

Influence of splanchnic vascular infusion on the content of endotoxins in plasma and the translocation of intestinal bacteria in rats with acute hemorrhage necrosis pancreatitis

Ren Yi Qin, Sheng Quan Zou, Zai De Wu and Fa Zu Qiu

Subject headings acute hemorrhage necrosis pancreatitis; microcirculation/splanchnic organ; endo toxins/plasma; intestinal bacterial trans-location

Qin RY, Zou SQ, Wu ZD, Qiu FZ. Influence of splanchnic vascular infusion on the content of endotoxins in plasma and the translocation of intestinal bacteria in rats with acute hemorrhage necrosis pancreatitis. *World J Gastroentero*, 2000;6(4):577-580

INTRODUCTION

The main reason for the death of the patient with acute hemorrhage necrosis pancreatitis (AHNP) is pancreatic infection and multi-organ failure caused by endotoxemia and intestinal bacterial translocation^[1-7]. However, the pathogenesis of endotoxemia and intestinal bacterial translocation remains a question^[8-10]; moreover, no effective method of prevention and cure for it has been found till now^[11-15]. In the present study, we infused low dose dopamine and low molecular weight dextran through the catheters to abdominal aorta and portal vein, and observed its influence on the endotoxin concentration in plasma and the rate of translocation of intestinal bacteria in AHNP rats.

MATERIALS AND METHODS

Animals

A total of 48 Sprague-Dawley rats weighing 295-320 g were divided into 4 groups (with 12 rats in each group): Group A (healthy rats), Group B (AHNP rats), Group C (femoral artery and femoral vein infused rats), and Group D (abdominal aorta and portal vein infused rats).

Department of Surgery, Tongji Hospital, Tongji Medical University, Wuhan 430030, Hubei Province, China

Dr. QIN Ren-Yi, male, born in 1963-10-04, in Zhun Yi City, Gui Zhou Province. Graduated from Gui Yang Medical College in 1986. Receiving the degree of Medical Doctor (Zhejiang Medical University) in 1995, he is now an Associate Professor and is doing Postdoctoral research since 1997, and has 21 paper published.

Supported by the China Postdoctoral Sciences Foundation No C.P.S.F 1996. 2[#]

Correspondence to: Dr. Ren-Yi Qin, Department of Surgery, Tongji Hospital, Tongji Medical University, Wuhan 430030, Hubei Province, China

Tel. 0086-27-8366-2389, Fax. 0086-27-8364-6605

Email. ryqin@tjh.tjmu.edu.cn

Received 2000-02-13 **Accepted** 2000-03-05

Experimental methods

Bacterial labeling Following the way previously described by Wells, we directly labeled O₅₅B₅ Escherichia coli (*E. coli*) with fluorescein isothiocyanate to prepare the solution of 5×10^6 cfu/L tracer.

Induction of AHNP model and blood vessels catheter insertion and infusion

Firstly, all rats were deprived of food 24 h before laparotomy, and were given a gavage of fluorescein-labeled *E. Coli* (0.7 mL/100 g) 12 h later, then they were kept for 12 h before being anesthetized ip with 2% pentobarbital sodium (0.15 mL/100 g). Secondly, after the abdominal hair was removed and the abdominal cavity was opened through a midline laparotomy, the pancreas was exposed and 5% taurodexychoic acid sodium solution (0.15 mL/100 g; Sigma) was slowly injected into the pancreatic duct with retrograde pressure. Five minutes later, hemorrhage, necrosis, swelling and exudation appeared in the pancreas. Thirdly, 2 h after the AHNP models were completed, the animals of group B were infused continuously with saline through the catheters which were connected to the femoral artery and femoral vein and portal vein (the catheter was inserted from the ileocolic vein to the main trunk of portal vein), the animals of group C were infused continuously and alternately with low dose dopamine ($5 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$) and low molecular weight dextran (1.5 mL through catheters inserted into femoral artery and femoral vein, while being infused continuously with saline through the portal vein. The animals of Group D were infused continuously and alternately with low dose dopamine and low molecular weight dextran through the portal vein and abdominal aorta (catheters were inserted from the femoral artery and above the junction between the abdominal aorta and abdominal cavity artery), while being infused continuously with saline through the femoral vein. The total amount of infusion (6 mL/100 g) was the same in each group. Lastly, after 4 h of sustained infusion, the abdominal cavity was found to be hemorrhagic on being re-opened, and it was more evident in both Group B and Group C than in Group D. The pancreas showed pathological changes such as hemorrhage, necrosis, swelling and exudation, while Group A (control group) had no pathological changes.

Indicators and methods

Plasma endotoxin One mL vein blood was put into the heparin-containing test tube under aseptic and non-pyrogenic conditions, and after being centrifuged 500 rpm at for 10 min, the plasma was absorbed and preserved in a refrigerator at -20°C . The content of plasma endotoxin was investigated by quantitative azostromatic coloration limulus test microassay (kit from Institute of Medicine in Shanghai).

Investigation of mesenteric lymph node (MLN) labeled-bacteria MLN of the ileocecum were excised, weighed, triturated and diluted into 10% tissue plasma and was observed under fluorescein microscope for the existence of labeled-bacteria. The bacterial translocation rate was also calculated.

The microcirculation of pancreas and mesentery

The diameter of pancreatic and mesenteric small vein at the ileocecum end was directly measured under the microscope and recorded on the video-camera.

Pathological changes in the intestinal mucosa:

Observed under light and transmission electron microscopy.

RESULTS

Changes in content of plasma endotoxin and rate of translocation of labeled-bacteria in MLN (Table 1)

Table 1 Changes in plasma endotoxin concentration ($\bar{x} \pm S_x$, EU/mL) and the rate of translocation of labeled-bacteria in MLN

Group	Plasma endotoxin (2 h)	Plasma endotoxin (6 h)	Rate of translocation of labeled-bacteria (%)
A	0.023 ± 0.004	0.033 ± 0.006	0
B	0.028 ± 0.002	0.340 ± 0.038	91
C	0.025 ± 0.007	0.270 ± 0.048	83
D	0.027 ± 0.001	0.103 ± 0.018	33

There was a significant statistical difference between group D and either group B or C ($P < 0.05$), regarding the content of plasma endotoxin and translocation rate of MLN labeled-bacteria. However, there is no statistical difference between group B and group C ($P > 0.05$), while a significant difference exists when group A is compared with either group B, C, or D ($P < 0.05$).

Microcirculatory changes in pancreas and mesentery (Table 2)

Table 2 Change in the mesenteric small vein diameter after 2 h and 4 h AHNP

Group	2 h MVD	6 h MVD
A	0.66 ± 0.04	0.63 ± 0.04
B	0.69 ± 0.05	1.03 ± 0.05
C	0.72 ± 0.03	1.09 ± 0.05
D	0.77 ± 0.07	0.68 ± 0.05

Between 2 h and 4 h after AHNP, in Group B and Group C, the diameter of pancreatic and mesenteric small vein increased significantly ($P < 0.05$), and the velocity of blood was observed to be retarded or even blocked, while there was no significant increase in the diameters of pancreatic and mesenteric small veins in group D ($P > 0.05$) and no statistical difference regarding the velocity of blood stream between group D and group A ($P > 0.05$).

Pathological changes in intestinal mucosa

Optical microscopic observation It was seen that a large-number of mucosal chorionic epithelium were exfoliated, the upper parts of villus intestina were in significant edema, the central chylectasia was expanded, the blood vessels congested, the proprietary membrane was in moderate edema and the inflammatory cells infiltrated in group B and group C. While the damage of mucosa in group D was alleviated as compared to group B or group C, it was seen that only the villus became shorter, the proprietary membrane was in edema and the inflammatory cells infiltrated.

Electronmicroscopic observation Rarefaction and exfoliation of the epithelium microvilli of intestinal mucosa, exudation of matrix vacuolar degeneration of mitochondria, swelling of endoplasmic reticulum, and break-down of epithelium bridges were observed in groups B and C; and in group D only slight derangement of intestinal mucosa epithelium and slight swelling of mitochondria and endoplasmic reticulum were seen.

DISCUSSION

An extensive amount of experimental and clinical work reveals that the disorder of pancreatic microcirculation, the production of many inflammatory mediators and cytokines and the translocation of intestinal bacteria are all thought to play a critical role in the pathogenesis of acute hemorrhage necrosis pancreatitis^[15-26]; furthermore, the disorder of splanchnic organic microcirculation, especially the disorder of pancreas microcirculation in AHNP is closely connected with the production of many inflammatory mediators and cytokines^[26-37]. Dopamine has been seen to possess complicate pharmacological functions^[38-40], in above $10 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$, alpha adrenergic receptors are additionally activated, causing splanchnic vascular contraction. At a dose range of $1-4 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$, the effect is predominantly on dopaminergic receptors, leading to splanchnic dilatation. At the $4-10 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$, beta adrenergic receptors are increasingly stimulated, which stops the increase in microvascular permeability caused by histamine and bradykinin. It also activates dopaminergic receptors leading to an increase of blood flow in splanchnic organs. Low molecular weight dextran can lower blood viscosity and hemagglutination leading to a halt in microvascular thrombogenesis in portal vein system^[41-42]. Therefore, low dose dopamine and low molecular weight dextran can be used to improve splanchnic

microcirculation. With the aid of catheters inserted into the aorta and portal vein, we infused low dose dopamine and low molecular weight dextran continuously and alternatively, which enhances the drug concentration in pancreas, liver and intestinal tract. The results have revealed that by this method the microcirculation was improved, and the content of endotoxin and the rate of intestinal bacterial translocation were decreased with increasing blood supply to the pancreas and intestinal tract of AHNP rats. This effect can be related to the following factors: Firstly, low dose dopamine and low molecular weight dextran can directly improve the ischemic status of intestinal tract and inhibit the damage of the barrier function of intestinal mucosa. It can also inhibit the pancreatic hemorrhage, necrosis and decrease the production of inflammatory mediators by improving microcirculation of pancreas, liver, and intestinal tract. On the other hand they can lighten the damage of inflammatory mediators and endotoxin on intestinal tract by enhancing the ability of the liver in clearing from inflammatory mediators and endotoxin. Hence, the method can alleviate the injury of intestinal mucosa and protect its barrier function, and inhibit endotoxemia and translocation of the intestinal bacteria, and so indicate that the disorder of microcirculation of pancreas and intestinal tract and liver are critically important to endotoxemia and the bacterial translocation from the intestine. The study also shows that the infusion through catheter to femoral vein and artery has no obvious influence on the content of endotoxin in plasma and the bacterial translocation in AHNP rats, the reason of which may be related to the low concentration of drugs in pancreas, liver and intestinal tract.

To achieve clearance of inflammatory mediators in patients with AHNP, in addition to drainage and removing the necrotic tissues by operation, we can also infuse low dose dopamine and low molecular weight dextran into the abdominal cavity after operation through catheter inserted either from the right gastroduodenal vein to the portal vein or from the femoral artery, which can improve the microcirculation disorder of pancreas, liver and intestinal tract. Moreover, we can infuse enzyme inhibitors and other anti-inflammatory mediators through the catheter to the portal vein, so as to eliminate inflammatory mediators before they reach the liver.

REFERENCES

- Runkel NS, Moody FG, Smith GS, Rodriguez LF, LaRocco MT, Miller TA. The role of the gut in the development of sepsis in acute pancreatitis. *J Surg Res*, 1991;51:18-23
- Runkel NS, Rodriguez LF, Moody FG. Mechanisms of sepsis in acute pancreatitis in opossums. *Am J Surg*, 1995;169:227-232
- Andren Sandberg A, Hojer H. Necrotizing acute pancreatitis induced by Salmonella infection. *Int J Pancreatol*, 1994;15:229-300
- Gianotti L, Munda R, Alexander JW, Tchervenkov JI, Babcock GF. Bacterial translocation: a potential source for infection in acute pancreatitis. *Pancreas*, 1993;8:551-558
- Iwasaki G, Takeda K, Sunamura M, Kobari M, Matsuno S. The role of intestinal flora in the pathogenesis of infection and aggravation of experimental acute pancreatitis in rats. *Nippon Geka Gakkai Zasshi*, 1994;95:669-77
- Shu ZJ, Li WQ, Wang XB, Wang ZM, Wang SH, Wang L, Du JX, Li JS. Gastrointestinal tract complications in severe acute pancreatitis. *World J Gastroenterol*, 1998;4(Suppl2):98-100
- Wu XN. Current concept of pathogenesis of severe acute pancreatitis. *World J Gastroenterol*, 2000;6:32
- Medich DS, Lee TK, Melhem MF, Rowe MI, Schraut WH, Lee KK. Pathogenesis of pancreatic sepsis. *Am J Surg*, 1993;165:46-50
- Kazantsev GB, Hecht DW, Rao R, Fedorak IJ, Gattuso P, Thompson K, Djuricin G, Prinz RA. Plasmid labeling confirms bacterial translocation in pancreatitis. *Am J Surg*, 1994;167:201-206
- Widdison AL, Karanjia ND, Reber HA. Routes of spread of pathogens into the pancreas in a feline model of acute pancreatitis. *Gut*, 1994;35:1306-10
- Sahin M, Yol S, Ciftci E, Baykan M, Ozer S, Aköz M, Yilmaz O, Kuru C. Does large bowel enema reduce septic complications in acute pancreatitis. *Am J Surg*, 1998;176:331-334
- Liu Q, Djuricin G, Nathan C, Gattuso P, Weinstein RA, Prinz RA. The effect of epidermal growth factor on the septic complications of acute pancreatitis. *J Surg Res*, 1997;69:171-177
- Foitzik T, Kruschewski M, Kroesen AJ, Hotz HG, Eibl G, Buhr HJ. Does glutamine reduce bacterial translocation? A study in two animal models with impaired gut barrier. *Int J Colorectal Dis*, 1999;14:143-149
- Marotta F, Geng TC, Wu CC, Barbi G. Bacterial translocation in the course of acute pancreatitis: beneficial role of non absorbable antibiotics and lactitol enemas. *Digestion*, 1996;57:446-452
- Wu XN. Management of severe acute pancreatitis. *World J Gastroenterol*, 1998;4:90-94
- Qamruddin AO, Chadwick PR. Preventing pancreatic infection in acute pancreatitis. *J Hosp Infect*, 2000;44:245-253
- Bhatia M, Brady M, Shokuh S, Christmas S, Neoptolemos JP, Slavin J. Inflammatory mediators in acute pancreatitis. *J Pathol*, 2000;190:117-125
- Kusnierz B, Naskalski JW. Mediators of the inflammatory response in the course of acute pancreatitis. *Przegl Lek*, 1999;56:532-536
- Norman J. The role of cytokines in the pathogenesis of acute pancreatitis. *Am J Surg*, 1998;175:76-83
- Zhao LG, Chen Q. Decisive factor and severity assessment in severe acute pancreatitis. *Xin Xiaohuabingsue Zazhi*, 1997;5:597-592
- Qin RY, Zou SQ, Wu ZD, Qiu FZ. Experimental research on production and uptake sites of TNF α in rats with acute hemorrhagic necrotic pancreatitis. *World J Gastroenterol*, 1998;4:144-146
- Hou YA, Wang Y, Xue JG. Hemorheological influences of abdominal irrigation of Chinese herbs in treatment of acute hemorrhagic necrotic pancreatitis in rabbits. *Xin Xiaohuabingxue Zazhi*, 1997;5:297-278
- Farkas G, Nagy Z, Marton J, Mandi Y. Relevance of cytokine production to infected pancreatic necrosis. *Acta Chir Hung*, 1997;36:86-88
- Lu XD, Liu GD, Chen YR. Experimental and clinical research of tumor necrosis factor alpha on acute pancreatitis. *Xin Xiaohuabingxue Zazhi*, 1997;5:534-534
- Andersson R, Wang X, Sun Z, Deng X, Soltesz V, Ihse I. Effect of a platelet activating factor antagonist on pancreatitis associated gut barrier dysfunction in rats. *Pancreas*, 1998;17:107-119
- Zhao LG, Wu XX, Han EK, Chen YL, Chen C, Xu DO. Protective effect of YHI and HHI-I against experimental acute pancreatitis in rabbits. *World J Gastroenterol*, 1998;4:256-259
- Wu, CT, Li ZL, Xiong DX. Relationship between enteric microecologic dysbiosis and bacterial translocation in acute necrotizing pancreatitis. *World J Gastroenterol*, 1998;4:242-245
- Sunamura M, Yamauchi J, Shibuya K, Chen HM, Ding L, Takeda K, Kobari M, Matsuno S. Pancreatic microcirculation in acute pancreatitis. *J Hepatobiliary Pancreat Surg*, 1998;5:62-68
- Hotz HG, Foitzik T, Rohweder J, Schulzke JD, Fromm M, Runkel NS, Buhr HJ. Intestinal microcirculation and gut permeability in acute pancreatitis: early changes and therapeutic implications. *J Gastrointest Surg*, 1998;2:518-525
- Sunamura M, Shibuya K, Yamauchi J, Matsuno S. Microcirculatory derangement and ischemia of the pancreas. *Nippon Geka Gakkai Zasshi*, 1999;100:342-346
- von Dobschuetz E, Hoffmann T, Messmer K. Inhibition of neutrophil proteinases by recombinant serpin Lex032 reduces capillary no reflow in ischemia/reperfusion induced acute pancreatitis. *J Pharmacol Exp Ther*, 1999;290:782-788

- 32 Flickinger BD, Olson MS. Localization of the platelet activating factor receptor to rat pancreatic microvascular endothelial cells. *Am J Pathol*, 1999;154:1353-1358
- 33 Frossard JL, Saluja A, Bhagat L, Lee HS, Bhatia M, Hofbauer B, Steer ML. The role of intercellular adhesion molecule 1 and neutrophils in acute pancreatitis and pancreatitis associated lung injury. *Gastroenterology*, 1999;116:694-701
- 34 Plusczyk T, Bersal B, Westermann S, Menger M, Feifel G. ET1 induces pancreatitis like microvascular deterioration and acinar cell injury. *J Surg Res*, 1999;85:301-310
- 35 Foitzik T, Hotz HG, Eibl G, Hotz B, Kirchengast M, Buhr HJ. Therapy for microcirculatory disorders in severe acute pancreatitis: effectiveness of platelet activating factor receptor blockade vs endothelin receptor blockade. *J Gastrointest Surg*, 1999;3:244-51
- 36 Foitzik T, Eibl G, Buhr HJ. Therapy for microcirculatory disorders in severe acute pancreatitis: comparison of delayed therapy with ICAM-1 antibodies and a specific endothelin A receptor antagonist. *J Gastrointest Surg*, 2000;4:240-247
- 37 Foitzik T, Hotz HG, Kinzig M, Sorgel F, Buhr HJ. Influence of changes in pancreatic tissue morphology and capillary blood flow on antibiotic tissue concentrations in the pancreas during the progression of acute pancreatitis. *Gut*, 1997;40:526-530
- 38 Wang WX, Zhao HP, Shou NY, Yang CW. Role of oxygen free radical and other inflammatory mediators in acute necrotic pancreatitis. *World J Gastroentero*, 1998;4:59-59
- 39 Lokhandwala MF, Barrett RJ. Cardiovascular dopamine receptors: physiological, pharmacological and therapeutic implications. *J Auton Pharmacol*, 1982;2:189-215
- 40 Karanjia ND, Lutrin FJ, Chang YB, Reber HA. Low dose dopamine protects against hemorrhagic pancreatitis in cats. *J Surg Res*, 1990;48:440-443
- 41 Harvey MH, Wedgwood KR, Reber HA. Vasoactive drugs, microvascular permeability, and hemorrhagic pancreatitis in cats. *Gastroenterology*, 1987;93:1296-1300
- 42 Karanjia ND, Lutrin FJ, Chang YB, Duong T, Reber HA. The anti-inflammatory effect of dopamine in alcoholic hemorrhagic pancreatitis in cats. Studies on the receptors and mechanisms of action. *Gastroenterology*, 1991;101:1635-1641

Edited by Zhou XH
proofread by Mittra S