

World Journal of *Gastroenterology*

World J Gastroenterol 2021 August 28; 27(32): 5297-5459



OPINION REVIEW

- 5297** Management of Flood syndrome: What can we do better?
Strainiene S, Peculyte M, Strainys T, Stundiene I, Savlan I, Liakina V, Valantinas J

REVIEW

- 5306** Radiomics and machine learning applications in rectal cancer: Current update and future perspectives
Stanzione A, Verde F, Romeo V, Boccadifuoco F, Mainenti PP, Maurea S
- 5322** Could the burden of pancreatic cancer originate in childhood?
Diaconescu S, Gilcă-Blanariu GE, Poamaneagra S, Marginean O, Paduraru G, Stefanescu G

MINIREVIEWS

- 5341** Application of artificial intelligence in preoperative imaging of hepatocellular carcinoma: Current status and future perspectives
Feng B, Ma XH, Wang S, Cai W, Liu XB, Zhao XM
- 5351** Artificial intelligence application in diagnostic gastrointestinal endoscopy - Deus ex machina?
Correia FP, Lourenço LC
- 5362** Faecal microbiota transplantation enhances efficacy of immune checkpoint inhibitors therapy against cancer
Kang YB, Cai Y
- 5376** Immune checkpoint inhibitor-related hepatotoxicity: A review
Remash D, Prince DS, McKenzie C, Strasser SI, Kao S, Liu K

ORIGINAL ARTICLE

Basic Study

- 5392** Therapeutic effect of *Cistanche deserticola* on defecation in senile constipation rat model through stem cell factor/C-kit signaling pathway
Zhang X, Zheng FJ, Zhang Z
- 5404** Recombinant angiopoietin-like protein 4 attenuates intestinal barrier structure and function injury after ischemia/reperfusion
Wang ZY, Lin JY, Feng YR, Liu DS, Zhao XZ, Li T, Li SY, Sun JC, Li SF, Jia WY, Jing HR

Retrospective Study

- 5424** Prolonged survival in patients with hand-foot skin reaction secondary to cooperative sorafenib treatment
Ochi M, Kamoshida T, Araki M, Ikegami T

- 5438** Contrast-enhanced ultrasound imaging for intestinal lymphoma

Cui NY, Gong XT, Tian YT, Wang Y, Zhang R, Liu MJ, Han J, Wang B, Yang D

Observational Study

- 5448** Intestinal ischemic manifestations of SARS-CoV-2: Results from the ABDOCOVID multicentre study

Norsa L, Bonaffini PA, Caldato M, Bonifacio C, Sonzogni A, Indriolo A, Valle C, Furfaro F, Bonanomi A, Franco PN, Gori M, Smania V, Scaramella L, Forzenigo L, Vecchi M, Solbiati M, Costantino G, Danese S, D'Antiga L, Sironi S, Elli L

ABOUT COVER

Editorial Board Member of *World Journal of Gastroenterology*, Luca Mastracci, MD, Associate Professor, Anatomic Pathology, Department of Surgical Sciences and Integrated Diagnostics (DISC), University of Genova, Ospedale Policlinico San Martino, Largo Rosanna Benzi 10, Genova 16132, Italy. luca.mastracci@unige.it

AIMS AND SCOPE

The primary aim of *World Journal of Gastroenterology* (WJG, *World J Gastroenterol*) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Jia-Hui Li; Production Department Director: Yu-Jie Ma; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Andrzej S Tarnawski, Subrata Ghosh

EDITORIAL BOARD MEMBERS

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

PUBLICATION DATE

August 28, 2021

COPYRIGHT

© 2021 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Basic Study

Therapeutic effect of *Cistanche deserticola* on defecation in senile constipation rat model through stem cell factor/C-kit signaling pathway

Xia Zhang, Fa-Juan Zheng, Zhen Zhang

ORCID number: Xia Zhang 0000-0002-1855-9940; Fa-Juan Zheng 0000-0002-3538-699X; Zhen Zhang 0000-0001-9948-9034.

Author contributions: Zhang Z designed the study; Zhang X and Zhang Z wrote the manuscript and were involved in revision, editing and review; Zhang X reformatted the manuscript; Zheng FJ was involved in the animal experiment process and data collation and input; All authors read and approved the final manuscript.

Supported by Natural Science Foundation of Chongqing, No. cstc2017jcyjAX0306.

Institutional animal care and use committee statement: The study was reviewed and approved by the Chongqing Hospital of Traditional Chinese Medicine Ethics Committee, No. Cstc2017jcyjAX0306.

Conflict-of-interest statement: There is no conflict of interest associated with any of the senior author or other coauthors contributed their efforts in this manuscript.

Data sharing statement: Technical appendix, statistical code, and

Xia Zhang, Fa-Juan Zheng, Department of Science and Education, Chongqing Hospital of Traditional Chinese Medicine, Chongqing 400021, China

Zhen Zhang, Department of Anorectal, Chongqing Hospital of Traditional Chinese Medicine, Chongqing 400021, China

Corresponding author: Zhen Zhang, MD, Deputy Director, Department of Anorectal, Chongqing Hospital of Traditional Chinese Medicine, No. 6 Seventh Branch of Panxi Road, Jiangbei District, Chongqing 400021, China. zhangzhen@cdutcm.edu.cn

Abstract

BACKGROUND

Constipation is one of the chronic gastrointestinal functional diseases. It seriously affects the quality of life. *Cistanche deserticola* (*C. deserticola*) can treat constipation obviously, but its mechanism has not been clarified. We supposed that mechanism of it improved the intestinal motility by stimulating interstitial Cajal cells (ICC). Activation of the C-kit receptor on the surface of ICC is closely related to ICC function, and the stem cell factor (SCF)/C-kit signaling pathways plays an important role on it. To investigate the mechanism of how *C. deserticola* treats constipation, this study aimed to establish a constipation model in rats and explore the role of SCF/C-kit signaling pathway in the treatment.

AIM

To explore the SCF/C-kit signaling pathways in the role of *C. deserticola* for treatment of constipation by a constipation rat model.

METHODS

Forty-eight 8-mo-old Sprague-Dawley rats were divided into 4 groups by random weight method: Normal group ($n = 12$), model group ($n = 12$), *C. deserticola* group ($n = 12$) and blocker group ($n = 12$). The normal group received normal saline by gavage; the model group received loperamide by gavage; the blocker group received loperamide and *C. deserticola* by gavage, and STI571 was injected by intraperitoneally. During treatment, the weight, fecal granules and fecal quality were recorded every 10 d. On day 20 after model induction, the colon tissues of each group were removed. Hematoxylin and eosin staining was used to observe pathological changes. Expression levels of SCF, C-kit and *Aquaporin* genes were

dataset available from the corresponding author at zhangzhen@cdutcm.edu.cn.

Participants gave informed consent for data sharing.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/License/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: China

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): 0
Grade C (Good): 0
Grade D (Fair): 0
Grade E (Poor): 0

Received: April 7, 2021

Peer-review started: April 7, 2021

First decision: May 27, 2021

Revised: June 3, 2021

Accepted: July 20, 2021

Article in press: July 20, 2021

Published online: August 28, 2021

P-Reviewer: Rangel-Corona R

S-Editor: Wu YXJ

L-Editor: Filipodia

P-Editor: Li JH



detected by immunohistochemistry, western blotting, and real-time-quantitative polymerase chain reaction. The colonic epithelial mitochondria and goblet cells were observed by transmission electron microscopy.

RESULTS

Compared with the normal group, as treatment progressed, the weight of rats in the model and blocker groups decreased significantly, the stool weight decreased, and the stool quality was dry ($P < 0.05$). *C. deserticola* reversed the decrease in body weight and stool weight and improved stool quality. Histopathological analysis indicated that the colonic mucosal epithelium in the model group was incomplete, and the arrangement of the glands was irregular or damaged. Treatment with *C. deserticola* improved the integrity and continuity of the epithelial cells and regular arrangement of the glands. The blocking agents inhibited the effects of *C. deserticola*. Immunohistochemistry and real-time-quantitative polymerase chain reaction showed that expression of SCF and C-kit protein or genes in the colonic tissue of the model group was decreased ($P < 0.05$), while treatment with *C. deserticola* increased protein or gene expression ($P < 0.05$). Western blotting showed that expression of aquaporin APQ3 was increased, while the expression of Cx43 decreased in the model group. Treatment with *C. deserticola* inhibited expression of APQ3 and promoted expression of Cx43. Transmission electron microscopy showed that the mitochondria of the colonic epithelium in the model group were swollen and arranged disorderly, and microvilli were sparse. The condition was better in the *C. deserticola* group. Mice treated with STI571 blocker confirmed that blocking the SCF/C-kit pathway inhibited the improvement of constipation by *C. deserticola*.

CONCLUSION

C. deserticola improved defecation in rats with constipation, and the SCF/C-kit signaling pathway, which is a key link of ICC function, played an important role of the treatment.

Key Words: *Cistanche deserticola*; Senile constipation; Stem cell factors; C-kit

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: We studied a possible mechanism of *Cistanche deserticola* (*C. deserticola*) in the treatment of constipation. The mechanism might improve colon motility through stem cell factor (SCF)/C-kit signaling pathway in colon Cajal stromal cells. Therefore, the constipation rat model was replicated, and then rats were treated by direct administration of *C. deserticola* and specific blocking of the SCF/C-kit signaling pathway. The defecation of rats, changes of colonic pathology and ultrastructure as well as the protein expression related to the SCF/C-kit signaling pathway were observed. Our results support that the SCF/C-kit signaling pathway plays an important mechanism on the therapeutic effect of *C. deserticola*.

Citation: Zhang X, Zheng FJ, Zhang Z. Therapeutic effect of *Cistanche deserticola* on defecation in senile constipation rat model through stem cell factor/C-kit signaling pathway. *World J Gastroenterol* 2021; 27(32): 5392-5403

URL: <https://www.wjgnet.com/1007-9327/full/v27/i32/5392.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v27.i32.5392>

INTRODUCTION

Senile constipation (SC) is one of the common chronic gastrointestinal functional diseases in geriatric patients, which induces or aggravates cerebrovascular events and other diseases and seriously affects the quality of life of the elderly population[1,2]. According to traditional Chinese medicine, the main etiology and pathogenesis of SC are gastrointestinal heat, deficiency of Yin and Jin and intestinal de-wetting[3,4], while

Cistanche deserticola (*C. deserticola*) can warm the kidneys and relax the bowels. Researchers have reported that the effect of intestinal motility improvement and treatment of SC is clear, but its mechanism has not been clarified[5,6]. Interstitial Cajal cells (ICCs) are a special type of mesenchymal cells in the gastrointestinal tract. Activation of the C-kit receptor on the surface of ICCs is closely related to cell proliferation, differentiation and function. ICCs participate in the pathogenesis of many gastrointestinal motor dysfunctions[7,8]. Activation of C-kit depends on the binding of its ligand stem cell factor (SCF)[9]. To investigate the effect and mechanism of action of *C. deserticola* on SC, this study aimed to establish an SC model in rats induced by loperidol to explore the role of the SCF/C-kit signaling pathway in the treatment of constipation by *C. deserticola*.

MATERIALS AND METHODS

Experimental materials

Experimental animals: The experimental animals were 48 male 8-mo-old Sprague-Dawley rats weighing 500-550 g; all purchased from Changsha Tianqin Biotechnology Co. Ltd. Rats were raised in the animal laboratory of Chongqing Weisiteng Biomedical Technology Co. Ltd., given normal day and night light, food and drink. All procedures complied with the management guidelines issued by the Ethics Committee of Chongqing Traditional Chinese Medicine Hospital.

Primary reagents: The primary reagents included loperamide (Sigma, St. Louis, MO, United States), *C. deserticola* from Hubei Jurui Traditional Chinese Medicine Decoction Co. Ltd., ST1571 from Chinese Selleck Co. Ltd., and SCF, C-kit, connexin 43 (Cx43), aquaporin 3 (AQP3) and β -actin antibodies (Abcam, Cambridge, MA, United States), PrimeScript II 1st Strand cDNA Synthesis Kit and Premix Taq™ (Ex Taq™ Version 2.0) (Takara Co. Ltd., Japan).

Methods

SC model preparation and grouping: To avoid the influence of confounding factors such as body weight on the experimental results, the rats were randomly grouped by the block random grouping method. Forty-eight rats were fed adaptively for 1 wk, and the rats were numbered in order of weight from light to heavy. According to the order of body weight, the rats were divided into 12 zones (the weight of rats in each zone was similar), with 4 rats in each zone. The 12 zones were divided into 4 groups of 12 (normal, model, *C. deserticola* and blocker). Rats in the model group were given loperamide (0.0625/100 g, 10 mL/kg) by gavage. The first gavage dose was doubled, and the drug was given at 10:00 every day, once a day for 20 consecutive days. The normal group was given the same amount of normal saline by gavage. In addition, the model group was treated with *C. deserticola* (0.156 g/100 g, 10 mL/kg) by gavage. The blocker group was given *C. deserticola* (0.156 g/100 g, 10 mL/kg) by gavage and ST1571 (25 mg/mL, 1 mL/kg) by intraperitoneal injection. During the experiment, body weight, stool weight and stool condition were recorded every 10 d. No accidental death occurred in any of the rats. Twenty days later, the rats were killed, and the colonic tissue was removed for examination and liquid nitrogen preservation. The remaining parts were fixed and embedded.

Histopathological analysis: The rats fasted overnight were anesthetized by intraperitoneal injection with 7% chloral hydrate (5 mL/kg) on the next day. The abdomen was opened to separate the colonic tissue, and 8-10 mm of colon was removed. The intestinal contents were removed by gently shaking in precooled polybutylene succinate and fixed for 24 h in 4% paraformaldehyde fixative solution. Conventional paraffin embedding was performed. Tissue sections (5 μ m) were removed for hematoxylin and eosin staining, and the histopathological morphology was observed under a microscope.

Detection of expression of SCF and C-kit by immunohistochemical staining: The paraffin tissues examined by histopathology above were sectioned, rehydrated with sectioned xylene dewaxed gradient ethanol, antigen remedied by heat and sealed for 1 h with goat serum at room temperature. SCF and C-kit primary antibodies were added dropwise, and tissues were kept overnight at 4 °C. After rinsing with polybutylene succinate, horseradish peroxidase was dropped to label secondary antibody, rinsed with polybutylene succinate for 5 min, drop 2,4-diaminobutyric acid chromogen solution, re-dye with hematoxylin, reverse gradient alcohol dehydration, transpar-

entized by xylene and neutral resin seal. Microscopically, the nuclei were purplish blue, and the positive result products were brownish yellow or yellow particles.

Detection of expression of *C-kit* mRNA in colonic tissue by real-time-quantitative polymerase chain reaction (RT-qPCR): Frozen rat colonic tissue was ground by homogenizer, and 1 mL RNAiso Plus was added. Total RNA of colonic tissue was extracted by chloroform/isopropanol method, and the RNA quality was detected by gel electrophoresis. After identified by Bio-Rad gel imager, 1 µg RNA was reverse transcribed into cDNA by Takara PrimeScript II 1st Strand cDNA Synthesis Kit. According to NCBI database Gene ID: 64030, the CDS region of the *C-kit* gene was found and combined with NCBI Primer-Blast design Primer sequence, which was synthesized by Sangon Biotech Co. Ltd, *C-kit* primer sequence sense strand 5'-AGGTGTACCGTTCCTGTCCC-3', antisense strand 5'-GGCTGGATTGCTCTTTGCTGT-3', β -actin sense strand 5'-CTTCCTTCCTGGGTATGGAATC-3', antisense strand 5'-CTGTGTTGGCATAGAGGTCTT-3'. PCR amplification was performed on a Bio-Rad CFX90 Real-Time PCR instrument. The conditions for PCR assay were initial denaturation at 95 °C for 5 min, 30 cycles of 95 °C for 30 s, 60 °C for 30 s and 72 °C for 1 min followed by final extension at 72 °C for 10 min. $2^{-\Delta\Delta Ct}$ ($\Delta Ct = Ct \text{ value} - \beta\text{-actin } Ct \text{ value}$) was taken to analyze relative gene expression.

Detection of protein expression level of Cx43 and AQP3 in colonic tissue by Western blotting: About 1 cm of frozen colonic tissue was collected and lysed with RIPA strong pyrolysis liquid (100 µL). The tissue was homogenized with a glass homogenizer 20 times and transferred to a 1.5 mL centrifuge tube. The supernatant was extracted after centrifugation at 12000 rpm at 4 °C for 5 min, and the concentration of total protein in the supernatant was determined by BCA method. We removed 25 µg denatured protein to perform sodium dodecyl sulfate-polyacrylamide gel electrophoresis and then transferred the proteins in the gel to polyvinylidene fluoride membrane at a constant current of 200 mA then sealed by 5% skim milk confining liquid for 2 h at room temperature. Corresponding primary antibodies were diluted with sealing fluid, incubated at 4 °C overnight, then washed the membrane resistance and added second antibodies. After adding ECL fluid, they were put under the exposure meter for test by optical density analysis.

Transmission electron microscopy: We took about 1 cm of proximal colonic tissue, fixed it in 2.5% glutaraldehyde for 24 h, dehydrated it with graded acetone, embedded it in Epon812 and cut 50 nm ultrathin sections on an ultramicrotome. The sections were stained with uranium dioxide acetate and lead citrate dyeing liquid at room temperature for 10 min. We observed colonic villi, epithelial cell mitochondria and goblet cells under a Philips Tecnai-10 transmission electron microscope.

Statistical analysis

SPSS 19.0 software was used for data analysis, and GraphPad Prism8.4 software was used for plotting the results. The Shapiro-Wilk's test was used for normality testing. Data with normal distribution were represented by mean \pm SD. Error items were used to represent the mean \pm SD of hemoglobin after treatment. Student's *t*-test was used for comparison between two groups. Analysis of variance was used for comparison between multiple groups. The Student-Newman-Keuls method was used for comparison between multiple groups in pairs, and Dunnett's test was used to measure the differences between each experimental group and control group in sequence. Associated data were measured by multiple time point, and repeated measurement data analysis of variance was conducted. If the sphericity test showed $P > 0.05$, repeated measurement data analysis of variance was used. If $P < 0.05$, the generalized estimation equation was taken. Data that did not conform to a normal distribution were represented by M (P25, P75), and the Kruskal-Wallis *H* rank sum test was used for comparison between groups. The χ^2 test was used to compare between groups when the data was unordered data, and Pearson's χ^2 test was used if the proportion of cells with theoretical frequency < 5 was $< 20\%$. If the proportion of cells with theoretical frequency < 5 was $\geq 20\%$, Fisher's exact test was used. $P < 0.05$ was used to determine that the difference was statistically significant.

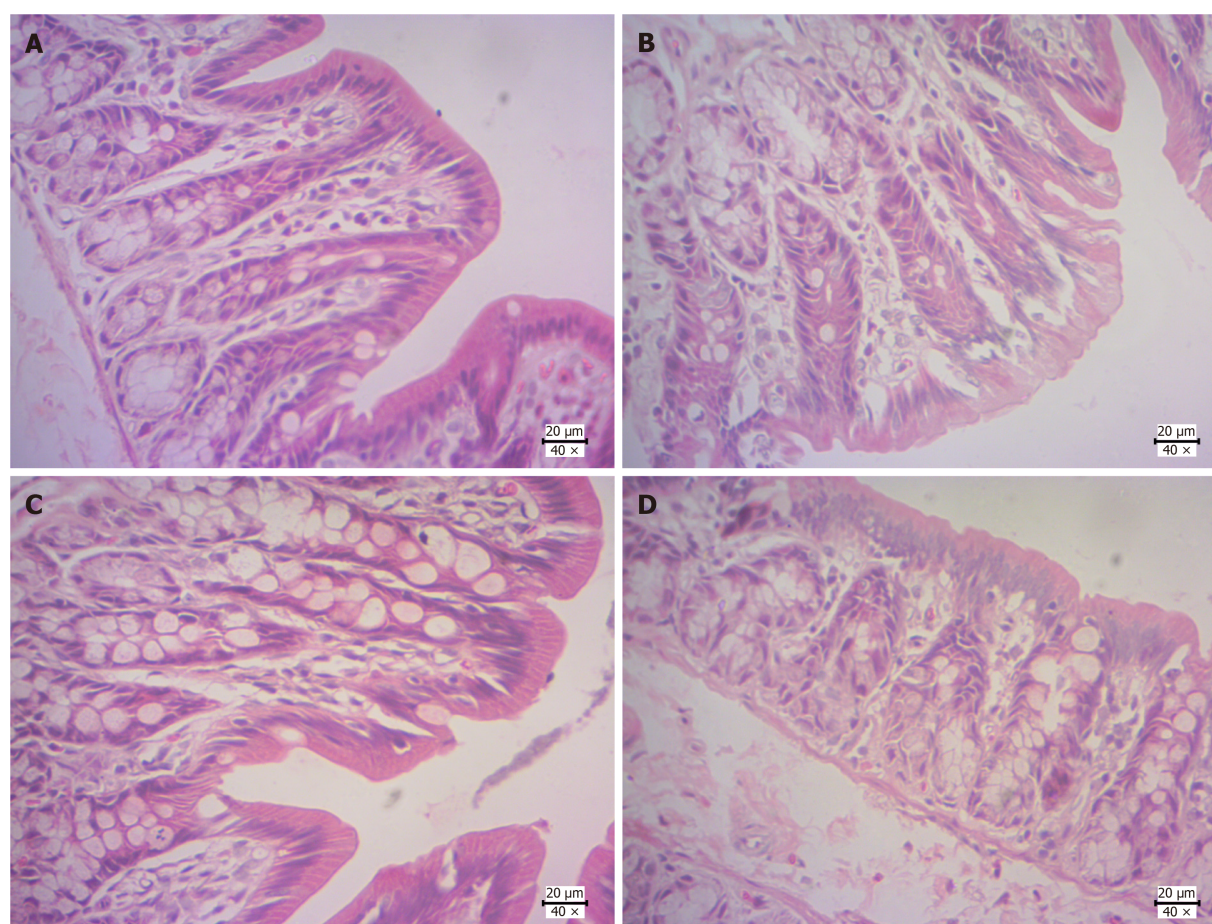


Figure 1 Colonic histopathological changes. A: Normal group; B: Model group; C: *Cistanche deserticola* group; D: Blocker group. Magnification: $\times 40$; Scale bar: 20 μm .

RESULTS

General conditions

One day after gavage of loperamide compared with the normal group, the body weight and stool weight in the model group were significantly decreased ($P < 0.05$), but there was no significant difference in stool quality score ($P > 0.05$). *C. deserticola* treatment in the model group slowed the decrease in body weight and stool weight ($P < 0.05$), but the body weight in the blocker group was not significantly reduced. After 10 d of treatment, compared with the normal group, the body weight and stool volume of rats in the model group were further significantly reduced, and the stools were dry and almost contained no water ($P < 0.05$). The body weight loss of rats was also significantly reduced by treatment with *C. deserticola* ($P < 0.05$). The body weight of rats in the blocker group was not significantly different from that in the model group ($P > 0.05$). On day 20 before sampling, the body weight of rats in the model group was significantly lower than that in the normal group, and the stool quality score was significantly increased ($P < 0.05$), while the weight loss caused by constipation was inhibited by *C. deserticola* treatment, and the stool quality score was decreased ($P < 0.05$).

Colonic histopathology

Hematoxylin and eosin staining of colonic tissue of rats is shown in Figure 1. In the normal group, intestinal villi were intact, intestinal mucosa was intact and continuous, cells and glands were arranged regularly. Mucosa and submucosa were not infiltrated by inflammatory cells, and tissue structure was normal. In the model group, intestinal villi and mucosa were largely destroyed, local villi were shed and loosely arranged, and part of the glandular structure disappeared. In the *C. deserticola* group, histopathological staining was less than that of the model group, but there was necrosis in some goblet cells. After intraperitoneal injection in the blocker group, compared with the *C. deserticola* group, there was still local loss of intestinal villi, and there was fracture and

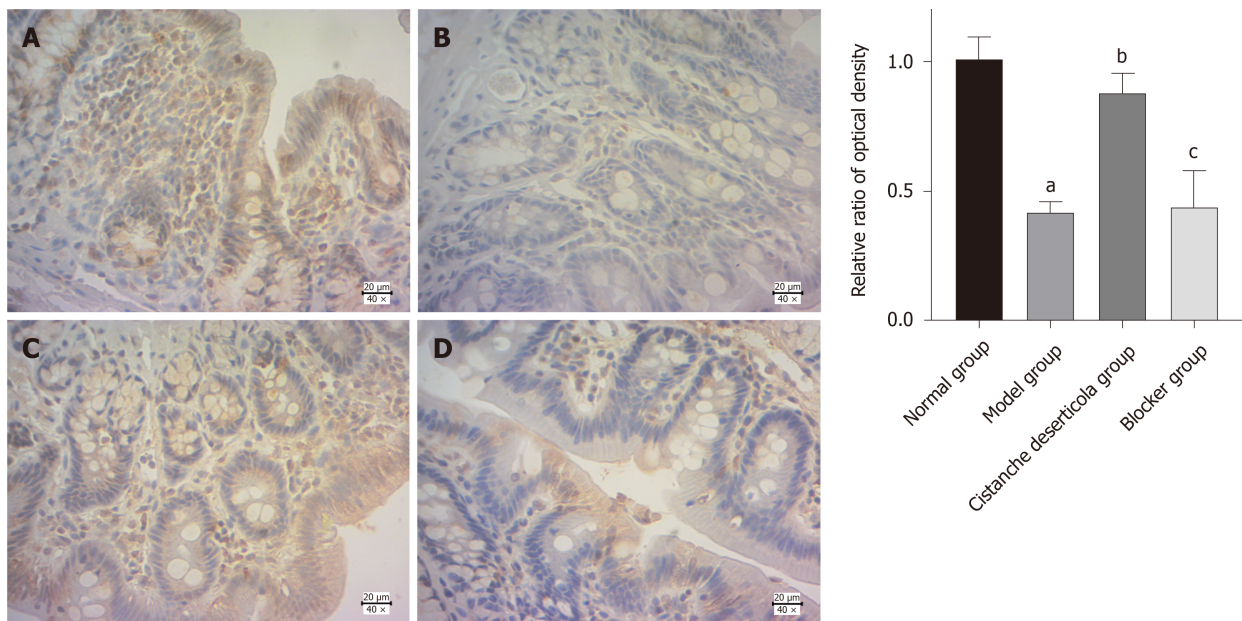


Figure 2 Expression of stem cell factor and relative optical density ratio in colonic tissue by immunohistochemical detection. A: Normal group; B: Model group; C: *Cistanche deserticola* group; D: Blocker group. Magnification: $\times 40$; Scale bar: 20 μm . ^a $P < 0.05$, the expressions of stem cell factor and C-kit in colon tissue of the model group were both significantly decreased compared with the normal group; ^b $P < 0.05$, the expression of stem cell factor and C-kit in the colon tissue of rats in the *Cistanche deserticola* group was significantly increased compared with model group. There was no difference in the expression of stem cell factor and C-kit in the colon tissue of rats between the blocker group and the model group.

loss of intestinal mucosa.

Detection of expression of stem cell factor and C-kit protein by immunohistochemical staining

Immunohistochemical staining showed that SCF and C-kit were expressed in the colon of the control group. Compared with the control group, expression of SCF and C-kit in the model group was significantly decreased ($P < 0.05$). Compared with the model group, expression of SCF and C-kit in the *C. deserticola* group was significantly increased ($P < 0.05$). There was no difference in expression of SCF and C-kit between the blocker and model groups ($P < 0.05$, Figures 2 and 3).

RT-qPCR

RT-qPCR showed that expression of *C-kit* mRNA in the model group was significantly lower than in the normal group (Figure 4, $P < 0.05$). Compared with the model group, *C-kit* in the *C. deserticola* group was significantly increased ($P < 0.05$). There was no significant difference in *C-kit* mRNA level between the model and blocker groups ($P > 0.05$). These results indicated that *C-kit* expression adjusted by *C. deserticola* participated in the improvement of constipation, and blockers inhibited the efficacy of *C. deserticola*.

Western blotting

Western blotting showed that AQP3 protein was expressed at a low level in the normal control group, while it was significantly increased in the model group compared with the control group (Figure 5) ($P < 0.05$). Expression of AQP3 protein was decreased after treatment with *C. deserticola*, and the inhibitory effect of *C. deserticola* on the expression of AQP3 was inhibited by blocker treatment ($P < 0.05$). Expression of Cx43 was the opposite of that of AQP3. These results suggested that the pharmacology effect of *C. deserticola* on rats with SC was achieved by promoting expression of Cx43 and inhibiting expression of AQP3 (Figures 5-7).

Transmission electron microscopy

After loperamide induction, mitochondrial swelling and autophagy occurred in some of the colonic mucosal epithelial cells in the model group. The number and length of intestinal microvilli decreased, and goblet cell pyknosis occurred (Figure 8). After treatment with *C. deserticola*, the number of intestinal microvilli increased. The microvilli were arranged neatly, and the structure was complete. The goblet cells

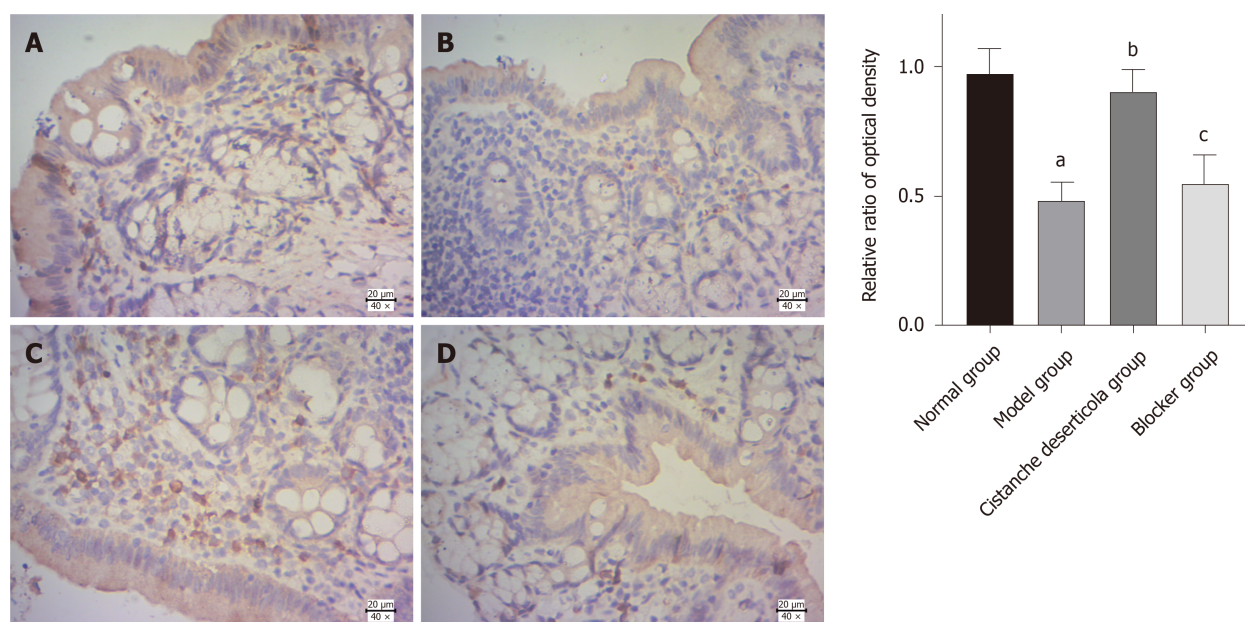


Figure 3 Expression of C-kit and relative optical density ratio in colonic tissue by immunohistochemical detection. A: Normal group; B: Model group; C: *Cistanche deserticola* group; D: Blocker group. Magnification: $\times 40$; Scale bar: 20 μm . ^a $P < 0.05$, compared with normal group; ^b $P < 0.05$, compared with model group; ^c $P > 0.05$, compared with model group.

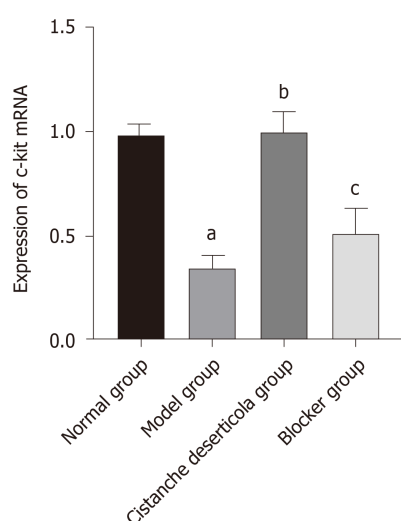


Figure 4 Expression of C-kit mRNA in colon tissue of rats in each group. ^a $P < 0.05$, compared with normal group; ^b $P < 0.05$, compared with model group; ^c $P > 0.05$, compared with model group.

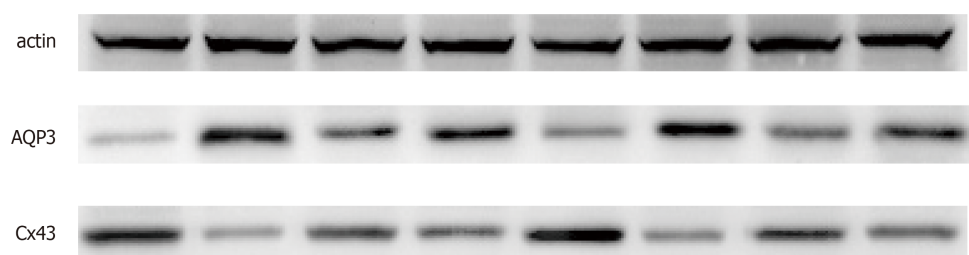


Figure 5 Connexin 43 and aquaporin 3 protein expression detected by western blotting. AQP3: Aquaporin 3; Cx43: Connexin 43.

regained some structural integrity but still did not return to normal length. After inhibitor treatment, the number of microvilli in colonic epithelial cells decreased significantly compared with those in the *C. deserticola* group, with loose arrangement,

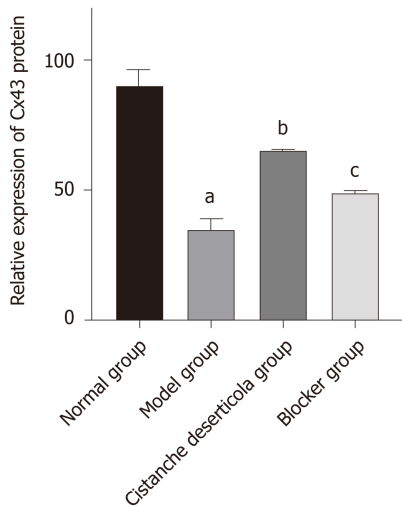


Figure 6 Relative expression of connexin 43 protein. ^a $P < 0.05$, compared with normal group; ^b $P < 0.05$, compared with model group; ^c $P > 0.05$, compared with model group. Cx43: Connexin 43.

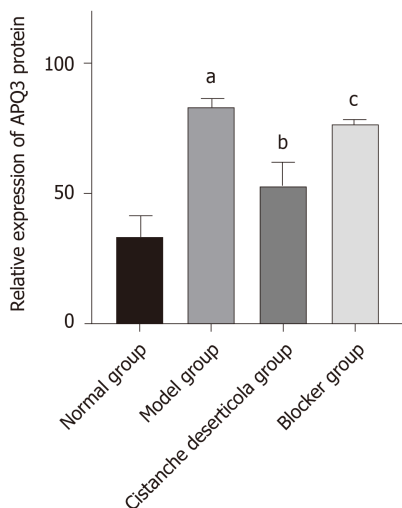


Figure 7 Relative expression of aquaporin 3 protein. ^a $P < 0.05$, compared with normal group; ^b $P < 0.05$, compared with model group; ^c $P > 0.05$, compared with model group. AQP3: Aquaporin 3.

shorter length, irregular morphology of mitochondria and severe pyknosis of goblet cells.

DISCUSSION

Constipation is common in the elderly population and is one of the common symptoms that affect the health status and quality of life of elderly people[10,11]. In terms of traditional Chinese medicine, Bai *et al*[12] believed that the basic pathogenesis of SC was dysfunction of large intestine conduction function, senile body failure, gradual loss of liver and kidney functions, lack of qi and blood and loss of body fluid, which led to dry stools, resulting in constipation[13]. SC puts pressure on patients and their families in many aspects, and it is important to explore its pathogenesis.

C. deserticola has the functions of tonifying kidney Yang, benefiting blood and regulating body immunity[14]. *C. deserticola* and Chinese herbal decoction containing *C. deserticola* have been widely used in the treatment of constipation, and the clinical effect is remarkable. Du *et al*[15] found in a rat model of Yang deficiency and constipation that *C. deserticola* improved the contraction amplitude of isolated colon, strengthened the contractility of the intestinal tract and returned the level of gastrointestinal hormones to normal. In addition, dietary fiber of *C. deserticola*, a

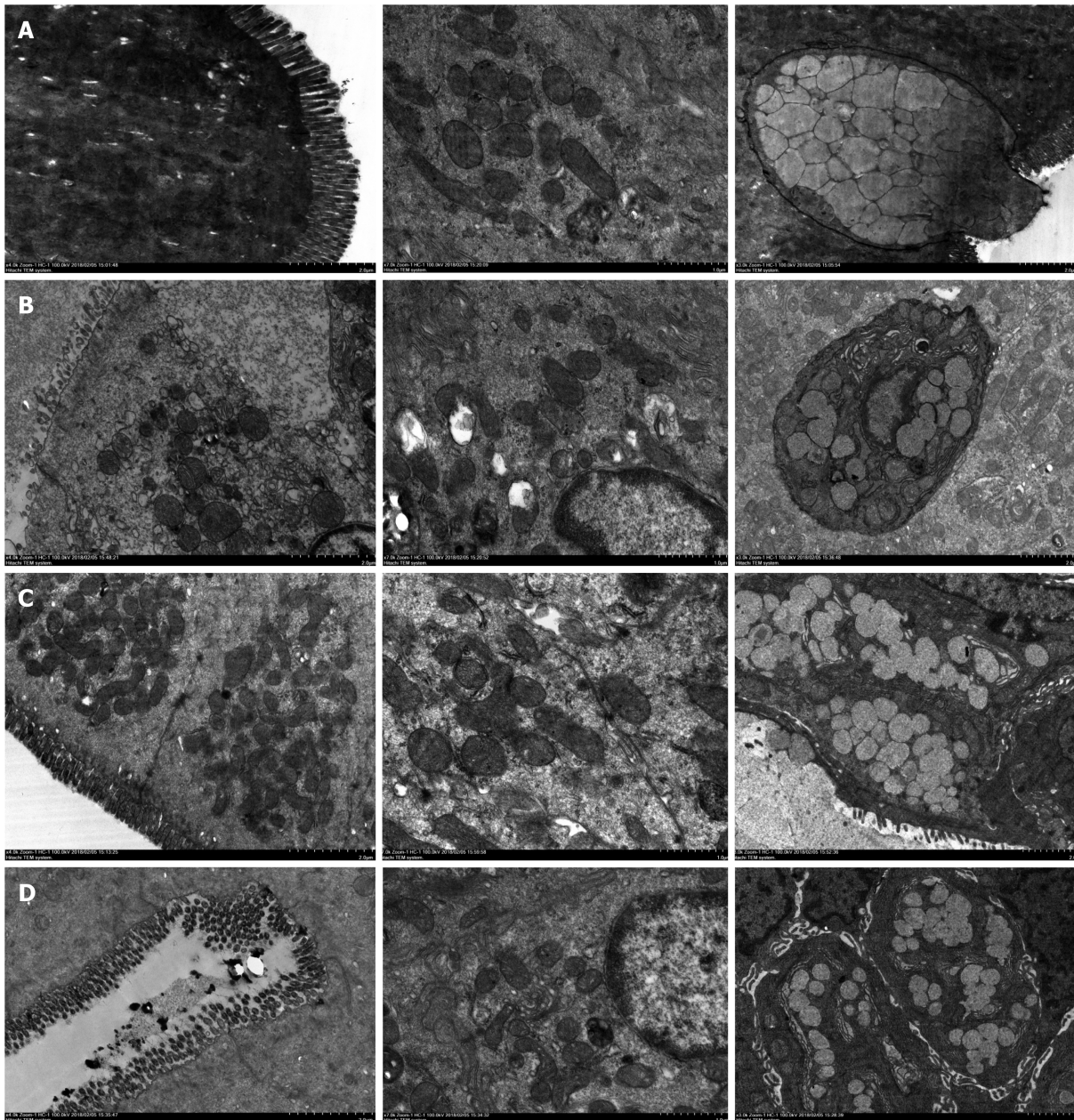


Figure 8 Observation of colonic villi, mitochondria of colonic epithelial cells and goblet cells by transmission electron microscopy. A: Normal group; B: Model group; C: *Cistanche deserticola* group; D: Blocker group.

processing byproduct of *C. deserticola*, improves the water-holding capacity of feces and has the advantage of low dose and good effect in nourishing intestines and improving defecation[16]. Our study found that *C. deserticola* treatment improved fecal moisture and colonic histopathology of rats with constipation.

ICCs are found throughout the intestinal tract and play a vital role in the neural control of intestinal movement as pacemakers and intestinal nerve transmission[17]. Proto-oncogene coding receptor tyrosine kinase, C-kit, is expressed by ICCs, and SCF is a ligand of C-kit[18]. The SCF/C-kit system is crucial in the development of ICCs. The SCF/C-kit signaling pathway has been analyzed during treatment of slow transfer constipation in rats by acupuncture[19]. Electroacupuncture at Tianshu point increased the number of colon ICCs in ACK2-treated mice, upregulated expression of SCF and C-kit proteins and reversed the phenotype of ICC. Prescription for invigorating qi and spleen can increase expression of ICC in colonic tissue and upregulate expression of C-kit and SCF mRNA, thus improving constipation symptoms[20]. In our study, immunohistochemistry showed that *C. deserticola* significantly upregulated expression of SCF and C-kit protein in a rat model of SC, and RT-qPCR showed that *C. deserticola* significantly upregulated expression of C-kit mRNA. C-kit blocker inhibited the upregulation of SCF and C-kit expression induced

by *C. deserticola*.

Fecal water content and colonic water transport are closely related to AQPs, which play an important role in intestinal water metabolism[21]. In a study of rats with slow transport constipation, Defecation decoction significantly improved constipation symptoms by regulating intestinal water absorption through downregulating AQP3 and AQP4[22]. In a rat model of functional constipation, the treatment mechanism of Oral Xiaofu Tongjie Fluid for functional constipation may be realized by regulating expression of vasoactive intestinal polypeptide and AQP3 in the colon[23]. Gap junctions play an essential role in mediating synchronous contraction of smooth muscle cells. Cx43 is the most important protein during that process[24]. Loss of Cx43 expression may be partially responsible for smooth muscle motor dysfunction[25]. Repair of the intestinal nervous system network structure in rats with slow transit constipation can improve the expression of Cx43 etc. and constipation symptoms[26]. These results indicate that the increased expression of AQP3 decreases fecal water content, leading to constipation, which may be an important mechanism of constipation. Expression of Cx43 may be related to contraction of intestinal smooth muscle and reduce the occurrence of constipation. In our study, western blotting showed that Cx43 expression was increased and AQP3 expression was inhibited by *C. deserticola* treatment, which resulted in increased smooth muscle contraction and decreased water absorption, and blocker treatment reversed these effects.

CONCLUSION

In conclusion, *C. deserticola* can inhibit expression of AQP3 and promote expression of Cx43 through the SCF/C-kit pathway, thereby improving the constipation induced by loperamine and reducing colonic tissue damage in aged rats. This study provided a good theoretical basis for clinical use of *C. deserticola*.

ARTICLE HIGHLIGHTS

Research background

Chronic constipation is a common functional gastrointestinal disease that seriously affects the quality of life, especially for senile patients. *Cistanche deserticola* (*C. deserticola*) is one kind of herb that can improve constipation obviously, but the mechanism of it is unclear. Since it increases the frequency of defecation, we suppose that its therapeutic effect is due to increased intestinal motility by an important signaling pathway, stem cell factor (SCF)/C-kit, located on the surface of interstitial Cajal cells.

Research motivation

The treatment of chronic constipation is not encouraging, and the available drugs cannot meet the clinical needs. New drugs are needed to safely increase intestinal motility and improve symptoms. New drugs should be based on in-depth studies of the mechanisms of some foods that are currently widely and safely used. *C. deserticola* is one kind of herb that has been used for thousands of years in Traditional Chinese Medicine for constipation. Benefits would be evident from more studies about the treatment mechanisms.

Research objectives

To investigate the mechanism of how *C. deserticola* treats constipation, this study aimed to establish a constipation model in rats and explore the role of the SCF/C-kit signaling pathway in the treatment.

Research methods

In the case of blank control group, a constipation rat model was first established. While these rats were treated with *C. deserticola*, a group of rats were specifically blocked from the target signaling pathway. The symptoms and defecation of these different groups of rats were observed, and the tissue and gene expression in which the target signaling pathway was located were observed to explain whether the SCF/C-kit signaling pathway plays a key role in the therapeutic effect of *C. deserticola*.

Research results

The model was successfully established, and the therapeutic effect of *C. deserticola* was also obvious. In the group where the target signaling pathway was blocked, the therapeutic effect of *C. deserticola* was significantly reduced, as reflected by histological and immunohistochemical changes as well as signal-pathway-related genes and proteins such as connexin 43, aquaporin 3 expression changes. *C. deserticola* can inhibit expression of aquaporin 3 and promote expression of connexin 43 through the SCF/C-kit pathway, thereby improving the constipation induced by loperamine and reducing colonic tissue damage in aged rats.

Research conclusions

This study provided a good theoretical basis for clinical use of *C. deserticola*. Furthermore, the SCF/C-kit signal pathway plays an important role of constipation treatment of *C. deserticola*. There is likely more mechanisms related to it, for the effect of *C. deserticola* was not blocked completely.

Research perspectives

The improvement of intestinal motility is the core point in the treatment of constipation. However, further research on intestinal dynamics is still needed. The research on intestinal dynamics of interstitial Cajal cells is still a focus of attention, but whether there is an unknown mechanism of its function is one of the directions of future research. In the meantime, herbs should also be more widely valued.

REFERENCES

- 1 Li XF. Effect evaluation of lactulose and bifidobacterium tetragalus in the treatment of elderly functional constipation. *Zhongguo Yaowu Yu Linchuang* 2019; **19**: 1294-1295
- 2 Liu X, Wang WH, Xia P, Cai FW. Clinical study of maren pill combined with prucalride in the treatment of elderly chronic constipation. *Xiandai Yaowu Yu Linchuang* 2019; **34**: 3329-3332
- 3 Zhang HL, Xia Y. Observation on the curative effect of Recipes for Nourishing Yin and Moistening Intestines on senile constipation. *Shiyong Zhongyiyiao Zazhi* 2019; **35**: 405-406
- 4 Ye SR. Experience in the treatment of senile constipation. *Zhongguo Zhongyiyiao Xinxu Zazhi* 2003; **S1**
- 5 Liu CY, Hu M. Clinical observation on 80 cases of elderly patients with intractable constipation treated by Cistanche Tongbian Decoction. *Hebei Zhongyi* 2010; **32**: 44-45
- 6 Tian PP. Clinical study on treatment of constipation in Parkinson's disease with Cistanche granules. Nanjing: Nanjing Zhongyiyao Daxue Xuebao 2016
- 7 Lorincz A, Redelman D, Horváth VJ, Bardsley MR, Chen H, Ordög T. Progenitors of interstitial cells of cajal in the postnatal murine stomach. *Gastroenterology* 2008; **134**: 1083-1093 [PMID: 18395089 DOI: 10.1053/j.gastro.2008.01.036]
- 8 Feng J, Gao J, Zhou S, Liu Y, Zhong Y, Shu Y, Meng MS, Yan J, Sun D, Fang Q. Role of stem cell factor in the regulation of ICC proliferation and detrusor contraction in rats with an underactive bladder. *Mol Med Rep* 2017; **16**: 1516-1522 [PMID: 28627603 DOI: 10.3892/mmr.2017.6749]
- 9 Yin J, Liang Y, Wang D, Yan Z, Yin H, Wu D, Su Q. Naringenin induces laxative effects by upregulating the expression levels of c-Kit and SCF, as well as those of aquaporin 3 in mice with loperamide-induced constipation. *Int J Mol Med* 2018; **41**: 649-658 [PMID: 29207043 DOI: 10.3892/ijmm.2017.3301]
- 10 Cai YQ, Wang HJ, Zhang X, Wang YH, Gu ZW, Wang HX, Liu Y. Investigation on the prevalence of constipation and its relationship with sub-health symptoms in the elderly in Nanjing city. *Zhonghua Laonian Yixue Zazhi* 2004; **23**: 267-269
- 11 Mugie SM, Benninga MA, Di Lorenzo C. Epidemiology of constipation in children and adults: a systematic review. *Best Pract Res Clin Gastroenterol* 2011; **25**: 3-18 [PMID: 21382575 DOI: 10.1016/j.bpg.2010.12.010]
- 12 Bai L, Wang CJ. Professor Wang Chuijie's experience in the treatment of senile constipation. *Jilin Zhongyiyao* 2008; **28**: 167-168
- 13 Bao GQ, Huang L. Treatment analysis of 173 cases of senile constipation by syndrome differentiation. *Heilongjiang Zhongyiyao* 2000; **5**: 22-22
- 14 Ding XH, Zhang J, Wang YC. Observation on the curative effect of Cistanche defaecation oral liquid on senile constipation. *Zhongguo Wuzhenxue Zazhi* 2009; **31**: 7609-7610
- 15 Du Q, Wu Z. Study on the dose-effect relationship and mechanism of the laxative effect of Cistanche deserticola based on the model of Yang deficiency and constipation. *Zhongnan Yaowu* 2016; **1**: 23-27
- 16 Wang LW, Sun J, Zhao B, Zhao MX. Study on the function of Cistanche deserticola dietary fiber to nourish intestines and defecate. *Shipin Anquan Zhiliang Jiance Xuebao* 2016; **7**: 3740-3744
- 17 Iino S, Ward SM, Sanders KM. Interstitial cells of Cajal are functionally innervated by excitatory motor neurones in the murine intestine. *J Physiol* 2004; **556**: 521-530 [PMID: 14754997 DOI: 10.1046/j.1365-2214.2004.01010.x]

- 10.1113/jphysiol.2003.058792]
- 18 **Torihashi S**, Ward SM, Nishikawa S, Nishi K, Kobayashi S, Sanders KM. c-kit-dependent development of interstitial cells and electrical activity in the murine gastrointestinal tract. *Cell Tissue Res* 1995; **280**: 97-111 [PMID: [7538451](#) DOI: [10.1007/BF00304515](#)]
 - 19 **HX**. Discussions on regulation of acupuncture on SCF/c-kit signaling pathway and mechanism of treatment of slow transverse constipation. *Nanjing: Nanjing University of Traditional Chinese Medicine* 2012
 - 20 **Wang JM**, Li M, Tang R, Wang PS, Xu LJ, Zhang R, Han Y. Effects of Recipe for invigorating qi, invigorating spleen and relieving constipation on ICC and SCF/ C-KIT signal pathway in colonic tissue of rats with slow transmission constipation. *Zhonghua Zhongyiyao Xuekan* 2019; **37**: 156-160+264
 - 21 **Ding YR**, Zheng PY, Li FG, Mei L, Huang H, Bai LM, Liu SM. Effects of Lactitol and Bifidobacterium infantis on AQP3 and ICC in rats with constipation. *Zhonghua Weishengwuxue He Miyanixue Zazhi* 2015; **35**: 890-895
 - 22 **Ji TL**. Effect of Defaecation Decoction on AQP3 and AQP4 in rats with slow transit constipation. *Nanjing: Nanjing University of Traditionl Chinese Medicine* 2017
 - 23 **Wang YJ**, Zhou YX, Zhang H, Yan SG, Xie P, Man SY, Li S. Effects of N Oral Xiaofu Tongjie Decoction on the Expression of VIP and AQP3 in Colonic Tissue of Rats with Functional Constipation. *Zhongguo Shiyan Fangjixue Zazhi* 2015; **9**: 116-119
 - 24 **Yu W**, Zeidel ML, Hill WG. Cellular expression profile for interstitial cells of cajal in bladder-a cell often misidentified as myocyte or myofibroblast. *PLoS One* 2012; **7**: e48897 [PMID: [23145014](#) DOI: [10.1371/journal.pone.0048897](#)]
 - 25 **Nemeth L**, Maddur S, Puri P. Immunolocalization of the gap junction protein Connexin43 in the interstitial cells of Cajal in the normal and Hirschsprung's disease bowel. *J Pediatr Surg* 2000; **35**: 823-828 [PMID: [10873019](#) DOI: [10.1053/jpsu.2000.6851](#)]
 - 26 **Kong JY**. Experimental study on the effect of Oral Xiaofu Tongjie Fluid on the repair of intestinal nervous system network structure in rats with slow transit constipation. *Chengdu Zongyiyao Daxue Xuebao* 2019



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

