

Injury-associated reacquiring of intestinal stem cell function

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Abstract

Epithelial layer of the intestine relies upon stem cells for maintaining homeostasis and regeneration. Two types of stem cells are currently defined in intestinal crypts: the cycling crypt base columnar cells and quiescent cells. Though several candidate markers and regulators of rapidly cycling and quiescent stem cells have been identified so far, the exact nature of quiescent cells is still questionable since investigations mainly focused on candidate markers rather than the label-retaining population itself. Recent results, however, have strengthened

the argument for functional plasticity. Using a lineage tracing strategy label-retaining cells (LRCs) of the intestinal epithelium were marked, then followed by a pulse-chase analysis it was found that during homeostasis, LRCs were Lgr5-positive and were destined to become Paneth and neuroendocrine cells. Nevertheless, it was demonstrated that LRCs are capable of clonogenic growth by recall to the self-renewing pool of stem cells in case of epithelial injury. These new findings highlight on the hierarchical and spatial organization of intestinal epithelial homeostasis and the important plasticity of progenitors during tissue regeneration, moreover, provide a motivation for studying their role in disorders like colorectal cancer.

Key words: Epithelium; Injury; Label-retaining cells; Stem cell; Lgr5; Intestine

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Core tip: The cellular plasticity and lineage reversibility of the epithelial layer may represent adaptive mechanisms for the self-preservation of the epithelial layer after injuries. Recent results revealed that a portion of Lgr5-expressing intestinal cells cycles less frequently, and upon physiological circumstances does not contribute to intestinal homeostasis, however, they reacquire stem cell function and can be recruited to serve as a functional clonogenic stem population after injury.

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COMMENTARY ON HOT TOPICS

The epithelial monolayer on the luminal border of the

intestinal wall has certain physiological functions, such as diffusion and absorption of small molecules, or acts as a physical, chemical and immunological barrier defence against luminal microbiota. Due to the high cellular turnover (2-3 d) the continuous replacement of shedding epithelial cells is required from a local stem cell pool even in the healthy intestine. Stem cells are located at the crypt base, and their progenies migrate towards the luminal surface where they undergo terminal differentiation^[1-3].

During the years, different methods have been developed to study the destiny, renewal, and differentiation potential of epithelial stem cells. The first functional demonstration of stemness in case of epithelial cells was performed using methods established originally for culturing epidermal keratinocytes under conditions where they were able to maintain and propagate for hundreds of colonies without losing stemness^[4,5]. Proliferative capacity assays and ultrastructural analyses of the intestinal crypt led to opening the hypothesis that crypt base columnar cells refer to intestinal stem cells^[6]. Consequentially, assignment of stemness favored cells at the +4 position, given their mode of chromosome segregation and higher resistance to cell death induced by deoxyribonucleotide-acid (DNA) damage^[7,8]. Lineage tracings of 0→+4 crypt basal cells with *Lgr5-Cre^{ER}* (leucine-rich repeat-containing G-protein coupled receptor 5), *Bmi1* (B cell-specific Moloney murine leukemia virus integration site 1), *mTERT* (mouse telomerase reverse transcriptase), and *Hopx-Cre^{ER}* (homeodomain-only protein homeobox) indicated that all crypt columnar basal cells display interconvertible multipotent intestinal stem cell characteristics^[9-13]. This fact is further illustrated by diptheria toxin-targeted ablation of *Lgr5+* cells, which does not influence intestinal homeostasis^[14]. Based on these results, the intestine is characterized by spatially separated interconvertible stem cells existing in quiescent and/or activated states.

Initially, it has been proposed that all *Lgr5+* intestinal stem cells are cycling rapidly^[9]. Recent data of Buczacki *et al.*^[15] revealed that in case of physiological circumstances approx. 20% of *Lgr5+* intestinal cells not only cycle less frequently, but differentiate into Paneth and neuroendocrine cells, as well do not contribute to intestinal homeostasis. However, this population of cells retain the ability to reacquire stem cell function, and also can be recruited to serve as a functional clonogenic stem population under conditions of regeneration.

DETECTION OF IMMORTAL DNA STRAND SEGREGATION

The Cairn's theory (*i.e.*, the immortal DNA strand hypothesis) describes a mechanism for adult stem cells to minimize genomic mutations^[16]. According to this hypothesis, in adult stem cells DNA division is asymmetric, and stem cells retain a distinct template set of DNA strands over successive generations. Adult

stem cells by non-random DNA division could transmit mutations arising from DNA replication errors onto terminally differentiating daughter cells, thus they may reduce the rate of accumulated mutations frequently leading to genetic disorders, like cancer. Further analysis of this hypothesis may yield insights into areas, such as tissue regeneration, cancer development or the process of aging.

One of the used methods to detect immortal DNA strand segregation is the label-retention assay^[17,18]. In this approach, symmetrically cycling cells are labeled by a repeated or continuous supply of tritiated thymidine or bromodeoxyuridine (BrdU) for one generation time, producing cells with hemi-labeled DNA. Further, asymmetric cell kinetics is induced, and simultaneously the tritiated thymidine or BrdU is removed. Then the cells continue cycling for a long chase period (usually of five generation time) during which the label is lost from all the cycling, transit amplifying cells, therefore only quiescent or slowly cycling cells (*i.e.*, stem cells) remain to be labeled. These cells are termed label-retaining cells (LRCs).

The other technique, the label-release assay, does not require the manipulation of cell division kinetics. In this technique non-immortal DNA strands are labeled and observed, and the captured event is the release of the labeled non-immortal DNA strands from the stem cell turning into the next cell cycle^[19].

EPITHELIAL STEM CELL MARKERS AND REGULATORS

Stem cells exhibit both the self renewal and the ability to give rise of differentiated progenies. Stem cells can be classified as pluripotent embryonic or as multi(uni)potent adult stem cells. In tissues with high cell turnover rates, such as the intestine adult stem cells are cycling asymmetrically^[20]. Currently two types of stem cells are defined in intestinal crypts, the cycling crypt base columnar cells and the quiescent (+4) cells^[21]. So far, several candidate markers and/or regulators of rapidly cycling and quiescent epithelial stem cells have already been identified.

CDX2

Caudal related homeobox protein 2 (CDX2) is a nuclear transcription factor with an important function in early differentiation and maintenance of the intestinal epithelial phenotype^[22], and controlling the expression of a number of downstream genes, some of which contribute to inflammation^[23]. Further, CDX2 was shown to inhibit *in vitro* cell growth and migration as well as dissemination of colon tumor cells *in vivo*^[24]. CDX2 could also be useful for detection of crypt epithelial stem cells^[22]. The induction of CDX2 in intestinal epithelium can lead to expression of Math1 (a basic helix-loop-helix transcription factor), which plays an important role in differentiation of stem cells into goblet cells^[25].

Musashi-1

Musashi-1, an RNA (ribonucleic acid)-binding protein, possesses a regulatory function of the asymmetric cell division in ectodermal precursor cells^[26], and exhibits importance in regulating the maintenance and differentiation of stem cells even in the intestine^[27,28]. On the other hand, Musashi-1+ cells might represent circulating smooth muscle cell precursors, as well^[29].

Lgr5

Lgr5, an orphan receptor, has been identified as a downstream target of the Wnt-pathway^[9]. In situ hybridization and lineage marking using an inducible Lgr5-Cre knockin and Rosa26-lacZ reporter demonstrated that Lgr5 is a marker for small and large bowel epithelial stem cells or long-lived multipotent progenitor cells^[9,30]. Lgr5 might also be involved in cancer stemness^[31]. Interestingly, knockout of the Lgr5 gene has no detectable effect on intestinal development, however, there are closely related receptors which may replace its function^[32].

Bmi1

Bmi1, originally described as a member of a nuclear protein complex in sarcoma and leukemic cells^[33,34], has been found to be expressed in distinct cells located near the bottom of crypts in the small intestine, predominantly in the +4 position^[10]. It has been recently reported that Bmi1+ cells found in the proximal small intestine can be clonally expanded, and repopulate Lgr5+ stem cells after conditional *Lgr5* cell deletion^[10,14].

Wip1

Wip1 (wildtype p53-induced phosphatase; PPM1D) was originally described as a human phosphatase that is induced in response to ionizing radiation in a p53-dependent manner in Burkitt lymphoma cells^[35]. Wip1, as a negative regulator of the nuclear factor (NF)- κ B p65 subunit regulates the homeostasis of intestinal stem cells, and co-localizes with LRCs in a supra-Paneth cell position^[36,37]. The loss of Wip1 gene can lower the threshold of p53-dependent apoptosis, so it prevents the conversion of adult stem cells into tumor-initiating ones^[36].

TERT

TERT is a ribonucleoprotein complex that maintains the telomeric ends of chromosomes, which could help to prevent cellular senescence being relevant for the self-renewal of adult stem cells^[38]. TERT is implicated in the direct regulation of epidermal stem cell proliferation and mobilization^[39,40]. The epithelial expression of TERT together with other growth factor receptors has been found to be increased in the crypt-base affected by chronic inflammation, which may lead to the pathologic survival and proliferation of epithelial cells carrying genetic defects^[41]. In mice, TERT expression has been found to be present in slowly cycling intestinal stem

cells^[11].

Bone morphogenetic protein type I receptor alpha

Bone morphogenetic protein type I receptor alpha (BMPRI α), a receptor of BMP-signaling pathway has been shown to be a negative regulator of intestinal stem cell proliferation^[42]. BMPRI α is highly expressed on LRCs, and the intestine-specific knockout of BMPRI α resulted in polyposis, probably due to an increase in intestinal stem cell self-renewal^[43].

Phosphatase and tensin homolog

Phosphatase and tensin homolog (PTEN), a known tumor suppressor has been found to limit the number of intestinal stem cells^[43]. Its inactive form is co-localized with BrdU+ LRCs^[44]. It has been proposed that BMP-signaling leads to the increase of PTEN activity, which as a negative regulator of phosphoinositol-3-kinase/Akt suppresses Wnt signaling, and thus controls stem cell proliferation^[43].

Ephrin ligands

The interplay of ephrin receptors and ephrin ligands (Eph) regulates cell migration and boundary formation during development and tumorigenesis^[45]. EphB2 and B3 were shown to be strongly expressed at the bottom of the crypts, even at cell positions 4-6, *i.e.*, the putative intestinal stem cell location^[46]. Moreover, gene expression analysis of laser microdissected crypt epithelial cells identified EphA6 as a potential stem cell marker^[47].

HOPX

HOPX, an unusual homeodomain protein, was originally described as a key regulator of cardiac development^[48]. In the intestine, LRCs in the +4 position can be marked by HOPX^[12]. HOPX-expressing cells give rise to columnar base cells and all mature intestinal epithelial lineages. These cells can convert to Lgr5+ population maintaining clonogenic growth^[12]. These findings support the presence of a bidirectional lineage relationship between active and quiescent stem cells in their niches.

Leucine-rich repeats and immunoglobulin-like domains protein 1

Leucine-rich repeats and immunoglobulin-like domains protein 1 (Lrig1), a negative-feedback regulator of the epidermal growth factor receptor family, was proposed to maintain epidermal stem cells in a quiescent, nondividing stage^[49]. In situ hybridisation has revealed that Lrig1 is highly expressed in the stem cell niche of the small intestine and colon, and it regulates proliferation within the niche by inhibiting EGFR-signaling^[50].

Interestingly, a recent report has demonstrated that several quiescent stem cell markers (*i.e.*, Bmi1, TERT, Hopx or Lrig1) are highly expressed by the Lgr5 population^[21]. The previously listed putative biomarkers

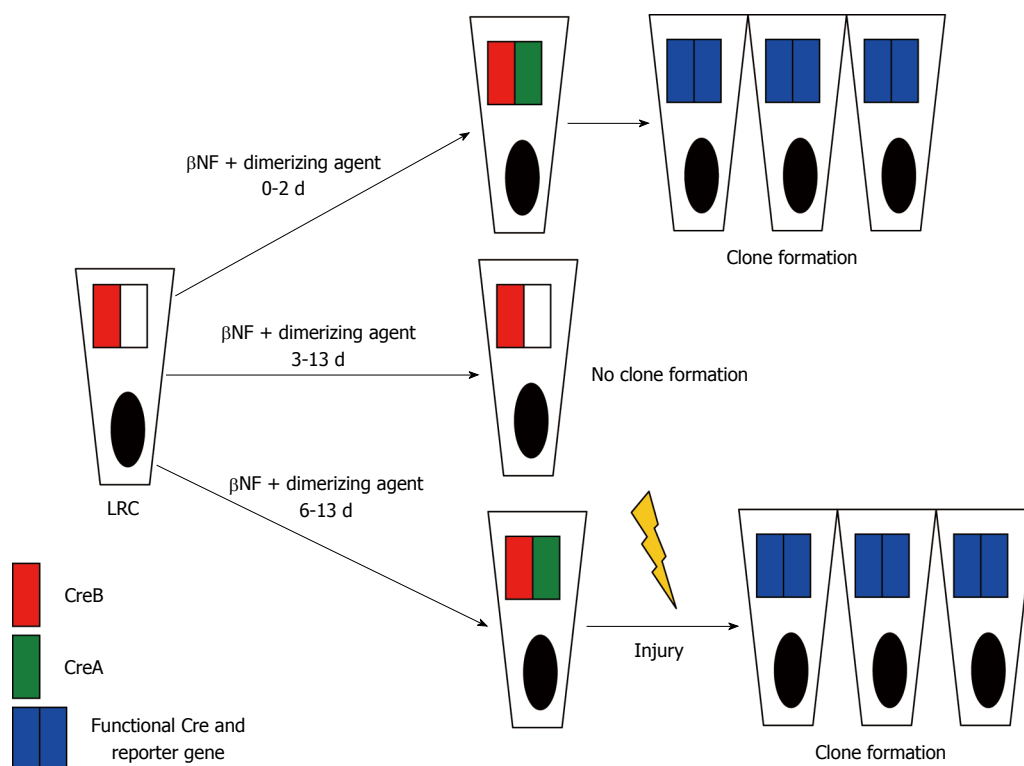


Figure 1 Schematic illustration of conditions for clone formation in the commented study. Dimerizing agent is competent for clone formation from 0 to 2 d post- β NF-induction (top line) but not between 3 and 13 d (middle line). Between 6-13 d both dimerizing agent and injury (hydroxyurea) are required for clone formation (bottom line). LRC: Label-retaining cell; β NF: β -naphthoflavone; Cre: Recombinase.

may provide useful cell surface/cytoplasmic/nuclear markers to study the intestinal stem cells, but remain to be carefully examined and functionally validated. On the whole, the exact nature of quiescent stem cells is still questionable and needs to be clarified, since so far candidate markers rather than the LRCs themselves have been investigated.

INJURY-ASSOCIATED REACQUIRING OF STEMNESS

It is widely accepted that adult intestinal stem cells are able to acquire different progeny fates. However, several recent studies have demonstrated that committed epithelial cells could have the capacity to reverse their destinies^[15,51]. Furthermore, the continuous debate regarding lineage and hierarchy in the intestinal epithelium is still existing. The possibility to reacquire stem cell function and the argument for functional plasticity have been strengthened by the recent results of Buczaccki *et al.*^[15].

Instead of focusing on different putative stem cell markers, for directly studying quiescence the authors^[15] identified cells retaining nuclear-localized fluorescent histone 2B-yellow fluorescent protein (H2B-YFP) fusion protein during a chase period following a pulse of induced expression. After the successful identification of YFP-LRCs, their transcriptional profiling proved that most of them belong to a subpopulation of Lgr5-

expressing cells. Next, it has been demonstrated that YFP-LRCs appear to exist mainly as a Paneth cell precursor population, however, they are also capable of enteroendocrine differentiation. During the most fascinating and unique step of the study only LRCs in the intestinal epithelium were marked by a smart lineage tracing strategy. They fused a fragment of Cre recombinase (CreA) to H2B under the control of a β -naphthoflavone (β NF)-inducible promoter expressed by intestinal epithelial cells. After a β NF pulse-chase analysis only LRCs retained H2B-CreA. Administration of an intravenous dimerization agent reunited the H2B-CreA fragment with its ubiquitously expressed counterpart CreB triggering lineage tracing in LRCs *via* recombination. By varying the interval between β NF induction and Cre dimerization further combined with intestinal injury the authors found that during homeostasis, LRCs were Lgr5+ and destined to become Paneth and neuroendocrine cells. After epithelial injury, however, they demonstrated that LRCs are capable of clonogenic growth by recall to the self-renewing pool of stem cells (Figure 1). In a mammalian system this is the first model of *in vivo* lineage tracing based on label retention.

The results of Buczaccki *et al.*^[15] seem to resolve the supposed paradox that quiescence markers are expressed within a population of Lgr5+ cells thought to be rapidly cycling ones^[21]. According to their results approx. 20% of Lgr5+ cells are largely quiescent and

continue to express Lgr5 before maturation to Paneth cell.

On the other hand, it has been recently shown that suppression of apoptosis (even caused by chemotherapeutic agents) may decrease the number of lineage tracing clones from Lgr5+ cells, whereas lineage tracing from other cells with stem characteristics (e.g., Bmi1-expression) may be increased^[52]. Based on these findings one can speculate that in a stem cell population apoptosis favors the proliferation of another stem cell population. Furthermore, environmental or genetic factors possibly can differentially influence progeny production by a given stem cell pool.

The debate about the origin, identity, and location of crypt epithelial stem cells is still continued by this new study of Buczacki *et al.*^[15]. Their findings highlight the hierarchical and spatial organization of intestinal epithelial homeostasis and the important plasticity of progenitors during tissue regeneration. The cellular plasticity and lineage reversibility may represent an adaptive mechanism after injuries, like intestinal inflammation for the self-preservation of the epithelial layer. Furthermore, in colorectal cancer determination the extent of quiescent cells retaining a maintained clonogenic potential may provide further insight into the etiology of tumorous colonic disorders and develop the basis of novel, efficient anti-cancer therapies.

REFERENCES

- 1 **Matsumoto T**, Okamoto R, Yajima T, Mori T, Okamoto S, Ikeda Y, Mukai M, Yamazaki M, Oshima S, Tsuchiya K, Nakamura T, Kanai T, Okano H, Inazawa J, Hibi T, Watanabe M. Increase of bone marrow-derived secretory lineage epithelial cells during regeneration in the human intestine. *Gastroenterology* 2005; **128**: 1851-1867 [PMID: 15940621]
- 2 **van der Flier LG**, Clevers H. Stem cells, self-renewal, and differentiation in the intestinal epithelium. *Annu Rev Physiol* 2009; **71**: 241-260 [PMID: 18808327 DOI: 10.1146/annurev.physiol.010908.163145]
- 3 **Ricci-Vitiani L**, Pagliuca A, Palio E, Zeuner A, De Maria R. Colon cancer stem cells. *Gut* 2008; **57**: 538-548 [PMID: 18334662 DOI: 10.1136/gut.2007.127837]
- 4 **Rheinwald JG**, Green H. Serial cultivation of strains of human epidermal keratinocytes: the formation of keratinizing colonies from single cells. *Cell* 1975; **6**: 331-343 [PMID: 1052771]
- 5 **Blanpain C**, Fuchs E. Stem cell plasticity. Plasticity of epithelial stem cells in tissue regeneration. *Science* 2014; **344**: 1242281 [PMID: 24926024 DOI: 10.1126/science.1242281]
- 6 **Cheng H**, Leblond CP. Origin, differentiation and renewal of the four main epithelial cell types in the mouse small intestine. I. Columnar cell. *Am J Anat* 1974; **141**: 461-479 [PMID: 4440632]
- 7 **Potten CS**, Hume WJ, Reid P, Cairns J. The segregation of DNA in epithelial stem cells. *Cell* 1978; **15**: 899-906 [PMID: 728994]
- 8 **Merritt AJ**, Potten CS, Kemp CJ, Hickman JA, Balmain A, Lane DP, Hall PA. The role of p53 in spontaneous and radiation-induced apoptosis in the gastrointestinal tract of normal and p53-deficient mice. *Cancer Res* 1994; **54**: 614-617 [PMID: 8306319]
- 9 **Barker N**, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haeghebarth A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* 2007; **449**: 1003-1007 [PMID: 17934449]
- 10 **Sangiorgi E**, Capecchi MR. Bmi1 is expressed in vivo in intestinal stem cells. *Nat Genet* 2008; **40**: 915-920 [PMID: 18536716 DOI: 10.1038/ng.165]
- 11 **Montgomery RK**, Carlone DL, Richmond CA, Farilla L, Kranendonk ME, Henderson DE, Baffour-Awuah NY, Ambruzz DM, Fogli LK, Algra S, Breault DT. Mouse telomerase reverse transcriptase (mTert) expression marks slowly cycling intestinal stem cells. *Proc Natl Acad Sci USA* 2011; **108**: 179-184 [PMID: 21173232 DOI: 10.1073/pnas.1013004108]
- 12 **Takeda N**, Jain R, LeBoeuf MR, Wang Q, Lu MM, Epstein JA. Interconversion between intestinal stem cell populations in distinct niches. *Science* 2011; **334**: 1420-1424 [PMID: 22075725 DOI: 10.1126/science.1213214]
- 13 **Ritsma L**, Ellenbroek SI, Zomer A, Snippert HJ, de Sauvage FJ, Simons BD, Clevers H, van Rheeën J. Intestinal crypt homeostasis revealed at single-stem-cell level by in vivo live imaging. *Nature* 2014; **507**: 362-365 [PMID: 24531760 DOI: 10.1038/nature12972]
- 14 **Tian H**, Biehs B, Warming S, Leong KG, Rangell L, Klein OD, de Sauvage FJ. A reserve stem cell population in small intestine renders Lgr5-positive cells dispensable. *Nature* 2011; **478**: 255-259 [PMID: 21927002 DOI: 10.1038/nature10408]
- 15 **Buczacki SJ**, Zecchini HI, Nicholson AM, Russell R, Vermeulen L, Kemp R, Winton DJ. Intestinal label-retaining cells are secretory precursors expressing Lgr5. *Nature* 2013; **495**: 65-69 [PMID: 23446353 DOI: 10.1038/nature11965]
- 16 **Cairns J**. Mutation selection and the natural history of cancer. *Nature* 1975; **255**: 197-200 [PMID: 1143315]
- 17 **Bickenbach JR**. Identification and behavior of label-retaining cells in oral mucosa and skin. *J Dent Res* 1981; **60** Spec No C: 1611-1620 [PMID: 6943171]
- 18 **Yurkewicz L**, Lauder JM, Marchi M, Giacobini E. 3H-thymidine long survival autoradiography as a method for dating the time of neuronal origin in the chick embryo: the locus coeruleus and cerebellar Purkinje cells. *J Comp Neurol* 1981; **203**: 257-267 [PMID: 7309923]
- 19 **Karpowicz P**, Morshead C, Kam A, Jervis E, Ramunas J, Cheng V, van der Kooy D. Support for the immortal strand hypothesis: neural stem cells partition DNA asymmetrically in vitro. *J Cell Biol* 2005; **170**: 721-732 [PMID: 16115957]
- 20 **Marshallman E**, Booth C, Potten CS. The intestinal epithelial stem cell. *Bioessays* 2002; **24**: 91-98 [PMID: 11782954]
- 21 **Muñoz J**, Stange DE, Schepers AG, van de Wetering M, Koo BK, Itzkovitz S, Volckmann R, Kung KS, Koster J, Radulescu S, Myant K, Versteeg R, Sansom OJ, van Es JH, Barker N, van Oudenaarden A, Mohammed S, Heck AJ, Clevers H. The Lgr5 intestinal stem cell signature: robust expression of proposed quiescent '+4' cell markers. *EMBO J* 2012; **31**: 3079-3091 [PMID: 22692129 DOI: 10.1038/emboj.2012.166]
- 22 **Suh E**, Traber PG. An intestine-specific homeobox gene regulates proliferation and differentiation. *Mol Cell Biol* 1996; **16**: 619-625 [PMID: 8552090]
- 23 **Beck F**, Chawengsaksophak K, Waring P, Playford RJ, Furness JB. Reprogramming of intestinal differentiation and intercalary regeneration in Cdx2 mutant mice. *Proc Natl Acad Sci USA* 1999; **96**: 7318-7323 [PMID: 10377412]
- 24 **Gross I**, Duluc I, Benamer T, Calon A, Martin E, Brabletz T, Kedinger M, Domon-Dell C, Freund JN. The intestine-specific homeobox gene Cdx2 decreases mobility and antagonizes dissemination of colon cancer cells. *Oncogene* 2008; **27**: 107-115 [PMID: 17599044]
- 25 **Mutoh H**, Sakamoto H, Hayakawa H, Arai Y, Satoh K, Nokubi M, Sugano K. The intestine-specific homeobox gene Cdx2 induces expression of the basic helix-loop-helix transcription factor Math1. *Differentiation* 2006; **74**: 313-321 [PMID: 16831200]
- 26 **Okano H**, Kawahara H, Toriya M, Nakao K, Shibata S, Imai T. Function of RNA-binding protein Musashi-1 in stem cells. *Exp Cell Res* 2005; **306**: 349-356 [PMID: 15925591]
- 27 **Potten CS**, Booth C, Tudor GL, Booth D, Brady G, Hurley P, Ashton G, Clarke R, Sakakibara S, Okano H. Identification of a putative intestinal stem cell and early lineage marker; musashi-1.

- Differentiation* 2003; **71**: 28-41 [PMID: 12558601]
- 28 **Wang X**, Hu JF, Tan Y, Cui J, Wang G, Msrny RJ, Li W. Cancer stem cell marker Musashi-1 rs2522137 genotype is associated with an increased risk of lung cancer. *PLoS One* 2014; **9**: e95915 [PMID: 24787949 DOI: 10.1371/journal.pone.0095915]
 - 29 **Bobryshev YV**, Tran D, Botelho NK, Lord RV, Orekhov AN. Musashi-1 expression in atherosclerotic arteries and its relevance to the origin of arterial smooth muscle cells: histopathological findings and speculations. *Atherosclerosis* 2011; **215**: 355-365 [PMID: 21296351 DOI: 10.1016/j.atherosclerosis.2011.01.013]
 - 30 **Montgomery RK**, Breault DT. Small intestinal stem cell markers. *J Anat* 2008; **213**: 52-58 [PMID: 18638070 DOI: 10.1111/j.1469-7580.2008.00925.x]
 - 31 **Nakata S**, Phillips E, Goidts V. Emerging role for leucine-rich repeat-containing G-protein-coupled receptors LGR5 and LGR4 in cancer stem cells. *Cancer Manag Res* 2014; **6**: 171-180 [PMID: 24711713 DOI: 10.2147/CMAR.S57846]
 - 32 **Morita H**, Mazerbourg S, Bouley DM, Luo CW, Kawamura K, Kuwabara Y, Baribault H, Tian H, Hsueh AJ. Neonatal lethality of LGR5 null mice is associated with ankyloglossia and gastrointestinal distension. *Mol Cell Biol* 2004; **24**: 9736-9743 [PMID: 15509778]
 - 33 **Satijn DP**, Gunster MJ, van der Vlag J, Hamer KM, Schul W, Alkema MJ, Saurin AJ, Freemont PS, van Driel R, Otte AP. RING1 is associated with the polycomb group protein complex and acts as a transcriptional repressor. *Mol Cell Biol* 1997; **17**: 4105-4113 [PMID: 9199346]
 - 34 **Lessard J**, Sauvageau G. Bmi-1 determines the proliferative capacity of normal and leukaemic stem cells. *Nature* 2003; **423**: 255-260 [PMID: 12714970]
 - 35 **Fiscella M**, Zhang H, Fan S, Sakaguchi K, Shen S, Mercer WE, Vande Woude GF, O'Connor PM, Appella E. Wip1, a novel human protein phosphatase that is induced in response to ionizing radiation in a p53-dependent manner. *Proc Natl Acad Sci USA* 1997; **94**: 6048-6053 [PMID: 9177166]
 - 36 **Demidov ON**, Timofeev O, Lwin HN, Kek C, Appella E, Bulavin DV. Wip1 phosphatase regulates p53-dependent apoptosis of stem cells and tumorigenesis in the mouse intestine. *Cell Stem Cell* 2007; **1**: 180-190 [PMID: 18371349 DOI: 10.1016/j.stem.2007.05.020]
 - 37 **Chew J**, Biswas S, Shreeram S, Humaidi M, Wong ET, Dhillon MK, Teo H, Hazra A, Fang CC, López-Collazo E, Bulavin DV, Tergaonkar V. WIP1 phosphatase is a negative regulator of NF-kappaB signalling. *Nat Cell Biol* 2009; **11**: 659-666 [PMID: 19377466 DOI: 10.1038/ncb1873]
 - 38 **Blackburn EH**. Structure and function of telomeres. *Nature* 1991; **350**: 569-573 [PMID: 1708110]
 - 39 **Flores I**, Cayuela ML, Blasco MA. Effects of telomerase and telomere length on epidermal stem cell behavior. *Science* 2005; **309**: 1253-1256 [PMID: 16037417]
 - 40 **Sarin KY**, Cheung P, Gilson D, Lee E, Tennen RI, Wang E, Artandi MK, Oro AE, Artandi SE. Conditional telomerase induction causes proliferation of hair follicle stem cells. *Nature* 2005; **436**: 1048-1052 [PMID: 16107853]
 - 41 **Sipos F**, Galamb O, Herszényi L, Molnár B, Solymosi N, Zágoni T, Berczi L, Tulassay Z. Elevated insulin-like growth factor 1 receptor, hepatocyte growth factor receptor and telomerase protein expression in mild ulcerative colitis. *Scand J Gastroenterol* 2008; **43**: 289-298 [PMID: 18938767]
 - 42 **Haramis AP**, Begthel H, van den Born M, van Es J, Jonkheer S, Offerhaus GJ, Clevers H. De novo crypt formation and juvenile polyposis on BMP inhibition in mouse intestine. *Science* 2004; **303**: 1684-1686 [PMID: 15017003]
 - 43 **He XC**, Zhang J, Tong WG, Tawfik O, Ross J, Scoville DH, Tian Q, Zeng X, He X, Wiedemann LM, Mishina Y, Li L. BMP signaling inhibits intestinal stem cell self-renewal through suppression of Wnt-beta-catenin signaling. *Nat Genet* 2004; **36**: 1117-1121 [PMID: 15378062]
 - 44 **He XC**, Yin T, Grindley JC, Tian Q, Sato T, Tao WA, Dirisina R, Porter-Westpfahl KS, Hembree M, Johnson T, Wiedemann LM, Barrett TA, Hood L, Wu H, Li L. PTEN-deficient intestinal stem cells initiate intestinal polyposis. *Nat Genet* 2007; **39**: 189-198 [PMID: 17237784]
 - 45 **Dodelet VC**, Pasquale EB. Eph receptors and ephrin ligands: embryogenesis to tumorigenesis. *Oncogene* 2000; **19**: 5614-5619 [PMID: 11114742]
 - 46 **Battle E**, Henderson JT, Begthel H, van den Born MM, Sancho E, Huls G, Meeldijk J, Robertson J, van de Wetering M, Pawson T, Clevers H. Beta-catenin and TCF mediate cell positioning in the intestinal epithelium by controlling the expression of EphB/ephrinB. *Cell* 2002; **111**: 251-263 [PMID: 12408869]
 - 47 **Giannakis M**, Stappenbeck TS, Mills JC, Leip DG, Lovett M, Clifton SW, Ippolito JE, Glasscock JI, Arumugam M, Brent MR, Gordon JI. Molecular properties of adult mouse gastric and intestinal epithelial progenitors in their niches. *J Biol Chem* 2006; **281**: 11292-11300 [PMID: 16464855]
 - 48 **Chen F**, Kook H, Milewski R, Gitler AD, Lu MM, Li J, Nazarian R, Schnepf R, Jen K, Biben C, Runke G, Mackay JP, Novotny J, Schwartz RJ, Harvey RP, Mullins MC, Epstein JA. Hop is an unusual homeobox gene that modulates cardiac development. *Cell* 2002; **110**: 713-723 [PMID: 12297045]
 - 49 **Jensen KB**, Watt FM. Single-cell expression profiling of human epidermal stem and transit-amplifying cells: Lrig1 is a regulator of stem cell quiescence. *Proc Natl Acad Sci USA* 2006; **103**: 11958-11963 [PMID: 16877544]
 - 50 **Wong VW**, Stange DE, Page ME, Buczacki S, Wabik A, Itami S, van de Wetering M, Poulsom R, Wright NA, Trotter MW, Watt FM, Winton DJ, Clevers H, Jensen KB. Lrig1 controls intestinal stem-cell homeostasis by negative regulation of ErbB signalling. *Nat Cell Biol* 2012; **14**: 401-408 [PMID: 22388892 DOI: 10.1038/ncb2464]
 - 51 **van Es JH**, Sato T, van de Wetering M, Lyubimova A, Nee AN, Gregorieff A, Sasaki N, Zeinstra L, van den Born M, Korving J, Martens AC, Barker N, van Oudenaarden A, Clevers H. Dll1+ secretory progenitor cells revert to stem cells upon crypt damage. *Nat Cell Biol* 2012; **14**: 1099-1104 [PMID: 23000963 DOI: 10.1038/ncb2581]
 - 52 **Zhu Y**, Huang YF, Kek C, Bulavin DV. Apoptosis differently affects lineage tracing of Lgr5 and Bmi1 intestinal stem cell populations. *Cell Stem Cell* 2013; **12**: 298-303 [PMID: 23415913 DOI: 10.1016/j.stem.2013.01.003]

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