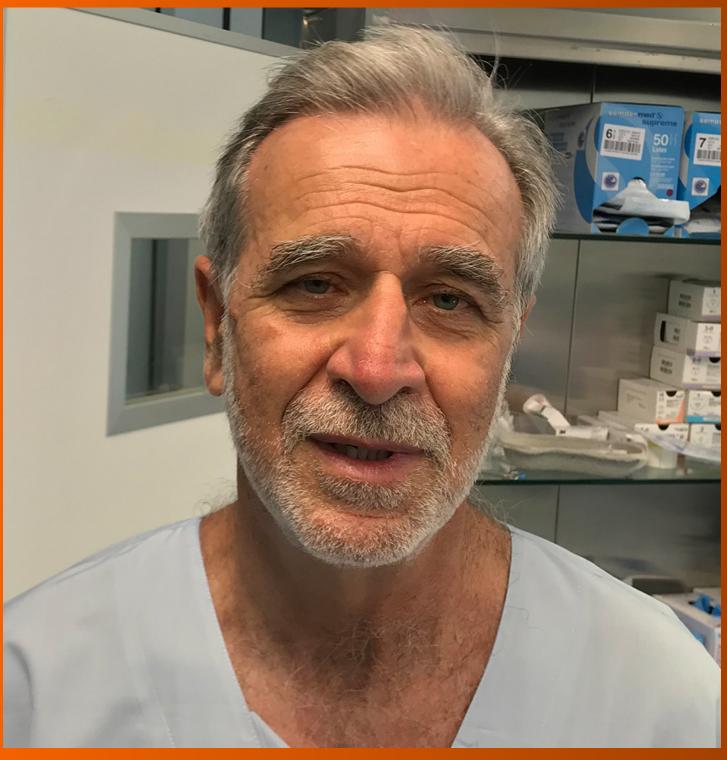
World Journal of Gastroenterology

World J Gastroenterol 2021 August 7; 27(29): 4746-4938





Contents

Weekly Volume 27 Number 29 August 7, 2021

REVIEW

Ischemic bowel disease in 2021 4746

Ahmed M

4763 Therapeutic implications of SARS-CoV-2 dysregulation of the gut-brain-lung axis

Johnson SD, Olwenyi OA, Bhyravbhatla N, Thurman M, Pandey K, Klug EA, Johnston M, Dyavar SR, Acharya A, Podany AT, Fletcher CV, Mohan M, Singh K, Byrareddy SN

4784 Hepatitis B virus infection modeling using multi-cellular organoids derived from human induced pluripotent stem cells

Cao D, Ge JY, Wang Y, Oda T, Zheng YW

MINIREVIEWS

4802 Artificial intelligence in colonoscopy

Joseph J, LePage EM, Cheney CP, Pawa R

4818 New hope for hepatitis C virus: Summary of global epidemiologic changes and novel innovations over 20 years

Dennis BB, Naji L, Jajarmi Y, Ahmed A, Kim D

4831 Impact of modern antiviral therapy of chronic hepatitis B and C on clinical outcomes of liver disease

Khoo T, Lam D, Olynyk JK

4846 Neurological and psychiatric effects of hepatitis C virus infection

Faccioli J, Nardelli S, Gioia S, Riggio O, Ridola L

4862 Muscular abnormalities in liver cirrhosis

Hari A

ORIGINAL ARTICLE

Basic Study

4879 Impact of Fusobacterium nucleatum in the gastrointestinal tract on natural killer cells

Kim YJ, Kim BK, Park SJ, Kim JH

Retrospective Study

4890 Poor performance of anti-mitochondrial antibodies for the diagnosis of primary biliary cholangitis in female Colombian patients: A single-center study

Guatibonza-García V, Gaete PV, Pérez-Londoño A, Puerto-Baracaldo DK, Gutiérrez-Romero SA, Mendivil CO, Tapias M

Contents

Weekly Volume 27 Number 29 August 7, 2021

Clinical Trials Study

4900 Effects of permissive hypocaloric vs standard enteral feeding on gastrointestinal function and outcomes in sepsis

Sun JK, Nie S, Chen YM, Zhou J, Wang X, Zhou SM, Mu XW

Observational Study

4913 Advanced glycation end product: A potential biomarker for risk stratification of non-alcoholic fatty liver disease in ELSA-Brasil study

Pereira ENGDS, Paula DP, Araujo BP, Fonseca MJMD, Diniz MFHS, Daliry A, Griep RH

CASE REPORT

4929 Autoimmune enteropathy and primary biliary cholangitis after proctocolectomy for ulcerative colitis: A case report and review of the literature

 Π

Zhou QY, Zhou WX, Sun XY, Wu B, Zheng WY, Li Y, Qian JM

Contents

Weekly Volume 27 Number 29 August 7, 2021

ABOUT COVER

Editorial Board Member of World Journal of Gastroenterology, Aldo Bove, MD, PhD, Assistant Professor of Surgery, Chief of Surgery "Pierangeli Hospital" Pescara, Department of Medicine, Dentistry and Biotechnology, University "G. D'Annunzio", Via dei Vestini, Chieti 66100, Italy. above@unich.it

AIMS AND SCOPE

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents[®]/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ying-Yi Yuan; Production Department Director: Xiang Li; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREOUENCY

Weekly

EDITORS-IN-CHIEF

Andrzei S Tarnawski, Subrata Ghosh

EDITORIAL BOARD MEMBERS

http://www.wignet.com/1007-9327/editorialboard.htm

PUBLICATION DATE

August 7, 2021

COPYRIGHT

© 2021 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

https://www.wjgnet.com/bpg/gerinfo/204

GUIDELINES FOR ETHICS DOCUMENTS

https://www.wjgnet.com/bpg/GerInfo/287

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

https://www.wjgnet.com/bpg/gerinfo/240

PUBLICATION ETHICS

https://www.wjgnet.com/bpg/GerInfo/288

PUBLICATION MISCONDUCT

https://www.wjgnet.com/bpg/gerinfo/208

ARTICLE PROCESSING CHARGE

https://www.wjgnet.com/bpg/gerinfo/242

STEPS FOR SUBMITTING MANUSCRIPTS

https://www.wjgnet.com/bpg/GerInfo/239

ONLINE SUBMISSION

https://www.f6publishing.com

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 August 7; 27(29): 4802-4817

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

Artificial intelligence in colonoscopy

Joel Joseph, Ella Marie LePage, Catherine Phillips Cheney, Rishi Pawa

ORCID number: Joel Joseph 0000-0002-6307-8505; Ella Marie LePage 0000-0002-0774-9212; Catherine Phillips Cheney 0000-0001-8168-9572; Rishi Pawa 0000-0001-6452-2359.

DOI: 10.3748/wjg.v27.i29.4802

Author contributions: Joseph J provided topic outlining, literature review and original draft preparation; LePage EM performed topic outlining, literature review and original draft preparation; Cheney CP performed literature review and original draft preparation; and Pawa R performed topic outlining, literature review, expertise and manuscript editing.

Conflict-of-interest statement: The authors declare that there are no any conflict of interests.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Joel Joseph, Ella Marie LePage, Department of Internal Medicine, Wake Forest Baptist Medical Center, Winston Salem, NC 27157, United States

Catherine Phillips Cheney, Department of Internal Medicine, Wake Forest School of Medicine, Winston Salem, NC 27157, United States

Rishi Pawa, Department of Internal Medicine, Section of Gastroenterology and Hepatology, Wake Forest Baptist Medical Center, Winston-Salem, NC 27157, United States

Corresponding author: Rishi Pawa, MBBS, Doctor, Department of Internal Medicine, Section of Gastroenterology and Hepatology, Wake Forest Baptist Medical Center, Medical Center Blvd, Winston-Salem, NC 27157, United States. rpawa@wakehealth.edu

Abstract

Colorectal cancer remains a leading cause of morbidity and mortality in the United States. Advances in artificial intelligence (AI), specifically computer aided detection and computer-aided diagnosis offer promising methods of increasing adenoma detection rates with the goal of removing more pre-cancerous polyps. Conversely, these methods also may allow for smaller non-cancerous lesions to be diagnosed in vivo and left in place, decreasing the risks that come with unnecessary polypectomies. This review will provide an overview of current advances in the use of AI in colonoscopy to aid in polyp detection and characterization as well as areas of developing research.

Key Words: Colonoscopy; Artificial intelligence; Computer-aided detection; Detection; Characterization; Computer-aided diagnosis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The rapidly evolving field of artificial intelligence (AI) has found many applications in the field of colonoscopy. Specifically, we describe the technologies that have been developed to detect and characterize colonic polyps with the goal of realtime analysis as well as minimizing the risks of avoidable polypectomies. Additionally, we discuss some of the future directions of AI in this area including advancements in robotic technology.

Citation: Joseph J, LePage EM, Cheney CP, Pawa R. Artificial intelligence in colonoscopy. World J Gastroenterol 2021; 27(29): 4802-4817



WJG https://www.wjgnet.com

Manuscript source: Invited

manuscript

Specialty type: Gastroenterology

and hepatology

Country/Territory of origin: United

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: January 28, 2021 Peer-review started: January 28,

First decision: May 2, 2021 Revised: May 12, 2021 Accepted: July 16, 2021 Article in press: July 16, 2021 Published online: August 7, 2021

P-Reviewer: Lam TYT S-Editor: Ma YJ L-Editor: A P-Editor: Xing YX



URL: https://www.wjgnet.com/1007-9327/full/v27/i29/4802.htm

DOI: https://dx.doi.org/10.3748/wjg.v27.i29.4802

INTRODUCTION

Colorectal cancer (CRC) is the third most common cancer and the second leading cause of cancer related death in men and women in the United States. The incidence of CRC has been declining for over 30 years due in part to screening colonoscopies that detect and remove pre-cancerous polyps[1].

The adenoma detection rate (ADR) is a metric used by endoscopists representing the percentage of time at least one adenoma is detected on screening colonoscopies [2]. Adenoma detection rates differ widely among endoscopists, between 7% to 52%, with higher ADRs associated with a decreased risk of CRC. It is now recommended that endoscopists target an ADR target $\geq 25\%$ [3]. Artificial intelligence (AI), specifically computer-aided detection (CADe) software is being studied to detect polyps during colonoscopy with the goal of increasing adenoma detection rates [2,4,5].

Broadly, artificial intelligence (AI) relates to the ability of a computer program to obtain outside data (e.g., images) and to subsequently take independent actions towards a particular goal (e.g., pattern identification). Machine learning is a form of AI that relies on the analysis of large datasets in order to make predictions that can be used for decision making. Deep learning, a subtype of machine learning, uses an artificial neural network comprised of layers of interconnected "computing units" that mimic biological neural connections and allow for complex "understanding" of input data. This neural network allows the computer program to learn independently from unstructured input data. Many times a deep learning program can process a large number of photos, independently identify patterns among them, and then use that information to make predictions about new images. This powerful technology that has been used to train machines in image and sound recognition is now being applied in the medical field in the form of computer-aided diagnosis and detection, which applies AI and computer vision technologies to the diagnosis of various pathologies. The technology is rapidly expanding in areas like colonoscopy where there is significant room to mitigate human error in visual diagnosis.

Many polyps detected and resected during colonoscopies are diminutive polyps (≤ 5 mm), and a significant number of these are non-neoplastic. Polypectomy increases the risk of complications during colonoscopy, including the risk for bleeding and perforation[6]. Computer-aided diagnosis (CAD) technology has been studied to characterize the histology of polyps in vivo.

The American Society for Gastrointestinal Endoscopy's Preservation and Incorporation of Valuable Endoscopic Innovations (PIVI) initiative has provided guidelines that aim to reduce the cost and need for pathological assessments in addition to reducing the risks associated with polypectomy. The PIVI's first guideline is known as "resect and discard", which entails resecting the diminutive polyp and discarding if the CAD technology used to characterize the polyp has a similar surveillance interval compared to the traditional pathology assessment (≥ 90%). The PIVI's second guideline allows the endoscopist to leave hyperplastic diminutive polyps in place in the rectosigmoid area if the CAD technology has a NPV ≥ 90% for characterizing adenoma histology[7].

COLORECTAL POLYP DETECTION

The first study to use CADe to detect colorectal polyps was published in 2003[8-11]. Karkanis et al[8] used wavelet transformation technology to detect polyps with a sensitivity of 93.6% and specificity of 99.3%. Years later, deep learning networks were applied to CADe, which has paved the way for in vivo and real-time analysis studies.

Urban *et al*[2] was the first to use CADe for polyp detection in real-time. The study assessed 9 standard colonoscopy videos. At the time of the colonoscopies, 28 polyps were identified and removed. Using CADe, an additional 17 polyps were detected, compared to only an additional 8 that were identified by an expert endoscopist[2,9]. Klare et al[12] then applied CADe in real time and in vivo to 55 colonoscopies. The ADR of the CADe was similar to the endoscopists' (29.1% and 30.9%, respectively). However, the CADe was inferior in detecting flat and small polyps.

More recently, there have been randomized control trials (RCTs) performed in realtime using colorectal polyp detection technology. Wang et al[4] included 1058 patients in a non-blinded study, 536 were randomized to colonoscopy and 522 were randomized to colonoscopy with CADe. The ADR was statistically superior in the colonoscopy with the CADe group compared to the control group (29.1% vs 20.3%). Moreover, CADe was better at detecting diminutive adenomas, but there was no difference in detection rate for polyps larger than 5 mm. Notably, the CADe did not miss any polyps, but had 39 false positives alarms for polyps[5]. Wang et al[4] then performed a double blinded RCT in which patients were randomized to a colonoscopy with sham system (n = 478) or a colonoscopy with CADe (n = 484) group. Results showed the ADR was 34% in the CADe group, which was superior to the control group, 28%. Gong et al[13] randomized 704 patients in a partially blinded RCT to CADe assisted colonoscopy or control standard colonoscopy. Similarly, the ADR was significantly better in the CADe group than the control group (16% compared to 8%, respectively). Repici et al[14] performed a similar nonblinded RCT and found an ADR of 54.8% in the colonoscopy with CADe group, which was significantly better than the ADR for the control group (40.4%). The CADe was also able to detect more adenomas that were < 10 mm in size compared to the control group. The authors also found that there was no significant difference in withdrawal time (excluding biopsy time) of the endoscope between the two groups. Liu et al[15] randomized 1026 patients to CADe or control groups. The ADR was significantly better in the CADe group (39%) than the control group (23%). The CADe did not miss any polyps and there were only 36 false positive alarms. It was also noted that the withdrawal times between groups were similar (CADe 6.16 minutes compared to control 6.11 min).

Su et al[16] created an automatic quality control system (AQCS) to improve aspects of colonoscopy using a deep learning model. They randomized 659 patients and found that the AQCS group had a superior ADR than the control group (28.9% vs 16.5%). However, they found the AQCS had a longer withdrawal time (excluding biopsy time) compared to the control group $(7.03 \pm 1.01 \text{ min } vs 5.68 \pm 1.26 \text{ min})$.

Recent meta-analyses have concluded that CADe was accurate at detecting adenomas[17,18]. Barua et al[17] included 5 RCTs (with a total of 4311 patients) and concluded the ADR was significantly better using CADe with colonoscopy (29.6%) than colonoscopy alone (19.3%), with a false positive alarm mean of 11.2%. Lui et al[18] analyzed 6 studies that used CADe and found the accuracy of CADe was 90% with sensitivity and specificity of 95% and 88% respectively. In both studies, colonoscopy with CADe improved detection of diminutive adenomas [17,18].

A significant limitation of CADe technology is the potential to have high false positive alarm rates. Even though Wang et al[5] and Liu et al[15] had low rates, Hassan et al[19] reported a total of 1092 false positive alarms, which averaged 27.3 per colonoscopy. Also, many of the studies assessing CADe had control groups with low ADRs between 8%-23% [5,13,15,16], which is below the recommended target ADR of ≥

Table 1 summarizes the key recent studies in colorectal polyp detection.

POLYP CHARACTERIZATION

Going beyond merely identifying polyps, AI has been applied to provide real-time in vivo diagnoses of neoplastic vs non-neoplastic lesions. A number of different modalities for this have been described over the years.

White light endoscopy

Traditional white light (WL) endoscopy is the most familiar modality used by endoscopists today. High-definition white light (HDWL) endoscopy is the most recent improvement in this area. Recent randomized controlled trials have demonstrated that HDWL is non-inferior to other modalities, specifically narrow band imaging (NBI) and chromoendoscopy[20-22]. In 2016, Rex et al[20] found that there was no statistically significant difference in the number of serrated lesions detected in 804 patients randomized to undergo either WL or NBI endoscopy. The same year, Klare et al[21] published the results of a trial that randomized 380 patients to either HDWL or NBI. They also found no statistically significant advantage of one modality over the other in distinguishing between neoplastic and non-neoplastic polyps. Yang et al[22] randomized 210 patients with ulcerative colitis to undergo colon cancer screening with HDWL or chromoendoscopy and found no significant difference in dysplasia detection rates between the two modalities (5.6% for HDWL vs 3.9% for chromoen-

Table 1 Col	Table 1 Colorectal polyp detection					
Ref.	Study design	Algorithm type	Dataset	Results		
Karkanis et al[8]	Retrospective	CADe (Wavelet Decomposition)	180 images	Sensitivity: 93.6%		
ut[0]				Specificity: 99.3%		
Urban et al [2]	Retrospective	CADe (DCNN)	8461 images &20 colonoscopy videos	Accuracy: 96.4%		
[4]			videos	False Positive: 7%		
Klare et al [12]	ProspectiveIn vivo	CADe	55 colonoscopies	ADR of: CAD 29.1% and Endoscopist 30.9%		
Wang et al [5]	Non-blinded RCT	CADe using Shanghai Wision Al Co. Ltd. (DCNN)	Randomized 522 patients to CADe and 536 to control group	ADR of CAD 29.1% vs control 20.3%		
Wang et al [4]	Double blinded RCT	CADe using EndoScreener (DCNN)	Randomized 484 patients to CAD and 478 to sham system	ADR of CAD 34% vs control 28%		
Gong et al [13]	Partially blinded RCT	CADe using ENDOANGEL (DCNN)	Randomized 355 patients to CAD and 349 to control	ADR of CAD 16% vs control 8%		
Repici <i>et al</i> [14]	Partially-blinded RCT	CADe using GI-Genius (CNN)	Randomized 341 patients to CAD and 344 to control	ADR of CAD 54.8% vs control 40.4%		
Liu et al[15]	Non-blinded RCT	CADe using Henan Xuanweitang Medical Information Technology Co. Ltd (convolutional 3D network)	Randomized 508 patients to CAD and 518 control	ADR of CAD 39% vs control 23%		
Su et al[16]	Partially blinded RCT	Automatic quality control system (ACQS)(DCNN)	Randomized 308 patients to AQCS and 315 to control	ADR of AQCS 28.9% vs control 16.5%		

CADe: Computer-aided detection; CAD: Computer-aided diagnosis; DCNN: Deep convolutional neural network; ADR: Adenoma detection rate.

doscopy).

In 2017, Komeda et al[23] designed and tested a CAD system based on a convolutional neural network to augment WL endoscopy. It functioned as an AI system that trained with previously collected colonoscopy images to assist endoscopists in detecting and diagnosing colon polyps during WL endoscopy. After training on 1200 images, their CAD-neural network system correctly differentiated between adenomatous and non-adenomatous polyps in 70% of newly presented cases[23].

This strategy of applying AI to the detection and classification of colorectal lesions using WL endoscopy has continued to be an area of active study. Researchers have shown that CAD, using both convolutional neural networks and deep learning models, promises to identify suspicious lesions and accurately classify them[24-26]. Zheng et al[24] developed a convoluted neural network (CNN) to be used with WL endoscopy. They trained their AI system with over 600 polyp-containing images from independent public databases and found that their diagnostic model had a sensitivity of 68.3% and a precision of 79.3% when applied to 196 new polyp-containing images [24]. However, they found significant variation in model performance depending on which image database was used to train the CNN, and they also only trained and tested their CNN on still images[24]. Going beyond polyp identification, Yang et al[26] developed a deep learning (DL) model to assist in classification of colorectal lesions during WL endoscopy. They trained their model on 3828 images and validated it on a set of 240 new images. When classifying lesions as neoplastic vs non-neoplastic, their model had a sensitivity of 95.4% and specificity of 30.1% [26]. Their model was also able to classify advanced lesions (high grade dysplasia and stages T1-T4 CRC vs nonadvanced (tubular adenomas and non-neoplastic lesions) with a sensitivity of 80.0% and specificity of 91.3%[26].

A recent prospective crossover study conducted by Wang et al[25] compared traditional WL colonoscopy to CAD-assisted colonoscopy in 369 patients. Patients requiring colonoscopy underwent either traditional WL colonoscopy or CAD-assisted colonoscopy immediately followed by the other, such that each study participant underwent both methods. They found that the adenoma miss rate was 40% for those undergoing traditional colonoscopy first vs 14% in those undergoing CAD-assisted colonoscopy first[25]. Polyp detection followed a similar trend with a miss rate of 46% in those undergoing traditional colonoscopy first vs 13% in those undergoing CADassisted colonoscopy first[25]. Interestingly, of the adenomas missed, participants undergoing CAD-assisted colonoscopy were less likely to have polyps under 5mm and under 10 mm missed, suggesting that CAD is particularly helpful in identifying smaller lesions[25].

The key studies for polyp characterization using white light endoscopy are summarized in Table 2.

Narrow band imaging

Narrow band imaging (NBI) is used to enhance visualization of vascular patterns in the epithelium of lesions to aid in the classification of polyps[27-29]. However, training and experience is needed to operate NBI; therefore, studies have applied computeraided diagnosis to NBI[27,29-34].

Tischendorf et al[29] was the first to apply CAD to magnified NBI. The CAD evaluated 209 polyps and assessed for three vessel features on each NBI image, then a support vector machine was used to classify the polyp as neoplastic or non-neoplastic. It had an accuracy of 85.3%, sensitivity of 90% and specificity of 70.2%. When compared to the consensus of the investigators (accuracy 91.9%, sensitivity 93.8%, and specificity 85.7%), the CAD was inferior. Gross et al[27] performed a similar study with 434 polyps, but assessed for nine vessel features. The CAD had an accuracy of 93.1%, sensitivity of 95%, specificity of 90.3%, and NPV of 92.4%, which was comparable to the results of the expert endoscopists and superior to the novice endoscopists.

Years later, Byrne et al[30] and Chen et al[31] improved CAD by creating deep learning models to analyze NBI images and categorize polyp histology. Both studies only included diminutive polyps (≤ 5 mm). Chen et al[31]'s deep learning model assessed still NBI images of 284 polyps and classified polyps with an accuracy of 90.1%, sensitivity of 96.3%, specificity of 78.1%, PPV of 89.6%, and NPV of 91.5%. The authors also studied the diagnosis time, which was statistically faster for the deep learning model than both expert and novice investigators[31]. Byrne et al[30]'s deep learning model assessed 125 endoscopy NBI video segments of polyps. Of the classified polyps, the accuracy was 94% with sensitivity 98%, specificity 83%, PPV 90%, and NPV 97%. However, the model was not able to classify 15% of polyps due to a lack of confidence. All of the CAD with NBI studies named above with the exception of Tischendorf's initial study met the PIVI criteria for resect and discard or diagnose and leave in situ[7,27,30,31].

Few prospective studies have been performed with CAD using NBI imaging to classify polyp histology. Kominami et al[32] evaluated 118 polyps with NBI. The CAD had an accuracy of 94.9%, sensitivity of 95.9%, specificity of 93.3%, PPV of 95.9%, and NPV 93.3%. The authors used this data to investigate colonoscopy surveillance interval which did not change in 38 of the 41 patients when using the CAD results to classify polyps. Mori et al[33] also performed a study assessing CAD when used with NBI on 466 diminutive polyps. The NPV for rectosigmoid neoplastic polyps ranged from 95.2% to 96.5% depending on the worst or best case scenario respectively.

Most recently, Song et al[35] created a CAD using a deep learning model and tested it in vivo by sending still NBI images during the colonoscopy to a computer. The CAD then categorized the histology in real time. The polyps were classified as serrated, benign adenoma, or deep submucosal cancer with an accuracy of 82.4%, which was superior to trainees (63.8%), but inferior to expert endoscopists (87.3%). The accuracy of trainee endoscopists improved with the addition of CAD to 82.7% showing that CAD can increase the accuracy in this group.

Table 3 provides a summary of the key recent studies of polyp characterization using narrow band imaging.

Laser-induced fluorescence spectroscopy

Another strategy currently under investigation to optically diagnose lesions during endoscopy is laser-induced fluorescence spectroscopy. This diagnostic method relies on low-power laser radiation to induce fluorescence in tissues that can differentiate normal from neoplastic lesions[36]. In recent years, CAD systems have been developed to analyze the fluorescent spectra produced when tissues are exposed to a laser. These CAD systems take advantage of the differences between the fluorescence of normal and pathological tissue to predict the likelihood that a lesion is abnormal.

Kuiper et al[37] and Rath et al[38] studied a laser-induced fluorescence spectroscopy system designed to be used in real-time to help clinicians make decisions regarding biopsy and resection of concerning lesions. However, it is important to note that the accuracy of the algorithm used by Kuiper et al[37] was 73.4% and the NPV only 74.4%, falling short of the performance thresholds of the American Society for Gastrointestinal Endoscopy's PIVI initiative for diminutive lesions. The pilot study conducted by Rath et al[38] was more promising, with an overall accuracy of 84.7%, sensitivity of 81.8%, specificity of 85.2%, and NPV of 96.1%. A 2017 randomized controlled trial by Min et al[39] was further able to demonstrate that use of linked color

Table 2 Wh	Table 2 White light endoscopy					
Ref.	Study design	Algorithm type	Dataset	Results		
Komeda et al[23]	Diagnostic model development	CAD-neural network combination to assist WL endoscopy	1200 training images then tested on 10 new images	Cross-validation accuracy: 0.751		
Zheng et al Diagnostic model		WL endoscopy using YOLO (CNN)	196 WL images from an independent public database	Accuracy: 79.3%		
[24]	development		independent public database	Sensitivity: 68.3%		
Wang et al [25]	Prospective crossover study	Traditional WL endoscopy vs CAD colonoscopy	369 patients from a single hospital in China	Adenoma miss rate of 13.9% in the CAD group vs 40% in the traditional group, P < 0.0001		
Yang et al [26]	Diagnostic model development	Validation of a deep learning model called "ResNet-152" to classify colorectal lesions	3828 WL colonoscopy images from 1339 patients	Mean model accuracy: 79.2% for advanced CRC, early CRC/HGD, TA, and non-neoplastic		
				AUC: 0.818		

CAD: Computer-aided diagnosis; WL: White light; CNN: Convoluted neural network.

imaging technology (which enhances the colors produced by laser endoscopic modalities) improved overall polyp detection rate when compared to traditional white light endoscopy (polyp detection rate 73% for WL and 91% with linked color imaging).

The recent key studies of laser-induced fluorescence spectroscopy are summarized in Table 4.

Autofluorescence endoscopy

Autofluorescence imaging (AFI) is a form of image enhanced endoscopy that differentiates tissues based on their various abilities to capture and reflect fluorescent light [40]. Similar to laser-induced fluorescence spectroscopy, this method takes advantage of endogenous reflective properties of various tissues (fluorophores), but instead of using a laser emitting an exact wavelength of light, AFI uses incoherent light sources. This technology aims to visually highlight tumors, which have more heterogeneous fluorescence on their surface compared to normal colonic mucosa. This imageenhancement displays normally fluorescing mucosa as green and abnormally fluorescing mucosa as red/purple.

In a 2019 study of 802 patients randomized to undergo either AFI endoscopy or white light endoscopy, Takeuchi et al[41] found that using AFI during endoscopy increased the number of flat neoplasms detected overall compared to WL, especially in the ascending colon. However, the overall detection rate of advanced neoplasms was not significantly improved with AF compared to WL[41]. In a meta-analysis of 11 studies, Wanders et al[42] calculated that the sensitivity and specificity of autofluorescence imaging for the optical diagnosis of colonic lesions were 86.7% and 65.9% respectively. This led the authors to conclude that AFI is not as reliable as other methods for visual diagnosis of colonic neoplasms. This was further confirmed in a 2018 meta-analysis by Imperatore et al[43] which found no significant difference between the dysplasia detection rates between AFI and WL (OR = 1.42, 95%CI: 0.74-4.11) when combining the results of two randomized controlled trials representing 92 patients undergoing surveillance colonoscopy.

More recently, researchers have developed CAD systems that can further characterize the images obtained during AFI endoscopy using software that can calculate the green to red light ratios of various tissues encountered during colonoscopy [44-47]. Such developments may help differentiate lesions from normal mucosa in cases where the green to red variation is less obvious, improving on the results of the studies looking at the use of AFI without the use of CAD.

Arita et al [44] created a color-contrast index (CCI) for AFI. Their CCI was developed from 54 colorectal lesions found in 43 patients who underwent either WL or AFI endoscopy. They found that as the CCI increased (i.e., greater contrast between the lesion and the adjacent normal tissue), so did the malignant potential of the assessed lesion (i.e., carcinomas had higher CCIs on average compared to adenomas)[44]. Aihara et al[45] expanded on this idea by using color-analysis software to calculate red/green ratios (RGR) for 102 Lesions in 32 patients undergoing AFI endoscopy. In their study, they were able to differentiate neoplastic from non-neoplastic lesions with sensitivity of 94.2%, specificity of 88.9%, PPV of 95.6%, and NPV of 85.2%[45]. In a similar study, Inomata et al[46] also calculated RGRs to distinguish between non-

Table 3 Narrow band imaging					
Ref.	Study design	Algorithm type	Dataset	Results	
Tischendorf et al[29]	Prospective Ex vivo	CAD - NBI (support vector machine)	209 polyp images	Accuracy: 85.3%	
				Sensitivity: 90%	
				Specificity: 70.2%	
Gross et al[27]	Prospective Ex vivo	CAD - NBI (support vector machine)	434 polyp images	Accuracy: 93.1%	
				Sensitivity: 95%	
				Specificity: 90.3%	
				NPV: 92.4%	
Chen et al[31]	Retrospective	CAD - NBI (DCNN)	284 polyp images	Accuracy: 90.1%	
				Sensitivity: 96.3%	
				Specificity: 78.1%	
				PPV: 89.6%	
				NPV: 91.5%	
Byrne et al[30]	Retrospective	CAD – NBI (DCNN)	125 polyp videos	Accuracy: 94%	
				Sensitivity: 98%	
				Specificity: 83%	
				PPV: 90%	
				NPV: 97%	
Kominami et al[32]	Prospective	CAD -NBI (support vector machine)	118 polyps	Accuracy: 94.9%	
				Sensitivity: 95.9%	
				Specificity: 93.3%	
				PPV: 95.9%	
				NPV: 93.3%	
Mori et al[33]	Prospective	CAD - NBI (support vector machine)	466 polyps	NPV: 95.2% to 96.5%	
Song et al[35]	Prospective In vivo	CAD -NBI (DCNN)	363 polyps	Accuracy: 82.4%	
	-			•	

CAD: Computer-aided diagnosis; NBI: Narrow band imaging; DCNN: Deep convolutional neural network.

neoplastic lesions, adenomas plus superficial cancers, and deep cancers. They were able to characterize hyperplastic polyps and neoplastic lesions, with sensitivity of 83.9%, specificity of 82.6%, PPV of 53.1%, and NPV of 95.6%[46]. Additionally, they were able to differentiate between adenomas plus superficial cancers and deep submucosal cancers with a sensitivity of 80.0%, specificity of 84.4%, PPV of 29.6%, and NPV of 98.1%[46].

In 2019, Horiuchi et al [47] developed software to calculate real-time RGR ratios with the specific goal of identifying diminutive neoplastic rectosigmoid polyps (≤ 5 mm). Using their CAD-assisted AFI endoscopy, they identified 429 diminutive polyps (258 rectosigmoid) in 95 patients. The endoscopists then confirmed whether the lesions identified with the CAD-assisted AFI were actually diminutive neoplastic polyps with trimodal imaging endoscopy (TME) combining findings of WL, AFI, and NBI. The CAD-assisted AFI software was able to identify diminutive neoplastic polyps with a sensitivity of 80.0%, specificity of 95.3%, PPV of 85.2%, and NPV of 93.4%[47].

Table 5 contains a summary of the key recent studies involving autofluorescence endoscopy.

Magnifying chromoendoscopy

4808

In the technique of magnifying chromoendoscopy, suspected colonic lesions are washed with proteinases and colored with an indigo carmine or crystal violet solution in order to allow better visualization of the surface under magnification up to 150 times[48]. In a systematic review, Brown et al[49] showed that chromoendoscopy

Table 4 Laser-induced fluorescence spectroscopy					
Ref.	Study design	Algorithm type	Dataset	Results	
Kuiper et Diagnostic model	Diagnostic performance of WavSTAT	87 patients	Accuracy: 73.4%		
al[<mark>37</mark>]	development			NPV: 74.4%	
	Diagnostic model	Diagnostic performance of WavSTAT for	27 patients	Accuracy: 84.7%	
[38] development		predicting polyp histology		Sensitivity: 81.8%	
				Specificity: 85.2%	
				NPV: 96.1%	
Min <i>et al</i> [39]	Randomized controlled trial	Linked color imaging with laser endoscopic system vs WL	141 patients from 3 hospitals in China	Polyp detection rate of 91% in the LCI group, 73% in the WL group, $P < 0.0001$	

WL: White light.

Table 5 Autofluorescence endoscopy				
Ref.	Study design	Algorithm type	Dataset	Results
Arita et al[44]	Diagnostic model development	Calculation of a color-contrast index (CCI) for AFI	43 patients who underwent both WL and AF endoscopy	Sensitivity: 95.3%
				Specificity: 63.6%
Aihara et al[45]	Diagnostic model development	CAD-assisted AF	32 patients undergoing colonoscopy in a Japanese hospital	Sensitivity: 94.2%
				Specificity: 88.9%
				PPV: 95.6%
				NPV: 85.2%
nomata <i>et al</i> 46]	Diagnostic model development	CAD-assisted AF	88 patients	Accuracy: 82.8%
				Sensitivity: 83.9%
				Specificity: 82.6%
				PPV: 53.1%
				NPV: 95.6%
Horiuchi et al 47]	Diagnostic model development	CAD-assisted AF	95 patients undergoing colonoscopy	Accuracy: 91.5%
				Sensitivity: 80.0%
				Specificity: 95.3%
				PPV: 85.2%
				NPV: 93.4%

AFI: Autofluorescence imaging; WL: White light; CAD: Computer-aided diagnosis; AF: Autofluorescence.

significantly increased both the number of patients with polyps (OR = 1.87, 95%CI: 1.51-2.3) and neoplasms detected (OR = 1.53, 95%CI: 1.31-1.79). Kudo et al[48] demonstrated that certain pit patterns on magnifying chromoendoscopy are associated with malignancy, showing sensitivity of 97.8%, specificity of 91.4%, and accuracy of 97.1%. Kanao et al[50] demonstrated that magnifying chromoendoscopy can be used to differentiate severely irregular lesions from those with only mild irregularities, a key distinction as 56.1% of the former are associated with deep submucosal invasive

adenomas vs only 6.7% of the latter.

Several automated computer-based systems have been developed for analysis of pit patterns. The system developed by Takemura et al[51] was able to accurately diagnose 132 out of 134 (98.5%) of images captured with magnifying chromoendoscopy demonstrating that artificial intelligence aided systems can reliably predict histological changes compared to endoscopists. Häfner et al[52] used texture analysis of magnified chromoendoscopic images to achieve accuracy as high as 99.59%. Recent work by Qi et al[53] further showed that artificial intelligence can be used to quantify colonic crypts and provide objective measures of area, density, eccentricity, solidity, straightness, and parallelism which in turn can be used to reduce variability compared to human

Recent studies on the use of magnifying chromoendoscopy in polyp characterization are summarized in Table 6.

Endocytoscopy

The technique of endocytoscopy uses a contact light microscope attached to a colonoscope to provide endoscopic images with ultra-magnification up to 520 times. The addition of staining allows for real-time histological diagnoses to be made. Studies have shown that the accuracy of this technique is comparable to traditional biopsy [54]. However, a major limitation is the need for expert experience in order to make real-time diagnoses. CAD has been developed in response to this shortcoming.

In 2015, Mori et al[54] described the use of an endocytoscopic imaging computeraided diagnostic system. In this study, 39 non-neoplastic and 176 neoplastic small colorectal polyps less than 10 mm in size were analyzed by artificial intelligence software and compared to the results of both expert and trainee endoscopists. They showed comparable sensitivity (92% vs 92.7%) and accuracy (89.2% vs 92.3%) of the computer-aided diagnostic system compared to experts[54]. Moreover, the artificial intelligence program performed significantly better than trainee endoscopists who only had sensitivity of 81.8% and accuracy of 80.4% [54]. Takeda et al [55] described the development of a CAD system that used 5543 endocytoscopic images for machine learning. Following this, 188 images of a mix of adenomas and invasive cancers were analyzed by the CAD system and compared to pathological diagnoses with 89.4% sensitivity, 98.9% specificity, 98.8% accuracy, 98.8% PPV, and 90.1% NPV[55]. Moreover, the system used a support vector machine to calculate the probability of results being classified accurately. The study specifically looked at high-confidence diagnoses described as those having a ≥ 90 % probability of being correct. Out of 188 images analyzed, 134 fell into this category with 98.1% sensitivity, 100% specificity, 99.3% accuracy, 100% PPV, and 98.8% NPV[55].

In 2018, Mori et al[33] showed the efficacy of real-time endocytoscopy with CAD in detecting diminutive polyps ≤ 5 mm in size. 466 polyps were assessed with 98.1% pathologic prediction rate, 93.8% sensitivity, 90.3% specificity, and 94.1% PPV. Moreover, they were able to demonstrate overall negative predictive value of 96.4% which is significantly above the threshold for a "diagnose-and-leave" treatment strategy. However, the study only demonstrated 65.8% NPV for lesions proximal to the recto-sigmoid area[33].

In 2020, Kudo et al[56] performed a study to evaluate the efficacy of an AI system that uses endocytoscopic images to look at cell nuclei, crypt structure, and microvessels. It was able to identify malignant lesions with 96.9% sensitivity, 100% specificity, 98% accuracy, 100% PPV, and 94.6% NPV. Compared to expert endoscopists with 92.8% sensitivity, 94.3% specificity, and 93.9% accuracy as well as trainee endoscopists with 70.8% sensitivity, 65.7% specificity, and 69% accuracy, this CAD system significantly outperformed both groups[56].

Table 7 summarizes the key recent studies in endocytoscopy.

Confocal endomicroscopy

Anatomic variation can hinder accurate traditional endoscopic biopsy. Confocal endomicroscopy produces high resolution magnification of the mucosal layer of the gastrointestinal tract using laser illumination with simultaneous detection of light reflected from the tissue through a narrow pinhole. By filtering out light that is scattered from angles outside of layers corresponding to mucosa in question, this technique allows for high spatial resolution and real-time endoscopic evaluation of targeted areas of tissue. Compared to normal colonic mucosa with well-organized crypt structures, malignancy causes irregularities and interruptions[57].

André et al[58] designed software that used probe-based confocal laser endomicroscopy to automatically classify colonic polyps. 135 images with both neoplastic and nonneoplastic polyps were analyzed by the automated software with 92.5% sensitivity,

Table 6 Magnifying chromoendoscopy					
Ref.	Study design	Algorithm type	Dataset	Results	
Takemura et al[51]	Partially blinded retrospective study	CAD using HuPAS	134 pit pattern images	Accuracy: 98.5%	
Häfner et al [52]	Partially blinded retrospective study	CAD using Dual-Tree Complex Wavelet Transform	484 RGB pit pattern images	Accuracy: 99.59%	
Qi et al[53]	Diagnostic model development	CAD using automated imaged analysis	79 colon samples (14 normal, 44 normal tissue adjacent to cancer, 21 malignant)	Automated segmentation achieved precision ratio of 0.69 and match ratio of 0.73	

CAD: Computer-aided diagnosis.

83.3% specificity, and 89.6% accuracy which was not significantly different compared to diagnosis by two expert endoscopists with 91.4% sensitivity, 85.7% specificity, and 89.6% accuracy. Ştefănescu et al[59] retrospectively analyzed 1035 endomicroscopic images processing them through a CAD system that allowed for feature identification via fractal analysis of glandular structures showing that homogeneity and feature number were significantly different in malignancy. In turn, this was used to design an artificial intelligence program with a diagnosis error rate of 15.5% [59].

One problem with early probe-based confocal endomicroscopic images is the need for high-level magnification which leads to a long learning-time for automated image interpretation. Taunk et al[60] showed the efficacy of a CAD algorithm that utilized a lower magnification method with a wider field-of-view necessitating less images. The algorithm demonstrated similar sensitivity, specificity, and accuracy compared to expert endoscopists (95% vs 98%, 94% vs 95%, and 94% vs 96% respectively) and significantly better performance than less experienced endoscopists who only had sensitivity of 60%, specificity of 85%, and accuracy of 73% [60].

The recent key studies on confocal endomicroscopy are summarized in Table 8.

FUTURE DIRECTIONS

Automated polyp detection and characterization

Many studies have been performed using AI to detect or classify the histology of colorectal polyps; however, little research has been done on simultaneous detection and classification of colorectal polyps using AI. Mori et al[33] combined previously studied CADe and CAD systems to develop AI technology that is able to detect then characterize the colorectal polyps[61,62]. White-light imaging was used to detect polyps with an accuracy of 94%[61,62]. Then, classification of the polyps was then performed using magnified NBI with a NPV of 95.2% [33,61]. Ozawa et al [34] created a deep convolutional neural network (DCNN) that detected polyps using white light or NBI with a sensitivity of 92% and a PPV of 86%. The DCNN then classified the polyps using either white light with an accuracy of 83% and NPV of 90% or NBI with an accuracy of 81% and NPV of 91%. Further research is needed in concurrent automated detection and characterization of colorectal polyps.

Robotics

Much of the work in robotics has centered around the use of self-propelling colonoscopes and less on polyp detection. Recent studies on the use of robotics in colonoscopy are summarized in Table 9.

Eickhoff et al[63] demonstrated the first use of a novel computer-assisted colonoscope in 2007 that would change shape at 16 different segments depending on insertion depth in an effort to decrease discomfort from colonoscope looping. While it required an endoscopist to steer the scope, the device used CAD to change shape as it was advanced. The device was able to intubate the cecum in 100% of patients with no complications at discharge, 48 h, and 30 d[63].

In 2016, Pullens et al[64] demonstrated the utility of colonoscopy with robotic steering and automated lumen centralization (RS-ALC). In a study of 18 endoscopists including 8 experts and 10 novices, the addition of RS-ALC significantly improved the time to intubate the cecum in novices (8 min 56 s compared to baseline 11 min 47 s without RS-ALC) as well as polyp detection rate (88.1% vs 78.6%)[64]. However,

Table 7 Endocytoscopy				
Ref.	Study design	Algorithm type	Dataset	Results
Mori et al	Pilot study	CAD using EC-CAD	176 colorectal polyps from 152 patients	Accuracy: 89.2%
[54]				Sensitivity: 92%
				Specificity 79.5%
Takeda et al	Retrospective study	CAD using EC-CAD	5543 endocytoscopy images for machine	Overall
[55]			learning. 200 test images	Accuracy: 94%
				Sensitivity: 89.4%
				Specificity: 98.9%
				PPV: 98.8%
				NPV: 90.1%
				High-confidence diagnosis
				Accuracy: 99.3%
				Sensitivity: 98.1%
				Specificity: 100%
				PPV: 100%
				NPV: 98.8%
Mori et al	Single-group, open-label,	Real-time CAD during	466 diminutive polyps from 325 patients	Accuracy: 98.1%
[33]	prospective study	colonoscopy		Sensitivity 93.8%
				Specificity 90.3%
				PPV 94.1%
				NPV 89.8%
Kudo et al	Retrospective study	CAD using EndoBRAIN	100 polyps from 89 patients	Accuracy: 98%
[56]				Sensitivity 96.9%
				Specificity 100%
				PPV 100%
				NPV 94.6%

CAD: Computer-aided diagnosis.

similar results were not seen with expert endoscopists whose time to intubate the cecum actually increased (13 min 1 s compared to baseline 2 min 9 s without RS-ALC) and polyp detection rate decreased (69% vs 80.9%)[64]. Slawinski et al[65] demonstrated the use of an in vivo autonomously controlled highly compliant magnetic flexible endoscope with diagnostic and therapeutic capability using an actuating permanent magnet in animal studies. They were able to conduct autonomous endoscopic retroflexion with 100% success in 30 attempts without perforation or trauma in pigs. However, diagnostic capability was worse than traditional endoscopy with an average detection miss rate of 21.7% and completion time of 575 s (compared to 5% and 257 s). When looking at lesion targeting alone, the robotic program took on average 251 s to identify lesions compared to only 32 s with traditional endoscopy[65].

In 2020, Formosa et al[66] showed the use of a sensor-enabled treaded robotic colonoscope with multiple degrees of freedom which was notable for containing all the functions of a traditional endoscope including direct visualization, channels for insufflation and irrigation as well as a tool port for endoscopy. It also had inertial measurement technology in addition to a magnetometer, motor encoders, and motor current sensors for future autonomous use. Ex-vivo porcine results showed locomotion ability up to 40 mm/s[66].

Table 8 Confocal endomicroscopy					
Ref.	Study design	Algorithm type	Dataset	Results	
Andréet al	Diagnostic model development	CAD using content based image retrieval (CBIR)	135 polyps from 71 patients	Accuracy: 89.6%	
[58]	development	approach		Sensitivity 92.5%	
				Specificity 83.3%	
Ştefănescu et al[59]	Diagnostic model development	CAD using NAVICAD and a two layer CNN	1035 endomicroscopy images including 725 for training, 155 for validation, and 155 for testing.	Testing decision accuracy error rate of 15.48% (24 out of 155 images)	
Taunk et al	Feasibility study	CAD using expectation-	189 endomicroscopy images from 26 patient	Accuracy: 94.2%	
[60]		maximization algorithm		Sensitivity 94.8%	
				Specificity 93.5%	

CAD: Computer-aided diagnosis; CNN: Convoluted neural network.

Table 9 R	Table 9 Robotics					
Ref.	Study design	Algorithm type	Dataset	Results		
Eickhoff et al[63]	Prospective, nonrandomized, unblinded feasibility study	CAD using NeoGuide Endoscopy System	10 patients	100% cecal intubation rate. Median time to cecum 20.5 min. 0 complications or adverse effects reported at discharge, 48 h, and 30 d		
Pullens et al[64]	Randomized control trial	CAD using automated lumen	8 expert endoscopists and 10 endoscopy-	Novice		
ալ04յ	with crossover design	vith crossover design automated lumen naïve novices performing endoscopy on a validated colon model with 21 polyps	Accuracy: 88.1%			
				Time to cecum: 8 min 56 s		
				Experts		
				Accuracy: 69%		
				Time to cecum: 13 min 1 s		

CAD: Computer-aided diagnosis.

CONCLUSION

Traditional colonoscopy has been shown to reduce colon cancer incidence by more than 80%[67]. The application of artificial intelligence, computer-aided detection, and computer-aided diagnosis to this field provides possibilities for improving an already powerful tool. Namely, the possibility of combining these technologies for real-time endoscopic detection and analysis of lesions overall makes the procedure less operator dependent. With the ability to identify smaller diminutive lesions as non-cancerous, these techniques also offer time and resource savings. Given the widespread use of colonoscopy as a screening test and an aging world population, this potentially translates to billions of dollars in cost reductions and even the possibility of extending screening intervals. Multiple modalities have shown to increase adenoma detection rates and have negative predictive values > 90%, meeting the goals of the ASGE's PIVI initiatives.

However, multiple challenges remain including a lack of large multicenter clinical trials and comparison of computer-aided detection and diagnosis modalities. Additionally, more widespread regulatory approval on government and payer levels is needed. There is still much room for clinical research following software development and more prospective studies evaluating the real-life application of these technologies in the endoscopy suite. Nonetheless, continued development of CADe, CAD and AI in colonoscopy offers patients the possibility of living longer and healthier lives.

REFERENCES

- American Cancer Society. Colorectal Cancer Facts & Figures 2020-2022. Atlanta: American Cancer Society, 2020
- Urban G, Tripathi P, Alkayali T, Mittal M, Jalali F, Karnes W, Baldi P. Deep Learning Localizes and Identifies Polyps in Real Time With 96% Accuracy in Screening Colonoscopy. Gastroenterology 2018; 155: 1069-1078.e8 [PMID: 29928897 DOI: 10.1053/j.gastro.2018.06.037]
- Rex DK, Schoenfeld PS, Cohen J, Pike IM, Adler DG, Fennerty MB, Lieb JG 2nd, Park WG, Rizk MK, Sawhney MS, Shaheen NJ, Wani S, Weinberg DS. Quality indicators for colonoscopy. Gastrointest Endosc 2015; 81: 31-53 [PMID: 25480100 DOI: 10.1016/j.gie.2014.07.058]
- Wang P, Liu X, Berzin TM, Glissen Brown JR, Liu P, Zhou C, Lei L, Li L, Guo Z, Lei S, Xiong F, Wang H, Song Y, Pan Y, Zhou G. Effect of a deep-learning computer-aided detection system on adenoma detection during colonoscopy (CADe-DB trial): a double-blind randomised study. Lancet Gastroenterol Hepatol 2020; 5: 343-351 [PMID: 31981517 DOI: 10.1016/S2468-1253(19)30411-X]
- Wang P, Berzin TM, Glissen Brown JR, Bharadwaj S, Becq A, Xiao X, Liu P, Li L, Song Y, Zhang D, Li Y, Xu G, Tu M, Liu X. Real-time automatic detection system increases colonoscopic polyp and adenoma detection rates: a prospective randomised controlled study. Gut 2019; 68: 1813-1819 [PMID: 30814121 DOI: 10.1136/gutjnl-2018-317500]
- ASGE Standards of Practice Committee, Fisher DA, Maple JT, Ben-Menachem T, Cash BD, Decker GA, Early DS, Evans JA, Fanelli RD, Fukami N, Hwang JH, Jain R, Jue TL, Khan KM, Malpas PM, Sharaf RN, Shergill AK, Dominitz JA. Complications of colonoscopy. Gastrointest Endosc 2011; 74: 745-752 [PMID: 21951473 DOI: 10.1016/j.gie.2011.07.025]
- Rex DK, Kahi C, O'Brien M, Levin TR, Pohl H, Rastogi A, Burgart L, Imperiale T, Ladabaum U, Cohen J, Lieberman DA. The American Society for Gastrointestinal Endoscopy PIVI (Preservation and Incorporation of Valuable Endoscopic Innovations) on real-time endoscopic assessment of the histology of diminutive colorectal polyps. Gastrointest Endosc 2011; 73: 419-422 [PMID: 21353837 DOI: 10.1016/j.gie.2011.01.023]
- Karkanis SA, Iakovidis DK, Maroulis DE, Karras DA, Tzivras M. Computer-aided tumor detection in endoscopic video using color wavelet features. IEEE Trans Inf Technol Biomed 2003; 7: 141-152 [PMID: 14518727 DOI: 10.1109/titb.2003.813794]
- Kim KO, Kim EY. Application of Artificial Intelligence in the Detection and Characterization of Colorectal Neoplasm. Gut Liver 2021; 15: 346-353 [PMID: 32773386 DOI: 10.5009/gnl20186]
- Kudo SE, Mori Y, Misawa M, Takeda K, Kudo T, Itoh H, Oda M, Mori K. Artificial intelligence and colonoscopy: Current status and future perspectives. Dig Endosc 2019; 31: 363-371 [PMID: 30624835 DOI: 10.1111/den.13340]
- Maroulis DE, Iakovidis DK, Karkanis SA, Karras DA. CoLD: a versatile detection system for colorectal lesions in endoscopy video-frames. Comput Methods Programs Biomed 2003; 70: 151-166 [PMID: 12507791 DOI: 10.1016/s0169-2607(02)00007-x]
- Klare P, Sander C, Prinzen M, Haller B, Nowack S, Abdelhafez M, Poszler A, Brown H, Wilhelm D, Schmid RM, von Delius S, Wittenberg T. Automated polyp detection in the colorectum: a prospective study (with videos). Gastrointest Endosc 2019; 89: 576-582.e1 [PMID: 30342029 DOI: 10.1016/j.gie.2018.09.042]
- Gong D, Wu L, Zhang J, Mu G, Shen L, Liu J, Wang Z, Zhou W, An P, Huang X, Jiang X, Li Y, Wan X, Hu S, Chen Y, Hu X, Xu Y, Zhu X, Li S, Yao L, He X, Chen D, Huang L, Wei X, Wang X, Yu H. Detection of colorectal adenomas with a real-time computer-aided system (ENDOANGEL): a randomised controlled study. Lancet Gastroenterol Hepatol 2020; 5: 352-361 [PMID: 31981518 DOI: 10.1016/S2468-1253(19)30413-31
- Repici A, Badalamenti M, Maselli R, Correale L, Radaelli F, Rondonotti E, Ferrara E, Spadaccini M, Alkandari A, Fugazza A, Anderloni A, Galtieri PA, Pellegatta G, Carrara S, Di Leo M, Craviotto V, Lamonaca L, Lorenzetti R, Andrealli A, Antonelli G, Wallace M, Sharma P, Rosch T, Hassan C. Efficacy of Real-Time Computer-Aided Detection of Colorectal Neoplasia in a Randomized Trial. Gastroenterology 2020; **159**: 512-520.e7 [PMID: 32371116 DOI: 10.1053/j.gastro.2020.04.062]
- Liu WN, Zhang YY, Bian XQ, Wang LJ, Yang Q, Zhang XD, Huang J. Study on detection rate of polyps and adenomas in artificial-intelligence-aided colonoscopy. Saudi J Gastroenterol 2020; 26: 13-19 [PMID: 31898644 DOI: 10.4103/sjg.SJG_377_19]
- Su JR, Li Z, Shao XJ, Ji CR, Ji R, Zhou RC, Li GC, Liu GQ, He YS, Zuo XL, Li YQ. Impact of a real-time automatic quality control system on colorectal polyp and adenoma detection: a prospective randomized controlled study (with videos). Gastrointest Endosc 2020; 91: 415-424.e4 [PMID: 31454493 DOI: 10.1016/j.gie.2019.08.026]
- Barua I, Vinsard DG, Jodal HC, Løberg M, Kalager M, Holme Ø, Misawa M, Bretthauer M, Mori Y. Artificial intelligence for polyp detection during colonoscopy: a systematic review and meta-analysis. Endoscopy 2021; 53: 277-284 [PMID: 32557490 DOI: 10.1055/a-1201-7165]
- Lui TKL, Guo CG, Leung WK. Accuracy of artificial intelligence on histology prediction and detection of colorectal polyps: a systematic review and meta-analysis. Gastrointest Endosc 2020; 92: 11-22.e6 [PMID: 32119938 DOI: 10.1016/j.gie.2020.02.033]
- Hassan C, Badalamenti M, Maselli R, Correale L, Iannone A, Radaelli F, Rondonotti E, Ferrara E, Spadaccini M, Alkandari A, Fugazza A, Anderloni A, Galtieri PA, Pellegatta G, Carrara S, Di Leo M, Craviotto V, Lamonaca L, Lorenzetti R, Andrealli A, Antonelli G, Wallace M, Sharma P, Rösch T, Repici A. Computer-aided detection-assisted colonoscopy: classification and relevance of false

4814

- positives. Gastrointest Endosc 2020; 92: 900-904.e4 [PMID: 32561410 DOI: 10.1016/j.gie.2020.06.021]
- Rex DK, Clodfelter R, Rahmani F, Fatima H, James-Stevenson TN, Tang JC, Kim HN, McHenry L, Kahi CJ, Rogers NA, Helper DJ, Sagi SV, Kessler WR, Wo JM, Fischer M, Kwo PY. Narrow-band imaging vs white light for the detection of proximal colon serrated lesions: a randomized, controlled trial. Gastrointest Endosc 2016; 83: 166-171 [PMID: 25952085 DOI: 10.1016/j.gie.2015.03.1915]
- Klare P. Haller B. Wormbt S. Nötzel E. Hartmann D. Albert J. Hausmann J. Einwächter H. Weber A. 21 Abdelhafez M, Schmid RM, von Delius S. Narrow-band imaging vs. high definition white light for optical diagnosis of small colorectal polyps: a randomized multicenter trial. Endoscopy 2016; 48: 909-915 [PMID: 27448051 DOI: 10.1055/s-0042-110650]
- Yang DH, Park SJ, Kim HS, Park YS, Park DI, Lee KM, Jung SA, Choi CH, Koo JS, Cheon JH, Yang SK, Kim WH, Kim J, Kim H, Ryan Choi CH; Korean Association for the Study of the Intestinal Diseases (KASID) study. High-Definition Chromoendoscopy Versus High-Definition White Light Colonoscopy for Neoplasia Surveillance in Ulcerative Colitis: A Randomized Controlled Trial. Am J Gastroenterol 2019; 114: 1642-1648 [PMID: 31567166 DOI: 10.14309/ajg.0000000000000341]
- Komeda Y, Handa H, Watanabe T, Nomura T, Kitahashi M, Sakurai T, Okamoto A, Minami T, Kono M, Arizumi T, Takenaka M, Hagiwara S, Matsui S, Nishida N, Kashida H, Kudo M. Computer-Aided Diagnosis Based on Convolutional Neural Network System for Colorectal Polyp Classification: Preliminary Experience. Oncology 2017; 93 Suppl 1: 30-34 [PMID: 29258081 DOI: 10.1159/000481227]
- Zheng Y, Zhang R, Yu R, Jiang Y, Mak TWC, Wong SH, Lau JYW, Poon CCY. Localisation of Colorectal Polyps by Convolutional Neural Network Features Learnt from White Light and Narrow Band Endoscopic Images of Multiple Databases. Annu Int Conf IEEE Eng Med Biol Soc 2018; 2018: 4142-4145 [PMID: 30441267 DOI: 10.1109/EMBC.2018.8513337]
- Wang P, Liu P, Glissen Brown JR, Berzin TM, Zhou G, Lei S, Liu X, Li L, Xiao X. Lower Adenoma 25 Miss Rate of Computer-Aided Detection-Assisted Colonoscopy vs Routine White-Light Colonoscopy in a Prospective Tandem Study. Gastroenterology 2020; 159: 1252-1261.e5 [PMID: 32562721 DOI: 10.1053/j.gastro.2020.06.023]
- Yang YJ, Cho BJ, Lee MJ, Kim JH, Lim H, Bang CS, Jeong HM, Hong JT, Baik GH. Automated Classification of Colorectal Neoplasms in White-Light Colonoscopy Images via Deep Learning. J Clin Med 2020; 9 [PMID: 32456309 DOI: 10.3390/jcm9051593]
- Gross S, Trautwein C, Behrens A, Winograd R, Palm S, Lutz HH, Schirin-Sokhan R, Hecker H, Aach T, Tischendorf JJ. Computer-based classification of small colorectal polyps by using narrow-band imaging with optical magnification. Gastrointest Endosc 2011; 74: 1354-1359 [PMID: 22000791 DOI: 10.1016/j.gie.2011.08.001]
- McGill SK, Evangelou E, Ioannidis JP, Soetikno RM, Kaltenbach T. Narrow band imaging to differentiate neoplastic and non-neoplastic colorectal polyps in real time: a meta-analysis of diagnostic operating characteristics. Gut 2013; 62: 1704-1713 [PMID: 23300139 DOI: 10.1136/gutinl-2012-303965]
- Tischendorf JJ, Gross S, Winograd R, Hecker H, Auer R, Behrens A, Trautwein C, Aach T, Stehle T. Computer-aided classification of colorectal polyps based on vascular patterns: a pilot study. Endoscopy 2010; 42: 203-207 [PMID: 20101564 DOI: 10.1055/s-0029-1243861]
- Byrne MF, Chapados N, Soudan F, Oertel C, Linares Pérez M, Kelly R, Iqbal N, Chandelier F, Rex 30 DK. Real-time differentiation of adenomatous and hyperplastic diminutive colorectal polyps during analysis of unaltered videos of standard colonoscopy using a deep learning model. Gut 2019; 68: 94-100 [PMID: 29066576 DOI: 10.1136/gutjnl-2017-314547]
- Chen PJ, Lin MC, Lai MJ, Lin JC, Lu HH, Tseng VS. Accurate Classification of Diminutive Colorectal Polyps Using Computer-Aided Analysis. Gastroenterology 2018; 154: 568-575 [PMID: 29042219 DOI: 10.1053/j.gastro.2017.10.010]
- Kominami Y, Yoshida S, Tanaka S, Sanomura Y, Hirakawa T, Raytchev B, Tamaki T, Koide T, Kaneda K, Chayama K. Computer-aided diagnosis of colorectal polyp histology by using a real-time image recognition system and narrow-band imaging magnifying colonoscopy. Gastrointest Endosc 2016; **83**: 643-649 [PMID: 26264431 DOI: 10.1016/j.gie.2015.08.004]
- Mori Y, Kudo SE, Misawa M, Saito Y, Ikematsu H, Hotta K, Ohtsuka K, Urushibara F, Kataoka S, Ogawa Y, Maeda Y, Takeda K, Nakamura H, Ichimasa K, Kudo T, Hayashi T, Wakamura K, Ishida F, Inoue H, Itoh H, Oda M, Mori K. Real-Time Use of Artificial Intelligence in Identification of Diminutive Polyps During Colonoscopy: A Prospective Study. Ann Intern Med 2018; 169: 357-366 [PMID: 30105375 DOI: 10.7326/M18-0249]
- Ozawa T, Ishihara S, Fujishiro M, Kumagai Y, Shichijo S, Tada T. Automated endoscopic detection and classification of colorectal polyps using convolutional neural networks. Therap Adv Gastroenterol 2020; 13: 1756284820910659 [PMID: 32231710 DOI: 10.1177/1756284820910659]
- Song EM, Park B, Ha CA, Hwang SW, Park SH, Yang DH, Ye BD, Myung SJ, Yang SK, Kim N, Byeon JS. Endoscopic diagnosis and treatment planning for colorectal polyps using a deep-learning model. Sci Rep 2020; 10: 30 [PMID: 31913337 DOI: 10.1038/s41598-019-56697-0]
- Kapadia CR, Cutruzzola FW, O'Brien KM, Stetz ML, Enriquez R, Deckelbaum LI. Laser-induced fluorescence spectroscopy of human colonic mucosa. Detection of adenomatous transformation. Gastroenterology 1990; 99: 150-157 [PMID: 2160898 DOI: 10.1016/0016-5085(90)91242-x]
- Kuiper T, Alderlieste YA, Tytgat KM, Vlug MS, Nabuurs JA, Bastiaansen BA, Löwenberg M,

4815

- Fockens P, Dekker E. Automatic optical diagnosis of small colorectal lesions by laser-induced autofluorescence. Endoscopy 2015; 47: 56-62 [PMID: 25264763 DOI: 10.1055/s-0034-1378112]
- 38 Rath T, Tontini GE, Vieth M, Nägel A, Neurath MF, Neumann H. In vivo real-time assessment of colorectal polyp histology using an optical biopsy forceps system based on laser-induced fluorescence spectroscopy. Endoscopy 2016; 48: 557-562 [PMID: 27009081 DOI: 10.1055/s-0042-102251]
- Min M, Deng P, Zhang W, Sun X, Liu Y, Nong B. Comparison of linked color imaging and whitelight colonoscopy for detection of colorectal polyps: a multicenter, randomized, crossover trial. Gastrointest Endosc 2017; 86: 724-730 [PMID: 28286095 DOI: 10.1016/j.gie.2017.02.035]
- Renkoski TE, Banerjee B, Graves LR, Rial NS, Reid SA, Tsikitis VL, Nfonsam VN, Tiwari P, Gavini H, Utzinger U. Ratio images and ultraviolet C excitation in autofluorescence imaging of neoplasms of the human colon. J Biomed Opt 2013; 18: 16005 [PMID: 23291657 DOI: 10.1117/1.JBO.18.1.016005
- Takeuchi Y, Sawaya M, Oka S, Tamai N, Kawamura T, Uraoka T, Ikematsu H, Moriyama T, Arao M, Ishikawa H, Ito Y, Matsuda T. Efficacy of autofluorescence imaging for flat neoplasm detection: a multicenter randomized controlled trial (A-FLAT trial). Gastrointest Endosc 2019; 89: 460-469 [PMID: 30452914 DOI: 10.1016/j.gie.2018.11.012]
- 42 Wanders LK, East JE, Uitentuis SE, Leeflang MM, Dekker E. Diagnostic performance of narrowed spectrum endoscopy, autofluorescence imaging, and confocal laser endomicroscopy for optical diagnosis of colonic polyps: a meta-analysis. Lancet Oncol 2013; 14: 1337-1347 [PMID: 24239209 DOI: 10.1016/S1470-2045(13)70509-6]
- Imperatore N, Castiglione F, Testa A, De Palma GD, Caporaso N, Cassese G, Rispo A. Augmented Endoscopy for Surveillance of Colonic Inflammatory Bowel Disease: Systematic Review With Network Meta-analysis. J Crohns Colitis 2019; 13: 714-724 [PMID: 30597029 DOI: 10.1093/ecco-jcc/jjv218]
- Arita K, Mitsuyama K, Kawano H, Hasegawa S, Maeyama Y, Masuda J, Akagi Y, Watanabe Y, Okabe Y, Tsuruta O, Sata M. Quantitative analysis of colorectal mucosal lesions by autofluorescence endoscopy: discrimination of carcinomas from other lesions. Oncol Rep 2011; 26: 43-48 [PMID: 21573495 DOI: 10.3892/or.2011.1287]
- Aihara H, Saito S, Inomata H, Ide D, Tamai N, Ohya TR, Kato T, Amitani S, Tajiri H. Computer-45 aided diagnosis of neoplastic colorectal lesions using 'real-time' numerical color analysis during autofluorescence endoscopy. Eur J Gastroenterol Hepatol 2013; 25: 488-494 [PMID: 23249604 DOI: 10.1097/MEG.0b013e32835c6d9al
- Inomata H, Tamai N, Aihara H, Sumiyama K, Saito S, Kato T, Tajiri H. Efficacy of a novel autofluorescence imaging system with computer-assisted color analysis for assessment of colorectal lesions. World J Gastroenterol 2013; 19: 7146-7153 [PMID: 24222959 DOI: 10.3748/wig.v19.i41.7146]
- Horiuchi H, Tamai N, Kamba S, Inomata H, Ohya TR, Sumiyama K. Real-time computer-aided diagnosis of diminutive rectosigmoid polyps using an auto-fluorescence imaging system and novel color intensity analysis software. Scand J Gastroenterol 2019; 54: 800-805 [PMID: 31195905 DOI: 10.1080/00365521.2019.16274071
- Kudo SE, Mori Y, Wakamura K, Ikehara N, Ichimasa K, Wada Y, Kutsukawa M, Misawa M, Kudo T, Hayashi T, Miyachi H, Inoue H, Hamatani S. Endocytoscopy can provide additional diagnostic ability to magnifying chromoendoscopy for colorectal neoplasms. J Gastroenterol Hepatol 2014; 29: 83-90 [PMID: 23980563 DOI: 10.1111/jgh.12374]
- Brown SR, Baraza W, Din S, Riley S. Chromoscopy vs conventional endoscopy for the detection of 49 polyps in the colon and rectum. Cochrane Database Syst Rev 2016; 4: CD006439 [PMID: 27056645 DOI: 10.1002/14651858.CD006439.pub4]
- Kanao H, Tanaka S, Oka S, Kaneko I, Yoshida S, Arihiro K, Yoshihara M, Chayama K. Clinical significance of type V(I) pit pattern subclassification in determining the depth of invasion of colorectal neoplasms. World J Gastroenterol 2008; 14: 211-217 [PMID: 18186557 DOI: 10.3748/wjg.14.211]
- Takemura Y, Yoshida S, Tanaka S, Onji K, Oka S, Tamaki T, Kaneda K, Yoshihara M, Chayama K. Quantitative analysis and development of a computer-aided system for identification of regular pit patterns of colorectal lesions. Gastrointest Endosc 2010; 72: 1047-1051 [PMID: 21034905 DOI: 10.1016/j.gie.2010.07.037]
- Häfner M, Gangl A, Kwitt R, Uhl A, Vécsei A, Wrba F. Improving pit-pattern classification of endoscopy images by a combination of experts. Med Image Comput Comput Assist Interv 2009; 12: 247-254 [PMID: 20425994 DOI: 10.1007/978-3-642-04268-3 31]
- 53 Qi X, Pan Y, Hu Z, Kang W, Willis JE, Olowe K, Sivak MV Jr, Rollins AM. Automated quantification of colonic crypt morphology using integrated microscopy and optical coherence tomography. J Biomed Opt 2008; 13: 054055 [PMID: 19021435 DOI: 10.1117/1.2993323]
- Mori Y, Kudo SE, Wakamura K, Misawa M, Ogawa Y, Kutsukawa M, Kudo T, Hayashi T, Miyachi H, Ishida F, Inoue H. Novel computer-aided diagnostic system for colorectal lesions by using endocytoscopy (with videos). Gastrointest Endosc 2015; 81: 621-629 [PMID: 25440671 DOI: 10.1016/j.gie.2014.09.008]
- Takeda K, Kudo SE, Mori Y, Misawa M, Kudo T, Wakamura K, Katagiri A, Baba T, Hidaka E, Ishida F, Inoue H, Oda M, Mori K. Accuracy of diagnosing invasive colorectal cancer using computer-aided endocytoscopy. Endoscopy 2017; 49: 798-802 [PMID: 28472832 DOI: 10.1055/s-0043-105486]

- Kudo SE, Misawa M, Mori Y, Hotta K, Ohtsuka K, Ikematsu H, Saito Y, Takeda K, Nakamura H, Ichimasa K, Ishigaki T, Toyoshima N, Kudo T, Hayashi T, Wakamura K, Baba T, Ishida F, Inoue H, Itoh H, Oda M, Mori K. Artificial Intelligence-assisted System Improves Endoscopic Identification of Colorectal Neoplasms. Clin Gastroenterol Hepatol 2020; 18: 1874-1881.e2 [PMID: 31525512 DOI: 10.1016/j.cgh.2019.09.009]
- ASGE Technology Committee. Confocal laser endomicroscopy. Gastrointest Endosc 2014; 80: 928-57 938 [PMID: 25442092 DOI: 10.1016/j.gie.2014.06.021]
- André B, Vercauteren T, Buchner AM, Krishna M, Ayache N, Wallace MB. Software for automated classification of probe-based confocal laser endomicroscopy videos of colorectal polyps. World J Gastroenterol 2012; 18: 5560-5569 [PMID: 23112548 DOI: 10.3748/wjg.v18.i39.5560]
- Ștefănescu D, Streba C, Cârțână ET, Săftoiu A, Gruionu G, Gruionu LG. Computer Aided Diagnosis for Confocal Laser Endomicroscopy in Advanced Colorectal Adenocarcinoma. PLoS One 2016; 11: e0154863 [PMID: 27144985 DOI: 10.1371/journal.pone.0154863]
- Taunk P, Atkinson CD, Lichtenstein D, Rodriguez-Diaz E, Singh SK. Computer-assisted assessment of colonic polyp histopathology using probe-based confocal laser endomicroscopy. Int J Colorectal Dis 2019; **34**: 2043-2051 [PMID: 31696259 DOI: 10.1007/s00384-019-03406-y]
- Mori Y, Kudo SE, Misawa M, Mori K. Simultaneous detection and characterization of diminutive polyps with the use of artificial intelligence during colonoscopy. VideoGIE 2019; 4: 7-10 [PMID: 30623149 DOI: 10.1016/j.vgie.2018.10.0061
- Misawa M, Kudo SE, Mori Y, Cho T, Kataoka S, Yamauchi A, Ogawa Y, Maeda Y, Takeda K, Ichimasa K, Nakamura H, Yagawa Y, Toyoshima N, Ogata N, Kudo T, Hisayuki T, Hayashi T, Wakamura K, Baba T, Ishida F, Itoh H, Roth H, Oda M, Mori K. Artificial Intelligence-Assisted Polyp Detection for Colonoscopy: Initial Experience. Gastroenterology 2018; 154: 2027-2029.e3 [PMID: 29653147 DOI: 10.1053/j.gastro.2018.04.003]
- Eickhoff A, van Dam J, Jakobs R, Kudis V, Hartmann D, Damian U, Weickert U, Schilling D, Riemann JF. Computer-assisted colonoscopy (the NeoGuide Endoscopy System): results of the first human clinical trial ("PACE study"). Am J Gastroenterol 2007; 102: 261-266 [PMID: 17156149 DOI: 10.1111/i.1572-0241.2006.01002.xl
- Pullens HJ, van der Stap N, Rozeboom ED, Schwartz MP, van der Heijden F, van Oijen MG, Siersema PD, Broeders IA. Colonoscopy with robotic steering and automated lumen centralization: a feasibility study in a colon model. Endoscopy 2016; 48: 286-290 [PMID: 26126158 DOI: 10.1055/s-0034-13925501
- Slawinski PR, Taddese AZ, Musto KB, Sarker S, Valdastri P, Obstein KL. Autonomously Controlled Magnetic Flexible Endoscope for Colon Exploration. Gastroenterology 2018; 154: 1577-1579.e1 [PMID: 29530377 DOI: 10.1053/j.gastro.2018.02.037]
- Formosa GA, Prendergast JM, Edmundowicz SA, Rentschler ME. Novel Optimization-Based Design and Surgical Evaluation of a Treaded Robotic Capsule Colonoscope. IEEE Transact Robotics 2020; **36**: 545-552 [DOI: 10.1109/TRO.2019.2949466]
- Kahi CJ, Imperiale TF, Juliar BE, Rex DK. Effect of screening colonoscopy on colorectal cancer incidence and mortality. Clin Gastroenterol Hepatol 2009; 7: 770-5; quiz 711 [PMID: 19268269] DOI: 10.1016/j.cgh.2008.12.030]



Published by Baishideng Publishing Group Inc

7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: https://www.f6publishing.com/helpdesk

https://www.wjgnet.com

