



GASTRIC CANCER

Phosphorylated vasodilator-stimulated phosphoprotein is localized on mitotic spindles of the gastric cancer cell line SGC-7901

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Supported by National Natural Science Foundation of China, No. 30340036 and 30470891; Startup Grant from Jiangsu University, and Grant of Zhenjiang Key Institute of Clinical Laboratory Medicine (SH2006066)

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Received: 2006-08-24

Accepted: 2006-11-09

spindles of the gastric cancer cell line SGC-7901. *World J Gastroenterol* 2006; 12(46): 7478-7481

<http://www.wjgnet.com/1007-9327/12/7478.asp>

Abstract

AIM: To elucidate the localization of vasodilator stimulated phosphoprotein (VASP), a cytoskeletal organizing protein and a substrate of protein kinases A and G in mitotic gastric cancer cells.

METHODS: Immunofluorescence microscopy was used to observe the localization of α -tubulin, VASP and Ser157 phosphorylated VASP (p-VASP) in interphase of mitotic gastric cancer of the cell line SGC-7901.

RESULTS: Immunofluorescence staining showed that p-VASP but not VASP was co-localized with α -tubulin on spindle poles and fibers in prophase, metaphase and anaphase of the mitotic process of the gastric cancer cell line SGC-7901. H89, an inhibitor of protein kinases A and G, had no effect on the localization of p-VASP on the spindles.

CONCLUSION: VASP may play a role in assembling and stabilizing the mitotic spindle of cells, and phosphorylation of the protein is the precondition for it to exert this function.

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Key words: Phosphorylated vasodilator-stimulated phosphoprotein; Localization; Mitotic spindle; Gastric cancer cell

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INTRODUCTION

The spindle is a vital apparatus of mitotic cells. The mechanism of its formation has attracted great research attention. In centrosome-containing cells, centrosomes are thought to be instrumental for organization of the spindle poles and determination of both the microtubule polarity and the spindle axis^[1]. The spindle poles often contain widely different amounts of known spindle-pole components, such as γ -tubulin. At the beginning of mitosis, γ -tubulin is recruited to the poles and forms a γ -tubulin ring complex (γ TuRC), which can collect both α and β tubulins to form spindle fibers^[2,3]. At the same time, accessory proteins also join the process. One of these types is microtubule-associated proteins (MAPs). Three major neural MAPs (MAP1, MAP2 and tau) and one non-neural MAP (MAP4) have been identified and well characterized at the biochemical level^[4,5]. These MAPs possess the ability to stimulate tubulin polymerization and to bind to microtubules *in vitro* and are believed to play an important part in the regulation of microtubule formation and stabilization *in vivo*^[6].

Vasodilator-stimulated phosphoprotein (VASP) is a proline-rich founding member of the Ena (drosophila enabled)/VASP family, a group of multifunctional proteins associated with regulation of actin. VASP was first found in platelets and is known to be a substrate for cAMP dependent protein kinase (PKA) and cGMP dependent protein kinase (PKG) in a variety of cells, such as vascular smooth muscle cells and endothelial cells^[7,8]. It is also known to be actin cytoskeletal regulatory proteins. In a variety of cells like vascular endothelial cells, platelets, smooth muscle cells, and fibroblasts, VASP has been found to be highly expressed where it can be found in focal adhesions, along stress fibers, and in the areas with highly dynamic membrane activity, such as extending lamellipodia and filopodia^[9-12]. *In vivo* evidence indicates that VASP is a crucial factor in the formation of actin filaments and the integration of signals transmitted between

cytoskeleton, cytoskeleton-membrane interface and the two cyclic nucleotide signal transduction pathways^[9]. Even though increasing data address the importance of VASP, the biological role of this protein still remains largely unknown. We have recently found that phosphorylated VASP (p-VASP) is co-located with α -tubulin on the spindles of mitotic cells, indicating that this cytoskeletal organizing protein may also have a role in assembling and stabilizing the mitotic spindles.

MATERIALS AND METHODS

Cell line

Human gastric epithelial cell line SGC-7901 was provided by Institute of Cell Biology of China (Shanghai, China).

Reagents

Dulbeccos's modified Eagle's medium (DMEM) was from Gibco (Grand Island, NY). Newborn calf serum (NBCS) was from Minhai Bio-engineering Co., LTD (Lanzhou, China). Antibodies against VASP (Cat. No. sc-46668) and phosphorylated VASP (Cat. No. sc-23506-R) were from Santa Cruz Biotechnology (Santa Cruz, CA). Antibody against α -tubulin (BM1452) was from Wuhan Boster Biological Technology Ltd (Wuhan, China). FITC- and TRITC- conjugated secondary antibodies were from Jackson ImmunoResearch Laboratories (West Grove, PA). Protein kinase A inhibitor H89 (Cat. No. 371963) was from Calbiochem (San Diego, CA). Nuclear fluorochrome Hoechst 33342 was from Sigma (St. Louis, Missouri).

Immunofluorescent microscopy

The cells grown on cover slips were incubated to reveal nuclei with 0.2 μ mol/L Hoechst 33 342 for 10 min. Then, the cells were fixed with freshly prepared 40 g/L paraformaldehyde in PBS at 4°C overnight (o/n). After being penetrated with 30 mL/L Triton \times 100 and blocked with 30 g/L BSA, the cells were incubated with primary antibodies at 4°C o/n, and then with FITC- or TRITC- conjugated secondary antibodies for 1 h at room temperature (RT), and washed three times after each time of incubation. For immunofluorescence co-staining of α -tubulin and p-VASP, the cells on cover slips were subsequently incubated with mouse monoclonal antibody against α -tubulin at 4°C o/n, rabbit polyclonal antibody against p-VASP for 2 h at RT, goat anti-mouse IgG conjugated with FITC for 1 h at RT, and finally TRITC-conjugated goat anti-rabbit IgG for 1 h at RT, and washed three times after each time of incubation. The cover slips were mounted in 900 mL/L glycerol and 100 mL/L Tris-HCl (25 mmol/L, pH 7.5). The fluorescence of FITC (green), TRITC (red), and Hoechst 33 342 (blue) was observed under a fluorescence microscope with a CCD camera (Leica).

RESULTS

Distribution of p-VASP and α -tubulin in interphase of SGC-7901 cells

Immunofluorescence staining showed that both p-VASP

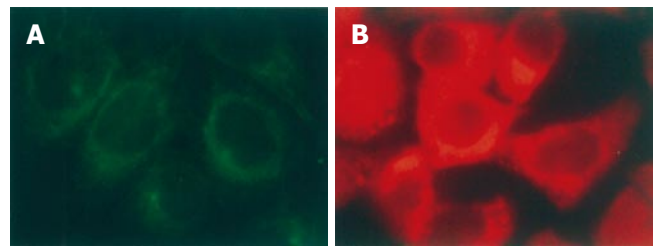


Figure 1 Distribution of α -tubulin (A) and p-VASP (B) in interphase of SGC-7901 cells (immunofluorescent staining \times 200). α -tubulin was stained with monoclonal antibody against the protein. p-VASP was stained with polyclonal antibody specifically against VASP phosphorylated at Ser157. Both α -tubulin and p-VASP were located in cytoplasm of the cells.

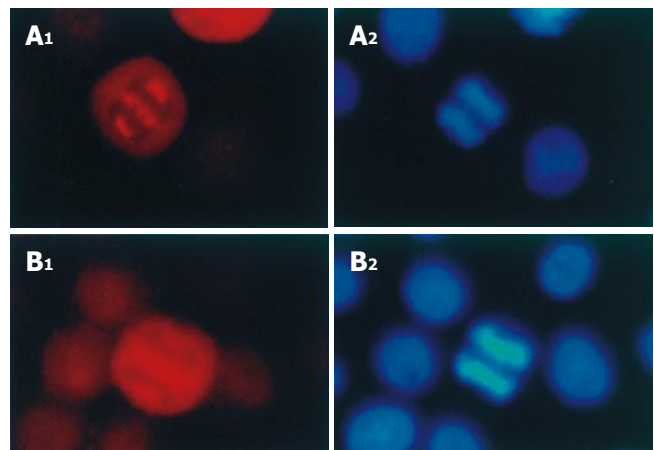


Figure 2 Distribution of p-VASP (A) and VASP (B) in mitotic SGC-7901 cells (\times 200). p-VASP and VASP in mitotic SGC-7901 cells were immunofluorescently stained with polyclonal and monoclonal antibodies respectively. The chromosomes were stained with Hoechst 33342.

and α -tubulin were mainly distributed in the cytoplasm of the cells in interphase (Figure 1A and B)

p-VASP but not VASP was located with α -tubulin on the spindle of mitotic cells

Immunofluorescence staining showed that p-VASP but not VASP was localized both on the spindle poles and on the spindle fibers of mitotic cells (Figure 2A and B). In prophase, the chromosomes became condensed with formation of p-VASP-containing spindle poles (Figure 3A). In metaphase, the chromosomes converged at the metaphase plate (Figure 3B) and the p-VASP-containing spindle poles were located at the opposite sides of chromosomes and the p-VASP-containing spindle fibers irradiated to the chromosomes (Figure 3B). In anaphase, daughter chromosomes were separated and moved toward the opposite p-VASP-containing spindle poles (Figure 3C). Both p-VASP-containing spindle poles and fibers could be seen in this phase (Figure 3C). In telophase, p-VASP-containing spindle disappeared and new daughter nuclei formed (Figure 3D). However, p-VASP could be detected at the cleavage furrow between two separating cells (Figure 3D). Immunofluorescence co-staining showed that α -tubulin was co-localized with p-VASP throughout all phases of mitosis (Figure 3A-D).

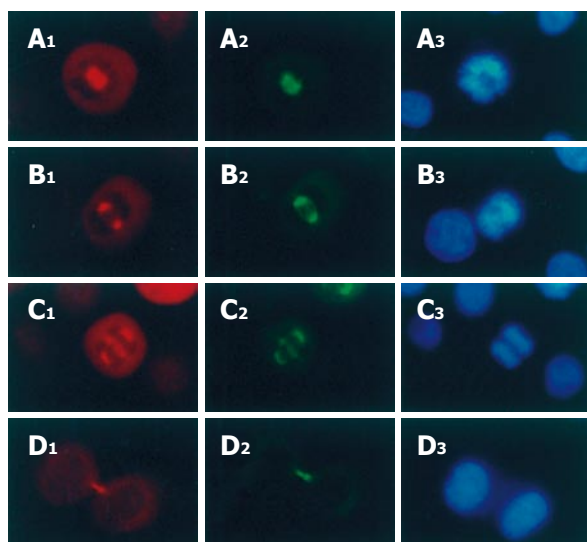


Figure 3 p-VASP co-localization with α -tubulin and chromosomes on spindles of mitotic SGC-7901 cells ($\times 200$). p-VASP and α -tubulin were stained with polyclonal and monoclonal antibodies respectively. **A1-D1**: p-VASP localization in prophase, metaphase, anaphase and telophase respectively; **A2-D2**: α -tubulin localization in the same phases; **A3-D3**: chromosomes of the cells were stained with Hoechst 33342.

PKA and PKG inhibitor had no effect on the distribution of p-VASP in mitotic spindle

H89, an inhibitor of PKA and PKG, had no obvious effect on the localization of p-VASP on the mitotic spindle. Immunofluorescent staining of p-VASP could still be seen on the spindles of mitotic cells treated with H89 (10 μ mol/L) for different periods of time (Figure 4).

DISCUSSION

Mitosis is an important event of the cell cycle. During this process, duplicated DNA genetic material is condensed to chromosomes moving into two daughter cells with high fidelity. In eukaryotic cells, the accurate segregation of chromosomes during cell division occurs on the spindle. The formation of the spindle is a complicated process that requires the participation of several types of proteins such as tubulins, spindle-microtubule motor proteins, and microtubule-associated proteins (MAPs)^[13,14]. Even though accumulating data address the dynamic process of spindle formation, the exact mechanism and process still need to be further clarified.

In this experiment, we showed for the first time that p-VASP was localized both on spindle poles and on spindle fibers of the mitotic cells. The localization of p-VASP exerted a dynamic change associated with stages of the cell cycle. In interphase, p-VASP was detected in the cytoplasm. In mitotic cells, p-VASP was localized on the mitotic spindle from prophase to anaphase. In prophase, p-VASP was localized on the poles of the spindle. p-VASP could be detected on both the poles and fibers of the spindles from metaphase to anaphase. The localization of p-VASP in mitotic cells was exactly coincident with the localization of alpha tubulin, suggesting that p-VASP might play a role in the formation of mitotic spindles, which is associated with tubulin.

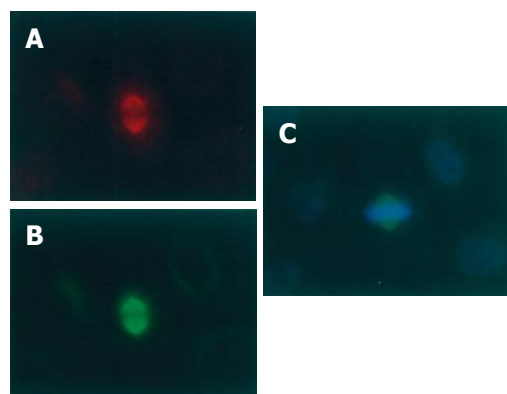


Figure 4 No effect of PKA and PKG inhibitor H89 on location of p-VASP on spindles of mitotic SGC-7901 cells ($\times 200$). SGC-7901 cells were treated with H89 (10 μ mol/L) for 6 h and immunofluorescently stained for p-VASP (**A**) and α -tubulin (**B**). The chromosomes were stained with Hoechst 33342 (**C**).

Microtubule-associated proteins (MAPs) play an important role in the formation of spindles, and can bind to tubulin and assist the assembling and stabilizing of spindles. The microtubule-binding domains of MAPs are divided into three sub-domains: a proline-rich region, an AP sequence region and a short hydrophobic tail^[4]. The importance of the proline-rich region in *in vitro* MAP-induced microtubule assembly has been widely recognized^[15-18]. Tokuraku *et al*^[6] showed that this proline-rich region plays a new role as the protofilament linker in *in vitro* MAP-induced microtubule assembly. Since microtubules consist of tubulin as its major component, the proline-rich region of MAPs may be one of the structural features that permit the protein be able to bind to tubulin. Interestingly, VASP also has a proline-rich region in its structure. This protein is organized in three domains, an Ena-VASP-homology 1 (EVH1) NH₂-terminal domain, a proline-rich central domain, and an EVH2 COOH-terminal domain. The proline-rich region comprises a (GP)₅ sequence presenting as a single or repeated motif in different proteins. It has been confirmed that profilin binds to VASP *in vitro* and cell extracts probably *via* the (GP)₅ repeats^[19]. Considering this structural feature of VASP, it is reasonable to speculate that p-VASP may be able to bind to tubulin and the possible function of the p-VASP-tubulin binding is to stimulate the polymerization of tubulin on spindle fibers and increase the stability of microtubules of the spindles, a role similar to that of MAPs.

Research data indicate that phosphorylation is necessary for the accessory proteins to exert their role in forming and stabilizing the spindles. For example, mutations that cause loss of function of the protein kinase aurora prevent centrosome separation in *Drosophila*^[20], while phosphorylation of Eg5 has been shown to be necessary for its localization on spindle poles^[21]. Since our results showed that p-VASP was a form of this protein detected on spindles, it is possible that phosphorylation is the precondition for VASP to exert its role in spindle formation and stabilization. VASP has a peptide chain containing 380 amino acid residues. Three of them are known as phosphorylation sites, of which two

serine residues are Ser157 and Ser239 respectively and the threonine residue is Thr278. Phosphorylation of VASP at these sites is directly regulated by PKA and PKG, and VASP has also been shown to be directly phosphorylated on Ser157 by protein kinase C (PKC)^[22]. It has been confirmed that VASP phosphorylation is a modulatory event for VASP-dependent regulation of the actin cytoskeleton. The phosphorylation can affect the cytoskeleton structure and may induce morphological changes of cells^[23-25]. Research data also indicate that phosphorylation of VASP on different amino acid residues has different functions. For example, serum-induced VASP phosphorylation at serine157 by both PKA and PKC promotes smooth muscle cell (SMC) proliferation, whereas VASP phosphorylation at serine239 induced by nitric oxide (NO)/PKG is required to induce the inhibitory effects of NO on SMC proliferation^[26]. One interesting phenomenon observed in our experiment was that PKA (also PKG) inhibitor H89 could not prevent the localization of p-VASP on the mitotic spindles, suggesting that VASP on the spindle is not the substrate of PKA and PKG.

In conclusion, p-VASP might play a role in mitotic spindle formation and stabilization, which implies a potential target for controlling the division and proliferation of gastric cancer cells. The exact mechanism and which protein kinase is responsible for the phosphorylation of VASP on the mitotic spindle need to be further investigated.

REFERENCES

1. Heald R, Tournebize R, Habermann A, Karsenti E, Hyman A. Spindle assembly in *Xenopus* egg extracts: respective roles of centrosomes and microtubule self-organization. *J Cell Biol* 1997; **138**: 615-628
2. Schnackenberg BJ, Khodjakov A, Rieder CL, Palazzo RE. The disassembly and reassembly of functional centrosomes in vitro. *Proc Natl Acad Sci USA* 1998; **95**: 9295-9300
3. Zheng Y, Wong ML, Alberts B, Mitchison T. Nucleation of microtubule assembly by a gamma-tubulin-containing ring complex. *Nature* 1995; **378**: 578-583
4. Aizawa H, Emori Y, Murofushi H, Kawasaki H, Sakai H, Suzuki K. Molecular cloning of a ubiquitously distributed microtubule-associated protein with Mr 190,000. *J Biol Chem* 1990; **265**: 13849-13855
5. Chapin SJ, Bulinski JC. Non-neuronal 210 x 10(3) Mr microtubule-associated protein (MAP4) contains a domain homologous to the microtubule-binding domains of neuronal MAP2 and tau. *J Cell Sci* 1991; **98** (Pt 1): 27-36
6. Tokuraku K, Katsuki M, Nakagawa H, Kotani S. A new model for microtubule-associated protein (MAP)-induced microtubule assembly. The Pro-rich region of MAP4 promotes nucleation of microtubule assembly in vitro. *Eur J Biochem* 1999; **259**: 158-166
7. Markert T, Krenn V, Leebmann J, Walter U. High expression of the focal adhesion- and microfilament-associated protein VASP in vascular smooth muscle and endothelial cells of the intact human vessel wall. *Basic Res Cardiol* 1996; **91**: 337-343
8. Draijer R, Vaandrager AB, Nolte C, de Jonge HR, Walter U, van Hinsbergh VW. Expression of cGMP-dependent protein kinase I and phosphorylation of its substrate, vasodilator-stimulated phosphoprotein, in human endothelial cells of different origin. *Circ Res* 1995; **77**: 897-905
9. Haffner C, Jarchau T, Reinhard M, Hoppe J, Lohmann SM, Walter U. Molecular cloning, structural analysis and functional expression of the proline-rich focal adhesion and microfilament-associated protein VASP. *EMBO J* 1995; **14**: 19-27
10. Reinhard M, Halbrügge M, Scheer U, Wiegand C, Jockusch BM, Walter U. The 46/50 kDa phosphoprotein VASP purified from human platelets is a novel protein associated with actin filaments and focal contacts. *EMBO J* 1992; **11**: 2063-2070
11. Rottner K, Behrendt B, Small JV, Wehland J. VASP dynamics during lamellipodia protrusion. *Nat Cell Biol* 1999; **1**: 321-322
12. Svitkina TM, Bulanova EA, Chaga OY, Vignjevic DM, Kojima S, Vasiliev JM, Borisy GG. Mechanism of filopodia initiation by reorganization of a dendritic network. *J Cell Biol* 2003; **160**: 409-421
13. Mack GJ, Compton DA. Analysis of mitotic microtubule-associated proteins using mass spectrometry identifies astrin, a spindle-associated protein. *Proc Natl Acad Sci USA* 2001; **98**: 14434-14439
14. Katsuki M, Tokuraku K, Murofushi H, Kotani S. Functional analysis of microtubule-binding domain of bovine MAP4. *Cell Struct Funct* 1999; **24**: 337-344
15. Ferralli J, Doll T, Matus A. Sequence analysis of MAP2 function in living cells. *J Cell Sci* 1994; **107** (Pt 11): 3115-3125
16. Gustke N, Trinczek B, Biernat J, Mandelkow EM, Mandelkow E. Domains of tau protein and interactions with microtubules. *Biochemistry* 1994; **33**: 9511-9522
17. Aizawa H, Emori Y, Mori A, Murofushi H, Sakai H, Suzuki K. Functional analyses of the domain structure of microtubule-associated protein-4 (MAP-U). *J Biol Chem* 1991; **266**: 9841-9846
18. Mori A, Aizawa H, Saido TC, Kawasaki H, Mizuno K, Murofushi H, Suzuki K, Sakai H. Site-specific phosphorylation by protein kinase C inhibits assembly-promoting activity of microtubule-associated protein 4. *Biochemistry* 1991; **30**: 9341-9346
19. Laurent V, Loisel TP, Harbeck B, Wehman A, Gröbe L, Jockusch BM, Wehland J, Gertler FB, Carlier MF. Role of proteins of the Ena/VASP family in actin-based motility of *Listeria monocytogenes*. *J Cell Biol* 1999; **144**: 1245-1258
20. Glover DM, Leibowitz MH, McLean DA, Parry H. Mutations in aurora prevent centrosome separation leading to the formation of monopolar spindles. *Cell* 1995; **81**: 95-105
21. Blangy A, Lane HA, d'Hérin P, Harper M, Kress M, Nigg EA. Phosphorylation by p34cdc2 regulates spindle association of human Eg5, a kinesin-related motor essential for bipolar spindle formation in vivo. *Cell* 1995; **83**: 1159-1169
22. Wentworth JK, Pula G, Poole AW. Vasodilator-stimulated phosphoprotein (VASP) is phosphorylated on Ser157 by protein kinase C-dependent and -independent mechanisms in thrombin-stimulated human platelets. *Biochem J* 2006; **393**: 555-564
23. Lawrence DW, Pryzwansky KB. The vasodilator-stimulated phosphoprotein is regulated by cyclic GMP-dependent protein kinase during neutrophil spreading. *J Immunol* 2001; **166**: 5550-5556
24. Smolenski A, Poller W, Walter U, Lohmann SM. Regulation of human endothelial cell focal adhesion sites and migration by cGMP-dependent protein kinase I. *J Biol Chem* 2000; **275**: 25723-25732
25. Wei L, Muller S, Ouyang J, Stoltz JF, Wang X. Changes of vasodilator-stimulated phosphoprotein (VASP) and its phosphorylation in endothelial cells exposed to laminar flow. *Clin Hemorheol Microcirc* 2003; **28**: 113-120
26. Chen L, Daum G, Chitaley K, Coats SA, Bowen-Pope DF, Eigenthaler M, Thumati NR, Walter U, Clowes AW. Vasodilator-stimulated phosphoprotein regulates proliferation and growth inhibition by nitric oxide in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 2004; **24**: 1403-1408

S- Editor Wang GP L- Editor Wang XL E- Editor Bai SH