



Observational Study

Determination of esophageal squamous cell carcinoma and gastric adenocarcinoma on raw tissue using Raman spectroscopy

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Abstract

BACKGROUND

Cancer detection is a global research focus, and novel, rapid, and label-free techniques are being developed for routine clinical practice. This has led to the development of new tools and techniques from the bench side to routine clinical practice. In this study, we present a method that uses Raman spectroscopy (RS) to detect cancer in unstained formalin-fixed, resected specimens of the esophagus and stomach. Our method can record a clear Raman-scattered light spectrum in these specimens, confirming that the Raman-scattered light spectrum changes because of the histological differences in the mucosal tissue.

AIM

To evaluate the use of Raman-scattered light spectrum for detecting endoscop-

ically resected specimens of esophageal squamous cell carcinoma (SCC) and gastric adenocarcinoma (AC).

METHODS

We created a Raman device that is suitable for observing living tissues, and attempted to acquire Raman-scattered light spectra in endoscopically resected specimens of six esophageal tissues and 12 gastric tissues. We evaluated formalin-fixed tissues using this technique and captured shifts at multiple locations based on feasibility, ranging from six to 19 locations 200 microns apart in the vertical and horizontal directions. Furthermore, a correlation between the obtained Raman scattered light spectra and histopathological diagnosis was performed.

RESULTS

We successfully obtained Raman scattered light spectra from all six esophageal and 12 gastric specimens. After data capture, the tissue specimens were sent for histopathological analysis for further processing because RS is a label-free methodology that does not cause tissue destruction or alterations. Based on data analysis of molecular-level substrates, we established cut-off values for the diagnosis of esophageal SCC and gastric AC. By analyzing specific Raman shifts, we developed an algorithm to identify the range of esophageal SCC and gastric AC with an accuracy close to that of histopathological diagnoses.

CONCLUSION

Our technique provides qualitative information for real-time morphological diagnosis. However, further *in vivo* evaluations require an excitation light source with low human toxicity and large amounts of data for validation.

Key Words: Raman spectroscopy; Squamous cell carcinoma; Adenocarcinoma; Esophagus; Stomach; Label-free cancer detection; Real-time diagnosis

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Core Tip: Cancer diagnosis is a critical step in patient management, and involves a combination of diagnostic modalities or a single investigation. Diagnostic techniques that provide comprehensive data on the disease process can be particularly valuable, and Raman spectroscopy (RS) is one such modality that offers detailed molecular-level information. In this study, we utilized RS to rapidly detect cancer in resected esophageal and stomach specimens, providing information beyond morphology. By providing detailed molecular-level data, RS can provide a more comprehensive understanding of disease processes and aid in accurate diagnosis.

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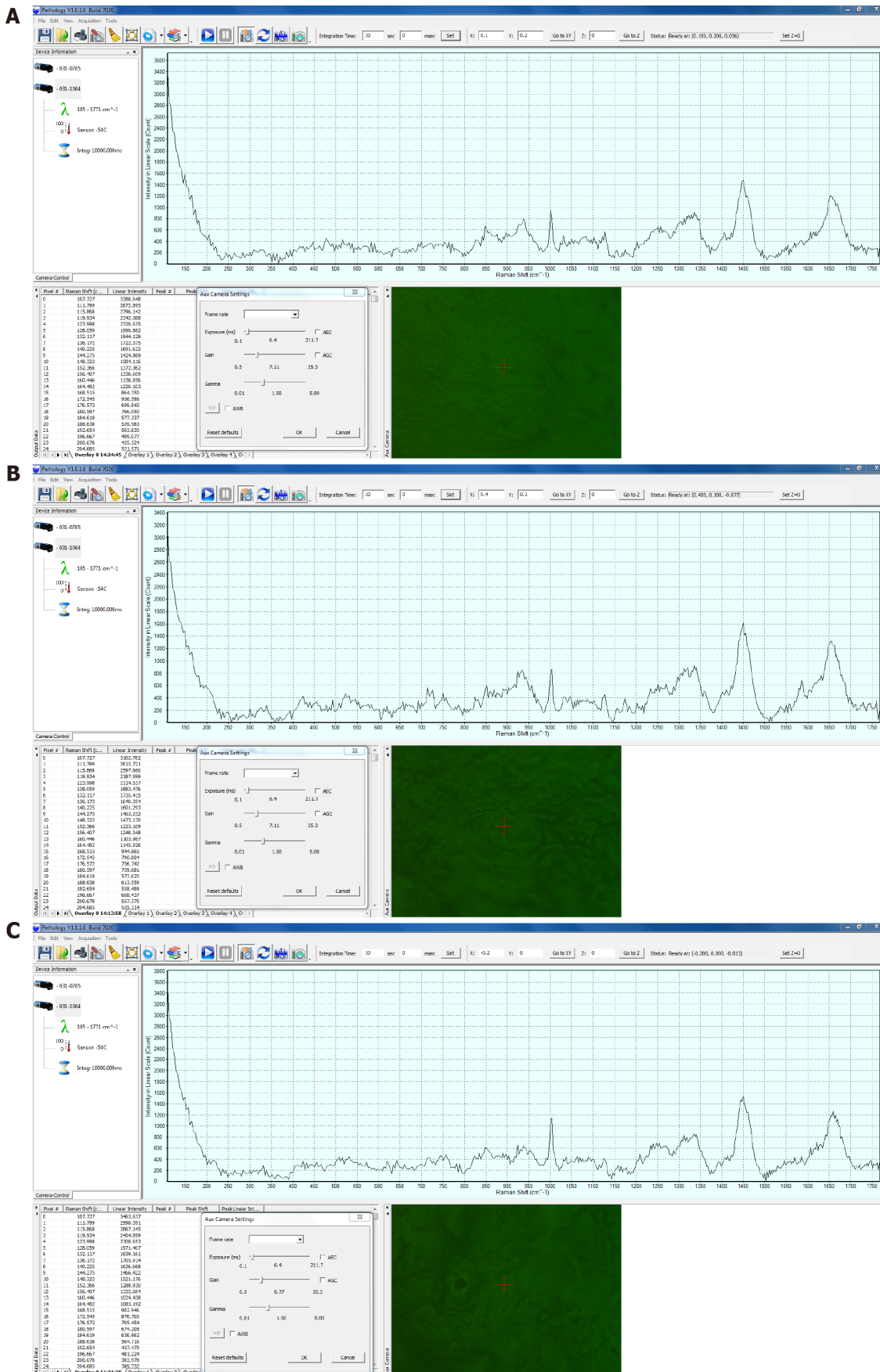
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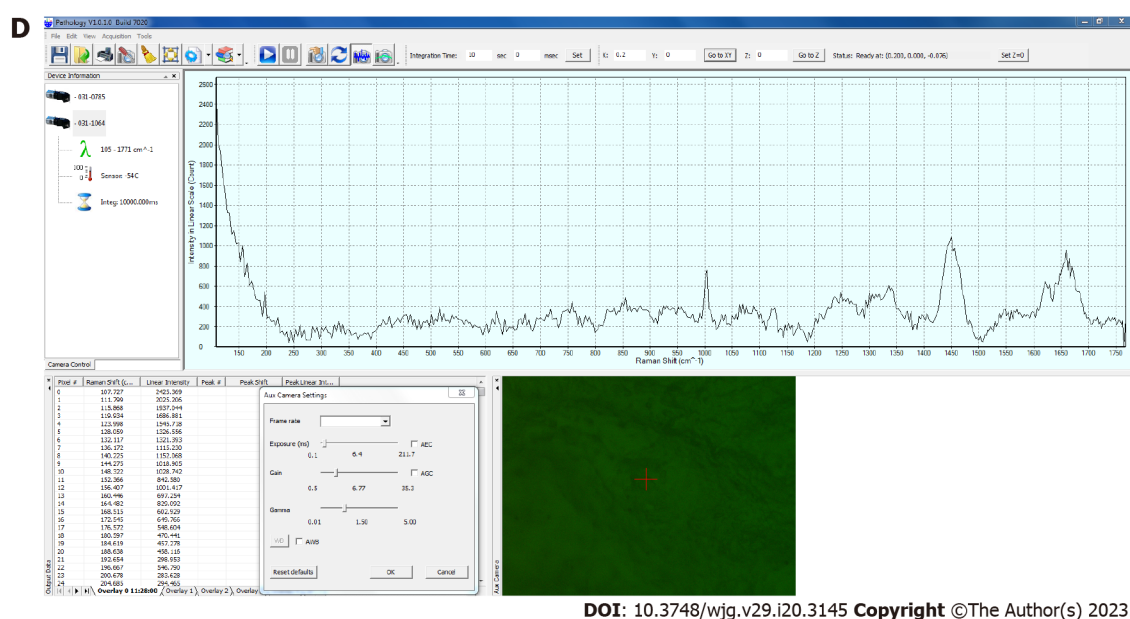
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INTRODUCTION

Gastrointestinal cancers are typically evaluated using various investigative modalities, like radiological imaging, endoscopy, and histopathology[1,2]. Although tissue diagnosis through histopathological evaluation remains the gold standard for diagnosis, it is primarily based on morphological interpretation, even in current molecular-level evaluation practices[3]. In addition to morphology, many other such modalities are rapidly making inroads into practice[3,4]. Unlike morphology alone, immuno-histochemistry, which has become almost synonymous with objective histopathological evaluation, is further aided by fluorescence *in situ* hybridization and sequencing. At the other end of the spectrum lies non-invasive/minimally invasive, non-label-based evaluation options, such as near-infrared spectroscopy and Raman spectroscopy (RS)[5].

Endoscopic biopsy is the preferred diagnostic method for gastrointestinal cancer[6]. Recent technological advances in endoscopy have enabled reliable diagnoses based on endoscopic findings alone, which have been further improved by the incorporation of artificial intelligence[5-7]. However, these





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Figure 1 Example of Raman spectra. Screenshots from the software window illustrating the Raman shift plotted against the intensity on a linear scale. A: Normal esophageal mucosa; B: Esophageal squamous cell carcinoma; C: Normal stomach mucosa; D: Stomach adenocarcinoma.

methodologies rely primarily on morphological assessments and do not consider molecular biological information. Tumor tissues contain abnormal molecules and proteins that are absent in normal tissues. The addition of molecular biological information to existing highly evolved morphological diagnostic methods may become an epoch-making technique with higher diagnostic accuracy[5,8,9].

Hence, we evaluated the biological samples using RS to obtain molecular information[8,10]. RS is a nondestructive inspection method that identifies a substance by measuring the type and amount of molecules contained in the substance by analyzing the wavelength of the reflected light obtained by irradiating the target substance with light, such as a laser[11-13]. Furthermore, RS can be used to evaluate substances with all types of properties, including solids[14-16], liquids[9,11], and gases[17], without sample pretreatment. Therefore, RS is an excellent inspection method for evaluating substances within a short time.

Therefore, attempts have been made to evaluate tissue samples using RS. Bergholt *et al*[4] presented a diagnostic technique for esophageal disease in the gastrointestinal tract using RS and published diagnostic techniques for gastric cancer. Moreover, Duraipandian *et al*[3] reported a diagnostic technique for gastric cancer using RS. However, a standard method for biological evaluations using RS has not yet been established.

A major challenge in using RS to evaluate biological samples is interference of autofluorescence[18]. To address this issue, we developed a novel micro-Raman device with unique features that enables effective evaluation of biological samples using RS.

We selected a near-infrared laser, which is not easily affected by autofluorescence, as the excitation light source. Because the Raman-scattered light intensity is inversely proportional to the square of the excitation light wavelength, the scattered light intensity of a long-wavelength near-infrared laser is low [18]. We developed a highly sensitive circuit to detect this weakly scattered light. We attempted to record the Raman-scattered light wavelengths from living esophageal tissues using a micro-Raman device with these characteristics.

MATERIALS AND METHODS

Study population and data collection

Sixteen patients aged 80 years or younger who underwent endoscopic submucosal dissection of the esophagus or stomach at the Digestive Disease Center of Showa University Koto Toyosu Hospital were recruited for this study. Written consent was obtained from all participants. Eleven participants were male and five were female, with an age range of 38-80 years. Eighteen specimens (esophagus, $n = 6$; stomach, $n = 12$) were analyzed (Table 1). This study was approved by the in-hospital clinical research review board (approval number: 18T5009) and conducted in accordance with the Declaration of Helsinki.

Table 1 Patients' background

Patient number	Patient	Clinical diagnosis	Number of lesions	Treatment date
1	50-year-old, female	Esophageal cancer	1	December 20, 2018
2	65-year-old, male	Esophageal cancer	1	December 20, 2018
3	61-year-old, male	Esophageal cancer	1	January 11, 2019
4	61-year-old, male	Esophageal cancer	2	January 18, 2019
5	78-year-old, female	Esophageal cancer	1	April 18, 2019
6	72-year-old, female	Stomach cancer	1	January 8, 2019
7	63-year-old, male	Stomach cancer	1	January 18, 2019
8	56-year-old, male	Stomach cancer	1	January 24, 2019
9	58-year-old, male	Stomach cancer	1	February 3, 2019
10	79-year-old, male	Stomach cancer	1	February 4, 2019
11	77-year-old, male	Stomach cancer	1	February 17, 2019
12	80-year-old, male	Stomach cancer	2	February 28, 2019
13	65-year-old, male	Stomach cancer	1	February 28, 2019
14	68-year-old, male	Stomach cancer	1	April 8, 2019
15	66-year-old, female	Stomach adenoma	1	May 19, 2019
16	38-year-old, female	Stomach submucosal tumor	1	April 9, 2019

Measurement protocol and data capture

Esophageal and stomach tissue samples (including mucosal and submucosal tissues) were collected immediately after endoscopic resection. The tissue was attached to a black rubber plate with a metal pin, and a 0.02 mm thick aluminum foil was sandwiched between the rubber plate and the esophageal tissue to prevent Raman scattered light from the rubber plate from being included in the measurements.

A linear or grid-like measurement grid was set up to include endoscopically diagnosed lesions in the mucosa and the surrounding normal mucosa. Multiple measurement points were selected at 5 mm or 10 mm intervals depending on the shape of the clinically determined lesion. Raman scattered light wavelengths were recorded from 19 or 6 locations at intervals of 200 μm in the vertical and horizontal directions for each selected point depending on the feasibility. To prevent the sample from drying out, distilled water was sprayed on the sample as necessary during the measurement.

A RS device (BaySpec Inc., San Jose, CA, United States) was used, with a computer-controlled stage, an objective lens of 20 \times magnification, a correction collar for near-infrared microscopy (LCPLN20XIR, Olympus Corporation, Tokyo, Japan), and an excitation laser wavelength of 1064 nm. The measurements were captured with a laser power of 200 mW and an exposure time of 10 s per point. Pathologic System Software Version 1.0.1.0 (BaySpec, Inc., San Jose CA, United States) was used and baseline correction was performed without smoothing the waveform. Examples of the Raman spectra of the esophageal and gastric mucosa are shown in [Figure 1](#).

Analysis

Fifteen types of Raman shifts corresponding to the constituent molecules of living tissues ([Table 2](#)) were selected for the analysis. From the recorded Raman-scattered light waveforms, the scattered light intensities of 15 types of Raman shifts were extracted, and the optimum combination matching the range of squamous cell carcinoma (SCC) of the esophagus and gastric adenocarcinoma (AC) by histopathological diagnosis was determined.

RESULTS

The study involved 16 patients, and 18 specimens were resected for histopathological analysis. The results showed that five of the specimens were SCC of the esophagus, 10 were well-differentiated tubular AC of the stomach, one was a gastric adenoma, and one was a gastric carcinoid tumor (NET G1) ([Table 3](#)).

Raman spectra were recorded for all six esophageal and 12 gastric specimens. All combinations of the scattered light intensities of the 15 Raman shifts were calculated to determine the combination that best matched the range of cancer by histopathological diagnosis. In esophageal squamous cell cancer, the

Table 2 Fifteen Raman shift parameters obtained from the raw tissues and the range of the shifts[7,9]

Number	Assigned peak	Shift start (cm ⁻¹)	Shift end (cm ⁻¹)
RS 1	Phenylalanine C-C twist	611	631
RS 2	Cholesterol	700	720
RS 3	Tryptophane ring breath	751	771
RS 4	Tyrosine ring breath	830	850
RS 5	Phenylalanine ring breath	993	1013
RS 6	Skeletal C-C	1060	1080
RS 7	Nucleotide O-P-O	1091	1111
RS 8	Skeletal C-C stretch	1123	1143
RS 9	Amide III beta-sheet	1244	1264
RS 10	Amide III delta (CH) ₂	1275	1295
RS 11	Amide III alpha-helix	1322	1342
RS 12	CH ₂ stretch	1408	1428
RS 13	CH ₃ deformation	1448	1468
RS 14	Phenylalanine C=C	1596	1616
RS 15	Amide I alpha-helix	1647	1667

Table 3 Esophageal and gastric samples and histopathological diagnosis along with TNM classification

Sample number	Patient	Pathological diagnosis (UICC TNM classification)
Eso-1	50-year-old, female	SCC, pT1a (LPM)
Eso-2	65-year-old, male	SCC, pT1a (LPM)
Eso-3	61-year-old, male	SCC, pT1a (MM)
Eso-4	61-year-old, male	SCC, pTis
Eso-5	61-year-old, male	SCC, pTis
Eso-6	78-year-old, female	SCC, pT1b (SM)
Sto-1	72-year-old, female	Tub1, pT1a
Sto-2	63-year-old, male	Tub1>tub2, pT1b2
Sto-3	56-year-old, male	Tub1, pT1b1
Sto-4	58-year-old, male	Tub1, pT1b1
Sto-5	79-year-old, male	Tub1, pT1b1
Sto-6	77-year-old, male	Tub1, pT1a
Sto-7	80-year-old, male	Tub1, pT1b1
Sto-8	80-year-old, male	Tub1, pT1a
Sto-9	65-year-old, male	Tub1, pT1a
Sto-10	68-year-old, male	Tub1, pT1a
Sto-11	66-year-old, female	Tubular adenoma
Sto-12	38-year-old, female	Carcinoid tumor (NET G1), pT1b

Tub1: Well differentiated tubular adenocarcinoma. UICC: Union for International Cancer Control; SCC: Squamous cell carcinoma; TNM: Tumor node metastasis.

Table 4 Raman shift peaks observed in the esophageal and the gastric samples

	Peak 1	Peak 2	Peak 3
Esophagus	RS 8	RS 1	RS 13
Stomach	RS12	RS 5	RS 15

Table 5 Cut-off ratios obtained from the esophagus and stomach samples

	Cut-off 1 (peak 2/peak 1)	Cut-off 2 (peak 3/peak 1)
Esophagus	0.65	2.0
Stomach	1.85	2.0

scattered light intensity of Skeletal C-C stretch was Peak 1, the scattered light intensity of Phenylalanine C-C twist was Peak 2, the scattered light intensity of CH₃ deformation was Peak 3 (Table 4), and Peak 2/Peak 1 was defined as Cut-off 1. Peak 3/Peak 1 was defined as Cut-off 2 (Table 5). The site with more than half of the measurement points having Cut-off 1 greater than 0.65 and Cut-off 2 greater than 2.0 was determined to be the best match for histopathological SCC of the esophagus (Figure 2).

For gastric AC, the scattered light intensity of the CH₂ stretch was Peak 1, Phenylalanine ring breadth was Peak 2, and Amide I alpha helix was Peak 3 (Table 4). The ratio of Peak 2/Peak 1 was defined as Cut-off 1 and Peak 3/Peak 1 was defined as Cut-off 2 (Table 5). The site with more than half of the measurement points with Cut-off 1 greater than 1.85 and Cut-off 2 greater than 2.0 was determined to be the best match for the histopathological diagnosis of gastric AC (Figure 3).

DISCUSSION

Patients with gastrointestinal cancers affecting the esophagus and stomach have symptoms, such as difficulty in swallowing, heartburn, and fullness[6]. Contrast examination or endoscopy is often performed in patients with a clinical evaluation suggestive of esophageal or gastric cancer[6]. Histopathological evaluation of the tissues obtained *via* endoscopic biopsy can be used to confirm the diagnosis[3, 4,6]. In recent years, the accuracy of endoscopic finding-based diagnoses has dramatically improved. One of the reasons for this is the better quality of images obtained through high-resolution camera optics[3,6].

It is highly possible that the ability to diagnose cancer will improve further if it is feasible to depict what cannot be delineated using conventional equipment by making the image richer in features. Another reason is the use of image classification algorithms based on artificial intelligence[13,16,19]. By recognizing and patterning endoscopic images using artificial intelligence, the existence of lesions is clarified, and diagnosis by doctors is supported. Since the diagnosis using artificial intelligence evaluates endoscopic images morphologically, it is possible to prevent the lesion from being overlooked [13,16]. However, its diagnostic ability is not higher than that of an experienced doctor. Another drawback is that diagnostic ability is greatly affected by the quality of the endoscopic image[5,9,14].

As mentioned above, the accuracy of endoscopic diagnosis of lesions in the gastrointestinal tract has greatly improved; however, it is still at the morphological interpretation level[6,7]. Histopathological diagnosis, which is the current definitive diagnostic method, is a form of morphological evaluation; moreover, qualitative information that assists in morphological evaluation is required to improve diagnostic accuracy beyond the existing practice. RS can be used to add qualitative information to the morphological information[2,20]. Short *et al*[1] published a technique for diagnosing lung cancer using bronchoscopy combined with RS. Lui *et al*[21] reported a method for diagnosing skin cancer using RS. Krishna *et al*[22] reported a method for diagnosing oral cancer using RS. Jermyn *et al*[20] reported that RS can be used to identify lesion areas during brain surgery. Furthermore, Bergholt *et al*[6] presented a diagnostic technique for esophageal disease in the gastrointestinal tract using RS, and published diagnostic techniques for gastric cancer[4]. Duraipandian *et al*[3] reported a diagnostic technique for gastric cancer using RS. Molckovsky *et al*[2] reported a diagnostic technique for colon cancer using RS. However, a standard method for analyzing living organisms and biological samples using RS is yet to be developed.

In this study, we selected 15 Raman shifts corresponding to constituent molecules in living tissues. The analysis results of the scattered light intensity of Raman shift of Skeletal C-C stretch, Phenylalanine C-C twist, and CH₃ deformation in SCC of the esophagus, and Raman shift of CH₂ stretch, Phenylalanine ring breadth, and Amide I alpha-helix in gastric AC. The results of the analysis of the scattered light intensity of the shift were in good agreement with the range of morphological cancer

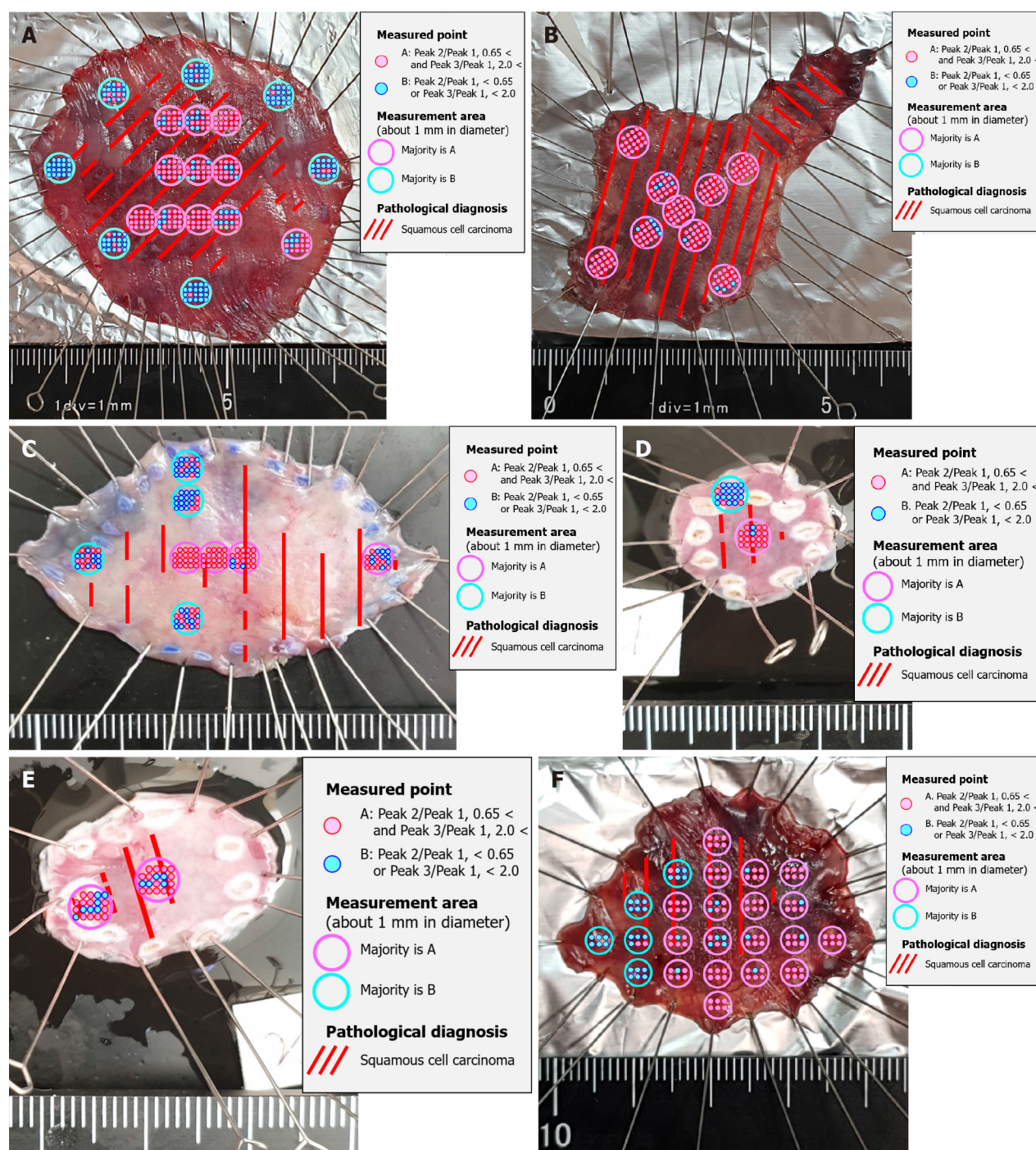
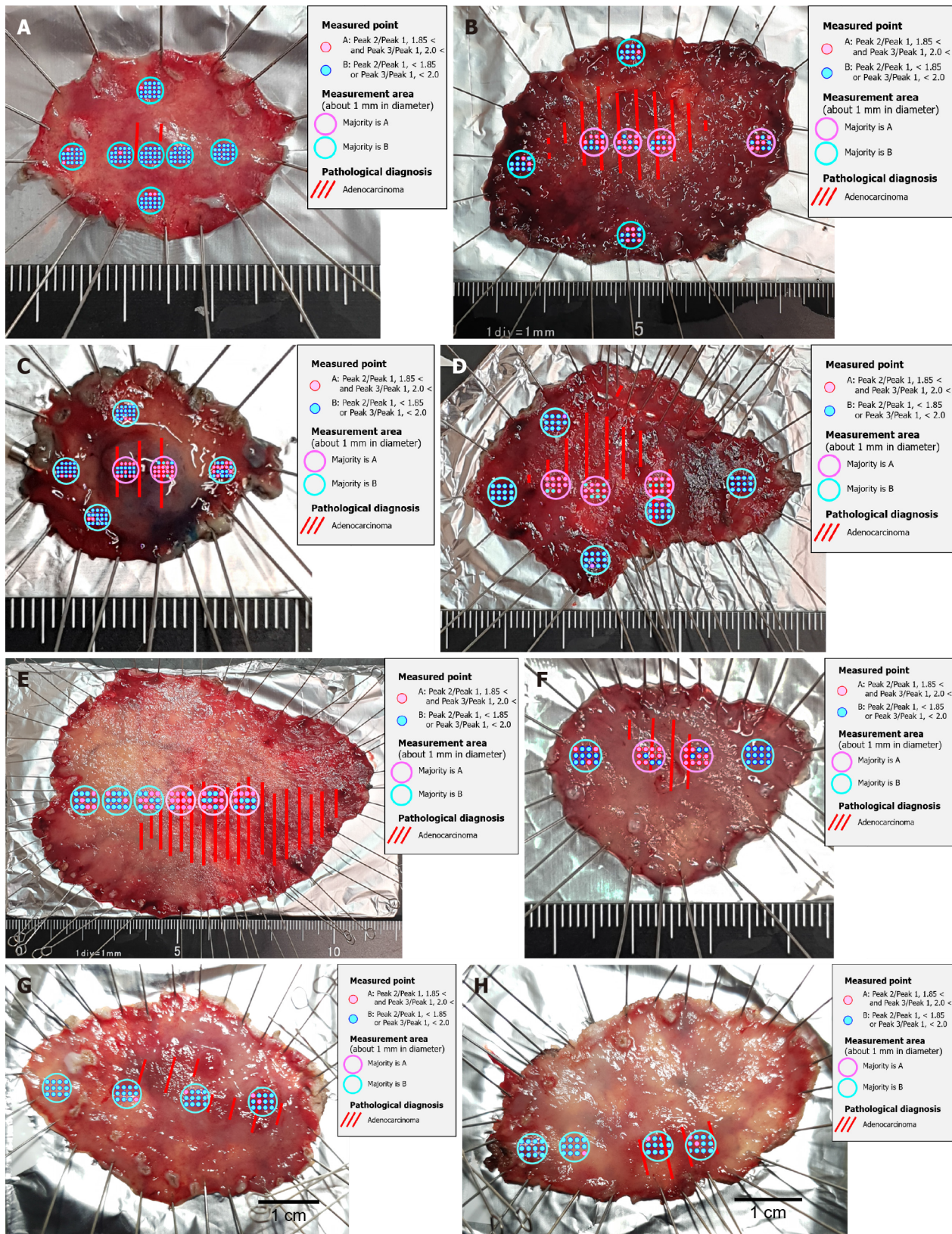
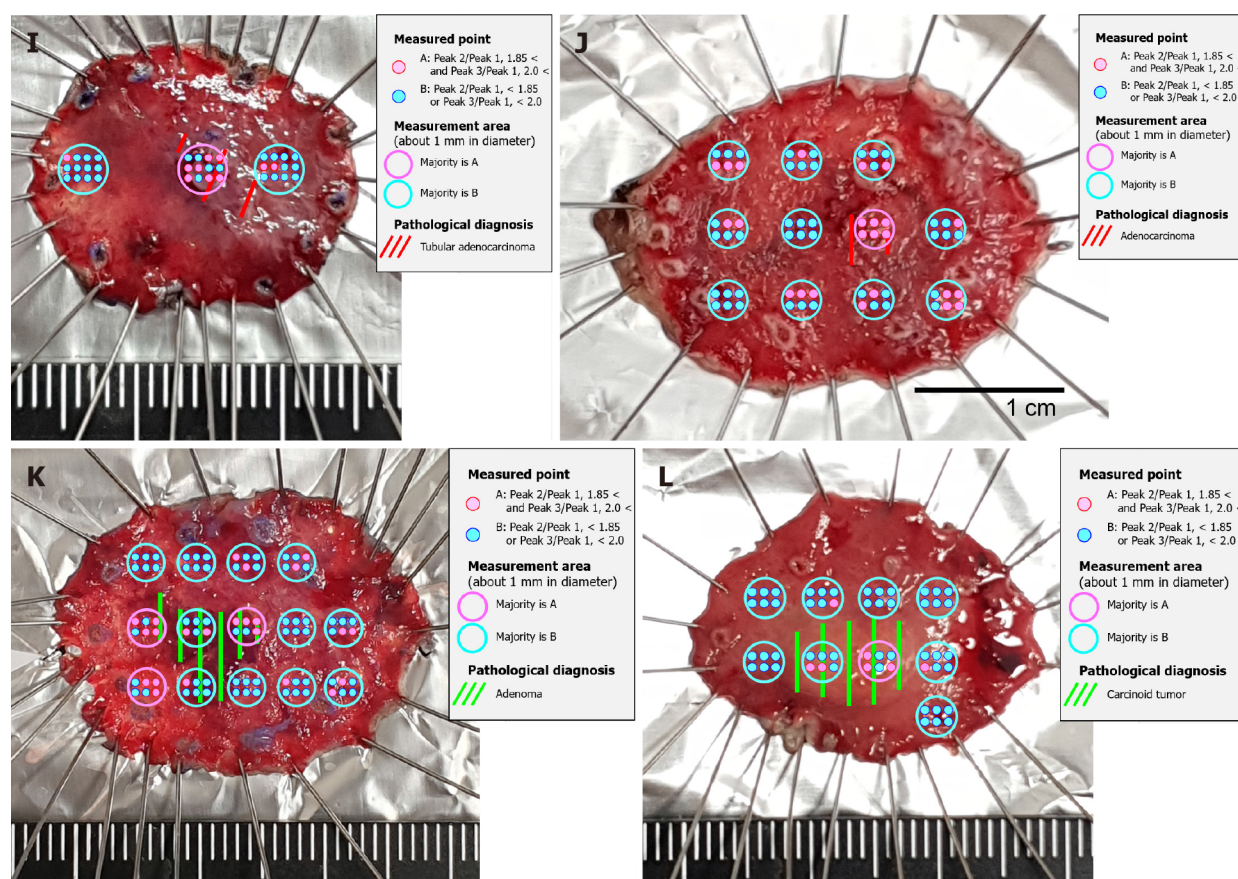


Figure 2 Comparison of Raman spectroscopic analysis and pathological diagnosis of sample Eso [squamous cell carcinoma, pT1a(LPM)-pT1b(SM)]. A: Eso-1; B: Eso-2; C: Eso-3; D: Eso-4; E: Eso-5; F: Eso-6.

diagnoses offered by histopathological evaluation. SCC of the esophagus and gastric AC are distinct types of cancers with different biological characteristics. Therefore, a significant Raman shift is expected to differ in the analysis conducted using RS. The technique presented in this study has the potential to provide qualitative evaluation in addition to the morphological evaluation currently used. In particular, it may offer valuable information regarding lesions that are challenging to assess using endoscopic or histopathological diagnoses alone.

However, at some sites, the results of this technique did not match the histopathological diagnoses. The limitations of this study include the small number of samples analyzed ($n = 18$), which included both the esophagus and stomach. Hence, there is no doubt that more samples must be analyzed in the future to confirm the accuracy of this technique. Using this technique, molecular information not solely dependent on morphological interpretation can be obtained. Furthermore, it may indicate the degree of mucosal abnormality from normal to cancerous. Evaluation of precancerous tissues may enable highly





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Figure 3 Comparison of Raman spectroscopic analysis and pathological diagnosis of sample Sto (well-differentiated tubular adenocarcinoma, pT1a-pT1b1; tubular adenoma; carcinoid tumor, NET G1). A: Sto-1; B: Sto-2; C: Sto-3; D: Sto-4; E: Sto-5; F: Sto-6; G: Sto-7; H: Sto-8; I: Sto-9; J: Sto-10; K: Sto-11; L: Sto-12.

accurate preventive medicine. We aim to further advance this research and establish a method for predicting the current and future states of the gastrointestinal mucosa with high accuracy by clarifying the correlation between the Raman-scattered light intensity and the tissue state at each Raman shift.

CONCLUSION

Based on the results, it was concluded that RS could accurately identify esophageal SCC and gastric AC by analyzing the scattered light intensities of the 15 types of Raman shifts. The optimum combinations of Raman shifts were identified as Peak 1, Peak 2, and Peak 3 for both cancer types, with specific cut-off values for each peak. Although it is currently used in endoscopic treatment, the qualitative diagnosis of lesions, risk of residual lesions, *etc.*, can be confirmed in a short time by analyzing excised specimens, and treatment can be provided as necessary.

ARTICLE HIGHLIGHTS

Research background

Cancer diagnosis plays an important role in patient management, many researchers focused on the cancer detection, and novel, rapid, and label-free techniques are being developed for routine clinical practice.

Research motivation

To address the issue of using Raman spectroscopy (RS) to evaluate biological samples is interference of autofluorescence.

Research objectives

Our study is to evaluate the use of Raman-scattered light spectrum for detecting endoscopically resected specimens of esophageal squamous cell carcinoma (SCC) and gastric adenocarcinoma (AC).

Research methods

We created a Raman device, which is suitable for observing living tissues, and we attempted to acquire Raman-scattered light spectra in endoscopically resected specimens of six esophageal tissues and 12 gastric tissues. Furthermore, we performed a correlation between the obtained Raman scattered light spectra and histopathological diagnosis.

Research results

We obtained Raman scattered light spectra from all six esophageal and 12 gastric specimens successfully, and developed an algorithm to identify the range of esophageal SCC and gastric AC with an accuracy close to that of histopathological diagnoses.

Research conclusions

In this study, we utilized RS to rapidly detect cancer in resected esophageal and stomach specimens, providing information beyond morphology.

Research perspectives

By providing detailed molecular level data, RS can provide a more comprehensive understanding of disease processes and aid in accurate diagnosis.

FOOTNOTES

Author contributions: Ito H and Miyazaki T designed the study; Ito H was the guarantor, performed measurement, analysis, and interpretation of the data, and drafted the initial manuscript; Uragami N and Miyazaki T contributed to the acquisition, analysis, and interpretation of the data; Uragami N, Shimamura Y, Ikeda H, Nishikawa Y, Onimaru M, Matsuo K, and Isozaki M participated in the collection of the sample; Yang W designed and produced the Raman spectroscopy; Issha K continued importing the Raman spectroscopy; Kimura S, Kawamura M, Yokoyama N, Kushima M, and Inoue H revised the article critically for important intellectual content; Kushima M performed histopathological diagnosis of samples.

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Informed consent statement: All the participants provided written informed consent to participate in the study.

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Data sharing statement: No additional data are available.

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REFERENCES

- 1 **Short MA**, Lam S, McWilliams A, Zhao J, Lui H, Zeng H. Development and preliminary results of an endoscopic Raman probe for potential *in vivo* diagnosis of lung cancers. *Opt Lett* 2008; **33**: 711-713 [PMID: [18382526](#) DOI: [10.1364/OL.33.000711](#)]
- 2 **Molckovsky A**, Song LM, Shim MG, Marcon NE, Wilson BC. Diagnostic potential of near-infrared Raman spectroscopy in the colon: differentiating adenomatous from hyperplastic polyps. *Gastrointest Endosc* 2003; **57**: 396-402 [PMID: [12612529](#) DOI: [10.1067/mge.2003.105](#)]
- 3 **Duraipandian S**, Sylvest Bergholt M, Zheng W, Yu Ho K, Teh M, Guan Yeoh K, Bok Yan So J, Shabbir A, Huang Z. Real-time Raman spectroscopy for *in vivo*, online gastric cancer diagnosis during clinical endoscopic examination. *J Biomed Opt* 2012; **17**: 081418 [PMID: [23224179](#) DOI: [10.1117/1.JBO.17.8.081418](#)]
- 4 **Bergholt MS**, Zheng W, Lin K, Ho KY, Teh M, Yeoh KG, So JB, Huang Z. Raman endoscopy for *in vivo* differentiation between benign and malignant ulcers in the stomach. *Analyst* 2010; **135**: 3162-3168 [PMID: [20941419](#) DOI: [10.1039/c0an00336k](#)]
- 5 **Chakraborty A**, Ghosh A, Barui A. Advances in surface-enhanced Raman spectroscopy for cancer diagnosis and staging. *J Raman Spectrosc* 2020; **51**: 7-36 [DOI: [10.1002/jrs.5726](#)]
- 6 **Bergholt MS**, Zheng W, Ho KY, Yeoh KG, Teh M, So JBY, Huang Z. Real-time depth-resolved Raman endoscopy for *in vivo* diagnosis of dysplasia in Barrett's esophagus. Proceedings of the SPIE BiOS; 2013 Mar 20; San Francisco, California, United States: Society of Photo-Optical Instrumentation Engineers, 2003
- 7 **Ito H**, Uragami N, Miyazaki T, Yang W, Issha K, Matsuo K, Kimura S, Arai Y, Tokunaga H, Okada S, Kawamura M, Yokoyama N, Kushima M, Inoue H, Fukagai T, Kamijo Y. Highly accurate colorectal cancer prediction model based on Raman spectroscopy using patient serum. *World J Gastrointest Oncol* 2020; **12**: 1311-1324 [PMID: [33250963](#) DOI: [10.4251/wjgo.v12.i11.1311](#)]
- 8 **Wang K**, Qiu Y, Wu C, Wen ZN, Li Y. Surface-enhanced Raman spectroscopy and multivariate analysis for the diagnosis of oral squamous cell carcinoma. *J Raman Spectrosc* 2023; **54**: 355-362 [DOI: [10.1002/jrs.6495](#)]
- 9 **Lei J**, Yang D, Li R, Dai Z, Zhang C, Yu Z, Wu S, Pang L, Liang S, Zhang Y. Label-free surface-enhanced Raman spectroscopy for diagnosis and analysis of serum samples with different types lung cancer. *Spectrochim Acta A Mol Biomol Spectrosc* 2021; **261**: 120021 [PMID: [34116414](#) DOI: [10.1016/j.saa.2021.120021](#)]
- 10 **Yue X**, Wang Y, Guo D, He G, Sui C, Qu Z, Zhao Y, Liu X. Surface-enhanced Raman spectroscopy is an accurate technique in diagnosis of breast cancer: A meta-analysis. *J Raman Spectrosc* 2022; **53**: 2058-2067 [DOI: [10.1002/jrs.6437](#)]
- 11 **Falamas A**, Faur CI, Baciut M, Rotaru H, Chirila M, Cinta Pinzaru S, Hedesiu M. Raman spectroscopic characterization of saliva for the discrimination of oral squamous cell carcinoma. *Anal Lett* 2020; **54** [DOI: [10.1080/00032719.2020.1719129](#)]
- 12 **Zhang H**, Cheng C, Gao R, Yan Z, Zhu Z, Yang B, Chen C, Lv X, Li H, Huang Z. Rapid identification of cervical adenocarcinoma and cervical squamous cell carcinoma tissue based on Raman spectroscopy combined with multiple machine learning algorithms. *Photodiagnosis Photodyn Ther* 2021; **33**: 102104 [PMID: [33212265](#) DOI: [10.1016/j.pdpdt.2020.102104](#)]
- 13 **Zhang L**, Wu Y, Zheng B, Su L, Chen Y, Ma S, Hu Q, Zou X, Yao L, Yang Y, Chen L, Mao Y, Ji M. Rapid histology of laryngeal squamous cell carcinoma with deep-learning based stimulated Raman scattering microscopy. *Theranostics* 2019; **9**: 2541-2554 [PMID: [31131052](#) DOI: [10.7150/thno.32655](#)]
- 14 **Zhan Q**, Li Y, Yuan Y, Liu J. The accuracy of Raman spectroscopy in the detection and diagnosis of oral cancer: A systematic review and meta-analysis. *J Raman Spectrosc* 2020; **51**: 2377-2397 [DOI: [10.1002/jrs.5940](#)]
- 15 **Zheng C**, Qing S, Wang J, Lü G, Li H, Lü X, Ma C, Tang J, Yue X. Diagnosis of cervical squamous cell carcinoma and cervical adenocarcinoma based on Raman spectroscopy and support vector machine. *Photodiagnosis Photodyn Ther* 2019; **27**: 156-161 [PMID: [31136828](#) DOI: [10.1016/j.pdpdt.2019.05.029](#)]
- 16 **Yu M**, Yan H, Xia J, Zhu L, Zhang T, Zhu Z, Lou X, Sun G, Dong M. Deep convolutional neural networks for tongue squamous cell carcinoma classification using Raman spectroscopy. *Photodiagnosis Photodyn Ther* 2019; **26**: 430-435 [PMID: [31082525](#) DOI: [10.1016/j.pdpdt.2019.05.008](#)]
- 17 **Bögözi T**, Popp J, Frosch T. Fiber-enhanced Raman multi-gas spectroscopy: what is the potential of its application to breath analysis? *Bioanalysis* 2015; **7**: 281-284 [PMID: [25697186](#) DOI: [10.4155/bio.14.299](#)]
- 18 **Wachsmann-Hogiu S**, Weeks T, Huser T. Chemical analysis *in vivo* and *in vitro* by Raman spectroscopy--from single cells to humans. *Curr Opin Biotechnol* 2009; **20**: 63-73 [PMID: [19268566](#) DOI: [10.1016/j.copbio.2009.02.006](#)]
- 19 **Xia J**, Zhu L, Yu M, Zhang T, Zhu Z, Lou X, Sun G, Dong M. Analysis and classification of oral tongue squamous cell carcinoma based on Raman spectroscopy and convolutional neural networks. *J Mod Opt* 2020; **67**: 481-489 [DOI: [10.1080/09500340.2020.1742395](#)]
- 20 **Jermyn M**, Mok K, Mercier J, Desroches J, Pichette J, Saint-Arnaud K, Bernstein L, Guiot MC, Petrecca K, Leblond F. Intraoperative brain cancer detection with Raman spectroscopy in humans. *Sci Transl Med* 2015; **7**: 274ra19 [PMID: [25673764](#) DOI: [10.1126/scitranslmed.aaa2384](#)]
- 21 **Lui H**, Zhao J, McLean D, Zeng H. Real-time Raman spectroscopy for *in vivo* skin cancer diagnosis. *Cancer Res* 2012; **72**: 2491-2500 [PMID: [22434431](#) DOI: [10.1158/0008-5472.CAN-11-4061](#)]
- 22 **Krishna H**, Majumder SK, Chaturvedi P, Sidramesh M, Gupta PK. *In vivo* Raman spectroscopy for detection of oral neoplasia: a pilot clinical study. *J Biophotonics* 2014; **7**: 690-702 [PMID: [23821433](#) DOI: [10.1002/jbio.201300030](#)]



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