

• BASIC RESEARCH •

Preventive effect of glutamine on intestinal barrier dysfunction induced by severe trauma

Jun-You Li, Yi Lu, Sen Hu, Dan Sun, Yong-Ming Yao

Jun-You Li, Yi Lu, Sen Hu, Dan Sun, Yong-Ming Yao, Burn Institute, Chinese PLA 304 Hospital, Beijing 100037, China
Supported by the Key Project of the "Tenth Five-Year Plan" of the Chinese PLA (01L081)

Correspondence to: Jun-You LI, Burn Institute, Chinese PLA 304 Hospital, 51 Fu Cheng Road, Beijing 100037, China. WuZG@A-1.net.cn
Telephon: +86-10-66867395 Fax: +86-10-68429998

Received 2001-06-03 Accepted 2001-11-15

Abstract

AIM: To investigate the mechanism underlying intestinal barrier function damage after severe trauma and the therapeutic effect of glutamine.

METHODS: Burned patients, and animal models of severe trauma replicated by hemorrhagic shock combined with endotoxin infusion and burn injury, were included in a serial experiment. Effects of oral glutamine on intestinal barrier function were observed in scalded rats. Parameters measured in these experiments were as follows: plasma levels of diamine oxidase (DAO), tumor necrosis factor (TNF α), endotoxin (LPS), and lactate as well as D-lactate by biochemical methods, lactose/mannitol (L/M) ratio in urine by SP-3400, and pathological examination of intestinal mucosa under light microscopy.

RESULTS: Plasma DAO activity was significantly increased after injury. There was a negative correlation between plasma DAO and intestinal mucosal DAO or pH_i ($r=-0.93$, plasma 0.80 ± 0.93 , 2.83 ± 1.71 , 1.14 ± 0.64 , 2.36 ± 2.06 and 2.49 ± 1.67 vs intestinal 0.52 ± 0.12 , 0.34 ± 0.03 , 0.45 ± 0.18 , 0.37 ± 0.26 and 0.41 ± 0.07 ; $r=-0.533$, plasma 0.87 ± 0.75 , 1.89 ± 1.13 , 1.21 ± 0.23 , 3.03 ± 2.61 and 4.70 ± 1.22 vs pH_i 7.03 ± 0.05 , 7.05 ± 0.06 , 7.14 ± 0.096 , 7.20 ± 0.08 and 7.05 ± 0.07 ; $P<0.01-0.05$). Positive correlations were found between DAO activity and plasma TNF α , LPS, lactate, L/M and D-lactate ($r=0.817$, 0.842 , 0.872 , and 0.951 ; plasma DAO 0.87 ± 0.75 , 1.89 ± 1.13 , 1.21 ± 0.23 , 3.03 ± 2.61 and 4.70 ± 1.22 vs TNF α 0.08 ± 0.02 , 0.03 ± 0.25 , 0.17 ± 0.09 , 0.34 ± 0.15 and 0.33 ± 0.18 ; vs LPS 0.14 ± 0.03 , 0.16 ± 0.04 , 0.21 ± 0.02 , 0.18 ± 0.16 and 0.37 ± 0.10 ; vs lactate 9.03 ± 2.19 , 18.30 ± 2.56 , 9.81 ± 2.83 , 12.01 ± 6.83 , 12.01 ± 6.84 and 43.61 ± 11.27 ; vs L/M 0.03 ± 0.01 , 0.41 ± 0.27 , 0.62 ± 0.20 , 1.70 ± 0.60 ; $r=0.774$, plasma DAO 1.25 ± 0.41 , 2.17 ± 0.71 , 2.29 ± 0.87 , 1.23 ± 0.55 and 1.11 ± 0.47 vs D-lactate 8.37 ± 2.48 , 18.25 ± 6.18 , 13.96 ± 4.94 , 8.93 ± 3.00 and 12.39 ± 4.94 ; all $P<0.01$), respectively. Damage of intestinal mucosa was found by pathological examination. Intestinal barrier function was improved to a certain extent by oral glutamine in scalded rats.

CONCLUSION: Intestinal barrier function was damaged in the early stage after trauma. Plasma DAO activity, D-lactate content, intestinal pH_i and urine L/M may be sensitive markers of intestinal mechanical injury, and glutamine may protect against intestinal barrier dysfunction after severe trauma.

Li JY, Lu Y, Hu S, Sun D, Yao YM. Preventive effect of glutamine on intestinal barrier dysfunction induced by severe trauma. *World J Gastroenterol* 2002;8(1):168-171

INTRODUCTION

It is generally accepted that the intestine may serve as an important organ in the development of severe complications under critically ill conditions, including trauma, burns, shock, etc.^[1-3] Hemorrhagic shock and/or gut ischemia-reperfusion injury commonly occur in the early stage after acute insults, leading to gut-derived sepsis as a result of gut barrier dysfunction^[4-10]. In order to investigate the mechanism underlying intestinal barrier function damage and its potential interventional measures, burned patients and animal models of severe trauma were employed in our current experiments^[6,11-15].

MATERIALS AND METHODS

Animal models

Animal models of severe trauma were replicated by hemorrhagic shock combined with endotoxin infusion. Male Wistar rats, weighing 190g-230g, were anaesthetized with intraperitoneal injection of 30g·L⁻¹ barbitone sodium (35mg·kg⁻¹), and the femoral artery and jugular vein were cannulated under aseptic conditions. The rats were then bled via the jugular vein catheter until a mean arterial pressure of 30-35 mmHg (4.6 kPa) was reached. At the end of shock, endotoxin (*E.coli*O55 B5, Sigma) was infused through tail vein at a dose of 2mg·kg⁻¹. A goat model of hemorrhagic shock combined with endotoxin challenge was established according to the previous report ($n=20$)^[5]. Animals received *E.coli*O26 B6 endotoxin via portal vein 24h after the recovery from shock, and the dosage was 30 ng·kg⁻¹·min⁻¹, which was given in a continuous infusion lasting for 5d. Wistar rats were divided randomly into three groups: normal controls, early feeding with standard feed phase Gln 0.5g after scalding, and animals (except control group) sustained a 30% TBSA full-thickness scald covering the back and flanks^[6]. Determination of plasma diamine oxidase in 21 burned patients (17 male and 4 female) at the age of 33±10 years, with burn area (64±21)%, and (35±20)% III/0. Plasma DAO activity was determined on day 1, 3, 7, 14 and 21 postburn. Blood and intestinal DAO levels were tested according to our previous report^[17]. Plasma lactate and D-lactate concentrations were determined by biochemical methods as described by Brandt *et al*^[18]. Microassay for quantitation of endotoxin in blood was made with new PCA treatment using chromogenic limulus amoebocyte lysate^[19].

Tumor necrosis factor (TNF α) assay. Plasma TNF content was measured by radioimmunoassay. Lactulose/mannitol (L/M) ratio tests in urine were made by SP-3400^[20].

Pathological examination. Tissue samples were examined under light microscopy.

Statistical Analyses

Data were expressed as the mean±standard error, and were statistically evaluated by Students *t* test and correlation analysis. Differences were considered to be significant with $P<0.05$.

RESULTS

Plasma DAO levels were elevated in double-peak patterns, one at early stage after trauma and another during invading infection in animal models. Similar results were also obtained in burned patients. Meanwhile, intestinal DAO levels were decreased to certain extent after trauma in animal model. There was a significantly negative correlation between plasma and intestinal DAO activity (Table 1; Figures 1 and 2).

Table 1 Changes in plasma DAO in trauma animal model ($\bar{x} \pm s \times 10^3 \text{U} \cdot \text{L}^{-1}$)

Animal model	Before injury	T (after injury)/h				
		2	6	24	48	72
Goat	0.9±0.8	1.9±1.1 ^a		1.2±0.2	3.0±2.6	4.7±1.2 ^b
Rats	1.3±0.4	2.2±0.7 ^b	2.3±0.87 ^b	1.2±0.6	1.1±0.5	
Scalded rats	0.8±0.9	0.8±1.8 ^b		1.1±0.6	2.4±2.1 ^a	
Burned pigs	4.1±1.4	4.7±1.5		4.7±1.4	4.8±1.1	5.8±1.4 ^a
Gun shooting	1.5±0.6	1.2±0.5	2.0±0.6 ^b	1.9±0.2 ^b		

Dog in hypothermia

^a $P < 0.05$; ^b $P < 0.01$, vs before injury.

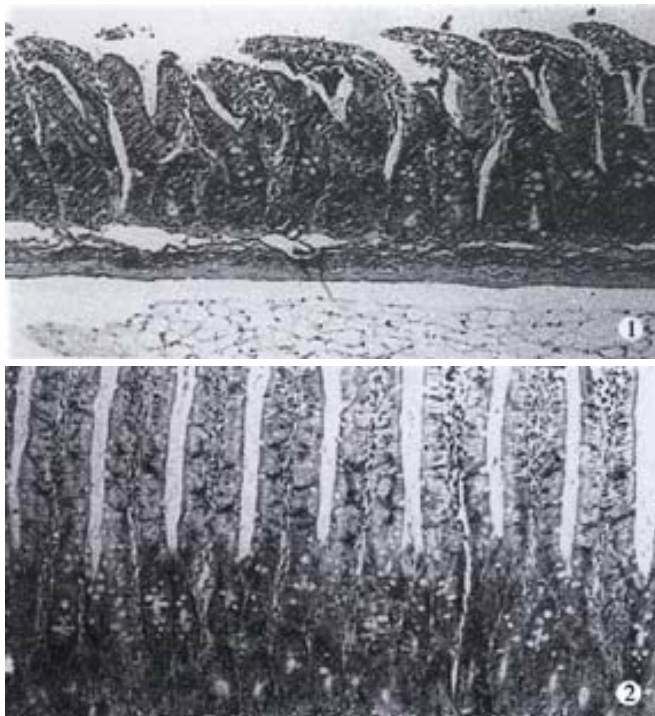


Figure 1 Pathological changes of intestinal mucosa after 8d in scalded rats. HE×100

Figure 2 Pathological changes of intestinal mucosa after 8d oral GLN in scalded rats. HE×100

Plasma DAO concentrations and the level of the related index, and plasma $\text{TNF}\alpha$ significantly increased at various intervals after trauma in goats. Plasma endotoxin levels notably increased at 24h and 72h after injury. Blood lactic acid significantly increased from 2h to 72h after trauma. The plasma DAO activity was obviously correlated with plasma $\text{TNF}\alpha$, LPS and lactate ($P < 0.01$; Table 2 and 3). Changes in DAO activity were significantly related with plasma $\text{TNF}\alpha$ and LPS levels in scalded rats. Plasma DAO activity and plasma D-lactate were significantly correlated in rats secondary to hemorrhage followed by endotoxin challenge ($r = 0.774$; $P < 0.01$).

Glutamine could protect against intestinal barrier function damage. The results indicated that plasma DAO activity was decreased in animals with early glutamine supplementation compared with those without glutamine treatment (10h after trauma $P < 0.05$).

Results of intestinal pathological examination. The pathologic examination of the intestine showed that the damage of epithelial cells of intestinal mucosa, hemorrhage and necrosis, accompanied by the inflammatory cell infiltration in intestinal wall in goats suffering from hemorrhagic shock combined with endotoxin infusion. It was revealed that there was disruption of intestinal mucosa after scald and gut ischemia-reperfusion combined with endotoxin challenge in rats, whereas oral glutamine supplementation could markedly improve intestinal mucosa following acute insults (Figure 2).

Table 2 Changes in parameters in goats after hemorrhagic shock combined with endotoxin infusion

	Before injury	T (after injury)/h			
		2	24	48	72
$\text{TNF}/\mu\text{g} \cdot \text{L}^{-1}$	0.08±0.02	0.03±0.25 ^a	0.17±0.09 ^a	0.34±0.15 ^a	0.33±0.18 ^a
$\text{LPS}/\times 10^3 \text{Eu} \cdot \text{L}^{-1}$	0.14±0.03	0.16±0.04	0.21±0.02 ^a	0.18±0.16	0.37±0.10 ^b
Lactate/ $\text{nmol} \cdot \text{L}^{-1}$	9.03±2.19	18.30±2.56 ^a	9.81±2.83	12.01±6.84	43.61±11.27 ^b
L/M rat	0.03±0.01	0.41±0.27	0.62±0.20		1.70±0.60 ^b
Intestinal pH	7.03±0.05	7.05±0.06 ^b	7.14±0.09 ^b	7.20±0.08 ^a	7.05±0.07 ^b

^a $P < 0.05$; ^b $P < 0.01$, vs before injury.

Table 3 Correlation analysis in goats subjected to hemorrhagic shock combined with endotoxin infusion

X	Y	r	P
DAO	$\text{TNF}\alpha$	0.817	<0.01
DAO	LPS	0.842	<0.01
DAO	Lactate	0.872	<0.01
DAO	L/m	0.951	<0.01
DAO	Intestinal pH	-0.553	<0.05

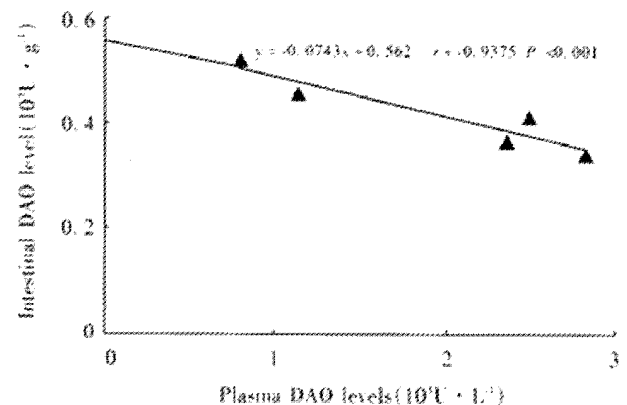


Figure 3 Relationship between intestinal and plasma DAO level.

DISCUSSION

DAO is located in the upper part of intestinal mucosa in human as well as in mammals, and is a highly active intracellular enzyme. Under certain circumstances, intestinal mucosa cells became necrosed and dropped into the intestinal cavity, leading to decrease in intestinal mucosal DAO, and increase in DAO activity inside the intestinal cavity. DAO can also enter into the mucosal space between cells, lymphatic vessel^[21-27] and blood flow, making plasma DAO markedly elevated. Gut as an important organ, may play an important role in the pathogenesis of serious complications. Intestinal mucosal surface layer with tight epithelial cells is an important component for intestinal barrier function, thus it can be seen that intestinal epithelial tissue integrated property is a key part to preserve intestinal barrier function. The changes in DAO activity is an ideal index to investigate intestinal barrier function damage after trauma, specially the changes in plasma DAO activity^[2,6,10,11]. Therefore, intestinal barrier function injury after severe trauma could result in bacterial/toxin translocation, in turn evoke systemic inflammatory response syndrome

and multiple organ dysfunction syndrome^[3,28-29].

The intestinal blood flow might keep relatively low despite of systemic circulation recovery during trauma, which was evident by significant decrease in intestinal mucosal pH_i^[30-33]. This may be the pathological basis for intestinal origin sepsis and multiple organ dysfunction syndrome. The changes in intestinal permeability were reflected by blood D-lactate and L/M rate in urine. Secretory IgA is an important component part for regulation intestinal immune function, it can prevent the bacteria from adhering to intestinal epithelial cells, and prevent gut-derived bacteria from invading through the intestinal barrier, which might reduce the toxicity of bacterial products to epithelial cells. Therefore, IgA may possess the beneficial effect on the preservation of intestinal mechanical barrier^[34-37]. From our data that changes in several indexes of intestinal barrier function in various animal models after trauma, it was shown that DAO was released to increase in blood, and decreased in intestinal tissues. Thus, determination of DAO activity might reflect the condition of intestinal injury and repaired process.

The change in plasma D-lactate, lactulose, mannitol and ratio of L/M could reflect the increased intestinal permeability. The intestinal IgA levels appear to be associated with the local immunological dysfunction^[38-41]. The change in intestinal pH_i showed that intestinal hemorrhagic injury may result in the release of intestinal mucosal enzyme, subsequently leading to significant elevation of plasma DAO activity. This study showed that there is a close relationship between plasma DAO and TNF α , LPS, D-lactate, lactate, L/M, and the change of intestinal pathology and intestinal barrier function index was similar, indicating that intestinal barrier function was damaged after trauma.

We also observed that the plasma DAO activity was increased 2h and 72h after trauma, especially at 72h, and change of plasma TNF α and LPS was similar to the former. These results suggest that stress injury can cause changes of intestine barrier function index following intestinal ischemia-reperfusion injury, repair or endo/inextra protection, but the trends and degrees of changes were varied. Therefore, it is important to reduce development of SIRS to MODS by protection of intestinal barrier function in early trauma, and prevention of translation of intestinal origin bacteria and toxins^[6,10,14,42-45].

Recent studies have shown that GLN may provide protection against intestinal barrier function injury after trauma^[46-52]. Blood DAO activity was reduced to different extents at 10h, 5d and 8d after early stage of oral GLN in scalded rats. Intestinal pathological examination showed that the damage of epithelial cells of intestinal mucosa could be markedly improved, and the close correlation between plasma DAO and LPS and TNF α could also be changed after scalding^[6]. The results indicate that GLN can protect against the intestinal barrier function damage after trauma.

REFERENCES

- Li JY, Hu S, Sun XQ, Yan M, Jin H, Jiang XG, Zhou BT, Sheng ZY. Study on pathophysiological changes during early stage of sepsis after trauma. *Zhong guo wei zhong bing ji jiu yi xue* 2001;13:291-294
- Li JY, Sun SR, Xue LB, Lu Y, Jing H, Gu ZR. Sequential changes in diamine oxidase activity after burns. *Zhong hua zheng xing shao shang za zhi* 1997;13:40-42
- Swank GM, Deitch EA. Role of the gut in multiple organ failure: bacterial translocation and permeability changes. *World J Surg* 1996;20:411-417
- Chen HL, Wu JZ, Pei DK, Guan FL. Damage of gut barrier in the pathogenesis of multiple organ dysfunction syndrome. *Zhong hua pu tong wai ke za zhi* 1998;13:50-53
- Hu S, Sheng ZY, Xue XB, Zhou BT, Lu JY, Jin H, Yu Y, Sun XQ, Li JY, Zheng YQ, Xiong DX. A serilized studies of animal models of posttraumatic multiple organ dysfunction syndrome. *Jie fang jun yi xue za zhi* 1996;21:5-9
- Li JY, Lu Y, Xue LB, Wan SY, Sun SR, Yu Y, Xu HJ, Sheng ZY. The effect of oral glutamine gut mucosa Function and structure in scalded rat. *Jie fang jun yi xue za zhi* 1996;21:91-93
- Yao YM, Sheng ZY, Wu Y, Yu Y, Lu LR. Relationship between plasma D-lactate levels and acute intestinal injury in rats following ischemia-reperfusion. *Zhong hua zheng xing shao shang za zhi* 1998;14:266-269
- Lu Y, Sheng ZY, Li JY, Sun XQ, Jin H, Jiang XG. The relationship between the ICAM-1 expression of hemangioendothelial cell and the dysfunction of murine small bowel in an intestinal ischemia-reperfusion model. *Zhong guo pu tong wai ke za zhi* 2000;15:145-147
- Cui XL, Sheng ZY, Guo ZR, He LX, Zao J, Ren XW, Sun SR, Jiang LX. Mechanisms of early gastro-intestinal ischemia after burn: hemodynamic and hemorrhologic features. *Zhong hua zheng xing shao shang za zhi* 1998;14:262-265
- Li JY, Lu Y, Fu XB, Jin H, Hu S, Sun XQ, Sheng ZY. The significance of changes in diamine oxidase activity in intestinal injury after trauma. *Zhong guo wei zhong bing ji jiu yi xue* 2000;12:482-484
- Li JY, Lu Y, Hu S, Jin H. An experimental study on monitoring of barrier function of small intestine. *Chuang shang wai ke za zhi* 2001;3:109-112
- Li JY, Lu Y, Jin H, Hu S, Sun XQ, Jiang XG, Sheng ZY. Sequential changes of diamine oxidase activity after trauma. *An ji suan he sheng wu zi yuan* 1999;21:17-19
- Fang WH, Yao YM, Shu HG, Yu Y, Wu Y, Zheng YQ. The time course and tissue distribution of endotoxin in rats after thermal injury. *Zhong hua zheng xing shao shang za zhi* 1999;15:298-300
- Fu XB, Yang YH, Sun XQ, Gu XM, Sheng ZY. Protective effects of endogenous basic fibroblast growth factor activated by 2.3 butanedion monoxide on functional changes of ischemic intestine, liver and kidney in rats. *Zhong guo wei zhong bing ji jiu yi xue* 2000;12:69-72
- Fu XB, Sheng ZY, Wang YP, Ye YX, Xu MH, Sun TZ, Zhou BT. Basic fibroblast growth factor reduces the gut and liver morphologic and functional injuries after ischemia and reperfusion. *J Trauma* 1997;42:1080-1087
- Wu CT, Li ZL. Effect of DAO on intestinal damage in acute necrotizing pancreatitis in dogs. *Shi jie hua ren xiao hua za zhi* 1999;7:64-65
- Li JY, Yu Y, Hao J, Jin H, Xu HJ. Determination of diamine oxidase activity in intestinal tissue and blood using spectrophotometry. *An ji suan he sheng wu zi yuan* 1996;18:28-30
- Sun XQ, Fu XB, Zhang R, Lu Y, Deng Q, Zhang C, Jiang XG, Sheng ZY. Plasma D-lactate levels as a useful marker of increased intestinal permeability after severe injuries. *Zhong guo wei zhong bing ji jiu yi xue* 2000;12:476-478
- Yao YM, Tian HM, Wang YP, Yu Y, Shi ZG, Sheng ZY. Microassay for quantitation of endotoxin in blood with new PCA treatment using chromogenic limulus amoebocyte lysate. *Shang hai yi xue jian yan za zhi* 1993;8:31-33
- Zhu XF, Xiong DX, Sheng ZY. Establish measuring Method of alterations in intestinal permeability. *Zhong guo wei sheng tai za zhi* 1994;6:36-38
- Hosoda N, Nishi M, Nakagawa M, Hiramatsu Y, Hioki K, Yamamoto D M. Structural and functional alterations in the gut of parenterally or enterally fed rats. *J Surg Res* 1989;47:129-133
- Yan BG, Yang ZC, Huang YS, Liu ZY, Li A. Effect of delayed rapid fluid resuscitation on plasma diamine oxidase in early postburn dogs. *Shi jie hua ren xiao hua za zhi* 2000;8:178-180
- Lu Y, Li JY, Sun SR, Jin H, Jiang XG, Sun XQ, Sheng ZY. Relationship between change of plasma diamine oxidase activity and gut injury in rats during gut ischemia-reperfusion. *An ji suan he sheng wu zi yuan* 2000;22:50-53
- Liu MH, Gao LX. The effect of oral glutamine supplementation on small intestinal function in burned rats. *Ying yang xue bao* 1995;17:17-21
- Dong HL, Cai JW, Si ZX. Effects of radiation on intestinal and plasma DAO activity in rats. *Shi jie xiao hua za zhi* 1998;6:698
- Bragg LE, Thompson JS, West WW. Intestinal diamine oxidase levels reflect ischemic injury. *J Surg Res* 1991;50:228-233
- Ci XL, Wang BS, Zhang SW, Zhang NN. Experimental treatment with herbal medicines for impaired intestinal mucosal barrier induced by endotoxin plus tumor necrosis factor-alpha. *Zhong guo wei zhong bing ji jiu yi xue* 1999;11:262-265
- Rush BF, Sori AJ, Murphy TF, Smith S, Flanagan JJ, Machiedo GW. Endotoxemia and bacteremia during hemorrhagic shock: The link between and sepsis? *Ann Surg* 1988;207:549-552
- Yao YM, Sheng ZY, Tian HM, Wang YP, Yu Y, Fu XB, Lu LY, Wang DW, Ma YY. Gut-derived endotoxemia and multiple system organ failure following gunshot wounds combined with hemorrhagic shock an experimental study in the dog. *J Trauma* 1995;38:742-746
- Hu S, Jin H, Lu Y, Li JY, Zhou BT. The changes of PH on gastroduodenal mucosa after hemorrhage shock and resuscitation in goats. *Zhong guo wei zhong bing ji jiu yi xue* 1997;9:708-710
- Murray MJ, Gonze MD, Nowak LR, Cobb CF. Serum D(-)-Lactate levels as an aid to diagnosing acute intestinal ischemia. *The American Journal of Surgery* 1994;167:575-578
- Sun XQ, Lu Y, Jin H, Li JY, Jiang XG, Hu S, Fu XB. Change of intestine mucosal barrier induced by intestine ischemia-reperfusion injury.

- Chuang shang wai ke za zhi* 1999;1:208-210
- 33 Murray MJ, Barbose JJ, Cobb CF. Serum D-lactate levels, as a predictor of acute intestinal ischemia in a rat model. *J Surg Res* 1993;54:507-509
 - 34 Doe WF. The intestinal immune system. *Gut* 1989;30:1679-1685
 - 35 Yu Y, Shi ZG, Chai JK, Guo ZY, Yao YM, Li G, Ho XX, Jiang W, Wang HB, Sheng ZY. The clinical significance of changes in IgA levels in intestinal tract after major burns. *Zhong guo wei zhong bing ji jiu yi xue* 1998;10:725-727
 - 36 Yu Y, Sheng ZY, Tian HM, Wang YP, Yu Y, Lu LR, Chang GY, Ma RS. An experimental study on injury of intestinal immuno-barrier in rat after scald. *Zhong hua zheng xing shao shang wai ke za zhi* 1996;12:86-89
 - 37 Biesbrock AR, Reddy MS, Levine MJ. Interaction of a salivary mucin-secretory immunoglobulin a complex with mucosal pathogens. *Infection and Immunity* 1991;59:3492-3497
 - 38 Harmatz PR, Cater EA, Sullivan D, Hatz RA, Baker R, Breazlale E, Grant K, Bloch KJ. Effect of thermal injury in the rat on transfer of IgA protein into bile. *Ann Surg* 1989;210:203-207
 - 39 Cappeler WA, Bloch KJ, Hatz RA, Carter EA, Fagundes J, Sullivan DA, Harmatz PR. Reduction in biliary IgA after burn injury. *Ann Surg* 1992;215:338-341
 - 40 Alverdy JC, Aoys E. The Effect of dexamethasone and endotoxin administration on biliary IgA and bacterial adherence. *J Surg Res* 1992;53:450-454
 - 41 Yu Y, Chai JK, Li JY, Jin H, Dong N. A preliminary study on the relationship between level of IgA in intestinal content and urine after burns. *An ji suan he sheng wu zi yuan* 2001;23:49-51
 - 42 Diebel LN, Liberati DM, Brown WJ, Dulchavsky SA, painter TM, Diglio CA, Montgomery PC. Secretory immunoglobulin A blocks hypoxia-augmented bacterial passage across madin-darby canine kidney cell monolayers. *J Trauma* 1997;43:759-763
 - 43 Noguchi Y, James JH, Fischer JE, Hasselgren PO. Increased glutamine consumption in small intestine epithelial cell during sepsis in rats. *Am J Surg* 1997;173:199-205
 - 44 Yu B, Wang SL, You ZY, Li A, Zhao Y. Effects of L-Glutamine on ghe tut nutrient metabolism in severe burnedminiswines. *Chang nei yu chang wai ying yang* 1995;2:7-10
 - 45 Li N, Liu FN, Li YS, Kang J, Li FJ, Li JS. The Changes of plasma glutamine and its influence on intestinal permeability after abdominal Surgery. *Chang nei chang wai ying yang* 1998;5:3-6
 - 46 Rhoads JM, Keku EO, Quinn J, Woosely J, Lecce JG. L-Glutamine stimulates jejunal sodium and chloride absorption in pig rotavirus enteritis. *Gastroenterology* 1991;100:683-691
 - 47 Yu B, Wang SL, You ZY, Zhao Y, Li A. Enhancement of gut absorptive function by early enteral feeding enriched with L-Glutamine in severe burned miniswines. *Zhong hua wai ke za zhi* 1995;33:742-744
 - 48 Wu F, Jin XQ, Wu SX, Li J, Xu JL. The experimental study of cholestasis due to parenteral nutrition. *Chang nei chang wai ying yang* 1998;5:96-98
 - 49 Yang JT, Wang ZG, Zhu PF. The Effects of glutamine supplementation on the cellular immunity in traumatized rats. *Ying yang xue bao* 1999;21:8-12
 - 50 Dai DW, Wu SM, Li M, Chen HJ, Cai W. Protective effect of Glutamine in the anoxia/reoxygenation induced injury of human intestinal epithelial cells *in vitro*. *An ji suan he sheng wu zi yuan* 1997;19:1-3
 - 51 Tu WF, Li JS, Zhu WM, Li ZD, Liu FN, Chen YM, XU JG, Shao HF, Xiao GX, LI A. Infiluence of glutamine on gut bacteria/endotoxin translocation in acute severe pancreatites. *Chang wai yu chang nei ying yang* 1999;6:29-32
 - 52 Yu B, Wang SL, You ZY, Zhao Y, Li A. Protection against intestinal injury with oral feeding L-glutamine in severe burned mini-swines. *Zhong hua wai ke za zhi* 1998;14:19-21

Edited by Ma JY