

TOPIC HIGHLIGHT

Jesus K Yamamoto-Furusho, Dr, Series Editor

Genetic factors associated with the development of inflammatory bowel disease

Jesus K Yamamoto-Furusho

Jesus K Yamamoto-Furusho, Inflammatory Bowel Disease Clinic, Department of Gastroenterology, Instituto Nacional de Ciencias Médicas y Nutrición, Vasco de Quiroga 15, Colonia Sección XVI, Tlalpan, C.P. 14000, México, D.F., México

Correspondence to: Jesús K Yamamoto-Furusho, MD, PhD, MSc, Head of Inflammatory Bowel Disease Clinic, Department of Gastroenterology, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán, Vasco de Quiroga 15, Col. Sección XVI, Tlalpan, México, D.F., Mexico. kazuofurusho@hotmail.com
Telephone: +52-55-55733418 Fax: +52-55-56550942

Received: April 19, 2007 Revised: July 28, 2007

Abstract

Crohn's disease (CD) and ulcerative colitis (UC) are complex polygenic disorders, characterized by several genes together with environmental factors contributing to the development of inflammatory bowel disease (IBD). Recent advances in research on genetic susceptibility have allowed the identification of diverse genes at different levels: (1) Innate immunity; (2) Antigen presentation molecules; (3) Epithelial integrity; (4) Drug transporter; (5) Cell adhesion. The application of genetic testing into clinical practice is close and all genetic markers may have several clinical implications: prediction of disease phenotype, molecular classification, prevention of complications, and prognosis.

© 2007 WJG. All rights reserved.

Key words: Genetic; Susceptibility; Inflammatory bowel disease; Ulcerative colitis; Crohn's disease

Yamamoto-Furusho JK. Genetic factors associated with the development of inflammatory bowel disease. *World J Gastroenterol* 2007; 13(42): 5594-5597

<http://www.wjgnet.com/1007-9327/13/5594.asp>

INTRODUCTION

Crohn's disease (CD) and ulcerative colitis (UC) are chronic relapsing inflammatory bowel diseases (IBD) of unknown etiology. CD and UC are complex polygenic disorders, characterized by several genes together with environmental factors contributing to the development of IBD.

A variety of epidemiological and clinical data suggest that genetic factors are intimately involved in the pathogenesis of IBD including familial aggregation pattern of disease with a much higher disease frequency in first degree relatives of affected individuals compared with the general population. Twin studies provide the argument for a genetic basis for IBD, with a much higher rate of disease concordance observed in monozygotic than in dizygotic twins and wide variations in the incidence and prevalence of IBD among different populations^[1].

IBD is now considered a non-Mendelian polygenic disorder with important environmental interactions (e.g., microbial factors, smoking).

There are two main approaches to identifying genes in complex multifactorial diseases: the positional cloning approach based on linkage studies, and the candidate gene approach based on association studies. Linkage analysis studies the cosegregation of the disease with a marker within the families. Linkage analysis allows scanning of the whole genome. Eleven of these total genome scans have been undertaken in IBD, resulting in a number of susceptibility regions on chromosomes 1, 3, 4, 5, 6, 7, 10, 12, 14, 16, 19 and X^[2]. According to their initial date of reporting and independent confirmations, the regions on chromosomes 16q, 12, 6, 14, 5, 19, 1, 16p and 10 have been renamed IBD 1 to IBD 9, respectively. However, new genes have been reported recently. All susceptibility genes discovered can be categorized into different levels of susceptibility: (1) Innate immunity; (2) Human leucocyte antigen (HLA) molecules; (3) Epithelial integrity (4); Drug transporter; and (5) Cell adhesion.

INNATE IMMUNITY

Understanding of innate immunity has progressed enormously with the discovery of many microbial sensors called pattern recognition receptors (PRRs). The toll-like receptor (TLR) and nucleotide oligomerization domain (NOD) receptor families of PRRs appear to play essential roles in mucosal homeostasis and their alterations contribute to the pathogenesis of IBD.

NOD2 is expressed constitutively in macrophages, neutrophils and dendritic cells^[3], as well as in Paneth cells and is induced in epithelial cells^[4]. NOD2 is a cytoplasmic protein that serves as a microbial sensor, and its leucine-rich repeat (LRR) domain is required for recognition of muramyl dipeptide (MDP), a fragment of peptidoglycan

present in bacterial cell walls. The ligand MDP ultimately leads to activation of the transcription factor nuclear factor kappa B (NF- κ B), and induction of proinflammatory cytokines^[5,6]. Specific mutations of the NOD2 gene have been definitively associated with increased susceptibility to ileal Crohn's disease in Western (but not Asian) populations: Arg702Trp, Gly908Arg, and leu1007fsinsC (a frame shift mutation that truncates the carboxy terminal 33 aminoacids)^[7,8]. Heterozygous carriage of the risk alleles confers a 2-4 fold increased risk, and homozygotes or compound heterozygotes have a 20-40 fold increased risk^[9]. More than 90% of all CD-associated mutations are located in the LRR domain, suggesting that these may affect the function of NOD2 with respect to bacterial recognition and signaling.

NOD1/ CARD4 (Caspase Recruitment Domain 4) plays a role in colonic epithelial defense against *E. coli* and *S. flexeneri* and mediates NF- κ B activation^[3,10]. Recently, genetic variants of NOD1 have been shown to be associated with disease susceptibility. In a recent study of 556 patients with IBD (294 CD and 252 UC), an association between the variant (rs695857) in nucleotides 30, 258 and 950 of NOD1 and the development of IBD was found. Another variant known as rs2907748 in nucleotides 30, 246 and 263 was also associated with the presence of UC and CD and even with the early onset of the disease (< 25 years of age)^[11].

TLRs are abundantly expressed on the surface of monocytes, macrophages, dendritic and epithelial cells. Alterations of TLR3 and TLR4 expression by intestinal epithelial cells have been described in IBD^[12], suggesting that there is differential expression of TLR family members. Two common polymorphisms of TLR4 (Asp299Gly and Thr399Ile) have been described in humans. Asp299Gly has been associated with reduced responsiveness following lipopolysaccharide stimulation^[13]. These polymorphisms have been associated with the development of CD and UC in Caucasian populations^[14-16]. Pierik *et al*^[17] showed that TLR1 R80T and TLR2 R753G polymorphisms were associated with pancolitis in UC patients, while a negative association was observed between TLR6 S249P and proctitis in patients with UC. These results suggest that TLR2 and its co-receptors TLR1 and TLR6 are involved in the initial immune response to bacteria in the pathogenesis of IBD.

ANTIGEN PRESENTATION MOLECULES

The major histocompatibility complex (MHC) region is the region studied most extensively. Human leucocyte antigen (HLA) class II molecules present partially digested antigen to the T-cell receptor and play a central role in the immune response. The mechanism by which classical HLA class II genes exert their influence in IBD is unknown. Different HLA molecules may bind preferentially to different peptides, or bind the same peptide with varying affinity. In IBD, cross reactivity (known as molecular mimicry) may exist between the peptides derived from bacterial luminal flora and from self antigens present in the gut. This may lead to the generation of auto reactive T cells which contribute to disease pathogenesis. HLA-DRB1

is the most extensively studied gene in IBD. In a meta-analysis made by Stokkers *et al*^[18], positive associations between UC and HLA-DR2, HLA-DRB1*1502 (OR = 3.74, CI: 2.2-6.38), HLA-DR9 (OR = 1.54, CI: 1.06-2.24) and HLA-DRB1*0103 (OR = 3.42, CI: 1.52-3.69) were found; a negative association was found with HLA-DR4 (OR = 0.54, CI: 0.43-0.68). Another study found that HLA-DRB1*0103 allele was associated with UC and its severe manifestations such as colectomy and pancolitis ($P = 0.003$, OR = 3.6, CI 95%: 1.46-8.9), while HLA-DRB1*15 allele was only associated with pancolitis in patients with UC ($P = 0.001$, OR = 8.5)^[19].

On the other hand, HLA class III genes have been associated with IBD. Several studies have shown the role of tumor necrosis factor α (TNF α) polymorphisms in IBD. There are specific genetic polymorphisms involving TNF α that influence the amount of cytokine produced. Bouma *et al*^[20] reported an association between the polymorphism of TNF α gene promoter region at -308 position and UC, and this finding was confirmed by other studies^[21, 22].

EPITHELIAL INTEGRITY

The organic cation transporter (OCTN) is a family of transporter proteins for organic cations, and may also transport carnitine, an essential cofactor of the metabolism of lipids. Carnitine is involved in the transport of long-chain fatty acids into the mitochondria. There is evidence that inhibition of fatty acid oxidation in the epithelium of the colonic mucosa is associated with the development of UC. There are two subtypes of this gene, OCTN1 and OCTN2, and some mutations have been reported in them: SLC22A4 1672C/T for OCTN1 and SLC22A5-207G/C for OCTN2, which are associated with the development of CD. The presence or combination of these mutations constitutes TC haplotype, which is associated with ileal, colonic and perianal affection and onset and the need of surgical treatment in CD^[23, 24].

DLG5 (Drosophila long disc homologue 5) gene is a member of the membrane associated guanylate kinase gene family which encodes cell scaffolding proteins and seems to play a role in the maintenance of intestinal epithelial cells, and its mutations have been involved in a rise in intestinal permeability^[25]. DLG5 is a widely expressed protein found in many tissues such as the placenta, small bowel, colon, heart, skeletal muscle, liver and pancreas. It is important in signal transduction and epithelial cell integrity. Four haplotypes have been identified, but only D haplotypes were associated with UC and CD in a European cohort^[26]. Another variant of this gene (rs37462) was found in Japanese people with CD^[27]. The haplotype characterized by the haplotype-tagging single nucleotide polymorphisms (SNP) G113A called haplotype D, was found substantially over-transmitted in patients with IBD.

DRUG TRANSPORTER

The multidrug-resistance (MDR-1) gene encodes the drug efflux pump P-glycoprotein 170 (Pgp-170). Various polymorphisms have been identified within MDR-1: a

mutation C3435T in exon 26 and a mutation G2677T in exon 21 have been correlated with altered Pgp expression and function in humans. Overexpression of MDR-1 leads to an increased efflux of drugs and decreased cytoplasmic drug concentrations. Several drugs, including glucocorticoids, are known Pgp-170 substrates. Farrell *et al*^[28] showed that MDR was significantly elevated in CD and UC patients who required bowel resection and proctocolectomy after failed medical therapy. Variant C3435T was related to the presence of pancolitis in patients with UC in Scotland^[29]. However, the frequency of SNPs is low and is different among populations, with the exception of three SNPs in exon 12 (C1236T), exon 20 (G2677T/A) and exon 26 (C3435T), and some of them are correlated with different diseases and clinical characteristics^[30].

CELL ADHESION

Cell surface adhesion molecules conveying leukocyte-endothelial interactions, govern homing of activated inflammatory cells into gut. Extravasation and migration into the site of inflammation are mediated by integrins and selectins, and these molecules are increased in IBD patients. There are targeting therapies against adhesion molecules in clinical trials to date including natalizumab (integrin $\alpha 4$ subunit) and MLN-02 (selective adhesion molecule blocker for integrin $\alpha 4\beta 7$). In Japanese patients with IBD, the intercellular adhesion molecule-1 (ICAM-1) K469 allele is associated with CD and UC^[31].

CONCLUSION

There are increasing numbers of genetic markers associated with the development of IBD at different levels: innate immunity, antigen presentation, epithelial integrity, drug transporter and cell adhesion that contribute, in genetic susceptibility, to the development of IBD in conjunction with environmental and immunological factors.

REFERENCES

- 1 Bonen DK, Cho JH. The genetics of inflammatory bowel disease. *Gastroenterology* 2003; **124**: 521-536
- 2 Vermeire S. Review article: genetic susceptibility and application of genetic testing in clinical management of inflammatory bowel disease. *Aliment Pharmacol Ther* 2006; **24** Suppl 3: 2-10
- 3 Ogura Y, Inohara N, Benito A, Chen FF, Yamaoka S, Nuñez G. Nod2, a Nod1/Apaf-1 family member that is restricted to monocytes and activates NF-kappaB. *J Biol Chem* 2001; **276**: 4812-4818
- 4 Ogura Y, Lala S, Xin W, Smith E, Dowds TA, Chen FF, Zimmermann E, Tretiakova M, Cho JH, Hart J, Greenson JK, Keshav S, Nuñez G. Expression of NOD2 in Paneth cells: a possible link to Crohn's ileitis. *Gut* 2003; **52**: 1591-1597
- 5 Inohara N, Ogura Y, Fontalba A, Gutierrez O, Pons F, Crespo J, Fukase K, Inamura S, Kusumoto S, Hashimoto M, Foster SJ, Moran AP, Fernandez-Luna JL, Nuñez G. Host recognition of bacterial muramyl dipeptide mediated through NOD2. Implications for Crohn's disease. *J Biol Chem* 2003; **278**: 5509-5512
- 6 Girardin SE, Boneca IG, Viala J, Chamaillard M, Labigne A, Thomas G, Philpott DJ, Sansonetti PJ. Nod2 is a general sensor of peptidoglycan through muramyl dipeptide (MDP) detection. *J Biol Chem* 2003; **278**: 8869-8872
- 7 Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, Achkar JP, Brant SR, Bayless TM, Kirschner BS, Hanauer SB, Nuñez G, Cho JH. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 2001; **411**: 603-606
- 8 Hugot JP, Chamaillard M, Zouali H, Lesage S, Cézard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, Binder V, Finkel Y, Cortot A, Modigliani R, Laurent-Puig P, Gower-Rousseau C, Macry J, Colombel JF, Sahbatou M, Thomas G. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 2001; **411**: 599-603
- 9 Cuthbert AP, Fisher SA, Mirza MM, King K, Hampe J, Croucher PJ, Mascheretti S, Sanderson J, Forbes A, Mansfield J, Schreiber S, Lewis CM, Mathew CG. The contribution of NOD2 gene mutations to the risk and site of disease in inflammatory bowel disease. *Gastroenterology* 2002; **122**: 867-874
- 10 Kim JG, Lee SJ, Kagnoff MF. Nod1 is an essential signal transducer in intestinal epithelial cells infected with bacteria that avoid recognition by toll-like receptors. *Infect Immun* 2004; **72**: 1487-1495
- 11 McGovern DP, Hysi P, Ahmad T, van Heel DA, Moffatt MF, Carey A, Cookson WO, Jewell DP. Association between a complex insertion/deletion polymorphism in NOD1 (CARD4) and susceptibility to inflammatory bowel disease. *Hum Mol Genet* 2005; **14**: 1245-1250
- 12 Cario E, Podolsky DK. Differential alteration in intestinal epithelial cell expression of toll-like receptor 3 (TLR3) and TLR4 in inflammatory bowel disease. *Infect Immun* 2000; **68**: 7010-7017
- 13 Arbour NC, Lorenz E, Schutte BC, Zabner J, Kline JN, Jones M, Frees K, Watt JL, Schwartz DA. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet* 2000; **25**: 187-191
- 14 Franchimont D, Vermeire S, El Housni H, Pierik M, Van Steen K, Gustot T, Quertinmont E, Abramowicz M, Van Gossum A, Devière J, Rutgeerts P. Deficient host-bacteria interactions in inflammatory bowel disease? The toll-like receptor (TLR)-4 Asp299gly polymorphism is associated with Crohn's disease and ulcerative colitis. *Gut* 2004; **53**: 987-992
- 15 Arnott ID, Ho GT, Nimmo ER, Satsangi J. Toll-like receptor 4 gene in IBD: further evidence for genetic heterogeneity in Europe. *Gut* 2005; **54**: 308; author reply 309
- 16 Lakatos PL, Lakatos L, Szalay F, Willheim-Polli C, Osterreicher C, Tulassay Z, Molnar T, Reinisch W, Papp J, Mozsik G, Ferenci P. Toll-like receptor 4 and NOD2/CARD15 mutations in Hungarian patients with Crohn's disease: phenotype-genotype correlations. *World J Gastroenterol* 2005; **11**: 1489-1495
- 17 Pierik M, Joossens S, Van Steen K, Van Schuerbeek N, Vlietinck R, Rutgeerts P, Vermeire S. Toll-like receptor-1, -2, and -6 polymorphisms influence disease extension in inflammatory bowel diseases. *Inflamm Bowel Dis* 2006; **12**: 1-8
- 18 Stokkers PC, Reitsma PH, Tytgat GN, van Deventer SJ. HLA-DR and -DQ phenotypes in inflammatory bowel disease: a meta-analysis. *Gut* 1999; **45**: 395-401
- 19 Yamamoto-Furusho JK, Uscanga LF, Vargas-Alarcón G, Ruiz-Morales JA, Higuera L, Cutiño T, Rodríguez-Pérez JM, Villarreal-Garza C, Granados J. Clinical and genetic heterogeneity in Mexican patients with ulcerative colitis. *Hum Immunol* 2003; **64**: 119-123
- 20 Bouma G, Xia B, Crusius JB, Bioque G, Koutroubakis I, Von Blomberg BM, Meuwissen SG, Peña AS. Distribution of four polymorphisms in the tumour necrosis factor (TNF) genes in patients with inflammatory bowel disease (IBD). *Clin Exp Immunol* 1996; **103**: 391-396
- 21 Hirv K, Seyfarth M, Uibo R, Kull K, Salupere R, Latza U, Rink L. Polymorphisms in tumour necrosis factor and adhesion molecule genes in patients with inflammatory bowel disease: associations with HLA-DR and -DQ alleles and subclinical markers. *Scand J Gastroenterol* 1999; **34**: 1025-1032
- 22 Yamamoto-Furusho JK, Uscanga LF, Vargas-Alarcón G, Rodríguez-Pérez JM, Zuñiga J, Granados J. Polymorphisms in the promoter region of tumor necrosis factor alpha (TNF-alpha)

- and the HLA-DRB1 locus in Mexican mestizo patients with ulcerative colitis. *Immunol Lett* 2004; **95**: 31-35
- 23 **Török HP**, Glas J, Tonenchi L, Lohse P, Müller-Myhsok B, Limbersky O, Neugebauer C, Schnitzler F, Seiderer J, Tillack C, Brand S, Brännler G, Jagiello P, Epplen JT, Griga T, Klein W, Schiemann U, Folwaczny M, Ochsenkühn T, Folwaczny C. Polymorphisms in the DLG5 and OCTN cation transporter genes in Crohn's disease. *Gut* 2005; **54**: 1421-1427
- 24 **Noble CL**, Nimmo ER, Drummond H, Ho GT, Tenesa A, Smith L, Anderson N, Arnott ID, Satsangi J. The contribution of OCTN1/2 variants within the IBD5 locus to disease susceptibility and severity in Crohn's disease. *Gastroenterology* 2005; **129**: 1854-1864
- 25 **Peeters M**, Geyens B, Claus D, Nevens H, Ghos Y, Verbeke G, Baert F, Vermeire S, Vlietinck R, Rutgeerts P. Clustering of increased small intestinal permeability in families with Crohn's disease. *Gastroenterology* 1997; **113**: 802-807
- 26 **Stoll M**, Corneliussen B, Costello CM, Waetzig GH, Mellgard B, Koch WA, Rosenstiel P, Albrecht M, Croucher PJ, Seegert D, Nikolaus S, Hampe J, Lengauer T, Pierrou S, Foelsch UR, Mathew CG, Lagerstrom-Fermer M, Schreiber S. Genetic variation in DLG5 is associated with inflammatory bowel disease. *Nat Genet* 2004; **36**: 476-480
- 27 **Yamazaki K**, Takazoe M, Tanaka T, Ichimori T, Saito S, Iida A, Onouchi Y, Hata A, Nakamura Y. Association analysis of SLC22A4, SLC22A5 and DLG5 in Japanese patients with Crohn disease. *J Hum Genet* 2004; **49**: 664-668
- 28 **Farrell RJ**, Murphy A, Long A, Donnelly S, Cherikuri A, O'Toole D, Mahmud N, Keeling PW, Weir DG, Kelleher D. High multidrug resistance (P-glycoprotein 170) expression in inflammatory bowel disease patients who fail medical therapy. *Gastroenterology* 2000; **118**: 279-288
- 29 **Ho GT**, Nimmo ER, Tenesa A, Fennell J, Drummond H, Mowat C, Arnott ID, Satsangi J. Allelic variations of the multidrug resistance gene determine susceptibility and disease behavior in ulcerative colitis. *Gastroenterology* 2005; **128**: 288-296
- 30 **Wang D**, Sadée W. Searching for polymorphisms that affect gene expression and mRNA processing: example ABCB1 (MDR1). *AAPS J* 2006; **8**: E515-E520
- 31 **Matsuzawa J**, Sugimura K, Matsuda Y, Takazoe M, Ishizuka K, Mochizuki T, Seki SS, Yoneyama O, Bannnai H, Suzuki K, Honma T, Asakura H. Association between K469E allele of intercellular adhesion molecule 1 gene and inflammatory bowel disease in a Japanese population. *Gut* 2003; **52**: 75-78

S- Editor Zhu LH L- Editor Alpini GD E- Editor Li HY