

• BASIC RESEARCH •

Protective effect of estradiol on hepatocytic oxidative damage

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Abstract

AIM: To examine the protective effect of estradiol on the cultured hepatocytes under oxidative stress.

METHODS: Hepatocytes of rat were isolated by using perfusion method, and oxidative stress was induced by a serum-free medium and FeNTA. MDA level was determined with TBA method. Cell damage was assessed by LDH assay. Apoptosis of hepatocytes was assessed with cytoflowmetric analysis. Expression of Bcl-xl in cultured hepatocytes was detected by Western blot. The radical-scavenging activity of estradiol was valued by its ability to scavenge the stable free radical of DDPH.

RESULTS: Oxidative stress increased LDH (from $168 \pm 25 \times 10^{-6} \text{IU} \cdot \text{cell}^{-1}$ to $780 \pm 62 \times 10^{-6} \text{IU} \cdot \text{cell}^{-1}$) and MDA (from $0.28 \pm 0.07 \times 10^{-6} \text{nmol} \cdot \text{cell}^{-1}$ to $1.35 \pm 0.12 \times 10^{-6} \text{nmol} \cdot \text{cell}^{-1}$) levels in cultured hepatocyte, and estradiol inhibited both LDH and MDA production in a dose dependent manner. In the presence of estradiol $10^{-6} \text{mol} \cdot \text{L}^{-1}$, $10^{-7} \text{mol} \cdot \text{L}^{-1}$ and $10^{-8} \text{mol} \cdot \text{L}^{-1}$, the LDH levels are $410 \pm 53 \times 10^{-6} \text{IU} \cdot \text{cell}^{-1}$ ($P < 0.01$ vs oxidative group), $530 \pm 37 \times 10^{-6} \text{IU} \cdot \text{cell}^{-1}$ ($P < 0.01$ vs oxidative group), $687 \pm 42 \times 10^{-6} \text{IU} \cdot \text{cell}^{-1}$ ($P < 0.05$ vs oxidative group) respectively, and the MDA level are $0.71 \pm 0.12 \times 10^{-6} \text{nmol} \cdot \text{cell}^{-1}$ ($P < 0.01$ vs oxidative group), $0.97 \pm 0.11 \times 10^{-6} \text{nmol} \cdot \text{cell}^{-1}$ ($P < 0.01$ vs oxidative group) and $1.27 \pm 0.19 \times 10^{-6} \text{nmol} \cdot \text{cell}^{-1}$ respectively. Estradiol suppressed apoptosis of hepatocytes induced by oxidative stress, administration of estradiol ($10^{-6} \text{mol} \cdot \text{L}^{-1}$) decreased the apoptotic rate of hepatocytes under oxidative stress from $18.6 \pm 1.2\%$ to $6.5 \pm 2.5\%$, $P < 0.01$. Bcl-xl expression was related to the degree of liver cell damage due to oxidative stress, and estradiol showed a protective action.

CONCLUSION: Estradiol protects hepatocytes from oxidative damage by means of its antioxidant activity.

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INTRODUCTION

Hepatic fibrosis is a common consequence of chronic liver injury from many causes^[1-7], and the critical event in hepatic fibrosis is the

activation of lipocyte (also known as the stellate, fat-storing or Ito cell) which is the main source of extracellular matrix in fibrosis formation^[8-20]. The putative impetus to lipocyte activation came from cytokines released from Kupffer cells or leukocytes in liver injury^[21]. An alternative way is that parenchymal-cell necrosis itself may activate lipocyte; such activation could be mediated by the lipid peroxides formed after the membrane of parenchymal cells is injured^[22-28]. Therefore, preventing hepatocyte from injury is a matter of primary importance in blocking the fibrogenic pathway. We have reported the inhibitory effect of estradiol on activation of rat lipocytes^[29,30] and the suppressive effect on fibrogenesis in rat model^[31]. The present study is initiated to investigate the role of estradiol on the hepatocyte under oxidative stress, and to elucidate the mechanism of its inhibitory effect on hepatic fibrogenesis.

MATERIALS AND METHODS

Hepatocyte isolation and induction of lipid peroxidation

Hepatocytes were isolated from the liver of male Wistar rats (500-600g) with *in situ* perfusion method as previously described. Inocula of 2×10^5 cells per well were introduced into 12-well plate, (Nunc). The cells were cultured in 1ml Williams medium E supplemented with $50 \text{ml} \cdot \text{L}^{-1}$ FBS, $105 \text{U} \cdot \text{L}^{-1}$ penicillin, $100 \text{mg} \cdot \text{L}^{-1}$ streptomycin, and $10 \text{g} \cdot \text{L}^{-1}$ L-glutamine at 37°C in $50 \text{ml} \cdot \text{L}^{-1}$ CO_2 atmosphere and 100% humidity. After 4h, the cell medium was removed and lipid peroxidation was induced by incubating hepatocytes in serum-free Williams medium E with $100 \mu\text{mol} \cdot \text{L}^{-1}$ FeNTA (ferric nitrilotriacetate). Three groups of hepatocyte were analysed in parallel in each experiment including the hepatocytes cultured under normal condition and the hepatocyte cultured under oxidative stress in the presence or absence of $17\text{-}\beta$ -estradiol (Sigma).

Detection of (MDA) and lactate dehydrogenase (LDH) level in culture medium

Lipid peroxidation in cultured hepatocyte was determined by detecting the level of malondialdehyde (MDA), end product of lipid peroxidation in culture medium. The cell medium was collected, centrifuged at 450g to remove cell debris, and the MDA contents was determined by using a colorimetric reaction with thiobarbituric acid. A calibration curve was constructed from the conversion of tetraethoxypropane to MDA. The degree of cell damage was assessed by detecting the lactate dehydrogenase (LDH) activity released from the cytosol of damaged cells into the supernatant. The LDH activity was determined by using LDH detection Kit (Sigma). The time course of the MDA and LDH levels in culture medium was constructed to examine the lipid peroxidation and cell damage of hepatocyte during cultivation.

Flowcytometric analysis for apoptosis in hepatocytes

Rat hepatocytes were isolated and inoculated in 24-well plate as described. After oxidative stress was induced for 24h, apoptotic hepatocytes were detected by flowcytometry. Apoptotic cells expose their phosphatidylserine (PS) in the outer leaflet of cell membrane.

The exposed PS can be revealed by FITC-conjugated annexin V. Annexin V Kits(BD pharmingen) was used ,and the samples were treated according to the instruction enclosed in the kit as mannul described and analysed on the flowcytometer(Coulter Epics).

Western blot analysis of Bcl-xl

Rat hepatocytes were cultured in 35mm diameter dishes(Nunc) which were divided into 3 groups: the normal group, the oxygen stress and the oxygen stress plus estradiol group. In the presence or absence of $10^{-6}\text{mol}\cdot\text{L}^{-1}$ estradiol for the indicated time period,the dishes were then washed twice with ice cold PBS and lysed directly in 1ml SDS loading buffer($50\text{mmol}\cdot\text{L}^{-1}$ Tris, pH6.7, $20\text{g}\cdot\text{L}^{-1}$ SDS, $100\text{g}\cdot\text{L}^{-1}$ glycerol, $0.6\text{g}\cdot\text{L}^{-1}$ bromophenol blue, $100\text{mmol}\cdot\text{L}^{-1}$ dithiothreitol). The samples were boiled for 5 min and applied to a standard $120\text{g}\cdot\text{L}^{-1}$ SDS polyacrylamide protein gel. After electrophoresis, protein transfer was performed onto Hybond-ECL(Amersham Pharmacia Biotech) using a semi-dry blotting apparatus. The membrane was treated first with $100\text{ml}\cdot\text{L}^{-1}$ non-fat milk in PBS at room temperature for 2h and next with the Bcl-xl or Bcl-2 monoclonal antibody (Tanslab; diluted 1:500)for two hours at room temperature. After washing, the membrane was then incubated with HRP conjugated goat antimouse IgG(Amersham Pharmacia Biotech; diluted 1:1000)for one hour at room temperature. Immunoreactive bands were visualized using the ECL western blotting detection system kit (Amersham Pharmacia Biotech) according to the manufacturer's recommended protocol. The membranes used for Bcl-xl or Bcl-2 detection were reprobed with actin polyanitbody and the corresponding secondary antibody to normalize the signal strength of Bcl-xl and Bcl-2.

Radical-scavenging activity of estradiol

The radical-scavenging activity of estradiol was determined from its ability to scavenge the stable free radical of 1,1-diphenyl-2-picrylhydrazyl (DPPH,Wako) and was compared with that of the well-known antioxidant, α -tocopherol. $5\mu\text{l}$ of estradiol or α -tocopherol ($2,4,6,8,10\mu\text{mol}\cdot\text{L}^{-1}$) was added to 2.5ml $100\mu\text{mol}\cdot\text{L}^{-1}$ DPPH, in $20\text{mmol}\cdot\text{L}^{-1}$ 2-(N-morpholino) ethanesulfonic acid (MES) (pH 5.5) and the change in optical absorbance at 517nm was measured 30min thereafter.

Statistical analysis

Experimental results were analyzed by Student's *t* test for multiple comparisons. *P* values less than 0.05 were considered to be statistically significant.

RESULTS

Effect of estradiol on MDA and LDH level in hepatocytes under oxidative stress

The MDA and LDH levels in the medium of hepatocytes cultured under normal condition and oxidative stress were shown respectively in Figure 1 and Figure 2. The MDA level increased rapidly 5h after the oxidative stress and was maintained at a considerably high concentration during the period of continued cultivation. In normal control, MDA was kept at a low concentration in the incubating time, indicating a low level of lipid peroxidation in the normal condition. LDH level increased steadily after oxidative stress, whereas the LDH level in normal control increased slightly during culture. These data revealed that oxidative stress initiate lipid peroxidation and cause cell membrane damage in hepatocytes under stress. Twenty-four' oxidative stress was selected as the time point to examine the effect of estradiol administration on the MDA and LDH level in hepatocytes under oxidative stress. As the data showed in Table 1,estradiol decreased the MDA and LDH level in the culture medium in a dose

dependent manner.That means estradiol could inhibit the lipid peroxidation and subsequent hepatocytic membrane damage under oxidative stress.

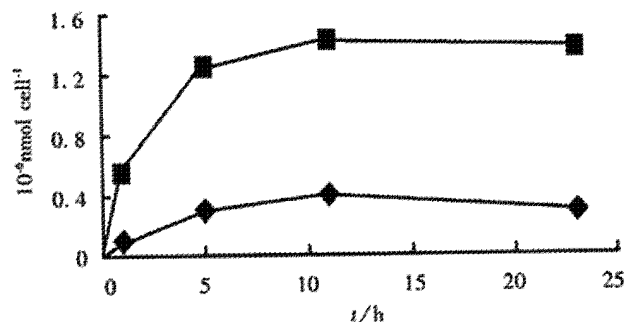


Figure 1 Effect of estradiol on MDA level in hepatocytes cultured under normal condition (◆) and oxidative stress (■)

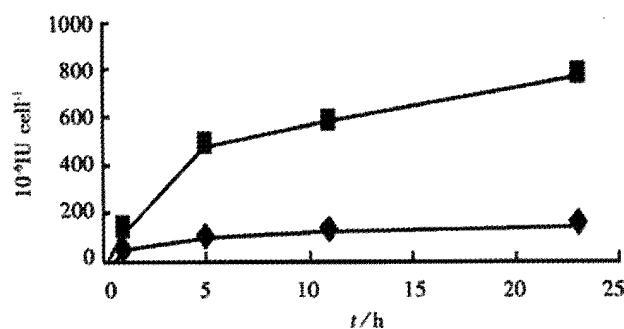


Figure 2 Effect of estradiol on LDH level in hepatocytes cultured under normal contion (◆) and oxidative stress (■)

Table 1 Effect of estradiol on MDA and LDH level in culture medium of hepatocytes under oxidative stress

Stress	MDA($10^{-6}\text{nmol}\cdot\text{cell}^{-1}$)	LDH ($10^3\text{IU}\cdot\text{cell}^{-1}$)
780±62		1.35±0.12
$10^{-8}\text{mol}\cdot\text{L}^{-1}$ estradiol	1.27±0.19	687±42 ^a
$10^{-7}\text{mol}\cdot\text{L}^{-1}$ estradiol	0.97±0.11 ^b	530±37 ^b
$10^{-6}\text{mol}\cdot\text{L}^{-1}$ estradiol	0.71±0.12 ^b	410±53 ^b
Normal control	0.28±0.07 ^b	168±25 ^b

^a*P*<0.05, ^b*P*<0.01 vs stress group.

Flowcyto-metric analysis for apoptotic hepatocytes

The aim of this investigation was to ascertain whether oxidative stress could induce apoptosis of hepatocytes, and whether estradiol could protect hepatocytes from such damage. The apoptotic rate of hepatocyte under normal culture condition was $5.9\pm1.7\%$, but in hepatocytes under oxidative stress it was $18.6\pm1.2\%$. With the administration of $10^{-6}\text{mol}\cdot\text{L}^{-1}$ estradiol,the apoptotic rate of hepatocytes under oxidative stress decreased to $6.5\pm2.5\%$. These data suggest that estradiol inhibit hepatocytic apoptosis induced by oxidative stress (results obtained from three distinct experiments).

Western blot analysis of Bcl-xl

Bcl-2 family has been investigated extensively for its proapoptotic or antiapoptotic property; Bcl-xl and Bcl-2 are well-known negative regulator of apoptosis^[32-36]. Western blot was used to examine the relationship between Bcl-xl and Bcl-2 level and estradiol administration. Estradiol applied in this experiment was $10^{-6}\text{mol}\cdot\text{L}^{-1}$. The result was depicted in Figure 3 (showing one representative result

from three independent experiments). Bcl-xl level was upregulated in hepatocytes under oxidative stress for 24h as compared with the normal control, while estradiol administration attenuated the increased expression of Bcl-xl induced by oxidative stress. After culturing for 48h, Bcl-xl expression was increased in hepatocytes both in the absence or presence of estradiol under oxidative stress. Bcl-xl level in hepatocytes was increased under oxidative stress. Bcl-2 expression was not detectable even though two antibodies from two different companies were used, indicating its low level in hepatocyte.

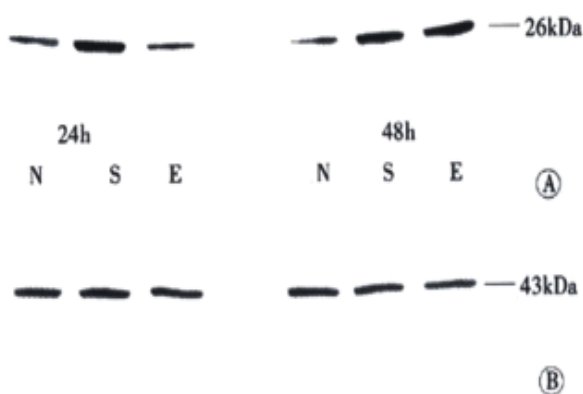


Figure 3 (A) Effect of estradiol on Bcl-xl expression in cultured hepatocytes for 24h and 48h. (B) Actin expression in cultured hepatocytes for 24h and 48h. N: Normal condition. S: Oxidative stress. E: Oxidative stress supplemented with 10^{-6} mol \cdot L $^{-1}$ estradiol.

Radical-scavenging activity of estradiol

Radical-scavenging activity of estradiol was determined by monitoring the decrease in absorbance at 517nm. Estradiol caused an immediate decrease in DPPH absorbance in a dose dependent manner (Figure 4). Its radical scavenging activity potency was approximately half that of α -tocopherol.

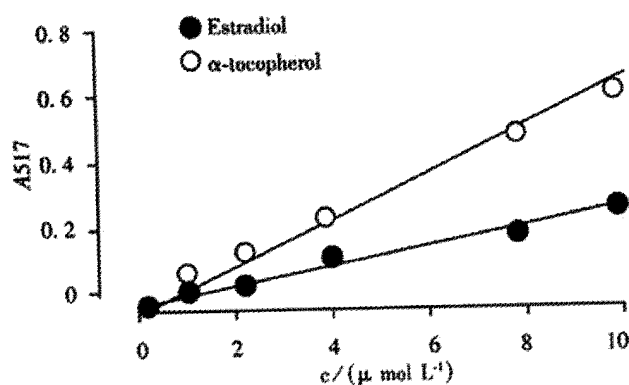


Figure 4 DPPH radical-scavenging activity of estradiol

DISCUSSION

Hepatic fibrosis and cirrhosis occur more frequently in men than in women. The ratio of male:female has been reported to range from 2.3:1 to 2.6:1 which indicates that sex hormones may play a role in the development of hepatic fibrosis and subsequent cirrhosis. We have found that estradiol treatment resulted in reduced hepatic fibrosis in male rat in which hepatic fibrosis had been induced by dimethylnitrosamine (DMN), and this phenomenon had been proven to be associated with the inhibitory effect of estradiol on the activation of lipocyte which is essential for hepatic fibrogenesis. Lipocyte can be activated by inflammatory factors released from Kupffer cells or free radicals and lipid peroxides formed in injured liver. Therefore, we examined the effect of estradiol on the cultured hepatocytes to evaluate its protective activity against oxidative damage of the liver. Estradiol is a steroidal compound that binds to specific intracellular receptors which act as transcription factors. Although the liver is not the classical sex hormone target, but livers in both men and women contain high affinity, low capacity estradiol receptors, and they are believed to respond to estradiol regulation. Another transcription independent activity of the molecule estradiol is its intrinsic antioxidant activity which makes it a potential chemical shield for cells^[37,38]. The neuroprotective effect of estradiol from oxidative damage has been extensively investigated in recent years^[39,40]. It is also noteworthy that oxygen-derived free radicals and lipid peroxidation have been implicated in hepatic injury^[41]. Our data also demonstrated that estradiol could inhibit the lipid peroxidation in hepatocyte and the free radicals induced-hepatocytic injury, and thus exerted its suppressive effect on liver fibrosis.

Hepatocytic damages induced by oxidative stress include two forms: cell necrosis and apoptosis. We also found that estradiol could inhibit the apoptosis of hepatocytes in the face of oxidative challenges. To elucidate the mechanism of its antiapoptotic effect, we examined the relationship between estradiol administration and the Bcl-2 and Bcl-x expression level by Western blotting. Figure 3 showed that Bcl-xl level in hepatocytes was increased under oxidative stress, while administration of estradiol attenuated the increased expression of Bcl-x induced by oxidative stress. These data suggested that Bcl-x expression responded directly to oxidative stress as a protective reaction, and estradiol could counteract the oxidative stress and stabilized the Bcl-xl expression before its antioxidant activity was depleted. In this study, Bcl-2 expression was not detectable even though two antibodies from different companies were used because of its low level in hepatocytes. But in another investigation we found that estradiol administration dramatically increased the Bcl-2 level in fibrotic liver tissue of rat induced by DMN injection (data not presented). Therefore, the mechanism of the antiapoptotic effect of estradiol is still elusive. The regulators, which participate in this pathway need further investigation. In conclusion, our findings suggested that estradiol could protect hepatocytes from oxidative stress, and the transcription-independent antioxidant activity of estradiol molecule may play a major role in this pathway.

REFERENCES

- 1 Wu CH. Fibrodynamics-elucidation of the mechanisms and sites of liver fibrogenesis. *World J Gastroenterol* 1999;5:388-390
- 2 Bissell DM. Hepatic fibrosis in wound repair: a progress report. *J Gastroenterol* 1998;33:295-302
- 3 Friedman SL. Molecular regulation of hepatic fibrosis, an integrated cellular response to tissue injury. *J Biol Chem* 2000;275:2247-2250
- 4 Brenner DA, Waterboer T, Choi SK, Lindquist JN, Stefanovic B, Burchardt E, Yamauchi M, Gillan A, Rippe RA. New aspects of hepatic fibrosis. *J Hepatol* 2000;32(Suppl 1):32-38
- 5 Albanis E, Friedman SL. Hepatic fibrosis. Pathogenesis and principles of therapy. *Clin Liver Dis* 2001;5:315-334
- 6 Santra A, Maiti A, Das S, Lahiri S, Chakaborty SK, Mazumder DN. Hepatic damage caused by chronic arsenic toxicity in experimental animals.

- J Toxicol Clin Toxicol* 2000;38:395-405
- 7 Maddrey WC. Alcohol-induced liver disease. *Clin Liver Dis* 2000;4: 115-131
- 8 Neubauer K, Saile B, Ramadori G. Liver fibrosis and altered matrix synthesis. *Can. J Gastroenterol* 2001;15:187-193
- 9 Shiba M, Shimizu I, Yasuda M, Ii K, Ito S. Expression of type I and type III collagens during the course of dimethylnitrosamine-induced hepatic fibrosis in rats. *Liver* 1998;18:196-204
- 10 Benyon RC, Arthur MJ. Extracellular matrix degradation and the role of hepatic stellate cells. *Semin Liver Dis* 2001;21:373-384
- 11 Ide M, Yamate J, Kuwamura M, Kotani T, Sakuma S, Takeya M. Immunohistochemical analysis of macrophages and myofibroblasts appearing in hepatic and renal fibrosis of dogs. *J Comp Pathol* 2001;124:60-69
- 12 Eng FJ, Friedman SL. Fibrogenesis I. New insights into hepatic stellate cell activation: the simple becomes complex. *Am J Physiol Gastrointest Liver Physiol* 2000;279:G7-G11
- 13 Eng FJ, Friedman SL. Transcriptional regulation in hepatic stellate cells. *Semin Liver Dis* 2001;21:385-395
- 14 Wei HS, Lu HM, Li DG, Zhan YT, Wang ZR, Huang X, Cheng JL, Xu QF. The regulatory role of AT 1 receptor on activated HSCs in hepatic fibrogenesis: effects of RAS inhibitors on hepatic fibrosis induced by CCl₄. *World J Gastroenterol* 2000;6:824-828
- 15 Iredale JP. Hepatic stellate cell behavior during resolution of liver injury. *Semin Liver Dis* 2001;21:427-436
- 16 Bataller R, Brenner DA. Hepatic stellate cells as a target for the treatment of liver fibrosis. *Semin Liver Dis* 2001;21:437-451
- 17 Pinzani M, Marra F, Carloni V. Signal transduction in hepatic stellate cells. *Liver* 1998;18:2-13
- 18 Du WD, Zhang YE, Zhai WR, Zhou XM. Dynamic changes of type I, III and IV collagen synthesis and distribution of collagen-producing cells in carbon tetrachloride induced rat liver fibrosis. *World J Gastroenterol* 1999;5:397-403
- 19 Wang LT, Zhang B, Chen JJ. Effect of anti-fibrosis compound on collagen expression of hepatic cells in experimental liver fibrosis of rats. *World J Gastroenterol* 2000;6:877-880
- 20 Liu HL, Li XH, Wang DY, Yang SP. Matrix metalloproteinase-2 and tissue inhibitor of metalloproteinase-1 expression in fibrotic rat liver. *World J Gastroenterol* 2000;6:881-884
- 21 Ramadori G, Armbrust T. Cytokines in the liver. *Eur J Gastroenterol Hepatol* 2001;13:777-784
- 22 Poli G. Pathogenesis of liver fibrosis: role of oxidative stress. *Mol Aspects Med* 2000;21:49-98
- 23 Lu LG, Zeng MD, Li JQ, Hua J, Fan JG, Fan ZP, Qiu DK. Effect of lipid on proliferation and activation of rat hepatic stellate cells (I). *World J Gastroenterol* 1998;4:497-499
- 24 Kim KY, Choi I, Kim SS. Progression of hepatic stellate cell activation is associated with the level of oxidative stress rather than cytokines during CCl₄-induced fibrogenesis. *Mol Cells* 2000;10:289-300
- 25 Hu YY, Liu CH, Wang RP, Liu C, Liu P, Zhu DY. Protective actions of salvianolic acid A on hepatocyte injured by peroxidation *in vitro*. *World J Gastroenterol* 2000;6:402-404
- 26 Chen PS, Zhai WR, Zhou XM, Zhang JS, Zhang YE, Ling YQ, Gu YH. Effects of hypoxia, hyperoxia on the regulation of expression and activity of matrix metalloproteinase-2 in hepatic stellate cells. *World J Gastroenterol* 2001;7:647-651
- 27 Greenwel P, Dominguez-Rosales JA, Mavi G, Rivas-Estilla AM, Rojkind M. Hydrogen peroxide: a link between acetaldehyde-elicited alpha1(I) collagen gene up-regulation and oxidative stress in mouse hepatic stellate cells. *Hepatology* 2000;31:109-116
- 28 Nalini G, Hariprasad C, Narayanan VA. Oxidative stress in alcoholic liver disease. *Indian J Med Res* 1999;110:200-203
- 29 Shimizu I, Mizobuchi Y, Yasuda M, Shiba M, Ma YR, Horie T, Liu F, Ito S. Inhibitory effect of oestradiol on activation of rat hepatic stellate cells *in vivo* and *in vitro*. *Gut* 1999;44:127-136
- 30 Shimizu I, Yasuda M, Mizobuchi Y, Ma YR, Liu F, Shiba M, Horie T, Ito S. Suppressive effect of oestradiol on chemical hepatocarcinogenesis in rats. *Gut* 1998;42:112-119
- 31 Yasuda M, Shimizu I, Shiba M, Ito S. Suppressive effects of estradiol on dimethylnitrosamine-induced fibrosis of the liver in rats. *Hepatology* 1999;29:719-727
- 32 Adams JM, Cory S. Life-or-death decisions by the Bcl-2 protein family. *Trends Biochem Sci* 2001;26:61-66
- 33 Zornig M, Hueber A, Baum W, Evan G. Apoptosis regulators and their role in tumorigenesis. *Biochim Biophys Acta* 2001;1551:F1-37
- 34 Budd RC. Activation-induced cell death. *Curr Opin Immunol* 2001;13: 356-362
- 35 Liu HF, Liu WW, Fang DC, Men RP. Expression of bcl-2 protein in gastric carcinoma and its significance. *World J Gastroenterol* 1998;4: 228-230
- 36 Grad JM, Zeng XR, Boise LH. Regulation of Bcl-xL: a little bit of this and a little bit of STAT. *Curr Opin Oncol* 2000;12:543-549
- 37 Huh K, Shin US, Choi JW, Lee SI. Effect of sex hormones on lipid peroxidation in rat liver. *Arch Pharm Res* 1994;17:109-114
- 38 Leal AM, Begona Ruiz-Larrea M, Martinez R, Lacort M. Cytoprotective actions of estrogens against tert-butyl hydroperoxide-induced toxicity in hepatocytes. *Biochem Pharmacol* 1998;56:1463-1469
- 39 Sawada H, Shimohama S. Neuroprotective effects of estradiol in mesencephalic dopaminergic neurons. *Neurosci Biobehav Rev* 2000;24: 143-147
- 40 Dare E, Gotz ME, Zhivotovsky B, Manzo L, Ceccatelli S. Antioxidants J811 and 17beta-estradiol protect cerebellar granule cells from methylmercury-induced apoptotic cell death. *J Neurosci Res* 2000; 62:557-565
- 41 Bruck R, Shirin H, Aeed H, Matas Z, Hochman A, Pines M, Avni Y. Prevention of hepatic cirrhosis in rats by hydroxyl radical scavengers. *J Hepatol* 2001;35:457-464

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