

WJG 20th Anniversary Special Issues (2): Hepatitis C virus

Hepatitis C-associated liver carcinogenesis: Role of PML nuclear bodies

Kerstin Herzer, Guido Gerken, Thomas G Hofmann

Kerstin Herzer, Guido Gerken, Department for Gastroenterology and Hepatology, University Hospital Essen, 45122 Essen, Germany

Kerstin Herzer, Department of General-, Visceral- und Transplantation Surgery, University Hospital Essen, 45122 Essen, Germany

Thomas G Hofmann, Cellular Senescence Group, DKFZ-ZMBH Alliance, Deutsches Krebsforschungszentrum (dkfz.), 69120 Heidelberg, Germany

Author contributions: Herzer K and Hofmann TG designed and wrote the manuscript; Gerken G contributed in supplementing the manuscript.

Correspondence to: Kerstin Herzer, MD, Department for Gastroenterology and Hepatology, University Hospital Essen, Hufeland str. 55, 45122 Essen, Germany. kerstin.herzer@uk-essen.de
Telephone: +49-201-7236579 Fax: +49-201-7236926

Received: November 14, 2013 Revised: December 31, 2013

Accepted: July 24, 2014

Published online: September 21, 2014

Abstract

Successful escape from immune response characterises chronic hepatitis C virus (HCV) infection, which results in persistence of infection in about 80% of the patients. The deleterious consequences are cirrhosis and hepatocellular carcinoma. HCV accounts the most frequent cause for hepatocellular carcinoma (HCC) and liver transplantation (LT) in the western world. The underlying molecular mechanisms how HCV promotes tumor development are largely unknown. There is some *in vitro* and *in vivo* evidence that HCV interferes with the tumor suppressor PML and may thereby importantly contribute to the HCV-associated pathogenesis with respect to the development of HCC. The tumor suppressor protein "promyelocytic leukemia" (PML) has been implicated in the regulation of important cellular processes like differentiation and apoptosis. In cancer biology, PML and its associated nuclear bodies (NBs) have initially attracted intense interest due to its role in the pathogenesis of acute promyelocytic leukemia

(APL). More recently, loss of PML has been implicated in human cancers of various histologic origins. Moreover, number and intensity of PML-NBs increase in response to interferons (IFNs) and there is evidence that PML-NBs may represent preferential targets in viral infections. Thus, PML could not only play a role in the mechanisms of the antiviral action of IFNs but may also be involved in a direct oncogenic effect of the HCV on hepatocytes. This review aims to summarise current knowledge about HCV-related liver carcinogenesis and to discuss a potential role of the nuclear body protein PML for this this hard-to-treat cancer.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Hepatitis C virus; Promyelocytic leukemia; Nuclear bodies; Hepatocellular carcinoma; Apoptosis

Core tip: Escape from immune response and non-responsiveness to interferon-therapy are frequently observed in hepatitis C virus (HCV) infection. HCV infection leads to severe liver disease like cirrhosis and hepatocellular carcinoma and is the major cause for hepatocellular carcinoma and liver transplantation in the western world. Interestingly, HCV interferes with tumor suppressor promyelocytic leukemia (PML) function in apoptosis. PML localises to multi-protein complexes termed nuclear bodies (NBs). The number and the size of PML NBs are increased upon interferon-treatment and PML NBs represent preferential targets in viral infection. Accordingly, PML plays an important role in the antiviral action of interferons.

Herzer K, Gerken G, Hofmann TG. Hepatitis C-associated liver carcinogenesis: Role of PML nuclear bodies. *World J Gastroenterol* 2014; 20(35): 12367-12371 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v20/i35/12367.htm> DOI: <http://dx.doi.org/10.3748/wjg.v20.i35.12367>

HEPATOCELLULAR CARCINOMA

The HCC, which accounts for 80%-90% of primary liver tumors is the fifth most common cancer worldwide and the third most common cause of cancer mortality^[1]. While therapeutic options are scarce, liver transplantation has become the treatment of choice for the early stage of unresectable HCC patients because it offers complete tumor excision along with the removal of the carcinogenic liver^[2].

Chronic viral hepatitis B and C infections are the most important risk factors, responsible for the majority of HCC worldwide^[3]. While cirrhosis accounts as precarcinogenic condition as such, hepatocarcinogenesis is a multistep process and hepatitis viruses are supposed to exert a direct oncogenic effect in hepatocytes^[4]. It has been proposed that HCC development requires a combination of two mechanisms implying cell lysis and stimulation of mitosis, leading to an accumulation of molecular events necessary for transformation, followed by increased neoplasia mediated by induction of recombinogenic proteins during chronic hepatitis. Thus, HCV-related HCC is a consequence of accumulative somatic mutations in the genome of virus-infected liver cells^[5]. However, the exact mechanisms are either diverse or not elucidated yet, making current available therapeutic modalities for HCC largely inadequate. So far, cytotoxic chemotherapy has proven ineffective and no first-line therapy has emerged for advanced HCC^[6]. Despite major efforts to improve diagnosis and treatment of HCC, therapeutic options remain limited to local ablative procedures like transarterial chemoembolization (TACE) and selective interne radiotherapy (SIRT) with liver transplantation (LT) being the only curative treatment approach. Thus, the need for novel therapeutic agents and strategies is obvious.

Chronic infection with hepatitis C virus (HCV) has been causally associated with HCC^[7]. Advances in imaging and treatment gave rise to some improvement in prognosis of patients with HCV-related HCC, but outcomes are still unsatisfactory. Reasons for this unfavourable prognosis are considered to include high intrahepatic tumor recurrence rates and sustained hepatic damage, both resulting from HCV infection. Underlying HCV-related hepatic damage may also impair liver function and consecutively clinical outcome. Thus, prevention of HCC recurrence and preservation of liver function are both central issues for improving prognosis of patients with HCV-related HCC.

The combination of pegylated interferone (PegIFN) plus ribavirin (RBV) was the standard of care for HCV infection for 10 years until 2011, when the first generation protease inhibitors boceprevir (BOC) and telaprevir (TVR) were approved. For the other genotypes, the PegIFN plus RBV treatment regimen remained the standard of care. This treatment fails to eradicate HCV in many chronically infected patients with additional side effects. In those patients for who PegIFN was effective in eradicating HCV, this treatment could have value in mini-

mizing hepatic necrosis, inflammation and fibrosis, and thus, reducing the incidence of HCC^[8]. Several studies have reported that IFN therapy, even after curative treatment for HCV-related HCC, could prevent HCC recurrence and improve survival^[9]. Moreover, PegIFN-alpha has been shown to effectively treat advanced hepatocellular carcinoma (HCC), even more so in combination with 5-fluorouracil (5-FU)^[10]. However, the mechanisms by which IFN suppresses HCC recurrence, including possible direct anti-tumor and anti-inflammatory effects, remain uncertain.

Recently we described that the IFN- α increases the expression of the tumor suppressor TNF-related apoptosis-inducing ligand (TRAIL) in hepatoma cells, a mechanism that is mediated by the promyelocytic leukemia protein (PML)^[11] indicating that enhanced expression of PML and TRAIL and subsequent activation of death receptors is a central feature in IFN-associated regulation of apoptosis in liver tumor cells^[12]. Thus, TRAIL and PML may account central components in IFN-mediated apoptosis regulation and prevention of hepatocarcinogenesis.

Since the recent approval of several new direct acting antivirals (DAAs), IFN might become dispensable and HCV therapy achieves currently a new milestone, as some highly effective antiviral substances are now available and more are awaited^[13]. Experts promise complete eradication of the virus in the coming years. Thus, 2014 is the beginning of a new era of treatment options for HCV infection and the consequences on the incidence of HCC can eagerly be awaited.

PML NUCLEAR BODIES

Tumor suppressor protein PML localizes predominantly to distinct nuclear domains termed PML-nuclear bodies (PML-NBs), multiprotein complexes implicated in apoptosis regulation, cellular senescence and antiviral response. PML-NBs are present in almost every human cell type and appear as discrete nuclear dots. More than 50 different proteins have been localized to PML NBs, which are defined by the two molecular marker proteins PML and Sp100. PML-NBs can contain ribosomal proteins, transcription factors, transcriptional cofactors, as well as oncoproteins and also viral proteins, which are transiently associated with PML-NBs^[14].

In leukemic blast of patients suffering from acute promyelocytic leukemia, the PML gene is reciprocally fused with the *retinoic acid receptor α* gene, thus resulting in expression of an oncogenic PML-RAR α fusion protein. PML-RAR α protein leads to disruption of PML-NBs in numerous tiny microspeckles, and its expression leads to cellular transformation and induction of leukemia. Treatment of APL patient with all-trans retinoic acid or arsenic trioxide (As₂O₃) causes reformation of PML-NBs and triggers either terminal differentiation, apoptosis or senescence of the blasts, which finally results in disease remission. Investigations of PML-RAR α transgenic and PML^{-/-} mice have demonstrated that PML inactivation results in impaired induction of apoptosis by multiple

apoptotic pathways, and *PML*^{-/-} mice are highly susceptible to developing tumors when challenged by carcinogens^[15]. Recently, comprehensive studies have shown that PML protein is frequently lost in human cancers of various histologic origins^[16]. These observations demonstrate that PML is crucial for critical tumor-suppressive pathways although its exact mechanism of action in most tumors is largely unknown.

Remarkably, the promyelocytic leukemia (*PML*) gene is a target of IFNs that is specifically up-regulated at the transcriptional level. Multiple isoforms of PML have been identified, most of which are found in the nucleus where they can assemble to PML-NBs^[17]. In addition, PML deficiency has been linked to increased susceptibility to viral pathogens^[18]. Recent reports suggest that suppression of PML by hepatitis B virus surface antigen interferes with cellular mechanisms responding to double-strand DNA damage repair or apoptosis induction, potentially contributing to hepatocarcinogenesis associated with chronic hepatitis B virus infection^[19,20].

Furthermore, it is well established that PML-NBs act as platforms which are involved in the control of p53 activity^[21]. PML isoform IV (PML-IV) serves as a p53 coactivator and colocalizes with numerous factors regulating p53 post-translational modification, in particular its phosphorylation and acetylation, in PML-NBs, thereby regulating p53 target gene expression and effector function. Although previous studies showed that HCV core protein interacts with p53 and affects its function, the reported effects of HCV core on p53 activity are not consistent^[22,23]. We were, however, able to observe that HCV core interacts with PML, thereby interfering with p53-phosphorylation and activation and, thus, induction of apoptosis which might result in HCV-associated hepatocarcinogenesis.

HCV INFECTION AND PML

Chronic infection with HCV is a major risk factor for the development of HCC and, moreover, is considered as the leading cause for liver transplantation in the western world^[24]. Some authors suggest the Hepatitis C virus-induced chronic inflammation and the effects of cytokines in the development of fibrosis and liver cell proliferation as major pathogenic mechanisms for HCV-associated HCC development. On the other side, the HCV is an RNA virus that does not integrate into the host genome but HCV proteins interact in the cytoplasm with many host-cell proteins, thereby influencing cell signalling, transcription, cell proliferation, apoptosis and translational regulation^[25]. Some convincing experimental evidence suggests that HCV contributes to HCC development by direct interference with pathways that promote the malignant transformation of hepatocytes^[26]. At least four of the HCV gene products, namely HCV core, NS3, NS4B and NS5A, have been shown to exhibit transformation potential in tissue culture^[26]. Several studies have investigated a potential role of the core protein in

regulating apoptotic cell death. It has been demonstrated to exert both, pro- and anti-apoptotic activities^[27] and in liver derived cells opposite and controversial effects were observed. It was demonstrated that the core protein might physically interact with the cytoplasmic domains of CD95, TNF-R1 and the lymphotoxin- β receptor and thereby enhances downstream signalling events of these receptors^[28]. Moreover, the core protein did also interact with the death domain of FADD and with c-FLIP^[29]. Because of the presence of a putative DNA-binding motif and nuclear localisation signals, a possible function of the core protein as a gene regulatory mediator has been suggested. However, interpretation of these data is difficult. In the different studies, only scarce attempts have been made in order to understand how HCV proteins could interfere with apoptosis at the molecular level.

As a central apoptosis-modulating mechanism, HCV core has also been reported to bind to p53. However, in the different reports it either activates or inactivates p53 and its downstream target p21. As mentioned above, our lab was able to shed some light on this controversy demonstrating that HCV core inhibits p53-mediated target gene expression through targeting predominantly the coactivator function of promyelocytic leukemia protein (PML)-IV^[30]. These findings strongly suggest that in hepatoma cells the HCV core protein predominantly interferes with p53 function through targeting PML and not only p53, as suggested by previous reports^[31-35]. Furthermore, these findings could provide an explanation for the contradictory reports about the effect of HCV core expression on p53 function, since they demonstrate a prominent role of PML in this scenario, which might be differentially expressed in the particular cell system used in these studies.

In an attempt to reduce potential artifacts caused by non-physiological overexpression in cell culture, HCV transgenic mice have been generated. In those mice, liver cell apoptosis was significantly reduced^[36]. These and other data suggest that HCV proteins might directly or indirectly inhibit apoptosis rather than facilitate apoptosis. Moreover, after crossing those mice with PML knock out mice, the resulting HCV-transgenic PML-deficient mice show a significantly higher sensitivity towards induction of liver tumors^[37]. These observations confirm a central role of PML in HCV-associated development of liver cancer.

CONCLUSION

While it is well established that PML mediates tumor suppressive functions, including induction of apoptosis and growth arrest, and PML has been reported to suppress the growth of breast and prostate cancer cells^[38,39], it is not known whether this protein modulates key tumor suppressive pathways in the pathogenesis of liver tumors. While HCV core impairs PML function but IFN- α induces PML expression, these observations might explain the anti-hepatocarcinogenic effect of therapeutically ap-

plied IFN in HCV infection: IFN -induced PML might compensate the PML which is trapped by the virus.

These data support the potential usefulness of PML in cancer gene therapy, including the treatment of various types of liver cancers, and suggest that the PML gene can be effective in tumor cells resistant to p53-mediated apoptosis, such as advanced liver cancer in adults. In addition, it will be interesting to treat tumors in HCV-transgenic PML-deficient mice with a liver-specific adenovirus recombinantly expressing PML. Speculations that PML-reconstitution has a not only therapeutic but also preventive effect on hepatocarcinogenesis is intriguing and thus, we would postulate a therapeutic potential for PML in this setting.

An immunohistologic study of PML expression in hepatocarcinogenesis describes differential overexpression of PML in dysplastic nodules versus in carcinoma suggesting a differential regulation during the multistep hepatocarcinogenesis^[40]. To further elucidate a functional role for PML in HCC, it would be interesting to address the question, whether PML expression in HCCs is associated with chronic HCV infection and whether velocity of disease progression correlates with alterations in PML expression or defects in the PML gene.

While the induction of PML appears important for enhanced expression of death receptors and apoptosis in liver cancer, the binding partners of PML appear to be quite diverse in different cell types. To understand its role in transcriptional control, a major focus in the future will therefore be to identify transcription factors with which PML interacts upon IFN stimulation.

The availability of both IFN-responsive and non-responsive lines together with availability of the HCV replicon system in combination with techniques such as array analyses will contribute to identify PML-regulated target genes, which are involved in HCV replication. Subsequently, additional tumor suppressive pathways that are regulated by PML upon IFN treatment might be identified in order to elucidate molecular mechanisms of HCV-associated hepatocarcinogenesis. Finally, new generation DAAs must be assessed with respect to their potential to influence the expression of tumor suppressors. Those studies will provide new insights into the understanding of the use of old versus new HCV therapeutics with respect to their potential for not only the treatment of HCV infection but also to prevent hepatocarcinogenesis.

REFERENCES

- 1 Teufel A, Marquardt JU, Galle PR, Wörns M. [Hepatocellular carcinoma: what's new?]. *Dtsch Med Wochenschr* 2012; **137**: 210-213 [PMID: 22250042 DOI: 10.1055/s-0031-1292890]
- 2 Cicinnati VR, Sotiropoulos GC, Beckebaum S. Established and emerging therapies for hepatocellular carcinoma. *Minerva Med* 2010; **101**: 405-418 [PMID: 21196900]
- 3 Moudgil V, Redhu D, Dhanda S, Singh J. A review of molecular mechanisms in the development of hepatocellular carcinoma by aflatoxin and hepatitis B and C viruses. *J Environ Pathol Toxicol Oncol* 2013; **32**: 165-175 [PMID: 24099430]
- 4 Teufel A, Staib F, Kanzler S, Weinmann A, Schulze-Bergkamen H, Galle PR. Genetics of hepatocellular carcinoma. *World J Gastroenterol* 2007; **13**: 2271-2282 [PMID: 17511024]
- 5 Teufel A, Marquardt JU, Galle PR. Novel insights in the genetics of HCC recurrence and advances in transcriptomic data integration. *J Hepatol* 2012; **56**: 279-281 [PMID: 21782765 DOI: 10.1016/j.jhep.2011.05.035]
- 6 Colombo M, Raoul JL, Lencioni R, Galle PR, Zucman-Rossi J, Bañares R, Seehofer D, Neuhaus P, Johnson P. Multi-disciplinary strategies to improve treatment outcomes in hepatocellular carcinoma: a European perspective. *Eur J Gastroenterol Hepatol* 2013; **25**: 639-651 [PMID: 23628963 DOI: 10.1097/MEG.0b013e32835e33bb]
- 7 Jeong SW, Jang JY, Chung RT. Hepatitis C virus and hepatocarcinogenesis. *Clin Mol Hepatol* 2012; **18**: 347-356 [PMID: 23323249 DOI: 10.3350/cmh.2012.18.4.347]
- 8 Hiramatsu N, Oze T, Takehara T. Suppression of hepatocellular carcinoma development in hepatitis C patients given interferon-based anti-viral therapy. *Hepatol Res* 2014; Epub ahead of print [PMID: 25052449 DOI: 10.1111/hepr.12393]
- 9 Khaliq S, Latief N, Jahan S. Role of different regions of the hepatitis C virus genome in the therapeutic response to interferon-based treatment. *Arch Virol* 2014; **159**: 1-15 [PMID: 23851652 DOI: 10.1007/s00705-013-1780-x]
- 10 Murakami E, Aikata H, Miyaki D, Nagaoki Y, Katamura Y, Kawaoka T, Takaki S, Hiramatsu A, Waki K, Takahashi S, Kimura T, Kenjo M, Nagata Y, Ishikawa M, Kakizawa H, Awai K, Chayama K. Hepatic arterial infusion chemotherapy using 5-fluorouracil and systemic interferon- α for advanced hepatocellular carcinoma in combination with or without three-dimensional conformal radiotherapy to venous tumor thrombosis in hepatic vein or inferior vena cava. *Hepatol Res* 2012; **42**: 442-453 [PMID: 22176468 DOI: 10.1111/j.1872-034X.2011.00943.x]
- 11 Herzer K, Hofmann TG, Teufel A, Schimanski CC, Moehler M, Kanzler S, Schulze-Bergkamen H, Galle PR. IFN- α -induced apoptosis in hepatocellular carcinoma involves promyelocytic leukemia protein and TRAIL independently of p53. *Cancer Res* 2009; **69**: 855-862 [PMID: 19141642 DOI: 10.1158/0008-5472.CAN-08-2831]
- 12 Herr I, Schemmer P, Büchler MW. On the TRAIL to therapeutic intervention in liver disease. *Hepatology* 2007; **46**: 266-274 [PMID: 17596886 DOI: 10.1002/hep.21740]
- 13 Kamal SM. Pharmacogenetics of hepatitis C: transition from interferon-based therapies to direct-acting antiviral agents. *Hepat Med* 2014; **6**: 61-77 [PMID: 25114601 DOI: 10.2147/HMER.S41127]
- 14 Hofmann TG, Will H. Body language: the function of PML nuclear bodies in apoptosis regulation. *Cell Death Differ* 2003; **10**: 1290-1299 [PMID: 12934066 DOI: 10.1038/sj.cdd.4401313]
- 15 Wang ZG, Ruggero D, Ronchetti S, Zhong S, Gaboli M, Rivi R, Pandolfi PP. PML is essential for multiple apoptotic pathways. *Nat Genet* 1998; **20**: 266-272 [PMID: 9806545 DOI: 10.1038/3073]
- 16 Gurrieri C, Capodiceci P, Bernardi R, Scaglioni PP, Nafa K, Rush LJ, Verbel DA, Cordon-Cardo C, Pandolfi PP. Loss of the tumor suppressor PML in human cancers of multiple histologic origins. *J Natl Cancer Inst* 2004; **96**: 269-279 [PMID: 14970276]
- 17 El Bougrini J, Dianoux L, Chelbi-Alix MK. PML positively regulates interferon gamma signaling. *Biochimie* 2011; **93**: 389-398 [PMID: 21115099 DOI: 10.1016/j.biochi.2010.11.005]
- 18 Lunardi A, Gaboli M, Giorgio M, Rivi R, Bygrave A, Antoniou M, Drabek D, Dzierzak E, Fagioli M, Salmena L, Botto M, Cordon-Cardo C, Luzzatto L, Pelicci PG, Grosveld F, Pandolfi PP. A Role for PML in Innate Immunity. *Genes Cancer* 2011; **2**: 10-19 [PMID: 21779477 DOI: 10.1177/1947601911402682]
- 19 Chung YL, Wu ML. Promyelocytic leukaemia protein links DNA damage response and repair to hepatitis B virus-related hepatocarcinogenesis. *J Pathol* 2013; **230**: 377-387 [PMID: 23620081 DOI: 10.1002/path.4195]
- 20 Chung YL. Defective DNA damage response and repair

- in liver cells expressing hepatitis B virus surface antigen. *FASEB J* 2013; **27**: 2316-2327 [PMID: 23444429 DOI: 10.1096/fj.12-226639]
- 21 **Krieghoff-Henning E**, Hofmann TG. Role of nuclear bodies in apoptosis signalling. *Biochim Biophys Acta* 2008; **1783**: 2185-2194 [PMID: 18680765 DOI: 10.1016/j.bbamcr.2008.07.002]
 - 22 **Nishimura T**, Kohara M, Izumi K, Kasama Y, Hirata Y, Huang Y, Shuda M, Mukaidani C, Takano T, Tokunaga Y, Nuriya H, Satoh M, Saito M, Kai C, Tsukiyama-Kohara K. Hepatitis C virus impairs p53 via persistent overexpression of 3beta-hydroxysterol Delta24-reductase. *J Biol Chem* 2009; **284**: 36442-36452 [PMID: 19861417 DOI: 10.1074/jbc.M109.043232]
 - 23 **Long J**, Wang Y, Li M, Tong WM, Jia JD, Huang J. Correlation of TP53 mutations with HCV positivity in hepatocarcinogenesis: identification of a novel TP53 microindel in hepatocellular carcinoma with HCV infection. *Oncol Rep* 2013; **30**: 119-124 [PMID: 23624687 DOI: 10.3892/or.2013.2430]
 - 24 **Villanueva A**, Forns X, Llovet JM. Molecular epidemiology in HCV-related hepatocellular carcinoma: first steps. *J Hepatol* 2012; **57**: 213-214 [PMID: 22282033 DOI: 10.1016/j.jhep.2012.01.002]
 - 25 **Bartenschlager R**, Frese M, Pietschmann T. Novel insights into hepatitis C virus replication and persistence. *Adv Virus Res* 2004; **63**: 71-180 [PMID: 15530561 DOI: 10.1016/S0065-3527(04)63002-8]
 - 26 **Selimovic D**, El-Khattouti A, Ghozlan H, Haikel Y, Abdelkader O, Hassan M. Hepatitis C virus-related hepatocellular carcinoma: An insight into molecular mechanisms and therapeutic strategies. *World J Hepatol* 2012; **4**: 342-355 [PMID: 23355912 DOI: 10.4254/wjh.v4.i12.342]
 - 27 **Jahan S**, Ashfaq UA, Khaliq S, Samreen B, Afzal N. Dual behavior of HCV Core gene in regulation of apoptosis is important in progression of HCC. *Infect Genet Evol* 2012; **12**: 236-239 [PMID: 22266242 DOI: 10.1016/j.meegid.2012.01.006]
 - 28 **You LR**, Chen CM, Lee YH. Hepatitis C virus core protein enhances NF-kappaB signal pathway triggering by lymphotoxin-beta receptor ligand and tumor necrosis factor alpha. *J Virol* 1999; **73**: 1672-1681 [PMID: 9882379]
 - 29 **Saito K**, Meyer K, Warner R, Basu A, Ray RB, Ray R. Hepatitis C virus core protein inhibits tumor necrosis factor alpha-mediated apoptosis by a protective effect involving cellular FLICE inhibitory protein. *J Virol* 2006; **80**: 4372-4379 [PMID: 16611896 DOI: 10.1128/JVI.80.9.4372-4379.2006]
 - 30 **Herzer K**, Weyer S, Krammer PH, Galle PR, Hofmann TG. Hepatitis C virus core protein inhibits tumor suppressor protein promyelocytic leukemia function in human hepatoma cells. *Cancer Res* 2005; **65**: 10830-10837 [PMID: 16322229 DOI: 10.1158/0008-5472.CAN-05-0880]
 - 31 **Kao CF**, Chen SY, Chen JY, Wu Lee YH. Modulation of p53 transcription regulatory activity and post-translational modification by hepatitis C virus core protein. *Oncogene* 2004; **23**: 2472-2483 [PMID: 14968111]
 - 32 **Otsuka M**, Kato N, Lan K, Yoshida H, Kato J, Goto T, Shiratori Y, Omata M. Hepatitis C virus core protein enhances p53 function through augmentation of DNA binding affinity and transcriptional ability. *J Biol Chem* 2000; **275**: 34122-34130 [PMID: 10924497]
 - 33 **Honda M**, Kaneko S, Shimazaki T, Matsushita E, Kobayashi K, Ping LH, Zhang HC, Lemon SM. Hepatitis C virus core protein induces apoptosis and impairs cell-cycle regulation in stably transformed Chinese hamster ovary cells. *Hepatology* 2000; **31**: 1351-1359 [PMID: 10827163]
 - 34 **Ray RB**, Meyer K, Ray R. Suppression of apoptotic cell death by hepatitis C virus core protein. *Virology* 1996; **226**: 176-182 [PMID: 8955036]
 - 35 **Ray RB**, Steele R, Meyer K, Ray R. Transcriptional repression of p53 promoter by hepatitis C virus core protein. *J Biol Chem* 1997; **272**: 10983-10986 [PMID: 9110985]
 - 36 **Disson O**, Haouzi D, Desagher S, Loesch K, Hahne M, Kremer EJ, Jacquet C, Lemon SM, Hibner U, Lerat H. Impaired clearance of virus-infected hepatocytes in transgenic mice expressing the hepatitis C virus polyprotein. *Gastroenterology* 2004; **126**: 859-872 [PMID: 14988840]
 - 37 **Herzer K**, Carbow A, Sydor S, Sowa JP, Biesterfeld S, Hofmann TG, Galle PR, Gerken G, Canbay A. Deficiency of the promyelocytic leukemia protein fosters hepatitis C-associated hepatocarcinogenesis in mice. *PLoS One* 2012; **7**: e44474 [PMID: 22984515 DOI: 10.1371/journal.pone.0044474]
 - 38 **Chatterjee A**, Chatterjee U, Ghosh MK. Activation of protein kinase CK2 attenuates FOXO3a functioning in a PML-dependent manner: implications in human prostate cancer. *Cell Death Dis* 2013; **4**: e543 [PMID: 23492774 DOI: 10.1038/cddis.2013.63]
 - 39 **Carracedo A**, Weiss D, Leliaert AK, Bhasin M, de Boer VC, Laurent G, Adams AC, Sundvall M, Song SJ, Ito K, Finley LS, Egia A, Libermann T, Gerhart-Hines Z, Puigserver P, Haigis MC, Maratos-Flier E, Richardson AL, Schafer ZT, Pandolfi PP. A metabolic prosurvival role for PML in breast cancer. *J Clin Invest* 2012; **122**: 3088-3100 [PMID: 22886304 DOI: 10.1172/JCI62129]
 - 40 **Chan JY**, Chin W, Liew CT, Chang KS, Johnson PJ. Altered expression of the growth and transformation suppressor PML gene in human hepatocellular carcinomas and in hepatitis tissues. *Eur J Cancer* 1998; **34**: 1015-1022 [PMID: 9849449]

P- Reviewer: Morris-Stiff G, Feo F, Yu TH **S- Editor:** Gou SX
L- Editor: A **E- Editor:** Zhang DN





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327

