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REVIEW

### Molecular insights into clinical trials for immune checkpoint inhibitors in colorectal cancer: Unravelling challenges and future directions

Samantha Sharma, Naresh Singh, Anita Ahmed Turk, Isabella Wan, Akshay Guttikonda, Julia Lily Dong, Xinna Zhang, Mateusz Opyrchal

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### Abstract

Colorectal cancer (CRC) is a complex disease with diverse etiologies and clinical outcomes. Despite considerable progress in development of CRC therapeutics, challenges remain regarding the diagnosis and management of advanced stage metastatic CRC (mCRC). In particular, the five-year survival rate is very low since mCRC is currently rarely curable. Over the past decade, cancer treatment has significantly improved with the introduction of cancer immunotherapies, specifically immune checkpoint inhibitors. Therapies aimed at blocking immune checkpoints such as PD-1, PD-L1, and CTLA-4 target inhibitory pathways of the immune system, and thereby enhance anti-tumor immunity. These therapies thus have shown promising results in many clinical trials alone or in combination. The efficacy and safety of immunotherapy, either alone or in combination with CRC, have been investigated in several clinical trials. Clinical trials, including KEYNOTE-164 and CheckMate 142, have led to Food and Drug Administration approval of the PD-1 inhibitors pembrolizumab and nivolumab, respectively, for the treatment of patients with unresectable or metastatic microsatellite instabilityhigh or deficient mismatch repair CRC. Unfortunately, these drugs benefit only a small percentage of patients, with the benefits of immunotherapy remaining elusive for the vast majority of CRC patients. To this end, primary and secondary resistance to immunotherapy remains a significant issue, and further research is necessary to optimize the use of immunotherapy in CRC and identify biomarkers



to predict the response. This review provides a comprehensive overview of the clinical trials involving immune checkpoint inhibitors in CRC. The underlying rationale, challenges faced, and potential future steps to improve the prognosis and enhance the likelihood of successful trials in this field are discussed.

Key Words: Colorectal cancer; Immune checkpoint inhibitors; Clinical trials; Immunotherapy; Microsatellite instability; Microsatellite stability; DNA mismatch repair

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**Core Tip:** Colorectal cancer (CRC) often eludes early detection, limiting the efficacy of existing chemotherapy and targeted therapies. This article delves into the realm of immune checkpoint inhibitors in CRC, dissecting their mechanisms and outcomes through a comprehensive review of clinical trials. It sheds light on the underlying rationale, challenges faced, and potential strategies to improve prognosis and trial success in this critical domain. Notably, while microsatellite instabilityhigh CRC exhibits heightened responsiveness to checkpoint inhibitors, the article underscores potential breakthroughs in treating microsatellite stable CRC-the predominant cases-providing insights into bettering prognosis and trial outcomes in CRC treatment.

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### INTRODUCTION

Colorectal cancer (CRC) is a prevalent malignancy recognized worldwide for its intricate pathogenesis, diverse etiologies, and clinical outcomes[1,2]. Approximately 147950 new cases are expected to be diagnosed in 2023, along with an estimated number of 53200 deaths due to CRC[3]. Moreover, the incidence of early onset CRC is increasing as well[4]. CRC arises from the malignant transformation of epithelial cells lining the colon or rectum. The development of CRC is influenced by a multitude of risk factors, including advanced age, dietary choices, obesity, and inflammatory bowel disease[5-7]. The molecular pathogenesis of CRC is complex, with genetic and epigenetic alterations that drive tumorigenesis and contribute to disease progression[8-12]. These alterations intricately disrupt essential signaling pathways, such as WNT/β-catenin pathway, KRAS/BRAF/MEK/ERK pathway, and PI3K/AKT/mTOR pathway governing critical cellular processes, including cell proliferation, differentiation, and survival[8,9,13-15].

Currently, several approaches are employed for CRC treatment, including surgical procedures, chemotherapy, radiation therapy, targeted therapy, and immunotherapy[16]. However, following preliminary diagnosis, the 5-year survival rate of CRC patients is 65.0%, which significantly decreases to approximately 13% for metastatic CRC (mCRC) [17]. Treatment of advanced or mCRC is hindered by several challenges. Treatment options are particularly limited for patients who have exhausted multiple lines of treatment. Additionally, CRC tumors can develop resistance to chemotherapy, diminishing treatment efficacy over time[18,19]. The toxicity associated with chemotherapy and targeted therapies further complicates treatment and affects patients' quality of life. mCRC also has poor prognosis. Tumor heterogeneity adds another layer of complexity, contributing to treatment resistance and variability in patient responses[20,21]. To this end, immunotherapy targeting immune checkpoints such as PD-1/PD-L1 axis and CTLA-4 shows promise in treating advanced CRC, particularly in CRC tumors with microsatellite instability-high (MSI-H) or deficient mismatch repair (dMMR)[22]. Combination therapies, involving immunotherapy with chemotherapy, targeted therapies, and other immunomodulators, further offer the potential of synergistic effects and enhanced treatment efficacy [23,24]. Ongoing research efforts on predictive biomarkers, such as tumor mutational burden (TMB) and immune cell infiltration patterns, aim to identify patients most likely to benefit from immunotherapy [25,26]. Hence, immunotherapy holds promise as a transformative approach for the management of advanced or mCRC with durable responses and improved patient outcomes.

Immunotherapy using immune checkpoint inhibitions (ICIs) has recently emerged as a promising therapeutic approach for various cancers, including CRC. Understanding the immune infiltration patterns in CRC patients with microsatellite-stable (MSS) vs microsatellite-instability (MSI) phenotype is crucial for developing immunotherapeutic strategies. While MSI-H tumors may benefit from immunotherapy due to their higher immune infiltration and mutational load, MSS tumors may still require alternative or combination approaches to enhance the antitumor immune response and improve treatment outcomes [27,28]. To this end, ongoing research efforts aim to unravel the complexities of immune infiltration in different CRC subtypes to guide the development of more effective and personalized therapeutic interventions. Initial studies conducted between 2010 and 2013 showed limited clinical activity of ICIs in patients who were not selected based on specific biomarkers or treatment history [29-33]. Eventually, several promising findings have led to the approval of ICIs for MSI-H or dMMR CRC. Nonetheless, a low response to immunotherapy remains a

significant challenge in the treatment of MSS or proficient MMR (pMMR) CRC, highlighting the need for further research to enhance effectiveness and identify biomarkers to improve treatment outcomes[34,35]. Newer immunotherapeutic approaches have been investigated for CRC treatment, including cancer vaccines, adoptive T-cell therapy, and oncolytic viruses[36,37]. These approaches aim to stimulate the immune response against cancer cells by various means, including inducing antigen-specific T cell responses, genetically modifying T-cells to recognize and attack cancer cells, and using viruses to selectively target and destroy cancer cells[38,39]. A multitude of clinical trials, spanning both ongoing and concluding studies, have been conducted to explore the efficacy and safety of diverse drugs and combination therapies for CRC treatment. This review provides insights into the current landscape, challenges, and potential advancements in this field. CRC clinical trials involving ICIs and their mechanistic actions are outlined, treatment strategies and the future trajectory of CRC therapeutics are discussed.

### MOLECULAR INSIGHTS AND THERAPEUTIC PROGRESS IN CRC

Molecular characterization of CRC has identified two major subtypes, MSS and MSI CRC that account for approximately 85% and 15% of all CRC cases, respectively[40-42]. Clinical and pathological features of MSS CRC differ from those of MSI CRC[2]. Specifically, MSS CRC is typically associated with older age, male sex, and distal colon location, whereas MSI CRC is associated with younger age, female sex, proximal colon location, and better prognosis[41]. Furthermore, the MMR status and CRC are intricately linked due to their role in maintaining genome integrity and preventing the accumulation of mutations that can lead to cancer[43,44]. MSI-H CRC tends to have a dMMR status, a higher mutational load, and a distinct molecular profile compared to MSS CRC, which has a pMMR[43-45]. In particular, MSI and MMR status are predictive biomarkers for response to ICIs therapy[34,42]. The consensus molecular subtype (CMS) classification system divides CRC into four distinct subtypes based on gene expression profiles: CMS1 (immune), CMS2 (canonical), CMS3 (metabolic), and CMS4 (mesenchymal)[46]. Each CMS subtype has a distinct molecular signature and clinical phenotype. The system thus offers a clear biological understanding and serves as a foundation for future clinical stratification and targeted interventions based on specific subtypes.

Over the years, substantial progress has been achieved in the development of CRC therapeutics, resulting in enhanced survival rates attributed to advancements in primary and adjuvant treatment modalities[47]. Notably, the inclusion of chemotherapy as a neoadjuvant or adjuvant intervention has emerged as a strategic approach aimed at mitigating the tumor burden, reduction, and stabilization[48-51]. Chemotherapy maintains its pivotal status in current treatment strategies. However, the utility of chemotherapy is curtailed by a restricted therapeutic range, significant adverse reactions, and the frequent occurrence of acquired resistance[52]. Several chemotherapy agents, radiotherapies, and other physical forces also induce destruction of cells and tissues, leading to death of immune cells and subsequently enhancing therapeutic outcomes[53-56]. Immunotherapy with checkpoint inhibitors has provided a significant improvement in cancer treatment, demonstrating high efficacy and manageable side effects in various tumor types[57-62]. However, the success of immunotherapy in CRC patients remains limited, with only a small subset of cases characterized by MSI-H or dMMR benefiting from treatment[63,64]. Thus, despite over 50 decades of research on immunotherapy for CRC treatment and major advancements, significant challenges remain in the diagnosis and management of CRC, particularly in the context of advanced or metastatic disease[27] (Figure 1).

Immuno-oncology is an emerging field of cancer treatment that involves harnessing the patient's immune system to recognize and eliminate cancer cells[64]. Immunotherapy can potentially improve treatment outcomes for patients with a wide range of malignancies[38]. Immunotherapy is often considered more beneficial than chemotherapy due to its ability to induce durable immune responses. Unlike chemotherapy, which primarily induces short-term cytotoxic effects on cancer cells and eliminates immunosuppressive cells[65-67], immunotherapy activates the immune system, particularly cytotoxic T-cells, to recognize and target cancer cells[38]. Consequently, immunotherapy offers the potential for sustained protection against cancer by maintaining an immunological "memory" that can detect and eliminate cancer cells in case of re-encounter[68,69]. Immunotherapy for cancer has thus brought about revolutionary transformations in the field of oncology, extending the survival of individuals diagnosed with aggressive life-threatening malignancies[57-61]. CRC patients with MSI-H or dMMR status show higher mutation rates, more neoantigens, and increased tumor-infiltrating lymphocytes (TILs), particularly cytotoxic T-cells[70], fostering a robust antitumor immune response. In contrast, MSS tumors have an immunosuppressive microenvironment with regulatory T-cells (Tregs) and other immunosuppressive cell types that hinder effector T cell activity[71,72]. Eventually, the prognostic value of the immunoscore was initially established in individuals with colon cancer, showing its ability to assess prognosis based on factors such as the density, type, and localization of infiltrating immune cells[73].

ICIs are drugs that blocks certain key proteins on the surface of immune cells, particularly T-cells, and cancer cells, and release the brakes on the immune response. The development of ICIs has been a breakthrough in the field of cancer immunotherapy[36,69,74-76]. These proteins, known as immune checkpoints, play crucial roles in regulating the immune response. By blocking these checkpoints, ICIs enhance the ability of the immune system to recognize and attack cancer cells, thereby boosting the body's natural anti-cancer response[38]. ICIs have become a cornerstone of cancer therapy, with a wide range of approved agents available for multiple malignancies, leading to increased utilization in various treatment settings, including (neo)adjuvant and maintenance therapy. Thus, ICIs are accessible to nearly half of metastatic cancer patients in economically developed countries[77-79].

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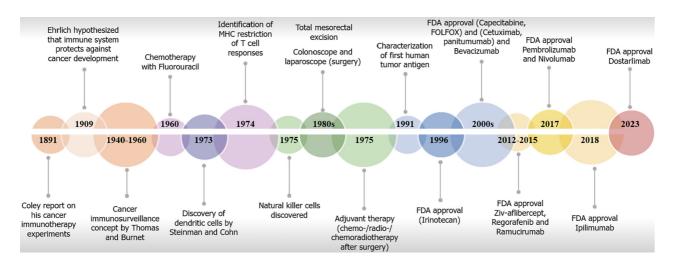


Figure 1 Timeline with key milestones in immuno-oncology research and United States Food and Drug Administration-approved immune checkpoint inhibitors in colorectal cancer.

### UNDERSTANDING THE MECHANISM OF IMMUNE CHECKPOINT INHIBITORS IN CRC

A solid understanding of the molecular drivers of CRC and identification of biomarkers of treatment response are essential for improving immunotherapy outcomes in patients with this disease[28,73,80-85]. A key element is the high TMB caused by genetic or sporadic mutations in MMR genes (such as MLH1, MSH2, MSH6, and PMS2), resulting in a deficiency of MMR proteins. This leads to an accumulation of genetic mutations in microsatellites[15,41], as observed in MSI-H or dMMR CRC tumors, compared to MSI-low (MSI-L) or pMMR. Consequently, enhanced immunogenicity is observed in such CRC cases, characterized by a higher count of neoantigens and substantial immune cell infiltration, including high numbers of tumor-infiltrating immune cells, such as CD8+ and CD4+ T-cells and macrophages[40,73,86-90]. Additionally, these tumors exhibit a microenvironment enriched with type I interferons, which distinguishes them from other CRC subtypes[87]. This immune-rich trait has been linked to improved rates of response to ICIs that block the PD-1/PD-L1 axis and T-cell activation[40,91]. In contrast, CRC tumors exhibiting pMMR along with MSS exhibit a low burden of mutations and low infiltration of CD4 and CD8 immune cells, resulting in evasion of the immune response.

ICIs enhance the recognition and elimination of cancer cells by activating the immune system, resulting in a more potent and sustained anticancer immune response. Understanding the mechanisms of synchronization with the disease pathophysiology is crucial for optimizing the therapeutic potential of ICIs and improving patient outcomes. The main mechanisms of action of ICIs include blockade of the PD-1/PD-L1 pathway and CTLA-4, which regulates T-cell activity and is often upregulated in tumors to evade the immune system (Figure 2)[38]. Indeed, PD-1 and CTLA-4 serve as negative regulators of T-cell activation and exert their biological effects at specific anatomical locations and at various points throughout the lifespan of T-cells[92]. The varied and late-onset autoimmune manifestations observed in Pdcd1-/- mice differ significantly from those in Ctla4-/- animals, highlighting that the PD1 axis governs T-cell biology in a distinct manner compared to CTLA4[92,93].

The PD-1/PD-L1 axis plays a role in autoimmunity by negatively regulating T-cell activation[94]. Functional loss of PD1 protein results in splenomegaly in mice models[94]. Additionally, mouse models lacking the PD-1 gene exhibit cardiac inflammation, leading to dilated cardiomyopathy and accelerated type 1 diabetes mellitus[95,96]. The PD1 axis is crucial for regulating differentiated effectors in T-cells[93,97,98]. Upon binding to PD-L1, PD1 exerts inhibitory intracellular signaling, leading to T-cell exhaustion, and eventually suppressing the immune response[99-101]. In addition to its role in regulating conventional T-cells, PD-L1 on antigen presenting cells (APCs) also plays a role in controlling Treg differentiation and immunosuppressive activity[102]. Tumor cells upregulate PD-L1 surface expression to take advantage of the PD-1/PD-L1 axis and escape immune response.

Anti-PD-1 antibodies such as pembrolizumab and nivolumab are ICIs used in cancer immunotherapy. Anti-PD-1 antibodies not only enhance the activity of cytotoxic T-cells but also affect the overall tumor microenvironment (TME) as well. These antibodies can alter the balance of immune cell populations by reducing the number of immunosuppressive cells such as Tregs and myeloid-derived suppressor cells[103]. This shift contributes to a more favorable immunological milieu for anti-tumor responses[104]. Anti-PD-1 antibodies promote increased production of pro-inflammatory cytokines, such as IFN-γ. These cytokines play key roles in amplifying the anti-tumor immune response by activating other immune cells and enhancing the recognition and elimination of cancer cells. In contrast, anti-PD-L1 antibodies target PD-L1 ligands on cancer cells. By blocking the interaction between PD-L1 and PD-1 on T-cells, these antibodies disrupt a key immune evasion mechanism employed by cancer cells. Anti-PD-1/PD-L1 antibodies help overcome adaptive immune resistance by enabling T-cells to recognize and target cancer cells more effectively. This leads to continuous adaptation of the immune response against evolving tumor cells. Immunotherapeutic responses are often associated with the expression of specific immunological biomarkers[105-107]. For anti-PD-1/PD-L1 therapy, the expression of PD-L1 on tumor cells is a commonly used biomarker. Tumors with high PD-L1 expression may have a higher likelihood of responding to anti-PD-1 antibodies. The presence of TILs is also considered a positive prognostic indicator of immuno-

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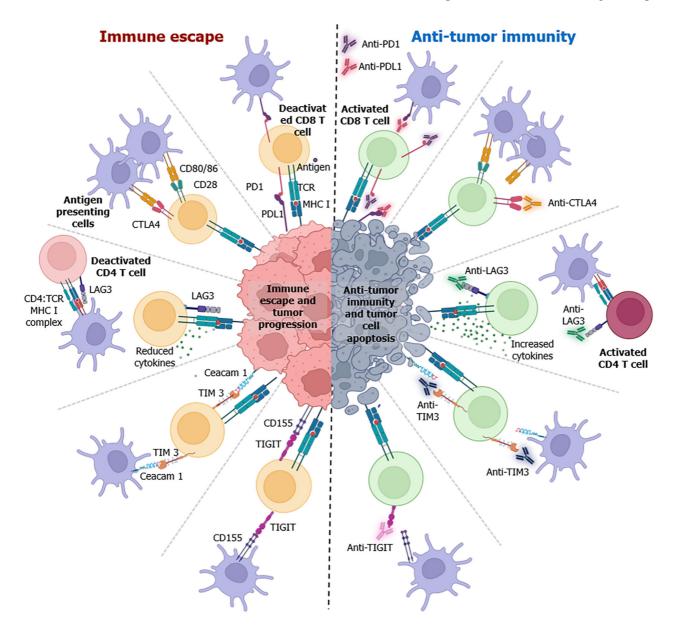


Figure 2 This figure presents a schematic diagram of the intricate interplay between immune checkpoints, immune cells, and malignant cells. It also elucidates the underlying molecular mechanisms employed in immune-checkpoint blockade. In the tumor microenvironment, the interaction of immune checkpoints leads to immune suppression and facilitates tumor progression (depicted on the left side of the diagram). Conversely, the administration of immune checkpoint inhibitions reverses the immune escape mechanism, fostering increased anti-tumor immunity and triggering tumor apoptosis (depicted on the right side of the diagram). FDA: Food and Drug Administration.

therapy response.

CTLA-4, a vital immune checkpoint, exhibits low basal levels in conventional T-cell. However, its expression is significantly induced after antigen activation. Activated T-cells expressing CTLA4 impede the interaction between B7-1 and B7-2 molecules on APCs and CD28, and thereby induce anergy and reduce T-cell activation [108-110]. TCR signaling studies affirm CTLA4's role in inhibiting T-cell activation and proliferation[111-113]. Ctla4-knockout mice were found to develop T-cell mediated autoimmune disease, which is mitigated by treatment with the CTLA4: Fc fusion protein (CTLA4Ig)[114-116]. Notably, CD4+ CD25+ Tregs, which are known for their immunosuppressive function, constitutively express CTLA4 and are necessary for the release of anti-inflammatory cytokines from Tregs[117,118]. These findings confirm that CTLA4 is a T-cell activation inhibitor with potential as a therapeutic agent against cancer [119]. Pre-clinical studies using anti-CTLA-4 antibodies aimed to prevent inhibitory signals, allowing for a more effective CD28 interaction with B7[120]. However, the results were found to depend on tissue specificity and tumor size[119,121]. Additionally, blocking CTLA4 enhances T-cell responses to tumor-associated neoantigens, and a high neoantigen burden predicts a positive response to anti-CTLA4 therapy.

Blocking CTLA-4 with anti-CTLA-4 antibodies promotes a sustained and enhanced T-cell activation. CD28 is a costimulator of T-cell activation that benefits from the increased availability of anti-CTLA-4 antibodies and facilitates enhanced binding to B7. This then amplifies co-stimulatory signals, promoting T-cell proliferation and function[120]. These antibodies also modify the TME by decreasing immunosuppressive cells, such as Tregs, creating a more favorable setting for anti-tumor immune responses [122,123]. Additionally, anti-CTLA-4 antibodies induce antibody-dependent cell-

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mediated cytotoxicity, with immune cells, particularly natural killer cells, recognizing and eliminating target cells marked with therapeutic antibodies[124]. Often used in combination with other ICIs, such as anti-PD-1 or anti-PD-L1 antibodies, this approach targets multiple checkpoints simultaneously, enhancing the overall anti-tumor immune response. Anti-CTLA-4 therapy induces systemic immune activation, affecting not only the tumor site but also distant metastases, contributing to the potential for durable responses and efficacy.

ICIs can also enhance antigen presentation by dendritic cells (DCs), and thus facilitate the priming of T-cells to initiate a robust and targeted immune response against cancer[125,126]. Activated DCs present tumor antigens to T-cells effectively to promote T-cell activation and proliferation. Adaptive immune resistance involves a dynamic interplay between the immune system and cancer cells. Successful ICIs therapy is associated with memory T-cell generation. Memory T-cells contribute to long-term immune memory by enabling the immune system to respond rapidly to cancer cell recurrence. Understanding these immunological nuances provides insights into how ICIs contribute to unleashing and potentiating the ability of the immune system to recognize and eliminate cancer cells. Despite its success in MSI-H CRC, the clinical efficacy of immunotherapy in MSS CRC remains very limited[28]. Recent studies have found that characteristics such as high levels of TILs and expression of immune checkpoint molecules such as PD-L1 may help identify patients with MSS CRC who are more likely to benefit from ICI treatment.

### CLINICAL TRIALS INVOLVING CHECKPOINT INHIBITORS IN CRC

The emergence of checkpoint inhibitors has brought a remarkable shift in our approach to treatment of cancers, including CRC. Notably, these inhibitors have exhibited encouraging treatment outcomes in specific subsets of patients, such as those with MSI-H or dMMR tumors, which are characterized by augmented levels of TILs and heightened susceptibility to immune checkpoint blockade. This section focuses on noteworthy clinical studies on application, efficacy, and potential benefits of ICIs in CRC. In 2014, the Food and Drug Administration (FDA) approved Pembrolizumab, a PD-1 immune checkpoint inhibitor for melanoma treatment[127]. Tumor cells evade the immune system through the PD-1 pathway where PDL1 and PDL2 Ligands on tumor cells binds to the PD-1 receptors on T cells to inactivate T cells. Pembrolizumab binds to these PD-1 receptor and blocks their interaction with PDL1 and PDL2, thereby restoring the immune response [128]. Subsequently, in 2020, the FDA approved this drug for patients with unresectable or metastatic MSI-H or dMMR CRC, based on results from key clinical trials. Phase 2 open-label, multicenter trial (NCT01876511) was conducted to evaluate the safety, efficacy, and tolerability of pembrolizumab in MSI-H-positive patients [129]. Trials have shown no dose-limiting toxicities associated with pembrolizumab, with a promising disease control rate of 80% in patients with MSI-positive CRC, suggesting the potential of Pembrolizumab in CRC treatment. Subsequently, another trial (NCT02460198) postulated the efficacy of pembrolizumab in patients with unresectable tumors who underwent standard chemotherapy[130,131]. The results showed a promising overall progression response rate of 32 to 34 months in both cohorts. This study demonstrated the potential of pembrolizumab as an effective treatment option for patients with dMMR and MSI-H mCRC.

Chemotherapy has been used over the years for the treatment of patients with CRC to shrink tumor volume[132]. In 2015, a phase 3 clinical trial (NCT02563002) was conducted to test pembrolizumab as a first-line treatment in comparison with standard chemotherapy treatment in mCRC patients with MSI-H or dMMR tumors[133,134]. The results showed a significant improvement in the progression-free survival (PFS) rate of 16.5 months in comparison to the standard chemotherapy group at 8.2 months. The trial demonstrated pembrolizumab monotherapy to be superior to standard chemotherapy in terms of PFS and overall response rate (ORR) for patients with MSI-H or dMMR mCRC as a standard care option. The potential efficacy of pembrolizumab in these patients was demonstrated by increased production of neoantigens resulting from an elevated mutational burden. This, in turn, leads to a heightened recognition of tumor cells by cytotoxic T cells, which are primed by blocking the interaction between PD-1 and PD-L1. These results thus led to a paradigm shift in the treatment of this patient population.

The evolution of pembrolizumab has led to the development of more PD-1-targeting drugs whose efficacy and safety profiles were assessed. Nivolumab, another PD-1 monoclonal antibody, was approved in 2017 for use in mCRC treatment [28]. Both drugs exhibited similar modes of action in blocking PD1 and inducing increased CTLs cytotoxicity. However, these two antibodies also exhibited significant structural differences in their binding to PD-1[135]. The epitope region of pembrolizumab displayed a considerably larger overlap with the PD-L1 binding site compared to that of nivolumab. Notably, the binding sites of pembrolizumab and nivolumab on PD-1 showed almost no convergence[135]. A study published in 2017 compared the effectiveness of drugs with comparable effectiveness, which may potentially be interchangeable. The effectiveness of nivolumab has been studied in NCT02060188 MSI-H or dMMR mCRC patients with or without the CTLA-4 inhibitor drug Ipilimumab[136-138]. The treatment showed promising results, with a disease control rate of 80% with nivolumab alone. Combination treatment with a CTLA-4 inhibitor was effective in 51 of the 74 patients who achieved disease control for a minimum of 12 wk. However, further studies are still needed to determine the optimal treatment duration for pembrolizumab and to identify predictive biomarkers of response to immunotherapy in this population. Overall, Nivolumab plus ipilimumab combination therapy is a promising treatment with a better disease control rate and objective response.

Atezolizumab, a monoclonal antibody targeting PD-L1, was approved by the FDA in 2016 for the treatment of nonsmall cell lung cancer tumors[139]. The mechanism of action of Atezolizumab differs from those of pembrolizumab and nivolumab. Instead of binding to PD-1, atezolizumab binds to PD-L1 on tumor cells, and thereby provides a mode of action similar to that of the PD-1 antibody. A clinical trial (NCT02788279) was conducted to evaluate the efficacy of this drug alone and in combination with a MEK inhibitor, cobimetinib, compared to regorafenib (a multi-kinase inhibitor)

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[140]. This combination is used as a therapeutic alternative to a MEK inhibitor that increases T-cell proliferation and CD8+ T cell infiltration, and PD-1 treatment to upregulate the PD-1/PD-L1 interaction, thereby downregulating the immunosuppressive TME. The results of the trial showed an improved PFS rate 1.91 of 2 months after cobimetinib treatment. In conclusion, combination of atezolizumab and cobimetinib improved PFS and ORR compared to regorafenib as a second-line treatment for patients with mCRC, yet did not significantly improve OS. These results underscore the potential of combination therapy and suggest improved investigations of combined immunotherapy drugs as promising targeted treatment approaches for mCRC. Combination therapy with MEK inhibition and PD-L1 blockade led to impressive long-term survival rates. MEK inhibitors act during the post-naïve stage of T-cell differentiation. MEK inhibition counteracts the expression of Nur77, which is associated with exhaustive T cell death induced by antigen-specific CD8<sup>+</sup> T cells, thereby rescuing T cell exhaustion[16].

Another PD-1 monoclonal antibody, dostarlimab, was approved in 2021 and tested in clinical trials (NCT04165772) in patients with MSI-H or dMMR CRC[141]. A high-resolution structure revealed that Dostarlimab binds to the flexible loops of PD-1, including the BC, C'D, and FG loops, in contrast to the binding modes of Pembrolizumab or Nivolumab [94]. The initial findings of the trials were published for 12 patients. Accordingly, all the patients (100%; 95%CI: 74-100) showed a complete clinical response[142]. The response was confirmed using magnetic resonance imaging, which showed no evidence of tumors. At that time, none of the patients had received chemoradiotherapy or undergone surgery. No cases of progression or recurrence were observed during follow-up (6-25 months). The study listed no grade 3 or higher AEs during the trial period. This study thus demonstrated the high sensitivity of dMMR locally advanced rectal cancer to single-agent PD-1 blockade. Despite these promising results, a longer follow-up period and a larger sample size are still needed to assess the duration of the response.

Following the improved success of ICIs in CRC patients with MSI-H or dMMR tumors, researchers have investigated their efficacy in MSS or pMMR CRC patients as well. These data suggested that more than 85% of CRC patients with MSS tumors show resistance to ICIs therapy. A clinical trial (NCT01876511) illustrating the potential of pembrolizumab in MSI-H or dMMR CRC patients has shown no measurable responses in any of the 18 patients with pMMR CRC, as defined by the RECIST criteria[129]. To overcome these limitations, combination treatments have been used to improve the drug responses. A clinical trial was conducted using PD-L1 with a multi kinase inhibitor in patients with MSS or pMMR CRC. Regorafenib targets stromal and oncogenic receptor tyrosine kinases, and shows anti-angiogenic activity due to its dual-target VEGFR2-TIE2 tyrosine kinase inhibition[143]. The NCT04126733 trial conducted between 2019 and 2022 included 94 CRC patients with MSS or pMMR[144]. The results showed an ORR of 7% and overall survival (OS) rate of 11.9%. The relatively reduced success observed in patients with MSS or pMMR CRC indicates the need for further advancement of drug efficacy to provide better outcomes in MSS tumors.

In 2016, a combination trial (NCT01988896) was performed in CRC patients with BRAF/KRAS mutations using cobimetinib and atezolizumab to study OS and PFS in MSS or pMMR CRCs patients[145]. These mutations are rarely identified but are more frequently found in patients with MSS CRC. BRAF and KRAS mutations are mutually exclusive, and BRAF-mt induces aberrant and inappropriate activation of the MAPK/ERK pathway, making it a good candidate for combination therapy with ICIs and kinases or MEK inhibitors[146]. As expected, the combination provided relatively better outcomes in patients with BRAF/KRAS mutations, with an OS rate of 43%. These findings provide compelling evidence that MAPK pathway blockade therapy combined with ICIs is promising for improving treatment efficacy in mCRC patients with MSS/pMMR BRAF mutations.

Recent developments have improved targeted immunotherapies using engineered ICIs to increase the success rate in patients with MSS or pMMR CRC[147]. A phase 1 clinical trial was conducted to categorize the adverse effects and doselimiting toxicity of botensilimab, an Fc-engineered anti-CTLA-4 monoclonal antibody, in patients with MSS CRC. This Fc engineering promotes intratumoral Treg depletion and reduces complement fixation. This modification provides optimized T-cell priming, activation, and memory formation by strengthening antigen-presenting cell/T-cell coengagement. The trial showed an ORR of 22% (95%CI: 12-35) and a disease control rate of 73% (95%CI: 42-75) for patients with non-hepatic disease in refractory CRC. The trial showed the efficacy of the anti-tumor activity in MSS CRC patients with active liver metastatic disease, and a phase 2 trial (NCT05608044) for MSS CRC has begun to study its potential in controlling tumor progression.

The success of ICIs in patients with MSI-H CRC has been hindered by their reduced potential in MSS CRC. The results from clinical trials in patients with MSS CRC undergoing immune checkpoint blockade immunotherapy suggest the need for the development of new pre-clinical mouse models to replicate the microenvironment of human CRC, and potentiate new targeted therapies to improve patient survival.

### LIMITATIONS OF USING IMMUNE CHECKPOINT INHIBITORS FOR IMMUNOTHERAPY

Although ICI therapy holds benefits, patients also often experience autoimmune-like effects known as immune-related adverse events (irAEs). This is likely the result of generalized, non-antigen-specific activation of the immune system following a checkpoint blockade. Inhibition of a naturally occurring central immune checkpoint releases potent immune effector mechanisms that may not adhere to the usual boundaries of immune tolerance towards self-tissues[148]. Human loss-of-function mutations in CTLA4 and its regulatory partner, LPS Responsive Beige-Like Anchor Protein, mimic the immune-related side effects of anti-CTLA4 therapy[149,150]. irAEs have been reported in 15%–90% of patients[57], with severe events requiring intervention being observed in 30% and 15% of patients treated with CTLA4 and PD1 axis inhibitors, respectively[151]. This immune checkpoint inhibitor leads to toxicity in naïve T cells and accumulation of memory T cells in peripheral organs[152,153]. Compared with PD-1, CTLA4 therapy possess with severe autoimmune

complications, as seen in pre-clinical and clinical trials[154].

Colitis is a frequent complication observed in ICIs therapy[155]. Anti-CTLA4 treatment resulted in a potentially higher colitis rate, ranging from 5.7% to 22% of patients, compared to 0.7% to 1.6% with anti-PD-1 agents[156]. The development of ICI-mediated colitis and diarrhea (IMC) may involve cytotoxic CD8+ T cells. Recent analysis of single-cell RNA sequences from patients with IMC revealed an expansion of tissue-resident memory CD8+ T cells into inflammatory populations within the colon tissue, suggesting that the activation or alteration of CD8+ T cell populations could be a potential mechanism for colitis induced by ICIs[157]. Therefore, with the potential use of immune checkpoint blockade, the current research should aim to identify potential predictive markers for organ-specific toxicities caused by immuno-therapy.

### IMMUNE PROFILING OF MSI AND MSS CRC INFLUENCE ICIS SUCCESS

Patients comprising MSI-H or dMMR tumors have a significantly high overall mutation burden, with approximately 12 mutations per million DNA bases. In contrast, pMMR/MSI-L tumors have a relatively reduced tumor burden, with fewer than eight mutations per million DNA bases[158]. This phenomenon is primarily attributed to somatic defects in the function of MMR genes, with the most prevalent mechanism being hypermethylation of the MLH1 promoter, which serves as a prognostic marker. In MSI, frameshift mutations in protein-coding sequences can create diverse peptides that serve as potential necepitopes, which are recognized as foreign by the immune system. Mutant peptides form complexes with major histocompatibility complex class I molecules and act as foreign neoantigens that initiate immune cell priming and infiltration. Within the TME, tumor-associated macrophages play a crucial role in influencing tumor growth and progression. Recent study has demonstrated a frameshift mutation in the TGFβRII producing an immunogenic peptide called p538[159]. This peptide is present in over 90% of tumors with dMMR, indicating its broad relevance in the field. These tumors exhibit robust infiltration by immune cells, particularly CD8+ TILs, Th1 CD4+ TILs, and macrophages[73]. Furthermore, the microenvironment of these tumors is notably enriched in type I interferons, which distinguishes them from other CRCs types.

Approximately 15% of all CRCs exhibit MSI-H or dMMR characteristics[160]. Patients with MSI-H or dMMR tumors before ICIs therapy continue to have a poor prognosis. Cancers show significantly upregulated expression of PD1, PDL1, and CTLA4, rendering them potentially susceptible to ICIs[87]. In contrast, MSS or pMMR tumors lacking neoantigens are characterized by reduced T cell infiltration and elevated levels of immunosuppressive ligands. These characteristics offer insights into the disagreement between MSI-H or dMMR and MSS or pMMR CRCs in ICIs responses and could potentially serve as prognostic biomarkers for patient selection. As shown in previously described clinical trials, immunotherapy as a neoadjuvant approach has not shown any clinical benefit in patients with MSS or pMMR CRC, including individuals with mCRC.

Contrastingly, the MSS CRC, majorly referred to as an "immune cold" cancer type, are predisposed by various molecular factors contributing to the underlying resistance to immunotherapy[161]. MSS-CRC is characterized by larger chromosomal aberrations that mark the phenotype of MSS-CRC, resulting in a lower tumor mutation burden and reduced neoantigen configuration. This framework partially elucidates the disparate clinical responses to ICIs observed in these CRC subgroups. The MSS CRC TME hosts more tumor-associated macrophages, which have been associated with poor prognosis in most studies. Notably, a pioneer study identified that increased  $\beta$ -catenin activation (a downstream effect of APC mutation) resulted in reduced infiltration of CD8+ and CD103+ DCs, orchestrating an immune suppressive environment *via* T-cell exclusion[16]. The APC protein is mutated in more than 70% and 20% of MSS and MSI-H CRCs, respectively, driving the distinct oncogenesis mechanisms and subsequent "immune hot" and "immune cold" TME[162]. These differences are reflected in clinical trials with ICIs, where MSS tumors have very low response rates compared to MSI tumors (Table 1).

### IMPLICATION OF PRE-CLINICAL MOUSE MODELS OF CRC IN DRUG DEVELOPMENT AND IMMUNOTHERAPY

Mouse- and cell-based models have been used for decades to investigate the molecular origins of CRC, and more recently, to identify drug and immune responses in specific CRC types (Figure 1). These efforts have yielded tremendous improvement in our basic understanding of the disease and TME. However, despite recent approvals, the majority of patients continue to have limited immunotherapy options. A recurring challenge highlighted in the literature is the absence of a mouse model that precisely replicates the progression of human CRC from adenoma and adenocarcinoma to metastasis, including changes in the microenvironment. Initial mouse models lacked significant penetrance of the metastatic phenotype, often forming tumors in the small intestine rather than in the colon, unless induced by laparoscopy [163,164]. In 2013, the National Institute of Health formally concluded the Mouse Models in Human Cancer Consortium, leading researchers to explore alternative models, such as patient-derived xenografts and patient-derived organoids, to study the disease. The significance of the TME in metastasis remains a focus of current research, and the potential of checkpoint inhibitors and other immunological and inhibitor therapies are being explored. However, an ideal model still does not exist, highlighting the importance of an immunocompetent autochthonous model. Notably, single-cell RNA sequencing of mouse tumors to understand the mechanisms underlying immune-modulating therapies could help draw more impactful conclusions[165]. Despite these limitations, the diversity of the methods employed by researchers with

Table 1 List of clinical trials with immune check point inhibition therapy					
Trial identified (number of patients)	Treatment groups	Patients enrolled	Primary and secondary outcomes		
Checkpoint inhibit	or: Pembrolizumab				
NCT01876511 ( <i>n</i> = 113)	Pembrolizumab	Cohort A: MSI positive (pMMR) CRC	Cohort A: ORR: 54.0% (95%CI: 37.0–69.0); PFS: 70% (95%CI: 57–86); OS: NA (95%CI: 151.86-NA)		
		Cohort B: MSI negative (dMMR) CRC	Cohort B: ORR: 0% (95%CI: 0.0–14.0); PFS: 16% (95%CI: 6–41); OS: 36.71 (95%CI: 21.29-69.43)		
NCT02460198 (n = 124)	Pembrolizumab	mCRC with dMMR or MSI-H status Cohort A: Participants must have received prior treatment with standard therapies	Cohort A: ORR: 32.8 (95%CI: 21.3 to 46.0); PFS: 2.3 (95%CI: 2.1–8.1); OS: 31.4 (95%CI: 21.4–58)		
		Cohort B: Participants must have undergone at least one line of systemic standard of care therapy	Cohort B: ORR: 34.9 (95%CI: 23.3–48.0); PFS: 4.1 (95%CI: 2.1–18.9); OS: 47 (19.2–NA)		
NCT02563002 ( <i>n</i> = 307)	Arm A: Pembrolizumab	mCRC with high MSI-H or dMMR	Arm A: ORR: 45.1% (95%CI: 37.1-53.3); PFS: 16.5 (95%CI: 5.4-38.1); OS: NA (95%CI: 49.2-NA)		
	Arm B: mFOLFOX6/FOLFIRI/Bevacizumab/Cetuximab/Pembrol- izumab		Arm B: ORR: 33.1% (95%CI: 25.8–41.1); PFS: 8.2 (95%CI: 6.1–10.2); OS: 27.6 (95%CI: 27.6–NA)		
Checkpoint inhibit	or: Nivolumab + Regorafenib				
NCT04126733 ( <i>n</i> = 94)	Regorafenib and Nivolumab	Patients with pMMR or MSS CRC	ORR: 7% (95%CI: 2.4–15.9); PFS: 1.8 (95%CI: 1.8–2.4); OS: 11.9 (95%CI: 7.0 to NA)		
Checkpoint inhibit	Checkpoint inhibitor: Nivolumab + Ipilimumab				
NCT02060188 ( <i>n</i> = 119)	Arm A: Nivolumab	MSI-H or dMMR mCRC	Arm A: No results posted		
- 119)	Arm B: Nivolumab + Ipilimumab		Arm B: ORR: 55% (95%CI: 45.2%-63.8%); PFS: 71% (95%CI: 61.4 to 78.7); OS: 85% (95%CI: 77.0 to 90.2)		
	Arm C: Cobimetinib + Nivolumab + Ipilimumab		Arm C: No results posted		
	Arm D: Nivolumab + Daratumumab		Arm D: No results posted		
	Arm E: Nivolumab + BMS-986016		Arm E: No results posted		
Checkpoint inhibit	Checkpoint inhibitor: Atezolizumab				
NCT02788279 ( <i>n</i> = 363)	Arm A: Atezolizumab	Patients with mCRC (MSI or MSS status unknown)	Arm A: PFS: 1.94 (95%CI: 1.91 to 2.10); OS: 7.10 (95%CI: 6.05-10.05)		
	Arm B: Cobimetinib + Atezolizumab		Arm B: PFS: 1.91 (95%CI: 1.87 to 1.97); OS: 8.87 (95%CI: 7.00-10.61)		
	Arm C: Regorafenib		Arm C: PFS: 2 (95%CI: 1.87-3.61); OS: 8.51 (95%CI: 6.41-10.71)		
NCT01988896 ( <i>n</i> = 84)	Atezolizumab + Cobimetinib	Patients having BRAF/KRAS mutation in mCRC	ORR: 8% (7/84) (6 patients: MSS, 1 patient: MSI)		
NCT01633970 ( $n = 10$ )	Arm A: Atezolizumab + Bevacizumab	No results posted	No results posted		
= 10)	Arm B: Atezolizumab + Bevacizumab + FOLFOX				
	Arm C: Atezolizumab + Carboplatin + Paclitaxel				

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	Arm D: Atezolizumab + Carboplatin + Pemetrexed				
	Arm E: Atezolizumab + Carboplatin + Nab-paclitaxel				
	Arm F: Atezolizumab + Nab-paclitaxel				
Checkpoint inhibitor: Durvalumab + Tremelimumab					
NCT03122509 ( <i>n</i> = 25)	Arm A: Durvalumab + Tremelimumab + Radiotherapy	Metastatic Colorectal Cancer (MSI or MSS status unknown)	Arm A: ORR: 8%; Stable response: 12%; Progressive disease: 76%		
	Arm B: Durvalumab + Tremelimumab + Ablation		Arm B: No participants enrolled		
Checkpoint inhibitor: Dostarlimab					
NCT04165772 ( <i>n</i> = 200)	Arm A: Dostarlimab	Patients with dMMR rectal adenocarcinoma	Arm A: Complete response: 12 patients (100%; 95%CI: 74–100)		
	Arm B: Dostarlimab + Capecitabine or 5-FU + IMRT		Arm B: No participants enrolled		

CRC: Colorectal cancer; mCRC: Metastatic colorectal cancer; MSI: Microsatellite-instability; MSI-H: Microsatellite instability-high; pMMR: Proficient mismatch repair; dMMR: Deficient mismatch repair; MSS: Microsatellite-stable; 5-FU: 5-fluorouracil IMRT: Intensity modulated radiotherapy.

mice, the adaptability of the system, and the deductive formation of CRC images from diverse models remain impressive.

Various transplantation techniques have been introduced over the years to replicate complex TME in mouse models. The subcutaneous injection model has been widely used [166], yet it has limitations, such as creating an ectopic environment and lacking accuracy in mimicking the metastatic spread of cancer [167]. The orthotopic transplantation model has emerged as a promising alternative to address these issues. This involves a precise injection into the intestinal region, such as the cecal wall, colon, or rectum. Among these, the orthotopic CRC model using cecal wall injections has been widely adopted. However, it is essential to recognize that even this model has constraints as it does not faithfully replicate the anatomical location where tumors typically occur in humans and exhibits a microenvironment distinct from the rest of the colon[168].

Genetically engineered mouse models (GEMMs) are based on genetic engineering techniques, particularly the Cre recombinase lox-P system, to simulate tumorigenesis by modifying the structure of the genome [169]. GEMMs, including those incorporating mutations in genes, such as APC, KRAS, p53, and MSH2, provide valuable insights into the molecular mechanisms underlying CRC and play a pivotal role in the study of CRC development and therapeutic strategies. APC mutations activate Wnt/b-catenin, causing increased b-catenin levels and tumor development[170,171], restricting tumor growth within the intestines, and mimicking human CRC. In addition, compared with previous single-mutation GEMMs for CRC, transgenic mice established via combined APC/KRAS mutations have been shown to represent CRC initiation, progression, and metastasis more accurately into nearby tissues[163,172]. Advantages include specificity in mirroring human CRC growth, representation of TMEs, and the ability to visualize the CRC through colonoscopy. Immunotherapeutic studies using transgenic mice have revealed promising avenues for targeted treatments. However, limitations such as extended metastatic development time and limited colon-specific models warrant further refinement for comprehensive CRC research [173]. As a result, transgenic mice are often ineffective in representing the later stages of tumor development owing to highly variable metastasis [166]. Additionally, there are few current transgenic mouse models that lead to specific CRC development in the colon, as the majority of pre-existing models lead to CRC development within the small intestine or other nearby tissues, contributing to the development of familial cancers rather than specific GEMM mutation-derived CRC[174].

The observation that tumors in mice have a narrower phenotype than human tumors suggests that the mice themselves need to be subtyped before drawing comparisons with human subtypes. This recognition is crucial, especially considering that murine backgrounds are often congenic (with the same genetic makeup) and artificially altered for experimental purposes. In summary, the critical importance of selecting appropriate preclinical models in CRC research requires better understanding. Mouse models are indispensable tools for discovering effective therapeutic interventions. The evaluation of various transplantation methods, with particular emphasis on the orthotopic CRC model via cecal wall injection, provides a nuanced understanding of their utility while acknowledging the inherent limitations associated with this approach. This recognition is vital for refining experimental design and interpretation to better translate findings from preclinical studies to human clinical scenarios.

### FUTURE DIRECTIONS

Overall, immunotherapy, particularly with the use of ICIs like pembrolizumab and nivolumab has demonstrated significant clinical benefit in MSI-H CRC, while its efficacy in MSS CRC remains limited [91,130,133,138]. However, even in MSI-H tumors, the upregulation of immune checkpoint proteins, the presence of other immunosuppressive mechanisms within the TME, and the heterogeneity of MSI-H CRC tumors within the primary tumor and across



metastatic sites can contribute to varied responses to immunotherapy. Understanding these factors and further research on the mechanisms of immune resistance in patients with MSI-H CRC are essential to improve the outcomes of immunotherapy in this patient population.

In the field of ICI therapy for CRC, various research avenues to enhance treatment efficacy and broaden its scope of application are being actively pursued. Therefore, there is a need to identify response biomarkers and devise novel treatment approaches to address these challenges. One area of focus is the investigation of combination therapies in which ICIs are used in conjunction with chemotherapy, targeted therapies, and other immunotherapies. Similarly, combining immunotherapy with targeted therapy directed against specific signaling pathways, such as the MAPK pathway, may also improve treatment outcomes [136-138,140,175]. Here, the goal is to enhance the response rates and improve patient survival by leveraging the synergistic effects of different treatment modalities. Another important research avenue is the discovery of reliable biomarkers that can accurately predict patient response to ICIs. Although PD-L1 expression is currently used as a biomarker for some cancer types [176,177], its predictive value for CRC is limited [129, 178]. For example, tumors with elevated PD-L1 expression may be more responsive to anti-PD-1/PD-L1 therapy, whereas tumors with low PD-L1 expression may require combination therapy to achieve a response. Similarly, tumors with specific genetic mutations such as BRAF V600E may require targeted therapy in addition to immunotherapy to achieve a response. Mutations in genes such as JAK1, JAK2, and B2M may contribute to treatment resistance [179-181]. Truncating mutations in B2M affect antigen presentation, and can lead to pembrolizumab resistance. Evidence indicates that a high somatic mutational load and neoantigen density are associated with an improved response to immune checkpoint blockade in various cancers. This is attributed to the increased presence of mutation-associated neoantigens, which contribute to greater T cell diversity [182]. Moreover, there are also ongoing research efforts on the development of novel immunotherapeutic agents such as bispecific antibodies, chimeric antigen receptor (CAR) T-cells, and vaccines, which may provide new treatment options for CRC patients[183-188]. Efforts are underway to identify additional biomarkers that assist patient selection and treatment decisions. These targeted therapies, when combined with combination therapies, have shown considerable potential in enhancing treatment efficacy and overcoming drug resistance. By simultaneously targeting different pathways implicated in CRC progression, these approaches aim to maximize therapeutic benefits while minimizing adverse effects. Future research should focus on identifying optimal drug combinations, elucidating synergistic interactions, and refining treatment regimens to improve patient response.

A recent study revealed that immune cells form multicellular hubs in CRC samples that are spatially organized and functionally distinct from the surrounding immune cells[85]. The findings indicated that these immune hubs are composed of different cell types, including T-cells, B-cells, and myeloid cells, and are enriched in specific functional pathways related to the immune response and cell-cell communication. Researchers have also observed that the distribution and composition of immune hubs vary between patients and may be influenced by factors such as tumor stage and treatment history. Furthermore, the findings also demonstrated that the presence of immune hubs was associated with better clinical outcomes in CRC patients, suggesting a crucial role in the immune response to cancer. The authors proposed that targeting immune hubs could be a promising strategy for enhancing the efficacy of immunotherapy in CRC[85].

Additionally, there is a growing interest in exploring the use of ICIs in the early stages of CRC, such as adjuvant therapy following surgery. Early detection and intervention are pivotal for improving CRC outcomes. Emerging technologies, such as liquid biopsies and advanced imaging modalities, hold promise for the detection of CRC at earlier stages when treatment options are more effective [189-191]. Additionally, minimally invasive surgical techniques and organpreserving approaches offer less invasive alternatives for managing early stage CRC, reducing morbidity, and improving the quality of life of patients[192].

Current research focuses on understanding the intricate mechanisms underlying drug resistance, including genetic mutations, TME interactions, and adaptive signaling pathways. Strategies for overcoming resistance include developing combination therapies that target multiple pathways, repurposing existing drugs, and developing novel agents to evade resistance mechanisms. Precision medicine approaches such as tumor molecular profiling and real-time monitoring facilitate the early detection of resistance mechanisms, allowing prompt adjustments to treatment strategies[193,194]. Furthermore, biomarker research in CRC is rapidly evolving with the aim of identifying molecular signatures crucial for diagnosis, prognosis, and treatment decisions. Biomarkers, such as mutations in genes such as KRAS and BRAF, not only influence tumor behavior, but also affect responses to targeted therapies, notably anti-EGFR antibodies[24]. Additionally, MSI status serves not only as a guide for immunotherapy but also as a valuable prognostic indicator as well. Liquid biopsies offer a noninvasive method to analyze circulating tumor DNA and to monitor disease progression and treatment responses[189]. Epigenetic alterations, such as DNA methylation patterns and microRNA expression profiles, are promising diagnostic and prognostic markers[24,195]. Traditional biomarkers, such as carcinoembryonic antigen, provide insights into tumor burden and treatment response, while gene expression signatures, such as Oncotype DX and ColoPrint, offer predictive value for treatment outcomes and recurrence risk assessment[196]. Integrating these diverse biomarkers into clinical practice can help personalize treatment strategies, optimize patient management, and ultimately enhance the survival outcomes for CRC patients. However, drug resistance remains a significant challenge, compromising the efficacy of chemotherapy, targeted therapies, and immunotherapy [132]. CRC cells develop resistance to chemotherapeutic drugs such as fluoropyrimidines and oxaliplatin through mechanisms such as altered drug metabolism and enhanced drug efflux [197,198]. Similarly, targeted therapies may encounter resistance due to secondary mutations or activation of alternative signaling pathways. Understanding and overcoming these mechanisms are crucial for advancing CRC treatment and improving patient prognosis. Finally, new targets and agents beyond the PD-1/PD-L1 pathway are being investigated. Promising preclinical data to have led to clinical trials of molecules targeting additional T cell checkpoint inhibitors, such as TIM3, LAG3, and TIGIT, in various advanced malignancies, including CRC (Figure 2)[94, 199-202]. In addition to immune checkpoint blockade, molecules that enhance T-cell differentiation, survival, and prolif-

eration are being investigated as standalone treatments or in combination with checkpoint inhibitors. These molecules, including CD27, OX40, 4-1BB, and others, act as antibody agonists for the costimulatory group within the TNF receptor superfamily[203-209].

Personalized medicine has revolutionized CRC treatment by tailoring interventions to individual patient characteristics, including genetic and molecular factors. Over the years, personalized medicine has gained traction and treatment approaches have been tailored based on individual patient characteristics. This involves integrating tumor genetic profiling, immune profiling, and other personalized medicine strategies to identify the most effective treatment options for each patient [194,210,211]. Comprehensive genomic profiling and clinical data integration enable the identification of actionable targets and personalized treatment regimens. Artificial intelligence enhances data interpretation and improves the accuracy of treatment response prediction. Liquid biopsies provide a noninvasive method for monitoring disease progression and identifying therapeutic targets. Personalized medicine integrates liquid biopsy-based monitoring into treatment management, allowing for real-time therapy adjustments. This approach promises to optimize treatment strategies and improve the clinical outcomes for CRC patients with CRC. Advancements in biomarker research coupled with efforts to overcome drug resistance and implement personalized medicine offer a multifaceted approach to CRC management that holds great promise for enhancing patient care and outcomes in the future.

Moreover, additional novel strategies for CRC treatment such as mRNA vaccines, TILs therapy, CAR-T therapy, oncolytic virus therapy, bispecific T-cell engagers, and combination strategies aim to improve treatment outcomes by specifically considering metastatic location and TME regulation. Novel agents and therapeutic strategies are being developed to expand the range of options available for immune modulation of this disease. Accordingly, in addition to the development of new biomarkers and therapeutic strategies, there is also a need for better pre-clinical mouse models that can potentially or closely replicate the human CRC microenvironment, thus providing a better opportunity to unmask novel approaches for treatment. As the field of immunotherapy evolves, these directions hold great promise for advancing immune checkpoint inhibitor therapy and other immunotherapeutic approaches for CRC, ultimately resulting in improved patient outcomes.

### CONCLUSION

Immunotherapy has significantly reshaped the CRC treatment landscape, particularly for patients with MSI-H or dMMR tumors. Key accomplishments include the FDA approval of PD-1 inhibitors, such as pembrolizumab and nivolumab, for these patient subsets. Pembrolizumab has demonstrated promising outcomes both as a monotherapy and in combination with chemotherapy, surpassing standard treatments in terms of PFS and ORR[133,134]. Furthermore, combination therapies have shown promise, such as the use of nivolumab with ipilimumab (a CTLA-4 inhibitor), which has demonstrated improved disease control rates [136-138]. Additionally, atezolizumab in combination with Cobimetinib has shown enhanced PFS rates in second-line treatments, although further studies are needed to establish its effects on OS [145]. A recent study established Dostarlimab as a drug with 100% effectiveness against MSI-H or dMMR CRC tumors. Despite the preliminary success of immuno-oncology, challenges persist for CRC treatment, particularly those pertaining to the extension of immunotherapeutic benefits to MSS or pMMR tumors, which commonly exhibit resistance to ICIs. In this regard, irAEs associated with ICIs should be managed effectively, which requires identification of predictive biomarkers and the development of mitigation strategies. Combination therapies, as exemplified by the synergistic effects observed with nivolumab and ipilimumab, require further investigation to optimize their performance and to identify their underlying mechanisms. Exploring novel therapeutic targets beyond immune checkpoint blockade, including targeted therapies and engineered immunotherapies, holds promise for overcoming resistance mechanisms. Addressing these challenges requires interdisciplinary collaboration, ongoing preclinical and clinical research, and rigorous validation through well-controlled trials. By overcoming these obstacles, advancements in CRC treatment can be realized, leading to improved clinical outcomes and enhanced quality of life in affected patients. In conclusion, immunotherapy has revolutionized CRC treatment, resulting in improved outcomes and survival rates in MSI-H or dMMR patients. However, challenges persist in extending these benefits to patients with MSS or pMMR and in the management of irAEs. Future research should focus on optimizing combination therapies, identifying predictive biomarkers, and mitigating treatment-related toxicities to realize the full potential of immunotherapy in CRC management.

### FOOTNOTES

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Author contributions: Sharma S and Singh N contributed equally to the study's conception, design, and wrote the manuscript as co-first authors; Turk AA offered insights into the current clinical aspects of colorectal cancer patients; Wan I, Guttikonda A, and Dong JL assisted in drafting the manuscript; Zhang X and Opyrchal M supervised the review framework as senior authors, offered valuable feedback, and contributed to the writing of the manuscript.

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### REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of 1 Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Li K, Luo H, Huang L, Zhu X. Microsatellite instability: a review of what the oncologist should know. Cancer Cell Int 2020; 20: 16 [PMID: 2 31956294 DOI: 10.1186/s12935-019-1091-8]
- Siegel RL, Wagle NS, Cercek A, Smith RA, Jemal A. Colorectal cancer statistics, 2023. CA Cancer J Clin 2023; 73: 233-254 [PMID: 3 36856579 DOI: 10.3322/caac.21772]
- Sinicrope FA. Increasing Incidence of Early-Onset Colorectal Cancer. N Engl J Med 2022; 386: 1547-1558 [PMID: 35443109 DOI: 4 10.1056/NEJMra2200869
- 5 Ewing I, Hurley JJ, Josephides E, Millar A. The molecular genetics of colorectal cancer. Frontline Gastroenterol 2014; 5: 26-30 [PMID: 24416503 DOI: 10.1136/flgastro-2013-100329]
- 6 Islami F, Goding Sauer A, Miller KD, Siegel RL, Fedewa SA, Jacobs EJ, McCullough ML, Patel AV, Ma J, Soerjomataram I, Flanders WD, Brawley OW, Gapstur SM, Jemal A. Proportion and number of cancer cases and deaths attributable to potentially modifiable risk factors in the United States. CA Cancer J Clin 2018; 68: 31-54 [PMID: 29160902 DOI: 10.3322/caac.21440]
- Dekker E, Tanis PJ, Vleugels JLA, Kasi PM, Wallace MB. Colorectal cancer. Lancet 2019; 394: 1467-1480 [PMID: 31631858 DOI: 7 10.1016/S0140-6736(19)32319-0
- 8 Pearlman R, Frankel WL, Swanson B, Zhao W, Yilmaz A, Miller K, Bacher J, Bigley C, Nelsen L, Goodfellow PJ, Goldberg RM, Paskett E, Shields PG, Freudenheim JL, Stanich PP, Lattimer I, Arnold M, Liyanarachchi S, Kalady M, Heald B, Greenwood C, Paquette I, Prues M, Draper DJ, Lindeman C, Kuebler JP, Reynolds K, Brell JM, Shaper AA, Mahesh S, Buie N, Weeman K, Shine K, Haut M, Edwards J, Bastola S, Wickham K, Khanduja KS, Zacks R, Pritchard CC, Shirts BH, Jacobson A, Allen B, de la Chapelle A, Hampel H; Ohio Colorectal Cancer Prevention Initiative Study Group. Prevalence and Spectrum of Germline Cancer Susceptibility Gene Mutations Among Patients With Early-Onset Colorectal Cancer. JAMA Oncol 2017; 3: 464-471 [PMID: 27978560 DOI: 10.1001/jamaoncol.2016.5194]
- 9 Barnetson RA, Tenesa A, Farrington SM, Nicholl ID, Cetnarskyj R, Porteous ME, Campbell H, Dunlop MG. Identification and survival of carriers of mutations in DNA mismatch-repair genes in colon cancer. N Engl J Med 2006; 354: 2751-2763 [PMID: 16807412 DOI: 10.1056/NEJMoa053493
- Hampel H, Frankel WL, Martin E, Arnold M, Khanduja K, Kuebler P, Clendenning M, Sotamaa K, Prior T, Westman JA, Panescu J, Fix D, 10 Lockman J, LaJeunesse J, Comeras I, de la Chapelle A. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. J Clin Oncol 2008; 26: 5783-5788 [PMID: 18809606 DOI: 10.1200/JCO.2008.17.5950]
- 11 Hampel H, Frankel WL, Martin E, Arnold M, Khanduja K, Kuebler P, Nakagawa H, Sotamaa K, Prior TW, Westman J, Panescu J, Fix D, Lockman J, Comeras I, de la Chapelle A. Screening for the Lynch syndrome (hereditary nonpolyposis colorectal cancer). N Engl J Med 2005; 352: 1851-1860 [PMID: 15872200 DOI: 10.1056/NEJMoa043146]
- Yurgelun MB, Allen B, Kaldate RR, Bowles KR, Judkins T, Kaushik P, Roa BB, Wenstrup RJ, Hartman AR, Syngal S. Identification of a 12 Variety of Mutations in Cancer Predisposition Genes in Patients With Suspected Lynch Syndrome. Gastroenterology 2015; 149: 604-13.e20 [PMID: 25980754 DOI: 10.1053/j.gastro.2015.05.006]
- Sinicrope FA. Lynch Syndrome-Associated Colorectal Cancer. N Engl J Med 2018; 379: 764-773 [PMID: 30134129 DOI: 13 10.1056/NEJMcp1714533]
- 14 Koveitypour Z, Panahi F, Vakilian M, Peymani M, Seyed Forootan F, Nasr Esfahani MH, Ghaedi K. Signaling pathways involved in colorectal cancer progression. Cell Biosci 2019; 9: 97 [PMID: 31827763 DOI: 10.1186/s13578-019-0361-4]
- Arends MJ. Pathways of colorectal carcinogenesis. Appl Immunohistochem Mol Morphol 2013; 21: 97-102 [PMID: 23417071 DOI: 15 10.1097/PAI.0b013e31827ea79e]
- Van der Jeught K, Xu HC, Li YJ, Lu XB, Ji G. Drug resistance and new therapies in colorectal cancer. World J Gastroenterol 2018; 24: 3834-16 3848 [PMID: 30228778 DOI: 10.3748/wjg.v24.i34.3834]
- National Cancer Institute. All Cancer Sites Combined Recent Trends in SEER Age-Adjusted Incidence Rates, 2000-2020. [cited 23 August 17 2023]. Available from: https://seer.cancer.gov/statistics-network/explorer/application.html
- Longley DB, Johnston PG. Molecular mechanisms of drug resistance. J Pathol 2005; 205: 275-292 [PMID: 15641020 DOI: 18 10.1002/path.1706
- Salonga D, Danenberg KD, Johnson M, Metzger R, Groshen S, Tsao-Wei DD, Lenz HJ, Leichman CG, Leichman L, Diasio RB, Danenberg 19 PV. Colorectal tumors responding to 5-fluorouracil have low gene expression levels of dihydropyrimidine dehydrogenase, thymidylate synthase, and thymidine phosphorylase. Clin Cancer Res 2000; 6: 1322-1327 [PMID: 10778957]
- 20 Källberg J, Harrison A, March V, Berzina S, Nemazanyy I, Kepp O, Kroemer G, Mouillet-Richard S, Laurent-Puig P, Taly V, Xiao W.



Intratumor heterogeneity and cell secretome promote chemotherapy resistance and progression of colorectal cancer. Cell Death Dis 2023; 14: 306 [PMID: 37142595 DOI: 10.1038/s41419-023-05806-z]

- 21 Molinari C, Marisi G, Passardi A, Matteucci L, De Maio G, Ulivi P. Heterogeneity in Colorectal Cancer: A Challenge for Personalized Medicine? Int J Mol Sci 2018; 19 [PMID: 30477151 DOI: 10.3390/ijms19123733]
- Golshani G, Zhang Y. Advances in immunotherapy for colorectal cancer: a review. Therap Adv Gastroenterol 2020; 13: 1756284820917527 22 [PMID: 32536977 DOI: 10.1177/1756284820917527]
- Péraudeau E, Renoux B, Emambux S, Poinot P, Châtre R, Thoreau F, Riss Yaw B, Tougeron D, Clarhaut J, Papot S. Combination of Targeted 23 Therapies for Colorectal Cancer Treatment. Mol Pharm 2023; 20: 4537-4545 [PMID: 37579031 DOI: 10.1021/acs.molpharmaceut.3c00224]
- Alese OB, Wu C, Chapin WJ, Ulanja MB, Zheng-Lin B, Amankwah M, Eads J. Update on Emerging Therapies for Advanced Colorectal 24 Cancer. Am Soc Clin Oncol Educ Book 2023; 43: e389574 [PMID: 37155942 DOI: 10.1200/EDBK\_389574]
- Samstein RM, Lee CH, Shoushtari AN, Hellmann MD, Shen R, Janjigian YY, Barron DA, Zehir A, Jordan EJ, Omuro A, Kaley TJ, Kendall 25 SM, Motzer RJ, Hakimi AA, Voss MH, Russo P, Rosenberg J, Iyer G, Bochner BH, Bajorin DF, Al-Ahmadie HA, Chaft JE, Rudin CM, Riely GJ, Baxi S, Ho AL, Wong RJ, Pfister DG, Wolchok JD, Barker CA, Gutin PH, Brennan CW, Tabar V, Mellinghoff IK, DeAngelis LM, Ariyan CE, Lee N, Tap WD, Gounder MM, D'Angelo SP, Saltz L, Stadler ZK, Scher HI, Baselga J, Razavi P, Klebanoff CA, Yaeger R, Segal NH, Ku GY, DeMatteo RP, Ladanyi M, Rizvi NA, Berger MF, Riaz N, Solit DB, Chan TA, Morris LGT. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. Nat Genet 2019; 51: 202-206 [PMID: 30643254 DOI: 10.1038/s41588-018-0312-8]
- 26 Zheng M. Tumor mutation burden for predicting immune checkpoint blockade response: the more, the better. J Immunother Cancer 2022; 10 [PMID: 35101940 DOI: 10.1136/jitc-2021-003087]
- 27 Ganesh K. Optimizing immunotherapy for colorectal cancer. Nat Rev Gastroenterol Hepatol 2022; 19: 93-94 [PMID: 34907331 DOI: 10.1038/s41575-021-00569-4]
- 28 Ganesh K, Stadler ZK, Cercek A, Mendelsohn RB, Shia J, Segal NH, Diaz LA Jr. Immunotherapy in colorectal cancer: rationale, challenges and potential. Nat Rev Gastroenterol Hepatol 2019; 16: 361-375 [PMID: 30886395 DOI: 10.1038/s41575-019-0126-x]
- Brahmer JR, Drake CG, Wollner I, Powderly JD, Picus J, Sharfman WH, Stankevich E, Pons A, Salay TM, McMiller TL, Gilson MM, Wang 29 C, Selby M, Taube JM, Anders R, Chen L, Korman AJ, Pardoll DM, Lowy I, Topalian SL. Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: safety, clinical activity, pharmacodynamics, and immunologic correlates. J Clin Oncol 2010; 28: 3167-3175 [PMID: 20516446 DOI: 10.1200/JCO.2009.26.7609]
- Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K, Pitot HC, Hamid O, Bhatia 30 S, Martins R, Eaton K, Chen S, Salay TM, Alaparthy S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll DM, Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012; 366: 2455-2465 [PMID: 22658128 DOI: 10.1056/NEJMoa1200694]
- 31 Chung KY, Gore I, Fong L, Venook A, Beck SB, Dorazio P, Criscitiello PJ, Healey DI, Huang B, Gomez-Navarro J, Saltz LB. Phase II study of the anti-cytotoxic T-lymphocyte-associated antigen 4 monoclonal antibody, tremelimumab, in patients with refractory metastatic colorectal cancer. J Clin Oncol 2010; 28: 3485-3490 [PMID: 20498386 DOI: 10.1200/JCO.2010.28.3994]
- Lipson EJ, Sharfman WH, Drake CG, Wollner I, Taube JM, Anders RA, Xu H, Yao S, Pons A, Chen L, Pardoll DM, Brahmer JR, Topalian 32 SL. Durable cancer regression off-treatment and effective reinduction therapy with an anti-PD-1 antibody. Clin Cancer Res 2013; 19: 462-468 [PMID: 23169436 DOI: 10.1158/1078-0432.CCR-12-2625]
- 33 Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Carvajal RD, Sosman JA, Atkins MB, Leming PD, Spigel DR, Antonia SJ, Horn L, Drake CG, Pardoll DM, Chen L, Sharfman WH, Anders RA, Taube JM, McMiller TL, Xu H, Korman AJ, Jure-Kunkel M, Agrawal S, McDonald D, Kollia GD, Gupta A, Wigginton JM, Sznol M. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 2012; 366: 2443-2454 [PMID: 22658127 DOI: 10.1056/NEJMoa1200690]
- Janjigian YY, Sanchez-Vega F, Jonsson P, Chatila WK, Hechtman JF, Ku GY, Riches JC, Tuvy Y, Kundra R, Bouvier N, Vakiani E, Gao J, 34 Heins ZJ, Gross BE, Kelsen DP, Zhang L, Strong VE, Schattner M, Gerdes H, Coit DG, Bains M, Stadler ZK, Rusch VW, Jones DR, Molena D, Shia J, Robson ME, Capanu M, Middha S, Zehir A, Hyman DM, Scaltriti M, Ladanyi M, Rosen N, Ilson DH, Berger MF, Tang L, Taylor BS, Solit DB, Schultz N. Genetic Predictors of Response to Systemic Therapy in Esophagogastric Cancer. Cancer Discov 2018; 8: 49-58 [PMID: 29122777 DOI: 10.1158/2159-8290.CD-17-0787]
- Bai R, Lv Z, Xu D, Cui J. Predictive biomarkers for cancer immunotherapy with immune checkpoint inhibitors. Biomark Res 2020; 8: 34 35 [PMID: 32864131 DOI: 10.1186/s40364-020-00209-0]
- Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. Science 2018; 359: 1350-1355 [PMID: 29567705 DOI: 36 10.1126/science.aar4060]
- Galluzzi L, Chan TA, Kroemer G, Wolchok JD, López-Soto A. The hallmarks of successful anticancer immunotherapy. Sci Transl Med 2018; 37 **10** [PMID: 30232229 DOI: 10.1126/scitranslmed.aat7807]
- Waldman AD, Fritz JM, Lenardo MJ. A guide to cancer immunotherapy: from T cell basic science to clinical practice. Nat Rev Immunol 2020; 38 **20**: 651-668 [PMID: 32433532 DOI: 10.1038/s41577-020-0306-5]
- Sumransub N, Vantanasiri K, Prakash A, Lou E. Advances and new frontiers for immunotherapy in colorectal cancer: Setting the stage for 39 neoadjuvant success? Mol Ther Oncolytics 2021; 22: 1-12 [PMID: 34307839 DOI: 10.1016/j.omto.2021.05.001]
- Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, Lu S, Kemberling H, Wilt C, Luber BS, Wong F, Azad NS, Rucki AA, 40 Laheru D, Donehower R, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Greten TF, Duffy AG, Ciombor KK, Eyring AD, Lam BH, Joe A, Kang SP, Holdhoff M, Danilova L, Cope L, Meyer C, Zhou S, Goldberg RM, Armstrong DK, Bever KM, Fader AN, Taube J, Housseau F, Spetzler D, Xiao N, Pardoll DM, Papadopoulos N, Kinzler KW, Eshleman JR, Vogelstein B, Anders RA, Diaz LA Jr. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science 2017; 357: 409-413 [PMID: 28596308 DOI: 10.1126/science.aan6733]
- Boland CR, Goel A. Microsatellite instability in colorectal cancer. Gastroenterology 2010; 138: 2073-2087.e3 [PMID: 20420947 DOI: 41 10.1053/j.gastro.2009.12.064]
- Vilar E, Gruber SB. Microsatellite instability in colorectal cancer-the stable evidence. Nat Rev Clin Oncol 2010; 7: 153-162 [PMID: 20142816 42 DOI: 10.1038/nrclinonc.2009.237]
- 43 Jin Z, Sanhueza CT, Johnson B, Nagorney DM, Larson DW, Mara KC, Harmsen WC, Smyrk TC, Grothey A, Hubbard JM. Outcome of Mismatch Repair-Deficient Metastatic Colorectal Cancer: The Mayo Clinic Experience. Oncologist 2018; 23: 1083-1091 [PMID: 29674439 DOI: 10.1634/theoncologist.2017-0289]
- Li SKH, Martin A. Mismatch Repair and Colon Cancer: Mechanisms and Therapies Explored. Trends Mol Med 2016; 22: 274-289 [PMID: 44 26970951 DOI: 10.1016/j.molmed.2016.02.003]



- Battaglin F, Naseem M, Lenz HJ, Salem ME. Microsatellite instability in colorectal cancer: overview of its clinical significance and novel 45 perspectives. Clin Adv Hematol Oncol 2018; 16: 735-745 [PMID: 30543589]
- Guinney J, Dienstmann R, Wang X, de Reyniès A, Schlicker A, Soneson C, Marisa L, Roepman P, Nyamundanda G, Angelino P, Bot BM, 46 Morris JS, Simon IM, Gerster S, Fessler E, De Sousa E Melo F, Missiaglia E, Ramay H, Barras D, Homicsko K, Maru D, Manyam GC, Broom B, Boige V, Perez-Villamil B, Laderas T, Salazar R, Gray JW, Hanahan D, Tabernero J, Bernards R, Friend SH, Laurent-Puig P, Medema JP, Sadanandam A, Wessels L, Delorenzi M, Kopetz S, Vermeulen L, Tejpar S. The consensus molecular subtypes of colorectal cancer. Nat Med 2015; 21: 1350-1356 [PMID: 26457759 DOI: 10.1038/nm.3967]
- Kuipers EJ, Grady WM, Lieberman D, Seufferlein T, Sung JJ, Boelens PG, van de Velde CJ, Watanabe T. Colorectal cancer. Nat Rev Dis 47 Primers 2015; 1: 15065 [PMID: 27189416 DOI: 10.1038/nrdp.2015.65]
- Labianca R, Nordlinger B, Beretta GD, Mosconi S, Mandalà M, Cervantes A, Arnold D; ESMO Guidelines Working Group. Early colon 48 cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2013; 24 Suppl 6: vi64-vi72 [PMID: 24078664 DOI: 10.1093/annonc/mdt354]
- 49 Van Cutsem E, Cervantes A, Nordlinger B, Arnold D; ESMO Guidelines Working Group. Metastatic colorectal cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2014; 25 Suppl 3: iii1-iii9 [PMID: 25190710 DOI: 10.1093/annonc/mdu260]
- Brown KGM, Solomon MJ, Mahon K, O'Shannassy S. Management of colorectal cancer. BMJ 2019; 366: 14561 [PMID: 31439545 DOI: 50 10.1136/bmj.14561]
- 51 Benson AB, Venook AP, Al-Hawary MM, Arain MA, Chen YJ, Ciombor KK, Cohen S, Cooper HS, Deming D, Farkas L, Garrido-Laguna I, Grem JL, Gunn A, Hecht JR, Hoffe S, Hubbard J, Hunt S, Johung KL, Kirilcuk N, Krishnamurthi S, Messersmith WA, Meyerhardt J, Miller ED, Mulcahy MF, Nurkin S, Overman MJ, Parikh A, Patel H, Pedersen K, Saltz L, Schneider C, Shibata D, Skibber JM, Sofocleous CT, Stoffel EM, Stotsky-Himelfarb E, Willett CG, Gregory KM, Gurski LA. Colon Cancer, Version 2.2021, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw 2021; 19: 329-359 [PMID: 33724754 DOI: 10.6004/jnccn.2021.0012]
- 52 Schirrmacher V. From chemotherapy to biological therapy: A review of novel concepts to reduce the side effects of systemic cancer treatment (Review). Int J Oncol 2019; 54: 407-419 [PMID: 30570109 DOI: 10.3892/ijo.2018.4661]
- 53 Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunogenic cell death in cancer and infectious disease. Nat Rev Immunol 2017; 17: 97-111 [PMID: 27748397 DOI: 10.1038/nri.2016.107]
- Rodriguez-Ruiz ME, Vitale I, Harrington KJ, Melero I, Galluzzi L. Immunological impact of cell death signaling driven by radiation on the 54 tumor microenvironment. Nat Immunol 2020; 21: 120-134 [PMID: 31873291 DOI: 10.1038/s41590-019-0561-4]
- 55 Bracci L, Schiavoni G, Sistigu A, Belardelli F. Immune-based mechanisms of cytotoxic chemotherapy: implications for the design of novel and rationale-based combined treatments against cancer. Cell Death Differ 2014; 21: 15-25 [PMID: 23787994 DOI: 10.1038/cdd.2013.67]
- Zhai J, Gu X, Liu Y, Hu Y, Jiang Y, Zhang Z. Chemotherapeutic and targeted drugs-induced immunogenic cell death in cancer models and 56 antitumor therapy: An update review. Front Pharmacol 2023; 14: 1152934 [PMID: 37153795 DOI: 10.3389/fphar.2023.1152934]
- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van 57 den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A, Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med 2010; 363: 711-723 [PMID: 20525992 DOI: 10.1056/NEJMoa1003466]
- Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, Tykodi SS, Sosman JA, Procopio G, Plimack ER, Castellano D, 58 Choueiri TK, Gurney H, Donskov F, Bono P, Wagstaff J, Gauler TC, Ueda T, Tomita Y, Schutz FA, Kollmannsberger C, Larkin J, Ravaud A, Simon JS, Xu LA, Waxman IM, Sharma P; CheckMate 025 Investigators. Nivolumab versus Everolimus in Advanced Renal-Cell Carcinoma. N Engl J Med 2015; 373: 1803-1813 [PMID: 26406148 DOI: 10.1056/NEJMoa1510665]
- Ribas A. Releasing the Brakes on Cancer Immunotherapy. N Engl J Med 2015; 373: 1490-1492 [PMID: 26348216 DOI: 59 10.1056/NEJMp1510079]
- Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, Diéras V, Hegg R, Im SA, Shaw Wright G, Henschel V, Molinero L, 60 Chui SY, Funke R, Husain A, Winer EP, Loi S, Emens LA; IMpassion130 Trial Investigators. Atezolizumab and Nab-Paclitaxel in Advanced Triple-Negative Breast Cancer. N Engl J Med 2018; 379: 2108-2121 [PMID: 30345906 DOI: 10.1056/NEJMoa1809615]
- Gandhi L, Rodríguez-Abreu D, Gadgeel S, Esteban E, Felip E, De Angelis F, Domine M, Clingan P, Hochmair MJ, Powell SF, Cheng SY, 61 Bischoff HG, Peled N, Grossi F, Jennens RR, Reck M, Hui R, Garon EB, Boyer M, Rubio-Viqueira B, Novello S, Kurata T, Gray JE, Vida J, Wei Z, Yang J, Raftopoulos H, Pietanza MC, Garassino MC; KEYNOTE-189 Investigators. Pembrolizumab plus Chemotherapy in Metastatic Non-Small-Cell Lung Cancer. N Engl J Med 2018; 378: 2078-2092 [PMID: 29658856 DOI: 10.1056/NEJMoa1801005]
- 62 Forde PM, Chaft JE, Pardoll DM. Neoadjuvant PD-1 Blockade in Resectable Lung Cancer. N Engl J Med 2018; 379: e14 [PMID: 30157404 DOI: 10.1056/NEJMc1808251]
- Franke AJ, Skelton WP, Starr JS, Parekh H, Lee JJ, Overman MJ, Allegra C, George TJ. Immunotherapy for Colorectal Cancer: A Review of 63 Current and Novel Therapeutic Approaches. J Natl Cancer Inst 2019; 111: 1131-1141 [PMID: 31322663 DOI: 10.1093/jnci/djz093]
- Sharma P, Allison JP. The future of immune checkpoint therapy. Science 2015; 348: 56-61 [PMID: 25838373 DOI: 10.1126/science.aa8172] 64
- Suzuki E, Kapoor V, Jassar AS, Kaiser LR, Albelda SM. Gemcitabine selectively eliminates splenic Gr-1+/CD11b+ myeloid suppressor cells 65 in tumor-bearing animals and enhances antitumor immune activity. Clin Cancer Res 2005; 11: 6713-6721 [PMID: 16166452 DOI: 10.1158/1078-0432.CCR-05-0883]
- Vincent J, Mignot G, Chalmin F, Ladoire S, Bruchard M, Chevriaux A, Martin F, Apetoh L, Rébé C, Ghiringhelli F. 5-Fluorouracil selectively 66 kills tumor-associated myeloid-derived suppressor cells resulting in enhanced T cell-dependent antitumor immunity. Cancer Res 2010; 70: 3052-3061 [PMID: 20388795 DOI: 10.1158/0008-5472.CAN-09-3690]
- Zitvogel L, Kepp O, Kroemer G. Immune parameters affecting the efficacy of chemotherapeutic regimens. Nat Rev Clin Oncol 2011; 8: 151-67 160 [PMID: 21364688 DOI: 10.1038/nrclinonc.2010.223]
- Kwon ED, Hurwitz AA, Foster BA, Madias C, Feldhaus AL, Greenberg NM, Burg MB, Allison JP. Manipulation of T cell costimulatory and 68 inhibitory signals for immunotherapy of prostate cancer. Proc Natl Acad Sci U S A 1997; 94: 8099-8103 [PMID: 9223321 DOI: 10.1073/pnas.94.15.8099]
- Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. Science 1996; 271: 1734-1736 [PMID: 69 8596936 DOI: 10.1126/science.271.5256.1734]
- Sahin IH, Akce M, Alese O, Shaib W, Lesinski GB, El-Rayes B, Wu C. Immune checkpoint inhibitors for the treatment of MSI-H/MMR-D 70 colorectal cancer and a perspective on resistance mechanisms. Br J Cancer 2019; 121: 809-818 [PMID: 31607751 DOI:



### 10.1038/s41416-019-0599-y]

- 71 Aristin Revilla S, Kranenburg O, Coffer PJ. Colorectal Cancer-Infiltrating Regulatory T Cells: Functional Heterogeneity, Metabolic Adaptation, and Therapeutic Targeting. Front Immunol 2022; 13: 903564 [PMID: 35874729 DOI: 10.3389/fimmu.2022.903564]
- Olguín JE, Medina-Andrade I, Rodríguez T, Rodríguez-Sosa M, Terrazas LI. Relevance of Regulatory T Cells during Colorectal Cancer 72 Development. Cancers (Basel) 2020; 12 [PMID: 32674255 DOI: 10.3390/cancers12071888]
- Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, 73 Bruneval P, Cugnenc PH, Trajanoski Z, Fridman WH, Pagès F. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 2006; 313: 1960-1964 [PMID: 17008531 DOI: 10.1126/science.1129139]
- Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, Fitz LJ, Malenkovich N, Okazaki T, Byrne MC, Horton HF, Fouser L, 74 Carter L, Ling V, Bowman MR, Carreno BM, Collins M, Wood CR, Honjo T. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. J Exp Med 2000; 192: 1027-1034 [PMID: 11015443 DOI: 10.1084/jem.192.7.1027
- Buchbinder EI, Desai A. CTLA-4 and PD-1 Pathways: Similarities, Differences, and Implications of Their Inhibition. Am J Clin Oncol 2016; 75 **39**: 98-106 [PMID: 26558876 DOI: 10.1097/COC.00000000000239]
- Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 2012; 12: 252-264 [PMID: 22437870 DOI: 76 10.1038/nrc32391
- 77 Vilgelm AE, Johnson DB, Richmond A. Combinatorial approach to cancer immunotherapy: strength in numbers. J Leukoc Biol 2016; 100: 275-290 [PMID: 27256570 DOI: 10.1189/jlb.5RI0116-013RR]
- Haslam A, Gill J, Prasad V. Estimation of the Percentage of US Patients With Cancer Who Are Eligible for Immune Checkpoint Inhibitor 78 Drugs. JAMA Netw Open 2020; 3: e200423 [PMID: 32150268 DOI: 10.1001/jamanetworkopen.2020.0423]
- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Sosman JA, Atkins MB, Leming PD, Spigel DR, 79 Antonia SJ, Drilon A, Wolchok JD, Carvajal RD, McHenry MB, Hosein F, Harbison CT, Grosso JF, Sznol M. Five-Year Survival and Correlates Among Patients With Advanced Melanoma, Renal Cell Carcinoma, or Non-Small Cell Lung Cancer Treated With Nivolumab. JAMA Oncol 2019; 5: 1411-1420 [PMID: 31343665 DOI: 10.1001/jamaoncol.2019.2187]
- 80 van der Stok EP, Spaander MCW, Grünhagen DJ, Verhoef C, Kuipers EJ. Surveillance after curative treatment for colorectal cancer. Nat Rev Clin Oncol 2017; 14: 297-315 [PMID: 27995949 DOI: 10.1038/nrclinonc.2016.199]
- Sánchez-Gundín J, Fernández-Carballido AM, Martínez-Valdivieso L, Barreda-Hernández D, Torres-Suárez AI. New Trends in the 81 Therapeutic Approach to Metastatic Colorectal Cancer. Int J Med Sci 2018; 15: 659-665 [PMID: 29910669 DOI: 10.7150/ijms.24453]
- Wolf AMD, Fontham ETH, Church TR, Flowers CR, Guerra CE, LaMonte SJ, Etzioni R, McKenna MT, Oeffinger KC, Shih YT, Walter LC, 82 Andrews KS, Brawley OW, Brooks D, Fedewa SA, Manassaram-Baptiste D, Siegel RL, Wender RC, Smith RA. Colorectal cancer screening for average-risk adults: 2018 guideline update from the American Cancer Society. CA Cancer J Clin 2018; 68: 250-281 [PMID: 29846947 DOI: 10.3322/caac.21457]
- 83 Keum N, Giovannucci E. Global burden of colorectal cancer: emerging trends, risk factors and prevention strategies. Nat Rev Gastroenterol Hepatol 2019; 16: 713-732 [PMID: 31455888 DOI: 10.1038/s41575-019-0189-8]
- Lee YT, Tan YJ, Oon CE. Molecular targeted therapy: Treating cancer with specificity. Eur J Pharmacol 2018; 834: 188-196 [PMID: 84 30031797 DOI: 10.1016/j.ejphar.2018.07.034]
- Pelka K, Hofree M, Chen JH, Sarkizova S, Pirl JD, Jorgji V, Bejnood A, Dionne D, Ge WH, Xu KH, Chao SX, Zollinger DR, Lieb DJ, Reeves 85 JW, Fuhrman CA, Hoang ML, Delorey T, Nguyen LT, Waldman J, Klapholz M, Wakiro I, Cohen O, Albers J, Smillie CS, Cuoco MS, Wu J, Su MJ, Yeung J, Vijaykumar B, Magnuson AM, Asinovski N, Moll T, Goder-Reiser MN, Applebaum AS, Brais LK, DelloStritto LK, Denning SL, Phillips ST, Hill EK, Meehan JK, Frederick DT, Sharova T, Kanodia A, Todres EZ, Jané-Valbuena J, Biton M, Izar B, Lambden CD, Clancy TE, Bleday R, Melnitchouk N, Irani J, Kunitake H, Berger DL, Srivastava A, Hornick JL, Ogino S, Rotem A, Vigneau S, Johnson BE, Corcoran RB, Sharpe AH, Kuchroo VK, Ng K, Giannakis M, Nieman LT, Boland GM, Aguirre AJ, Anderson AC, Rozenblatt-Rosen O, Regev A, Hacohen N. Spatially organized multicellular immune hubs in human colorectal cancer. Cell 2021; 184: 4734-4752.e20 [PMID: 34450029 DOI: 10.1016/j.cell.2021.08.003]
- Giannakis M, Mu XJ, Shukla SA, Qian ZR, Cohen O, Nishihara R, Bahl S, Cao Y, Amin-Mansour A, Yamauchi M, Sukawa Y, Stewart C, 86 Rosenberg M, Mima K, Inamura K, Nosho K, Nowak JA, Lawrence MS, Giovannucci EL, Chan AT, Ng K, Meyerhardt JA, Van Allen EM, Getz G, Gabriel SB, Lander ES, Wu CJ, Fuchs CS, Ogino S, Garraway LA. Genomic Correlates of Immune-Cell Infiltrates in Colorectal Carcinoma. Cell Rep 2016; 15: 857-865 [PMID: 27149842 DOI: 10.1016/j.celrep.2016.03.075]
- Llosa NJ, Cruise M, Tam A, Wicks EC, Hechenbleikner EM, Taube JM, Blosser RL, Fan H, Wang H, Luber BS, Zhang M, Papadopoulos N, 87 Kinzler KW, Vogelstein B, Sears CL, Anders RA, Pardoll DM, Housseau F. The vigorous immune microenvironment of microsatellite instable colon cancer is balanced by multiple counter-inhibitory checkpoints. Cancer Discov 2015; 5: 43-51 [PMID: 25358689 DOI: 10.1158/2159-8290.CD-14-0863]
- 88 Dolcetti R, Viel A, Doglioni C, Russo A, Guidoboni M, Capozzi E, Vecchiato N, Macrì E, Fornasarig M, Boiocchi M. High prevalence of activated intraepithelial cytotoxic T lymphocytes and increased neoplastic cell apoptosis in colorectal carcinomas with microsatellite instability. Am J Pathol 1999; 154: 1805-1813 [PMID: 10362805 DOI: 10.1016/S0002-9440(10)65436-3]
- 89 Nagorsen D, Voigt S, Berg E, Stein H, Thiel E, Loddenkemper C. Tumor-infiltrating macrophages and dendritic cells in human colorectal cancer: relation to local regulatory T cells, systemic T-cell response against tumor-associated antigens and survival. J Transl Med 2007; 5: 62 [PMID: 18047662 DOI: 10.1186/1479-5876-5-62]
- Smyrk TC, Watson P, Kaul K, Lynch HT. Tumor-infiltrating lymphocytes are a marker for microsatellite instability in colorectal carcinoma. 90 Cancer 2001; 91: 2417-2422 [PMID: 11413533]
- 91 André T, Shiu KK, Kim TW, Jensen BV, Jensen LH, Punt C, Smith D, Garcia-Carbonero R, Benavides M, Gibbs P, de la Fouchardiere C, Rivera F, Elez E, Bendell J, Le DT, Yoshino T, Van Cutsem E, Yang P, Farooqui MZH, Marinello P, Diaz LA Jr; KEYNOTE-177 Investigators. Pembrolizumab in Microsatellite-Instability-High Advanced Colorectal Cancer. N Engl J Med 2020; 383: 2207-2218 [PMID: 33264544 DOI: 10.1056/NEJMoa2017699]
- Fife BT, Bluestone JA. Control of peripheral T-cell tolerance and autoimmunity via the CTLA-4 and PD-1 pathways. Immunol Rev 2008; 224: 92 166-182 [PMID: 18759926 DOI: 10.1111/j.1600-065X.2008.00662.x]
- Parry RV, Chemnitz JM, Frauwirth KA, Lanfranco AR, Braunstein I, Kobayashi SV, Linsley PS, Thompson CB, Riley JL. CTLA-4 and PD-1 93 receptors inhibit T-cell activation by distinct mechanisms. Mol Cell Biol 2005; 25: 9543-9553 [PMID: 16227604 DOI: 10.1128/MCB.25.21.9543-9553.2005



- Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: Co-inhibitory Receptors with Specialized Functions in Immune Regulation. 94 Immunity 2016; 44: 989-1004 [PMID: 27192565 DOI: 10.1016/j.immuni.2016.05.001]
- Nishimura H, Okazaki T, Tanaka Y, Nakatani K, Hara M, Matsumori A, Sasayama S, Mizoguchi A, Hiai H, Minato N, Honjo T. Autoimmune 95 dilated cardiomyopathy in PD-1 receptor-deficient mice. Science 2001; 291: 319-322 [PMID: 11209085 DOI: 10.1126/science.291.5502.319]
- 96 Wang J, Yoshida T, Nakaki F, Hiai H, Okazaki T, Honjo T. Establishment of NOD-Pdcd1-/- mice as an efficient animal model of type I diabetes. Proc Natl Acad Sci U S A 2005; 102: 11823-11828 [PMID: 16087865 DOI: 10.1073/pnas.0505497102]
- Kopetz S, Grothey A, Yaeger R, Van Cutsem E, Desai J, Yoshino T, Wasan H, Ciardiello F, Loupakis F, Hong YS, Steeghs N, Guren TK, 97 Arkenau HT, Garcia-Alfonso P, Pfeiffer P, Orlov S, Lonardi S, Elez E, Kim TW, Schellens JHM, Guo C, Krishnan A, Dekervel J, Morris V, Calvo Ferrandiz A, Tarpgaard LS, Braun M, Gollerkeri A, Keir C, Maharry K, Pickard M, Christy-Bittel J, Anderson L, Sandor V, Tabernero J. Encorafenib, Binimetinib, and Cetuximab in BRAF V600E-Mutated Colorectal Cancer. N Engl J Med 2019; 381: 1632-1643 [PMID: 31566309 DOI: 10.1056/NEJMoa1908075]
- 98 Nishimura H, Minato N, Nakano T, Honjo T. Immunological studies on PD-1 deficient mice: implication of PD-1 as a negative regulator for B cell responses. Int Immunol 1998; 10: 1563-1572 [PMID: 9796923 DOI: 10.1093/intimm/10.10.1563]
- 99 Barber DL, Wherry EJ, Masopust D, Zhu B, Allison JP, Sharpe AH, Freeman GJ, Ahmed R. Restoring function in exhausted CD8 T cells during chronic viral infection. Nature 2006; 439: 682-687 [PMID: 16382236 DOI: 10.1038/nature04444]
- Blank C, Brown I, Peterson AC, Spiotto M, Iwai Y, Honjo T, Gajewski TF. PD-L1/B7H-1 inhibits the effector phase of tumor rejection by T 100 cell receptor (TCR) transgenic CD8+ T cells. Cancer Res 2004; 64: 1140-1145 [PMID: 14871849 DOI: 10.1158/0008-5472.can-03-3259]
- Wei SC, Levine JH, Cogdill AP, Zhao Y, Anang NAS, Andrews MC, Sharma P, Wang J, Wargo JA, Pe'er D, Allison JP. Distinct Cellular 101 Mechanisms Underlie Anti-CTLA-4 and Anti-PD-1 Checkpoint Blockade. Cell 2017; 170: 1120-1133.e17 [PMID: 28803728 DOI: 10.1016/j.cell.2017.07.024]
- Francisco LM, Salinas VH, Brown KE, Vanguri VK, Freeman GJ, Kuchroo VK, Sharpe AH. PD-L1 regulates the development, maintenance, and function of induced regulatory T cells. J Exp Med 2009; 206: 3015-3029 [PMID: 20008522 DOI: 10.1084/jem.20090847]
- Hou A, Hou K, Huang Q, Lei Y, Chen W. Targeting Myeloid-Derived Suppressor Cell, a Promising Strategy to Overcome Resistance to 103 Immune Checkpoint Inhibitors. Front Immunol 2020; 11: 783 [PMID: 32508809 DOI: 10.3389/fimmu.2020.00783]
- Lindau D, Gielen P, Kroesen M, Wesseling P, Adema GJ. The immunosuppressive tumour network: myeloid-derived suppressor cells, 104 regulatory T cells and natural killer T cells. Immunology 2013; 138: 105-115 [PMID: 23216602 DOI: 10.1111/imm.12036]
- Hamanishi J, Mandai M, Iwasaki M, Okazaki T, Tanaka Y, Yamaguchi K, Higuchi T, Yagi H, Takakura K, Minato N, Honjo T, Fujii S. 105 Programmed cell death 1 ligand 1 and tumor-infiltrating CD8+ T lymphocytes are prognostic factors of human ovarian cancer. Proc Natl Acad Sci U S A 2007; 104: 3360-3365 [PMID: 17360651 DOI: 10.1073/pnas.0611533104]
- 106 Thompson RH, Dong H, Lohse CM, Leibovich BC, Blute ML, Cheville JC, Kwon ED. PD-1 is expressed by tumor-infiltrating immune cells and is associated with poor outcome for patients with renal cell carcinoma. Clin Cancer Res 2007; 13: 1757-1761 [PMID: 17363529 DOI: 10.1158/1078-0432.CCR-06-2599]
- Thompson RH, Kuntz SM, Leibovich BC, Dong H, Lohse CM, Webster WS, Sengupta S, Frank I, Parker AS, Zincke H, Blute ML, Sebo TJ, 107 Cheville JC, Kwon ED. Tumor B7-H1 is associated with poor prognosis in renal cell carcinoma patients with long-term follow-up. Cancer Res 2006; 66: 3381-3385 [PMID: 16585157 DOI: 10.1158/0008-5472.CAN-05-4303]
- 108 Linsley PS, Brady W, Urnes M, Grosmaire LS, Damle NK, Ledbetter JA. CTLA-4 is a second receptor for the B cell activation antigen B7. J Exp Med 1991; 174: 561-569 [PMID: 1714933 DOI: 10.1084/jem.174.3.561]
- Linsley PS, Greene JL, Brady W, Bajorath J, Ledbetter JA, Peach R. Human B7-1 (CD80) and B7-2 (CD86) bind with similar avidities but 109 distinct kinetics to CD28 and CTLA-4 receptors. Immunity 1994; 1: 793-801 [PMID: 7534620 DOI: 10.1016/s1074-7613(94)80021-9]
- Pentcheva-Hoang T, Egen JG, Wojnoonski K, Allison JP. B7-1 and B7-2 selectively recruit CTLA-4 and CD28 to the immunological 110 synapse. Immunity 2004; 21: 401-413 [PMID: 15357951 DOI: 10.1016/j.immuni.2004.06.017]
- Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. J Exp Med 1995; 182: 459-465 111 [PMID: 7543139 DOI: 10.1084/jem.182.2.459]
- Krummel MF, Allison JP. CTLA-4 engagement inhibits IL-2 accumulation and cell cycle progression upon activation of resting T cells. J Exp 112 Med 1996; 183: 2533-2540 [PMID: 8676074 DOI: 10.1084/jem.183.6.2533]
- 113 Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, Thompson CB, Bluestone JA. CTLA-4 can function as a negative regulator of T cell activation. Immunity 1994; 1: 405-413 [PMID: 7882171 DOI: 10.1016/1074-7613(94)90071-x]
- Waterhouse P, Penninger JM, Timms E, Wakeham A, Shahinian A, Lee KP, Thompson CB, Griesser H, Mak TW. Lymphoproliferative 114 disorders with early lethality in mice deficient in Ctla-4. Science 1995; 270: 985-988 [PMID: 7481803 DOI: 10.1126/science.270.5238.985]
- Mandelbrot DA, McAdam AJ, Sharpe AH. B7-1 or B7-2 is required to produce the lymphoproliferative phenotype in mice lacking cytotoxic T 115 lymphocyte-associated antigen 4 (CTLA-4). J Exp Med 1999; 189: 435-440 [PMID: 9892625 DOI: 10.1084/jem.189.2.435]
- Tivol EA, Boyd SD, McKeon S, Borriello F, Nickerson P, Strom TB, Sharpe AH. CTLA4Ig prevents lymphoproliferation and fatal multiorgan 116 tissue destruction in CTLA-4-deficient mice. J Immunol 1997; 158: 5091-5094 [PMID: 9164923]
- Tai X, Van Laethem F, Pobezinsky L, Guinter T, Sharrow SO, Adams A, Granger L, Kruhlak M, Lindsten T, Thompson CB, Feigenbaum L, 117 Singer A. Basis of CTLA-4 function in regulatory and conventional CD4(+) T cells. Blood 2012; 119: 5155-5163 [PMID: 22403258 DOI: 10.1182/blood-2011-11-388918
- Takahashi T, Tagami T, Yamazaki S, Uede T, Shimizu J, Sakaguchi N, Mak TW, Sakaguchi S. Immunologic self-tolerance maintained by 118 CD25(+)CD4(+) regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. J Exp Med 2000; 192: 303-310 [PMID: 10899917 DOI: 10.1084/jem.192.2.303]
- Grosso JF, Jure-Kunkel MN. CTLA-4 blockade in tumor models: an overview of preclinical and translational research. Cancer Immun 2013; 119 13: 5 [PMID: 23390376]
- Intlekofer AM, Thompson CB. At the bench: preclinical rationale for CTLA-4 and PD-1 blockade as cancer immunotherapy. J Leukoc Biol 120 2013; 94: 25-39 [PMID: 23625198 DOI: 10.1189/jlb.1212621]
- Yang YF, Zou JP, Mu J, Wijesuriya R, Ono S, Walunas T, Bluestone J, Fujiwara H, Hamaoka T. Enhanced induction of antitumor T-cell 121 responses by cytotoxic T lymphocyte-associated molecule-4 blockade: the effect is manifested only at the restricted tumor-bearing stages. Cancer Res 1997; 57: 4036-4041 [PMID: 9307290]
- Peggs KS, Quezada SA, Chambers CA, Korman AJ, Allison JP. Blockade of CTLA-4 on both effector and regulatory T cell compartments 122 contributes to the antitumor activity of anti-CTLA-4 antibodies. J Exp Med 2009; 206: 1717-1725 [PMID: 19581407 DOI:



Sharma S et al. Unlocking CRC treatment: Immune checkpoint insights

10.1084/jem.20082492]

- 123 Sharma N, Vacher J, Allison JP. TLR1/2 ligand enhances antitumor efficacy of CTLA-4 blockade by increasing intratumoral Treg depletion. Proc Natl Acad Sci U S A 2019; 116: 10453-10462 [PMID: 31076558 DOI: 10.1073/pnas.1819004116]
- Jie HB, Schuler PJ, Lee SC, Srivastava RM, Argiris A, Ferrone S, Whiteside TL, Ferris RL. CTLA-4+ Regulatory T Cells Increased in 124 Cetuximab-Treated Head and Neck Cancer Patients Suppress NK Cell Cytotoxicity and Correlate with Poor Prognosis. Cancer Res 2015; 75: 2200-2210 [PMID: 25832655 DOI: 10.1158/0008-5472.CAN-14-2788]
- Hou TZ, Qureshi OS, Wang CJ, Baker J, Young SP, Walker LS, Sansom DM. A transendocytosis model of CTLA-4 function predicts its 125 suppressive behavior on regulatory T cells. J Immunol 2015; 194: 2148-2159 [PMID: 25632005 DOI: 10.4049/jimmunol.1401876]
- Qureshi OS, Zheng Y, Nakamura K, Attridge K, Manzotti C, Schmidt EM, Baker J, Jeffery LE, Kaur S, Briggs Z, Hou TZ, Futter CE, 126 Anderson G, Walker LS, Sansom DM. Trans-endocytosis of CD80 and CD86: a molecular basis for the cell-extrinsic function of CTLA-4. Science 2011; 332: 600-603 [PMID: 21474713 DOI: 10.1126/science.1202947]
- 127 Raedler LA. Keytruda (Pembrolizumab): First PD-1 Inhibitor Approved for Previously Treated Unresectable or Metastatic Melanoma. Am Health Drug Benefits 2015; 8: 96-100 [PMID: 26629272]
- Ghosh C, Luong G, Sun Y. A snapshot of the PD-1/PD-L1 pathway. J Cancer 2021; 12: 2735-2746 [PMID: 33854633 DOI: 128 10.7150/jca.57334]
- Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, Skora AD, Luber BS, Azad NS, Laheru D, Biedrzycki B, Donehower RC, 129 Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Duffy SM, Goldberg RM, de la Chapelle A, Koshiji M, Bhaijee F, Huebner T, Hruban RH, Wood LD, Cuka N, Pardoll DM, Papadopoulos N, Kinzler KW, Zhou S, Cornish TC, Taube JM, Anders RA, Eshleman JR, Vogelstein B, Diaz LA Jr. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N Engl J Med 2015; 372: 2509-2520 [PMID: 26028255 DOI: 10.1056/NEJMoa1500596]
- Le DT, Kim TW, Van Cutsem E, Geva R, Jäger D, Hara H, Burge M, O'Neil B, Kavan P, Yoshino T, Guimbaud R, Taniguchi H, Elez E, Al-130 Batran SE, Boland PM, Crocenzi T, Atreya CE, Cui Y, Dai T, Marinello P, Diaz LA Jr, André T. Phase II Open-Label Study of Pembrolizumab in Treatment-Refractory, Microsatellite Instability-High/Mismatch Repair-Deficient Metastatic Colorectal Cancer: KEYNOTE-164. J Clin Oncol 2020; 38: 11-19 [PMID: 31725351 DOI: 10.1200/JCO.19.02107]
- van Vugt MJH, Stone JA, De Greef RHJMM, Snyder ES, Lipka L, Turner DC, Chain A, Lala M, Li M, Robey SH, Kondic AG, De Alwis D, 131 Mayawala K, Jain L, Freshwater T. Immunogenicity of pembrolizumab in patients with advanced tumors. J Immunother Cancer 2019; 7: 212 [PMID: 31395089 DOI: 10.1186/s40425-019-0663-4]
- Hammond WA, Swaika A, Mody K. Pharmacologic resistance in colorectal cancer: a review. Ther Adv Med Oncol 2016; 8: 57-84 [PMID: 132 26753006 DOI: 10.1177/1758834015614530]
- Diaz LA Jr, Shiu KK, Kim TW, Jensen BV, Jensen LH, Punt C, Smith D, Garcia-Carbonero R, Benavides M, Gibbs P, de la Fourchardiere C, 133 Rivera F, Elez E, Le DT, Yoshino T, Zhong WY, Fogelman D, Marinello P, Andre T; KEYNOTE-177 Investigators. Pembrolizumab versus chemotherapy for microsatellite instability-high or mismatch repair-deficient metastatic colorectal cancer (KEYNOTE-177): final analysis of a randomised, open-label, phase 3 study. Lancet Oncol 2022; 23: 659-670 [PMID: 35427471 DOI: 10.1016/S1470-2045(22)00197-8]
- Andre T, Amonkar M, Norquist JM, Shiu KK, Kim TW, Jensen BV, Jensen LH, Punt CJA, Smith D, Garcia-Carbonero R, Sevilla I, De La 134 Fouchardiere C, Rivera F, Elez E, Diaz LA Jr, Yoshino T, Van Cutsem E, Yang P, Farooqui M, Le DT. Health-related quality of life in patients with microsatellite instability-high or mismatch repair deficient metastatic colorectal cancer treated with first-line pembrolizumab versus chemotherapy (KEYNOTE-177): an open-label, randomised, phase 3 trial. Lancet Oncol 2021; 22: 665-677 [PMID: 33812497 DOI: 10.1016/S1470-2045(21)00064-4]
- Fessas P, Lee H, Ikemizu S, Janowitz T. A molecular and preclinical comparison of the PD-1-targeted T-cell checkpoint inhibitors nivolumab 135 and pembrolizumab. Semin Oncol 2017; 44: 136-140 [PMID: 28923212 DOI: 10.1053/j.seminoncol.2017.06.002]
- Overman MJ, Lonardi S, Wong KYM, Lenz HJ, Gelsomino F, Aglietta M, Morse MA, Van Cutsem E, McDermott R, Hill A, Sawyer MB, 136 Hendlisz A, Neyns B, Svrcek M, Moss RA, Ledeine JM, Cao ZA, Kamble S, Kopetz S, André T. Durable Clinical Benefit With Nivolumab Plus Ipilimumab in DNA Mismatch Repair-Deficient/Microsatellite Instability-High Metastatic Colorectal Cancer. J Clin Oncol 2018; 36: 773-779 [PMID: 29355075 DOI: 10.1200/JCO.2017.76.9901]
- 137 Lenz HJ, Van Cutsem E, Luisa Limon M, Wong KYM, Hendlisz A, Aglietta M, García-Alfonso P, Neyns B, Luppi G, Cardin DB, Dragovich T, Shah U, Abdullaev S, Gricar J, Ledeine JM, Overman MJ, Lonardi S. First-Line Nivolumab Plus Low-Dose Ipilimumab for Microsatellite Instability-High/Mismatch Repair-Deficient Metastatic Colorectal Cancer: The Phase II CheckMate 142 Study. J Clin Oncol 2022; 40: 161-170 [PMID: 34637336 DOI: 10.1200/JCO.21.01015]
- Overman MJ, McDermott R, Leach JL, Lonardi S, Lenz HJ, Morse MA, Desai J, Hill A, Axelson M, Moss RA, Goldberg MV, Cao ZA, 138 Ledeine JM, Maglinte GA, Kopetz S, André T. Nivolumab in patients with metastatic DNA mismatch repair-deficient or microsatellite instability-high colorectal cancer (CheckMate 142): an open-label, multicentre, phase 2 study. Lancet Oncol 2017; 18: 1182-1191 [PMID: 28734759 DOI: 10.1016/S1470-2045(17)30422-9]
- Shah NJ, Kelly WJ, Liu SV, Choquette K, Spira A. Product review on the Anti-PD-L1 antibody atezolizumab. Hum Vaccin Immunother 2018; 139 14: 269-276 [PMID: 29194007 DOI: 10.1080/21645515.2017.1403694]
- Eng C, Kim TW, Bendell J, Argilés G, Tebbutt NC, Di Bartolomeo M, Falcone A, Fakih M, Kozloff M, Segal NH, Sobrero A, Yan Y, Chang 140 I, Uyei A, Roberts L, Ciardiello F; IMblaze370 Investigators. Atezolizumab with or without cobimetinib versus regorafenib in previously treated metastatic colorectal cancer (IMblaze370): a multicentre, open-label, phase 3, randomised, controlled trial. Lancet Oncol 2019; 20: 849-861 [PMID: 31003911 DOI: 10.1016/S1470-2045(19)30027-0]
- André T, Berton D, Curigliano G, Sabatier R, Tinker AV, Oaknin A, Ellard S, de Braud F, Arkenau HT, Trigo J, Gravina A, Kristeleit R, 141 Moreno V, Abdeddaim C, Vano YA, Samouëlian V, Miller R, Boni V, Torres AA, Gilbert L, Brown J, Dewal N, Dabrowski C, Antony G, Zografos E, Veneris J, Banerjee S. Antitumor Activity and Safety of Dostarlimab Monotherapy in Patients With Mismatch Repair Deficient Solid Tumors: A Nonrandomized Controlled Trial. JAMA Netw Open 2023; 6: e2341165 [PMID: 37917058 DOI: 10.1001/jamanetworkopen.2023.41165]
- Cercek A, Lumish M, Sinopoli J, Weiss J, Shia J, Lamendola-Essel M, El Dika IH, Segal N, Shcherba M, Sugarman R, Stadler Z, Yaeger R, 142 Smith JJ, Rousseau B, Argiles G, Patel M, Desai A, Saltz LB, Widmar M, Iyer K, Zhang J, Gianino N, Crane C, Romesser PB, Pappou EP, Paty P, Garcia-Aguilar J, Gonen M, Gollub M, Weiser MR, Schalper KA, Diaz LA Jr. PD-1 Blockade in Mismatch Repair-Deficient, Locally Advanced Rectal Cancer. N Engl J Med 2022; 386: 2363-2376 [PMID: 35660797 DOI: 10.1056/NEJMoa2201445]
- Bajbouj K, Qaisar R, Alshura MA, Ibrahim Z, Alebaji MB, Al Ani AW, Janajrah HM, Bilalaga MM, Omara AI, Abou Assaleh RS, Saber-143 Ayad MM, Elmoselhi AB. Synergistic Anti-Angiogenic Effect of Combined VEGFR Kinase Inhibitors, Lenvatinib, and Regorafenib: A



Therapeutic Potential for Breast Cancer. Int J Mol Sci 2022; 23 [PMID: 35457226 DOI: 10.3390/ijms23084408]

- Fakih M, Raghav KPS, Chang DZ, Larson T, Cohn AL, Huyck TK, Cosgrove D, Fiorillo JA, Tam R, D'Adamo D, Sharma N, Brennan BJ, 144 Wang YA, Coppieters S, Zebger-Gong H, Weispfenning A, Seidel H, Ploeger BA, Mueller U, Oliveira CSV, Paulson AS. Regorafenib plus nivolumab in patients with mismatch repair-proficient/microsatellite stable metastatic colorectal cancer: a single-arm, open-label, multicentre phase 2 study. EClinicalMedicine 2023; 58: 101917 [PMID: 37090438 DOI: 10.1016/j.eclinm.2023.101917]
- Bendell J, Ciardiello F, Tabernero J, Tebbutt N, Eng C, Bartolomeo MD, Falcone A, Fakih M, Kozloff M, Segal N, Sobrero A, Shi Y, Roberts 145 L, Yan Y, Chang I, Uyei A, Kim T. Efficacy and safety results from IMblaze370, a randomised Phase III study comparing atezolizumab+cobimetinib and atezolizumab monotherapy vs regorafenib in chemotherapy-refractory metastatic colorectal cancer. Ann Oncol 2018; 29: v123 [DOI: 10.1093/annonc/mdy208.003]
- Khaddour K, Maahs L, Avila-Rodriguez AM, Maamar Y, Samaan S, Ansstas G. Melanoma Targeted Therapies beyond BRAF-Mutant 146 Melanoma: Potential Druggable Mutations and Novel Treatment Approaches. Cancers (Basel) 2021; 13 [PMID: 34831002 DOI: 10.3390/cancers13225847
- El-Khoueiry AB, Fakih M, Gordon MS, Tsimberidou AM, Bullock AJ, Wilky BA, Trent JC, Margolin KA, Mahadevan D, Balmanoukian AS, 147 Sanborn RE, Schwartz GK, Bockorny B, Moser JC, Grossman JE, Feliu WIO, Rosenthal K, O'Day S, Lenz HJ, and Schlechter BL. Results from a phase 1a/1b study of botensilimab (BOT), a novel innate/adaptive immune activator, plus balstilimab (BAL; anti-PD-1 antibody) in metastatic heavily pretreated microsatellite stable colorectal cancer (MSS CRC). J Clin Oncol 2023; 41 : Suppl 4: LBA8-LBA [DOI: 10.1200/JCO.2023.41.4\_suppl.LBA8]
- Fritz JM, Lenardo MJ. Development of immune checkpoint therapy for cancer. J Exp Med 2019; 216: 1244-1254 [PMID: 31068379 DOI: 148 10.1084/jem.20182395]
- Lo B, Zhang K, Lu W, Zheng L, Zhang Q, Kanellopoulou C, Zhang Y, Liu Z, Fritz JM, Marsh R, Husami A, Kissell D, Nortman S, Chaturvedi 149 V, Haines H, Young LR, Mo J, Filipovich AH, Bleesing JJ, Mustillo P, Stephens M, Rueda CM, Chougnet CA, Hoebe K, McElwee J, Hughes JD, Karakoc-Aydiner E, Matthews HF, Price S, Su HC, Rao VK, Lenardo MJ, Jordan MB. AUTOIMMUNE DISEASE. Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy. Science 2015; 349: 436-440 [PMID: 26206937 DOI: 10.1126/science.aaa1663
- Kuehn HS, Ouvang W, Lo B, Deenick EK, Niemela JE, Avery DT, Schickel JN, Tran DQ, Stoddard J, Zhang Y, Frucht DM, Dumitriu B, 150 Scheinberg P, Folio LR, Frein CA, Price S, Koh C, Heller T, Seroogy CM, Huttenlocher A, Rao VK, Su HC, Kleiner D, Notarangelo LD, Rampertaap Y, Olivier KN, McElwee J, Hughes J, Pittaluga S, Oliveira JB, Meffre E, Fleisher TA, Holland SM, Lenardo MJ, Tangye SG, Uzel G. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. Science 2014; 345: 1623-1627 [PMID: 25213377 DOI: 10.1126/science.1255904]
- Michot JM, Bigenwald C, Champiat S, Collins M, Carbonnel F, Postel-Vinay S, Berdelou A, Varga A, Bahleda R, Hollebecque A, Massard C, 151 Fuerea A, Ribrag V, Gazzah A, Armand JP, Amellal N, Angevin E, Noel N, Boutros C, Mateus C, Robert C, Soria JC, Marabelle A, Lambotte O. Immune-related adverse events with immune checkpoint blockade: a comprehensive review. Eur J Cancer 2016; 54: 139-148 [PMID: 26765102 DOI: 10.1016/j.ejca.2015.11.016]
- Kumar V, Chaudhary N, Garg M, Floudas CS, Soni P, Chandra AB. Current Diagnosis and Management of Immune Related Adverse Events 152 (irAEs) Induced by Immune Checkpoint Inhibitor Therapy. Front Pharmacol 2017; 8: 49 [PMID: 28228726 DOI: 10.3389/fphar.2017.00049]
- Geisler AN, Phillips GS, Barrios DM, Wu J, Leung DYM, Moy AP, Kern JA, Lacouture ME. Immune checkpoint inhibitor-related 153 dermatologic adverse events. J Am Acad Dermatol 2020; 83: 1255-1268 [PMID: 32454097 DOI: 10.1016/j.jaad.2020.03.132]
- Wang PF, Chen Y, Song SY, Wang TJ, Ji WJ, Li SW, Liu N, Yan CX. Immune-Related Adverse Events Associated with Anti-PD-1/PD-L1 154 Treatment for Malignancies: A Meta-Analysis. Front Pharmacol 2017; 8: 730 [PMID: 29093678 DOI: 10.3389/fphar.2017.00730]
- Weingarden AR, Rubin SJS, Gubatan J. Immune checkpoint inhibitor-mediated colitis in gastrointestinal malignancies and inflammatory 155 bowel disease. World J Gastrointest Oncol 2021; 13: 772-798 [PMID: 34457186 DOI: 10.4251/wjgo.v13.i8.772]
- Hashash JG, Francis FF, Farraye FA. Diagnosis and Management of Immune Checkpoint Inhibitor Colitis. Gastroenterol Hepatol (NY) 2021; 156 17: 358-366 [PMID: 34602898]
- Luoma AM, Suo S, Williams HL, Sharova T, Sullivan K, Manos M, Bowling P, Hodi FS, Rahma O, Sullivan RJ, Boland GM, Nowak JA, 157 Dougan SK, Dougan M, Yuan GC, Wucherpfennig KW. Molecular Pathways of Colon Inflammation Induced by Cancer Immunotherapy. Cell 2020; 182: 655-671.e22 [PMID: 32603654 DOI: 10.1016/j.cell.2020.06.001]
- Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer. Nature 2012; 487: 330-337 158 [PMID: 22810696 DOI: 10.1038/nature11252]
- Saeterdal I, Bjørheim J, Lislerud K, Gjertsen MK, Bukholm IK, Olsen OC, Nesland JM, Eriksen JA, Møller M, Lindblom A, Gaudernack G. 159 Frameshift-mutation-derived peptides as tumor-specific antigens in inherited and spontaneous colorectal cancer. Proc Natl Acad Sci USA 2001; 98: 13255-13260 [PMID: 11687624 DOI: 10.1073/pnas.231326898]
- Mulet-Margalef N, Linares J, Badia-Ramentol J, Jimeno M, Sanz Monte C, Manzano Mozo JL, Calon A. Challenges and Therapeutic 160 Opportunities in the dMMR/MSI-H Colorectal Cancer Landscape. Cancers (Basel) 2023; 15 [PMID: 36831367 DOI: 10.3390/cancers15041022]
- Wu Y, Zhuang J, Qu Z, Yang X, Han S. Advances in immunotyping of colorectal cancer. Front Immunol 2023; 14: 1259461 [PMID: 161 37876934 DOI: 10.3389/fimmu.2023.1259461]
- Sahin IH, Ciombor KK, Diaz LA, Yu J, Kim R. Immunotherapy for Microsatellite Stable Colorectal Cancers: Challenges and Novel 162 Therapeutic Avenues. Am Soc Clin Oncol Educ Book 2022; 42: 1-12 [PMID: 35658496 DOI: 10.1200/EDBK\_349811]
- 163 Hung KE, Maricevich MA, Richard LG, Chen WY, Richardson MP, Kunin A, Bronson RT, Mahmood U, Kucherlapati R. Development of a mouse model for sporadic and metastatic colon tumors and its use in assessing drug treatment. Proc Natl Acad Sci USA 2010; 107: 1565-1570 [PMID: 20080688 DOI: 10.1073/pnas.0908682107]
- Roper J, Tammela T, Akkad A, Almeqdadi M, Santos SB, Jacks T, Yilmaz ÖH. Colonoscopy-based colorectal cancer modeling in mice with 164 CRISPR-Cas9 genome editing and organoid transplantation. Nat Protoc 2018; 13: 217-234 [PMID: 29300388 DOI: 10.1038/nprot.2017.136]
- Svoronos N, Perales-Puchalt A, Allegrezza MJ, Rutkowski MR, Payne KK, Tesone AJ, Nguyen JM, Curiel TJ, Cadungog MG, Singhal S, 165 Eruslanov EB, Zhang P, Tchou J, Zhang R, Conejo-Garcia JR. Tumor Cell-Independent Estrogen Signaling Drives Disease Progression through Mobilization of Myeloid-Derived Suppressor Cells. Cancer Discov 2017; 7: 72-85 [PMID: 27694385 DOI: 10.1158/2159-8290.CD-16-0502]
- Oliveira RC, Abrantes AM, Tralhão JG, Botelho MF. The role of mouse models in colorectal cancer research-The need and the importance of 166 the orthotopic models. Animal Model Exp Med 2020; 3: 1-8 [PMID: 32318654 DOI: 10.1002/ame2.12102]



- Bürtin F, Mullins CS, Linnebacher M. Mouse models of colorectal cancer: Past, present and future perspectives. World J Gastroenterol 2020; 167 26: 1394-1426 [PMID: 32308343 DOI: 10.3748/wjg.v26.i13.1394]
- Tseng W, Leong X, Engleman E. Orthotopic mouse model of colorectal cancer. J Vis Exp 2007; 484 [PMID: 18989400 DOI: 10.3791/484] 168
- Kim H, Kim M, Im SK, Fang S. Mouse Cre-LoxP system: general principles to determine tissue-specific roles of target genes. Lab Anim Res 169 2018; 34: 147-159 [PMID: 30671100 DOI: 10.5625/lar.2018.34.4.147]
- Jiang L, Hermeking H. miR-34a and miR-34b/c Suppress Intestinal Tumorigenesis. Cancer Res 2017; 77: 2746-2758 [PMID: 28363996 DOI: 170 10.1158/0008-5472.CAN-16-2183
- Parker TW, Neufeld KL. APC controls Wnt-induced β-catenin destruction complex recruitment in human colonocytes. Sci Rep 2020; 10: 171 2957 [PMID: 32076059 DOI: 10.1038/s41598-020-59899-z]
- Tetteh PW, Kretzschmar K, Begthel H, van den Born M, Korving J, Morsink F, Farin H, van Es JH, Offerhaus GJ, Clevers H. Generation of 172 an inducible colon-specific Cre enzyme mouse line for colon cancer research. Proc Natl Acad Sci USA 2016; 113: 11859-11864 [PMID: 27708166 DOI: 10.1073/pnas.1614057113]
- Li C, Lau HC, Zhang X, Yu J. Mouse Models for Application in Colorectal Cancer: Understanding the Pathogenesis and Relevance to the 173 Human Condition. Biomedicines 2022; 10 [PMID: 35885015 DOI: 10.3390/biomedicines10071710]
- 174 Xue Y, Johnson R, Desmet M, Snyder PW, Fleet JC. Generation of a transgenic mouse for colorectal cancer research with intestinal cre expression limited to the large intestine. Mol Cancer Res 2010; 8: 1095-1104 [PMID: 20663863 DOI: 10.1158/1541-7786.MCR-10-0195]
- Rivera F, Karthaus M, Hecht JR, Sevilla I, Forget F, Fasola G, Canon JL, Guan X, Demonty G, Schwartzberg LS. Final analysis of the 175 randomised PEAK trial: overall survival and tumour responses during first-line treatment with mFOLFOX6 plus either panitumumab or bevacizumab in patients with metastatic colorectal carcinoma. Int J Colorectal Dis 2017; 32: 1179-1190 [PMID: 28424871 DOI: 10.1007/s00384-017-2800-1]
- Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, Patnaik A, Aggarwal C, Gubens M, Horn L, Carcereny E, Ahn MJ, Felip 176 E, Lee JS, Hellmann MD, Hamid O, Goldman JW, Soria JC, Dolled-Filhart M, Rutledge RZ, Zhang J, Lunceford JK, Rangwala R, Lubiniecki GM, Roach C, Emancipator K, Gandhi L; KEYNOTE-001 Investigators. Pembrolizumab for the treatment of non-small-cell lung cancer. N Engl J Med 2015; 372: 2018-2028 [PMID: 25891174 DOI: 10.1056/NEJMoa1501824]
- Herson M, Jørgensen JT. Companion and Complementary Diagnostics-Focus on PD-L1 Expression Assays for PD-1/PD-L1 Checkpoint 177 Inhibitors in Non-Small Cell Lung Cancer. Ther Drug Monit 2018; 40: 9-16 [PMID: 29084031 DOI: 10.1097/FTD.00000000000460]
- André T, Overman M, Lonardi S, Aglietta M, McDermott R, Wong KYM, Morse M, Hendlisz A, Moss RA, Ledeine JM, Tang H, Cao ZA, 178 Kopetz S. Analysis of tumor PD-L1 expression and biomarkers in relation to clinical activity in patients (pts) with deficient DNA mismatch repair (dMMR)/high microsatellite instability (MSI-H) metastatic colorectal cancer (mCRC) treated with nivolumab (NIVO) + ipilimumab (IPI): CheckMate 142. Ann Oncol 2017; 28 [DOI: 10.1093/annonc/mdx393.011]
- Koelzer VH, Baker K, Kassahn D, Baumhoer D, Zlobec I. Prognostic impact of β-2-microglobulin expression in colorectal cancers stratified 179 by mismatch repair status. J Clin Pathol 2012; 65: 996-1002 [PMID: 22859396 DOI: 10.1136/jclinpath-2012-200742]
- Shin DS, Zaretsky JM, Escuin-Ordinas H, Garcia-Diaz A, Hu-Lieskovan S, Kalbasi A, Grasso CS, Hugo W, Sandoval S, Torrejon DY, 180 Palaskas N, Rodriguez GA, Parisi G, Azhdam A, Chmielowski B, Cherry G, Seja E, Berent-Maoz B, Shintaku IP, Le DT, Pardoll DM, Diaz LA Jr, Tumeh PC, Graeber TG, Lo RS, Comin-Anduix B, Ribas A. Primary Resistance to PD-1 Blockade Mediated by JAK1/2 Mutations. Cancer Discov 2017; 7: 188-201 [PMID: 27903500 DOI: 10.1158/2159-8290.CD-16-1223]
- Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinas H, Hugo W, Hu-Lieskovan S, Torrejon DY, Abril-Rodriguez G, Sandoval S, Barthly 181 L, Saco J, Homet Moreno B, Mezzadra R, Chmielowski B, Ruchalski K, Shintaku IP, Sanchez PJ, Puig-Saus C, Cherry G, Seja E, Kong X, Pang J, Berent-Maoz B, Comin-Anduix B, Graeber TG, Tumeh PC, Schumacher TN, Lo RS, Ribas A. Mutations Associated with Acquired Resistance to PD-1 Blockade in Melanoma. N Engl J Med 2016; 375: 819-829 [PMID: 27433843 DOI: 10.1056/NEJMoa1604958]
- Li B, Li T, Pignon JC, Wang B, Wang J, Shukla SA, Dou R, Chen Q, Hodi FS, Choueiri TK, Wu C, Hacohen N, Signoretti S, Liu JS, Liu XS. 182 Landscape of tumor-infiltrating T cell repertoire of human cancers. Nat Genet 2016; 48: 725-732 [PMID: 27240091 DOI: 10.1038/ng.3581]
- Aurisicchio L, Fridman A, Mauro D, Sheloditna R, Chiappori A, Bagchi A, Ciliberto G. Safety, tolerability and immunogenicity of V934/ 183 V935 hTERT vaccination in cancer patients with selected solid tumors: a phase I study. J Transl Med 2020; 18: 39 [PMID: 32000810 DOI: 10.1186/s12967-020-02228-91
- Bartnik A, Nirmal AJ, Yang SY. Peptide Vaccine Therapy in Colorectal Cancer. Vaccines (Basel) 2012; 1: 1-16 [PMID: 26343847 DOI: 184 10.3390/vaccines1010001
- Heery CR, Singh BH, Rauckhorst M, Marté JL, Donahue RN, Grenga I, Rodell TC, Dahut W, Arlen PM, Madan RA, Schlom J, Gulley JL. 185 Phase I Trial of a Yeast-Based Therapeutic Cancer Vaccine (GI-6301) Targeting the Transcription Factor Brachyury. Cancer Immunol Res 2015; 3: 1248-1256 [PMID: 26130065 DOI: 10.1158/2326-6066.CIR-15-0119]
- Li M, Yuan YH, Han Y, Liu YX, Yan L, Wang Y, Gu J. Expression profile of cancer-testis genes in 121 human colorectal cancer tissue and 186 adjacent normal tissue. Clin Cancer Res 2005; 11: 1809-1814 [PMID: 15756003 DOI: 10.1158/1078-0432.CCR-04-1365]
- Liu C, Xie Y, Sun B, Geng F, Zhang F, Guo Q, Wu H, Yu B, Wu J, Yu X, Kong W, Zhang H. MUC1- and Survivin-based DNA Vaccine 187 Combining Immunoadjuvants CpG and interleukin-2 in a Bicistronic Expression Plasmid Generates Specific Immune Responses and Antitumour Effects in a Murine Colorectal Carcinoma Model. Scand J Immunol 2018; 87: 63-72 [PMID: 29193199 DOI: 10.1111/sji.12633]
- Snook AE, Baybutt TR, Xiang B, Abraham TS, Flickinger JC Jr, Hyslop T, Zhan T, Kraft WK, Sato T, Waldman SA. Split tolerance permits 188 safe Ad5-GUCY2C-PADRE vaccine-induced T-cell responses in colon cancer patients. J Immunother Cancer 2019; 7: 104 [PMID: 31010434 DOI: 10.1186/s40425-019-0576-2]
- Connal S, Cameron JM, Sala A, Brennan PM, Palmer DS, Palmer JD, Perlow H, Baker MJ. Liquid biopsies: the future of cancer early 189 detection. J Transl Med 2023; 21: 118 [PMID: 36774504 DOI: 10.1186/s12967-023-03960-8]
- Fukumoto M, Kurohara A, Akagi N, Yoshida D, Yoshida S. Ga-67 visualization of the coexistence of two mucosa-associated lymphoid tissue 190 (MALT) lymphomas in the thyroid and stomach. Clin Nucl Med 1998; 23: 484 [PMID: 9676964 DOI: 10.1097/00003072-199807000-00024]
- Tang Q, Wang J, Frank A, Lin J, Li Z, Chen CW, Jin L, Wu T, Greenwald BD, Mashimo H, Chen Y. Depth-resolved imaging of colon tumor 191 using optical coherence tomography and fluorescence laminar optical tomography. Biomed Opt Express 2016; 7: 5218-5232 [PMID: 28018738 DOI: 10.1364/BOE.7.005218]
- 192 Jensen CD, Corley DA, Quinn VP, Doubeni CA, Zauber AG, Lee JK, Zhao WK, Marks AR, Schottinger JE, Ghai NR, Lee AT, Contreras R, Klabunde CN, Quesenberry CP, Levin TR, Mysliwiec PA. Fecal Immunochemical Test Program Performance Over 4 Rounds of Annual Screening: A Retrospective Cohort Study. Ann Intern Med 2016; 164: 456-463 [PMID: 26811150 DOI: 10.7326/M15-0983]



- Louie BH, Kato S, Kim KH, Lim HJ, Lee S, Okamura R, Fanta PT, Kurzrock R. Precision medicine-based therapies in advanced colorectal 193 cancer: The University of California San Diego Molecular Tumor Board experience. Mol Oncol 2022; 16: 2575-2584 [PMID: 35238467 DOI: 10.1002/1878-0261.13202
- Malone ER, Oliva M, Sabatini PJB, Stockley TL, Siu LL. Molecular profiling for precision cancer therapies. Genome Med 2020; 12: 8 [PMID: 194 31937368 DOI: 10.1186/s13073-019-0703-1]
- 195 Jung G, Hernández-Illán E, Moreira L, Balaguer F, Goel A. Epigenetics of colorectal cancer: biomarker and therapeutic potential. Nat Rev Gastroenterol Hepatol 2020; 17: 111-130 [PMID: 31900466 DOI: 10.1038/s41575-019-0230-y]
- Gao S, Tibiche C, Zou J, Zaman N, Trifiro M, O'Connor-McCourt M, Wang E. Identification and Construction of Combinatory Cancer 196 Hallmark-Based Gene Signature Sets to Predict Recurrence and Chemotherapy Benefit in Stage II Colorectal Cancer. JAMA Oncol 2016; 2: 37-45 [PMID: 26502222 DOI: 10.1001/jamaoncol.2015.3413]
- 197 Azwar S, Seow HF, Abdullah M, Faisal Jabar M, Mohtarrudin N. Recent Updates on Mechanisms of Resistance to 5-Fluorouracil and Reversal Strategies in Colon Cancer Treatment. Biology (Basel) 2021; 10 [PMID: 34571731 DOI: 10.3390/biology10090854]
- Blondy S, David V, Verdier M, Mathonnet M, Perraud A, Christou N. 5-Fluorouracil resistance mechanisms in colorectal cancer: From 198 classical pathways to promising processes. Cancer Sci 2020; 111: 3142-3154 [PMID: 32536012 DOI: 10.1111/cas.14532]
- 199 Grosso JF, Kelleher CC, Harris TJ, Maris CH, Hipkiss EL, De Marzo A, Anders R, Netto G, Getnet D, Bruno TC, Goldberg MV, Pardoll DM, Drake CG. LAG-3 regulates CD8+ T cell accumulation and effector function in murine self- and tumor-tolerance systems. J Clin Invest 2007; 117: 3383-3392 [PMID: 17932562 DOI: 10.1172/JCI31184]
- 200 Ngiow SF, von Scheidt B, Akiba H, Yagita H, Teng MW, Smyth MJ. Anti-TIM3 antibody promotes T cell IFN-y-mediated antitumor immunity and suppresses established tumors. Cancer Res 2011; 71: 3540-3551 [PMID: 21430066 DOI: 10.1158/0008-5472.CAN-11-0096]
- 201 Sakuishi K, Apetoh L, Sullivan JM, Blazar BR, Kuchroo VK, Anderson AC. Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity. J Exp Med 2010; 207: 2187-2194 [PMID: 20819927 DOI: 10.1084/jem.20100643]
- 202 Woo SR, Turnis ME, Goldberg MV, Bankoti J, Selby M, Nirschl CJ, Bettini ML, Gravano DM, Vogel P, Liu CL, Tangsombatvisit S, Grosso JF, Netto G, Smeltzer MP, Chaux A, Utz PJ, Workman CJ, Pardoll DM, Korman AJ, Drake CG, Vignali DA. Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape. Cancer Res 2012; 72: 917-927 [PMID: 22186141 DOI: 10.1158/0008-5472.CAN-11-1620]
- 203 Curigliano G, Gelderblom H, Mach N, Doi T, Tai D, Forde PM, Sarantopoulos J, Bedard PL, Lin CC, Hodi FS, Wilgenhof S, Santoro A, Sabatos-Peyton CA, Longmire TA, Xyrafas A, Sun H, Gutzwiller S, Manenti L, Naing A. Phase I/Ib Clinical Trial of Sabatolimab, an Anti-TIM-3 Antibody, Alone and in Combination with Spartalizumab, an Anti-PD-1 Antibody, in Advanced Solid Tumors. Clin Cancer Res 2021; 27: 3620-3629 [PMID: 33883177 DOI: 10.1158/1078-0432.CCR-20-4746]
- Curti BD, Kovacsovics-Bankowski M, Morris N, Walker E, Chisholm L, Floyd K, Walker J, Gonzalez I, Meeuwsen T, Fox BA, Moudgil T, 204 Miller W, Haley D, Coffey T, Fisher B, Delanty-Miller L, Rymarchyk N, Kelly T, Crocenzi T, Bernstein E, Sanborn R, Urba WJ, Weinberg AD. OX40 is a potent immune-stimulating target in late-stage cancer patients. Cancer Res 2013; 73: 7189-7198 [PMID: 24177180 DOI: 10.1158/0008-5472.CAN-12-4174]
- 205 Harding JJ, Moreno V, Bang YJ, Hong MH, Patnaik A, Trigo J, Szpurka AM, Yamamoto N, Doi T, Fu S, Calderon B, Velez de Mendizabal N, Calvo E, Yu D, Gandhi L, Liu ZT, Galvao VR, Leow CC, de Miguel MJ. Blocking TIM-3 in Treatment-refractory Advanced Solid Tumors: A Phase Ia/b Study of LY3321367 with or without an Anti-PD-L1 Antibody. Clin Cancer Res 2021; 27: 2168-2178 [PMID: 33514524 DOI: 10.1158/1078-0432.CCR-20-4405
- Mettu NB, Ulahannan SV, Bendell JC, Garrido-Laguna I, Strickler JH, Moore KN, Stagg R, Kapoun AM, Faoro L, Sharma S. A Phase 1a/b 206 Open-Label, Dose-Escalation Study of Etigilimab Alone or in Combination with Nivolumab in Patients with Locally Advanced or Metastatic Solid Tumors. Clin Cancer Res 2022; 28: 882-892 [PMID: 34844977 DOI: 10.1158/1078-0432.CCR-21-2780]
- Brenner D, Blaser H, Mak TW. Regulation of tumour necrosis factor signalling: live or let die. Nat Rev Immunol 2015; 15: 362-374 [PMID: 207 26008591 DOI: 10.1038/nri3834]
- Croft M, Benedict CA, Ware CF. Clinical targeting of the TNF and TNFR superfamilies. Nat Rev Drug Discov 2013; 12: 147-168 [PMID: 208 23334208 DOI: 10.1038/nrd3930]
- Ward-Kavanagh LK, Lin WW, Šedý JR, Ware CF. The TNF Receptor Superfamily in Co-stimulating and Co-inhibitory Responses. Immunity 209 2016; 44: 1005-1019 [PMID: 27192566 DOI: 10.1016/j.immuni.2016.04.019]
- Lyons YA, Wu SY, Overwijk WW, Baggerly KA, Sood AK. Immune cell profiling in cancer: molecular approaches to cell-specific 210 identification. NPJ Precis Oncol 2017; 1: 26 [PMID: 29872708 DOI: 10.1038/s41698-017-0031-0]
- Mukherjee S. Genomics-Guided Immunotherapy for Precision Medicine in Cancer. Cancer Biother Radiopharm 2019; 34: 487-497 [PMID: 211 31314580 DOI: 10.1089/cbr.2018.2758]





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