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REVIEW

Novel nutraceutic therapies for the treatment of metabolic syndrome

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Abstract

Nutraceutic therapies such as berberine, bitter melon,

Gymnema sylvestre, Irvingia gabonensis, resveratrol and ursolic acid have been shown to help control metabolic syndrome (MetS). The effect of berberine on glucose and lipid metabolism, hypertension, obesity and MetS has been evaluated in animal models and humans. Most clinical trials involving bitter melon have been conducted to evaluate its effect on glucose metabolism; nevertheless, some studies have reported favorable effects on lipids and blood pressure although there is little information about its effect on body weight. Gymnema sylvestre helps to decrease body weight and blood sugar levels; however, there is limited information on dyslipidemia and hypertension. Clinical trials of Irvingia gabonensis have shown important effects decreasing glucose and cholesterol concentrations as well decreasing body weight. Resveratrol acts through different mechanisms to decrease blood pressure, lipids, glucose and weight, showing its effects on the population with MetS. Finally, there is evidence of positive effects with ursolic acid in in vitro and in vivo studies on glucose and lipid metabolism and on body weight and visceral fat. Therefore, a review of the beneficial effects and limitations of the above-mentioned nutraceutic therapies is presented.

Key words: Nutraceutics; Metabolic syndrome; Berberine; Bitter melon; *Gymnema silvestre*; *Irvingia gabonensis*; Resveratrol; Ursolic acid

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Core tip: Metabolic syndrome (MetS) is a cluster of endocrine problems including obesity, dysglycemia, dyslipidemia, and hypertension. Unfortunately, there is no unique treatment to control it. Nutraceutic therapies such as berberine, bitter melon, *Gymnema sylvestre*, *Irvingia gabonensis*, resveratrol and ursolic acid have demonstrated some improvement in anthropometric parameters and cardiometabolic risk factors and could



be considered as treatment for patients with MetS. This review attempts to demonstrate the beneficial effects and limitations of some of these novel nutraceutic therapies.

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INTRODUCTION

Metabolic syndrome (MetS) is a cluster of endocrine disturbances including typically obesity, dysglycemia, dyslipidemia, and hypertension, predisposing individuals to increased risk for atherosclerosis, cardiovascular events, and eventually type 2 diabetes mellitus (T2DM)^[1]. However, a number of other parameters that appear to be related to MetS, including nonalcoholic fatty liver disease, should be evaluated in some specific cases to help determine the risk of complications^[2,3]. Prevalence of MetS is increasing significantly and is becoming a worldwide health problem^[4]. Unfortunately, there is no a single treatment to control MetS; frequently, the option is to treat each component separately. Therefore, any substance that helps to control all the characteristic disturbances of MetS must be considered and studied in depth^[5]. Nutraceutic therapies such as berberine, bitter melon, Gymnema sylvestre (G. sylvestre), Irvingia gabonensis (I. gabonensis), resveratrol and ursolic acid, which are currently being studied in our Research Institute, among many therapies, have demonstrated to improve some anthropometric parameters and cardiometabolic risk factors. In this regard, they could be considered as treatment for patients with MetS. The aim of this review is to show the beneficial effects and limitations of some of these novel nutraceutic therapies.

BERBERINE

Berberine is an isoquinoline quaternary alkaloid (or a 5,6 dihydrodibenzo[a,g]quinolizinium derivative) isolated from many medicinal plants such as *Hydrastis canadensis*, *Berberis aristata*, *Coptis chinensis*, *Coptis rhizome*, *Coptis japonica*, *Phellondendron amurense* and *Phellondendron chinense schneid*^[6]. Berberine is traditionally used for its supposed antimicrobial effects and as treatment for diabetes in traditional Chinese, Indian and Middle Eastern folk medicine^[7] and has definite potential as a drug included in a wide spectrum of clinical applications.

During approximately 500 A.D., Hongjing Tao recorded the anti-diabetes activity of Rhizoma coptidis for the first time in a book entitled "Note of Elite

Physicians". In 1988, the hypoglycemic effect of berberine was revealed when berberine was used to treat diarrhea in diabetic patients in China. Since that time, many physicians in China have used berberine as an anti-hyperglycemic agent. There are a substantial number of clinical reports regarding the hypoglycemic action of berberine in Chinese literature reports^[8]. A meta-analysis of berberine reported beneficial effects on blood glucose control in the treatment of T2DM patients similar to those obtained with conventional oral antidiabetic treatments^[9]. One study confirms that administration of berberine (0.5 g three times/d) at the beginning of each major meal was able to reduce fasting blood glucose as well as postprandial blood glucose in adult patients with newly diagnosed T2DM. Glycated hemoglobin A1c (A1C) level was decreased by 2.0% with berberine treatment, which is comparable to that of metformin. In poorly controlled diabetic patients^[8], berberine regulates glucose metabolism possibly through multiple mechanisms and signal pathways such as increasing insulin sensitivity, activating the adenosine monophosphate- (AMP-) activated protein kinase (AMPK) pathway, modulating gut microbiota, inhibiting liver gluconeogenesis, stimulating glycolysis in peripheral tissue cells, promoting intestinal glucagonlike protein-1 secretion, upregulating hepatic lowdensity lipoprotein receptor mRNA expression, and increasing glucose transporter^[10].

The effects of berberine on lipid metabolism have been evaluated in animals and humans. A systematic review and meta-analysis of randomized controlled trials with berberine show that its administration produced a significant reduction in total cholesterol (mean difference -0.61 mmol/L; 95%CI: -0.83 to -0.39), triglycerides (mean difference -0.50 mmol/L; 95%CI: -0.69 to -0.31), and low-density lipoprotein cholesterol (LDL-C) (mean difference -0.65 mmol/L; 95%CI: -0.76 to -0.54) levels, with a remarkable increase in high-density lipoprotein cholesterol (HDL-C) (mean difference 0.05 mmol/L; 95%CI: 0.02 to 0.09)[11]. The lipid-lowering effect of berberine appears to be mainly due to stabilization of hepatic LDL receptor (LDL-R) in an extracellular signalregulated kinase (ERK)-dependent manner and also by increasing transcriptional activity of LDL-R promoter by c-Jun N-terminal kinase (JNK) pathway. Berberine also activates AMPK while blocking the AMPK/ERK pathway, resulting in inhibition of lipid synthesis^[7].

Few reports in the literature affirm that berberine is able to decrease blood pressure in humans; however, vasorelaxant effects of berberine have been observed in different rat models^[7]. Vasodilatator effect of berberine is the result of its action on both endothelium and vascular smooth muscle. Other mechanisms suggested to be involved in the vasorelaxant effect of berberine are angiotensin-converting enzyme (ACE) inhibitor effect and direct release of nitric oxide (NO)/cyclic guanosine monophosphate (cGMP) from rat aortic rings, all-adrenoreceptor antagonistic action in rat and rabbit aorta, potentiation of acetylcholine, activation

of K⁺ channels and inhibition of intracellular calcium release, and blocking of L-type calcium channels^[7]. A recent study showed that berberine could delay the onset and attenuate the severity of hypertension as well as to ameliorate hypertension-induced renal damage in spontaneously hypertensive rats (SHR). Furthermore, berberine could inhibit the activities of the reninangiotensin system and pre-inflammatory cytokines such as interleukin (IL)-6, IL-17 and IL-23, which are involved in the pathophysiology of hypertension^[12].

Several clinical studies have reported the effect of berberine on obesity indicators such as body weight reduction, waist circumference or body mass index (BMI). A study in 116 patients with T2DM and dyslipidemia showed that berberine (1.0 g daily) compared with placebo for 3 mo decreased body weight from 68.7 ± 11.3 to 66.4 ± 11.8 kg^[13]. This effect could be due to an inhibition of adipogenesis that may contribute to the anti-obesity activity of berberine. Since then, it has been shown to suppress adipocyte differentiation and reduce lipid accumulation in (3T3-L1) adipocytes. In cells treated with berberine, expression of several lipogenic genes including peroxisome proliferator-activated receptor gamma (PPAR_γ), enhancer-binding protein alpha (EBPα), sterol regulatory element-binding protein 1 (SREBP-1c), fatty acid synthase, acetyl coenzyme A carboxylase, acyl-CoA synthase, lipoprotein lipase, and cluster of differentiation 36 were all suppressed[8].

The above-mentioned findings show that berberine has excellent potential for prevention and treatment of MetS. A randomized, double-blind, placebo-controlled clinical trial carried out by our research group in 24 patients with a diagnosis of MetS showed that, after berberine administration, patients had a remission of 36% (P=0.037) in the presence of MetS and a significant decrease in waist circumference in females ($106\pm4~\rm cm~vs~103\pm3~\rm cm,~P<0.05$), systolic blood pressure ($123\pm7~\rm mmHg~vs~115\pm9~\rm mmHg,~P<0.01$), and triglycerides ($2.4\pm0.7~\rm mmol/L~vs~1.4\pm0.5~\rm mmol/L~vs~$

There is no effective dose for berberine; however, the therapeutic dosage for most clinical situations is 0.2-1.5 g/d for the treatment of various diseases, especially for T2DM^[7].

Berberine has been shown to be safe in the majority of clinical trials. In a low percentage of patients, berberine has been reported to cause nausea, vomiting, constipation, hypertension, respiratory failure and paresthesias; however, clinical evidence of such adverse effects is not often reported in the literature^[7].

The diverse pharmacological properties exhibited by berberine indicate that the alkaloid has definite potential as a drug in a wide spectrum of clinical applications that include MetS.

BITTER MELON

Bitter melon, also known as Momordica charantia, is a

common tropical vegetable that has also been used in traditional medicine. The plant grows in tropical areas of Asia, Amazon, East Africa, India and the Caribbean^[15]. Approximately 228 different compounds with possible medicinal properties have been isolated from bitter melon fruit, seeds, leaves, pericaps and endosperm. Among these, the most actively studied constituents shown to improve glycemic control include charantin, vicine, momordicin, and polypeptide-p. Polypeptide-p closely resembles bovine insulin with the exception of one extra amino acid, methionine^[16].

Several mechanisms of action have been proposed for its effects on glucose, lipids and blood pressure. Studies have shown that bitter melon inhibits the absorption of glucose by inhibiting α -glucosidase, reduces Na⁺/K⁺-dependent absorption of glucose by the intestinal mucosa and also suppresses disaccharidase activity in the intestine[15,17]. Bitter melon repairs damaged β-cells, stimulates insulin secretion, and enhances insulin sensitivity. Enhancement in insulin sensitivity may be due to multiple factors such as inhibition of protein tyrosine phosphatase 1B (PTP-1B) activity in skeletal muscle, increase in the number and translocation of glucose transporter type 4 (GLUT4) receptors, increase in the rate of phosphorylation of insulin receptor substrate and enhancement in the activity of AMPK. AMPK inhibits cholesterol synthesis in liver by activating 3-hydroxy-3-methylglutaryl-coenzyme reductase. It also stimulates the synthesis and release of thyroid hormones and adiponectin^[17]. Other proposed mechanisms for actions include decreased hepatic gluconeogenesis and increased hepatic glycogen synthesis^[18]. PPARs are nuclear receptors that control lipid and carbohydrate metabolism. These receptors are regarded as important targets for treating MetS. In animal models, bitter melon upregulated PPARy- and PPAR α -mediated pathways^[18].

The hypoglycemic, hypolipidemic and antihypertensive effects of bitter melon have been reported in animal models and clinical trials. Male db/db mice (an animal model of obesity, diabetes, and dyslipidemia) were given sterile tap water as a control or bitter melon daily at a dosage of 150 mg/kg body weight for 5 wk. A1C levels were higher in control mice compared with the bitter melon-treated mice. Additionally, bitter melon reduced PTP-1B activity in skeletal muscle cytosol^[19]. Normal and streptozotocin-induced diabetic rats were fed either with basal diet or a diet containing 10% bitter melon powder. Specific activities of intestinal disaccharidases were significantly increased during diabetes. Bitter melon supplementation in the diet clearly indicated amelioration in the activities of maltase and lactase during diabetes^[20]. The effect of bitter melon at 10% level in the diet was evaluated in streptozotocininduced diabetic rats. Amelioration of approximately 30% in fasting blood glucose was observed^[21]. The aqueous extract powder of the fruit of bitter melon at a dose of 20 mg/kg body weight was also found to reduce fasting blood glucose by 48% in diabetic rats^[22].

To date, most published human clinical trials on bitter melon have focused on blood glucose control. A randomized, double-blind, active-control trial was conducted to assess the efficacy and safety of three doses of bitter melon compared with metformin. Patients were randomized into four groups to receive bitter melon 500 mg/d, 1000 mg/d, and 2000 mg/d or metformin 1000 mg/d. All patients were followed for 4 wk. There was a significant decline in fructosamine in the metformin group (16.8 \pm 40.6 μ mol/L) and the bitter melon 2000 mg/d group (-10.2 \pm 23.3 μ mol/L) $^{[23]}$. After adding bitter melon (800-1600 mg/d) to the current regimens (sulfonylureas and/or metformin) of 42 diabetic patients, fasting plasma glucose was reduced by 26.9 \pm 40.8 mg/dL (P < 0.001) $^{[24]}$.

The effect of bitter melon on blood pressure and lipids has been evaluated in several experimental studies and only one clinical trial has aimed to investigate its effects on MetS. Acute intravenous administration of bitter melon aqueous extract produced dose-dependent, significant reductions in systemic arterial blood pressure and heart rates of normal and hypertensive Dahl saltsensitive rats^[25]. In another study, normal Sprague Dawley rats were divided into control and three test groups. Rats were administered one of three bitter melon preparations in food for 52 d: Chinese or Indian commercial preparations or an extract of bitter melon. All test groups lowered systolic, but not diastolic, blood pressure. Only the group with the extract significantly lowered ACE activity[26]. The methanol extract of bitter melon fruit was administered to diabetic rats for 30 d. A significant decrease in triglyceride and LDL-C and a significant increase in HDL-C were observed^[27]. Bitter melon lowered plasma apolipoprotein B-100 and apolipoprotein B-48 levels in mice fed a high-fat diet and inhibited lipogenesis by downregulating lipogenic gene expression in adipose tissue of diet-induced obese mice^[17].

A preliminary open-label, single arm, uncontrolled supplementation trial was carried out in 42 participants to evaluate the effect of bitter melon supplementation (4.8 g/d for 3 mo) on MetS. Decrease in the incidence of MetS rate at the end of the supplementation period was significantly different from that at baseline (19.0%, P=0.021). The difference remained significant for 1 mo after cessation of supplementation (P=0.047). Except for waist circumference (-2.09 cm, P<0.05), the remaining four risk factors of MetS did not show significant decreases after bitter melon supplementation^[18].

An effective dose for bitter melon has not been established. In animal models the dose range has oscillated from 20 to 150 mg/kg body weight, whereas in clinical trials the dose has varied from 500 mg to 4800 mg per day^[18,19,22,24].

Few side effects have been associated with the use of bitter melon. The most commonly observed adverse effects include mild diarrhea and abdominal pain, which subside after discontinuation. Bitter melon use

is also contraindicated during pregnancy because of its abortifacient properties^[16].

Although the effect of bitter melon on glucose, blood pressure and lipids has been evaluated in several studies with significant results, only one clinical trial has assessed its effect on waist circumference as a primary outcome. Therefore, its effects on body weight remain to be studied in future clinical trials. The multiple mechanisms behind the hypoglycemic, hypolipidemic and antihypertensive effects of bitter melon and the results reported in previous studies provide a firm base for further well-designed randomized controlled trials to evaluate the efficacy of bitter melon on MetS.

G. SYLVESTRE

G. sylvestre is a medicinal plant belonging to the Asclepiadacea family popularly known as "gurmar" in Hindi, which means "sugar destroying". It is a woody climber that grows in tropical forests in India and South East Asia. Its leaves exhibit a broad range of therapeutic effects due to its active ingredients referred to as gymnemic acids. These are a mixture of at least 17 different saponins, acidic glycosides and anthroquinones^[28]. In Indian medicine it is used for its main antidiabetic effects; however, other important metabolic effects have emerged from various studies with potential for treating MetS^[29,30].

G. sylvestre helps to promote weight loss possibly through its ability to reduce cravings for sweets and also controls blood sugar levels. Chewing the leaves, rinsing the mouth with aqueous extracts, or topical application to the tongue selectively and reversibly inhibit the sensation of sweetness. Some investigations have suggested that gymnemic acid binds to the receptor located on the taste buds of the tongue and prevents activation by sugar molecules as well as suppressing sugar uptake, presumably by blocking sucrose receptors by one of its molecules, the gurmarin peptide^[31,32].

G. sylvestre has also been found to be useful against obesity in accordance with recent preclinical studies in a murine model of obesity where the anti-obesity effect of ethanolic or water-soluble fraction of G. sylvestre extract (120 mg/kg, orally for 21 d) was demonstrated in a high-fat diet (HFD)-induced murine model of obesity^[33]. Another study with a standardized ethanolic G. sylvestre extract (200 mg/kg) administered for 28 d resulted in a significant reduction of BMI, organ weight and visceral fat pad weight, among other metabolic parameters^[34]. G. sylvestre has also shown a decrease in body weight without rebound on Otsuka Long-Evans Tokushima Fatty rats^[35]. Decreasing body weight in humans has been demonstrated in studies using G. sylvestre only in combination with various dietary supplements. Therefore, the resulting weight loss cannot be attributed to only G. sylvestre^[36,37].

Researchers have recently established that *G. sylvestre* does not block only sweet receptors on the taste buds of the mouth. It has the same inhibitory

activity on sodium-dependent glucose transporter 1 found in high levels in brush-border membranes of intestinal epithelial cells $^{[38]}$.

The ability of G. sylvestre to lower blood glucose concentrations has been tested as a hypoglycemic agent in combination with insulin in humans, with encouraging results. A preliminary study shows that administration of 200 mg/d of G. sylvestre extract decreased the required insulin dose by 50% and lowered A1C in both type 1 and T2DM. It also increased the number of beta cells in the pancreas and therefore the internal production of insulin. When 400 mg/d of this extract is taken with conventional hypoglycemic drugs such as glyburide or tolbutamide, some patients were able to reduce the dose of the drug or even discontinue its use^[39,40]. *In* vivo studies with oral administration of an extract of G. sylvestre, Om Santal Adivasi (OSA®) (1 g/d for 60 d) induced a significant increase in circulating insulin and C-peptide, which were associated with significant reductions in fasting and postprandial blood glucose. In vitro measurements using isolated human islets of Langerhans demonstrated direct stimulatory effects of OSA® on insulin secretion in human cells, consistent with an in vivo mode of action through enhancing insulin secretion. As a result, it also stabilizes blood sugar and decreases insulin doses. In fact, one patient with a disease duration of 10 years and another patient with a duration of 2 years and who were both using a total of 20 U of insulin a day were able to completely discontinue insulin at this point in the study[41].

Individual chemical components of extract of G. sylvestre have also been shown to be potent and selective antagonists $in\ vitro$ and $in\ vivo$ for the β isoform of liver X receptor $^{[42]}$ in rats in whom G. sylvestre was administered at a dose of 200 mg/kg. Significant reductions in lipid levels and an increase in HDL-C have been reported $^{[43]}$.

Compounds from the leaves of G. sylvestre may act as an endothelial synthase (eNOS) agonist. To further confirm the results, animal studies were performed with G. sylvestre leaves to demonstrate its future usefulness, not only in controlling blood glucose levels in diabetic patients but also to help avoid diabetic complications such as vascular diseases that occur due to decreased availability of NO^[44]. One of the most active constituents of G. sylvestre is deacyl gymnemic acid (DAGA), which is associated with decreases in homeostasis model asesment (HOMA) insulin resistance, a surrogate marker of insulin resistance, suggesting treatment with DAGA at a dose of 200 mg/kg has beneficial effects on improvement in insulin sensitivity[30]. Conversely, in another study, systolic blood pressure was increased in SHR fed a high sucrose diet, but the clinical importance of this finding is unknown^[37].

Clinical studies investigating antidiabetic effects have typically used 200-1000 mg extract daily, standardized to contain 25% gymnemic acids^[30,39,41].

Adverse effects have not been reported in long-term studies in patients with type 1 diabetes^[45]. However,

at high doses, hypoglycemia, weakness, excessive sweating and muscular dystrophy may occur^[46]. On the other hand, due to its lipophilic character, *G. sylvestre* may inhibit intestinal absorption of oleic acid^[47]. However, the United Nations Organizations has reported only one case of toxic hepatitis due to the use of *G. sylvestre*. Additional studies are needed to support its toxic effect^[48]. The above-mentioned evidence supports the possibility of treating MetS with *G. sylvestre*, although more studies are needed.

I. GABONENSIS

I. gabonensis belongs to the family Irvingiaceae. The tree of Irvingia, commonly known as mango bush, wild bush, dikanut or African mango, is native to Central and Occidental Africa^[49]. Both the fruit and seeds of *I. gabonensis* are widely consumed in Africa as part of its gastronomy. It has recently been reported that roots, leaves and an extract of the seeds have medicinal properties.

I. gabonensis has been used for the treatment of diarrhea and to shorten the time of lactation in women. It is also administered for the treatment of colicky pain and dysentery. The tree bark has antibiotic properties and helps to heal dermal wounds produced by burning. It has also been administered for the treatment of toothache.

The use of an extract of *I. gabonensis* seeds has been studied as a source of dietary fiber useful to decrease glucose and cholesterol concentrations in diseases such as diabetes mellitus. Gastric emptying is delayed and absorption of glucose at the intestinal level is reduced, leading to better insulin sensitivity in tissues. This extract has also demonstrated to modify distribution of phospholipids, which lowers the plasma concentrations of total cholesterol and triglycerides^[50]. Although the use of the extract of *I. gabonensis* has increased, no pharmacokinetic data have been reported.

Different studies have been carried out to determine the composition, antioxidant capacity, mechanism of action and effects of *I. gabonensis*. One study that aimed to identify the principal components of an extract of *I. gabonensis* seeds through high-resolution liquid chromatography coupled to mass spectrophotometry demonstrated that its principal components are ellagic acid, mono-, di-, and tri-O-methyl-ellagic and some long-chain glucosides^[51].

In relation to its antioxidant activity, a study was carried out to evaluate the antioxidant capacity of 14 species from Cameroon including I. gabonensis. Using different methanol extracts and two different assays to determine antioxidant capacity - the Folin assay and the ferric reduction potential assay - it was found that I. gabonensis has an elevated antioxidant concentration of approximately 202 mmol/100 $g^{[52]}$.

Another experimental study was carried out with the aim of investigating the effect of an extract of *I. gabonensis* on inhibition of intracellular triglycerides



and the activity of the enzyme glycerol-3-phosphate in adipocytes 3T3-L1 of a murine model. Expression of some proteins typical of adipogenesis, leptin and adiponectin was also studied. Adipocytes were cultivated for 8 d after initiation of their differentiation and were treated with 0-250 $\mu mol/L$ of $\it I.$ gabonensis for 12 and 24 h at 37 $^{\circ}{\rm C}$ in an incubator with humidity at 5%. The results showed that $\it I.$ gabonensis significantly inhibits adipogenesis in adipocytes. This effect appears to be mediated through a decrease in the expression of the PPAR $_{\rm Y}$ ($\it P<0.05$) and leptin ($\it P<0.05$). An increase in adiponectin expression was also found ($\it P<0.05$) $^{[53]}$.

An experimental study carried out in diabetic rats fed for 4 wk with a typical rat diet supplemented with $I.\ gabonensis$ or cellulose found that both types of diets significantly reduced glucose, cholesterol and triglycerides concentrations and also increased HDL-C ($P < 0.05^{[54]}$.

These results agree with results reported in another experimental study where the potential of a seed extract of I. gabonensis was studied to decrease hyperglycemia and hyperlipidemia in a group of diabetic rats administered a diet supplemented with I. gabonensis for 4 wk. The results showed a significant decrease in glucose concentrations, food intake, total cholesterol, triglycerides and LDL-C levels. A significant increase in HDL-C was also reported (P < 0.05)^[49].

A study in which the effect of the administration of a viscous presentation of *I. gabonensis* seeds in diabetic rats was evaluated for 3 wk at a dose of 2 g/kg every 12 h showed that the extract decreased glucose concentrations (P < 0.05), decreased activity of the enzymes pyruvate kinase and lactate dehydrogenase (P < 0.05) and increased the activity of the enzyme glucose-6-phosphatase (P < 0.05) compared with the control group^[55].

Another experimental study was carried out to evaluate the long-term effect of an aqueous extract of the bark of I. gabonensis administered daily to rabbits for 24 wk. At the end of the study, glucose concentration and body weight significantly decreased (P < 0.05)^[56].

Some clinical trials have been conducted to determine if I. gabonensis has an effect on body weight, glucose and lipid concentrations. A double-blind clinical trial carried out in 40 obese subjects who received I. gabonensis or placebo at a dose of 1.05 g three times/d for 1 mo showed that the administration of the extract of I. gabonensis decreased on average 5.25 kg of body weight (P < 0.001). The subjects also showed a significant decrease of total cholesterol, LDL-C and triglycerides concentrations and increased their HDL-C^[57].

Another clinical trial was conducted in 102 overweight or obese subjects who were randomized into two groups: One group who received 150 mg of *I. gabonensis* 30 min prior to breakfast and dinner and the other received placebo at the same dose for 10 wk. The results showed a significant diminution on body weight, fat mass and waist circumference.

Significant differences were also found in plasma concentrations of total cholesterol, LDL-C, glucose, C-reactive protein, leptin and a significant increase was shown for adiponectin and HDL-C concentrations in the I. gabonensis group vs placebo^[58].

An approved dose has not yet been established for its use. A systematic review of three randomized controlled trials that evaluated the efficacy of I. gabonensis supplementation in the management of overweight and obesity found that the daily dosages differed from approximately 200 mg to approximately 3150 mg^[59].

Adverse events reported in some clinical trials regarding the use of I. gabonensis are headache, dry mouth, diarrhea, sleep disturbances, and constipation^[60]. Acute toxicity studies have not reported any deaths after the 7-d administration of I. gabonenis at a dose of 1600 mg/kg in rats^[59].

The different studies performed either in animal models or as clinical trials suggest that the administration of *I. gabonensis* may be a promising option for the prevention and treatment of MetS.

RESVERATROL

As a chemical compound, resveratrol (3,5,4-trihydro-xystilbene) has been described since the 1940s when it was isolated for the first time from the roots of a white hellebore. Years later, it was extracted from the dried roots of a plant called *Polygonum cuspidatum*, which is often used in traditional Chinese medicine^[61].

Today it is known that resveratrol can be found in different quantities in > 70 plants and is also present in some foods and beverages such as nuts, berries, grapes, peanuts and their derivatives such as red wine. The quantity of resveratrol depends of different factors such crop type, geographical origin, and climate^[62].

In plants, resveratrol acts as a phytoalexin, a toxic compound produced by plants as a defense mechanism in response to the presence of pests and other stressful situations such as climate.

Resveratrol can be found in two different isomeric forms: *cis* and *trans*, the *cis* form being the more common used form due to its pharmacological properties^[63].

Despite the multiple therapeutic effects attributed to resveratrol, its pharmacokinetic characteristics are not favorable because of its poor bioavailability. It is rapidly metabolized and excreted $^{[64]}$.

There is no evidence of the existence of specific receptors for resveratrol. However, resveratrol seems to accumulate in different tissues, mainly related with its absorption and metabolism such as duodenum, colon, liver and kidney^[65-67].

Although most of the studies carried out with resveratrol are in regard to its cardioprotective effect, there is evidence that resveratrol has other pharmacological properties in a wide range of chronic diseases such as cancer, T2DM, and degenerative diseases such as Alzheimer's as well as having antithrombotic,



antiosteoporotic and antimicrobial effects^[63,67]. Resveratrol acts through different mechanisms. Similar to other polyphenols, resveratrol has an important antioxidant activity and interacts with different receptors, kinases and enzymes^[68]. Some studies carried out in *in vivo* models reveal that resveratrol activates sirtuin 1 (SIRT1) and AMPK, both molecules implicated in metabolism regulation; therefore, resveratrol could be a new alternative for the prevention and treatment of MetS^[69].

Activation of SIRT1 by resveratrol decreases the activity of PPAR γ and therefore adipogenesis, which decreases the number of adipocytes and thus obesity. Resveratrol also increases phosphorylation of the coactivator type 1α of PPAR (PGC- 1α) and cyclic adenine monophosphate (cAMP), which increases lipolysis. Resveratrol also enhances the activity of AMPK, which decreases the activity of acetyl CoA carboxylase by its phosphorylation, resulting in a decrease of lipogenesis that contributes to the control of obesity and dyslipidemia. Increase of the activity of AMPK stimulates phosphorylation of the myocyte enhancer factor 2 (MEF2), which results in a higher expression of GLUT4 and therefore a lower resistance to insulin and a diminution of glucose.

Finally, resveratrol increases the activity of endothelial eNOS and therefore the NO concentrations, which contributes to the vasodilation and indirectly to decreased blood pressure^[70]. All these effects have been confirmed in different studies, both in animal models and in clinical trials.

A clinical trial was conducted in 11 males with obesity but without any other metabolic alteration. Patients received resveratrol or homologated placebo at a dose of 150 mg/d for 30 d. Results show that resveratrol activated AMPK at the muscular level and increases levels of SIRT1 and PGC-1 α , resulting in higher lipolysis of adipose tissue. A decrease in glucose, insulin and HOMA index was also demonstrated [71]. A meta-analysis carried out with 11 clinical trials found that resveratrol administrated at different doses for at least 2 wk in patients with diabetes decreases fasting glucose, insulin, A1C and insulin resistance evaluated through HOMA index, but this meta-analysis did not find any differences in patients without diabetes [72].

Although the information obtained about the effects of resveratrol on cholesterol and triglycerides concentrations is inconclusive, some studies performed in animal models with MetS have shown that resveratrol at different doses reduces atherosclerotic plaque formation, total cholesterol and triglycerides^[73,61]. Clinical trials reported in obese patients have not found any significant differences in lipid profile after resveratrol administration^[71,74].

Our research group^[75] conducted a randomized, double-blind, placebo-controlled clinical trial in 24 patients with a diagnosis of MetS in accordance with the International Diabetes Federation modified criteria. Resveratrol or homologated placebo was administrated for 90 d at a dose of 500 mg three times per day. After

resveratrol administration, significant differences were found in total weight (94.4 \pm 13.2 kg vs 90.5 \pm 12.3 kg, P = 0.007), BMI (35.6 \pm 3.2 kg/m² vs 34.3 \pm 3.0 kg/m², P = 0.006), fat mass (41.2 \pm 7.9 kg vs 38.8 \pm 6.0 kg, P = 0.001), and waist circumference (109 \pm 9 cm vs 105 \pm 10 cm, P = 0.004). There were also significant differences in area under the curve (AUC) of insulin (48418 \pm 22707 pmol/L vs 26473 \pm 8273 pmol/L, P = 0.003) and total insulin secretion evaluated through insulinogenic index (0.48 \pm 0.22 pmol/L vs 0.28 \pm 0.08 pmol/L, P = 0.004).

An approved dose has not yet been established for its use. In a meta-analysis where the effect of resveratrol on glucose control and insulin sensitivity was evaluated, a dose range from 8 to 1500 mg/dL was found^[72].

Some adverse effects reported due to the use of resveratrol are headache, abdominal pain and general malaise^[75]. At high doses (2000 mg twice daily for 1 wk), a clinical trial reported statistically, but not clinically significant, increased serum bilirubin and potassium concentrations^[76]. Daily dosing of 100 mg for 4 wk did not change these values^[77].

These results lead to the conclusion that resveratrol could be an option for the treatment of MetS due to the decrease of obesity and by controlling the hypersecretion of insulin characteristic of this group of patients.

URSOLIC ACID

Ursolic acid is a pentacyclic triterpene carboxylic acid present as a free acid or as an aglycone part of saponins^[78] and can be obtained naturally or synthetically^[79]. It is also known as urson, prunol, micromerol or malol^[80]. This compound was considered inactive; however, in recent years interest has been sparked due to the multiple and varied effects of ursolic acid^[79,81]. Evidence for this substance appears promising for the treatment of MetS.

The main sources of ursolic acid include components of certain fruits, herbs and plants. Ursolic acid is found in apple peel, cranberry juice and grape skin. It is also found in some common spices like rosemary, thyme and oregano and has been identified in Ayurvedic herbs such as Holy Basil, some traditional Chinese medicinal herbs including Jujuba zizyphus, and in yerba mate and sage. Ursolic acid also is found in some herbs that have attributed antidiabetic effects and is found in small amounts in the leaves of some plants^[82,83].

Ursolic acid is formed by 30 carbons distributed in five rings of six carbons and has an hydroxyl group at carbon 3, a carboxyl group at carbon 28 and a double bond at carbon 12 and 13. Its chemical formula is $C_{30}H_{48}O_3^{[84]}$. Some structurally related compounds of ursolic acid include its isomer, oleanolic acid, in addition to corosolic, maslinic, latanolic, pomolic, camarinic and pomolic acids^[85]. These compounds share common characteristics of pentacyclic triterpenoids with

apparently similar effects, although differing from each other in strength^[85].

Physicochemical properties of ursolic acid give it great stability. Ursolic acid has a molecular weight of 456.70032 g/mol. Its melting point is 269-271 $^{\circ}\mathrm{C}$. It has an optical activity of +34° at a concentration of 0.20 g/100 mL in methanol and a molar solubility in pure water at pH 7 and 25 $^{\circ}\mathrm{C}$ of 1.11 \times 10⁻⁵ mg/L^[80,84].

Evidence demonstrates positive effects *in vitro* and *in vivo* through various mechanisms in glucose and lipid metabolism as well as in body weight and visceral fat usually altered in MetS.

Ursolic acid inhibits the enzyme PTP1B, promoting phosphorylation of the insulin receptor *in vitro*, thereby stimulating glucose uptake^[86,87]. PTPB1 is an enzyme associated with the endoplasmic reticulum and plays a key role in signaling metabolic pathways that interacts and dephosphorylates insulin receptor and leptin, causing downregulation signaling of both receptors in modulating the mitogenic actions of insulin^[88].

Translocation of GLUT4 is increased by ursolic acid as part of the action on the insulin receptor and manages to improve glucose uptake. GLUT4 is the principal glucose transporter protein and thus plays a key role in regulating whole body glucose homeostasis^[88].

Ursolic acid appears to inhibit the α -amylase enzyme, an enzyme that hydrolyzes α -links of large polysaccharides such as starch and glycogen to yield glucose and maltose. Inhibition of α -amylase has been shown to lower blood glucose levels due to lowering the breakdown and absorption of starch^[89].

Ursolic acid reduces the activity of aldose reductase and sorbitol dehydrogenase^[90,91]. These enzymes catalyze the reduction of hexoses. In the presence of hyperglycemia, aldose reductase converts glucose to sorbitol. The latter is metabolized to fructose by sorbitol dehydrogenase. During this process, the production of sorbitol and fructose occurs. Reduced nicotinamide adenine dinucleotide phosphate is decreased and nicotinamide adenine dinucleotide phosphate is increased^[91]. Sorbitol increases intracellular osmotic pressure and damages tissues by cell edema; fructose causes protein fructosylation^[90].

The increase in the glyoxalase system produced by ursolic acid represents the decrease of cytotoxicity and chronic complications caused by methylglyoxal, a toxic metabolite produced as a by-product of metabolism. This detoxification reaction is carried out by the glyoxalase system^[92].

Administration of ursolic acid was associated with decreased adipocyte differentiation^[93]. Adipocytes synthesize and release a wide variety of peptide and non-peptide substances and also store and mobilize triglycerides, cholesterol and retinoids. Lipid-laden adipocytes can be emptied and extended, forming cells that resemble their predecessors not only in appearance but also for its potential for multiplication. This change reflects fully differentiated adipocyte regression to an earlier or less mature, but complete, stage^[93].

Overregulation of the c-Cbl associated protein (CAP) was observed in adipocytes treated with ursolic acid^[94]. CAP is expressed only in insulin-sensitive tissues (adipose, liver and muscle). Increase in transcription of CAP is directly related to greater sensitivity to insulin in adipocytes. It is postulated that CAP would facilitate phosphorylation of c-Cbl by the insulin receptor, allowing the union of c-Cbl to the insulin-dependent tyrosine kinase. The relationship of CAP is an example of a direct molecular link between PPAR γ sensitivity and insulin in adipose tissue^[94].

Through the activation of protein kinase A, ursolic acid appears to increase lipolysis *in vitro* as well as to decrease hormone-sensitive lipase and perilipin activity [93]. Lipolysis favors the production of energy from fatty acids into the mitochondria, enabling the generation of free fatty acids from triglycerides stored in adipocytes of white adipose tissue. As a result, there is an activation of fatty acids as well as a translocation to the mitochondria from tissues such as muscle and brown adipose tissue. As a final result, the production of energy occurs from β -oxidation of fatty acids in mitochondria and in some cases in the peroxisome [93].

There is no established dose for ursolic acid. Animal studies have found benefits with ursolic acid at 0.05%-0.2% of the diet^[86-93], which is about 10-40 mg/kg based on their weight and food intake. In clinical trials, a 150-mg dose one to three times a day has been used, providing a maximum of 450 mg and revealing some biological activity.

No adverse effects have been associated with ursolic acid in humans. However, studies in animals have reported that ursolic acid at very high doses resulted in a decrease of sperm motility, cell death and DNA damage^[95]. Due to the beneficial effects of ursolic acid on several components of the MetS, its clinical administration should be further studied.

CONCLUSION

Nutraceutic therapies such as berberine, bitter melon, *G. sylvestre, I. gabonensis,* resveratrol and ursolic acid have demonstrated substantial scientific information regarding their safety and beneficial effects to be comprehensively considered for treating patients with MetS. Berberine and resveratrol, which already have been studied in patients with MetS, have demonstrated valuable results. For the remainder of the nutraceutics presented in this review, it may be necessary to perform more in-depth studies to be clinically recommended.

REFERENCES

- Moreira GC, Cipullo JP, Ciorlia LA, Cesarino CB, Vilela-Martin JF. Prevalence of metabolic syndrome: association with risk factors and cardiovascular complications in an urban population. *PLoS One* 2014; 9: e105056 [PMID: 25180496 DOI: 10.1371/journal. pone.0105056]
- 2 Alberti KG, Zimmet P, Shaw J. Metabolic syndrome--a new worldwide definition. A Consensus Statement from the International



- Diabetes Federation. *Diabet Med* 2006; **23**: 469-480 [PMID: 16681555 DOI: 10.1111/j.1464-5491.2006.01858.x]
- Tarantino G, Finelli C. What about non-alcoholic fatty liver disease as a new criterion to define metabolic syndrome? World J Gastroenterol 2013; 19: 3375-3384 [PMID: 23801829 DOI: 10.3748/wjg.v19.i22.3375]
- 4 González-Ortiz M, Martínez-Abundis E, Jacques-Camarena O, Hernández-González SO, Valera-González IG, Ramos-Zavala MG. Prevalence of metabolic syndrome in adults with excess of adiposity: comparison of the Adult Treatment Panel III criteria with the International Diabetes Federation definition. *Acta Diabetol* 2006; 43: 84-86 [PMID: 17143786 DOI: 10.1007/s00592-006-0218-2]
- 5 González-Ortiz M, Martínez-Abundis E, Robles-Cervantes JA, Ramos-Zavala MG. Effect of exenatide on fat deposition and a metabolic profile in patients with metabolic syndrome. *Metab Syndr Relat Disord* 2011; 9: 31-34 [PMID: 20874425 DOI: 10.1089/met.2010.0025]
- Tillhon M, Guamán Ortiz LM, Lombardi P, Scovassi AI. Berberine: new perspectives for old remedies. *Biochem Pharmacol* 2012; 84: 1260-1267 [PMID: 22842630 DOI: 10.1016/j.bcp.2012.07.018]
- 7 Derosa G, Maffioli P, Cicero AF. Berberine on metabolic and cardiovascular risk factors: an analysis from preclinical evidences to clinical trials. *Expert Opin Biol Ther* 2012; 12: 1113-1124 [PMID: 22780092 DOI: 10.1517/14712598.2012.704014]
- 8 Yin J, Zhang H, Ye J. Traditional chinese medicine in treatment of metabolic syndrome. *Endocr Metab Immune Disord Drug Targets* 2008; 8: 99-111 [PMID: 18537696 DOI: 10.2174/18715300878453 4330]
- 9 Dong H, Wang N, Zhao L, Lu F. Berberine in the treatment of type 2 diabetes mellitus: a systemic review and meta-analysis. *Evid Based Complement Alternat Med* 2012; 2012: 591654 [PMID: 23118793 DOI: 10.1155/2012/591654]
- Pang B, Zhao LH, Zhou Q, Zhao TY, Wang H, Gu CJ, Tong XL. Application of berberine on treating type 2 diabetes mellitus. Int J Endocrinol 2015; 2015: 905749 [PMID: 25861268 DOI: 10.1155/2015/905749]
- 11 Dong H, Zhao Y, Zhao L, Lu F. The effects of berberine on blood lipids: a systemic review and meta-analysis of randomized controlled trials. *Planta Med* 2013; 79: 437-446 [PMID: 23512497 DOI: 10.1055/s-0032-1328321]
- 12 Guo Z, Sun H, Zhang H, Zhang Y. Anti-hypertensive and renoprotective effects of berberine in spontaneously hypertensive rats. Clin Exp Hypertens 2015; 37: 332-339 [PMID: 25867076 DOI: 10.3109/10641963.2014.972560]
- 13 Zhang Y, Li X, Zou D, Liu W, Yang J, Zhu N, Huo L, Wang M, Hong J, Wu P, Ren G, Ning G. Treatment of type 2 diabetes and dyslipidemia with the natural plant alkaloid berberine. *J Clin Endocrinol Metab* 2008; 93: 2559-2565 [PMID: 18397984 DOI: 10.1210/jc.2007-2404]
- Pérez-Rubio KG, González-Ortiz M, Martínez-Abundis E, Robles-Cervantes JA, Espinel-Bermúdez MC. Effect of berberine administration on metabolic syndrome, insulin sensitivity, and insulin secretion. *Metab Syndr Relat Disord* 2013; 11: 366-369 [PMID: 23808999 DOI: 10.1089/met.2012.0183]
- 15 Grover JK, Yadav SP. Pharmacological actions and potential uses of Momordica charantia: a review. *J Ethnopharmacol* 2004; 93: 123-132 [PMID: 15182917 DOI: 10.1016/j.jep.2004.03.035]
- Efird JT, Choi YM, Davies SW, Mehra S, Anderson EJ, Katunga LA. Potential for improved glycemic control with dietary Momordica charantia in patients with insulin resistance and prediabetes. *Int J Environ Res Public Health* 2014; 11: 2328-2345 [PMID: 24566057 DOI: 10.3390/ijerph110202328]
- 17 Chaturvedi P. Antidiabetic potentials of Momordica charantia: multiple mechanisms behind the effects. *J Med Food* 2012; 15: 101-107 [PMID: 22191631 DOI: 10.1089/jmf.2010.0258]
- 18 **Tsai CH**, Chen EC, Tsay HS, Huang CJ. Wild bitter gourd improves metabolic syndrome: a preliminary dietary supplementation trial. *Nutr J* 2012; **11**: 4 [PMID: 22243626 DOI: 10.1186/1475-2891-11-4]
- 19 Klomann SD, Mueller AS, Pallauf J, Krawinkel MB. Antidiabetic

- effects of bitter gourd extracts in insulin-resistant db/db mice. *Br J Nutr* 2010; **104**: 1613-1620 [PMID: 20615270 DOI: 10.1017/S0007114510002680]
- 20 Kumar Shetty A, Suresh Kumar G, Veerayya Salimath P. Bitter gourd (Momordica charantia) modulates activities of intestinal and renal disaccharidases in streptozotocin-induced diabetic rats. *Mol Nutr Food Res* 2005; 49: 791-796 [PMID: 16007724 DOI: 10.1002/ mnfr.200500035]
- 21 Shetty AK, Kumar GS, Sambaiah K, Salimath PV. Effect of bitter gourd (Momordica charantia) on glycaemic status in streptozotocin induced diabetic rats. *Plant Foods Hum Nutr* 2005; 60: 109-112 [PMID: 16187012 DOI: 10.1007/s11130-005-6837-x]
- Virdi J, Sivakami S, Shahani S, Suthar AC, Banavalikar MM, Biyani MK. Antihyperglycemic effects of three extracts from Momordica charantia. *J Ethnopharmacol* 2003; 88: 107-111 [PMID: 12902059 DOI: 10.1016/S0378-8741(03)00184-3]
- Fuangchan A, Sonthisombat P, Seubnukarn T, Chanouan R, Chotchaisuwat P, Sirigulsatien V, Ingkaninan K, Plianbangchang P, Haines ST. Hypoglycemic effect of bitter melon compared with metformin in newly diagnosed type 2 diabetes patients. *J Ethnopharmacol* 2011; 134: 422-428 [PMID: 21211558 DOI: 10.1016/j.jep.2010.12.045]
- 24 Fuangchan A, Seubnukarn T, Jungpattanawadee D, Sonthisombat P, Ingkaninan K, Plianbangchang P, Haines ST. Retrospective study on the use of bitter melon for type 2 diabetes at Dansai Crown Prince Hospital, Thailand. *Srinagarind Med J* 2009; 24: 332-338
- 25 Ojewole JA, Adewole SO, Olayiwola G. Hypoglycaemic and hypotensive effects of Momordica charantia Linn (Cucurbitaceae) whole-plant aqueous extract in rats. *Cardiovasc J S Afr* 2006; 17: 227-232 [PMID: 17117226]
- 26 Clouatre DL, Rao SN, Preuss HG. Bitter melon extracts in diabetic and normal rats favorably influence blood glucose and blood pressure regulation. *J Med Food* 2011; 14: 1496-1504 [PMID: 21861717 DOI: 10.1089/jmf.2010.0276]
- 27 Chaturvedi P, George S, Milinganyo M, Tripathi YB. Effect of Momordica charantia on lipid profile and oral glucose tolerance in diabetic rats. *Phytother Res* 2004; 18: 954-956 [PMID: 15597317 DOI: 10.1002/ptr.1589]
- Sinsheimer JE, Rao GS. Constituents from Gymnema sylvestre leaves. VI. Acylated genins of the gymnemic acids--isolated and preliminary characterization. *J Pharm Sci* 1970; 59: 629-632 [PMID: 5450284 DOI: 10.1002/jps.2600590511]
- Chakraborty D, Ghosh S, Bishayee K, Mukherjee A, Sikdar S, Khuda-Bukhsh AR. Antihyperglycemic drug Gymnema sylvestre also shows anticancer potentials in human melanoma A375 cells via reactive oxygen species generation and mitochondria-dependent caspase pathway. *Integr Cancer Ther* 2013; 12: 433-441 [PMID: 23615751 DOI: 10.1177/1534735413485419]
- 30 Bhansali S, Shafiq N, Pandhi P, Singh AP, Singh I, Singh PK, Sharma S, Malhotra S. Effect of a deacyl gymnemic acid on glucose homeostasis & amp; metabolic parameters in a rat model of metabolic syndrome. *Indian J Med Res* 2013; 137: 1174-1179 [PMID: 23852298 DOI: 10.1016/0378-8741(83)90021-1]
- 31 Schroeder JA, Flannery-Schroeder E. Use of the Herb Gymnema sylvestre to Illustrate the Principles of Gustatory Sensation: An Undergraduate Neuroscience Laboratory Exercise. J Undergrad Neurosci Educ 2005; 3: A59-A62 [PMID: 23493970]
- Warren RP, Warren RM, Weninger MG. Inhibition of the sweet taste by Gymnema sylvestre. *Nature* 1969; 223: 94-95 [PMID: 5792442 DOI: 10.1038/223094a0]
- 33 Kumar V, Bhandari U, Tripathi CD, Khanna G. Anti-obesity effect of Gymnema sylvestre extract on high fat diet-induced obesity in Wistar rats. *Drug Res* (Stuttg) 2013; 63: 625-632 [PMID: 23842942 DOI: 10.1055/s-0033-1349852]
- 34 Kumar V, Bhandari U, Tripathi CD, Khanna G. Evaluation of antiobesity and cardioprotective effect of Gymnema sylvestre extract in murine model. *Indian J Pharmacol* 2012; 44: 607-613 [PMID: 23112423 DOI: 10.4103/0253-7613.100387]
- Nielubowicz J. Magic of the physician's word. Pol Tyg Lek 1991;
 46: 679-684 [PMID: 1669131 DOI: 10.1007/s11010-005-9049-7]



- 36 Woodgate DE, Conquer JA. Effects of a stimulant-free dietary supplement on body weight and fat loss in obese adults: a sixweek exploratory study. Curr Ther Res Clin Exp 2003; 64: 248-262 [PMID: 24944372 DOI: 10.1016/S0011-393X(03)00058-4]
- 37 Preuss HG, Jarrell ST, Scheckenbach R, Lieberman S, Anderson RA. Comparative effects of chromium, vanadium and gymnema sylvestre on sugar-induced blood pressure elevations in SHR. *J Am Coll Nutr* 1998; 17: 116-123 [PMID: 9550454 DOI: 10.1080/07315 724.1998.10718736]
- 38 Wang Y, Dawid C, Kottra G, Daniel H, Hofmann T. Gymnemic acids inhibit sodium-dependent glucose transporter 1. *J Agric Food Chem* 2014; 62: 5925-5931 [PMID: 24856809 DOI: 10.1021/jf501766u]
- 39 Baskaran K, Kizar Ahamath B, Radha Shanmugasundaram K, Shanmugasundaram ER. Antidiabetic effect of a leaf extract from Gymnema sylvestre in non-insulin-dependent diabetes mellitus patients. *J Ethnopharmacol* 1990; 30: 295-300 [PMID: 2259217 DOI: 10.1016/0378-8741(90)90108-6]
- 40 Shanmugasundaram ER, Rajeswari G, Baskaran K, Rajesh Kumar BR, Radha Shanmugasundaram K, Kizar Ahmath B. Use of Gymnema sylvestre leaf extract in the control of blood glucose in insulin-dependent diabetes mellitus. *J Ethnopharmacol* 1990; 30: 281-294 [PMID: 2259216 DOI: 10.1016/0378-8741(90)90107-5]
- 41 Al-Romaiyan A, Liu B, Asare-Anane H, Maity CR, Chatterjee SK, Koley N, Biswas T, Chatterji AK, Huang GC, Amiel SA, Persaud SJ, Jones PM. A novel Gymnema sylvestre extract stimulates insulin secretion from human islets in vivo and in vitro. *Phytother Res* 2010; 24: 1370-1376 [PMID: 20812281 DOI: 10.1002/ptr.3125]
- 42 Renga B, Festa C, De Marino S, Di Micco S, D'Auria MV, Bifulco G, Fiorucci S, Zampella A. Molecular decodification of gymnemic acids from Gymnema sylvestre. Discovery of a new class of liver X receptor antagonists. *Steroids* 2015; 96: 121-131 [PMID: 25668616 DOI: 10.1016/j.steroids.2015.01.024]
- 43 Rachh PR, Rachh MR, Ghadiya NR, Modi DC, Modi KP, Patel NM, Rupareliya MT. Antihyperlipidemic Activity of Gymenma sylvestre R. Br. Leaf Extract on Rats Fed with High Cholesterol Diet. *Inter J Pharmacol* 2010; 6: 138-141 [DOI: 10.3923/ijp.2010.138.141]
- 44 Khan KA, Shabnam D, Mohammed AS. Molecular docking and preclinical studies of Gymnema sylvestre on endothelial nitric oxide synthase (enos) in Type-2 diabetes related complications. *J Young Pharm* 2014; 6: 25-32 [DOI: 10.5530/jyp.2014.4.5]
- 45 Thakur GS, Sharma R, Sanodiya BS, Pandey M, Prasad GBKS, Bisen PS. Gymnema sylvestre: An Alternative Therapeutic Agent for Management of Diabetes. *JAPS* 2012; 2: 1-6 [DOI: 10.7324/ JAPS.2012.21201]
- 46 Tiwari P, Mishra BN, Sangwan NS. Phytochemical and pharmacological properties of Gymnema sylvestre: an important medicinal plant. *Biomed Res Int* 2014; 2014: 1-18 [DOI: 10.1155/2014/830285]
- 47 El Shafey AAM, El-Ezabi MM, Seleim MME, Ouda HHM, Ibrahim DS. Effect of Gymnema sylvestre R. Br. leaves extract on certain physiological parameters of diabetic rats. *J King Saud Univ* 2013; 25: 135-141 [DOI: 10.1016/j.jksus.2012.11.001]
- 48 Song C, Yang Z, Zhong M, Chen Z. Sericin protects against diabetes-induced injuries in sciatic nerve and related nerve cells. *Neural Regen Res* 2013; 8: 506-513 [PMID: 25206693 DOI: 10.3969/j.issn.1673-5374.2013.06.003]
- 49 Dzeufiet PD, Ngeutse FD, Dimo T, Tédong L, Ngueguim TF Tchamadeu MC, Nkouambou NC, Sokeng SD, Kamtchouing P. Hypoglycemic and hypolipidemic effects of Irvingia gabonensis in diabetic rats. *Pharmacologyonline* 2009; 2: 957-962
- 50 Ainge L, Brown N. Bush Mango (Irvingia gabonensis and I. wombolu). In: Clark LE, Sunderland TC. The Key Non-Timber Forest Products of Central Africa: State of the Knowledge. USAID, Bureau for Arica, Office of Sustainable Development, 2004: 15-35
- 51 Sun J, Chen P. Ultra high-performance liquid chromatography with high-resolution mass spectrometry analysis of African mango (Irvingia gabonensis) seeds, extract, and related dietary supplements. *J Agric Food Chem* 2012; 60: 8703-8709 [PMID: 22880691 DOI: 10.1021/jf301703u]

- 52 Agbor GA, Oben JE, Ngogang JY, Xinxing C, Vinson JA. Antioxidant capacity of some herbs/spices from cameroon: a comparative study of two methods. *J Agric Food Chem* 2005; 53: 6819-6824 [PMID: 16104805 DOI: 10.1021/jf050445c]
- Oben JE, Ngondi JL, Blum K. Inhibition of Irvingia gabonensis seed extract (OB131) on adipogenesis as mediated via down regulation of the PPARgamma and leptin genes and up-regulation of the adiponectin gene. *Lipids Health Dis* 2008; 7: 44 [PMID: 19014517 DOI: 10.1186/1476-511X-7-44]
- Omoruyi F, Adamson I. Effect of supplements of dikanut (Irvingia gabonensis) and cellulose on plasma lipids and composition of hepatic phospholipids in streptozotocin-induced diabetic rats. *Nutrition Research* 1994; 14: 537-544 [DOI: 10.1016/ S0271-5317(05)80217-9]
- Ozula RI, Eriyamremu GE, Okene EO, Ochei U. Hypoglycaemic effects of viscous preparation of Irvingia gabonensis (Dikanut) seeds in Streptozotocin-Induced Diabetic Wistar Rats. *Journal of Herbs, Spices & Medical Plants* 2014; 12: 1-9 [DOI: 10.1300/J044v12n04 01]
- 56 Omonkhua AA, Onoagbe IO. Effects of long-term oral administration of aqueous extracts of Irvingia gabonensis bark on blood glucose and liver profile of normal rabbits. *Journal of Medicinal Plants Research* 2012; 6: 2581-2589 [DOI: 10.5897/ JMPR11.561]
- 57 Ngondi JL, Oben JE, Minka SR. The effect of Irvingia gabonensis seeds on body weight and blood lipids of obese subjects in Cameroon. *Lipids Health Dis* 2005; 4: 12 [PMID: 15916709 DOI: 10.1186/1476-511X-4-12]
- Ngondi JL, Etoundi BC, Nyangono CB, Mbofung CM, Oben JE. IGOB131, a novel seed extract of the West African plant Irvingia gabonensis, significantly reduces body weight and improves metabolic parameters in overweight humans in a randomized double-blind placebo controlled investigation. *Lipids Health Dis* 2009; 8: 7 [PMID: 19254366 DOI: 10.1186/1476-511X-8-7]
- 59 Onakpoya I, Davies L, Posadzki P, Ernst E. The efficacy of Irvingia gabonensis supplementation in the management of overweight and obesity: a systematic review of randomized controlled trials. *J Diet Suppl* 2013; 10: 29-38 [PMID: 23419021]
- 60 African Mango. Accessed 2014 December 15. Available from: URL: http://www.drugs.com/npp/african-mango.html
- 61 Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. *Nat Rev Drug Discov* 2006; 5: 493-506 [PMID: 16732220 DOI: 10.1038/nrd2060]
- 62 Zhang J. Resveratrol inhibits insulin responses in a SirT1-independent pathway. *Biochem J* 2006; 397: 519-527 [PMID: 16626303 DOI: 10.1042/BJ20050977]
- 63 Millán-Parrila F, Serrano-San Miguel G. Resveratrol nuevos retos en el tratamiento anti-envejecimiento. Salud Estética 2008; 5: 1-5
- 64 Kaldas MI, Walle UK, Walle T. Resveratrol transport and metabolism by human intestinal Caco-2 cells. *J Pharm Pharmacol* 2003; 55: 307-312 [PMID: 12724035 DOI: 10.1211/002235702612]
- 65 Walle T. Bioavailability of resveratrol. *Ann N Y Acad Sci* 2011; 1215: 9-15 [PMID: 21261636 DOI: 10.1111/j.1749-6632.2010.05842.x]
- 66 Timmers S, Auwerx J, Schrauwen P. The journey of resveratrol from yeast to human. *Aging* (Albany NY) 2012; 4: 146-158 [PMID: 22436213]
- 67 Northrop MW, Piper GM. A study of diets of patients in a prenatal clinic with an attempt to correlate dietary adequacy with physical findings. *Northwest Med* 1947; 46: 294-298 [PMID: 20291649]
- 68 Beaudeux JL, Nivet-Antoine V, Giral P. Resveratrol: a relevant pharmacological approach for the treatment of metabolic syndrome? *Curr Opin Clin Nutr Metab Care* 2010; 13: 729-736 [PMID: 20823772 DOI: 10.1097/MCO.0b013e32833ef291]
- 69 Smoliga JM, Baur JA, Hausenblas HA. Resveratrol and health-a comprehensive review of human clinical trials. *Mol Nutr Food Res* 2011; 55: 1129-1141 [PMID: 21688389 DOI: 10.1002/mnfr 201100143]
- 70 Baile CA, Yang JY, Rayalam S, Hartzell DL, Lai CY, Andersen C, Della-Fera MA. Effect of resveratrol on fat mobilization. *Ann N Y Acad Sci* 2011; 1215: 40-47 [PMID: 21261640 DOI: 10.1111/j.1749-6632.2010.05845.x]



- 71 Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, Hoeks J, van der Krieken S, Ryu D, Kersten S, Moonen-Kornips E, Hesselink MK, Kunz I, Schrauwen-Hinderling VB, Blaak EE, Auwerx J, Schrauwen P. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metab* 2011; 14: 612-622 [PMID: 22055504 DOI: 10.1016/j.cmet.2011.10.002]
- 72 Liu K, Zhou R, Wang B, Mi MT. Effect of resveratrol on glucose control and insulin sensitivity: a meta-analysis of 11 randomized controlled trials. *Am J Clin Nutr* 2014; 99: 1510-1519 [PMID: 24695890 DOI: 10.3945/ajcn.113.082024]
- 73 Robich MP, Osipov RM, Chu LM, Han Y, Feng J, Nezafat R, Clements RT, Manning WJ, Sellke FW. Resveratrol modifies risk factors for coronary artery disease in swine with metabolic syndrome and myocardial ischemia. *Eur J Pharmacol* 2011; 664: 45-53 [PMID: 21575630 DOI: 10.1016/j.ejphar.2011.04.059]
- 74 Poulsen MM, Vestergaard PF, Clasen BF, Radko Y, Christensen LP, Stødkilde-Jørgensen H, Møller N, Jessen N, Pedersen SB, Jørgensen JO. High-dose resveratrol supplementation in obese men: an investigator-initiated, randomized, placebo-controlled clinical trial of substrate metabolism, insulin sensitivity, and body composition. *Diabetes* 2013; 62: 1186-1195 [PMID: 23193181 DOI: 10.2337/db12-0975]
- 75 Méndez-del Villar M, González-Ortiz M, Martínez-Abundis E, Pérez-Rubio KG, Lizárraga-Valdez R. Effect of resveratrol administration on metabolic syndrome, insulin sensitivity, and insulin secretion. *Metab Syndr Relat Disord* 2014; 12: 497-501 [PMID: 25137036 DOI: 10.1089/met.2014.0082]
- 76 la Porte C, Voduc N, Zhang G, Seguin I, Tardiff D, Singhal N, Cameron DW. Steady-State pharmacokinetics and tolerability of trans-resveratrol 2000 mg twice daily with food, quercetin and alcohol (ethanol) in healthy human subjects. *Clin Pharmacokinet* 2010; 49: 449-454 [PMID: 20528005 DOI: 10.2165/11531820-000 000000-00000]
- 77 Chow HH, Garland LL, Hsu CH, Vining DR, Chew WM, Miller JA, Perloff M, Crowell JA, Alberts DS. Resveratrol modulates drugand carcinogen-metabolizing enzymes in a healthy volunteer study. *Cancer Prev Res* (Phila) 2010; 3: 1168-1175 [PMID: 20716633 DOI: 10.1158/1940-6207.CAPR-09-0155]
- 78 Liu J. Pharmacology of oleanolic acid and ursolic acid. J Ethnopharmacol 1995; 49: 57-68 [PMID: 8847885 DOI: 10.1016/0 378-8741(95)90032-2]
- 79 Liu J. Oleanolic acid and ursolic acid: research perspectives. *J Ethnopharmacol* 2005; **100**: 92-94 [PMID: 15994040 DOI: 10.1016/j.jep.2005.05.024]
- 80 Checker R, Sandur SK, Sharma D, Patwardhan RS, Jayakumar S, Kohli V, Sethi G, Aggarwal BB, Sainis KB. Potent anti-inflammatory activity of ursolic acid, a triterpenoid antioxidant, is mediated through suppression of NF-κB, AP-1 and NF-AT. PLoS One 2012; 7: e31318 [PMID: 22363615 DOI: 10.1371/journal. pone.0031318]
- 81 Ikeda Y, Murakami A, Ohigashi H. Ursolic acid: an anti- and proinflammatory triterpenoid. *Mol Nutr Food Res* 2008; 52: 26-42 [PMID: 18203131 DOI: 10.1002/mnfr.200700389]
- 82 Kowalski R. Studies of selected plant raw materials as alternative sources of triterpenes of oleanolic and ursolic acid types. J Agric

- Food Chem 2007; **55**: 656-662 [PMID: 17263457 DOI: 10.1021/if0625858]
- 83 Jäger S, Trojan H, Kopp T, Laszczyk MN, Scheffler A. Pentacyclic triterpene distribution in various plants - rich sources for a new group of multi-potent plant extracts. *Molecules* 2009; 14: 2016-2031 [PMID: 19513002 DOI: 10.3390/molecules14062016]
- 84 **Pubchem Open Chemistry Database**. Bethesda: National Center for Biotechnology Information, U.S. National Library of Medicine; 2005. [accessed 2015 Jun 19]. Available from: URL: https://pubchem.ncbi.nlm.nih.gov/search/#collection=compounds&query_type=text&query="Ursolic acid"
- Ahmed Z, Ali D, Malik A. Structure determination of ursenetype triterpenes by NMR techniques. *Magn Reson Chem* 2006; 44: 717-719 [PMID: 16607673 DOI: 10.1002/mrc.1803]
- Zhang W, Hong D, Zhou Y, Zhang Y, Shen Q, Li JY, Hu LH, Li J. Ursolic acid and its derivative inhibit protein tyrosine phosphatase 1B, enhancing insulin receptor phosphorylation and stimulating glucose uptake. *Biochim Biophys Acta* 2006; 1760: 1505-1512 [PMID: 16828971 DOI: 10.1016/j.bbagen.2006.05.009]
- 87 Na M, Yang S, He L, Oh H, Kim BS, Oh WK, Kim BY, Ahn JS. Inhibition of protein tyrosine phosphatase 1B by ursane-type triterpenes isolated from Symplocos paniculata. *Planta Med* 2006; 72: 261-263 [PMID: 16534732 DOI: 10.1055/s-2005-873194]
- 88 He Y, Li W, Li Y, Zhang S, Wang Y, Sun C. Ursolic acid increases glucose uptake through the PI3K signaling pathway in adipocytes. PLoS One 2014; 9: e110711 [PMID: 25329874 DOI: 10.1371/journal.pone.0110711]
- 89 Ali H, Houghton PJ, Soumyanath A. alpha-Amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to Phyllanthus amarus. *J Ethnopharmacol* 2006; 107: 449-455 [PMID: 16678367 DOI: 10.1016/j.jep.2006.04.004]
- 90 Lee EH, Popov SA, Lee JY, Shpatov AV, Kukina TP, Kang SW, Pan CH, Um BH, Jung SH. Inhibitory effect of ursolic acid derivatives on recombinant human aldose reductase. *Bioorg Khim* 2011; 37: 637-644 [PMID: 22332359 DOI: 10.1134/S1068162011050050]
- 91 Jang SM, Kim MJ, Choi MS, Kwon EY, Lee MK. Inhibitory effects of ursolic acid on hepatic polyol pathway and glucose production in streptozotocin-induced diabetic mice. *Metabolism* 2010; 59: 512-519 [PMID: 19846180 DOI: 10.1016/j.metabol.2009.07.040]
- 92 Wang ZH, Hsu CC, Huang CN, Yin MC. Anti-glycative effects of oleanolic acid and ursolic acid in kidney of diabetic mice. Eur J Pharmacol 2010; 628: 255-260 [PMID: 19917277 DOI: 10.1016/ j.ejphar.2009.11.019]
- 93 Li Y, Kang Z, Li S, Kong T, Liu X, Sun C. Ursolic acid stimulates lipolysis in primary-cultured rat adipocytes. *Mol Nutr Food Res* 2010; 54: 1609-1617 [PMID: 20521271 DOI: 10.1002/mnfr.200900564]
- 94 Li D, Wang GL, Shan MY, Liu JH, Wang L, Zhu DZ. Effects of ursolic acid on c-Cbl-associated protein expression in 3T3-L1 adipocytes with insulin resistance. *Zhongxiyi Jiehe Xuebao* 2012; 10: 886-893 [PMID: 22883405 DOI: 10.3736/jcim20120809]
- Wang XH, Zhou SY, Qian ZZ, Zhang HL, Qiu LH, Song Z, Zhao J, Wang P, Hao XS, Wang HQ. Evaluation of toxicity and single-dose pharmacokinetics of intravenous ursolic acid liposomes in healthy adult volunteers and patients with advanced solid tumors. *Expert Opin Drug Metab Toxicol* 2013; 9: 117-125 [PMID: 23134084 DOI: 10.1517/17425255.2013.738667]







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