World Journal of Diabetes

World J Diabetes 2023 March 15; 14(3): 130-351





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INDEXING/ABSTRACTING

The WID is now abstracted and indexed in Science Citation Index Expanded (SCIE, also known as SciSearch®), Current Contents/Clinical Medicine, Journal Citation Reports/Science Edition, PubMed, PubMed Central, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 Edition of Journal Citation Reports® cites the 2021 impact factor (IF) for WJD as 4.560; IF without journal self cites: 4.450; 5-year IF: 5.370; Journal Citation Indicator: 0.62; Ranking: 62 among 146 journals in endocrinology and metabolism; and Quartile category: Q2.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yu-Xi Chen; Production Department Director: Xu Guo; Editorial Office Director: Jia-Ping Yan.

NAME OF JOURNAL

World Journal of Diabetes

ISSN

ISSN 1948-9358 (online)

LAUNCH DATE

June 15, 2010

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Michael Horowitz, Md. Shahidul Islam, Lu Cai

EDITORIAL BOARD MEMBERS

https://www.wignet.com/1948-9358/editorialboard.htm

PUBLICATION DATE

March 15, 2023

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INSTRUCTIONS TO AUTHORS

https://www.wjgnet.com/bpg/gerinfo/204

GUIDELINES FOR ETHICS DOCUMENTS

https://www.wjgnet.com/bpg/GerInfo/287

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

https://www.wjgnet.com/bpg/gerinfo/240

PUBLICATION ETHICS

https://www.wjgnet.com/bpg/GerInfo/288

PUBLICATION MISCONDUCT

https://www.wjgnet.com/bpg/gerinfo/208

ARTICLE PROCESSING CHARGE

https://www.wjgnet.com/bpg/gerinfo/242

STEPS FOR SUBMITTING MANUSCRIPTS

https://www.wjgnet.com/bpg/GerInfo/239

ONLINE SUBMISSION

https://www.f6publishing.com

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World J Diabetes 2023 March 15; 14(3): 130-146

DOI: 10.4239/wjd.v14.i3.130 ISSN 1948-9358 (online)

REVIEW

Pancreatic β-cell dysfunction in type 2 diabetes: Implications of inflammation and oxidative stress

Phiwayinkosi V Dludla, Sihle E Mabhida, Khanyisani Ziqubu, Bongani B Nkambule, Sithandiwe E Mazibuko-Mbeje, Sidney Hanser, Albert Kotze Basson, Carmen Pheiffer, Andre Pascal Kengne

Specialty type: Endocrinology and metabolism

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): D Grade E (Poor): 0

P-Reviewer: Adela R, India; Saisho Y, Japan

Received: October 12, 2022 Peer-review started: October 12,

First decision: November 6, 2022

Revised: November 26, 2022 Accepted: February 27, 2023 Article in press: February 27, 2023 Published online: March 15, 2023



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Abstract

Insulin resistance and pancreatic β-cell dysfunction are major pathological mechanisms implicated in the development and progression of type 2 diabetes (T2D). Beyond the detrimental effects of insulin resistance, inflammation and oxidative stress have emerged as critical features of T2D that define β -cell dysfunction. Predominant markers of inflammation such as C-reactive protein, tumor necrosis factor alpha, and interleukin- 1β are consistently associated with β cell failure in preclinical models and in people with T2D. Similarly, important markers of oxidative stress, such as increased reactive oxygen species and depleted intracellular antioxidants, are consistent with pancreatic β -cell damage in conditions of T2D. Such effects illustrate a pathological relationship between an abnormal inflammatory response and generation of oxidative stress during the

progression of T2D. The current review explores preclinical and clinical research on the pathological implications of inflammation and oxidative stress during the development of β -cell dysfunction in T2D. Moreover, important molecular mechanisms and relevant biomarkers involved in this process are discussed to divulge a pathological link between inflammation and oxidative stress during β -cell failure in T2D. Underpinning the clinical relevance of the review, a systematic analysis of evidence from randomized controlled trials is covered, on the potential therapeutic effects of some commonly used antidiabetic agents in modulating inflammatory makers to improve β -cell function.

Key Words: Type 2 diabetes; Insulin resistance; β-cell dysfunction; Inflammation; Oxidative stress

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Core Tip: Elevated markers of inflammation and oxidative stress are related to β -cell dysfunction, the intracellular defense (antioxidant) mechanisms responsible for ameliorating some of these effects are significantly depleted during type 2 diabetes (T2D). Thus, beyond lowering glucose levels like most antidiabetic drugs, future research should invest in developing therapeutic agents to ameliorate inflammation and oxidative stress to improve blood control in patients with T2D.

Citation: Dludla PV, Mabhida SE, Ziqubu K, Nkambule BB, Mazibuko-Mbeje SE, Hanser S, Basson AK, Pheiffer C, Kengne AP. Pancreatic β-cell dysfunction in type 2 diabetes: Implications of inflammation and oxidative stress. *World J Diabetes* 2023; 14(3): 130-146

URL: https://www.wjgnet.com/1948-9358/full/v14/i3/130.htm

DOI: https://dx.doi.org/10.4239/wjd.v14.i3.130

INTRODUCTION

Type 2 diabetes (T2D) is among the leading causes of death worldwide[1]. Latest global estimates indicate that one in ten adults are currently living with diabetes, of which over 90% of cases are attributed to T2D[2]. Insulin resistance and β -cell dysfunction are considered the major pathophysiological derangements in T2D. Insulin resistance is primarily associated with T2D, however, people with T1D have also been shown to develop insulin resistance mainly because of certain genetic factors or lifestyle modifications[2,3]. Generally, β -cell dysfunction indicates a compromised state of insulin secretion, while insulin resistance refers to the inability of insulin to exert its effects on target organs[3]. In a complex mechanism, insulin resistance and β -cell dysfunction promote elevated blood glucose levels and further drive the pathogenesis of T2D[4]. Changes in β -cell function occur during the early stages of diabetes development (the prediabetic stage) and gradually become worse with disease progression[5,6]. Thus, it has become imperative to delineate the pathological mechanisms driving β -cell dysfunction to alleviate complications linked with T2D, including those that implicate inflammation and oxidative stress.

Inflammation has long been considered the main component of diabetes[7,8]. During T2D, elevated blood glucose levels lead to an undesired inflammatory response, which may be exacerbated by inflammatory intermediaries produced by adipocytes and macrophages in adipose tissue[9]. This process may initiate the low-grade, chronic inflammatory state that induces injury to the pancreatic β -cells, subsequently causing inadequate insulin production, and leading to hyperglycemia[9]. As a result, uncontrolled inflammation has been positioned among the foremost factors in the pathogenesis of T2D [7,8]. Several reviews on the role of inflammation during β -cell dysfunction in T2D have been conducted. For example, Jo and Fang[10] reviewed evidence indicating that malfunctioning of the essential components of the inflammation, including helper T cells, cytotoxic T cells, and regulatory T cells may underpin pancreatic β cell failure in T2D. Sun *et al*[11] recently discussed that aberrant epigenetic signatures, including DNA methylation, chromatin accessibility, histone alteration, and noncoding RNAs orchestrate β -cell malfunction during embryonic growth and postnatal development, thus contributing to β cell dysfunction. These findings further highlight the pathological link between impaired metabolic function and alterations in molecular mechanisms that may lead to β cell dysfunction in T2D[11-13].

Oxidative stress is another factor that is consistently associated with β -cell destruction during the development of T2D[14,15]. Oxidative stress normally arises due to the excessive production of free radicals, especially reactive oxygen species (ROS) that severely affect the neutralizing capacity of intracellular antioxidants[14,15]. Generally, oxidative stress may induce its destructive effects through

causing damage to DNA, proteins, and lipids. In fact, due to the dyslipidemic features of most patients with T2D, uncontrolled oxidative stress is associated with clustering of interconnected plasma lipid and lipoprotein anomalies that may aggravate diabetic complications[16]. Notably, due to the inherent low expression of antioxidant enzymes in pancreatic islets[17], the consequences of oxidative stress can have devastating effects on driving β-cell dysfunction during the development of diabetes[18]. Obesity or excessive fat accumulation within the pancreas are some of the major mechanisms that promote oxidative stress, insulin resistance and β-cell dysfunction in T2D[19,20]. Enhancement of intracellular antioxidants can be targeted to alleviate oxidative stress and improve β-cell function to combat diabetesassociated complications[21,22]. In fact, the pathological relationship between inflammation and oxidative stress can have devastating outcomes leading to the progression of T2D. These complications are distinctively linked with worsening of T2D-related abnormalities, including retinopathy, neuropathy, nephropathy, and damage to tissues[23].

The current review updates and critically discusses literature on the pathological implications of inflammation and oxidative stress during the development of β-cell dysfunction in T2D. Preclinical and clinical research, elucidating the mechanisms that orchestrate the link between inflammation and oxidative stress during the development and progression of T2D are discussed. Firstly, an overview on the link between insulin resistance and β -cell dysfunction is covered to highlight its detrimental effect during the worsening of T2D. Thereafter, different biomarkers of inflammation and oxidative stress are discussed for their relevance in monitoring disease severity. This information also remains important to develop therapeutic targets to alleviate β-cell dysfunction in T2D. To further contribute to the novelty and relevance of the discussed information, a systematic analysis of evidence from randomized controlled trials (RCTs) on the therapeutic effects of antidiabetic agents in modulating inflammatory or oxidative stress to improve β -cell function is also covered.

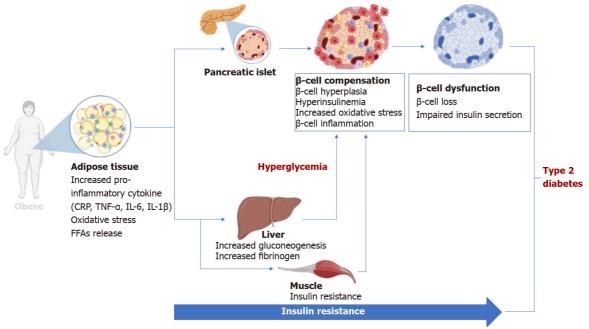
LITERATURE SEARCH AND CLINICAL STUDY SELECTION

Findings from preclinical studies, especially animal models, remain important to accurately decipher or characterize pathological mechanisms implicated in the development of disease. Consistent with the main objective of the current review, this infers describing the potential biological processes and molecular mechanisms involving inflammation and oxidative stress during pancreatic β-cell dysfunction in T2D. However, it also remains important to uncover clinical data on the therapeutic effects of commonly used antidiabetic drugs like metformin in modulating inflammation and oxidative stress to protect against β -cell dysfunction in T2D.

Thus, a systematic search of major electronic engines and databases was done from inception until 18 November 2022 for relevant RCTs. To prioritize clinical relevance of the review, the search was restricted to RCTs reporting on the link between inflammation or oxidative stress and β-cell function in T2D. Medical Subject-Heading and text words such as "inflammation", "oxidative stress", "β-cells", and "type 2 diabetes", including their analogous synonyms and related words were tailored for the individual search engine or database. There was no restriction on the type of antidiabetic drug, with all RCTs reporting on the modulation of these drugs on inflammation or oxidative stress markers in patients with T2D included. The search focused on inflammatory markers such as C-reactive protein (CRP), fibrinogen, interleukins (IL)-6/IL-1 β , and tumor necrosis factor alpha (TNF- α) as well as oxidative stress indicators like ROS, glutathione peroxidase (Gpx), superoxide dismutase (SOD), thioredoxin, and catalase (CAT) that have been linked with β -cell dysfunction in T2D.

INSULIN RESISTANCE AND PANCREATIC B-CELL DYSFUNCTION IN T2D

The pancreatic β-cells have the important function of producing and secreting insulin, a vital hormone that is necessary for the regulation of metabolism. Indeed, insulin is critical for the metabolic regulation of key energy substrates such as carbohydrates, lipids, and proteins. Insulin is required for the absorption of glucose from the blood stream into different cells, including cells from adipose tissue, skeletal muscle, and the liver. Thus, exploring the role of insulin in a broad spectrum of physiological processes including its production and regulation has relevance in understanding the development of T2D[24]. Notably, disturbances in insulin signaling through the inhibition of the insulin receptor substrate protein, phosphoinositide-3-kinase, and protein kinase B (AKT) leads to insulin resistance [25]. The latter is consistently associated with obesity, a major risk factor for T2D. Abnormal adipose tissue expansion causes elevated circulating levels of non-esterified fatty acids, glycerol, markers of oxidative stress, and pro-inflammatory cytokines, subsequently leading to the development of insulin resistance in individuals with obesity [26,27]. In fact, there is a close association between insulin resistance, obesity, and T2D[28]. Insulin resistance, obesity and pancreatic β-cell dysfunction are complex pathological mechanisms implicated in the progression of T2D (Figure 1). Both pathogenic states induce hyperglycemia and therefore increases insulin demand. Subsequently, β-cell dysfunction arises from insufficient glucose sensing to stimulate insulin secretion, hence increased glucose levels persist. This



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Figure 1 An overview of the pathological implications of adipose tissue hypertrophy and insulin resistance during the development and progressive loss of β-cell function in conditions of obesity to type 2 diabetes. Briefly, adipose tissue expansion (usually seen in obesity or type 2 diabetes) is associated with enhanced secretion of pro-inflammatory markers and generation of oxidative stress that directly or indirectly cause pancreatic β-cell loss, leading to impaired insulin secretion. CRP: C-reactive protein; TNF-a: Tumor necrosis factor-alpha; IL: Interleukin; FFAs: Free fatty acids.

process leads to the development of insulin resistance, increased glucose concentrations beyond the physiological state, thereby resulting in the manifestation of hyperglycemia. As a result, β-cells compensate for insulin resistance by hypersecretion of insulin, ultimately leading to β-cell failure[29]. βcell dysfunction follows insulin resistance during the development and progression of T2D. In fact, both pathological states influence each other and likely synergistically worsens diabetes [29]. Maintaining β cell function and insulin signaling in patients with T2D is vital for controlling glucose homeostasis. As such, alleviating the detrimental effects of inflammation has become a critical feature to protect and maintain β -cell function in conditions of T2D[4,30].

INFLAMMATION AND PANCREATIC B-CELL DYSFUNCTION IN T2D

Inflammation is generally classified as a localized response to cellular or tissue injury that is consistent with increased blood flow, leucocyte intrusion, and enhanced production of diverse chemical mediators. This response is necessary to prompt the removal of toxic agents and the restoration of injured tissue. It is now well accepted that chronic inflammation is coupled with insulin resistance and β -cell dysfunction in patients with T2D[4,30]. Adipose tissue expansion is known to play a major role during this process, with inflammation characterized by enhanced levels of macrophages and increased secretion of inflammatory cytokines [31,32]. Briefly, TNF- α , as well as IL-1 β , and IL-6 are considered some of the prominent pro-inflammatory markers in the pathogenesis of T2D[31-33]. Beyond the elevated markers of inflammation, immune dysfunction is also an essential component of inflammation that has been implicated in β-cell failure in T2D.

CRP and β-cell dysfunction in T2D

Epidemiologic studies have reported a close relationship between elevated biomarkers of inflammation and the worsening of T2D and its complications [34,35]. Obesity, which is common in T2D, is wellacknowledged to be the main driver of the pathological consequences inflammation[36]. Obesity is responsible for the initiation of chronic systemic inflammation, with this feature characterized by the activation of the innate immune system in adipose tissue. This outcome prompts the systemic acutephase response which is distinguished by elevation of acute-phase protein levels. For example, CRP is considered one of the predominant markers of systemic inflammation. Indeed, previous research have indicated that elevated levels of high-sensitive CRP (hs-CRP) are strongly associated with advanced βcell dysfunction and insulin resistance in patients with T2D[37,38]. Although considered of hepatic origin, macrophages and T cells are known to be the main activators CRP levels in response to inflammation. Conventionally exploited as an indicator of infection and cardiovascular events[39], accumulating evidence show that CRP plays a vital role in diverse inflammatory mechanisms including the complement pathway, cell death, autophagy, nitric oxide modulation, and the production of cytokines, especially IL-6 and TNF-α[40-42]. Serum concentrations of CRP and other pro-inflammatory cytokines like IL-6 and TNF- α , have been found to be significantly elevated in rats with T2D[43]. Interestingly, such effects occurred concurrently with damage to islets, islet atrophy and β-cell failure in these diabetic animals. Mechanistically, CRP has been reported to promote tissue injury and accelerate apoptosis, through inducing pro-inflammatory mechanisms involving toll-like receptor 4, nuclear factor kappa B (NF-κB), transforming growth factor-β, and the extracellular signal-regulated kinase (ERK) pathway in preclinical models of diabetes [44,45]. Although not directly reporting on β -cell failure, studies in humans have linked enhanced circulatory levels of CRP with systemic inflammation, development and progression of T2D[46,47]. For example, Weber and colleagues showed that worsened low-grade inflammation (through increased levels of hs-CRP) and poor glycemic control were accompanied by reduced β -cell function in patients with T2D[48].

Fibrinogen and β-cell dysfunction in T2D

It has been more than two decades since it was reported that increased plasma levels of fibrinogen are associated with clinical complications of T2D[49]. Fibrinogen is a glycoprotein secreted by the liver and its levels in circulation depict systemic inflammation and tissue injury[50]. The secretion of this protein is crucial for coagulation, revascularization and wound restoration, and this process is mainly facilitated through its enzymatic conversion to fibrin by thrombin. In T2D, fibrinogen levels are positively associated with vascular complications [51-53]. Patients with T2D display elevated levels of fibrinogen, which is linked with poor blood glucose control and increased cardiovascular risk [54,55]. Beyond its role in systemic inflammation[56], fibrinogen levels are increased in rats treated with streptozotocin, a chemical substance known to destroy pancreatic β-cells [57]. Fibrinogen directly promotes profibrogenic and proinflammatory functions in pancreatic stellate cells[58]. These effects are consistent with the activation of pro-inflammatory mechanisms such as activation of NF-κB, three classes of mitogenactivated ERK, c-Jun N-terminal kinase (JNK) and p38 mitogen-activated protein kinase (MAPK) in pancreatic cells. Although clinical evidence directly implicating fibrinogen in β-cell dysfunction is limited, enhanced levels of this protein (hyperfibrinogenemia) has been identified in patients with diabetes[59,60].

IL-6 and IL-1β and β-cell dysfunction in T2D

ILs are known for their diverse biological functions and are of pathological importance during the development of many diseases including T2D. Different interleukin family members have been studied and increasing evidence indicate the significance of these proteins in connecting innate immunity with diverse diseases including inflammatory conditions[41]. Varied IL, including both IL-6 and IL-1β, play a central role in modulating inflammatory responses, especially during the development of metabolic disease[61,62]. Briefly, IL-1β, which is produced by stimulated macrophages is crucial for innate immune regulation (which is considered the first line of defense against invading pathogens). Through its interaction with pattern recognition receptors, the 1β-processing platform is vital for initiating signaling pathways that induce the inflammatory response and regulates adaptive immunity[63]. Notably, IL-6 can induce differentiation of naïve CD4+ T cells, further playing a major role in modulating the acquired immune response[61]. Abnormally regulated levels of both IL-6 and IL-1β are associated with interruption of immunological tolerance and is thus pathologically implicated in the development of autoimmune and chronic inflammatory diseases[62,64-66].

Evidence suggests that IL-6 protects β-cells against oxidative damage through the effective modulation of autophagy and enhancing the antioxidant response. In fact, it was demonstrated that IL-6 couples' autophagy to antioxidant responses leading to the reduction of ROS in β-cells and human islets [67]; whereas β-cell-specific blockage of IL-6 signaling in vivo causes mice to be more vulnerable to oxidative damage and cell death in response to exposure to selective β-cell toxins such as streptozotocin and alloxan[67]. Similarly, others have reported that pretreating β-cells with IL-6 blocks apoptosis induced by pro-inflammatory cytokines, mainly through effective regulation of autophagy [68]. These results are also consistent with reduced IL-6 pathway signaling in islets from donors with T2D[68]. Although such protective effects are noted, others argue that β-cell specific production of IL-6 is consistent with the development of diabetes, downplaying the potential advantage of targeting IL-6 as a therapeutic target for diabetes[69]. However, unlike IL-6, IL-1β has been associated with the inhibition of β-cell function and activation of fatty acid synthase-triggered apoptosis in part by interacting with the transcription factor NF-κB[70]. IL-1β-secreting β cells were identified in pancreatic sections of patients with T2D but not in nondiabetic control subjects[70].

TNF-α and β-cell dysfunction in T2D

A landmark study by Hotamisligil et al[71] was instrumental in demonstrating the close relationship between TNF- α and the progression of diabetes. Their results showed that elevated levels of TNF- α within adipose tissue of rats was consistent with the development of insulin resistance, while the inhibition of this pro-inflammatory cytokine was associated with improved glucose control and insulin sensitivity. TNF-α belongs to a superfamily of type II transmembrane proteins comprising of the TNF homology domain and is acknowledged to play a major role in diverse cellular functions, especially the processes of immune response and inflammation. In various preclinical models of diabetes, including the genetically modified mouse model of T2D (db/db), the pancreatic islet cells display increased levels of chemokines and pro-inflammatory cytokines like TNF-α, when compared to nondiabetic controls [72]. In a Kurdish population, genetic polymorphisms of TNF- α have been linked with genetic predisposition to T2D[73]. Recombinant human TNF-α administration has been shown to acutely reduce basal plasma insulin levels but does not affect glucose-stimulated insulin secretion in patients with T2D[74]. Therapeutic targeted reduction in TNF- α levels has been associated with improved insulin sensitivity in patients with T2D[75]. More importantly, it is evident that TNF-α-activated pathways are responsible for inducing apoptotic cell death in pancreatic β-cells. Stephens et al [76] demonstrated that caspase activation is the prevailing mechanism of TNF-α-induced cell death in NIT-1 cells (an insulin-secreting mouse cell line). Others have shown that TNF- α , in combination with another pro-inflammatory cytokine, interferon-γ (IFN-γ) can induce pancreatic β-cell apoptosis by destructing highly controlled Bcell lymphoma 2 member proteins that are essential for efficient mitochondrial function[77]. Preclinical studies have laid an important foundation to clarify the pathological role of TNF- α in causing β -cell dysfunction in T2D[78], and such information is consistent with insulin resistance and progression of diabetes in clinical settings[79,80]. Although evidence on the detrimental effects of IL-1β, together with other pro-inflammatory markers like IFN- γ and TNF- α in pancreatic β -cell dysfunction is acknowledged [78,81].

T-cells and β-cell dysfunction in T2D

It is acknowledged that both innate and adaptive immune cells play a major role during pancreatic islet inflammation[82]. Innate immune cells produce cytokines that directly and indirectly modulate insulin secretion, whilst also stimulating inflammatory reactions. Macrophages and neutrophils, which physiologically inhabit the pancreatic tissue, can also partake in tissue homeostasis, including harmful activation of immune responses [82,83]. T cells are one of the significant white blood cells of the immune system and are crucial in the adaptive immune response. T-cells consist of two main subtypes, which are CD8+ "killer" and CD4+ "helper" T cells [84]. Regulatory T cells are yet another different subset of T cells that are required to support the mechanism of tolerance, whereby immune cells can recognize and differentiate between parent and invading cells. It is well known that diabetes impairs T cell function [85,86], although the precise mechanisms involved remain to be fully established. In fact, activated Thelper (TH)1 CD4+ T cells and CD8+ cytotoxic T cells have long been implicated in the destruction of pancreatic β-cells in diabetes [87]. Apparently, CD4+ T cells can be activated by IL-12 produced from macrophages and dendritic cells, and this consequence occurs as part of a vicious process involving cytotoxic T cells and recruitment by the pancreatic islets[87]. Notably, CD4+ T cells are instrumental in improving immune responses and their activation can lead to their differentiation into specific subtypes depending on the disease state[84]. Well-known subsets of CD4+ T cells include TH1, which are understood to promote β-cell damage by accelerating apoptosis [88]. Using electron microscopy, it was demonstrated that CD4+ T helper cells exhibit a much higher arrest (a cell jamming process) in the exocrine tissue than islet specific CD8+ T cells in diabetic mice [89]. With the overwhelming evidence supporting the notion that, like autoimmune diabetes, CD4+ (TH1 cell)-mediated inflammation and apoptosis may be the prominent features responsible for β-cell dysfunction and the aggravation T2Dassociated complications [86,88,90,91]. Figure 2 highlights some of the pathophysiological mechanisms implicated in immune and T-cell activation during inflammation-mediated β-cell dysfunction in conditions of T2D.

ANTIDIABETICS AND OTHER AGENTS POTENTIALLY REGULATE INFLAMMATORY MARKERS TO IMPROVE B-CELL FUNCTION

It remains important to decipher the therapeutic mechanisms through which commonly used antidiabetic agents alleviate β-cell insult in conditions of T2D. Table 1 summarizes evidence from RCTs reporting on the effects of antidiabetics and other agents on β-cell function in patients with T2D. Here, the systematic search was focused on establishing the therapeutic link between β -cell function and regulation of circulating levels of prominent inflammatory makers, including hs-CRP, fibrinogen, IL-6, TNF- α , and T-cells. It emerged as early as in 1993 that metformin administration (up to a maximum of 850 mg three times a day for 12 wk) could improve glycemic control and β-cell function but had no effect on plasma fibringen concentrations and platelet function in patients with T2D[92]. Metformin is the most used antidiabetic drug with widely reported pleiotropic effects against complications linked with T2D[93,94]. Interestingly, predominantly included RCTs in Table 1 reported on the therapeutic effects of metformin in controlling basic metabolic profiles together with improving β-cell function and modulating inflammatory markers in patients with T2D. Evidence from different research groups published over the years (2006-2018) has indeed confirmed that metformin administration (from a dose between 1700-2000 mg/d) for a treatment duration of at least 6 mo could improve glycemic control,

Table 1 Effects of antidiabetics and other agents on β-cell dysfunction in type 2 diabetes through the regulation of inflammatory markers

| Ref. | Study population | Intervention | Findings |
|---|---|---|--|
| Nagi and Yudkin[92], 1993 | Patients with T2D (<i>n</i> = 27), with an average age between 48 and 56 yr | Received metformin up to a maximum of 850 mg three times a day, for 12 wk | Improved glycemic control and β -cell function, while ameliorating insulin resistance and risk factors for cardiovascular disease, including plasminogen activator inhibitor-1. But had no effect on plasma fibrinogen concentrations and platelet function |
| Tsunekawa <i>et al</i> [102], 2003 | Patients with T2D (<i>n</i> = 17), with an average age of 67 yr | Received glimepiride started from 1 mg daily and increased up to 6 mg daily for 12 wk | Alleviated insulin resistance by decreasing plasma TNF- $\!\alpha$ levels and reducing those of adiponectin |
| Dominguez <i>et al</i> [103], 2005 | Patients with T2D (<i>n</i> = 10), with an average age of 53 yr | Received etanercept treatment at 25 mg subcutaneously twice weekly for 4 wk | Reduced plasma levels of CRP and interleukin-6 decreased, while also improving β -cell function |
| Pfützner <i>et al</i> [37], 2006 | Patients with T2D (<i>n</i> = 4270), with an average age of 64 yr | Received a combination therapy of peroxisome proliferator activated receptor g agonists and metformin. Disease duration was 5.4 ± 5.6 yr | Increased hs-CRP levels were associated beta-cell dysfunction but showed no correlation with disease duration or glucose control. Patients receiving combination therapy presented the lowest hs-CRP mean values |
| Hamann <i>et al</i> [95], 2008 | Patients with T2D (<i>n</i> = 294), with an average age of 58 yr | Received maximum tolerated doses of rosiglitazone 8 mg plus metformin 2 g/d during the first 12 wk of double-blind treatment for 52 wk | Fixed-dose combination therapy with rosiglitazone/metformin lowered glycated HbA1c and hs-CRP levels over one year of treatment. This was followed by improved beta-cell function suggest and glycaemic control |
| Pfützner <i>et al</i> [96], 2011 | Patients with T2D (<i>n</i> = 146), with an average age of 59 yr | Received a fixed dose combination of 15 mg of pioglitazone with 850 mg of metformin given twice daily for 24 wk | Improved biomarkers of lipid metabolism, β -cell function, activity of the visceral adipose tissue, and chronic systemic inflammation. This was consistent with reduced hs-CRP and increased adiponectin levels |
| Bellia <i>et al</i> [104], 2012 | Patients with T2D (<i>n</i> = 27), with an average age of 56 yr | Received receive either rosuvastatin 20 mg daily or simvastatin 20 mg daily for 6 mo | Effectively reduced hs-CRP levels, but significantly diminished glycemic control and insulin secretion, without affecting insulin sensitivity |
| Derosa <i>et al</i> [97], 2012 | Patients with T2D (<i>n</i> = 167), with an average age of 53 yr | Received metformin gradually titrated until a mean dosage of 2500 ± 500 mg/d was reached for 8 ± 2 mo. Thereafter, patients were randomly assigned to take, vildagliptin at 50 mg twice a day for 12 mo | A combination of metformin and vildagliptin showed better effect in reducing body weight, glycemic control, Homeostatic Model Assessment for Insulin Resistance and improving β -cell function. However, no significant effect was observed for TNF- α levels |
| Brooks-Worrell and Palmer [105], 2013 | Patients with T2D (<i>n</i> = 26), with an average age of between 54 and 58 yr | Received rosiglitazone at 4 mg once/day and increased to twice/day if glycaemic control (HbA1c 70%) not achieved. Glyburide was at 2.5 mg and increased to twice per day up to a maximum of 10 mg twice/day if glycaemic control not achieved | Rosiglitazone reduced islet-specific T cell responses and improved glucagon-stimulated- β -cell secretion, consistent to decreasing in interferon gamma production. This was accompanied by increased adiponectin levels in comparison to glyburide-treated patients |
| Gagnon <i>et al</i> [106], 2014 | Patients with T2D (<i>n</i> = 35), with an average age of 54 yr | Received a combination of calcium carbonate (1200 mg) and cholecalciferol [2000-6000 IU to target 25(OH)D 0.75 nmol/L] for 6 mo | Treatment did not affect glucose tolerance, inflammatory markers (including hs-CRP levels) and β -cell function in patients with T2D, but improved insulin sensitivity in subjects with prediabetes |
| Zografou <i>et al</i> [98], 2015 | Patients with T2D (<i>n</i> = 64), with an average age between 52 and 56 yr | Received metformin at 1700 mg/d plus vildagliptin at 100 mg/d for 6 mo | A combination of metformin and vildagliptin reduced hs-CRP and improved glycemic control and $\beta\text{-cell}$ function |
| Tao et al[99], 2018 | Patients with T2D (<i>n</i> = 21), with an average of 29 yr | Received metformin at 2000 mg/d or saxagliptin at 5 mg/d for 24 wk | Treatment was comparatively effective at reducing body mass index and hs-CRP levels. This was parallel to improved glycemic control, lipid profiles and β -cell function |
| Zakerkish <i>et al</i> [107], 2019 | Patients with T2D (<i>n</i> = 50), with an average of 55 yr | Received Iranian propolis extract at 1000 mg/d for 90 d (3 mo) | Reduction on hs-CRP corresponded with beneficial effects of the extract in decreasing post prandial blood glucose, serum insulin, insulin resistance, and other inflammatory cytokines like TNF- α |

T2D: Type 2 diabetes; TNF-α: Tumor necrosis factor-alpha; hs-CRP: Highly sensitive C-reactive protein; HbA1C: Hemoglobin A1C; CRP: C-reactive protein.

> lipid profiles and $\beta\text{-cell}$ function in part by reducing pro-inflammatory markers like hs-CRP and TNF- $\!\alpha$ in patients with T2D[37,95-99]. Notably, evidence from these RCTs suggest that metformin may be most effective in improving β -cell function in patients with T2D when combined with other antidiabetic drugs such as rosiglitazone, pioglitazone and vildagliptin (Table 1). This highlights the potential enhanced effects of metformin when combined with other therapies in treating compilations of T2D, as discussed elsewhere[100,101].



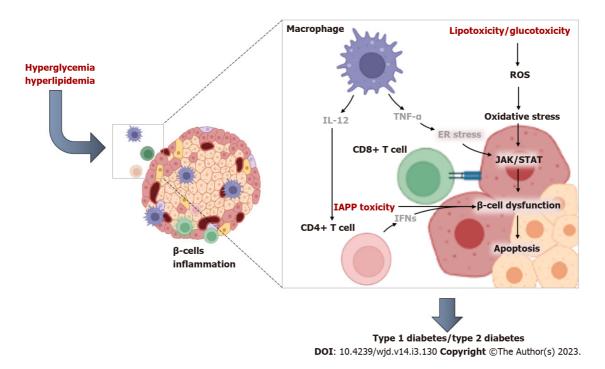


Figure 2 An overview of the pathological mechanisms linking impaired immune function and inflammation during β-cell dysfunction in conditions of type 2 diabetes (characterized by hyperglycemia and hyperlipidemia). Briefly, CD4+ T cells can be activated by interleukin-12 produced from macrophages and dendritic cells, and this consequence occurs as part of a vicious process involving cytotoxic T cells and recruitment by the pancreatic islets. Notably, elevated levels of tumor necrosis factor-alpha are linked with activation of pro-inflammatory signals such as Janus kinase/signal transducer and activator of transcription that promote β-cell failure. IL-12: Interleukin-12; TNF-α: Tumor necrosis factor-alpha; INFs: Interferons; IAPP: Islet amyloid polypeptide; ROS: Reactive oxygen species; ER stress: Endoplasmic reticulum stress; JAK/STAT: Janus kinase/signal transducer and activator of transcription.

Furthermore, evidence summarized in Table 1 indicate that other antidiabetic therapies can also improve β -cell function while modulating inflammatory markers in patients with T2D. For example, Tsunekawa et al[102] showed that administration of glimepiride (started from 1 mg daily and increased up to 6 mg daily for 12 wk) could ameliorate insulin resistance by decreasing plasma levels of TNF-α in patients with T2D. Dominguez *et al*[103] demonstrated that the TNF-α blocker, etanercept (received at 25 mg subcutaneously twice weekly for 4 wk) could reduce plasma levels of CRP and IL-6 to improve βcell function in patients with T2D. Such potential beneficial effects in improving β -cell function through the modulation of pro-inflammatory markers (especially hs-CRP and TNF-α) were confirmed through daily administration of other therapeutic drugs like rosiglitazone (at 4 mg), rosuvastatin or simvastatin (at 20 mg), a combination of calcium carbonate (at 1200 mg) and cholecalciferol (at 75 nmol/L), and even extracts like Iranian propolis extract (at 1000 mg) in patients with T2D[104-107]. These studies further indicate that interventions that block pro-inflammatory markers, especially hs-CRP and TNF- α levels, are likely to improve β -cell function in patients with T2D.

OXIDATIVE STRESS AND PANCREATIC B-CELL DYSFUNCTION IN T2D

Oxidative stress has emerged as a critical feature involved in health (physiology) or disease (pathophysiology)[108]. Oxidative stress is caused by the excessive production of free radical molecules (especially ROS) in response to severely diminished intracellular antioxidants. While ROS may be necessary for intracellular signaling [109], even during pathological conditions like cancer [110], this process is associated with unprecedented damage to many cellular processes during conditions like diabetes [23]. If uncontrolled, excess production of ROS can cause damage to DNA, cellular lipids and proteins, resulting in deteriorated metabolic function[14]. Although it can be sourced from different cellular compartments, the mitochondrial electron transport chain remains the major source of ROS within preclinal models and human systems [111]. The pancreatic β -cells contain mitochondria, which is vital for the regulation of glucose-stimulated insulin release by coupling glucose metabolism to insulin exocytosis[112]. Sustained exposure to hyperglycemic conditions has been associated with impaired βcell dysfunction through diverse biochemical and molecular mechanisms that implicate impaired oxidation phosphorylation, enhanced production of advanced glycation end products, and abnormal activation of protein kinase C, as well as the polyol and hexosamine pathways[14]. Defects in mitochondrial function are consistent with impaired metabolic function, which can ultimately result in accelerated apoptosis and β-cell death[111,113,114]. Likewise, severely depleted levels of intracellular antioxidants, such as Gpx, SOD, thioredoxin, and CAT have been linked with β -cell dysfunction in T2D, as highlighted in Figure 3.

Elevated markers of oxidative stress linked with β-cell dysfunction in T2D

Pancreatic β-cells are susceptible to oxidative damage through enhanced production of ROS. Glucose exposure can significantly increase ROS production and in turn cause damage to cultured β-cell-derived cells[115]. These effects were linked with activation of pro-inflammatory mechanisms involving JNK and MAPKs, leading to accelerated apoptosis and reduced β-cell mass[115]. This outcome further highlights the strong association between oxidative stress and inflammation during β-cell insults in conditions such as glucotoxicity. Diverse ROS molecules are actively studied for their detrimental effects in aggravating diabetes-associated complications, and these include superoxide (O2") and hydroxyl (OH) radicals, as well as hydrogen peroxide (H₂O₂)[14,108]. These ROS can cause chain activation of other free radicals, further driving the pathological features of oxidative stress. Nicotinamide adenine dinucleotide phosphate oxidases (NOX), existing in different isoforms depending on the specific type of tissue, is the prominent enzyme responsible for the generation of ROS, especially O₂-, although H₂O₂ can also be produced[116]. Previous evidence indicated that NOX-derived ROS generation was responsible for accelerated apoptosis in cultured pancreatic β-cells (NIT-1 cells)[117]. Consistent with other findings[115], these effects were associated with inflammatory pathways involving activation of JNK and inhibition of AKT, which are required for insulin signaling. In fact, through its interaction with mammalian target of rapamycin complex 1, AKT plays a major role during β-cell cycle development in part by regulating the activity of cyclins D2, D3 and cdk4/cyclin D[118]. Other findings have supported the detrimental effects of ROS in mediating pancreatic β -cell death, especially through the activation of stress-activated protein kinases, mitochondrial dysfunction, p38 and JNK, and MAPKs resulting in reduced glucose-stimulated insulin secretion[119-121].

Several studies have investigated the correlation between markers of oxidative stress and β-cell dysfunction in individuals with T2D or related metabolic complications. To highlight the close association between inflammation and oxidative stress, it was demonstrated that markers of Th1/Th2 cytokines and oxidative stress markers were significantly increased in patients with T2D when compared to controls[122]. These findings were consistent with reduced levels of nuclear factorerythroid factor 2-related factor 2 (Nrf2) and its downstream targets in peripheral blood mononuclear cells of diabetic patients. In many experimental models of T2D, Nrf2 is considered a master regulator of cellular survival, and its increased levels are instrumental to counteract the damaging effects of oxidative stress[123,124]. Reviewed evidence have further indicated that Nrf2 is essential for maintenance of β -cell mass to support the survival, function, and proliferation of β -cells [125]; whereas activation or upregulation of Nrf2 is necessary to diffuse inflammation, improve insulin sensitivity, decrease body weight, and protect against β-cell insult[125]. Thus, different therapeutic agents are entering clinical trials and being tested for their beneficial effects on β-cell survival and function by lowering markers of oxidative stress and promoting the antioxidant response in patients with metabolic disease and T2D[126-129].

Impaired levels of intracellular antioxidants linked with β-cell dysfunction in T2D

Pancreatic β cells are known to exhibit intrinsically low intracellular antioxidative capacity when compared to other tissues within the body [130]. Notably, Gpx, SOD, thioredoxins, and CAT are some of the prominent intracellular antioxidants that are important in protecting against oxidative insults to pancreatic β-cells[21]. The reduced expression of these antioxidants within the pancreas is pathologically implicated in ROS-induced β -cell damage. Importantly, it has been demonstrated that increasing intrinsic antioxidant defenses, through over-expressing Gpx-1 could protect β-cells from db/db mice against hyperglycemic insult[131]. Gpx is known for its high affinity to neutralize lipid hydroperoxides, with its low serum levels linked with oxidative stress and the progression of T2D[132]. The other isoforms of this enzyme, like Gpx-4, can salvage pancreatic β-cell death by reducing pro-inflammatory cytokines in pancreatic islets isolated from rats[133]. Over-expression of this glutathione-derived enzyme was very useful in alleviating dysregulated islet insulin production and secretion, mainly though acting on pancreatic and duodenal homeobox 1 (PDX-1) and uncoupling protein 2 (UCP-2), in high fat diet-fed mice[134]. PDX-1 is a transcriptional factor necessary for pancreatic development while UCP-2 is important for the detoxification of ROS through improved mitochondrial function. All these findings, highlight the significant role Gpx plays in detoxifying oxidative stress to improve β-cell function and insulin secretion under hyperglycemic conditions[135]. Ultimately, the genetic elimination of both Gpx-1 and SOD-1 can exert different influences on murine islet function and pancreatic integrity [136]. Even worse, this results in significantly reduced plasma insulin concentrations and islet β -cells mass, which also correlate with increased blood glucose, and blocked glucose-stimulated insulin secretion. These effects are orchestrated mainly through elevation of ROS levels within the pancreatic islets, especially the concentrations of O_2^- and H_2O_2 , leading to p53 phosphorylation[136]. While this is related to the diminished role of SOD, known for its neutralizing effects on ROS by eliminating O₂; phosphorylation of p53 has evolved the capability to incorporate unique environmental signals that facilitate DNA damage[137].

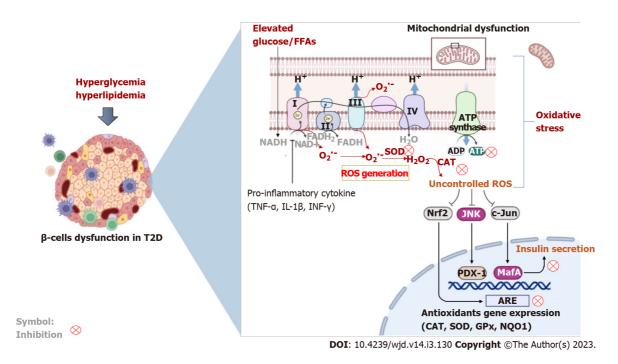


Figure 3 An overview of some pathophysiological mechanisms that highlight the interrelated link between inflammation and oxidative stress during β-cell dysfunction in type 2 diabetes. Briefly, elevated levels of glucose (hyperglycemia) and lipids (hyperlipidemia) are consistent with an abnormal inflammatory response (due to increased levels of tumor necrosis factor-alpha, interferon-gamma, and interleukin 1 beta) and impaired mitochondrial electron transport chain that causes enhanced generation of reactive oxygen species implicated in β-cell dysfunction. Both inflammation and oxidative stress are responsible for activating cellular destructive mechanisms such as the c-Jun N-terminal kinase pathway and suppressing intracellular antioxidant responses (e.g., nuclear factor erythroid 2-related factor 2, catalase, superoxide dismutase and others), leading to accelerated β-cell injury. ROS: Reactive oxygen species; SOD: Superoxide dismutase; CAT: Catalase; Gpx: Glutathione peroxidase; NQO1: NAD(P)H quinone dehydrogenase 1; NAD(H): Nicotinamide adenine dinucleotide; FADH: Flavin adenine dinucleotide; Nrf2: Nuclear factor erythroid 2-related factor 2; JNK: c-Jun N-terminal kinase; PDX-1: Pancreatic duodenal homeobox 1; ARE: Antioxidant response element; TNF-α: Tumor necrosis factor-alpha; IL-1β: Interleukin 1 beta; INF-γ: Interferon-gamma; ADP: Adenosine diphosphate; ATP: Adenosine triphosphate; ROS: Reactive oxygen species; T2D: Type 2 diabetes; FFAs: Free fatty acids.

Beyond the effects of Gpx1 and SOD, the thioredoxin reductase-dependent mechanism is another essential mechanism necessary for the detoxification of H_2O_2 -induced β -cell dysfunction[138]. Thioredoxins mainly act by reducing oxidized cysteine residues, with elevated levels of this enzyme linked to increased levels of circulating non-esterified fatty acids in patients with T2D[139]. Other findings indicate that elevated thioredoxin-interacting protein (TXNIP) levels correlate with increased β -cell apoptosis[140]; whereas TXNIP deficiency safeguards against T2D by promoting β -cell survival [141]. In fact, TXNIP is considered essential for the regulation of pancreatic β -cell function and other complications of T2D[142,143]. Another important enzyme, CAT, has long established to protect transgenic mouse β -cells by neutralizing H_2O_2 [144]. Indeed, CAT mainly acts by converting H_2O_2 to water and oxygen. Accumulating evidence indicate that increased levels of this enzyme is important for pancreatic β -cell protection, while maintaining insulin secreting function in conditions of T2D[144-146]. Such evidence is consistent with other reviews highlighting the susceptibility of islets to oxidative damage, and the importance of intracellular antioxidant enzymes in protecting β -cells against diabetic insults[21,147].

CONCLUSION

Currently, diverse pathological mechanisms are acknowledged to be involved in the development and progression of T2D. Although both T1D and T2D are associated with β -cell dysfunction, insulin resistance and obesity are the prominent characteristic features for T2D. During a state of obesity, adipose tissue expansion is linked with the production of an array of pro-inflammatory markers that are involved in accelerating β -cell dysfunction. Such effects are consistent with impaired immune response, further driving insulin resistance and elevated blood glucose levels. Oxidative stress also concurrently occurs with inflammation and can cause havoc in many biochemical processes leading to pancreatic β -cell death. Even worse, while sustainably elevated markers of inflammation and oxidative stress are related to β -cell dysfunction, the intracellular defense (antioxidant) mechanisms responsible for ameliorating some of these effects are significantly depleted during T2D. There is limited clinical evidence supporting the beneficial effects of commonly used antidiabetic therapies in enhancing

intracellular antioxidants to protect against β-cell dysfunction in T2D. The systematic analysis of RCTs supports the potential beneficial effects of metformin (especially when used in combination with other antidiabetic therapies like rosiglitazone, pioglitazone and vildagliptin) in improving β -cell function in part by reducing pro-inflammatory markers like hs-CRP and TNF-α in patients with T2D. Thus, beyond improving blood glucose control like with most antidiabetic drugs, future research should invest in developing therapies that can promote intracellular antioxidants and reduce markers of inflammation and oxidative stress to limit pancreatic β -cell failure in patients with T2D.

ACKNOWLEDGEMENTS

The work by Khanyisani Ziqubu, reported herein was made possible through partial funding by the South African Medical Research Council through its Division of Research Capacity Development under the Researcher Development Award Programme. The content hereof is the sole responsibility of the authors and do not necessarily represent the official views of the funders.

FOOTNOTES

Author contributions: Dludla PV, Mabhida SE, Ziqubu K, Nkambule BB, Mazibuko-Mbeje SE, Hanser S, Basson AK, Pheiffer C, Kengne AP contributed to writing and final approval of the manuscript.

Supported by the Biomedical Research and Innovation Platform, of the South African Medical Research Council (SAMRC), and the National Research Foundation (grant No. 132534 and 141929). The content hereof is the sole responsibility of the authors and do not necessarily represent the official views of the SAMRC or the funders.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

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S-Editor: Wang JJ L-Editor: A P-Editor: Wang JJ

REFERENCES

- World Health Organization. The top ten leading causes of death. [cited 10 September 2022]. Available from: https:// www.who.int/news-room/fact-sheets/detail/the-top-10-causes-of-death
- International Diabetes Federation. IDF Diabetes Atlas Tenth Edition. [cited 10 September 2022]. Available from: https://diabetesatlas.org
- Mazidi M, Kengne AP, Katsiki N, Mikhailidis DP, Banach M. Lipid accumulation product and triglycerides/glucose index are useful predictors of insulin resistance. J Diabetes Complications 2018; 32: 266-270 [PMID: 29395839 DOI: 10.1016/j.jdiacomp.2017.10.007
- Cerf ME. Beta cell dysfunction and insulin resistance. Front Endocrinol (Lausanne) 2013; 4: 37 [PMID: 23542897 DOI: 10.3389/fendo.2013.00037]
- Saisho Y. β-cell dysfunction: Its critical role in prevention and management of type 2 diabetes. World J Diabetes 2015; 6: 109-124 [PMID: 25685282 DOI: 10.4239/wjd.v6.i1.109]
- Do OH, Gunton JE, Gaisano HY, Thorn P. Changes in beta cell function occur in prediabetes and early disease in the Lepr (db) mouse model of diabetes. Diabetologia 2016; 59: 1222-1230 [PMID: 27048248 DOI: 10.1007/s00125-016-3942-3]
- Pickup JC, Crook MA. Is type II diabetes mellitus a disease of the innate immune system? Diabetologia 1998; 41: 1241-1248 [PMID: 9794114 DOI: 10.1007/s001250051058]
- Lontchi-Yimagou E, Sobngwi E, Matsha TE, Kengne AP. Diabetes mellitus and inflammation. Curr Diab Rep 2013; 13: 435-444 [PMID: 23494755 DOI: 10.1007/s11892-013-0375-y]

- Berbudi A, Rahmadika N, Tjahjadi AI, Ruslami R. Type 2 Diabetes and its Impact on the Immune System. Curr Diabetes Rev 2020; 16: 442-449 [PMID: 31657690 DOI: 10.2174/1573399815666191024085838]
- Jo S, Fang S. Therapeutic Strategies for Diabetes: Immune Modulation in Pancreatic β Cells. Front Endocrinol (Lausanne) 2021; 12: 716692 [PMID: 34484126 DOI: 10.3389/fendo.2021.716692]
- Sun X, Wang L, Obayomi SMB, Wei Z. Epigenetic Regulation of β Cell Identity and Dysfunction. Front Endocrinol (Lausanne) 2021; 12: 725131 [PMID: 34630329 DOI: 10.3389/fendo.2021.725131]
- Cuenco J, Dalmas E. Islet Inflammation and β Cell Dysfunction in Type 2 Diabetes. Handb Exp Pharmacol 2022; 274: 227-251 [PMID: 35044537 DOI: 10.1007/164_2021_571]
- Narasimhan A, Flores RR, Robbins PD, Niedernhofer LJ. Role of Cellular Senescence in Type II Diabetes. Endocrinology 2021; 162 [PMID: 34363464 DOI: 10.1210/endocr/bqab136]
- Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res 2010; 107: 1058-1070 [PMID: 21030723 14 DOI: 10.1161/CIRCRESAHA.110.223545]
- 15 Asmat U, Abad K, Ismail K. Diabetes mellitus and oxidative stress-A concise review. Saudi Pharm J 2016; 24: 547-553 [PMID: 27752226 DOI: 10.1016/j.jsps.2015.03.013]
- Krauss RM. Lipids and lipoproteins in patients with type 2 diabetes. Diabetes Care 2004; 27: 1496-1504 [PMID: 15161808 DOI: 10.2337/diacare.27.6.1496]
- Lenzen S, Drinkgern J, Tiedge M. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. Free Radic Biol Med 1996; 20: 463-466 [PMID: 8720919 DOI: 10.1016/0891-5849(96)02051-5]
- Drews G, Krippeit-Drews P, Düfer M. Oxidative stress and beta-cell dysfunction. Pflugers Arch 2010; 460: 703-718 [PMID: 20652307 DOI: 10.1007/s00424-010-0862-9]
- Rohm TV, Meier DT, Olefsky JM, Donath MY. Inflammation in obesity, diabetes, and related disorders. Immunity 2022; 55: 31-55 [PMID: 35021057 DOI: 10.1016/j.immuni.2021.12.013]
- Singh RG, Yoon HD, Wu LM, Lu J, Plank LD, Petrov MS. Ectopic fat accumulation in the pancreas and its clinical relevance: A systematic review, meta-analysis, and meta-regression. Metabolism 2017; 69: 1-13 [PMID: 28285638 DOI: 10.1016/j.metabol.2016.12.012]
- Lei XG, Vatamaniuk MZ. Two tales of antioxidant enzymes on β cells and diabetes. Antioxid Redox Signal 2011; 14: 489-503 [PMID: 20618069 DOI: 10.1089/ars.2010.3416]
- Wang J, Wang H. Oxidative Stress in Pancreatic Beta Cell Regeneration. Oxid Med Cell Longev 2017; 2017: 1930261 [PMID: 28845211 DOI: 10.1155/2017/1930261]
- Halim M, Halim A. The effects of inflammation, aging and oxidative stress on the pathogenesis of diabetes mellitus (type 2 diabetes). Diabetes Metab Syndr 2019; 13: 1165-1172 [PMID: 31336460 DOI: 10.1016/j.dsx.2019.01.040]
- Wilcox G. Insulin and insulin resistance. Clin Biochem Rev 2005; 26: 19-39 [PMID: 16278749]
- Boucher J, Kleinridders A, Kahn CR. Insulin receptor signaling in normal and insulin-resistant states. Cold Spring Harb Perspect Biol 2014; 6 [PMID: 24384568 DOI: 10.1101/cshperspect.a009191]
- Wondmkun YT. Obesity, Insulin Resistance, and Type 2 Diabetes: Associations and Therapeutic Implications. Diabetes Metab Syndr Obes 2020; 13: 3611-3616 [PMID: 33116712 DOI: 10.2147/DMSO.S275898]
- 27 Dludla PV, Nkambule BB, Jack B, Mkandla Z, Mutize T, Silvestri S, Orlando P, Tiano L, Louw J, Mazibuko-Mbeje SE. Inflammation and Oxidative Stress in an Obese State and the Protective Effects of Gallic Acid. Nutrients 2018; 11 [PMID: 30577684 DOI: 10.3390/nu110100231
- Golacki J, Matuszek M, Matyjaszek-Matuszek B. Link between Insulin Resistance and Obesity-From Diagnosis to 28 Treatment. Diagnostics (Basel) 2022; 12 [PMID: 35885586 DOI: 10.3390/diagnostics12071681]
- Kasuga M. Insulin resistance and pancreatic beta cell failure. J Clin Invest 2006; 116: 1756-1760 [PMID: 16823472 DOI: 10.1172/JCI291891
- Böni-Schnetzler M, Meier DT. Islet inflammation in type 2 diabetes. Semin Immunopathol 2019; 41: 501-513 [PMID: 30 30989320 DOI: 10.1007/s00281-019-00745-4]
- Cerf ME. Beta Cell Physiological Dynamics and Dysfunctional Transitions in Response to Islet Inflammation in Obesity and Diabetes. Metabolites 2020; 10 [PMID: 33182622 DOI: 10.3390/metabo10110452]
- Ying W, Fu W, Lee YS, Olefsky JM. The role of macrophages in obesity-associated islet inflammation and β-cell abnormalities. Nat Rev Endocrinol 2020; 16: 81-90 [PMID: 31836875 DOI: 10.1038/s41574-019-0286-3]
- Hardy OT, Czech MP, Corvera S. What causes the insulin resistance underlying obesity? Curr Opin Endocrinol Diabetes Obes 2012; 19: 81-87 [PMID: 22327367 DOI: 10.1097/MED.0b013e3283514e13]
- Tsalamandris S, Antonopoulos AS, Oikonomou E, Papamikroulis GA, Vogiatzi G, Papaioannou S, Deftereos S, Tousoulis D. The Role of Inflammation in Diabetes: Current Concepts and Future Perspectives. Eur Cardiol 2019; 14: 50-59 [PMID: 31131037 DOI: 10.15420/ecr.2018.33.1]
- Rodríguez-Hernández H, Simental-Mendía LE, Rodríguez-Ramírez G, Reyes-Romero MA. Obesity and inflammation: epidemiology, risk factors, and markers of inflammation. Int J Endocrinol 2013; 2013: 678159 [PMID: 23690772 DOI: 10.1155/2013/6781591
- 36 Daousi C, Casson IF, Gill GV, MacFarlane IA, Wilding JP, Pinkney JH. Prevalence of obesity in type 2 diabetes in secondary care: association with cardiovascular risk factors. Postgrad Med J 2006; 82: 280-284 [PMID: 16597817 DOI: 10.1136/pmj.2005.039032]
- Pfützner A, Standl E, Strotmann HJ, Schulze J, Hohberg C, Lübben G, Pahler S, Schöndorf T, Forst T. Association of high-sensitive C-reactive protein with advanced stage beta-cell dysfunction and insulin resistance in patients with type 2 diabetes mellitus. Clin Chem Lab Med 2006; 44: 556-560 [PMID: 16681424 DOI: 10.1515/CCLM.2006.108]
- PrayGod G, Filteau S, Range N, Kitilya B, Kavishe BB, Ramaiya K, Jeremiah K, Rehman AM, Changalucha J, Olsen MF, Andersen AB, Friis H, Krogh-Madsen R, Faurholt-Jepsen D. β-cell dysfunction and insulin resistance in relation to pre-diabetes and diabetes among adults in north-western Tanzania: a cross-sectional study. Trop Med Int Health 2021; 26: 435-443 [PMID: 33406288 DOI: 10.1111/tmi.13545]
- Ndevahoma F, Nkambule BB, Dludla PV, Mukesi M, Natanael KN, Nyambuya TM. The effect of underlying

- inflammation on iron metabolism, cardiovascular risk and renal function in patients with type 2 diabetes. EJHaem 2021; 2: 357-365 [PMID: 35844722 DOI: 10.1002/jha2.257]
- Sproston NR, Ashworth JJ. Role of C-Reactive Protein at Sites of Inflammation and Infection. Front Immunol 2018; 9: 754 [PMID: 29706967 DOI: 10.3389/fimmu.2018.00754]
- Kaneko N, Kurata M, Yamamoto T, Morikawa S, Masumoto J. The role of interleukin-1 in general pathology. Inflamm Regen 2019; 39: 12 [PMID: 31182982 DOI: 10.1186/s41232-019-0101-5]
- Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L. Inflammatory responses and inflammationassociated diseases in organs. Oncotarget 2018; 9: 7204-7218 [PMID: 29467962 DOI: 10.18632/oncotarget.23208]
- 43 Sharma AK, Bharti S, Ojha S, Bhatia J, Kumar N, Ray R, Kumari S, Arya DS. Up-regulation of PPARγ, heat shock protein-27 and -72 by naringin attenuates insulin resistance, β-cell dysfunction, hepatic steatosis and kidney damage in a rat model of type 2 diabetes. Br J Nutr 2011; 106: 1713-1723 [PMID: 21736771 DOI: 10.1017/S000711451100225X]
- Sun W, Wu Y, Gao M, Tian Y, Qi P, Shen Y, Huang L, Shi L, Wang Y, Liu X. C-reactive protein promotes inflammation through TLR4/NF-κB/TGF-β pathway in HL-1 cells. Biosci Rep 2019; 39 [PMID: 31391207 DOI: 10.1042/BSR20190888]
- Zhang L, Shen ZY, Wang K, Li W, Shi JM, Osoro EK, Ullah N, Zhou Y, Ji SR. C-reactive protein exacerbates epithelialmesenchymal transition through Wnt/β-catenin and ERK signaling in streptozocin-induced diabetic nephropathy. FASEB J 2019; **33**: 6551-6563 [PMID: 30794428 DOI: 10.1096/fj.201801865RR]
- Pradhan AD, Manson JE, Rifai N, Buring JE, Ridker PM. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. JAMA 2001; 286: 327-334 [PMID: 11466099 DOI: 10.1001/jama.286.3.327]
- Hu FB, Meigs JB, Li TY, Rifai N, Manson JE. Inflammatory markers and risk of developing type 2 diabetes in women. Diabetes 2004; 53: 693-700 [PMID: 14988254 DOI: 10.2337/diabetes.53.3.693]
- Weber KS, Nowotny B, Strassburger K, Pacini G, Müssig K, Szendroedi J, Herder C, Roden M; GDS Group. The Role of Markers of Low-Grade Inflammation for the Early Time Course of Glycemic Control, Glucose Disappearance Rate, and β -Cell Function in Recently Diagnosed Type 1 and Type 2 Diabetes. Diabetes Care 2015; 38: 1758-1767 [PMID: 26153272 DOI: 10.2337/dc15-0169]
- Barazzoni R, Zanetti M, Davanzo G, Kiwanuka E, Carraro P, Tiengo A, Tessari P. Increased fibrinogen production in type 2 diabetic patients without detectable vascular complications: correlation with plasma glucagon concentrations. J Clin Endocrinol Metab 2000; 85: 3121-3125 [PMID: 10999796 DOI: 10.1210/jcem.85.9.6779]
- Sarangi R, Padhi S, Mohapatra S, Swain S, Padhy RK, Mandal MK, Patro SK, Kumar S. Serum high sensitivity Creactive protein, nitric oxide metabolites, plasma fibrinogen, and lipid parameters in Indian type 2 diabetic males. Diabetes Metab Syndr 2012; 6: 9-14 [PMID: 23014248 DOI: 10.1016/j.dsx.2012.05.015]
- Emanuele N, Azad N, Abraira C, Henderson W, Colwell J, Levin S, Nuttall F, Comstock J, Sawin C, Silbert C, Marcovina S, Lee HS. Effect of intensive glycemic control on fibrinogen, lipids, and lipoproteins: Veterans Affairs Cooperative Study in Type II Diabetes Mellitus. Arch Intern Med 1998; 158: 2485-2490 [PMID: 9855387 DOI:
- Deng Y, Papageorgiou DP, Li X, Perakakis N, Mantzoros CS, Dao M, Karniadakis GE. Quantifying Fibrinogen-Dependent Aggregation of Red Blood Cells in Type 2 Diabetes Mellitus. Biophys J 2020; 119: 900-912 [PMID: 32814061 DOI: 10.1016/j.bpj.2020.07.0261
- Le DS, Miles R, Savage PJ, Cornell E, Tracy RP, Knowler WC, Krakoff J. The association of plasma fibringen concentration with diabetic microvascular complications in young adults with early-onset of type 2 diabetes. Diabetes Res Clin Pract 2008; 82: 317-323 [PMID: 18922595 DOI: 10.1016/j.diabres.2008.08.019]
- Bembde AS. A study of plasma fibrinogen level in type-2 diabetes mellitus and its relation to glycemic control. Indian J Hematol Blood Transfus 2012; 28: 105-108 [PMID: 23730017 DOI: 10.1007/s12288-011-0116-9]
- Chen QF, Cao D, Ye TT, Deng HH, Zhu H. Peripheral Arterial Disease in Type 2 Diabetes Is Associated with an Increase in Fibrinogen Levels. Int J Endocrinol 2018; 2018: 3709534 [PMID: 30532778 DOI: 10.1155/2018/3709534]
- Sarangi S, Mahapatra S, Padhi S. Association of plasma fibrinogen and serum high-sensitivity C-reactive protein in type 2 diabetes mellitus. J Curr Res Sci Med 2015; 1: 21-26
- Iwai S, Okazaki M, Nara K, Murakami H, Maruyama M, Kiuchi Y, Oguchi K. Altered fibrinogen and prothrombin mRNA expression in streptozotocin-induced diabetic rats. Showa Univ J Med Sci 2000; 12: 295-302 [DOI: 10.15369/sujms1989.12.295]
- Masamune A, Kikuta K, Watanabe T, Satoh K, Hirota M, Hamada S, Shimosegawa T. Fibrinogen induces cytokine and collagen production in pancreatic stellate cells. Gut 2009; 58: 550-559 [PMID: 19052021 DOI: 10.1136/gut.2008.154401]
- Guo Q, Zhang B, Dong X, Xie Q, Guo E, Huang H, Wu Y. Elevated levels of plasma fibrinogen in patients with pancreatic cancer: possible role of a distant metastasis predictor. Pancreas 2009; 38: e75-e79 [PMID: 19276866 DOI: 10.1097/MPA.0b013e3181987d86
- Ye X, Huai J, Chen R, Ding J, Chen Y, Cai Z. Correlation of fibrinogen-like protein 2 with disease progression in patients with severe acute pancreatitis. Exp Ther Med 2014; 7: 85-89 [PMID: 24348769 DOI: 10.3892/etm.2013.1354]
- Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. Cold Spring Harb Perspect Biol 2014; 6: a016295 [PMID: 25190079 DOI: 10.1101/cshperspect.a016295]
- Mahlangu T, Dludla PV, Nyambuya TM, Mxinwa V, Mazibuko-Mbeje SE, Cirilli I, Marcheggiani F, Tiano L, Louw J, Nkambule BB. A systematic review on the functional role of Th1/Th2 cytokines in type 2 diabetes and related metabolic complications. Cytokine 2020; 126: 154892 [PMID: 31704479 DOI: 10.1016/j.cyto.2019.154892]
- Sahoo M, Ceballos-Olvera I, del Barrio L, Re F. Role of the inflammasome, IL-1β, and IL-18 in bacterial infections. ScientificWorldJournal 2011; 11: 2037-2050 [PMID: 22125454 DOI: 10.1100/2011/212680]
- Rosenblum MD, Remedios KA, Abbas AK. Mechanisms of human autoimmunity. J Clin Invest 2015; 125: 2228-2233 [PMID: 25893595 DOI: 10.1172/JCI78088]
- Mahlangu TJ, Dludla PV, Mxinwa V, Mkandla Z, Tiano L, Louw J, Mutize T, Nyambuya TM, Nkambule BB. Elevated T-helper 2 cytokine levels in high fat diet-fed C57BL/6 mice are attenuated by short-term 6-week treatment with a combination of low-dose aspirin and metformin. Cytokine 2020; 128: 154999 [PMID: 32014718 DOI:



10.1016/j.cvto.2020.1549991

- Martín-Sánchez F, Diamond C, Zeitler M, Gomez AI, Baroja-Mazo A, Bagnall J, Spiller D, White M, Daniels MJ, Mortellaro A, Peñalver M, Paszek P, Steringer JP, Nickel W, Brough D, Pelegrín P. Inflammasome-dependent IL-1β release depends upon membrane permeabilisation. Cell Death Differ 2016; 23: 1219-1231 [PMID: 26868913 DOI: 10.1038/cdd.2015.176]
- Marasco MR, Conteh AM, Reissaus CA, Cupit JE 5th, Appleman EM, Mirmira RG, Linnemann AK. Interleukin-6 Reduces β-Cell Oxidative Stress by Linking Autophagy With the Antioxidant Response. Diabetes 2018; 67: 1576-1588 [PMID: 29784660 DOI: 10.2337/db17-1280]
- Linnemann AK, Blumer J, Marasco MR, Battiola TJ, Umhoefer HM, Han JY, Lamming DW, Davis DB. Interleukin 6 protects pancreatic β cells from apoptosis by stimulation of autophagy. FASEB J 2017; 31: 4140-4152 [PMID: 28592636 DOI: 10.1096/fj.201700061RR]
- Van Belle TL, Pagni PP, Liao J, Sachithanantham S, Dave A, Bel Hani A, Manenkova Y, Amirian N, Yang C, Morin B, Zhang H, Campbell IL, von Herrath MG. Beta-cell specific production of IL6 in conjunction with a mainly intracellular but not mainly surface viral protein causes diabetes. J Autoimmun 2014; 55: 24-32 [PMID: 24582317 DOI: 10.1016/j.jaut.2014.02.002]
- Maedler K, Sergeev P, Ris F, Oberholzer J, Joller-Jemelka HI, Spinas GA, Kaiser N, Halban PA, Donath MY. Glucoseinduced beta cell production of IL-1 beta contributes to glucotoxicity in human pancreatic islets. J Clin Invest 2002; 110: 851-860 [PMID: 12235117 DOI: 10.1172/JCI15318]
- 71 Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor-alpha: direct role in obesitylinked insulin resistance. Science 1993; 259: 87-91 [PMID: 7678183 DOI: 10.1126/science.7678183]
- Ikeda H. KK mouse. Diabetes Res Clin Pract 1994; 24 Suppl: S313-S316 [PMID: 7859626 DOI: 10.1016/0168-8227(94)90268-21
- Darogha SN. Serum levels of TNF-a and IFN-g gene polymorphism in type 2 diabetes mellitus in kurdish patients. Cell Mol Biol (Noisy-le-grand) 2021; 67: 171-177 [PMID: 34817320 DOI: 10.14715/cmb/2021.67.2.27]
- **Ibfelt T**, Fischer CP, Plomgaard P, van Hall G, Pedersen BK. The acute effects of low-dose TNF-α on glucose metabolism and β -cell function in humans. *Mediators Inflamm* 2014; **2014**: 295478 [PMID: 24692847 DOI:
- Imanparast F, Javaheri J, Kamankesh F, Rafiei F, Salehi A, Mollaaliakbari Z, Rezaei F, Rahimi A, Abbasi E. The effects of chromium and vitamin D3 co-supplementation on insulin resistance and tumor necrosis factor-alpha in type 2 diabetes: a randomized placebo-controlled trial. Appl Physiol Nutr Metab 2020; 45: 471-477 [PMID: 31593637 DOI:
- 76 Stephens LA, Thomas HE, Ming L, Grell M, Darwiche R, Volodin L, Kay TW. Tumor necrosis factor-alpha-activated cell death pathways in NIT-1 insulinoma cells and primary pancreatic beta cells. Endocrinology 1999; 140: 3219-3227 [PMID: 10385418 DOI: 10.1210/endo.140.7.6873]
- Barthson J, Germano CM, Moore F, Maida A, Drucker DJ, Marchetti P, Gysemans C, Mathieu C, Nuñez G, Jurisicova A, Eizirik DL, Gurzov EN. Cytokines tumor necrosis factor-α and interferon-γ induce pancreatic β-cell apoptosis through STAT1-mediated Bim protein activation. J Biol Chem 2011; 286: 39632-39643 [PMID: 21937453 DOI: 10.1074/jbc.M111.253591]
- Wang C, Guan Y, Yang J. Cytokines in the Progression of Pancreatic β-Cell Dysfunction. Int J Endocrinol 2010; 2010: 515136 [PMID: 21113299 DOI: 10.1155/2010/515136]
- Blank SE, Johnson EC, Weeks DK, Wysham CH. Circulating dendritic cell number and intracellular TNF-α production in women with type 2 diabetes. Acta Diabetol 2012; 49 Suppl 1: S25-S32 [PMID: 20449757 DOI: 10.1007/s00592-010-0190-81
- Houjeghani S, Kheirouri S, Faraji E, Jafarabadi MA. l-Carnosine supplementation attenuated fasting glucose, triglycerides, advanced glycation end products, and tumor necrosis factor-α levels in patients with type 2 diabetes: a double-blind placebo-controlled randomized clinical trial. Nutr Res 2018; 49: 96-106 [PMID: 29420997 DOI: 10.1016/i.nutres.2017.11.0031
- Alfadul H, Sabico S, Al-Daghri NM. The role of interleukin-1β in type 2 diabetes mellitus: A systematic review and meta-analysis. Front Endocrinol (Lausanne) 2022; 13: 901616 [PMID: 35966098 DOI: 10.3389/fendo.2022.901616]
- Citro A, Campo F, Dugnani E, Piemonti L. Innate Immunity Mediated Inflammation and Beta Cell Function: Neighbors or Enemies? Front Endocrinol (Lausanne) 2020; 11: 606332 [PMID: 33628197 DOI: 10.3389/fendo.2020.606332]
- 83 Hilhorst M, Shirai T, Berry G, Goronzy JJ, Weyand CM. T cell-macrophage interactions and granuloma formation in vasculitis. Front Immunol 2014; 5: 432 [PMID: 25309534 DOI: 10.3389/fimmu.2014.00432]
- Luckheeram RV, Zhou R, Verma AD, Xia B. CD4⁺T cells: differentiation and functions. Clin Dev Immunol 2012; 2012: 925135 [PMID: 22474485 DOI: 10.1155/2012/925135]
- Nyambuya TM, Dludla PV, Mxinwa V, Nkambule BB. T-cell activation and cardiovascular risk in adults with type 2 diabetes mellitus: A systematic review and meta-analysis. Clin Immunol 2020; 210: 108313 [PMID: 31765833 DOI: 10.1016/j.clim.2019.108313]
- Xia C, Rao X, Zhong J. Role of T Lymphocytes in Type 2 Diabetes and Diabetes-Associated Inflammation. J Diabetes Res 2017; **2017**: 6494795 [PMID: 28251163 DOI: 10.1155/2017/6494795]
- Yoon JW, Jun HS. Autoimmune destruction of pancreatic beta cells. Am J Ther 2005; 12: 580-591 [PMID: 16280652 DOI: 10.1097/01.mjt.0000178767.67857.63]
- Pakala SV, Kurrer MO, Katz JD. T helper 2 (Th2) T cells induce acute pancreatitis and diabetes in immune-compromised nonobese diabetic (NOD) mice. J Exp Med 1997; 186: 299-306 [PMID: 9221759 DOI: 10.1084/jem.186.2.299]
- Espinosa-Carrasco G, Le Saout C, Fontanaud P, Stratmann T, Mollard P, Schaeffer M, Hernandez J. CD4(+) T Helper Cells Play a Key Role in Maintaining Diabetogenic CD8(+) T Cell Function in the Pancreas. Front Immunol 2017; 8: 2001 [PMID: 29403481 DOI: 10.3389/fimmu.2017.02001]
- Almawi WY, Tamim H, Azar ST. Clinical review 103: T helper type 1 and 2 cytokines mediate the onset and progression of type I (insulin-dependent) diabetes. J Clin Endocrinol Metab 1999; 84: 1497-1502 [PMID: 10323367 DOI:

- 10.1210/icem.84.5.56991
- Mxinwa V, Dludla PV, Nyambuya TM, Mokgalaboni K, Mazibuko-Mbeje SE, Nkambule BB. Natural killer cell levels in adults living with type 2 diabetes: a systematic review and meta-analysis of clinical studies. BMC Immunol 2020; 21: 51 [PMID: 32907543 DOI: 10.1186/s12865-020-00378-5]
- Nagi DK, Yudkin JS. Effects of metformin on insulin resistance, risk factors for cardiovascular disease, and plasminogen activator inhibitor in NIDDM subjects. A study of two ethnic groups. Diabetes Care 1993; 16: 621-629 [PMID: 8462390 DOI: 10.2337/diacare.16.4.6211
- Prattichizzo F, Giuliani A, Mensà E, Sabbatinelli J, De Nigris V, Rippo MR, La Sala L, Procopio AD, Olivieri F, Ceriello A. Pleiotropic effects of metformin: Shaping the microbiome to manage type 2 diabetes and postpone ageing. Ageing Res Rev 2018; 48: 87-98 [PMID: 30336272 DOI: 10.1016/j.arr.2018.10.003]
- Dludla PV, Nyambuya TM, Johnson R, Silvestri S, Orlando P, Mazibuko-Mbeje SE, Gabuza KB, Mxinwa V, Mokgalaboni K, Tiano L, Muller CJF, Louw J, Nkambule BB. Metformin and heart failure-related outcomes in patients with or without diabetes: a systematic review of randomized controlled trials. Heart Fail Rev 2021; 26: 1437-1445 [PMID: 32157481 DOI: 10.1007/s10741-020-09942-y]
- 95 Hamann A, Garcia-Puig J, Paul G, Donaldson J, Stewart M. Comparison of fixed-dose rosiglitazone/metformin combination therapy with sulphonylurea plus metformin in overweight individuals with Type 2 diabetes inadequately controlled on metformin alone. Exp Clin Endocrinol Diabetes 2008; 116: 6-13 [PMID: 18095238 DOI: 10.1055/s-2007-9844411
- Pfützner A, Schöndorf T, Tschöpe D, Lobmann R, Merke J, Müller J, Lehmann U, Fuchs W, Forst T. PIOfix-study: effects of pioglitazone/metformin fixed combination in comparison with a combination of metformin with glimepiride on diabetic dyslipidemia. Diabetes Technol Ther 2011; 13: 637-643 [PMID: 21457065 DOI: 10.1089/dia.2010.0233]
- Derosa G, Ragonesi PD, Carbone A, Fogari E, D'Angelo A, Cicero AF, Maffioli P. Vildagliptin action on some adipocytokine levels in type 2 diabetic patients: a 12-month, placebo-controlled study. Expert Opin Pharmacother 2012; 13: 2581-2591 [PMID: 23121473 DOI: 10.1517/14656566.2012.734499]
- Zografou I, Sampanis C, Gkaliagkousi E, Iliadis F, Papageorgiou A, Doukelis P, Vogiatzis K, Douma S. Effect of vildagliptin on hsCRP and arterial stiffness in patients with type 2 diabetes mellitus. Hormones (Athens) 2015; 14: 118-125 [PMID: 25402372 DOI: 10.14310/horm.2002.1512]
- Tao T, Wu P, Wang Y, Liu W. Comparison of glycemic control and β-cell function in new onset T2DM patients with PCOS of metformin and saxagliptin monotherapy or combination treatment. BMC Endocr Disord 2018; 18: 14 [PMID: 29482528 DOI: 10.1186/s12902-018-0243-5]
- Prattichizzo F, Ceriello A. Is time ready for combination therapy at diagnosis of type 2 diabetes? Diabetes Metab Res Rev 2021; **37**: e3460 [PMID: 34240524 DOI: 10.1002/dmrr.3460]
- Dludla PV, Silvestri S, Orlando P, Gabuza KB, Mazibuko-Mbeje SE, Nyambuya TM, Mxinwa V, Mokgalaboni K, Johnson R, Muller CJF, Tiano L, Louw J, Nkambule BB. Exploring the Comparative Efficacy of Metformin and Resveratrol in the Management of Diabetes-associated Complications: A Systematic Review of Preclinical Studies. Nutrients 2020; 12 [PMID: 32168855 DOI: 10.3390/nu12030739]
- Tsunekawa T, Hayashi T, Suzuki Y, Matsui-Hirai H, Kano H, Fukatsu A, Nomura N, Miyazaki A, Iguchi A. Plasma adiponectin plays an important role in improving insulin resistance with glimepiride in elderly type 2 diabetic subjects. Diabetes Care 2003; 26: 285-289 [PMID: 12547850 DOI: 10.2337/diacare.26.2.285]
- Dominguez H, Storgaard H, Rask-Madsen C, Steffen Hermann T, Ihlemann N, Baunbjerg Nielsen D, Spohr C, Kober L, Vaag A, Torp-Pedersen C. Metabolic and vascular effects of tumor necrosis factor-alpha blockade with etanercept in obese patients with type 2 diabetes. J Vasc Res 2005; 42: 517-525 [PMID: 16155368 DOI: 10.1159/000088261]
- Bellia A, Rizza S, Lombardo MF, Donadel G, Fabiano R, Andreadi K, Quon MJ, Sbraccia P, Federici M, Tesauro M, Cardillo C, Lauro D. Deterioration of glucose homeostasis in type 2 diabetic patients one year after beginning of statins therapy. Atherosclerosis 2012; 223: 197-203 [PMID: 22658255 DOI: 10.1016/j.atherosclerosis.2012.04.015]
- Brooks-Worrell BM, Palmer JP. Attenuation of islet-specific T cell responses is associated with C-peptide improvement in autoimmune type 2 diabetes patients. Clin Exp Immunol 2013; 171: 164-170 [PMID: 23286943 DOI: 10.1111/cei.12012]
- Gagnon C, Daly RM, Carpentier A, Lu ZX, Shore-Lorenti C, Sikaris K, Jean S, Ebeling PR. Effects of combined calcium and vitamin D supplementation on insulin secretion, insulin sensitivity and β-cell function in multi-ethnic vitamin Ddeficient adults at risk for type 2 diabetes: a pilot randomized, placebo-controlled trial. PLoS One 2014; 9: e109607 [PMID: 25299668 DOI: 10.1371/journal.pone.0109607]
- Zakerkish M, Jenabi M, Zaeemzadeh N, Hemmati AA, Neisi N. The Effect of Iranian Propolis on Glucose Metabolism, Lipid Profile, Insulin Resistance, Renal Function and Inflammatory Biomarkers in Patients with Type 2 Diabetes Mellitus: A Randomized Double-Blind Clinical Trial. Sci Rep 2019; 9: 7289 [PMID: 31086222 DOI: 10.1038/s41598-019-43838-8]
- Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, Bitto A. Oxidative Stress: Harms and Benefits for Human Health. Oxid Med Cell Longev 2017; 2017: 8416763 [PMID: 28819546 DOI: 10.1155/2017/84167631
- Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D, Abete P. Oxidative stress, aging, and diseases. Clin Interv Aging 2018; 13: 757-772 [PMID: 29731617 DOI: 10.2147/CIA.S158513]
- Aggarwal V, Tuli HS, Varol A, Thakral F, Yerer MB, Sak K, Varol M, Jain A, Khan MA, Sethi G. Role of Reactive Oxygen Species in Cancer Progression: Molecular Mechanisms and Recent Advancements. Biomolecules 2019; 9 [PMID: 31766246 DOI: 10.3390/biom9110735]
- Murphy MP. How mitochondria produce reactive oxygen species. Biochem J 2009; 417: 1-13 [PMID: 19061483 DOI: 10.1042/BJ20081386
- Diane A, Al-Shukri NA, Bin Abdul Mu-U-Min R, Al-Siddiqi HH. β-cell mitochondria in diabetes mellitus: a missing puzzle piece in the generation of hPSC-derived pancreatic β-cells? J Transl Med 2022; 20: 163 [PMID: 35397560 DOI: 10.1186/s12967-022-03327-51



- 113 Luc K, Schramm-Luc A, Guzik TJ, Mikolajczyk TP. Oxidative stress and inflammatory markers in prediabetes and diabetes. J Physiol Pharmacol 2019; 70 [PMID: 32084643 DOI: 10.26402/jpp.2019.6.01]
- Bigagli E, Lodovici M. Circulating Oxidative Stress Biomarkers in Clinical Studies on Type 2 Diabetes and Its Complications. Oxid Med Cell Longev 2019; 2019: 5953685 [PMID: 31214280 DOI: 10.1155/2019/5953685]
- 115 Hou N, Torii S, Saito N, Hosaka M, Takeuchi T. Reactive oxygen species-mediated pancreatic beta-cell death is regulated by interactions between stress-activated protein kinases, p38 and c-Jun N-terminal kinase, and mitogen-activated protein kinase phosphatases. Endocrinology 2008; 149: 1654-1665 [PMID: 18187551 DOI: 10.1210/en.2007-0988]
- Tarafdar A, Pula G. The Role of NADPH Oxidases and Oxidative Stress in Neurodegenerative Disorders. Int J Mol Sci 2018; 19 [PMID: 30513656 DOI: 10.3390/ijms19123824]
- Yuan H, Zhang X, Huang X, Lu Y, Tang W, Man Y, Wang S, Xi J, Li J. NADPH oxidase 2-derived reactive oxygen species mediate FFAs-induced dysfunction and apoptosis of β-cells via JNK, p38 MAPK and p53 pathways. PLoS One 2010; 5: e15726 [PMID: 21209957 DOI: 10.1371/journal.pone.0015726]
- Balcazar Morales N, Aguilar de Plata C. Role of AKT/mTORC1 pathway in pancreatic β-cell proliferation. Colomb Med (Cali) 2012; 43: 235-243 [PMID: 24893199]
- Pi J, Zhang Q, Andersen ME. Reactive Oxygen Species and Antioxidants in Pancreatic β-Cell Function Yin and Yang. In: Laher I, editor. Systems Biology of Free Radicals and Antioxidants. Berlin, Heidelberg: Springer; 2014
- Ahmed Alfar E, Kirova D, Konantz J, Birke S, Mansfeld J, Ninov N. Distinct Levels of Reactive Oxygen Species Coordinate Metabolic Activity with Beta-cell Mass Plasticity. Sci Rep 2017; 7: 3994 [PMID: 28652605 DOI: 10.1038/s41598-017-03873-9]
- Supale S, Li N, Brun T, Maechler P. Mitochondrial dysfunction in pancreatic β cells. *Trends Endocrinol Metab* 2012; 23: 477-487 [PMID: 22766318 DOI: 10.1016/j.tem.2012.06.002]
- Sireesh D, Dhamodharan U, Ezhilarasi K, Vijay V, Ramkumar KM. Association of NF-E2 Related Factor 2 (Nrf2) and inflammatory cytokines in recent onset Type 2 Diabetes Mellitus. Sci Rep 2018; 8: 5126 [PMID: 29572460 DOI: 10.1038/s41598-018-22913-6]
- Dludla PV, Muller CJ, Joubert E, Louw J, Essop MF, Gabuza KB, Ghoor S, Huisamen B, Johnson R. Aspalathin Protects the Heart against Hyperglycemia-Induced Oxidative Damage by Up-Regulating Nrf2 Expression. Molecules 2017; 22 [PMID: 28098811 DOI: 10.3390/molecules22010129]
- David JA, Rifkin WJ, Rabbani PS, Ceradini DJ. The Nrf2/Keap1/ARE Pathway and Oxidative Stress as a Therapeutic Target in Type II Diabetes Mellitus. J Diabetes Res 2017; 2017: 4826724 [PMID: 28913364 DOI: 10.1155/2017/4826724]
- Baumel-Alterzon S, Katz LS, Brill G, Garcia-Ocaña A, Scott DK. Nrf2: The Master and Captain of Beta Cell Fate. Trends Endocrinol Metab 2021; 32: 7-19 [PMID: 33243626 DOI: 10.1016/j.tem.2020.11.002]
- Constantin A, Dumitrescu M, Nemecz M, Picu A, Smeu B, Guja C, Alexandru N, Georgescu A, Tanko G. Sera of Obese Type 2 Diabetic Patients Undergoing Metabolic Surgery Instead of Conventional Treatment Exert Beneficial Effects on Beta Cell Survival and Function: Results of a Randomized Clinical Study. Obes Surg 2019; 29: 1485-1497 [PMID: 30701387 DOI: 10.1007/s11695-019-03710-01
- Eckhoff DE, Smyth CA, Eckstein C, Bilbao G, Young CJ, Thompson JA, Contreras JL. Suppression of the c-Jun Nterminal kinase pathway by 17beta-estradiol can preserve human islet functional mass from proinflammatory cytokineinduced destruction. Surgery 2003; 134: 169-179 [PMID: 12947315 DOI: 10.1067/msy.2003.219]
- Oscarsson J, Önnerhag K, Risérus U, Sundén M, Johansson L, Jansson PA, Moris L, Nilsson PM, Eriksson JW, Lind L. Effects of free omega-3 carboxylic acids and fenofibrate on liver fat content in patients with hypertriglyceridemia and nonalcoholic fatty liver disease: A double-blind, randomized, placebo-controlled study. J Clin Lipidol 2018; 12: 1390-1403.e4 [PMID: 30197273 DOI: 10.1016/j.jacl.2018.08.003]
- Dandona P, Ghanim H, Abuaysheh S, Green K, Dhindsa S, Makdissi A, Batra M, Kuhadiya ND, Chaudhuri A. Exenatide Increases IL-1RA Concentration and Induces Nrf-2-Keap-1-Regulated Antioxidant Enzymes: Relevance to β-Cell Function. J Clin Endocrinol Metab 2018; 103: 1180-1187 [PMID: 29346597 DOI: 10.1210/jc.2017-02343]
- Eguchi N, Vaziri ND, Dafoe DC, Ichii H. The Role of Oxidative Stress in Pancreatic β Cell Dysfunction in Diabetes. Int J Mol Sci 2021; 22 [PMID: 33546200 DOI: 10.3390/ijms22041509]
- Harmon JS, Bogdani M, Parazzoli SD, Mak SS, Oseid EA, Berghmans M, Leboeuf RC, Robertson RP. beta-Cell-specific overexpression of glutathione peroxidase preserves intranuclear MafA and reverses diabetes in db/db mice. Endocrinology 2009; **150**: 4855-4862 [PMID: 19819955 DOI: 10.1210/en.2009-0708]
- Goyal R, Singhai M, Faizy AF. Glutathione peroxidase activity in obese and nonobese diabetic patients and role of hyperglycemia in oxidative stress. J Midlife Health 2011; 2: 72-76 [PMID: 22408335 DOI: 10.4103/0976-7800.92529]
- Krümmel B, Plötz T, Jörns A, Lenzen S, Mehmeti I. The central role of glutathione peroxidase 4 in the regulation of ferroptosis and its implications for pro-inflammatory cytokine-mediated beta-cell death. Biochim Biophys Acta Mol Basis Dis 2021; 1867: 166114 [PMID: 33662571 DOI: 10.1016/j.bbadis.2021.166114]
- Wang XD, Vatamaniuk MZ, Wang SK, Roneker CA, Simmons RA, Lei XG. Molecular mechanisms for hyperinsulinaemia induced by overproduction of selenium-dependent glutathione peroxidase-1 in mice. Diabetologia 2008; **51**: 1515-1524 [PMID: 18560803 DOI: 10.1007/s00125-008-1055-3]
- Tanaka Y, Tran PO, Harmon J, Robertson RP. A role for glutathione peroxidase in protecting pancreatic beta cells against oxidative stress in a model of glucose toxicity. Proc Natl Acad Sci U S A 2002; 99: 12363-12368 [PMID: 12218186 DOI: 10.1073/pnas.192445199]
- Wang X, Vatamaniuk MZ, Roneker CA, Pepper MP, Hu LG, Simmons RA, Lei XG. Knockouts of SOD1 and GPX1 exert different impacts on murine islet function and pancreatic integrity. Antioxid Redox Signal 2011; 14: 391-401 [PMID: 20586612 DOI: 10.1089/ars.2010.3302]
- 137 Maclaine NJ, Hupp TR. The regulation of p53 by phosphorylation: a model for how distinct signals integrate into the p53 pathway. Aging (Albany NY) 2009; 1: 490-502 [PMID: 20157532 DOI: 10.18632/aging.100047]
- Stancill JS, Broniowska KA, Oleson BJ, Naatz A, Corbett JA. Pancreatic β-cells detoxify H(2)O(2) through the peroxiredoxin/thioredoxin antioxidant system. J Biol Chem 2019; 294: 4843-4853 [PMID: 30659092 DOI:



10.1074/jbc.RA118.006219]

- Kakisaka Y, Nakashima T, Sumida Y, Yoh T, Nakamura H, Yodoi J, Senmaru H. Elevation of serum thioredoxin levels in patients with type 2 diabetes. Horm Metab Res 2002; 34: 160-164 [PMID: 11972307 DOI: 10.1055/s-2002-23201]
- Panse M, Kluth O, Lorza-Gil E, Kaiser G, Mühlbauer E, Schürmann A, Häring HU, Ullrich S, Gerst F. Palmitate and insulin counteract glucose-induced thioredoxin interacting protein (TXNIP) expression in insulin secreting cells via distinct mechanisms. PLoS One 2018; 13: e0198016 [PMID: 29813102 DOI: 10.1371/journal.pone.0198016]
- Lei Z, Chen Y, Wang J, Zhang Y, Shi W, Wang X, Xing D, Li D, Jiao X. Txnip deficiency promotes β-cell proliferation in the HFD-induced obesity mouse model. Endocr Connect 2022; 11 [PMID: 35294398 DOI: 10.1530/EC-21-0641]
- Wondafrash DZ, Nire'a AT, Tafere GG, Desta DM, Berhe DA, Zewdie KA. Thioredoxin-Interacting Protein as a Novel Potential Therapeutic Target in Diabetes Mellitus and Its Underlying Complications. Diabetes Metab Syndr Obes 2020; 13: 43-51 [PMID: 32021350 DOI: 10.2147/DMSO.S232221]
- Gao Y, Chen S, Peng M, Wang Z, Ren L, Mu S, Zheng M. Correlation Between Thioredoxin-Interacting Protein and Nerve Conduction Velocity in Patients With Type 2 Diabetes Mellitus. Front Neurol 2020; 11: 733 [PMID: 32774321 DOI: 10.3389/fneur.2020.00733]
- Xu B, Moritz JT, Epstein PN. Overexpression of catalase provides partial protection to transgenic mouse beta cells. Free Radic Biol Med 1999; 27: 830-837 [PMID: 10515587 DOI: 10.1016/s0891-5849(99)00130-6]
- Kralik PM, Xu B, Epstein PN. Catalase transfection decreases hydrogen peroxide toxicity in a pancreatic beta cell line. Endocr Res 1998; 24: 79-87 [PMID: 9553756 DOI: 10.3109/07435809809031870]
- Wang N, Zhang J, Qin M, Yi W, Yu S, Chen Y, Guan J, Zhang R. Amelioration of streptozotocininduced pancreatic β cell damage by morin: Involvement of the AMPKFOXO3catalase signaling pathway. Int J Mol Med 2018; 41: 1409-1418 [PMID: 29286118 DOI: 10.3892/ijmm.2017.3357]
- Karunakaran U, Park KG. A systematic review of oxidative stress and safety of antioxidants in diabetes: focus on islets and their defense. Diabetes Metab J 2013; 37: 106-112 [PMID: 23641350 DOI: 10.4093/dmj.2013.37.2.106]



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