World Journal of Diabetes

World J Diabetes 2022 September 15; 13(9): 668-801





Contents

Monthly Volume 13 Number 9 September 15, 2022

REVIEW

668 Nonalcoholic fatty liver disease and diabetes

> Bellini MI, Urciuoli I, Del Gaudio G, Polti G, Iannetti G, Gangitano E, Lori E, Lubrano C, Cantisani V, Sorrenti S, D'Andrea V

- 683 A review of potential mechanisms and uses of SGLT2 inhibitors in ischemia-reperfusion phenomena Quentin V, Singh M, Nguyen LS
- 696 Evolving spectrum of diabetic wound: Mechanistic insights and therapeutic targets Chakraborty R, Borah P, Dutta PP, Sen S

MINIREVIEWS

- 717 Potential role of Limosilactobacillus fermentum as a probiotic with anti-diabetic properties: A review Lacerda DC, Trindade da Costa PC, Pontes PB, Carneiro dos Santos LA, Cruz Neto JPR, Silva Luis CC, de Sousa Brito VP, de Brito Alves JL
- COVID-19 associated diabetes mellitus: A review 729 Gavkare AM, Nanaware N, Rayate AS, Mumbre S, Nagoba BS
- 738 Effectiveness and safety of COVID-19 vaccines in patients with diabetes as a factor for vaccine hesitancy Vasilev G, Kabakchieva P, Miteva D, Batselova H, Velikova T

ORIGINAL ARTICLE

Basic Study

752 Hyperglycemia and reduced adiposity of streptozotocin-induced diabetic mice are not alleviated by oral benzylamine supplementation

Carpéné C, Stiliyanov Atanasov K, Les F, Mercader Barcelo J

765 Role of insulin in pancreatic microcirculatory oxygen profile and bioenergetics

Li BW, Li Y, Zhang X, Fu SJ, Wang B, Zhang XY, Liu XT, Wang Q, Li AL, Liu MM

Retrospective Study

Relationship between age of pregnant women with gestational diabetes mellitus and mode of delivery and 776 neonatal Apgar score

Gao L, Chen CR, Wang F, Ji Q, Chen KN, Yang Y, Liu HW

SCIENTOMETRICS

786 Mapping the global research landscape on insulin resistance: Visualization and bibliometric analysis Zyoud SH, Shakhshir M, Koni A, Abushanab AS, Shahwan M, Jairoun AA, Al Subu R, Abu Taha A, Al-Jabi SW

World Journal of Diabetes

Contents

Monthly Volume 13 Number 9 September 15, 2022

799 Different nutrient compositions in diet and taking hypoglycemic drugs can modulate gut microbial flora Lin ZJ, Zhang QW, Yu XL, Zhou B, Liu CW, He LP

II

Contents

Monthly Volume 13 Number 9 September 15, 2022

ABOUT COVER

Peer Reviewer of World Journal of Diabetes, Nahlah Elkudssiah Ismail, BPharm (Hons), PhD, Professor, Department of Pharmacy, MAHSA University, Kuala Lumpur 42610, Malaysia. elkudssiah77@yahoo.com

AIMS AND SCOPE

The primary aim of World Journal of Diabetes (WJD, World J Diabetes) is to provide scholars and readers from various fields of diabetes with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WID mainly publishes articles reporting research results and findings obtained in the field of diabetes and covering a wide range of topics including risk factors for diabetes, diabetes complications, experimental diabetes mellitus, type 1 diabetes mellitus, type 2 diabetes mellitus, gestational diabetes, diabetic angiopathies, diabetic cardiomyopathies, diabetic coma, diabetic ketoacidosis, diabetic nephropathies, diabetic neuropathies, Donohue syndrome, fetal macrosomia, and prediabetic state.

INDEXING/ABSTRACTING

The WID is now abstracted and indexed in Science Citation Index Expanded (SCIE, also known as SciSearch®), Current Contents/Clinical Medicine, Journal Citation Reports/Science Edition, PubMed, PubMed Central, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 Edition of Journal Citation Reports® cites the 2021 impact factor (IF) for WJD as 4.560; IF without journal self cites: 4.450; 5-year IF: 5.370; Journal Citation Indicator: 0.62; Ranking: 62 among 146 journals in endocrinology and metabolism; and Quartile category: Q2.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yu-Xi Chen; Production Department Director: Xu Guo; Editorial Office Director: Jia-Ping Yan.

NAME OF JOURNAL

World Journal of Diabetes

ISSN

ISSN 1948-9358 (online)

LAUNCH DATE

June 15, 2010

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Michael Horowitz, Md. Shahidul Islam, Lu Cai, Jian-Bo Xiao

EDITORIAL BOARD MEMBERS

https://www.wignet.com/1948-9358/editorialboard.htm

PUBLICATION DATE

September 15, 2022

COPYRIGHT

© 2022 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

https://www.wjgnet.com/bpg/gerinfo/204

GUIDELINES FOR ETHICS DOCUMENTS

https://www.wjgnet.com/bpg/GerInfo/287

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

https://www.wjgnet.com/bpg/gerinfo/240

PUBLICATION ETHICS

https://www.wjgnet.com/bpg/GerInfo/288

PUBLICATION MISCONDUCT

https://www.wjgnet.com/bpg/gerinfo/208

ARTICLE PROCESSING CHARGE

https://www.wjgnet.com/bpg/gerinfo/242

STEPS FOR SUBMITTING MANUSCRIPTS

https://www.wjgnet.com/bpg/GerInfo/239

ONLINE SUBMISSION

https://www.f6publishing.com

© 2022 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com





Submit a Manuscript: https://www.f6publishing.com

World J Diabetes 2022 September 15; 13(9): 752-764

DOI: 10.4239/wjd.v13.i9.752 ISSN 1948-9358 (online)

ORIGINAL ARTICLE

Basic Study

Hyperglycemia and reduced adiposity of streptozotocin-induced diabetic mice are not alleviated by oral benzylamine supplementation

Christian Carpéné, Kristiyan Stiliyanov Atanasov, Francisco Les, Josep Mercader Barcelo

Specialty type: Endocrinology and metabolism

Provenance and peer review:

Invited article; Externally peer reviewed

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C, C, C, C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Balbaa ME, Egypt; Cheng JT, Taiwan; Zan-Chao L, China

Received: March 11, 2022

Peer-review started: March 11, 2022 First decision: April 25, 2022 Revised: May 13, 2022 Accepted: August 22, 2022 Article in press: August 22, 2022 Published online: September 15,

2022



Christian Carpéné, Institut des Maladies Métaboliques et Cardiovasculaires, INSERM UMR 1297, Toulouse 31432, France

Kristiyan Stiliyanov Atanasov, Josep Mercader Barcelo, Molecular Biology and One Health research group, Department of Fundamental Biology and Health Sciences, University of the Balearic Islands, Palma 07122, Spain

Francisco Les, Department of Pharmacy, Faculty of Health Sciences, Universidad San Jorge, Villanueva de Gállego, Zaragoza 50830, Spain

Corresponding author: Christian Carpéné, PhD, Senior Researcher, Institut des Maladies Métaboliques et Cardiovasculaires, INSERM UMR 1297, Université Paul Sabatier Toulouse III, CHU Rangueil, Bat L4, BP 84225, Toulouse 31432, France. christian.carpene@inserm.fr

Abstract

BACKGROUND

Benzylamine (Bza) oral administration delays the onset of hyperglycemia in insulin-resistant *db*^{-/-} mice; a genetic model of obesity and type 2 diabetes.

To extend the antihyperglycemic properties of oral benzylamine to a model of insulin-deficient type 1 diabetes.

METHODS

Male Swiss mice were rendered diabetic by streptozotocin treatment (STZ) and divided in two groups: one received 0.5% Bza as drinking solution for 24 d (STZ Bza-drinking) while the other was drinking water ad libitum. Similar groups were constituted in age-matched, nondiabetic mice. Food intake, liquid intake, body weight gain and nonfasting blood glucose levels were followed during treatment. At the end of treatment, fasted glycemia, liver and white adipose tissue (WAT) mass were measured, while glucose uptake assays were performed in adipocytes.

RESULTS

STZ diabetic mice presented typical features of insulin-deficient diabetes: reduced body mass and increased blood glucose levels. These altered parameters were not normalized in the Bza-drinking group in spite of restored food and water intake.

Bza consumption could not reverse the severe fat depot atrophy of STZ diabetic mice. In the nondiabetic mice, no difference was found between control and Bza-drinking mice for any parameter. In isolated adipocytes, hexose uptake was partially activated by 0.1 mmol/L Bza in a manner that was obliterated in vitro by the amine oxidase inhibitor phenelzine and that remained unchanged after Bza supplementation. Oxidation of 0.1 mmol/L Bza in WAT was lower in STZ diabetic than in normoglycemic mice.

CONCLUSION

Bza supplementation could not normalize the altered glucose handling of STZ diabetic mice with severe WAT atrophy. Consequently, its antidiabetic potential in obese and diabetic rodents does not apply to lipoatrophic type 1 diabetic mice.

Key Words: Diabetes; Adipocytes; Amine oxidases; Insulin-like agents; Glucose transport; Polydipsia

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: In adipocytes, benzylamine (Bza) is oxidized by amine oxidases and stimulates glucose uptake. Bza oral administration alleviates insulin-resistant diabetes in obese and diabetic mice. It was investigated whether Bza was also antihyperglycemic in insulin-deficient type 1 diabetes. To this aim, a 0.5% Bza drinking solution was given to streptozotocin-induced diabetic mice. Oral Bza did not recover hyperglycemia and reduced adiposity of lipoatrophic and diabetic mice. A minimal level of adiposity was required to support benzylamine oxidation and to improve glucose utilization. Thus, the antidiabetic properties of Bza in obese and diabetic models, do not apply for diabetes with severe lipoatrophy.

Citation: Carpéné C, Stiliyanov Atanasov K, Les F, Mercader Barcelo J. Hyperglycemia and reduced adiposity of streptozotocin-induced diabetic mice are not alleviated by oral benzylamine supplementation. World J Diabetes 2022; 13(9): 752-764

URL: https://www.wjgnet.com/1948-9358/full/v13/i9/752.htm

DOI: https://dx.doi.org/10.4239/wjd.v13.i9.752

INTRODUCTION

A recent study indicates that orally given benzylamine (Bza) delays the onset of diabetes in obese and insulin-resistant db'-mice[1]. Supplementation with 0.5% Bza (5 g/L) in the drinking water impaired the increase in blood glucose, water intake and urine emission that occurs after weaning in this mouse model of insulin-resistant type 2 diabetes. The proposed mechanism of action for ingested Bza, which is naturally present in vegetables and edible plants, relies on its oxidation by an amine oxidase, which is a copper-containing enzyme highly expressed in fat cells[2,3]: The semicarbazide-sensitive amine oxidase (SSAO)[4] also known as amine oxidase copper containing 3[5] and identical to vascular adhesion protein (VAP-1)[6]. More precisely, it is hydrogen peroxide, one of the products of amine oxidation, and known from decades to stimulate glucose uptake in fat cells[7], that supports the insulin-mimetic actions of Bza in adipocytes, either in rodents[8] or in humans[9]. The *in vitro* insulin-like actions of Bza encompass activation of glucose uptake[10], induction of adipogenesis[11], stimulation of lipogenesis [12], and inhibition of lipolysis. They occur even in the absence of insulin[8]. It was therefore of interest to investigate whether an oral treatment with Bza is capable of alleviating the impaired glucose handling of insulin-deficient, type 1 diabetic states.

Type 1 diabetes is characterized by a deficiency in insulin resulting from endocrine pancreas injury. To treat this disease, it is necessary to permanently normalize the altered blood glucose homeostasis. Since insulin is the major regulator of blood glucose levels, many therapeutic beneficial approaches have consisted in providing this pancreatic hormone, via repeated injections, or even by more sophisticated administration modes using biotechnologies, islet transplants or cell therapies[13]. Whatever the mode of supply, insulin overdose has to be avoided to prevent the risk of fatal hypoglycemia and to limit the onset of insulin resistance. Of note, various pharmacological agents or naturally occurring molecules can act as insulin-like factors on the glucose utilization by peripheral tissues[14]. In this view, testing the putative antihyperglycemic effect of Bza in type 1 diabetic rodents remains a preclinical step that deserves descriptive studies.

Alongside its capacity to oxidize Bza[1], fat tissue is not quantitatively but qualitatively of paramount importance in the regulation of glucose disposal. Adipose tissue uses glucose for accumulating lipid stores, and it also acts as an endocrine organ secreting a variety of adipokines with hyperglycemic or hypoglycemic properties, even in the absence of exogenous insulin[15]. The lack of adipose tissue

(lipoatrophy), such as that obtained in several genetically modified mice, is accompanied with altered glucose homeostasis [16,17]. Similarly, diabetic type 1 models, such as streptozotocin (STZ) diabetic rodents, with destroyed endocrine pancreas, exhibit reduced fat stores[18,19]. In humans, successful treatment of type 1 diabetes is concomitant with both restoration of normal glucose levels and adipose tissue recovery[20].

More importantly, diabetic phenotypes of diverse animal models have been ameliorated when white adipose tissue (WAT) or brown adipose tissue (BAT) has been reintroduced in these models, irrespective of the method used. Nowadays, it is suggested that adipose tissue contributes to the correction of type 1 diabetes, since hyperglycemia was lowered in diabetic mice treated by conditioned media from adipose-derived stem cells[21], and since mitigation of diabetes was observed in STZ diabetic mice receiving BAT transplantation[19]. To date, the beneficial effects of ingested Bza on glucose and lipid handling have been studied in obese rodents only [1,22]. These studies have suggested that enhanced fat deposition contribute to the insulin-like effects observed in vivo. Again, these considerations reinforced our interest in investigating the effects of Bza in a lipoatrophic model of type 1

The capacity of Bza to activate glucose transport in rat or mouse adipocytes is potentiated by the presence of vanadium[10,23], a widely recognized insulin-like agent[24,25]. Accordingly, it has been already demonstrated that in vivo treatments with a combination of amine oxidase substrates and vanadium exert antidiabetic effects in diverse diabetic rodents, including the STZ diabetic rats[10,26]. However, we demonstrated in recent studies that Bza[9] or catecholamines[27] are capable of activating glucose transport in human adipocytes, even in the absence of vanadium, and that the synergism vanadate/amine is much more weak in human adipocytes than in the murine ones. All these observations prompted us to examine for the first time the influence of prolonged oral administration of Bza alone-without any added vanadate - in a model of type 1 diabetes, which is nonobese and insulindeficient; the STZ-induced diabetic mouse.

We investigated whether Bza alone was able, via oral consumption, to improve glucose handling in insulin-deficient STZ mice. The following results do not confirm our assumption, although they suggest that Bza action on glucose disposal requires a minimal amount of adipocytes prone to increase their glucose consumption when oxidizing this SSAO substrate.

MATERIALS AND METHODS

Chemicals

Benzylamine hydrochloride, STZ, bovine insulin, phenelzine, collagenase A, and most of the other reagents were from Sigma-Aldrich-Merck (Saint Quentin Fallavier, France). [3H]-2-Deoxyglucose (2-DG) was from Perkin Elmer (Boston, MA, USA). The glucometers and consumables for follow-up of fed blood glucose were provided by Pr. Valet P. (Univ Toulouse, France), and used as previously described [28].

Insulin-deficient type 1 diabetic mice

Male Swiss mice obtained from Charles River Laboratories (L'arbresle, France) were housed at constant temperature (20-22°C) and with a 12-h light-dark cycle. At the age of 2 mo, they received an intraperitoneal injection of STZ (40 mg/kg) diluted in citrate buffer (0.05 mmol/L, pH 4.5) for four consecutive days, as described previously [21]. A week later, mice receiving only citrate buffer (nondiabetic) and treated mice exhibiting blood glucose ≥ 300 mg/100 mL (STZ diabetic) were subdivided into four groups of eight males, with either free access to water (control) or a 0.5% Bza solution as drinking liquid (Bza-drinking) for 24 d. To measure plasma insulin levels at the beginning of treatment, blood samples were withdrawn from tail vein then centrifuged and analyzed using Ultrasensitive insulin-ELISA kit (Mercodia, Uppsala, Sweden). All the mice had free access to food and water and were treated in accordance with the ARRIVE guidelines (Animal Research: Reporting of In Vivo Experiments)[29]. During this period, nonfasting blood glucose levels were determined every 3 d at 12:00 h (equivalent in the used circadian rhythm to 4 h after lights turned on) using an Accu-Check glucometer (Roche Diagnostics) on a blood drop withdrawn from the tail vein. Mice were killed after overnight fasting at the end of treatment and organs were collected and weighed.

Adipocyte preparations

Adipocyte preparations were obtained by collagenase digestion of WAT immediately after removal from the epididymal, intra-abdominal and inguinal anatomical locations. WAT was cut into small pieces, digested at 37°C by collagenase under agitation in Krebs-Ringer buffered at pH 7.5 with 15 mmol/L sodium bicarbonate, 10 mmol/L HEPES, supplemented with 3.5% of bovine serum albumin, as previously described[1]. Preparations of buoyant adipocytes were isolated from the digested WAT by filtration through nylon stockings and two gentle buffer washes, as described previously [10]. In our digestion process, approximately 1 g WAT was necessary to obtain sufficient functional adipocytes for the subsequent hexose uptake assays. When total amount of dissected WAT exceeded 1 g, excess samples were snap-frozen at -80°C. This occurred for each of the normoglycemic mice but not for the lipoatrophic STZ-treated mice. In this case, pools of two mice were used to freeze approximately 200 mg WAT.

Glucose transport assays

The nonmetabolizable analog [3H]-2-DG was the only source of hexose for the cell preparations during glucose transport assays. It was added at a final concentration of 0.1 mmol/L after 45 min incubation of the fat cell suspension with the tested agents, as previously described [10]. Pyruvate (2 mmol/L) was also present in the medium throughout the experiments for energy supply. Radioactive 2-DG (100 µL; approximately 1300000 dpm/vial) was added to 400 µL fat cell suspension, and hexose uptake assays were stopped 10 min later with 100 μL 100 μmol/L cytochalasin B. Cell suspensions (200 μL) were immediately transferred to plastic centrifugation microtubes prefilled with dinonyl-phthalate (density 0.98 g/mL), then subjected to a 30 s spin. The upper part of the tubes, containing radiolabelled hexose internalized in intact fat cells floating above the silicon layer was counted in scintillation vials, as described previously[10]. The extracellular [3H]-2-DG present in the upper part of the tubes was determined in tubes receiving cytochalasin B prior to 2-DG. It averaged 1%-5% of the radioactivity found in control uptake, and was subtracted from all assays, as described previously [9].

Determination of benzylamine oxidation

Amine oxidase activity was determined at 37°C using [14C]-Bza as substrate, in homogenates of thawed WAT samples, as previously described[10]. Isotopic dilution of [14C]-Bza (final concentration: 0.1 mmol/L) was incubated for 30 min in 200 µL 200 mmol/L phosphate buffer with approximately 50 µg proteins, then the radiolabeled oxidation products were immediately extracted in toluene/ethyl acetate and counted as previously specified[9]. Results were expressed as nmol of deamination products/mg protein/min.

Statistical analysis

Results are presented as means ± SEM of (n) observations. All the statistical analyses for comparisons between parameters used ANOVA followed by post hoc Dunnett's multiple comparisons test, analyzed with Prism 6 for Mac OS X (GraphPad Software). Relative EC₅₀ values were calculated by nonlinear regression.

RESULTS

Bza supplementation normalizes increased food and water consumption of STZ-induced diabetic mice without restoring body weight gain

At the start of the experiment, the STZ-induced diabetic mice exhibited lower body weight when compared to age-matched normoglycemic mice (Figure 1). The body weight gain of the insulin-deficient mice was also limited during the treatment period and was not corrected by Bza supplementation. At the end of the experiment, the mean body weight of STZ mice remained lower than that of normoglycemic mice. Hence, Bza supplementation tended to limit body weight gain in both groups, but this trend did not reach significance (Figure 1A). No significant decrease in food consumption was found in the Bza-drinking normoglycemic mice. By contrast, the hyperphagic status of the STZ mice was alleviated by Bza supplementation (Figure 1B). A similar influence of Bza supplementation was found for water consumption. An almost normalization of the elevated daily water intake of STZ diabetic mice occurred in the group subjected to Bza drinking (Figure 1B).

Figure 1 also shows that the characteristic polydipsic feature that occurs in STZ-induced type 1 diabetes was of greater magnitude than the hyperphagy triggered by the noxious diabetogenic agent. The exaggerated liquid consumption of the diabetic group was increased by 5.7 times when compared to normoglycemic control while this increase only reached 1.7 times for food intake. The former defect was expected to traduce glycosuria[19,30], while the second likely corresponded to a lowered efficiency of the ingested carbohydrates that accompanies insulin deficiency[31].

In view of these alterations of food and water intake in STZ diabetic mice and their recovery after Bza drinking, the influence of Bza supplementation on blood glucose levels was examined in both fed and fasted conditions.

Influence of oral supplementation of Bza on blood glucose in nondiabetic and diabetic mice

Figure 2 shows the pattern of nonfasting glycemia during the treatment period for the four experimental groups. The unfasted blood glucose levels of the mice previously challenged with STZ were at least twice higher than those of the controls throughout the study (Figure 2A). Such strong hyperglycemia was mainly a consequence of the low circulating levels of insulin found at the start of treatment in the two groups of STZ diabetic mice (0.40 ± 0.04 and 0.38 ± 0.05 ng/mL) when compared to the nondiabetic mice (1.26 \pm 0.14 and 1.35 \pm 0.09 ng/mL, n = 8; P < 0.001). In the STZ diabetic mice, the blood glucose

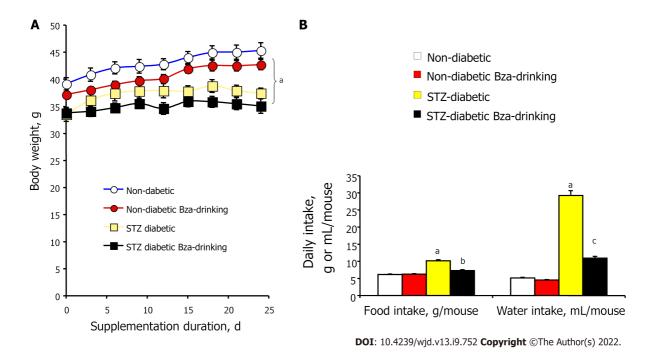


Figure 1 Influence of Bza supplementation on body weight gain, food intake and water consumption in normoglycemic and STZ-induced diabetic mice. A: Body weight of diabetic (squares) and nondiabetic mice (circles) drinking water (open symbols) or 0.5% Bza (Bza-drinking, closed symbols). Mean ± SEM of n = 8 males in each group. Significant difference at: ^aP < 0.001 between diabetic and nondiabetic mice, irrespective of the treatment; B: Average daily food intake and water intake. The mean daily consumption calculated throughout the treatment is expressed as g or mL/mouse, for each of the following groups: nondiabetic (white columns), nondiabetic Bza-drinking (red columns), STZ diabetic (yellow columns), STZ diabetic Bza-drinking (black columns). Each column is the mean ± SEM of at least 16 determinations. Different from nondiabetic rats at: ^aP < 0.01. Different from respective control significant at: ^bP < 0.01; ^cP < 0.001. STZ: Streptozotocin; Bza: Benzylamine.

levels remained elevated in both Bza-drinking and water-drinking groups (Figure 2A). In the normoglycemic mice, the nonfasting blood glucose was superimposed in the control and Bza-drinking groups and remained below 200 mg/100 mL. Thus, blood glucose levels were not significantly influenced by repeated Bza consumption.

To avoid any alteration in body weight gain and in glucose handling, the mice were subjected to overnight fasting only once, at the end of experiment. Fasting blood levels were expectedly lower than nonfasting blood glucose (Figure 2B). Again, the fasting values were superimposable in Bza-drinking mice and their respective controls, while the fasting blood glucose of STZ diabetic mice was higher than that in nondiabetic groups (Figure 2B). Thus, Bza supplementation did not exhibit any hypoglycemic or antihyperglycemic action in this animal model of severe type 1 diabetes.

These findings contrasted with the capacity of Bza to delay the onset of diabetes in the genetically obese and diabetic db'-mice[1]. Given the unexpected lack of efficiency of Bza consumption on glucose handling, it was poorly appropriate to delineate its putative mechanisms of action or to further examine other surrogate makers of diabetic state, as reported previously[1]. Instead, we verified whether the dose of Bza ingested was similar in the two diabetic models. Considering the daily liquid intake and the body mass of the STZ mice, it was calculated that these type 1 diabetic mice ingested 10850 ± 598 µmol/kg bw/d Bza throughout the treatment. This dose was similar to that used for Bza supplementation in young type 2 diabetic db^{-/-} mice[1], which ranged between 9300 and 10 100 µmol/kg bw/d. However, another difference between type 2 (insulin-resistant) and type 1 (insulin-deficient) diabetic mouse models lies in the occurrence of excessive fat depots in the former and a clearly emaciated state in the latter. Therefore, attention was focused on WAT in the STZ mice and their controls.

Comparison of fat stores between normoglycemic and STZ-induced diabetic mice

Smaller mass of subcutaneous and visceral WAT was a typical feature of STZ-induced diabetic mice when compared to normoglycemic controls (Figure 3). In the STZ diabetic mice, the low mass of fat pads was not modified by Bza drinking, whatever their anatomical location. Similarly, the normal adiposity of the nondiabetic mice was not modified after oral Bza supplementation.

When the mass of the dissected fat depots was normalized as percentage of body weight, such adiposomatic index[22] was significantly lower in diabetic than in nondiabetic mice (1.2 \pm 0.4% vs 3.7 \pm 0.6%, P < 0.001). Again, Bza supplementation did not modify adiposomatic index: 1.3 \pm 0.4% and 3.9 \pm 0.4%, in Bza-drinking diabetic and nondiabetic groups, respectively.

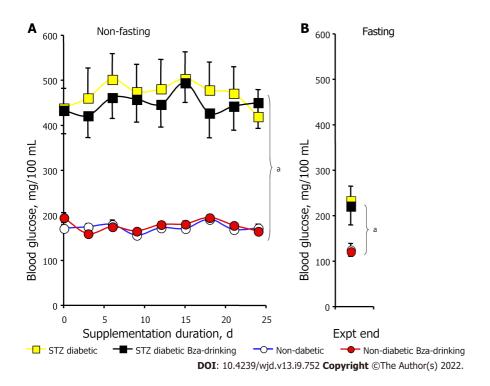


Figure 2 Non-fasting and fasting blood glucose in normoglycemic and STZ-induced diabetic mice during Bza supplementation. A: Blood glucose measured every three days at 12:00 in diabetic (squares) and nondiabetic mice (circles) drinking water (open symbols) or Bza 0.5% (Bza-drinking, closed symbols) for 24 d; B: Overnight fasted blood glucose levels at the end of experiment for the same groups of mice. Mean ± SEM of n = 8 males in each group. Different from nondiabetic mice at: ^aP < 0.001. No significant difference was found between Bza drinking and respective controls. STZ: Streptozotocin; Bza: Benzylamine; Expt end: The end of experiment.

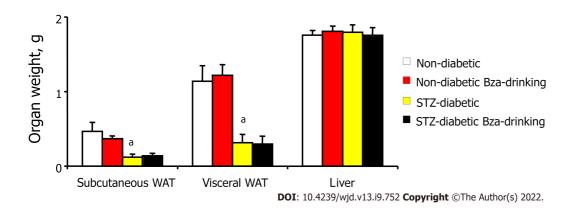


Figure 3 Lack of influence of Bza supplementation on organ weight in normoglycemic and STZ-induced diabetic mice. Mean ± SEM of the wet weight of subcutaneous or visceral white adipose tissues, and of liver for eight males in each group. Different from nondiabetic mice at: ^aP < 0.001. No significant difference was found between Bza-drinking and respective controls. STZ: Streptozotocin; Bza: Benzylamine; WAT: White adipose tissues.

757

In contrast, the weight of the liver was identical in the four experimental groups (Figure 3). However, when liver mass was expressed as ratio to body weight, the difference that appeared between diabetic and nondiabetic animals was opposite to that of the adiposomatic index. The liver represented 5.3 ± 0.2% of body mass in both STZ diabetic and STZ diabetic Bza-drinking mice (NS, n = 8). This proportion was smaller in nondiabetic mice (4.2 \pm 0.1%, P < 0.001), even after Bza drinking (4.5 \pm 0.2%).

These observations indicated that the STZ-induced diabetic mice did not normalize their reduced fat deposition and body weight gain after Bza supplementation, in spite of partial recovery of their altered food intake. Moreover, Bza supplementation was not efficient in normalizing the altered blood glucose control or relative hepatomegaly of the STZ mice, although limiting polydipsia. We have previously proposed that Bza oxidation occurring in the hypertrophied WAT of obese and diabetic $db^{-/-}$ mice supports its insulin-like in vitro effects by facilitating glucose utilization in adipocytes and contributes to its antihyperglycemic action[1]. Therefore, such in vitro effects were examined.

Effects of insulin and Bza on glucose transport in mouse adipocytes

Unfortunately, the WAT atrophy of the STZ diabetic mice did not allow the preparation of sufficient biological material for exploring the activation of 2-DG uptake in functional adipocytes from diabetic and Bza-drinking diabetic mice. There was only a pool of around 400 mg of WAT dissected from different anatomical locations in each STZ mouse, while 1-2 g was removed from each nondiabetic mouse. Consequently, sufficient adipocytes could be isolated from the latter samples only, and the subsequent hexose uptake assays were performed with adipocyte preparations that contained 18.0 ± 2.8 and 19.0 ± 2.5 mg lipid/400 µL in normoglycemic Bza-drinking and control mice, respectively. Thus, Figure 4A shows insulin stimulation of 2-DG uptake in nondiabetic mice only. As expected, insulin dose-dependently activated hexose uptake in adipocytes from control mice, and a tendency to improve insulin maximal effect was detected in Bza-drinking mice. EC₅₀ values of insulin were 0.4 and 2.3 nmol/L for Bza-drinking and control mice, respectively, without showing a significant difference between them. Figure 4B indicates that 0.1 mmol/L benzylamine was capable of reproducing one-third of the maximal insulin stimulation, in a manner that was blunted by the amine oxidase inhibitor phenelzine, which was inactive on basal or insulin-stimulated hexose uptake. The amine-oxidasedependent insulin-like effect of 0.1 mmol/L Bza was similar in control and Bza-drinking nondiabetic mice. There was no influence of oral Bza supplementation on the capacity of phenelzine to inhibit in *vitro* the insulin-like action of the amine (Figure 4B).

Oxidation of Bza in thawed preparations of adipose tissues

Amine oxidase activity was determined in homogenates from thawed WAT samples by measuring their capacity to oxidize 0.1 mmol/L [14C]-Bza. When expressed as nmol amine oxidized/mg protein/min, the activity was limited in WAT from STZ diabetic mice compared to normoglycemic ones, whether in the control or Bza-drinking groups (Figure 5). The reduced amount of WAT and its limited amine oxidase activity did not argue for a strong contribution of fat stores to the biotransformation of the Bza ingested by STZ diabetic mice.

DISCUSSION

At the first glance, the lack of antihyperglycemic effect of Bza drinking described here in STZ diabetic mice contrasts with its antidiabetic action observed in obese and diabetic db-/- mice[1]. As discussed below, all these findings converge to propose that the difference in Bza-drinking efficiency between the models of type 1 and type 2 diabetes is not related to insulin deficiency versus resistance, but rather to a difference in adiposity between the murine models.

Alongside bearing dramatically larger fat depots than their lean counterparts, the obese and diabetic db^{-/-} mice also possess higher levels of SSAO activity in their fat cells[1,32]. Thus, the antihyperglycemic effect of oral Bza reported for db-/- mice, and not for their lean littermates, could be related to the elevated amine oxidase activity found in the hypertrophied WAT of obese and diabetic animals[1]. In contrast, STZ diabetic rats exhibit lower monoamine oxidase (MAO) and SSAO activities in WAT than their normoglycemic controls[18]. The lack of antihyperglycemic effect of Bza supplementation in STZ mice reported here resembles the weak antidiabetic effect of prolonged administration of tyramine in STZ rats[18]. Tyramine, which is a substrate of both MAO and SSAO, can limit the hyperglycemic responses to a glucose load during a glucose tolerance test but cannot normalize the elevated fasting blood levels of these insulin-deficient rats. Tyramine or Bza can lower the elevated blood glucose of STZ-induced diabetic rats only when combined with vanadium[10,18,33].

Particular attention has been paid to studying the potential antidiabetic effects of amines alone since the synergism between vanadium and biogenic amines on the activation of glucose transport does not work well in human adipocytes [9,27]. Moreover, the potential antidiabetic use of vanadium derivatives is still limited by toxicological aspects. Several observations suggest that the beneficial effects of dietary amines on glucose handling in diabetic rodents (even when not combined with vanadium) rely upon the amount of SSAO present in WAT. The supplementation of drinking water with 0.4% methylamine (another SSAO substrate) has been reported to increase epididymal WAT mass and to improve glucose tolerance in transgenic mice overexpressing a human form of SSAO/VAP-1, while it is inefficient in nontransgenic mice[34]. Oral Bza also improves glucose handling in high-fat diet fed mice, characterized by increased adiposity [22]. Here, we suppose that it is the lipoatrophy of STZ diabetic mice (and not their lack of insulin) that prevented the occurrence of an antihyperglycemic action of Bza.

The sole beneficial effect of Bza drinking seen in the STZ diabetic mice was an almost total recovery of their characteristic hyperphagic and polydipsic behavior [31]. It could be supposed that urinary glucose leak of STZ mice was partially rescued by Bza drinking. Unfortunately, individual metabolic cages were not available for this study and we could not determine daily urine emission or glucosuria. However, water intake reduction occurred without correction of hyperglycemia. This indicated that renal glucose leak, if any, was not sufficiently rescued by Bza drinking to influence the overall glucose homeostasis, while this was the case for db^{-1} mice[1]. Food intake was also reduced in Bza-drinking STZ diabetic mice, but without notable decrease in body weight gain. Thus, food efficiency was increased by Bza drinking.

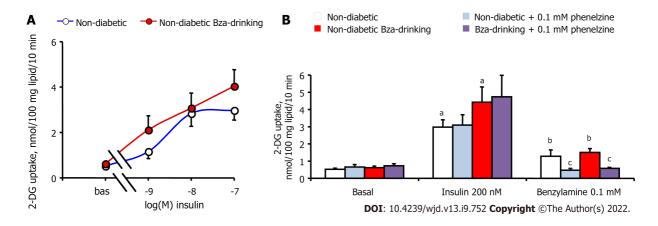


Figure 4 Direct stimulation by insulin and Bza of hexose uptake in adipocytes: lack of influence of Bza supplementation and in vitro inhibition by phenelzine. A: Radiolabeled 2-deoxyglucose (2-DG) uptake was determined in basal condition or in response to increasing doses of insulin in adipocytes from nondiabetic mice of the water-drinking (open circles) or Bza-drinking (red circles) group, while it could not be determined in diabetic mice due to the scarcity of adipocytes isolated from their emaciated fat depots, in both control and Bza-drinking groups. Mean ± SEM of eight adipocyte preparations. B: 2-DG uptake was determined after 45 min incubation without (basal) or with 200 nmol/L insulin and 0.1 mmol/L Bza. The stimulated hexose uptake was significantly different from basal at: ^aP < 0.001; ^bP < 0.01. Phenelzine was added at 0.1 mmol/L in the incubation medium of adipocytes from control nondiabetic mice (blue columns) or from Bza-drinking nondiabetic mice (purple columns). Phenelzine inhibited significantly Bza-induced hexose uptake at: °P < 0.01. STZ: Streptozotocin; Bza: Benzylamine; WAT: White adipose tissues

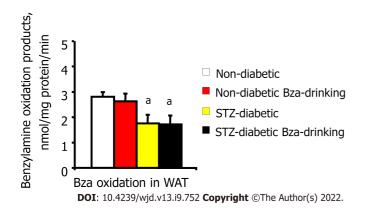


Figure 5 Bza oxidation in adipose tissue of normoglycemic and STZ-induced diabetic mice: lack of influence of dietary Bza consumption. Radiolabeled Bza was present at 0.1 mmol/L during 30 min incubation at 37°C with WAT homogenates from nondiabetic (white columns), nondiabetic Bza-drinking (red columns), STZ diabetic (yellow columns), STZ diabetic Bza-drinking (black columns). Mean ± SEM of eight determinations for nondiabetic mice and four determinations for lipoatrophic STZ diabetic mice. Different from nondiabetic at: ^aP < 0.02. No significant influence of Bza-treatment was detected. STZ: Streptozotocin; Bza: Benzylamine; WAT: White adipose tissues.

However, we cannot propose any underlying mechanism for this effect.

Indeed, it cannot be excluded that mechanisms other than oxidation by amine oxidases might be involved in the in vivo effect of Bza on food and water intake. Raimondi and coworkers have reported that Bza, like methylamine, rapidly induces hypophagia in mice via a modulation of neuronal channels, which is reinforced by SSAO inhibition[35,36]. This suggests that adipose SSAO is likely not the sole target of ingested Bza. Regarding activation of glucose uptake in adipocytes, the effect of Bza is impaired when its oxidation by SSAO is blocked. Surprisingly, the opposite occurred regarding its central effects on food and water intake. When Bza degradation by SSAO is blocked, its half-life is increased and its capacity to modulate the neuronal channels depicted by the group of Raimondi is improved[35,36]. Since there is practically no WAT in the STZ-diabetic mice, and since they have little adipose SSAO, we propose that the limitation of hyperphagia and polydipsia observed in these animals is likely due to a central effect distinct from oxidation by peripheral tissues.

Although the liver is another of the organs reached by ingested Bza, it is not a major site for its biotransformation or detoxification because Bza is metabolized to only a small extent by hepatic subcellular fractions, as observed by Mutlib et al [37]. By contrast, these authors reported that, when orally given to rats, Bza undergoes oxidative deamination and generates benzaldehyde, then hippuric acid, which is the major metabolite. These authors also observed that Bza was fairly stable in rat plasma despite of the presence of a soluble form of SSAO. Although circulating SSAO activity is known to increase with diabetes [18,38-40], it is low when compared to the levels of SSAO found in WAT[1]. A putative mediation of the amine effects via modulation of insulin secretion can be ruled out because, in another model of insulin-deficient diabetes, the alloxan-injected rat, oral administration of tyramine reduced the hyperglycemia by 35%-43% in a manner that was more dependent on insulin-like than on insulin-releasing actions[41].

A limitation of the study was that insulin plasma levels were not determined throughout the treatment since such measurements were performed only at the beginning. However, since circulating insulin was dramatically decreased by STZ challenge, and since the overt hyperglycemia was not corrected by Bza drinking, it was hypothesized that pancreatic injury was not recovered. The hyperinsulinemic levels of the insulin-resistant $db^{-/-}$ mice remained unchanged after Bza supplementation[1]. Similarly, no change in plasma insulin was found in the $db^{+/+}$ lean control after Bza drinking. Nonetheless, it has been reported that methylamine (another SSAO substrate) limits the insulin degradation by adipocytes [42]. If one supposes that increasing the ability of insulin to stimulate glucose transport is one of the mechanisms involved in the antidiabetic effect of Bza, this can explain why Bza was active in insulin-resistant but not in insulin-deficient diabetes models. Such a paradigm of insulinsensitizer capacity might provide an alternative to our interpretations based on the necessary abundance of SSAO and WAT to support peripheral glucose disposal. However, it requires to be demonstrated by further investigations, while we report in the current study that Bza alone activated 2-DG uptake in adipocytes, being therefore able to act as an insulin mimicker even in the absence of insulin.

Whether the in vitro SSAO-mediated insulin-like effect of Bza is solely responsible for the antihyperglycemic effect of Bza drinking is far from being demonstrated here. However, this assumption agrees with the conclusions of independent studies showing that treatment of diabetic rodents with SSAO inhibitors prevents diabetic complications but is not antihyperglycemic at all[43-45]. All these observations bring evidence that adipose cells are predominantly involved in Bza oxidation, as a consequence of their high SSAO expression[3], although they do not rule out other concomitant

We designed the current study to achieve a similar daily amount of Bza ingested by the STZ diabetic mice to that ingested by the obese and type 2 diabetic $db^{-/-}$ mice[1]. The results showed that such an objective was reached. However, similar amine intake did not result in a similar beneficial influence on glucose disposal in the two models. In the STZ diabetic mice, the lipoatrophy and lower richness of WAT in amine oxidase activity gave less probability for an adipocyte-dependent metabolism of the ingested amine and subsequent insulin-like actions. Another apparent weakness of the present study was that the nondiabetic Swiss mice did not ingest the same daily amount of Bza than those subjected to the STZ diabetogenic challenge. Our experiments showed that the polydipsia of the STZ diabetic mice was early rescued, after the first week of Bza supplementation. They also showed that, among the Bzadrinking groups, the accumulated fluid intake of the STZ diabetic mice was about twice that of the normoglycemic mice. It could be easily justified post hoc that, considering the initial polydipsia of diabetic mice, it would have been preferable to double the Bza concentration in the solution given to the Bza-drinking nondiabetic group. Hence, it cannot be excluded that such a high dose of Bza would have reduced liquid consumption in the nondiabetic mice also. By assumption, such an adverse effect on liquid consumption remains unlikely since, as with other organic amines, Bza has a taste varying from almond to fish waste [46], which is not supposed to be repellent for rodents. In reality, achieving exactly the same oral dose of Bza for diabetic and nondiabetic animals would have required weekly pairadjustments, which are difficult to achieve, and would not have yielded more information about the mechanisms of action. The unchanged lipoatrophy, together with the early recovery of polydipsia in the Bza-drinking group, converge to indicate that the antipolydipsic effect of the amine is mediated by a central effect, distinct from that observed in adipocytes.

The in vitro insulin-like effect of submillimolar dose of Bza on glucose transport in adipocytes, and its blockade by phenelzine, reinforced our hypothesis of enhancement of peripheral glucose disposal, although it could not be evidenced in lipoatrophic Bza-drinking STZ mice. Phenelzine, which is a combined MAO and SSAO inhibitor, was used because both MAO and SSAO substrates mimic insulinlike effects in adipocytes[33]. It blocked Bza-stimulated hexose uptake, but not basal or insulinstimulated hexose uptake. No resistance to the selective blockade by phenelzine appeared in the fat cells from Bza-drinking nondiabetic mice, indicating that continuous supplementation with the substrate did not dramatically downregulate the amine oxidase activities. These hexose uptake assays, which could be performed on nondiabetic mice only, confirmed that, even in the absence of insulin, Bza oxidation activates hexose uptake in adipocytes from Swiss white mice as well as in other rodents [43]. According to the literature, the increase of glucose transport by SSAO activation is limited to adipocytes, and only rare reports have extended this hydrogen-peroxide-dependent insulin-like action to other cell types [47]. Unfortunately, the insufficient number of adipocytes isolated from the atrophied WAT of STZ mice hampered the verification of glucose transport responsiveness to insulin and Bza in the type 1 diabetic state. Even if such insulin mimicry also occurred in adipocytes from insulin-deficient mice, it was too limited to modify the glucose handling, when considering the low mass of WAT, as attested by the significantly lower adiposomatic index found in STZ-treated mice. The limited oxidative metabolism of Bza found in WAT of STZ mice was likely unable to contribute to a replenishment of the atrophied fat depots via the increase of glucose utilization demonstrated in adipocytes of the normoglycemic controls. Being poorly biotransformed by the limited fat stores of STZ diabetic mice, the ingested Bza could not increase glucose entry in adipocytes and thereby did not contribute to glucose disposal. We presume that such a lack of Bza action explains how its consumption did not decrease elevated blood glucose. Such inefficiency does not preclude future improvements of the antidiabetic therapeutic applications of other amine substrates. However, our findings limit the relevance of Bza consumption to alleviate the complications of type 1 diabetes, especially when accompanied with lipoatrophy. Nevertheless, Bza and its derivatives remain potential antihyperglycemic agents since a recent integrated network pharmacology analysis has revealed that Bza derivatives contribute to the anti-insulin resistance effects of Moringa oleifera[48], one of the most potent antidiabetic medicinal plants[30,49,50].

CONCLUSION

Although Bza drinking is devoid of beneficial in vivo effects on the type 1 diabetes at doses that limit the onset of type 2 diabetes in genetically obese db^{-/-} mice, the present findings reinforce the hypothesis that oxidation of Bza at the level of adipocytes contributes to peripheral glucose uptake and improves glucose homeostasis. When no sufficient WAT is present (in STZ diabetic mice), the antihyperglycemic effect of Bza is hampered. In contrast, when Bza can be readily oxidized in WAT, it improves glucose tolerance at the expense of an enlargement of fat stores (in $db^{-/-}$ mice). The *in vitro* experiments confirmed the capacity of submillimolar doses of Bza to activate glucose transport in adipocytes. They also show that such SSAO-dependent insulin mimicry is not altered by chronic administration of the substrate.

ARTICLE HIGHLIGHTS

Research background

Oral administration of benzylamine (Bza) exerts antihyperglycemic effects in obese and diabetic rodent models. This effect has been proposed to depend on the insulin-like action of Bza in adipose cells. The amine oxidation catalyzed by amine oxidases abundantly present in adipocytes generates hydrogen peroxide, which activates glucose transport.

Research motivation

To extrapolate the potential antihyperglycemic properties of Bza found in obese and diabetic models to the treatment of insulin-deficient type 1 diabetic states. Bza administration might facilitate glucose utilization to increase lipogenic and adipogenic activities in the adipose tissue and thereby improve glucose disposal even in the absence of insulin.

Research objectives

To evaluate the impact of Bza supplementation on hyperglycemia, polydipsia and hyperphagia in type 1 diabetic mouse, and to demonstrate that Bza metabolism by adipose tissue supports these antidiabetic effects.

Research methods

Bza solution (5 g/L, Bza-drinking) replaced drinking water in streptozotocin (STZ)-induced, insulindeficient diabetic mice. Similar comparison between control and Bza-drinking groups was performed in normoglycemic mice. Nonfasting blood glucose, water and food intake were periodically recorded in the four groups. Adiposity was determined at the end of a 24-d treatment. Glucose transport in freshly isolated adipocytes was assessed ex vivo by determining the uptake of the nonmetabolizable radiolabeled 2-deoxyglucose.

Research results

Chronic Bza intake did not normalize hyperglycemia in STZ diabetic mice, despite it alleviating excessive water and food consumption. Bza intake had no effect on the limited body weight of the STZ diabetic mice and could not restore their dramatically reduced adipose tissue mass. In normoglycemic mice, the Bza-drinking group did not show altered body weight, or food or water consumption. However, when directly given in vitro to adipocytes isolated from nondiabetic mice, Bza was efficient in activating glucose uptake in both control and Bza-drinking groups.

Research conclusions

The capacity of Bza supplementation to reduce hyperglycemia, previously reported in obese and diabetic rodents, was not detectable in the emaciated and insulin-deficient STZ diabetic mice. However, the capacity of Bza to activate glucose transport in adipocytes was confirmed in nonobese, nondiabetic mice. It is likely that the adipose tissue atrophy induced by STZ challenge hampered the lipogenic and adipogenic action of Bza in this severe model of lipoatrophic, insulin-deficient diabetes.

Research perspectives

The current findings and their interpretations considerably limit the field of applications of oral Bza since this molecule did not work as an antidiabetic agent in rodents with reduced adiposity, as it is the case in type 1 STZ diabetic and lipoatrophic mice. Nevertheless, since SSAO substrates exhibit a direct action on glucose handling by fat cells, they still have potential interest for therapeutic use to combat other diabetic states.

ACKNOWLEDGEMENTS

We thank Thomas Cadoudal (I2MC, Toulouse, France) for managing STZ treatment and subsequent hyperglycemic and insulin deficiency screening. The authors also thank Ana Gomez-Ruiz and Sophie Fonvieille (I2MC, Toulouse, France) for help in the follow-up, and for disposable plastic supply, respectively. Special thanks to Philippe Valet and Anne Bouloumié (Univ. Toulouse) for sharing their invaluable knowledge about obese and diabetic rodents.

FOOTNOTES

Author contributions: Carpéné C designed the studies, isolated cells for in vitro experiments, reviewed the literature, designed the figures, wrote and revised the manuscript; Mercader Barceló J performed animal treatments, noninvasive and ex vivo explorations, Stiliyanov Atanasov K was involved in data mining, Les F contributed to statistical analysis, literature review and revised the manuscript.

Institutional review board statement: The study was approved by the I2MC Institutional Review Board: Institut des maladies métaboliques et cadiovasculaires (http://www.i2mc.inserm.fr/accueil).

Institutional animal care and use committee statement: Mice were housed and manipulated according to the INSERM guidelines and European Directive 2010/63/UE by competent and expert technicians or researchers in animal care facilities with agreement number A 31 555 011. The experimental protocol was approved by the local ethical committee CREFRE.

Conflict-of-interest statement: The authors declare no competing financial interests.

Data sharing statement: No additional data are available.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guideline.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: France

ORCID number: Christian Carpéné 0000-0001-7765-6647; Kristiyan Stiliyanov Atanasov 0000-0003-3202-9716; Francisco Les 0000-0002-7972-7119; Josep Mercader Barcelo 0000--0002-5293-2809.

 $\textbf{S-Editor:} \ Chang \ KL$ L-Editor: Kerr C P-Editor: Chang KL

REFERENCES

- Iffiú-Soltesz Z, Wanecq E, Tóthfalusi L, Szökő É, Carpéné C. Oral Supplementation with Benzylamine Delays the Onset of Diabetes in Obese and Diabetic db-/- Mice. Nutrients 2021; 13 [PMID: 34444782 DOI: 10.3390/nu13082622]
- Moldes M, Fève B, Pairault J. Molecular cloning of a major mRNA species in murine 3T3 adipocyte lineage. differentiation-dependent expression, regulation, and identification as semicarbazide-sensitive amine oxidase. J Biol Chem 1999; **274**: 9515-9523 [PMID: 10092636 DOI: 10.1074/jbc.274.14.9515]



- 3 Shen SH, Wertz DL, Klinman JP. Implication for functions of the ectopic adipocyte copper amine oxidase (AOC3) from purified enzyme and cell-based kinetic studies. PLoS One 2012; 7: e29270 [PMID: 22238597 DOI: 10.1371/journal.pone.0029270]
- 4 Olivieri A, Tipton KF, O'Sullivan J. Characterization of the in vitro binding and inhibition kinetics of primary amine oxidase/vascular adhesion protein-1 by glucosamine. Biochim Biophys Acta 2012; 1820: 482-487 [PMID: 22202180 DOI: 10.1016/j.bbagen.2011.12.009]
- Romauch M. Zinc-α2-glycoprotein as an inhibitor of amine oxidase copper-containing 3. Open Biol 2020; 10: 190035 [PMID: 32315567 DOI: 10.1098/rsob.190035]
- Salmi M, Jalkanen S. Vascular Adhesion Protein-1: A Cell Surface Amine Oxidase in Translation. Antioxid Redox Signal 2019; **30**: 314-332 [PMID: 29065711 DOI: 10.1089/ars.2017.7418]
- Czech MP. Differential effects of sulfhydryl reagents on activation and deactivation of the fat cell hexose transport system. *J Biol Chem* 1976; **251**: 1164-1170 [PMID: 1249070]
- Fontana E, Boucher J, Marti L, Lizcano JM, Testar X, Zorzano A, Carpéné C. Amine oxidase substrates mimic several of the insulin effects on adipocyte differentiation in 3T3 F442A cells. Biochem J 2001; 356: 769-777 [PMID: 11389684 DOI: 10.1042/0264-6021:3560769]
- Morin N, Lizcano JM, Fontana E, Marti L, Smih F, Rouet P, Prévot D, Zorzano A, Unzeta M, Carpéné C. Semicarbazidesensitive amine oxidase substrates stimulate glucose transport and inhibit lipolysis in human adipocytes. J Pharmacol Exp Ther 2001; 297: 563-572 [PMID: 11303044]
- Marti L, Abella A, Carpéné C, Palacín M, Testar X, Zorzano A. Combined treatment with benzylamine and low dosages of vanadate enhances glucose tolerance and reduces hyperglycemia in streptozotocin-induced diabetic rats. Diabetes 2001; 50: 2061-2068 [PMID: 11522672 DOI: 10.2337/diabetes.50.9.2061]
- Mercier N, Moldes M, El Hadri K, Fève B. Semicarbazide-sensitive amine oxidase activation promotes adipose conversion of 3T3-L1 cells. Biochem J 2001; 358: 335-342 [PMID: 11513731 DOI: 10.1042/0264-6021:3580335]
- 12 Yang H, Ralle M, Wolfgang MJ, Dhawan N, Burkhead JL, Rodriguez S, Kaplan JH, Wong GW, Haughey N, Lutsenko S. Copper-dependent amino oxidase 3 governs selection of metabolic fuels in adipocytes. PLoS Biol 2018; 16: e2006519 [PMID: 30199530 DOI: 10.1371/journal.pbio.2006519]
- Paget MB, Murray HE, Bailey CJ, Downing R. From insulin injections to islet transplantation: An overview of the journey. Diabetes Obes Metab 2022; 24 Suppl 1: 5-16 [PMID: 34431589 DOI: 10.1111/dom.14526]
- Khalivulla SI, Mohammed A, Mallikarjuna K. Novel Phytochemical Constituents and their Potential to Manage Diabetes. Curr Pharm Des 2021; 27: 775-788 [PMID: 33355047 DOI: 10.2174/1381612826666201222154159]
- Fujikawa T, Chuang JC, Sakata I, Ramadori G, Coppari R. Leptin therapy improves insulin-deficient type 1 diabetes by CNS-dependent mechanisms in mice. Proc Natl Acad Sci U S A 2010; 107: 17391-17396 [PMID: 20855609 DOI: 10.1073/pnas.1008025107]
- Savage DB. Mouse models of inherited lipodystrophy. Dis Model Mech 2009; 2: 554-562 [PMID: 19892886 DOI:
- Gao H, Guo Y, Yan Q, Yang W, Li R, Lin S, Bai X, Liu C, Chen D, Cao H, Xiao G. Lipoatrophy and metabolic disturbance in mice with adipose-specific deletion of kindlin-2. JCI Insight 2019; 4 [PMID: 31292295 DOI: 10.1172/jci.insight.128405]
- Visentin V, Bour S, Boucher J, Prévot D, Valet P, Ordener C, Parini A, Carpéné C. Glucose handling in streptozotocininduced diabetic rats is improved by tyramine but not by the amine oxidase inhibitor semicarbazide. Eur J Pharmacol 2005; **522**: 139-146 [PMID: 16202994 DOI: 10.1016/j.ejphar.2005.08.051]
- Gunawardana SC, Piston DW. Reversal of type 1 diabetes in mice by brown adipose tissue transplant. Diabetes 2012; 61: 674-682 [PMID: 22315305 DOI: 10.2337/db11-0510]
- Conway B, Miller RG, Costacou T, Fried L, Kelsey S, Evans RW, Orchard TJ. Adiposity and mortality in type 1 diabetes. Int J Obes (Lond) 2009; **33**: 796-805 [PMID: 19451912 DOI: 10.1038/ijo.2009.75]
- Dias I, Pinheiro D, Ribeiro Silva K, Stumbo AC, Thole A, Cortez E, de Carvalho L, Carvalho SN. Secretome effect of adipose tissue-derived stem cells cultured two-dimensionally and three-dimensionally in mice with streptozocin induced type 1 diabetes. Curr Res Pharmacol Drug Discov 2021; 2: 100069 [PMID: 34988430 DOI: 10.1016/j.crphar.2021.100069]
- Iffiú-Soltész Z, Wanecq E, Lomba A, Portillo MP, Pellati F, Szöko E, Bour S, Woodley J, Milagro FI, Alfredo Martinez J, Valet P, Carpéné C. Chronic benzylamine administration in the drinking water improves glucose tolerance, reduces body weight gain and circulating cholesterol in high-fat diet-fed mice. Pharmacol Res 2010; 61: 355-363 [PMID: 20045461 DOI: 10.1016/j.phrs.2009.12.014]
- Yraola F, García-Vicente S, Marti L, Albericio F, Zorzano A, Royo M. Understanding the mechanism of action of the novel SSAO substrate (C7NH10)6(V10O28).2H2O, a prodrug of peroxovanadate insulin mimetics. Chem Biol Drug Des 2007; **69**: 423-428 [PMID: 17581236 DOI: 10.1111/j.1747-0285.2007.00516.x]
- Shechter Y, Karlish SJ. Insulin-like stimulation of glucose oxidation in rat adipocytes by vanadyl (IV) ions. Nature 1980; **284**: 556-558 [PMID: 6988725 DOI: 10.1038/284556a0]
- Srivastava AK, Mehdi MZ. Insulino-mimetic and anti-diabetic effects of vanadium compounds. Diabet Med 2005; 22: 2-13 [PMID: 15606684 DOI: 10.1111/j.1464-5491.2004.01381.x]
- Zorzano A, Palacín M, Marti L, García-Vicente S. Arylalkylamine vanadium salts as new anti-diabetic compounds. J Inorg Biochem 2009; 103: 559-566 [PMID: 19246098 DOI: 10.1016/j.jinorgbio.2009.01.015]
- Carpéné C, Boulet N, Grolleau JL, Morin N. High doses of catecholamines activate glucose transport in human adipocytes independently from adrenoceptor stimulation or vanadium addition. World J Diabetes 2022; 13: 37-53 [PMID: 35070058 DOI: 10.4239/wid.v13.i1.371
- Rancoule C, Attané C, Grès S, Fournel A, Dusaulcy R, Bertrand C, Vinel C, Tréguer K, Prentki M, Valet P, Saulnier-Blache JS. Lysophosphatidic acid impairs glucose homeostasis and inhibits insulin secretion in high-fat diet obese mice. Diabetologia 2013; **56**: 1394-1402 [PMID: 23508306 DOI: 10.1007/s00125-013-2891-3]
- Kilkenny C, Browne W, Cuthill IC, Emerson M, Altman DG; NC3Rs Reporting Guidelines Working Group. Animal research: reporting in vivo experiments: the ARRIVE guidelines. Br J Pharmacol 2010; 160: 1577-1579 [PMID: 20649561

763



- DOI: 10.1111/j.1476-5381.2010.00872.x]
- Jaiswal D, Kumar Rai P, Kumar A, Mehta S, Watal G. Effect of Moringa oleifera Lam. leaves aqueous extract therapy on hyperglycemic rats. J Ethnopharmacol 2009; 123: 392-396 [PMID: 19501271 DOI: 10.1016/j.jep.2009.03.036]
- Granneman JG, Stricker EM. Food intake and gastric emptying in rats with streptozotocin-induced diabetes. Am J Physiol 1984; **247**: R1054-R1061 [PMID: 6239554 DOI: 10.1152/ajpregu.1984.247.6.R1054]
- Cioni L, De Siena G, Ghelardini C, Sernissi O, Alfarano C, Pirisino R, Raimondi L. Activity and expression of semicarbazide-sensitive benzylamine oxidase in a rodent model of diabetes: interactive effects with methylamine and alphaaminoguanidine. Eur J Pharmacol 2006; 529: 179-187 [PMID: 16325802 DOI: 10.1016/j.ejphar.2005.10.052]
- Marti L, Morin N, Enrique-Tarancon G, Prevot D, Lafontan M, Testar X, Zorzano A, Carpéné C. Tyramine and vanadate synergistically stimulate glucose transport in rat adipocytes by amine oxidase-dependent generation of hydrogen peroxide. J Pharmacol Exp Ther 1998; 285: 342-349 [PMID: 9536030]
- Stolen CM, Madanat R, Marti L, Kari S, Yegutkin GG, Sariola H, Zorzano A, Jalkanen S. Semicarbazide sensitive amine oxidase overexpression has dual consequences: insulin mimicry and diabetes-like complications. FASEB J 2004; 18: 702-704 [PMID: 14977883 DOI: 10.1096/fj.03-0562fje]
- Banchelli G, Ghelardini C, Raimondi L, Galeotti N, Pirisino R. Selective inhibition of amine oxidases differently potentiate the hypophagic effect of benzylamine in mice. Eur J Pharmacol 2001; 413: 91-99 [PMID: 11173067 DOI: 10.1016/s0014-2999(01)00739-7]
- Pirisino R, Ghelardini C, Banchelli G, Galeotti N, Raimondi L. Methylamine and benzylamine induced hypophagia in mice: modulation by semicarbazide-sensitive benzylamine oxidase inhibitors and aODN towards Kv1.1 channels. Br J Pharmacol 2001; 134: 880-886 [PMID: 11606329 DOI: 10.1038/sj.bjp.0704316]
- Mutlib AE, Dickenson P, Chen SY, Espina RJ, Daniels JS, Gan LS. Bioactivation of benzylamine to reactive intermediates in rodents: formation of glutathione, glutamate, and peptide conjugates. Chem Res Toxicol 2002; 15: 1190-1207 [PMID: 12230413 DOI: 10.1021/tx020063q]
- Mészáros Z, Szombathy T, Raimondi L, Karádi I, Romics L, Magyar K. Elevated serum semicarbazide-sensitive amine oxidase activity in non-insulin-dependent diabetes mellitus: correlation with body mass index and serum triglyceride. Metabolism 1999; **48**: 113-117 [PMID: 9920154 DOI: 10.1016/s0026-0495(99)90019-7]
- Salmi M, Stolen C, Jousilahti P, Yegutkin GG, Tapanainen P, Janatuinen T, Knip M, Jalkanen S, Salomaa V. Insulinregulated increase of soluble vascular adhesion protein-1 in diabetes. Am J Pathol 2002; 161: 2255-2262 [PMID: 12466139] DOI: 10.1016/S0002-9440(10)64501-41
- Januszewski AS, Mason N, Karschimkus CS, Rowley KG, Best JD, O'Neal DN, Jenkins AJ. Plasma semicarbazidesensitive amine oxidase activity in type 1 diabetes is related to vascular and renal function but not to glycaemia. Diab Vasc Dis Res 2014; 11: 262-269 [PMID: 24853908 DOI: 10.1177/1479164114532963]
- Lino CdS, Sales TdP, Gomes PB, Amaral JFd, Alexandre FSA, Silveira ER, Ferreira JM, Daniel F. de Sousa, Queiroz MGRd, Sousa FCFd, Brito GAC, Brito SMRC, Viana GSB. Anti-diabetic activity of a fraction from Cissus verticillata and tyramine, its main bioactive constituent, in alloxan-induced diabetic rats. Am J Pharmacol Toxicol 2007; 2: 178-188 [DOI: 10.3844/ajptsp.2007.178.188]
- 42 Kahn CR, Baird KL. Pathways of insulin degradation in isolated adipocytes: evaluation by gel filtration and differential precipitation. Metabolism 1985; 34: 354-363 [PMID: 3884964 DOI: 10.1016/0026-0495(85)90225-2]
- Yu PH, Wang M, Fan H, Deng Y, Gubisne-Haberle D. Involvement of SSAO-mediated deamination in adipose glucose transport and weight gain in obese diabetic KKAy mice. Am J Physiol Endocrinol Metab 2004; 286: E634-E641 [PMID: 14656718 DOI: 10.1152/ajpendo.00272.2003]
- Papukashvili D, Rcheulishvili N, Deng Y. Beneficial Impact of Semicarbazide-Sensitive Amine Oxidase Inhibition on the Potential Cytotoxicity of Creatine Supplementation in Type 2 Diabetes Mellitus. Molecules 2020; 25 [PMID: 32349282 DOI: 10.3390/molecules25092029]
- Tékus V, Horváth ÁI, Csekő K, Szabadfi K, Kovács-Valasek A, Dányádi B, Deres L, Halmosi R, Sághy É, Varga ZV, Adeghate E, Kőszegi T, Mátyus P, Gábriel R, Ferdinandy P, Pintér E, Helyes Z. Protective effects of the novel amineoxidase inhibitor multi-target drug SZV 1287 on streptozotocin-induced beta cell damage and diabetic complications in rats. Biomed Pharmacother 2021; 134: 111105 [PMID: 33338750 DOI: 10.1016/j.biopha.2020.111105]
- Olivieri A, Rico D, Khiari Z, Henehan G, O'Sullivan J, Tipton K. From caffeine to fish waste: amine compounds present in food and drugs and their interactions with primary amine oxidase. J Neural Transm (Vienna) 2011; 118: 1079-1089 [PMID: 21373760 DOI: 10.1007/s00702-011-0611-z]
- 47 El Hadri K, Moldes M, Mercier N, Andreani M, Pairault J, Feve B. Semicarbazide-sensitive amine oxidase in vascular smooth muscle cells: differentiation-dependent expression and role in glucose uptake. Arterioscler Thromb Vasc Biol 2002; 22: 89-94 [PMID: 11788466 DOI: 10.1161/hq0102.101550]
- Huang Q, Liu R, Liu J, Huang Q, Liu S, Jiang Y. Integrated Network Pharmacology Analysis and Experimental Validation to Reveal the Mechanism of Anti-Insulin Resistance Effects of Moringa oleifera Seeds. Drug Des Devel Ther 2020; 14: 4069-4084 [PMID: 33116398 DOI: 10.2147/DDDT.S265198]
- Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. J Ethnopharmacol 2003; 84: 105-108 [PMID: 12499084 DOI: 10.1016/s0378-8741(02)00144-7]
- Gupta R, Mathur M, Bajaj VK, Katariya P, Yadav S, Kamal R, Gupta RS. Evaluation of antidiabetic and antioxidant activity of Moringa oleifera in experimental diabetes. J Diabetes 2012; 4: 164-171 [PMID: 22103446 DOI: 10.1111/j.1753-0407.2011.00173.x]

764



Published by Baishideng Publishing Group Inc

7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: https://www.f6publishing.com/helpdesk

https://www.wjgnet.com

