

Studies on specific interaction of beta-2-glycoprotein I with HBsAg

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Supported by the National Natural Science Foundation of China, No.30070338

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Received: 2002-11-05 **Accepted:** 2002-12-12

Abstract

AIM: To observe the binding activity of beta-2-glycoprotein I (β_2 GPI) to hepatitis B surface antigen (HBsAg) and the possible roles of β_2 GPI in hepatitis B virus (HBV) infection.

METHODS: The rationale of ELISA methods and ELISA-based research method and ligand-blotting technique were used to detect the specific interaction of β_2 GPI with HBsAg.

RESULTS: With the increase of rHBsAg, the binding of β_2 GPI to rHBsAg elevated, and these changes had statistic significance. When we added non-biotinylated β_2 GPI, the OD value significantly decreased though they still were positively relevant to rHBsAg, suggesting non-biotinylated β_2 GPI competed with biotinylated β_2 GPI to saturate the binding sites on rHBsAg. Meanwhile BSA was used as negative control to substitute for rHBsAg coating the plates. The results indicated no interaction between β_2 GPI and BSA, suggesting the affinity of β_2 GPI to rHBsAg was specific. The ligand blotting indicated that β_2 GPI might bind to rHBsAg no matter whether it was under reduced condition or not.

CONCLUSION: The binding of β_2 GPI to HBsAg suggests that β_2 GPI may be a carrier of HBV and that β_2 GPI may play important roles in HBV infection.

Gao PJ, Piao YF, Liu XD, Qu LK, Shi Y, Wang XC, Yang HY. Studies on specific interaction of beta-2-glycoprotein I with HBsAg. *World J Gastroenterol* 2003; 9(9): 2114-2116
<http://www.wjgnet.com/1007-9327/9/2114.asp>

INTRODUCTION

β_2 GPI (beta-2-glycoprotein I) is a plasma glycoprotein circulating as a free protein and is associated to lipoproteins. This protein is also referred to as apolipoprotein H (Apo H). Human β_2 GPI is a single-chain molecule consisting of 326 amino acid residues and 5 carbohydrate chain and has a molecule mass of approximately 55 kDa^[1-3]. The amino acid sequences of β_2 GPI in human, bovine, mouse, and rat appear to be highly conserved^[4-6]. The protein contains 5 internal repeat

unit of 60 amino acid residues, each with 2 internal disulfide bonds, known as Sushi domain^[7].

Although quite a lot is known about the structure of β_2 GPI, its biological function remains unclear. It is known that β_2 GPI can bind to negatively charged substances such as DNA and heparin and negatively charged phospholipids etc *in vivo*^[8,9], but the meaning of such interaction is still unclear. It is known that β_2 GPI may serve as a major factor in clearing the plasma liposome^[10] as well as an anticoagulant in blood^[11]. β_2 GPI may also modulate the function of kidney and placenta. The abnormality of plasma β_2 GPI level has been shown to be associated with many diseases such as arterial and venous thrombosis, recurrent abortion and alcoholic liver disease^[12,13]. In addition, it has been found that, in diabetes or atherosclerosis patients, the concentration of plasma β_2 GPI increases and distribution of the protein among different types of lipoprotein is perturbed^[14,15]. Recent research on β_2 GPI has given a further impetus to the discovery that lipid-associated β_2 GPI can bind to some pathogenic antigens or proteins such as HBV Dane particles, protein p18, p26, gp160 of HIV and so on^[16], and perhaps hepatitis virus antigen^[17,18]. These findings highlighted a potentially critical role of β_2 GPI in the mechanism of hepatitis B, AIDS, and systemic lupus erythematosus, etc.

The lipid-binding and transportation functions are considered as a basic mechanism related to its physiological and pathogenic functions. It has been demonstrated by several labs that β_2 GPI prefers to bind negatively charged phospholipids^[9,19-21]. It was reported that β_2 GPI could be removed from the membranes with a weakly acidic buffer and it was partly associated with chylomicrons and high-density lipoproteins, both of which were targeted to hepatocytes during the normal course of lipid metabolism^[22]. In the present paper, the characteristics of β_2 GPI interacting with HBsAg were further examined by several measurements. As a result, the standpoints that β_2 GPI might participate in HBV infection were held out accordingly. Our results may also help to explain how β_2 GPI facilitates HBV transportation, location and the important roles of β_2 GPI in HBV infection.

MATERIALS AND METHODS

Reagents

TEMED, APS, PMSF, and aprotinin purchased from Sigma Chemical Co., acrylamide and biacrylamide purchased from Serva, human β_2 GPI provided by Prof. Zhang GR from Jilin University, rHBsAg offered by Zhang HY from Jilin Institute of Family Plan, Hybond obtained from Amersham, SDS, NP-40, Coomassie brilliant blue obtained from Fluka (Finland). The other chemicals used were of analytical grade made in China.

ELISA-based determination of β_2 GPI-rHBsAg interaction

rHBsAg was diluted into 0.125 μ g/ml, 0.5 μ g/ml and 2 μ g/ml in 0.05 NaHCO₃ (pH 9.4) and added to 96-well plates respectively at 4 °C overnight (8 wells/group). Non-specific sites were blocked with PBS containing 1 % BSA for 1.5 h at 37 °C. Plates were washed three times between the different incubation steps. Biotinylated β_2 GPI was added to every well followed by adding HRP-Avidin and substrate consequentially. Simultaneously, 40 ng of non-labeled β_2 GPI was added to

4well/group as a competitor. OPD was used to develop color and the intensity of color was quantified at 492 nm with Model 550 Microplate Reader (Bio-Rad). In addition, BSA was used as negative control (2 ug/ml, 1 ug/ml, 0.2 ug/ml), to observe its binding to β_2 GPI.

SDS-PAGE and ligand blotting

The β_2 GPI protein was performed by SDS-PAGE using the vertical electrophoretic apparatus (Bio-Rad) on 12 % acrylamide under reduced and non-reduced condition. The gels were stained with Coomassie brilliant blue to visualize the proteins. The proteins separated were transferred onto nitrocellulose using an electric transfer system at 50 V for 16 min, followed by immunoblot analysis employing biotinylated rHBsAg as a detecting probe and DAB as a developing system. The blotting buffer consisted of phosphate-buffered saline (10 mM phosphate, pH7.5, 138 mM NaCl, 2.7 mM KCl) containing 5 % dried skimmed milk powder and 1 % Tween 20 detergent (Sigma). Biotinylated rHBsAg was used at a dilution of 1/200 and HRP-avidin (Huamei Company), a dilution of 1/200. The density was analyzed with Luzex-F Image-analysis System.

Statistical analysis

Results from quantitative parameters were presented as mean \pm SD and comparisons of which were performed using the Student's *t* test.

RESULTS

ELISA-based determination of β_2 GPI- rHBsAg interaction

rHBsAg was diluted into 0.125 μ g/ml, 0.5 μ g/ml and 2 μ g/ml and used as capture antibody to coat the plates, then biotinylated β_2 GPI was added to interact with rHBsAg, simultaneously we used non- biotinylated β_2 GPI to observe the competitive binding interaction. The following figure indicated that with the increase of rHBsAg the binding of β_2 GPI to rHBsAg elevated and these changes had statistic significance (*vs* 0.125 μ g/ml group, $P < 0.05$). When we added non- biotinylated β_2 GPI, the OD value significantly decreased though they still were positively relevant to rHBsAg, suggesting that non-biotinylated β_2 GPI competed with biotinylated β_2 GPI to saturate the binding sites on rHBsAg (Figure 1). Meanwhile BSA was used as negative control to substitute for rHBsAg coating the plates, the results indicated no interaction between β_2 GPI and BSA, (Figure 2) suggesting the affinity of β_2 GPI to rHBsAg was specific.

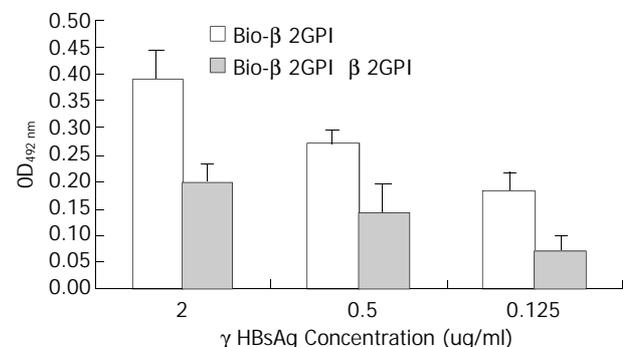


Figure 1 Comparative binding reaction of β_2 GPI to rHBsAg by ELISA-based determination of protein interaction.

Binding of β_2 GPI to rHBsAg observed by ligand blotting

The discrepancy between ligand blotting and Western blotting was that the former made use of ligand-receptor reaction, while the latter, the antigen-antibody interaction. The image showing -50 kDa protein bands in both lane 1 and lane 2 represented

reduced (with DTT) and non-reduced (without DTT) condition, respectively (Figure 3). The results indicated that β_2 GPI might bind to rHBsAg no matter whether it was under reduced condition or not, which manifested difference from that reported by Mehdi *et al*^[17]. The subsequent digital scanning by Luzex-F Imager confirmed no quantitative changes in density between lane 1 and lane 2.

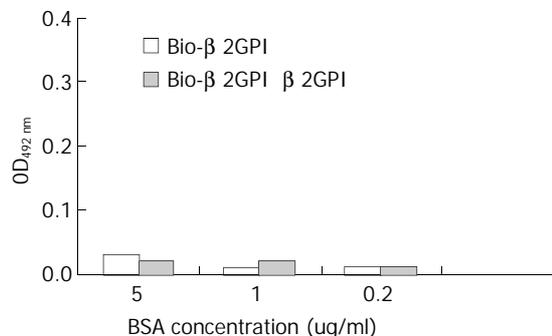


Figure 2 Comparative binding reaction of β_2 GPI to BSA by ELISA-based determination of protein interaction.

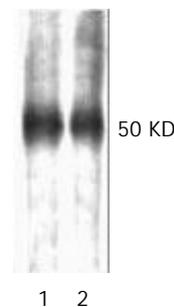


Figure 3 Reduced (lane1) and non-reduced (lane2) human β_2 GPI (1 μ g/lane) was separated with 12 % SDS-PAGE and applied to Hybond nitrocellulose membrane, blocked with 5 % skimmed milk, then probed with biotinylated rHBsAg, color was developed with HRP-avidin in DAB solution. Single band at -50 kDa was observed, no significant difference in color density was observed, which was further confirmed by digital scanning.

DISCUSSION

Hepatitis B virus (HBV) is a member of the hepadnavirus family, which includes duck HBV, woodchuck hepatitis virus and ground squirrel hepatitis virus. These viruses are highly infectious for their host animals, targeting primarily, though not exclusively, the liver. Although many researches have been done, it is not clear which of these virus' components is responsible for its attachment to a target cell^[23]. As a step toward identifying the mechanism of HBV targeting hepatocytes, Haider Mehdi reported that a 50 kDa protein known as β_2 GPI, which could be removed from the membranes with a weakly acidic buffer, might participate in the infection of HBV. Examination of human serum revealed that β_2 GPI was a serum protein. Isolation of plasma lipoproteins revealed that β_2 GPI was in part associated with chylomicrons and high-density lipoproteins, both of which are targeted to hepatocytes during the normal course of lipid metabolism^[21]. In this study, we further examined the possible roles of β_2 GPI in HBV infection. A ligand-blotting system was used to identify and partially characterize a rHBsAg-binding protein associated with hepatocyte membrane, plasma and lipoproteins. Immunoassay was performed to characterize the properties of rHBsAg binding to β_2 GPI and to demonstrate the specificity of this

interaction.

β_2 GPI is a glycoprotein with four N-linked carbohydrate chains^[2,3] present at concentrations of approximately 200 μ g/ml in serum^[17]. The 326-residue mature protein is composed of a 61 amino acid motif repeated four times, followed by one longer, modified repeat. Each of the first four repeats contains four cysteines in conserved positions, at least one of which is critical for its rHBsAg-binding activity. This pattern of disulfide bonds is similar to the short consensus repeat units found in a family of approximately 20 proteins which includes many of the complement control proteins. It is interesting that the measles virus receptor, CD46^[24,25] also belongs to this short consensus repeat family.

The work described in the present report was to reveal the possible roles of β_2 GPI in HBV infection. As we have known that β_2 GPI may target to hepatocytes in the form of chylomicrons and high-density lipoproteins. The result of the present paper suggested that β_2 GPI might specifically bind to HBsAg no matter whether it was under reduced condition or not. We might well reckon that β_2 GPI might contribute to transportation of HBV to hepatocytes as a kind of carrier. Since β_2 GPI is associated with lipoproteins, particularly chylomicrons and HDL, it is possible that HBV binds to β_2 GPI on the surface of these lipoprotein particles. In the bloodstream, chylomicrons are partially degraded by lipoprotein lipase, resulting in chylomicron remnants, which are taken up by hepatocytes. HDLs are also taken up by hepatocytes in the process of "reverse cholesterol transport". They may have been associated with the process of HBV infection. It is tempted to speculate that infectious HBV might bind to chylomicron or HDL particles by interacting with β_2 GPI and be taken into hepatocytes as a "hitchhiker" along with these lipoproteins. Our results showed that β_2 GPI might bind to rHBsAg no matter whether it was under reduced condition or not manifested difference from that reported by Mehdi *et al*^[17]. In Mehdi experimental system, they used human serum to observe the relationship between β_2 GPI and HBsAg, while we used purified β_2 GPI. There might exist differences between the two kinds of β_2 GPI sources in their first, second or third structure, which might lead to the differences of sensitivity to reduced-agents. Moreover, the genetic heterogeneity of β_2 GPI could also cause mutations of sequences of some domains, which eventually affect the binding of β_2 GPI to rHBsAg. Related issues should be further investigated.

To sum up, if HBV infection to hepatocytes involves β_2 GPI on chylomicrons and/or HDL, it is a novel mechanism for virus attachment and entry.

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