

# Observation of DNA damage of human hepatoma cells irradiated by heavy ions using comet assay

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## Abstract

**AIM:** Now many countries have developed cancer therapy with heavy ions, especially in GSI (Gesellschaft für Schwerionenforschung mbH, Darmstadt, Germany), remarkable results have obtained, but due to the complexity of particle track structure, the basic theory still needs further researching. In this paper, the genotoxic effects of heavy ions irradiation on SMMC-7721 cells were measured using the single cell gel electrophoresis (comet assay). The information about the DNA damage made by other radiations such as X-ray,  $\gamma$ -ray, UV and fast neutron irradiation is very plentiful, while little work have been done on the heavy ions so far. Hereby we tried to detect the reaction of liver cancer cells to heavy ion using comet assay, meanwhile to establish a database for clinic therapy of cancer with the heavy ions.

**METHODS:** The human hepatoma cells were chosen as the test cell line irradiated by 80Mev/u  $^{20}\text{Ne}^{10+}$  on HIRFL (China), the radiation-doses were 0, 0.5, 1, 2, 4 and 8 Gy, and then comet assay was used immediately to detect the DNA damages, 100-150 cells per dose-sample (30-50 cells were randomly observed at constant depth of the gel). The tail length and the quantity of the cells with the tail were put down. EXCEL was used for statistical analysis.

**RESULTS:** We obtained clear images by comet assay and found that SMMC-7721 cells were all damaged apparently from the dose 0.5Gy to 8Gy ( $t$ -test:  $P < 0.001$ , vs control). The tail length and tail moment increased as the doses increased, and the number of cells with tails increased with increasing doses. When doses were higher than 2Gy, nearly 100 % cells were damaged. Furthermore, both tail length and tail moment, showed linear equation.

**CONCLUSION:** From the clear comet assay images, our experiment proves comet assay can be used to measure DNA damages by heavy ions. Meanwhile DNA damages have a positive correlation with the dose changes of heavy ions and SMMC-7721 cells have a great radiosensitivity to  $^{20}\text{Ne}^{10+}$ . Different reactions to the change of doses indicate that comet assay is a useful tool to detect DNA damage induced by heavy ions.

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## INTRODUCTION

During the past few decades, radiation research has developed into specialized sub-disciplines, from basic physics and chemistry to tumor biology and experimental radiation therapy<sup>[1]</sup>. Although the radiobiological effects are extensively investigated for X-ray and  $\gamma$ -ray, little work has been directed towards heavy ion beams. With the exploration of the outer space, the research of high linear energy transfer (LET) has attracted more and more attention. Since heavy ions were first applied in the mid-1970s to cure cancer at Bevalac of Lawrence Berkeley Laboratory (LBL), United States, promising results have been reported when compared with the conventional radiotherapy for soft tissue sarcoma, bone sarcoma and prostate cancer<sup>[2]</sup>. Now scientists in many countries (GSI in Germany, HIMAC in Japan<sup>[25,26]</sup>, HIRFL in China) have designed accelerators to deliver beams of ions for the treatment and started basic researches of cancer therapy with heavy ions such as carbon, neon, oxygen and argon.

The aim of our present study was to investigate DNA damages induced by heavy ions by comet assay. The theoretical value was then compared to responses to external X-ray or  $\gamma$ -ray and other irradiations, so that we could establish the data base for clinical therapy.

Comet assay, the alkaline version in particular, has become a very popular method for the analysis of DNA damage caused by various chemical and physical agents because of its simplicity and rapidity<sup>[4-10]</sup>. DNA damages consisted of DNA strand breakage, alkali-labile sites and incomplete excision repair sites<sup>[11]</sup>. Although the direct DNA-breaking capacity can be estimated by alkaline elution, nick translation and alkaline single cell gel electrophoresis (SCGE), SCGE has been shown to be more sensitive than the former two. It had been proved that the sensitivity of SCGE is significantly higher than that of cytokinesis-blocked micronucleus (CBMN) test<sup>[12]</sup>. The most important point is that comet assay is an electrophoretic technique, which allows measurements of DNA damages as well as DNA repair rates on an individual cell. Therefore its contribution to DNA damages by irradiating cells with heavy ions at once or after a while can be reflected as initial damages or residual DNA damages, if time is allowed for enzymatic repairs of initial DNA strand break. In our lab, we focused on the radiobiological effects of heavy ions on tumor cells. Previous experiments were mainly on cell survival measurements and could not explain the underlying radiosensitization mechanism at molecular level<sup>[2]</sup>. To verify the radiobiological effects of heavy ions on cellular DNA, SCGE also called comet assay was used to directly measure DNA damages in cells.

## MATERIALS AND METHODS

### Cells and cell culture

Human hepatoma cells SMMC-7721, purchased from Second Military Medical University in Shanghai, were cultivated in

RMIP-1640 medium (Gibco product) supplemented with 15 % calf serum in a standard incubator at 37 °C. One passage of cells every 2-3 days and change of the medium everyday were performed, to ensure the cells growth in good conditions. Two days before the irradiation, the cells were shifted to  $\Phi$  35 mm petri-dishes, each had 2 ml cell suspension, and the density of the cells was  $5 \times 10^4$  cells/ml. Each dose had 5 petri-dishes. Before irradiation, cells in each petri-dish were examined under the inverted light microscope to select materials good in growth and even in density, and medium 1640 in petri-dishes was removed, only Dulbecc' s phosphate -buffered saline medium (PBS) was left to keep the moisture of the cells when irradiated.

### Selection of ion beams

Irradiation was performed using  $^{20}\text{Ne}^{10+}$  with energy of 80 Mev/u and intensity of 0.00136nA ( $2.1 \times 10^6$ p/s). The cells were irradiated at the doses of 0, 0.5, 1, 2, 4, 8Gy. The doses of cells were measured by an air ionization chamber.

### Preparation of single cell suspension and comet assay

As soon as irradiation ended, the cells were washed and collected, the final concentration of cells was adjusted to  $(5-10) \times 10^6$  by adding Dulbecc' s phosphate -buffered saline medium (PBS) to the single cell suspension.

The alkaline version of comet assay was carried out based on the work of Ostling and Johansson with some minor modifications as followings: On the day of electrophoresis, an aliquot of 10  $\mu$ l freshly prepared suspension of cells was mixed with 30  $\mu$ l 0.5 % low- melting-agarose in Dulbecc' s PBS (pH 7.4). The mixture was layered on top of an ordinary microscope slide precoated with 0.5 % normal-melting -agorose, which was allowed to dry at room temperature protected from dust and other particles. After low-melting-agarose was solidified in a refrigerator for 10 minutes, the coverslip was carefully removed and the slide was gently immersed in a freshly prepared lysing solution (2.5MNaCl, 10mM Tris, 1 % sodium lauryl sarcosinate, 100mM Na<sub>2</sub>EDTA, with 1 % Triton-100 and 10 % DMSO added just before use). From this moment until the end of neutralization, all steps needed to avoid the sunlight.

After lysis for 1-1.5 h, the microscopy slides were transferred to electrophoresis session, 18 microscopy slides from 6 samples (3 slides/each sample) were randomly placed in a electrophoretic unit.

After 20-30 minutes of DNA unwinding in electrophoresis buffer (1 mM EDTA-Na<sub>2</sub>, 300 mM NaOH, pH>13), single cell gel electrophoresis was performed in the same buffer (20 min, 20 V, 300 nA). After electrophoresis the slides were neutralized with 0.4 M Tris buffer (pH7.5).

### Evaluation of DNA damage

The microscopy slides were stained with ethidium bromide in water (40  $\mu$ g/ml, 50  $\mu$ l/slide). After application of a coverslip was removed, each slide was examined at 10 $\times$ 20 magnification in a fluorescence microscope (excitation filter: 400nm, barrier filter: 590nm). 100-150 cells per dose-sample (30-50 cell were randomly observed at constant depth of the gel, avoiding the edges of the gel on each of three replicate slides). The tail length and the quantity of the cells with the tail were put down, at the same time, photos were taken with 135# black and white film (ISO 400). Then analysis was done using EXCEL.

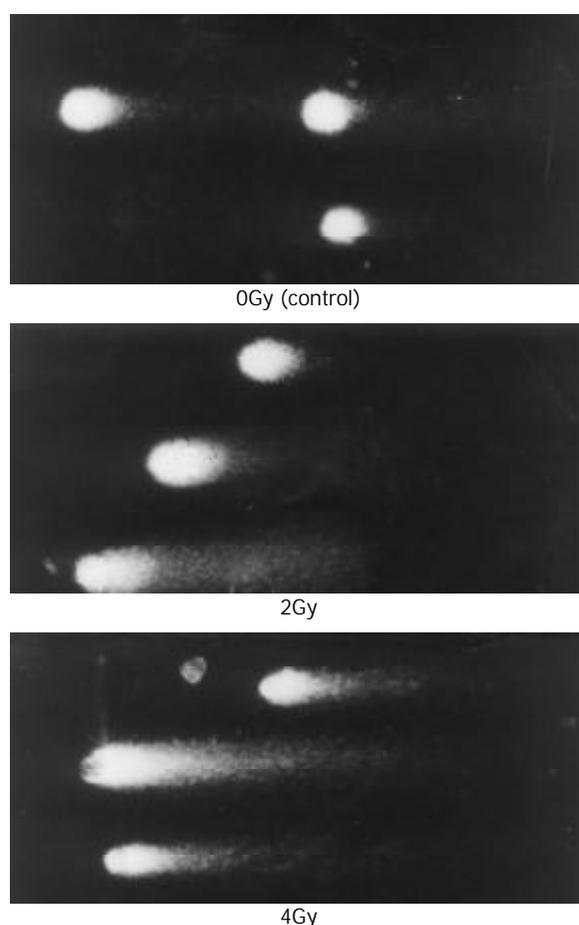
## RESULTS AND DISCUSSION

### Formation of comet assay images-DNA loops and alkaline unwinding

The comet assay is attractive for many reasons. Apart from

being a quick, simple, sensitive, reliable and fairly inexpensive way of measuring, it also produces appealing images.

There are two explanations about what the comet tails consist of. One is that it is a fragment DNA, the other is that the length of such a fragment is about 1  $\mu$ m, but the length of the tail of a comet is a few percentages of it<sup>[13-16]</sup>, as to our experiment the longest mean length of tail was no more than 200  $\mu$ m. Nuclear DNA is not a tangle of string, even after treatment with detergents and a strong salt solution, as in the SCGE procedure, the nuclear (or nucleoid) had a structure, the DNA was organized as loops, which retained the super coils that were contained in the nucleosome. Cook *et al*<sup>[17]</sup> deduced the presence of supercoiled loops and then they observed that, when DNA was broken by irradiation, supercoiling was relaxed and loops spilled out into a 'halo' around the nucleoid. By analogy, it is assumed that the Comet tail is made up of relaxed loops, and that the number of loops in the tail indicates the number of DNA breaks.



**Figure 1** Comet assay image at different doses.

The alkaline comet assay can detect DNA breaks including single and double DNA strand breaks, and AP sites, which are alkali-labile and probably converted to breaks while DNA is in the electrophoretic solution at high pH<sup>[14-16]</sup>. The present comet assay is generally practiced including incubation of DNA at high pH before and during electrophoresis, different from the original work of Ostling and Johansson who employed near-neutral pH. Collins AR<sup>[13]</sup> proved that both the neutral and alkaline methods could detect low levels of DNA damage, however, the breaks by higher levels of damage were more clearly resolved from the head under alkaline conditions<sup>[18]</sup>. Thus in our experiment, the alkaline version was used.

Furthermore, breaks will be transiently present when cells repair lesions via base excision or nucleotide excision so that

a high level of breaks in the Comet assay may indicate either severe damage or efficient repair<sup>[13]</sup>. In fact, much useful information can be obtained by exploiting cellular repair to produce DNA breaks and thus to reveal or amplify the effect of radiation that otherwise may not show positive effects by the comet assay. This will be discussed in our later work on the repair effects of heavy ions.

**DNA damage caused by heavy ions**

The SCGE test or comet assay is a straightforward visual method for the detection of DNA damage of cells in interphase. This technique is especially sensitive in detecting DNA single-strand breaks, alkali-labile damage and excision repair sites in individual cells<sup>[13-16]</sup>. The Comet assay has been widely applied in the following fields: radiation biology<sup>[5-6,9-12,19]</sup>, excisable DNA damage, DNA cross-links<sup>[20]</sup>, oxidative damage<sup>[21,22]</sup>, genetic toxicology and apoptosis<sup>[23,24]</sup>.

Ionizing radiation is a ubiquitous environmental agent. Its physiochemical interaction with cellular DNA produces a variety of primary lesions, such as single-strand breaks (SSB), double-strand breaks (DSB), DNA-DNA and DNA-protein crosslinks, and damage to purine and pyrimidine bases. And using ionizing radiation may avoid complications of drug metabolism, intracellular distribution, membrane permeability and drug efflux. Although the technique of SCGE is very sensitive to ionizing radiation, information about DNA damage made by other radiations such as X-ray<sup>[10,12]</sup>,  $\gamma$ -ray<sup>[8,9]</sup>, UV<sup>[9]</sup> and fast neutron<sup>[19]</sup> irradiation is very plentiful, little work has been done on heavy ions so far.

Microdosimetric considerations suggest that, in a given type of radiation, the yield DNA damage must be proportional to dose, so that besides the influence of changing repair efficiency, the magnitude of the dose might not be expected to be critical in comparison of the results. Heavy ion is a kind of high LET (Linear Energy Transfer) irradiation, as emphasized long ago by Lea, high-LET radiation could, through the increased frequency of DSB in close proximity, cause interactive damages and misrepair<sup>[27-28]</sup>. We anticipated that heavy ions probably had strong effects on the cellular DNA, but we wonder if it can make the linear equation after irradiation by heavy ions.

In our experiment, the data for DNA damages induced by heavy ions at the doses of 0-8Gy are presented in Table 1. The dose-response curves for tail length and tail moment (the fraction of DNA in the tail multiplied by tail length) are shown in Figure 2. We could see tail length and tail moment showed linear equation. It proved that SMMC-7721 cells have high radiosensitivity to heavy ions and comet assay is very useful to detect DNA damages induced by heavy ions.

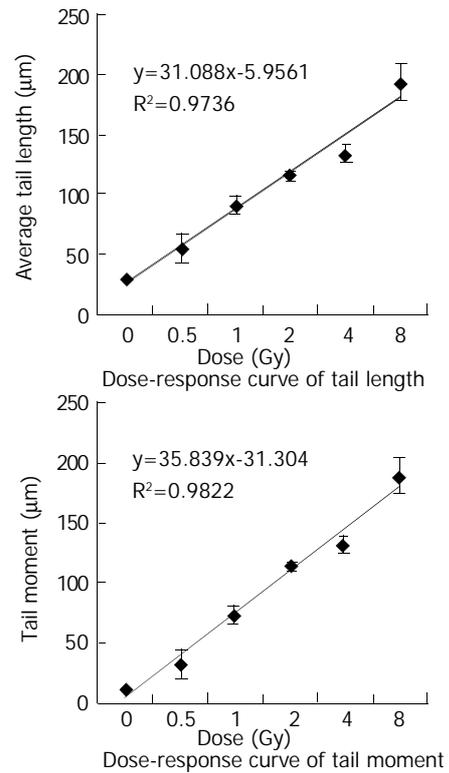
Figure 3 shows the change of tail DNA as the dose increased. It was found that almost 100 % cells were damaged when the dose reached at 2Gy. But the details were unknown about how badly DNA was damaged when the dose was higher than 2Gy. To completely evaluate DNA damages by different doses, comets of every dose-sample were sorted visually into classes 0-4, representing increasing amount of damage. Figure 4, result shows that with increase of the dose, slighter damage of DNA tail (class 0-2) converted to more severe change of DNA (DNA migrated from the head to form longer and longer tail).

We should pay more attention to the dose of 2Gy, which is the conventional choice of  $\gamma$ -ray radiotherapy. At the dose of 2 Gy, 100 % cells were damaged, with different grade of DNA damage (classes 1-4). Among them nearly 25 % cells were badly damaged. It is known that a central phenomenon in radiobiology is the efficiency of densely ionizing radiation for cellular effects. As in our experiment, chromosomal aberrations or cell killing occurred on these badly damaged cells<sup>[27-30, 33-35]</sup>. Additionally, we noticed that nearly 80 % cells

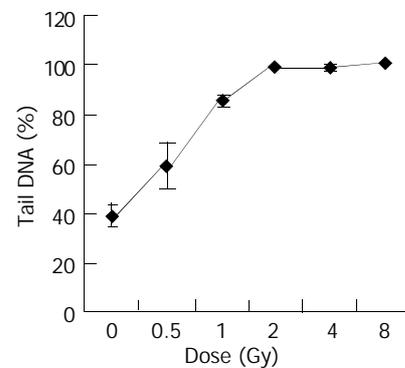
were most severely damaged (class 4) at the dose of 8Gy. It showed that 8Gy might be or near the highest dose that the cells could withstand.

**Table 1** Values of damages detected by Comet assay after 80Mev/u <sup>20</sup>Ne<sup>10+</sup> irradiation

Dose Gy	Tail length mean $\pm$ S.E. $\mu$ m	Tail DNA mean $\pm$ S.E. %	Tail moment mean $\pm$ S.E.	t-test P
0	29.44 $\pm$ 1.46	38.50 $\pm$ 3.50	11.53 $\pm$ 1.45	
0.5	54.18 $\pm$ 11.74	59.21 $\pm$ 9.21	33.02 $\pm$ 11.80	2.06227E-07
1	90.16 $\pm$ 6.66	85.54 $\pm$ 2.21	76.96 $\pm$ 3.44	8.7291E-21
2	115.09 $\pm$ 3.26	100.00 $\pm$ 0.00	115.09 $\pm$ 3.25	6.97944E-31
4	134.17 $\pm$ 8.18	98.86 $\pm$ 1.07	134.17 $\pm$ 8.10	4.48617E-68
8	194.08 $\pm$ 15.58	100.00 $\pm$ 0.00	194.08 $\pm$ 15.58	6.4087E-104

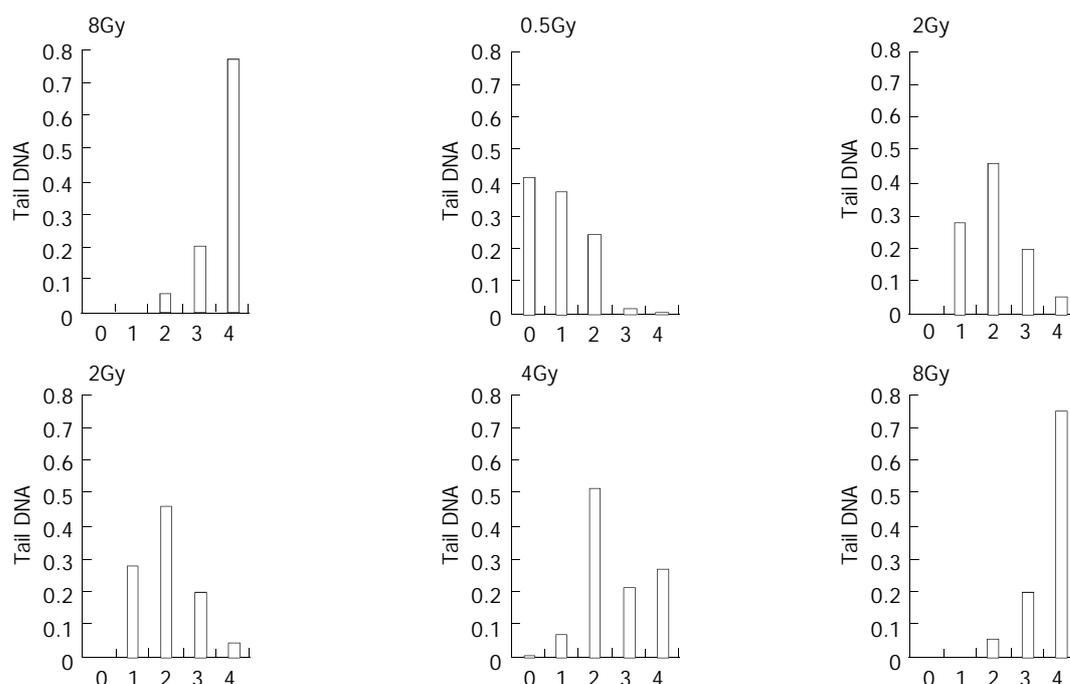


**Figure 2** Curve of tail length and tail moment.



**Figure 3** Dose-response curve of tail DNA.

One thing to be mentioned here is that the relative biological effectiveness (RBE) tends to increase with linear energy transfer (LET). For very heavy ions with LET in excess of about 100-200 keV/ $\mu$ m, a more complex dependence on particle track structure emerges<sup>[31,32]</sup>. Therefore, the study on particle track structure is very important for further research.



**Figure 4** Comet class at the dose of 0-8Gy of heavy ions irradiation.

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