. Biochemical modulation of fluorouracil: evidence of significant improvement of survival and quality of life in patients with advanced colorectal carcinoma. *J Clin Oncol* 1989; **7**: 1407-1418
- 13 Carrato A, Gallego-Plazas J, Guillen-Ponce C. Adjuvant therapy of resected gastric cancer is necessary. *Semin Oncol* 2005; **32**: S105-S108
- 14 Palli D, Russo A, Masala G, Saieva C, Guarrera S, Carturan

- S, Munnia A, Matullo G, Peluso M. DNA adduct levels and DNA repair polymorphisms in traffic-exposed workers and a general population sample. *Int J Cancer* 2001; **94**: 121-127
- 15 **Duell EJ**, Wiencke JK, Cheng TJ, Varkonyi A, Zuo ZF, Ashok TD, Mark EJ, Wain JC, Christiani DC, Kelsey KT. Polymorphisms in the DNA repair genes XRCC1 and ERCC2 and biomarkers of DNA damage in human blood mononuclear cells. *Carcinogenesis* 2000; **21**: 965-971
- 16 **Moller P**, Knudsen LE, Frenz G, Dybdahl M, Wallin H, Nexø BA. Seasonal variation of DNA damage and repair in patients with non-melanoma skin cancer and referents with and without psoriasis. *Mutat Res* 1998; **407**: 25-34
- 17 **Qiao Y**, Spitz MR, Guo Z, Hadeyati M, Grossman L, Kraemer KH, Wei Q. Rapid assessment of repair of ultraviolet DNA damage with a modified host-cell reactivation assay using a luciferase reporter gene and correlation with polymorphisms of DNA repair genes in normal human lymphocytes. *Mutat Res* 2002; **509**: 165-174
- 18 **Miller MC 3rd**, Mohrenweiser HW, Bell DA. Genetic variability in susceptibility and response to toxicants. *Toxicol Lett* 2001; **120**: 269-280
- 19 **Eisen JA**, Hanawalt PC. A phylogenomic study of DNA repair genes, proteins, and processes. *Mutat Res* 1999; **435**: 171-213
- 20 **Zienolddiny S**, Campa D, Lind H, Ryberg D, Skaug V, Stangeland L, Phillips DH, Canzian F, Haugen A. Polymorphisms of DNA repair genes and risk of non-small cell lung cancer. *Carcinogenesis* 2006; **27**: 560-567
- 21 **Matullo G**, Palli D, Peluso M, Guarrera S, Carturan S, Celentano E, Krogh V, Munnia A, Tumino R, Polidoro S, Piazza A, Vineis P. XRCC1, XRCC3, XPD gene polymorphisms, smoking and (32)P-DNA adducts in a sample of healthy subjects. *Carcinogenesis* 2001; **22**: 1437-1445

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