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Editorial Board Member of World Journal of Gastroenterology, Júlio Maria Fonseca Chebli, MD, PhD, Associate Professor, Professor, Research Scientist, Department of Medicine, Inflammatory Bowel disease Center, University Hospital of the Federal University, Juiz de Fora 36036-247, Minas Gerais, Brazil. julio.chebli@medicina.ufjf.br

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MINIREVIEWS

Recent advances in age-related metabolic dysfunction-associated steatotic liver disease

Qian-Jun He, Yi-Fei Li, Ling-Tong Zhao, Chun-Tong Lin, Chun-Yan Yu, Dan Wang

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Qian-Jun He, Ling-Tong Zhao, Chun-Tong Lin, Chun-Yan Yu, Dan Wang, College of Basic Medicine, Beihua University, Jilin 132013, Jilin Province, China

Yi-Fei Li, Department of Traumatic Surgery, Jilin Central Hospital, Jilin 132001, Jilin Province, China

Corresponding author: Dan Wang, PhD, Professor, College of Basic Medicine, Beihua University, No. 3999 Binjiangdonglu, Jilin 132013, Jilin Province, China. jlj11215@163.com

Abstract

Metabolic dysfunction-associated steatotic liver disease (MASLD) affects approximately 25% of the world's population and has become a leading cause of chronic liver disease. In recent years, an increasing amount of data suggests that MASLD is associated with aging. As the population ages, age-related MASLD will become a major global health problem. Targeting an aging will become a new approach to the treatment of MASLD. This paper reviews the current studies on the role of aging-related factors and therapeutic targets in MASLD, including: Oxidative stress, autophagy, mitochondrial homeostasis, bile acid metabolism homeostasis, and dysbiosis. The aim is to identify effective therapeutic targets for age-related MASLD and its progression.

Key Words: Metabolic dysfunction-associated steatotic liver disease; Aging; Mitochondrial homeostasis; Bile acid homeostasis; Dysbiosis

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Core Tip: Aging is closely associated with metabolic dysfunction-associated steatotic liver disease (MASLD). We will focus on the main features of age-related MASLD, the mechanisms by which age-related factors, such as oxidative stress, autophagy, mitochondrial homeostasis, bile acid metabolism and bacterial dysbiosis induce MASLD, as well as the study of related therapeutic targets.

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INTRODUCTION

Aging is broadly defined as the time-dependent decline in function that affects most organisms. López-Otín *et al*[1], in conjunction with recent research findings, has revised the concept of "aging hallmarks" by incorporating an additional 12 aging features, including macroautophagy deficiencies, chronic inflammation, and dysbiosis. Owing to advancements in living standards, economic growth, and healthcare infrastructure, population aging has emerged as the primary demographic phenomenon globally. Currently, the proportion of elderly people over the age of 60 worldwide is about one ninth, with projections indicating that by the year 2050, this ratio will escalate to one-fifth[2]. Human aging is characterized by molecular, structural, and functional alterations in various organ systems, including the liver. Within the intensifying milieu of aging, age-associated liver dysfunction has become a compelling clinical challenge that demands urgent attention and resolution.

Cellular damage, if left unattended or irreparable, can lead to cellular apoptosis or senescence, which is the fundamental cellular process employed by the organism in its defense against cancer. Similarly, following exposure to damage and stress signals, aging can irreversibly arrest the G0/G1 phase of the cell cycle. This phenomenon restricts the proliferative potential of damaged cells[3], ultimately leading to changes in the microenvironment and tissue homeostasis. Upon entering a state of cellular senescence, cells undergo morphological transformations characterized by permanent cell cycle arrest, displaying distinctive traits such as altered secretion profiles, macromolecular damage, and metabolic shifts. These alterations include an upregulated expression of senescence-associated β -galactosidase, acquisition of the senescence-associated secretory phenotype (SASP), elevated levels of P16INK4a (P16) and P21Cip1/Waf1 (P21), and increased levels of reactive oxygen species (ROS)[4]. In particular, the senescence of hepatic cells can potentially contribute to intracellular lipid accumulation, fibrosis, and inflammation, while concurrently secreting age-associated inflammatory mediators.

In 2023, the global hepatology community renamed nonalcoholic fatty liver disease (NAFLD) as metabolic dysfunction-associated steatotic liver disease (MASLD). MASLD is the most common chronic liver disease worldwide. Patients presenting hepatic steatosis and at least one of five cardiometabolic risk factors are diagnosed with MASLD. Generally, the initial hepatic steatosis observed in MASLD is considered reversible. However, persistent metabolic disturbances can lead to aberrant liver cell metabolism, resulting in the accumulation of lipids, such as fatty acids, cholesterol, and other lipid metabolites. This, in turn, induces the onset of MASLD. Excessive lipid accumulation in the liver can lead to lipotoxicity, mitochondrial dysfunction, increased ROS levels, and inflammation. These events signify the transition from MASLD to metabolic dysfunction-associated steatohepatitis (MASH) or the progression to more severe stages[5]. Younossi *et al*[6] conducted an extensive analysis involving 8515431 samples from 22 countries and demonstrated that the global prevalence of MASLD was 25.24%. The highest rates of prevalence were identified in the Middle East and South America, with Africa exhibiting the lowest prevalence. Notably, MASLD exhibited strong associations with the components of metabolic syndrome, including obesity, diabetes, and dyslipidemia. The median age of the MASLD population in the United States in 2015 was 50 years old. Researchers predict that it will increase to 55 years old by 2030. This indicates that with an increase in population aging, the incidence of MASLD-related liver diseases and mortality rates will increase in the United States[7].

RELATIONSHIP BETWEEN AGING AND MASLD

The aging phenotype exhibits remarkable stability, displaying resistance to mitotic stimuli and apoptosis^[8]. In healthy cells, senescence may be triggered primarily by factors such as oxidative stress, mitochondrial imbalance, and chronic inflammation either independently or synergistically. The progressive accumulation of senescent cells can elicit bystander effects, ultimately leading to organ aging and functional impairments^[4]. Age dependence is the main risk factor for chronic liver diseases, including MASLD and MASH. The accumulation of aging cells drives liver steatosis, and liver cell aging is closely related to the progression of MASLD[9]. The intricate mechanisms governing the pathogenesis of MASLD are not completely clear. Elevated hepatic lipid accumulation is a significant risk factor for MASLD, with hepatic cell senescence playing a contributory role in the development of hepatic steatosis. This can be best described by the "twohit" hypothesis. "First hit" denotes the initial occurrence of excessive hepatic lipid accumulation due to alterations in lipid metabolism, which leads to the development of nonalcoholic fatty liver (NAFL). The "second hit" entails the induction of hepatic cell damage, including oxidative stress, inflammatory cytokine release, and mitochondrial dysfunction, building upon the backdrop of hepatic lipotoxicity. This second hit can precipitate the progression of NAFL towards MASH and, in more severe cases, fibrosis, cirrhosis, hepatocellular carcinoma, and even death[10]. In recent years, the "multiple-hit" hypothesis has emerged as a more elaborate concept for elucidating the pathogenesis of MASLD. This theory underscores the pivotal roles played by gut microbiota (GM), insulin resistance, and adipose tissue-derived adipokines. These factors, through their interconnected relationships along the gut-liver axis, interact synergistically and causally, contributing to

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the progression of MASLD[11].

Oxidative stress

In response to oxidative stress, organisms generate substances, including ROS or free radicals, that can inflict damage on cellular constituents such as the cell membrane, proteins, and DNA. This damage can ultimately lead to cellular senescence or even apoptosis. The acceleration of oxidative stress due to hepatocyte aging reportedly augments lipid accumulation in the liver, suggesting the pivotal role of oxidative stress in the etiology and progression of MASLD. Oxidative stress initiates DNA damage, as well as instigates autophagy and the secretion of SASP. These processes are concomitant with the activation of the p53-p21 and p16-Rb pathways, leading to premature cellular senescence. Concurrently, oxidative stress exacerbates disruptions in lipid metabolism, promotes inflammatory responses, and induces hepatocyte damage[4]. During the normal aging process of the liver, hepatic stellate cells help drive macrophage differentiation towards the M1 phenotype, exerting anti-tumor effects. Conversely, a study has found that p53-knockout hepatic stellate cells promote macrophage differentiation towards the pro-tumor M2 phenotype, thereby inducing the progression of liver fibrosis, cirrhosis, and even HCC[12].

MASLD is characterized by increased levels of senescent cells compared with the control group. Additionally, patients with MASLD demonstrate significantly higher telomere shortening, along with increased cell cycle arrest, which coincides with an increased expression of p21[13]. Reportedly, Krüppel-like factor 16 (KLF16) can bind to the promoter of peroxisome proliferator-activated receptor alpha (PPARa), thereby activating it. Upregulation of KLF16 expression expedites fatty acid β -oxidation and mitigates oxidative stress responses in db/db and high-fat diet (HFD) mice. This, in turn, results in a reduction in hepatic lipid accumulation and an improvement in MASLD[14]. Oxidative stress induced cellular aging can alter the liver microenvironment, leading to disease progression from simple steatosis to inflammation and fibrosis, as well as hepatocellular carcinoma.

Autophagy

In hepatic cells, autophagy is subject to modulation by both individual and organ-level aging processes, with pronounced attenuation of hepatic autophagy evident in the aging state[15]. The process of aging decreases the quantity and efficiency of autophagosomes, subsequently causing a buildup of lipid droplets and resultant liver damage. Autophagy inhibition is posited as a potential risk factor in the progression of age-related MASLD. An animal study has revealed that the administration of plasma from younger mice restores autophagic activity in aged mice, effectively mitigating liver aging, lipid accumulation, and fibrosis[16]. Using Atg7-deficient mice, researchers have substantiated that the autophagic impairment in liver sinusoidal endothelial cells not only accelerates liver inflammation and fibrosis in the early phases of MASLD but also exacerbates hepatic inflammation and fibrosis during the advanced stages[17].

Mice deficient in Omi/HtrA2 exhibit premature aging symptoms and age-related autophagy inhibition, which results in hepatic dysfunction[18]. Therefore, augmenting autophagy may mitigate aging and hepatic steatosis, thus alleviating MASLD. Mice overexpressing Omi/HtrA2 have demonstrated enhanced autophagic activity, diminished hepatic steatosis, and elevated hepatic fatty acid β -oxidation, which ameliorated HFD-induced MASLD along with hepatic inflammation[19]. P62/sequestosome 1 facilitates the phosphorylation of unc-51 Like autophagy activating kinase 1, thereby promoting autophagy activation and triggering NFE2L2/NRF2 activation. This protects mouse liver cells against lipotoxic damage[20]. AMPK activation is a regulator of autophagy and aging-related changes in AMPK activation may impact autophagic processes, which results in decreased formation of autophagosomes and further hastening of the aging process[21]. In hepatic cells, PPAR δ activates the autophagy-lysosome pathway *via* the AMPK/mammalian target of rapamycin (mTOR) signaling to induce fatty acid β -oxidation, which reduces hepatic lipid levels[22]. Autophagy changes may be an important target for the treatment of age-related MASLD.

Mitochondrial homeostasis

In mouse liver, increased mitochondrial ROS levels are linked with the aging process. Oxidative stress triggers mutations in the mitochondrial DNA, leading to the accumulation of mitochondrial DNA fragments within the cell nucleus. This accumulation subsequently contributes to mitochondrial dysfunction[23]. It has been shown that the absence of the gene encoding the nonhomologous end-joining enzyme known as DNA ligase IV (DNL4) exacerbates linear mitochondrial DNA (mtDNA) aggregation in the nucleus. Cheng *et al*[24], proposed that linear nuclear mtDNA fragments accelerate aging in yme1-1 mutant cells by affecting nuclear DNA replication, recombination, repair, and transcription. In addition, mice deficient in the antioxidant enzyme superoxide dismutase 1 (SOD1) demonstrate premature aging, along with hepatic damage. Investigations have unveiled that SOD1-deficient (SOD1-/-) mice display shifts in the composition of their GM, including alterations in the ratio of Firmicutes and Bacteroidetes, a significant reduction in lactobacilli, increased hepatic metabolites, and manifestation of a systemic aging phenotype[25]. Moreover, cellular senescence results in mitochondrial dysfunction, which induces respiratory chain disturbances, membrane potential anomalies, and concomitant ROS generation, all of which further induce the development of MASLD[26].

Free fatty acids in the liver are primarily metabolized *via* two pathways: mitochondrial β-oxidation and esterification into triglycerides. The preservation of mitochondrial homeostasis is a pivotal factor in hepatic lipid metabolism. However, excessive free fatty acids burden the process of mitochondrial β-oxidation, subsequently leading to an imbalance in mitochondrial homeostasis. This imbalance further exacerbates the accumulation of lipids within the hepatic cells, ultimately contributing to the development of MASLD[27]. Omi/HtrA2 is a mitochondrial serine protease and a pro-apoptotic factor that plays a pivotal role in maintaining mitochondrial homeostasis[28]. Within the liver, Omi/HtrA2 mediates mitochondrial stability and autophagy, thus contributing to the amelioration of MASLD. Reportedly, the overexpression of HtrA2/Omi in the mouse liver enhances the expression of genes related to mitochondrial fatty acid β-

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oxidation, reduces hepatic lipid accumulation, and regulates glucose homeostasis^[19]. KLF16 tightly links hepatic lipid metabolism to mitochondrial homeostasis by regulating the transcriptional activity of PPARa. Knockdown of hepatic KLF16 also leads to increased mitochondrial stress and promotes the development of hepatic steatosis and insulin resistance in mice, whereas hepatic-specific PPARa overexpression effectively ameliorates hepatic steatosis induced by KLF16 deficiency and improves mitochondrial imbalance and insulin resistance [14]. The mitochondrial homeostasis is closely related associated with hepatic lipid metabolism and exacerbates the development of MASLD.

Bile acid-mediated metabolism homeostasis

Metabolic irregularities serve as both inducers and outcomes of the aging process and are intricately intertwined in the development of various diseases^[29]. Hepatic metabolism declines with aging and is accompanied by a reduction in enzymatic activity, owing to which the elderly are more susceptible to lipid accumulation. Bile acids (Bas), in conjunction with their homologous receptors such as the farnesoid X receptor (FXR) and Takeda G-protein-coupled receptor 5, play integral roles in numerous signaling pathways that are closely linked with MASLD. These pathways encompass BA selfregulation, glucose and lipid metabolism, energy modulation, cell proliferation, detoxification, and immune regulation [30]. BA analogs targeting FXR, TGR5, or both have been shown to effectively mitigate the progression of MASH[31].

BAs regulate metabolism by interacting with nuclear receptors and tightly modulating the diversity and relative abundance of the GM. The GM, along with their metabolic byproducts, may mediate MASLD by inducing endotoxemia, triggering insulin resistance, increasing short-chain fatty acids, elevating endogenous ethanol production, altering choline and BA metabolism, and impacting the host's immune response[32]. Therefore, the disruption of intracellular BA homeostasis may be a pivotal factor influencing the development of MASLD.

Dysbiosis

Aging results in the restructuring of the GM and is characterized by a decreased Firmicutes-to-Bacteroidetes ratio and reduced overall microbial diversity[33]. The lipopolysaccharide (LPS)/toll-like receptor 4 (TLR4) signaling pathway plays a crucial role in mediating the pathological mechanisms of MASLD. Patients with MASLD demonstrate an overgrowth of intestinal bacteria, which disrupts the intestinal barrier function. This altered gut barrier permeability leads to the translocation of LPS that triggers the activation of the LPS/TLR4/nuclear factor-κB (NF-κB) signaling pathway. That mediates the progression of MASLD to MASH[34]. A study has demonstrated that TLR4-deficient mice showed amelioration in insulin resistance and hepatic steatosis induced by HFD[35]. However, another study revealed that TLR5-deficient mice exhibited characteristics of metabolic syndrome, such as obesity, insulin resistance, and hepatic steatosis. Furthermore, transplanting the GM from TLR5-deficient mice into healthy mice exhibited the performance of metabolic syndrome in healthy mice[36].

Reportedly, GM additionally stimulates the generation of endogenous ethanol[37]. Dysbiosis within the gut microbiome can lead to the overgrowth of ethanol-producing bacteria, resulting in an increase in endogenous ethanol levels and subsequent induction of MASH. This has been demonstrated by Yuan et al [38], who isolated a high-alcoholproducing strain of Klebsiella pneumoniae (HiAlc Kpn) from the fecal samples of patients with Auto-brewery syndrome(ABS)/MASH and found that orally administering the strain to healthy sterile mice induced hepatic steatosis. Their results revealed that HiAlc-Kpn induced mouse MASLD model, the high-alcohol-producing strain of K. pneumoniae, upon colonization in the mouse gut, induced endogenous ethanol production that subsequently impaired the intestinal mucosal barrier. This resulted in heightened intestinal permeability in mice, which exacerbated inflammation in MASLD. Additionally, the transplantation of GM from younger mice to older mice could reverse age-related changes in the gut, eyes, and brain. Aged mice receiving young donor microbiota had reduced cortical and callosal microglia, reduced expression of inflammatory complement protein C3 in the retina, and reduced circulating concentrations of lipopolysaccharide (LPS)-binding protein (LBP), to levels comparable to young mice[39]. Furthermore, Hoyles et al[40] transplanted GM from patients with MASLD into mice maintained on a normal diet and found that the mice developed hepatic steatosis and their gut microbial characteristics realigned to those observed in MASLD.

POTENTIAL THERAPEUTIC TARGETS FOR MASLD

Regarding MASH treatment, this article predominantly focuses on clinical trials assessing agents targeting metabolic pathways, cellular stress responses, and interactions with GM.

Drugs targeting metabolism

FXR agonists: FXR plays a pivotal role in regulating lipid metabolism, BA homeostasis, and glucose equilibrium. The dysregulation of FXR function has been implicated in the pathogenesis of MASLD, cholestasis, and chronic inflammatory disorders that affect the liver and gastrointestinal tract. Despite a multitude of clinical trials targeting FXR for the management of MASLD/MASH, to date, only obeticholic acid has received approval for the treatment of primary biliary cholangitis. MET409[41] is another agent that has a unique non-BA composition and sustained pharmacokinetic/ pharmacodynamic properties. A study has assessed the efficacy of oral MET409 administered once daily over 12 wk in patients with MASH. By the end of the 12-week treatment cycle, MET409 remarkably reduced hepatic fat levels, with an average reduction of 55% (80 mg) and 38% (50 mg) compared with the 6% reduction observed with placebo (P < 0.001). Vonafexor (EYP001a) is a second-generation synthetic, non steroidal, non-bile salt, orally active carboxylic acid FXR agonist currently in development. In patients with MASH and suspected fibrosis, vonafexor has demonstrated efficacy in reducing hepatic fat content and improving liver enzymes. Specifically, 50.0% and 39.3% of patients treated with VONA-

100 mg and VONA-200 mg once daily, respectively, demonstrated a reduction in hepatic fat levels of > 30%, whereas only 12.5% of patients treated with placebo demonstrated this effect[42].

PPARs agonists: PPARs represent a subgroup of nuclear transcription factors activated by ligands and are categorically classified within the nuclear receptor superfamily. Post activation, PPARs form heterodimers with retinoid X receptors (RXRs). The resultant PPARy-RXR heterodimer binds to peroxisome proliferator response elements that are situated upstream of target gene promoters, consequently modulating the transcription of these specific target genes. The PPARs comprise PPAR α , PPAR β/δ , and PPAR γ that function as sensitive receptors for fatty acids and their derivatives, exerting crucial roles in lipid metabolism[43]. Notably, they can effectively mitigate hepatic steatosis, inflammation, and fibrosis in preclinical models of MASLD, thus underscoring their potential as promising targets for MASLD treatment. Selective agonists targeting PPARα and PPARγ have already demonstrated clinical efficacy, while clinical trials are currently assessing PPAR γ agonists such as pioglitazone; dual PPAR α/δ agonists such as chiglitazar, saroglitazar magnesium, and elafibranor; and pan-PPAR agonists such as lanifibranor. The activation of intestinal PPARa signaling plays a role in upregulating the expression of fatty acid-binding protein 1, thus facilitating intestinal fatty acid uptake and potentially contributing to the progression of MASH[44]. Elafibranor can improve steatosis, mitigate inflammation, and attenuate fibrosis in rodent models of MASLD/MASH[45]. Moreover, human liver in vitro models have unveiled that PPAR agonists can effectively diminish the increase in lipid levels, quell the secretion of inflammatory chemokines, and modulate the expression of pro-fibrotic genes *via* diverse mechanisms[46].

Glucagon-like peptide-1 agonists: The discovery of glucagon-like peptide-1 (GLP-1) represents a pivotal milestone in the field of biology, as the molecule significantly affects the regulation of blood glucose levels and the management of body weight. Cotadutide, an agonist of GLP-1R/GcgR, improves MASH and liver fibrosis by regulating mitochondrial function and lipid biosynthesis. Notably, in C6BL29/J mice exposed to an Amylin liver MASH diet for 57 wk, cotadutide displayed greater efficacy in treating MASH than cotadutide combined with obeticholic acid^[47]. Reportedly, the dual agonist GLP-1-Fc-FGF21 D1 exhibits notable and sustained hypoglycemic effects in diabetic mouse models. Moreover, in an HFD-induced ob/ob mouse model, GLP-1-Fc-FGF21 D1 has demonstrated robust anti-MASH properties via significant enhancements in liver function, alterations in serum and hepatic lipid profiles, and reduction in the MASLD activity score. Remarkably, its therapeutic efficacy surpasses that of singular FGF21 or GLP-1 analogs[48].

Thyroid hormone receptor-beta agonist: Activation of hepatic thyroid hormone receptor-beta (THR-β) reduces systemic lipid levels, enhances BA synthesis, and promotes lipid oxidation. Resmetirom (MGL-3196) is a liver-targeted and selective THR-β agonist. Studies have demonstrated that administering resmetirom (MGL-3196) to mice with dietinduced fibrotic DIO-MASH can lead to substantial reductions in liver weight, hepatic steatosis, plasma ALT levels, and hepatic and plasma cholesterol levels, as well as a decrease in blood glucose levels. Moreover, the treatment remarkably improved the MASLD activity score, with no discernible impact on body weight[49]. Compared with the placebo group, the resmetirom group was more effective in mitigating hepatic steatosis and ameliorating liver enzyme levels as well as inflammatory markers, resulting in a pronounced improvement in MASH as evidenced by liver biopsy assessments[50]. Furthermore, both 80 and 100 mg resmetirom result in favorable tolerability profiles, devoid of any severe or serious adverse events. The Phase 3 MASH clinical trial (NCT03900429) is now underway and is poised to enroll 900 volunteers with liver biopsy MASH with fibrosis grades I-III in a multinational, double-blind, randomized, placebo-controlled study to address the progression of MASH, cirrhosis, and/or hepatic decompensation (Figure 1).

Other agents: Sodium-dependent glucose co-transporter 2 (SGLT2) inhibitors are a novel class of antidiabetic drugs that inhibit glucose reabsorption in the proximal renal tubules, and have been shown to be effective in reducing hepatic fat content and AST/ALT levels and ameliorating hepatic fibrosis in several studies. Specifically, empagliflozin prevents MASLD progression in ApoE (-/-) mice by inducing autophagy via increased AMPK phosphorylation, reduced mTOR activity, and elevated LC3B expression. Furthermore, it mitigates endoplasmic reticulum stress and inhibits hepatocyte apoptosis[51]. Dapagliflozin, another SGLT2 inhibitor, has been found to activate AMPK and reduce mTOR phosphorylation in Zucker diabetic fatty rats. This effect has additionally been replicated in LO2 cells and HepG2 cells stimulated with palmitic acid. Consequently, the activation of AMPK promotes fatty acid oxidation and induces autophagy, ultimately improving hepatic steatosis[52]. Reportedly, fructose is catalyzed by ketohexokinase to produce fructose-1-phosphate, a metabolite that is primarily metabolized within the liver. Inhibition of fructose metabolism using ketohexokinase inhibitors can mitigate hepatic injury and fibrosis in both murine models and human subjects [53]. High fructose intake induces de novo lipid biosynthesis in the liver. This process does not depend on ATP citrate lyase (ACLY) but rather on the intestinal microflora which metabolizes fructose to acetate and converts the latter to acetyl coenzyme A (acetyl-CoA). Altered intestinal permeability, gut dysbiosis, and increased fructose intake exacerbate hepatic lipid accumulation and contribute to the development of MASLD in elderly patients[54].

Cell stress and cellular senescence

ROS inhibition: Excessive oxidative stress culminates in hepatocyte senescence, thereby instigating the accrual of hepatic fat[55]. Furthermore, hepatic lipotoxicity triggers oxidative stress and the release of inflammatory cytokines, thereby fostering the progression of MASH and, in more severe cases, fibrosis. Senolytics are a class of selective drugs that kill aging cells and can be used for the targeted intervention of cellular senescence. Quercetin, as an example, can elevate hepatic levels of SOD, catalase, and glutathione, concurrently decreasing interleukin (IL)-1 β , IL-6, tumor necrosis factor- α , and hepatic lipid accumulation in db/db mice. Furthermore, it can activate the FXR1/TGR5 signaling pathway, thereby contributing to the amelioration of MASLD[56]. In addition, quercetin demonstrates the ability to regulate GM dysbiosis and attenuate endotoxemia-induced upregulation of the TLR-4 pathway. This results in the inhibition of inflammasome

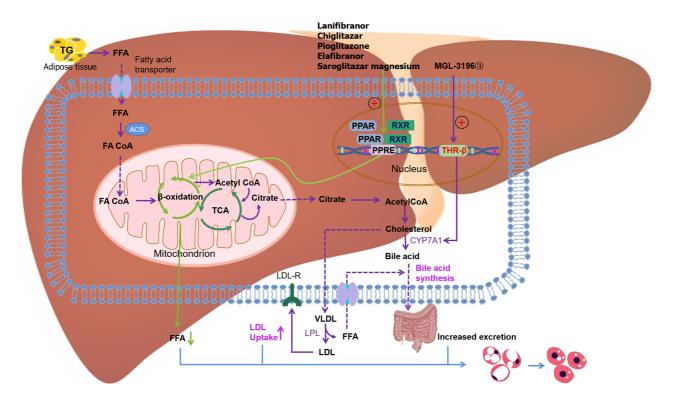


Figure 1 Signaling pathways of proliferator-activated receptor alpha and thyroid hormone receptor-beta and drugs targeting these pathways. PPAR: Proliferator-activated receptor; TG: Triglyceride; FFA: Free fatty acid; ACS: Acyl coenzyme A synthetase; FA CoA: Fatty acyl coenzyme A; TCA: Tricarboxylic acid cycle; RXR: Retinoid X receptor; PPRE: PPAR reaction element; THR: Thyroid hormone receptor-beta; LDL: Low-density lipoprotein; LDL-R: Lowdensity lipoprotein receptor; VLDL: Very low-density lipoprotein.

activation and stress pathway activation, which reinstates the equilibrium in lipid metabolism gene expression [57]. An ongoing clinical trial (NCT05506488) assessing a combination of dasatinib and quercetin for the clearance of senescent cells offers a potential avenue for addressing MASLD-associated fibrosis. N-acetylcysteine (NAC), an antioxidant with the capacity to reduce ROS levels and induce cellular apoptosis, can significantly reduce obesity, dyslipidemia, hepatic dysfunction, and GM dysbiosis induced by HFD in murine models. However, it is noteworthy that in these mice, using antibiotics for GM depletion resulted in a resurgence of hepatic steatosis and liver injury [58]. Another study has found that supplementing NAC to mice with diet-induced obesity and non-alcoholic steatohepatitis increases the CD4+ T cell population within liver tumor cells. Additionally, it elevates the levels of immunotherapeutic agents M30 and aOX40, thus effectively inhibiting the progression of liver tumors[59].

Myeloperoxidase inhibitor: Recent studies have revealed that patients with MASLD have elevated levels of plasma myeloperoxidase (MPO) and increased hepatic MPO expression compared with healthy controls. Notably, this elevation is more pronounced in patients with MASH. Studies have demonstrated that treatment with the MPO inhibitor AZM198 results in significantly reduced MASH-induced liver damage and fibrosis as well as decreased serum ALT levels and amelioration of hepatic steatosis[60]. Specifically in the elderly population, MASLD was associated with notable alterations in GM abundance, which resulted in compromised gut barrier function and heightened susceptibility to intestinal inflammation and subsequent systemic inflammatory responses. The administration of AZM198 improved the Firmicutes-to-Bacteroidetes ratio and regulated GM composition. Reportedly, TXNIP-deficient mice demonstrated decreased expression of inflammatory factors, reduced LPS levels, improved liver health, and restored intestinal barrier function. Notably, TXNIP is significantly upregulated in the intestinal mucosa of MASH mice. Moreover, studies have demonstrated that inhibiting the activation of the TXNIP-NLRP3 axis can effectively reduce MPO activity and oxidative stress, leading to the restoration of intestinal barrier function in the context of MASH. MPO Inhibitors reduce liver lipid accumulation, inflammation, and fibrosis and ameliorate the development of MASH.

PTUPB inhibitor: PTUPB, a dual inhibitor of soluble epoxide hydrolase and cyclooxygenase-2, exerts its effects by suppressing the PI3K/AKT/mTOR pathway via the modulation of Sirt3[61]. This leads to increased autophagy and decreased hepatocyte senescence, thus inducing the progression of MASLD. Reportedly, PTUPB can mitigate liver injury in HFD-induced MASLD murine models by inducing collagen deposition and lipid accrual; reducing hepatic triglyceride levels; and suppressing the expression of liver aging-associated molecules, such as p16, p53, and p21. Aspirin, an oral irreversible inhibitor of the cyclooxygenases COX-1 and COX-2, prevents the development of MASLD and MASH-related HCC[62].

Other agents: Lubiprostone functions by selectively stimulating type 2 chloride channels located on the apical cell membrane of gastrointestinal epithelial cells, leading to increased intestinal permeability. Research findings have

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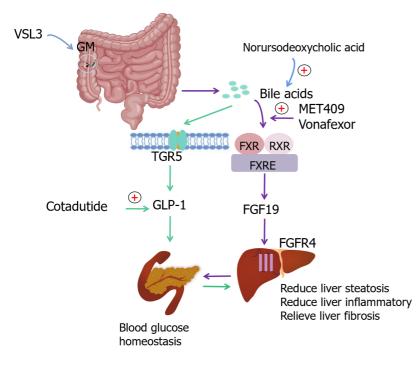


Figure 2 Targets related to lipids, bile acids, glucose homeostasis and intestinal microbiota in age-related metabolic dysfunctionassociated steatotic liver disease. GM: Gut microbiota; FXR: Farnesoid X receptor; RXR: Retinoid X receptor; FXRE: FXR reaction element; FGF19: Fibroblast growth factor 19; FGFR4: Fibroblast growth factor receptor 4; TGR5: Takeda G protein-coupled receptor 5; GLP-1: Glucagon-like peptide-1.

indicated that in patients with constipation and MASLD, lubiprostone reduces liver enzyme levels and exhibits favorable tolerability[63]. Heat shock protein (HSP) 47 is a collagen-specific molecular chaperone residing within the endoplasmic reticulum and plays a pivotal role in ensuring the correct folding, assembly, and extracellular secretion of collagen proteins within the extracellular matrix (ECM). The anomalous accumulation of collagen proteins within the ECM disrupts its structural integrity, thereby precipitating fibrotic processes[64]. These findings suggest that targeting HSP47 is of paramount importance for the treatment of liver fibrosis[65].

Drugs targeting the gut-liver axis

Probiotics: The GM assumes a crucial role in MASLD. Dysbiosis of the GM, attributed to oxidative stress, lifestyle choices, and excessive antibiotic use, leads to impaired intestinal permeability. An increased permeability facilitates the induction of pro-inflammatory cytokines and interferon-mediated factors through the activation of pattern recognition receptors by microbiota, bacterial byproducts, and LPS, thereby significantly contributing to the pathogenesis of MASLD [66]. Aging is concomitant with the deterioration of multiple physiological functions and the exacerbation of inflammatory processes. Age-related alterations in the gastrointestinal tract contribute to an elevated incidence of gastrointestinal inflammatory disorders. Aging is associated with increased production of ROS, which leads to lipid accumulation, DNA damage, and concomitant cellular functional impairments. Furthermore, modifications in GM composition result in the endogenous production of ethanol, which consequently compromises the integrity of the intestinal barrier and incites ROS accumulation in hepatic stellate cells and Kupffer cells[37]. Studies have shown that probiotics can restore homeostasis in the GM and mitigate oxidative stress[67]. Specifically, members of the Lactobacillus genus have demonstrated the ability to modulate the expression of inflammatory cytokines, including but not limited to IL-6, IL-1 β , IL-1 α , IL-12, and interferon- γ , both in serum and colonic tissues. This immunomodulatory effect is attributed to their capacity to inhibit NF-KB activation via the G protein-coupled receptor 109A pathway, thereby fostering improvements in immune function among aging mice[68]. In animal models of chronic liver injury, probiotics have demonstrated a protective role against hepatic steatosis and liver inflammation by modulating and potentially restoring the GM. Studies evaluating patients with MASH/MASLD have revealed that the probiotic formulation VSL3 displays inherent antiinflammatory properties and insulin-sensitizing effects, indicating its potential for the treatment of liver fibrosis[69]. Evidence suggests that GM can ameliorate an array of biomarkers associated with inflammation, blood glucose regulation, insulin resistance, lipid anomalies, obesity, and hepatic impairment, including reductions in liver enzymes, hepatic steatosis, and fibrosis. Probiotics exert their influence on the immune system, potentially improving MASLD outcomes, either by fortifying the intestinal barrier or preventing the formation of hepatotoxic metabolites[70] (Figure 2).

Weight loss surgery: Invasive weight loss surgeries may be associated with reduced tolerance among patients but demonstrate notable efficacy, particularly in severely obese individuals. A study assessing patients with severe obesity and MASH who underwent bariatric surgery revealed that 5 years post-surgery, 84% of the patients experienced resolution of inflammation, 70% demonstrated improvements in fibrosis, and 56% exhibited regression of liver fibrosis [71]. Bariatric surgery further modifies the ileal milieu, thereby inducing alterations in the GM or its metabolic products,

ultimately ameliorating MASLD.

CONCLUSION

The escalating trend of population aging underscores the increasing importance of investigating age-related diseases. Although research on MASLD has proliferated in recent years, the intricate mechanisms underlying the condition remain only partially elucidated. The factors precipitating aging exhibit reciprocal interactions with the pathogenesis and progression of MASLD. Oxidative stress and dysbiosis of the GM, for instance, can trigger both the aging process and inducing MASLD, which is frequently encountered in the elderly population. It is conceivable that such convergent targets, including oxidative stress and GM, may hold promise in formulating strategies for the management of agerelated MASLD.

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FOOTNOTES

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Country/Territory of origin: China

ORCID number: Qian-Jun He 0009-0004-0466-8515; Yi-Fei Li 0009-0005-9840-5229; Ling-Tong Zhao 0009-0008-1474-1391; Chun-Tong Lin 0009-0002-3778-0656; Chun-Yan Yu 0000-0002-5274-3370; Dan Wang 0000-0001-7756-710X.

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REFERENCES

- 1 López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: An expanding universe. Cell 2023; 186: 243-278 [PMID: 36599349 DOI: 10.1016/j.cell.2022.11.001]
- National Institutes of Health. National Institute on Aging, Global Health & Aging 2011. Available from: https://www.nia.nih.gov/research/ 2 publication/global-health-and-aging/health-and-work
- 3 Collado M, Blasco MA, Serrano M. Cellular senescence in cancer and aging. Cell 2007; 130: 223-233 [PMID: 17662938 DOI: 10.1016/j.cell.2007.07.003
- Huang W, Hickson LJ, Eirin A, Kirkland JL, Lerman LO. Cellular senescence: the good, the bad and the unknown. Nat Rev Nephrol 2022; 18: 4 611-627 [PMID: 35922662 DOI: 10.1038/s41581-022-00601-z]
- Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. Nat Med 5 2018; 24: 908-922 [PMID: 29967350 DOI: 10.1038/s41591-018-0104-9]
- Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic 6 assessment of prevalence, incidence, and outcomes. Hepatology 2016; 64: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- Estes C, Razavi H, Loomba R, Younossi Z, Sanyal AJ. Modeling the epidemic of nonalcoholic fatty liver disease demonstrates an exponential 7 increase in burden of disease. Hepatology 2018; 67: 123-133 [PMID: 28802062 DOI: 10.1002/hep.29466]
- Campisi J, d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol 2007; 8: 729-740 [PMID: 8 17667954 DOI: 10.1038/nrm2233]
- Radonjić T, Dukić M, Jovanović I, Zdravković M, Mandić O, Popadić V, Popović M, Nikolić N, Klašnja S, Divac A, Todorović Z, Branković 9 M. Aging of Liver in Its Different Diseases. Int J Mol Sci 2022; 23 [PMID: 36361873 DOI: 10.3390/ijms232113085]



- Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, Harrison SA, Brunt EM, Sanyal AJ. The diagnosis and management of 10 nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. Hepatology 2018; 67: 328-357 [PMID: 28714183 DOI: 10.1002/hep.29367]
- Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). Metabolism 2016; 65: 11 1038-1048 [PMID: 26823198 DOI: 10.1016/j.metabol.2015.12.012]
- Lujambio A, Akkari L, Simon J, Grace D, Tschaharganeh DF, Bolden JE, Zhao Z, Thapar V, Joyce JA, Krizhanovsky V, Lowe SW. Non-cell-12 autonomous tumor suppression by p53. Cell 2013; 153: 449-460 [PMID: 23562644 DOI: 10.1016/j.cell.2013.03.020]
- Laish I, Mannasse-Green B, Hadary R, Biron-Shental T, Konikoff FM, Amiel A, Kitay-Cohen Y. Telomere Dysfunction in Nonalcoholic Fatty 13 Liver Disease and Cryptogenic Cirrhosis. Cytogenet Genome Res 2016; 150: 93-99 [PMID: 28006764 DOI: 10.1159/000454654]
- Sun N, Shen C, Zhang L, Wu X, Yu Y, Yang X, Yang C, Zhong C, Gao Z, Miao W, Yang Z, Gao W, Hu L, Williams K, Liu C, Chang Y, Gao 14 Y. Hepatic Krüppel-like factor 16 (KLF16) targets PPARa to improve steatohepatitis and insulin resistance. Gut 2021; 70: 2183-2195 [PMID: 33257471 DOI: 10.1136/gutjnl-2020-321774]
- 15 Rubinsztein DC, Mariño G, Kroemer G. Autophagy and aging. Cell 2011; 146: 682-695 [PMID: 21884931 DOI: 10.1016/j.cell.2011.07.030]
- Liu A, Guo E, Yang J, Yang Y, Liu S, Jiang X, Hu Q, Dirsch O, Dahmen U, Zhang C, Gewirtz DA, Fang H. Young plasma reverses age-16 dependent alterations in hepatic function through the restoration of autophagy. Aging Cell 2018; 17 [PMID: 29210183 DOI: 10.1111/acel.12708]
- Hammoutene A, Biquard L, Lasselin J, Kheloufi M, Tanguy M, Vion AC, Mérian J, Colnot N, Loyer X, Tedgui A, Codogno P, Lotersztajn S, 17 Paradis V, Boulanger CM, Rautou PE. A defect in endothelial autophagy occurs in patients with non-alcoholic steatohepatitis and promotes inflammation and fibrosis. J Hepatol 2020; 72: 528-538 [PMID: 31726115 DOI: 10.1016/j.jhep.2019.10.028]
- 18 Xu J, Jiao K, Liu X, Sun Q, Wang K, Xu H, Zhang S, Wu Y, Wu L, Liu D, Wang W, Liu H. Omi/HtrA2 Participates in Age-Related Autophagic Deficiency in Rat Liver. Aging Dis 2018; 9: 1031-1042 [PMID: 30574416 DOI: 10.14336/AD.2018.0221]
- 19 Zhou W, Deng X, Zhu X, Yan Q, Zhou N, Du S, Li X. HtrA2/Omi mitigates NAFLD in high-fat-fed mice by ameliorating mitochondrial dysfunction and restoring autophagic flux. Cell Death Discov 2022; 8: 218 [PMID: 35449197 DOI: 10.1038/s41420-022-01022-4]
- 20 Lee DH, Park JS, Lee YS, Han J, Lee DK, Kwon SW, Han DH, Lee YH, Bae SH. SQSTM1/p62 activates NFE2L2/NRF2 via ULK1-mediated autophagic KEAP1 degradation and protects mouse liver from lipotoxicity. Autophagy 2020; 16: 1949-1973 [PMID: 31913745 DOI: 10.1080/15548627.2020.1712108]
- Salminen A, Kaarniranta K, Kauppinen A. Age-related changes in AMPK activation: Role for AMPK phosphatases and inhibitory 21 phosphorylation by upstream signaling pathways. Ageing Res Rev 2016; 28: 15-26 [PMID: 27060201 DOI: 10.1016/j.arr.2016.04.003]
- Tong L, Wang L, Yao S, Jin L, Yang J, Zhang Y, Ning G, Zhang Z. PPAR& attenuates hepatic steatosis through autophagy-mediated fatty acid 22 oxidation. Cell Death Dis 2019; 10: 197 [PMID: 30814493 DOI: 10.1038/s41419-019-1458-8]
- 23 Martínez-Cisuelo V, Gómez J, García-Junceda I, Naudí A, Cabré R, Mota-Martorell N, López-Torres M, González-Sánchez M, Pamplona R, Barja G. Rapamycin reverses age-related increases in mitochondrial ROS production at complex I, oxidative stress, accumulation of mtDNA fragments inside nuclear DNA, and lipofuscin level, and increases autophagy, in the liver of middle-aged mice. Exp Gerontol 2016; 83: 130-138 [PMID: 27498120 DOI: 10.1016/j.exger.2016.08.002]
- Cheng X, Ivessa AS. Accumulation of linear mitochondrial DNA fragments in the nucleus shortens the chronological life span of yeast. Eur J 24 Cell Biol 2012; 91: 782-788 [PMID: 22857949 DOI: 10.1016/j.ejcb.2012.06.005]
- Sagi H, Shibuya S, Kato T, Nakanishi Y, Tsuboi A, Moriya S, Ohno H, Miyamoto H, Kodama H, Shimizu T. SOD1 deficiency alters 25 gastrointestinal microbiota and metabolites in mice. Exp Gerontol 2020; 130: 110795 [PMID: 31805337 DOI: 10.1016/j.exger.2019.110795]
- 26 Miwa S, Kashyap S, Chini E, von Zglinicki T. Mitochondrial dysfunction in cell senescence and aging. J Clin Invest 2022; 132 [PMID: 35775483 DOI: 10.1172/JCI158447]
- Di Ciaula A, Passarella S, Shanmugam H, Noviello M, Bonfrate L, Wang DQ, Portincasa P. Nonalcoholic Fatty Liver Disease (NAFLD). 27 Mitochondria as Players and Targets of Therapies? Int J Mol Sci 2021; 22 [PMID: 34065331 DOI: 10.3390/ijms22105375]
- Hegde R, Srinivasula SM, Zhang Z, Wassell R, Mukattash R, Cilenti L, DuBois G, Lazebnik Y, Zervos AS, Fernandes-Alnemri T, Alnemri 28 ES. Identification of Omi/HtrA2 as a mitochondrial apoptotic serine protease that disrupts inhibitor of apoptosis protein-caspase interaction. J Biol Chem 2002; 277: 432-438 [PMID: 11606597 DOI: 10.1074/jbc.M109721200]
- Khosla S, Farr JN, Tchkonia T, Kirkland JL. The role of cellular senescence in ageing and endocrine disease. Nat Rev Endocrinol 2020; 16: 29 263-275 [PMID: 32161396 DOI: 10.1038/s41574-020-0335-y]
- Perino A, Demagny H, Velazquez-Villegas L, Schoonjans K. Molecular Physiology of Bile Acid Signaling in Health, Disease, and Aging. 30 Physiol Rev 2021; 101: 683-731 [PMID: 32790577 DOI: 10.1152/physrev.00049.2019]
- Xue R, Su L, Lai S, Wang Y, Zhao D, Fan J, Chen W, Hylemon PB, Zhou H. Bile Acid Receptors and the Gut-Liver Axis in Nonalcoholic 31 Fatty Liver Disease. Cells 2021; 10 [PMID: 34831031 DOI: 10.3390/cells10112806]
- Chen J, Vitetta L. Gut Microbiota Metabolites in NAFLD Pathogenesis and Therapeutic Implications. Int J Mol Sci 2020; 21 [PMID: 32 32717871 DOI: 10.3390/ijms21155214]
- Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, Marchesi JR, Falush D, Dinan T, Fitzgerald G, Stanton C, 33 van Sinderen D, O'Connor M, Harnedy N, O'Connor K, Henry C, O'Mahony D, Fitzgerald AP, Shanahan F, Twomey C, Hill C, Ross RP, O'Toole PW. Composition, variability, and temporal stability of the intestinal microbiota of the elderly. Proc Natl Acad Sci US A 2011; 108 Suppl 1: 4586-4591 [PMID: 20571116 DOI: 10.1073/pnas.1000097107]
- Soares JB, Pimentel-Nunes P, Roncon-Albuquerque R, Leite-Moreira A. The role of lipopolysaccharide/toll-like receptor 4 signaling in 34 chronic liver diseases. Hepatol Int 2010; 4: 659-672 [PMID: 21286336 DOI: 10.1007/s12072-010-9219-x]
- Jia L, Vianna CR, Fukuda M, Berglund ED, Liu C, Tao C, Sun K, Liu T, Harper MJ, Lee CE, Lee S, Scherer PE, Elmquist JK. Hepatocyte 35 Toll-like receptor 4 regulates obesity-induced inflammation and insulin resistance. Nat Commun 2014; 5: 3878 [PMID: 24815961 DOI: 10.1038/ncomms4878]
- Vijay-Kumar M, Aitken JD, Carvalho FA, Cullender TC, Mwangi S, Srinivasan S, Sitaraman SV, Knight R, Ley RE, Gewirtz AT. Metabolic 36 syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. Science 2010; 328: 228-231 [PMID: 20203013 DOI: 10.1126/science.1179721
- Zhu L, Baker SS, Gill C, Liu W, Alkhouri R, Baker RD, Gill SR. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) 37 patients: a connection between endogenous alcohol and NASH. Hepatology 2013; 57: 601-609 [PMID: 23055155 DOI: 10.1002/hep.26093]
- Yuan J, Chen C, Cui J, Lu J, Yan C, Wei X, Zhao X, Li N, Li S, Xue G, Cheng W, Li B, Li H, Lin W, Tian C, Zhao J, Han J, An D, Zhang Q, 38



Wei H, Zheng M, Ma X, Li W, Chen X, Zhang Z, Zeng H, Ying S, Wu J, Yang R, Liu D. Fatty Liver Disease Caused by High-Alcohol-Producing Klebsiella pneumoniae. Cell Metab 2019; 30: 675-688.e7 [PMID: 31543403 DOI: 10.1016/j.cmet.2019.08.018]

- 39 Parker A, Romano S, Ansorge R, Aboelnour A, Le Gall G, Savva GM, Pontifex MG, Telatin A, Baker D, Jones E, Vauzour D, Rudder S, Blackshaw LA, Jeffery G, Carding SR. Fecal microbiota transfer between young and aged mice reverses hallmarks of the aging gut, eye, and brain. Microbiome 2022; 10: 68 [PMID: 35501923 DOI: 10.1186/s40168-022-01243-w]
- Hoyles L, Fernández-Real JM, Federici M, Serino M, Abbott J, Charpentier J, Heymes C, Luque JL, Anthony E, Barton RH, Chilloux J, 40 Myridakis A, Martinez-Gili L, Moreno-Navarrete JM, Benhamed F, Azalbert V, Blasco-Baque V, Puig J, Xifra G, Ricart W, Tomlinson C, Woodbridge M, Cardellini M, Davato F, Cardolini I, Porzio O, Gentileschi P, Lopez F, Foufelle F, Butcher SA, Holmes E, Nicholson JK, Postic C, Burcelin R, Dumas ME. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. Nat Med 2018; 24: 1070-1080 [PMID: 29942096 DOI: 10.1038/s41591-018-0061-3]
- 41 Harrison SA, Bashir MR, Lee KJ, Shim-Lopez J, Lee J, Wagner B, Smith ND, Chen HC, Lawitz EJ. A structurally optimized FXR agonist, MET409, reduced liver fat content over 12 weeks in patients with non-alcoholic steatohepatitis. J Hepatol 2021; 75: 25-33 [PMID: 33581174 DOI: 10.1016/j.jhep.2021.01.047]
- Ratziu V, Harrison SA, Loustaud-Ratti V, Bureau C, Lawitz E, Abdelmalek M, Alkhouri N, Francque S, Girma H, Darteil R, Couchoux H, 42 Wolf M, Sanyal A, Vonderscher J, Scalfaro P. Hepatic and renal improvements with FXR agonist vonafexor in individuals with suspected fibrotic NASH. J Hepatol 2023; 78: 479-492 [PMID: 36334688 DOI: 10.1016/j.jhep.2022.10.023]
- Pyper SR, Viswakarma N, Yu S, Reddy JK. PPARalpha: energy combustion, hypolipidemia, inflammation and cancer. Nucl Recept Signal 43 2010; 8: e002 [PMID: 20414453 DOI: 10.1621/nrs.08002]
- 44 Yan T, Luo Y, Yan N, Hamada K, Zhao N, Xia Y, Wang P, Zhao C, Qi D, Yang S, Sun L, Cai J, Wang Q, Jiang C, Gavrilova O, Krausz KW, Patel DP, Yu X, Wu X, Hao H, Liu W, Qu A, Gonzalez FJ. Intestinal peroxisome proliferator-activated receptor a-fatty acid-binding protein 1 axis modulates nonalcoholic steatohepatitis. Hepatology 2023; 77: 239-255 [PMID: 35460276 DOI: 10.1002/hep.32538]
- 45 Staels B, Rubenstrunk A, Noel B, Rigou G, Delataille P, Millatt LJ, Baron M, Lucas A, Tailleux A, Hum DW, Ratziu V, Cariou B, Hanf R. Hepatoprotective effects of the dual peroxisome proliferator-activated receptor alpha/delta agonist, GFT505, in rodent models of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. Hepatology 2013; 58: 1941-1952 [PMID: 23703580 DOI: 10.1002/hep.26461]
- Boeckmans J, Natale A, Rombaut M, Buyl K, Cami B, De Boe V, Heymans A, Rogiers V, De Kock J, Vanhaecke T, Rodrigues RM. Human 46 hepatic in vitro models reveal distinct anti-NASH potencies of PPAR agonists. Cell Biol Toxicol 2021; 37: 293-311 [PMID: 32613381 DOI: 10.1007/s10565-020-09544-2]
- 47 Boland ML, Laker RC, Mather K, Nawrocki A, Oldham S, Boland BB, Lewis H, Conway J, Naylor J, Guionaud S, Feigh M, Veidal SS, Lantier L, McGuinness OP, Grimsby J, Rondinone CM, Jermutus L, Larsen MR, Trevaskis JL, Rhodes CJ. Resolution of NASH and hepatic fibrosis by the GLP-1R/GcgR dual-agonist Cotadutide via modulating mitochondrial function and lipogenesis. Nat Metab 2020; 2: 413-431 [PMID: 32478287 DOI: 10.1038/s42255-020-0209-6]
- Pan Q, Lin S, Li Y, Liu L, Li X, Gao X, Yan J, Gu B, Chen X, Li W, Tang X, Chen C, Guo L. A novel GLP-1 and FGF21 dual agonist has 48 therapeutic potential for diabetes and non-alcoholic steatohepatitis. EBioMedicine 2021; 63: 103202 [PMID: 33421947 DOI: 10.1016/j.ebiom.2020.103202]
- Kannt A, Wohlfart P, Madsen AN, Veidal SS, Feigh M, Schmoll D. Activation of thyroid hormone receptor- β improved disease activity and 49 metabolism independent of body weight in a mouse model of non-alcoholic steatohepatitis and fibrosis. Br J Pharmacol 2021; 178: 2412-2423 [PMID: 33655500 DOI: 10.1111/bph.15427]
- Harrison SA, Bashir M, Moussa SE, McCarty K, Pablo Frias J, Taub R, Alkhouri N. Effects of Resmetirom on Noninvasive Endpoints in a 50 36-Week Phase 2 Active Treatment Extension Study in Patients With NASH. Hepatol Commun 2021; 5: 573-588 [PMID: 33860116 DOI: 10.1002/hep4.1657]
- Nasiri-Ansari N, Nikolopoulou C, Papoutsi K, Kyrou I, Mantzoros CS, Kyriakopoulos G, Chatzigeorgiou A, Kalotychou V, Randeva MS, 51 Chatha K, Kontzoglou K, Kaltsas G, Papavassiliou AG, Randeva HS, Kassi E. Empagliflozin Attenuates Non-Alcoholic Fatty Liver Disease (NAFLD) in High Fat Diet Fed ApoE((-/-)) Mice by Activating Autophagy and Reducing ER Stress and Apoptosis. Int J Mol Sci 2021; 22 [PMID: 33467546 DOI: 10.3390/ijms22020818]
- Li L, Li Q, Huang W, Han Y, Tan H, An M, Xiang Q, Zhou R, Yang L, Cheng Y. Dapagliflozin Alleviates Hepatic Steatosis by Restoring 52 Autophagy via the AMPK-mTOR Pathway. Front Pharmacol 2021; 12: 589273 [PMID: 34093169 DOI: 10.3389/fphar.2021.589273]
- Shepherd EL, Saborano R, Northall E, Matsuda K, Ogino H, Yashiro H, Pickens J, Feaver RE, Cole BK, Hoang SA, Lawson MJ, Olson M, 53 Figler RA, Reardon JE, Nishigaki N, Wamhoff BR, Günther UL, Hirschfield G, Erion DM, Lalor PF. Ketohexokinase inhibition improves NASH by reducing fructose-induced steatosis and fibrogenesis. JHEP Rep 2021; 3: 100217 [PMID: 33490936 DOI: 10.1016/j.jhepr.2020.100217]
- Zhao S, Jang C, Liu J, Uehara K, Gilbert M, Izzo L, Zeng X, Trefely S, Fernandez S, Carrer A, Miller KD, Schug ZT, Snyder NW, Gade TP, 54 Titchenell PM, Rabinowitz JD, Wellen KE. Dietary fructose feeds hepatic lipogenesis via microbiota-derived acetate. Nature 2020; 579: 586-591 [PMID: 32214246 DOI: 10.1038/s41586-020-2101-7]
- Ogrodnik M, Miwa S, Tchkonia T, Tiniakos D, Wilson CL, Lahat A, Day CP, Burt A, Palmer A, Anstee QM, Grellscheid SN, Hoeijmakers 55 JHJ, Barnhoorn S, Mann DA, Bird TG, Vermeij WP, Kirkland JL, Passos JF, von Zglinicki T, Jurk D. Cellular senescence drives agedependent hepatic steatosis. Nat Commun 2017; 8: 15691 [PMID: 28608850 DOI: 10.1038/ncomms15691]
- Yang H, Yang T, Heng C, Zhou Y, Jiang Z, Qian X, Du L, Mao S, Yin X, Lu Q. Quercetin improves nonalcoholic fatty liver by ameliorating 56 inflammation, oxidative stress, and lipid metabolism in db/db mice. Phytother Res 2019; 33: 3140-3152 [PMID: 31452288 DOI: 10.1002/ptr.6486]
- 57 Porras D, Nistal E, Martínez-Flórez S, Pisonero-Vaquero S, Olcoz JL, Jover R, González-Gallego J, García-Mediavilla MV, Sánchez-Campos S. Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. Free Radic Biol Med 2017; 102: 188-202 [PMID: 27890642 DOI: 10.1016/j.freeradbiomed.2016.11.037]
- Ding Q, Guo R, Pei L, Lai S, Li J, Yin Y, Xu T, Yang W, Song Q, Han Q, Dou X, Li S. N-Acetylcysteine alleviates high fat diet-induced 58 hepatic steatosis and liver injury via regulating the intestinal microecology in mice. Food Funct 2022; 13: 3368-3380 [PMID: 35229847 DOI: 10.1039/d1fo03952k
- Heinrich B, Brown ZJ, Diggs LP, Vormehr M, Ma C, Subramanyam V, Rosato U, Ruf B, Walz JS, McVey JC, Wabitsch S, Fu Q, Yu SJ, 59 Zhang Q, Lai CW, Sahin U, Greten TF. Steatohepatitis Impairs T-cell-Directed Immunotherapies Against Liver Tumors in Mice. Gastroenterology 2021; 160: 331-345.e6 [PMID: 33010248 DOI: 10.1053/j.gastro.2020.09.031]



- Koop AC, Thiele ND, Steins D, Michaëlsson E, Wehmeyer M, Scheja L, Steglich B, Huber S, Schulze Zur Wiesch J, Lohse AW, Heeren J, 60 Kluwe J. Therapeutic Targeting of Myeloperoxidase Attenuates NASH in Mice. Hepatol Commun 2020; 4: 1441-1458 [PMID: 33024915 DOI: 10.1002/hep4.1566]
- Zhang CY, Tan XH, Yang HH, Jin L, Hong JR, Zhou Y, Huang XT. COX-2/sEH Dual Inhibitor Alleviates Hepatocyte Senescence in NAFLD 61 Mice by Restoring Autophagy through Sirt1/PI3K/AKT/mTOR. Int J Mol Sci 2022; 23 [PMID: 35897843 DOI: 10.3390/ijms23158267]
- Lange NF, Radu P, Dufour JF. Prevention of NAFLD-associated HCC: Role of lifestyle and chemoprevention. J Hepatol 2021; 75: 1217-1227 62 [PMID: 34339764 DOI: 10.1016/j.jhep.2021.07.025]
- Kessoku T, Imajo K, Kobayashi T, Ozaki A, Iwaki M, Honda Y, Kato T, Ogawa Y, Tomeno W, Kato S, Higurashi T, Yoneda M, Kirikoshi H, 63 Kubota K, Taguri M, Yamanaka T, Usuda H, Wada K, Kobayashi N, Saito S, Nakajima A. Lubiprostone in patients with non-alcoholic fatty liver disease: a randomised, double-blind, placebo-controlled, phase 2a trial. Lancet Gastroenterol Hepatol 2020; 5: 996-1007 [PMID: 32805205 DOI: 10.1016/S2468-1253(20)30216-8]
- 64 Friedman SL. Mechanisms of hepatic fibrogenesis. Gastroenterology 2008; 134: 1655-1669 [PMID: 18471545 DOI: 10.1053/j.gastro.2008.03.003]
- Abd El-Fattah EE, Zakaria AY. Targeting HSP47 and HSP70: promising therapeutic approaches in liver fibrosis management. J Transl Med 65 2022; **20**: 544 [PMID: 36435779 DOI: 10.1186/s12967-022-03759-z]
- Bai L, Li H. Innate immune regulatory networks in hepatic lipid metabolism. J Mol Med (Berl) 2019; 97: 593-604 [PMID: 30891617 DOI: 66 10.1007/s00109-019-01765-1]
- Campagnoli LIM, Marchesi N, Vairetti M, Pascale A, Ferrigno A, Barbieri A. Age-Related NAFLD: The Use of Probiotics as a Supportive 67 Therapeutic Intervention. Cells 2022; 11 [PMID: 36139402 DOI: 10.3390/cells11182827]
- Vemuri R, Gundamaraju R, Shinde T, Perera AP, Basheer W, Southam B, Gondalia SV, Karpe AV, Beale DJ, Tristram S, Ahuja KDK, Ball 68 M, Martoni CJ, Eri R. Lactobacillus acidophilus DDS-1 Modulates Intestinal-Specific Microbiota, Short-Chain Fatty Acid and Immunological Profiles in Aging Mice. Nutrients 2019; 11 [PMID: 31181695 DOI: 10.3390/nu11061297]
- 69 Velayudham A, Dolganiuc A, Ellis M, Petrasek J, Kodys K, Mandrekar P, Szabo G. VSL#3 probiotic treatment attenuates fibrosis without changes in steatohepatitis in a diet-induced nonalcoholic steatohepatitis model in mice. Hepatology 2009; 49: 989-997 [PMID: 19115316 DOI: 10.1002/hep.22711]
- Carpi RZ, Barbalho SM, Sloan KP, Laurindo LF, Gonzaga HF, Grippa PC, Zutin TLM, Girio RJS, Repetti CSF, Detregiachi CRP, Bueno 70 PCS, Mazuqueli Pereira ESB, Goulart RA, Haber JFDS. The Effects of Probiotics, Prebiotics and Synbiotics in Non-Alcoholic Fat Liver Disease (NAFLD) and Non-Alcoholic Steatohepatitis (NASH): A Systematic Review. Int J Mol Sci 2022; 23 [PMID: 35955942 DOI: 10.3390/iims231588051
- Lassailly G, Caiazzo R, Ntandja-Wandji LC, Gnemmi V, Baud G, Verkindt H, Ningarhari M, Louvet A, Leteurtre E, Raverdy V, Dharancy S, 71 Pattou F, Mathurin P. Bariatric Surgery Provides Long-term Resolution of Nonalcoholic Steatohepatitis and Regression of Fibrosis. Gastroenterology 2020; 159: 1290-1301.e5 [PMID: 32553765 DOI: 10.1053/j.gastro.2020.06.006]





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