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REVIEW

## Application of the woodchuck animal model for the treatment of hepatitis B virus-induced liver cancer

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Manasa Suresh declares no conflict of interest for this article. Stephan Menne serves occasionally as a paid scientific consultant to Northeastern Wildlife, Inc. (Harris, ID), the only commercial source for woodchucks within the United States

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## Abstract

This review describes woodchucks chronically infected with the woodchuck hepatitis virus (WHV) as an animal model for hepatocarcinogenesis and treatment of primary liver cancer or hepatocellular carcinoma (HCC) induced by the hepatitis B virus (HBV). Since laboratory animal models susceptible to HBV infection are limited, woodchucks experimentally infected with WHV, a hepatitis virus closely related to HBV, are increasingly used to enhance our understanding of virus-host interactions, immune response, and liver disease progression. A correlation of severe liver pathogenesis with high-level viral replication and deficient antiviral immunity has been established, which are present during chronic infection after WHV inoculation of neonatal woodchucks for modeling vertical HBV transmission in humans. HCC in chronic carrier woodchucks develops 17 to 36 mo after neonatal WHV infection and involves liver tumors that are comparable in size, morphology, and molecular gene signature to those of HBV-infected patients. Accordingly, woodchucks with WHV-induced liver tumors have been used for the improvement of imaging and ablation techniques of human HCC. In addition, drug efficacy studies in woodchucks with chronic WHV infection have revealed that prolonged treatment with nucleos(t)ide analogs, alone or in combination with other compounds, minimizes the risk of liver disease progression to HCC. More recently, woodchucks have been utilized in the delineation of mechanisms involved in innate and adaptive immune responses against WHV during acute, self-limited and chronic infections. Therapeutic interventions based on modulating the deficient host antiviral immunity have been explored in woodchucks for inducing functional cure in HBV-infected patients and for reducing or even delaying associated liver disease sequelae, including the onset of HCC. Therefore, woodchucks with chronic WHV infection constitute a well-characterized, fully immunocompetent animal model for HBV-induced liver cancer and for preclinical evaluation of the safety and efficacy of new modalities, which are based on chemo, gene, and immune therapy, for the prevention and treatment of HCC in patients for which current treatment options are dismal.



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**Core Tip:** Hepatitis B virus-induced liver tumors are hard to treat with currently available interventions and the prognosis of hepatocellular carcinoma (HCC) in patients remains still poor. Immunocompetent woodchucks are a useful animal model for human HCC, because multiple tumors at different stages develop spontaneously and secondary to viral infection. This similarity to human hepatocarcinogenesis and the animal's vascular architecture allowing catheterization with human-sized products have increased the preclinical use of this model to improve existing imaging (ultrasound, magnetic resonance imaging, and positron-emission tomography) and ablation techniques (embolization and radiotherapy) and to evaluate interventions (chemo, gene, and immune therapy) intended to treat human HCC.

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## INTRODUCTION

Infection of adult humans with the hepatitis B virus (HBV) usually leads to self-limited liver disease (*i.e.*, acute hepatitis B) and viral resolution, as the virus is controlled *via* a strong antiviral immune response[1,2]. Progression to chronic HBV infection is observed infrequently and occurs only in 5% of infected, healthy adults[3]. However, HBV infection acquired at birth by mother-to-child transfer or during early childhood in unvaccinated infants persists in 95% of individuals[3]. Persistent HBV infection then leads to chronic liver disease (i.e., chronic hepatitis B) that is associated with a diminished or impaired immune response unable to control the virus[1,2]. The immunodeficiencies developed overtime during the persistence of HBV infection are further responsible for the progression of liver disease to liver cirrhosis and hepatocellular carcinoma (HCC) later in life[1,4]. Estimates indicate that approximately 257 million people worldwide are chronic carriers of HBV[5]. Without antiviral treatment and/or liver transplantation, these individuals will die, because end-stage HCC has a low five-year survival rate of about 10%[6]. The therapeutic interventions available for the treatment of chronic HBV infection and associated liver disease sequelae are suboptimal, as they rarely induce viral clearance or significantly lower the risk of HCC development and either require lifelong administration or are associated sometimes with severe adverse effects[4,7-10]. HCC has a high mortality rate because it is frequently asymptomatic and medical attention is often sought when removal by surgery (i.e., hepatectomy) is limited or impossible[11,12]. The poor prognosis of HCC at an advanced stage is mainly due to its unresponsiveness to chemotherapy [11,13-16]. Although tyrosine kinase inhibitors such as sorafenib have demonstrated survival benefits among patients with advanced liver cancer, the prognosis of patients with HCC remains dismal, with tumor recurrence rates of 50% after three years[17]. Thus, chronic HBV infection is a major source of human HCC, which is the fifth most common cancer in the world and the third leading cause of cancer deaths[11,18-20]. Compared to uninfected individuals, the lifetime risk of developing HCC is significantly increased by 15- to 20-fold in patients positive for the HBV surface antigen (HBsAg), and can reach 100-fold in individuals with high levels of HBV replication and serum positivity for HBV e antigen (HBeAg)[20]. The HCC lifetime risk remains increased even after spontaneous clearance of the infection[21]. Therefore, the large reservoir of chronic HBV carriers could benefit immensely from the development of more effective and safer antiviral and anticancer therapies that cure the infection, eliminate the risk of liver disease progression, and/or eradicate established HCC.



Woodchuck hepatitis virus (WHV) infects naturally the Eastern woodchuck (Marmota monax) that habitats large areas within North America, including most eastern and midwestern states in the United States, southeastern Alaska, and southern Canada<sup>[22]</sup>. WHV was initially discovered in 1977 at the Philadelphia Zoo in a colony of woodchucks where several animals died due to chronic hepatitis B and HCC[23,24]. Subsequent studies revealed that WHV is closely related to HBV in regard to the nucleic acid sequence and organization of the genome, virion morphology, and mechanisms of infection and replication[23,25-28]. Consequently, WHV and HBV were classified as members of the genus Orthohepadnavirus within the Hepadnaviridae family [29]. Comparable to HBV infection in humans, WHV in woodchucks also causes agedependent acute, self-limited or chronic outcomes of infection[23,30-33].

Early progress in the development of the woodchuck as an animal model for HBV infection involved basic studies on virological response and liver tumor development that are associated with experimental WHV infection of neonatal and adult woodchucks. Thereafter, neonatal WHV inoculation progressing to chronic viral infection during adulthood has been initially applied for the evaluation of conventional vaccines and nucleos(t)ide analogs for safety and efficacy against HBV[30,34-37]. More recently, the neonatal inoculation model of chronic WHV infection has been increasingly used for the development of immunomodulators, including those stimulating pathogen recognition receptors (PRRs) or blocking immune checkpoint markers[34,38,39]. While some of these studies provided evidence for the prevention and treatment of liver disease progression[37,38], evaluation of interventions directly targeting liver tumors in woodchucks for the treatment of HCC is limited. Since immunopathogenesis and liver disease progression to HCC induced by WHV parallels HBV infection in humans more so than in any other animal model currently available for HBV research[30,34,38-40], woodchucks with established liver tumors have been further applied in the improvement of imaging and ablation techniques and in the evaluation of new therapeutic approaches for the treatment of human HCC. The purpose of this review is to highlight the woodchuck as an animal model for hepatitis virus-induced carcinogenesis and treatment of HCC in patients with chronic HBV infection.

#### WHV infection and liver disease progression

Inoculation of adult woodchucks with WHV almost always results in the acute, selflimited (i.e., resolved) outcome of infection[33,41-44]. Although virtually 100% of hepatocytes in the liver become infected with WHV[45], antiviral control is achieved by strong innate and adaptive immune responses. In the liver, innate immune response is activated within hours after experimental infection and partially inhibits WHV replication[46], although the infection expands further and reaches a peak thereafter. After a lack phase of immune response induction probably due to the "stealth-like behavior" of hepatitis viruses [38,47], a second, more marked, suppression of WHV replication is observed that is mediated by a non-cytolytic mechanism of viral clearance involving type I and II interferons (IFNs)[48]. IFN- $\alpha$  and IFN- $\beta$  are most likely produced by activated PRRs after sensing of viral DNA and RNA in the liver, while IFN- $\gamma$  is mainly secreted by natural killer (NK) cells. These antiviral cytokines inhibit the transcription of viral pre-genomic RNA from the episomal, covalentlyclosed circular (ccc) DNA genome in the nucleus of infected hepatocytes, block its packaging into nucleocapsids, prevent viral replication through upregulation of a ribonuclease, and/or impede synthesis of viral relaxed-circular (rc) DNA within these core particles during maturation, as shown for HBV in cell culture[49-53]and animal models<sup>54-56</sup>]. However, these antiviral cytokines do not affect the levels of WHV e and surface antigens (WHeAg and WHsAg) in the periphery of woodchucks[48,57]. This is followed by a cytolytic mechanism of viral clearance leading to a nearly complete loss of both serum viremia and antigenemia, as well as of intrahepatic WHV cccDNA[48,57]. This mechanism involves killing of infected hepatocytes via mainly cytolytic T lymphocytes (CTLs), apoptosis, and regeneration of hepatocytes, resulting in transient, moderate to marked hepatic inflammation and liver injury [48,58-60]. In addition, virus-neutralizing, protective antibodies against WHsAg, as well as antibodies against WHV core antigen (WHcAg) and WHeAg, are elicited by B-cells[48, 61]. The concerted actions of the immune system then lead to an almost complete shutdown of viral replication in the liver and clearance of the virus from the periphery, although residual amounts of replication-competent WHV and viral cccDNA often remain detectable in serum and in liver, spleen, and blood cells after resolution[45,61-64]. Truncated and thus replication-incompetent WHV DNA is found integrated into the chromosomal DNA of hepatocytes [65-67]. Such viral DNA is typically rearranged and targets different sites within the cellular DNA, suggesting



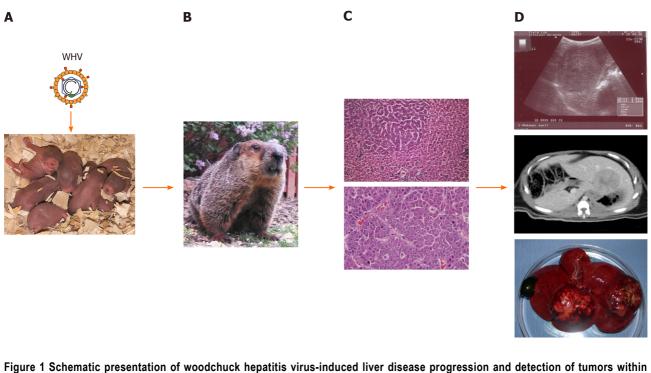
that these integration events may contribute to hepatocarcinogenesis. The presence of unintegrated and integrated virus appears to correlate with an overall lifetime risk of HCC development in 5%-20% of woodchucks after resolution of acute WHV infection [64,68].

This is in contrast to the inoculation of neonatal woodchucks with WHV (Figure 1), which leads to the chronic outcome of infection in approximately 60%-75% of animals later in life, and thus models the effect of age on the outcome of HBV infection in humans[31,33,41]. Persistent WHV infection in these animals involves an ongoing viral replication in liver, minimal to moderate hepatic inflammation and liver injury, and high levels of viral DNA and antigens in the periphery. Compared to the virion levels in patients with chronic HBV infection that are in the range of 109-1010 particles per mL [28], WHV virions often reach 10- to 100-fold greater concentrations in woodchucks with established chronic infection (i.e., 1010-1011 particles/mL), while subviral particles containing WHsAg are produced in large excess. Like in human HBV infection[69], a WHV core-related antigen (WHcrAg), including the classical WHcAg and WHeAg, and additionally, the WHV precore-related antigen (WPreC), is produced during infection in woodchucks, with elevated levels present in chronic WHV carriers[57]. The high loads of circulating WHeAg and WHsAg produced during chronic WHV infection in woodchucks are thought to be responsible for the immunological tolerance to the virus at the T- and B-cell level[30,34,39,40], and are further associated with the liver disease progression to chronic hepatitis B and liver cancer[31,70,71]. HCC develops in all animals over a median period of 2 to 2<sup>1</sup>/<sub>2</sub> years after neonatal inoculation, and the median life expectancy is approximately 6 mo that is similar to the situation in patients with HCC[37,68,72]. More specifically, HCC develops in 50% of woodchucks after 29 mo of chronic WHV infection, in 95% of animals after 3 years, and in 100% of animals by 5 years [73,74]. Thus, chronicity as an outcome of neonatal WHV infection appears to result from a suboptimal or unsuccessful immune response relatively early during the acute phase of infection[30,75,76]. During the later stage of chronic WHV infection, and comparable to chronic HBV infection in patients [1,2,77, 78], a limited type I but a moderate type II IFN response is present in liver[76,79]. Persistent WHV infection is further characterized by the inhibition of antigen presentation to immune cells[80], increases in hepatocyte cytotoxicity via perforingranzyme B and Fas ligand-Fas death pathways[81,82], induction of molecules linked to T-cell exhaustion (i.e., immune checkpoint markers)[79,83], and elevated levels of suppressor of cytokine signaling (SOC3)[79]. Since neutrophils accumulate in woodchuck liver[79], these cells may be responsible for the intrahepatic recruitment of mononuclear inflammatory cells via neutrophil-derived metalloproteinases, as observed in a transgenic mouse model of acute hepatitis B and in patients with chronic hepatitis B[84,85]. Liver disease then appears to progress to HCC due to the reduced immune-mediated clearance of WHV-infected hepatocytes by both non-cytolytic and cytolytic mechanisms[30,76], continuing chronic microinflammation [43,86-88], and viral integration events[67,72,89-91]. However, as described in more detail below, these deficiencies in humoral and cellular immune responses present in chronic WHV carrier woodchucks can be altered by different means leading to a functional cure (defined as a loss of viral DNA and surface antigen in serum, with or without seroconversion to virus-neutralizing antibodies[10]) that delays or even prevents HCC onset.

#### WHV-induced hepatocarcinogenesis

Virus-induced hepatocarcinogenesis in chronic WHV carrier woodchucks is a multistage process (Figure 2). Chronic hepatitis B in these animals is characterized by the mild infiltration of mononuclear cells into portal tracts, sometimes extending beyond the limiting plate[31]. Liver cells with cytoplasmic inclusions are present, which correspond to the ground glass hepatocytes found in the liver of patients with chronic HBV infection and that contain HBV surface antigen (HBsAg)[92]. In HBV transgenic mice, these HBsAg-containing ground glass hepatocytes cluster and form nodules and are seen as preneoplastic lesions[93]. In addition, scattered parenchymal hepatocellular necrosis with neutrophils and macrophages, as well as bile duct proliferation, are usually observed in woodchucks, and in some cases early fibrosis was noted but hepatic cirrhosis and ascites is essentially absent[32,86,87,94,95]. Clinical manifestation of cirrhosis, however, is also absent in a minority of human HCCs due to chronic HBV infection and approximately 20% of HCCs involve non-cirrhotic livers [96,97]. Neoplasia in woodchuck liver then progresses from foci of altered hepatocytes (FAHs) to neoplastic nodules and HCCs[88,98-100]. These altered hepatocytes often harbor viral DNA integrations[65], as also noted in HBV-infected patients[101,102]. They further have a selective regeneration or survival advantage [65] and may be able to escape immune surveillance due to limited intracellular WHV replication and/or





woodchuck liver. A: Neonatal woodchucks are experimentally infected with woodchuck hepatitis virus (WHV) to model vertical hepatitis B virus transmission in humans; B: WHV infection progresses to chronic hepatitis B in adult woodchucks after approximately 1 year; C: Chronic WHV carrier woodchucks develop liver tumors during the next 1-11/2 years. A focus of altered hepatocytes (FAH) in liver (top) and an undifferentiated liver tumor (bottom) are shown; D: Localization of liver tumors by ultrasonography (top) and computed tomography (middle). The liver of a woodchuck with two larger hepatocellular neoplasms (HCC) is shown (bottom). With permission from Elsevier, pictures shown in C were reprinted from: Tennant BC, Toshkov IA, Peek SF, Jacob JR, Menne S, Hornbuckle WE, Schinazi RD, Korba BE, Cote PJ, Gerin JL. Hepatocellular carcinoma in the woodchuck model of hepatitis B virus infection. Gastroenterology 2004; 127(5): S283-S293. Copyright ©American Gastroenterological Association 2004. Published by Elsevier[37]. WHV: Woodchuck hepatitis virus.

presentation of viral epitopes to immune cells. FAHs are detected as early as 6 mo after neonatal WHV inoculation, while small liver tumors occur as early as 3 mo thereafter<sup>[68]</sup>. Metastasis of HCC outside of the liver is essentially absent in woodchucks[32,87,94,103], except for rare cases of pulmonary metastasis in a few animals[86,94]. The hepatic neoplasms present in woodchucks are typically welldifferentiated, trabecular HCCs, although various histologic types are found in different animals or in different tumors in the same animal[32,94,104]. A comparison of intratumoral transcriptional profiles in woodchucks and HBV-infected patients established that WHV-induced HCC shares molecular characteristics with two subtypes of human HCC[79]. One HCC signature present in woodchucks correlated well with the human HCC subclass of poor prognosis ("poor survival subclass") that is characterized by low-level cluster of differentiation (CD) 8+ T-cell and NK-cell infiltration[105]. The second HCC signature in woodchucks was associated with the S2 subclass, a well-defined human HCC subtype[106], which is characterized by the activation of the MYC protooncogene, expression of alpha-fetoprotein (AFP) and epithelial cell adhesion molecule (EpCAM), and a relative suppression of IFNresponsive genes. The observation that HCC develops in all chronic WHV carrier woodchucks provides direct experimental evidence for the oncogenicity of WHV, and by analogy of HBV, as well as other hepatitis viruses naturally infecting several ground squirrel species[24,68]. However, infection with California ground squirrel hepatitis virus (GSHV) leads to less frequent liver cancer development and the HCC onset is much later seen than in chronic WHV infection[107]. This lower oncogenic potential of GSHV was further demonstrated in a comparative study of woodchucks infected as neonates with both WHV and GSHV, as GSHV-induced HCC developed at an later age than WHV-induced HCC in the same host[71]. Since immune cell infiltration into the liver is present during chronic WHV infection, as described above, this continuing chronic inflammatory response likely plays a role in the development of WHV-induced HCC, in addition to viral integration events and proteins, as also observed for HBV-induced HCC[108-111].

Important features of hepatitis virus-induced hepatocarcinogenesis have been investigated in woodchucks and are described here in more detail. Since replicationincompetent WHV DNA is integrated into the chromosomal DNA of woodchuck liver



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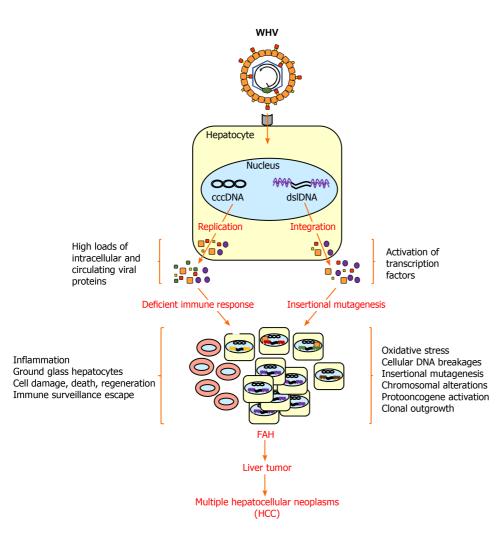


Figure 2 Woodchuck hepatitis virus-induced hepatocarcinogenesis in woodchucks. After infection of normal hepatocytes, woodchuck hepatitis virus (WHV) replicates via cccDNA and produces high loads of intracellular and circulating viral proteins (WHsAg, WHeAg and WPreC) that interfere with the antiviral immunity. The deficient immune response is unable to clear WHV from infected liver cells but causes inflammation. WHsAg accumulates in hepatocytes and gives rise to ground glass hepatocytes. WHV also integrates into the chromosomal DNA of hepatocytes via double-stranded linear (dsl) DNA leading to oxidative stress, oxidation-dependent cellular DNA breakages, insertional mutagenesis, chromosomal alterations, and protooncogene activation. WHsAg and WHV X antigen (WHxAg) are produced from viral DNA integrants. Integrant- and replication-derived viral proteins activate cellular proteins, such as transcription factors, that support the oncogenic process. The continued liver inflammation leads to cell degeneration and regeneration and facilities accumulation of genetic and epigenetic defects in hepatocytes. Individual hepatocytes with critical mutations and low WHV replication and/or antigen presentation escape immune surveillance and their clonal outgrowth results in FAHs that further develop into liver tumors and HCC. CccDNA: Covalently-closed circular DNA; Dsl DNA: Double-stranded linear DNA; FAH: Focus of altered hepatocytes; HCC: Hepatocellular carcinoma; WHV: Woodchuck hepatitis virus.

> tumor cells, which is comparable to the HBV DNA integration in human HCCs[112-115], a direct molecular role of hepatitis viruses in hepatocarcinogenesis is conceivable. The main substrate for integration is viral double-stranded linear (dsl) DNA, which is sometimes produced by the viral polymerase instead of rcDNA within the nucleocapsid[114]. Integration occurs after recycling of nucleocapsids to the nucleus for replenishment of the cccDNA pool[28]. Initial integration in hepatocytes, at least in vitro, is mediated by virus-induced oxidative stress resulting in oxidation-dependent cellular DNA breakages[116]. The integrated viral DNA cannot serve as the source of the progeny virus, but the produced RNA transcripts for the surface and X antigens can become abundantly or even predominantly present, when compared to the viral RNAs transcribed from the cccDNA genome[117]. Thus, integration-derived RNA transcripts may serve as a considerable source for viral antigens with similar function as replication-derived viral proteins and may influence the course of chronic infection and liver disease progression by interfering with the antiviral immunity[117,118].

> WHV DNA integrates into the woodchuck genome at multiple sites within hours after experimental infection [67]. Although it does not appear that there is a preferential integration site for hepatitis viruses [101,113], WHV integrates often into or near the MYC family of protooncogenes in most woodchuck HCCs[68]. Integration close to the N-MYC2 gene or in the b3n and win downstream loci then leads to activated N-



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MYC genes and overexpression of their transcripts in malignant hepatocytes[89,119-122]. In coordination with N-MYC, the insulin-like growth factor-2 (IGF2) is also overexpressed in woodchuck FAHs and HCCs[123,124]. IGF2 blocks apoptosis of malignant liver cells, and thus may allow hepatocytes which otherwise might die to survive, to form FAHs, and to progress to liver tumors[123]. WHV DNA integration further causes N-MYC2 rearrangements, especially in large but less differentiated liver tumors, suggesting that these genetic alterations provide initially a proliferative stimulus or growth advantage for transformed hepatocytes[104]. However, compared with woodchucks that naturally acquired WHV infection, animals experimentally infected with WHV as neonates have more WHV DNA integrations near the N-MYC2 Locus[121]. Although the exact role of N-MYC2 rearrangements and transcripts remains to be elucidated, it was shown that transgenic mice carrying the N-MYC2 gene under the control of WHV regulatory sequences develop liver cancer, including hepatocellular adenomas and HCCs[125].

Like in human HCCs[126], woodchuck liver tumors express small non-proteincoding RNAs or microRNAs at elevated levels, such as miR-17-92 polycistron and miR-21[127]. Knockdown of these microRNAs in human- and woodchuck-derived hepatoma cell lines resulted in a 55% reduction of cell proliferation and anchorageindependent growth, as well as in a suppression of cellular antiapoptotic function. This suggests that onco-microRNAs are involved in the maintenance of malignant hepatocyte transformation during hepatitis virus-induced hepatocarcinogenesis.

Among all viral proteins, the X antigen, a multifunctional transactivator of viral and cellular genes and essential for the establishment of WHV infection in woodchucks [128], has been implicated as a cofactor in the malignant transformation of hepatocytes [129]. HBV replication and liver cell transformation by the HBV X antigen (HBxAg) are associated with the induction of the mitotic polo-like kinase 1 (PLK1) and a parallel downregulation of chromatin remodeling components, including polycomb repressive complex 2 subunit (SUZ12) and zinc finger MYM-type protein 2 (ZMYM2 or ZNF198) [130]. This inverse relationship of PLK1 and SUZ12 was also identified in woodchuck liver tumors[131]. SUZ12 targets many hepatic cancer stem cell markers and proliferation genes. Since expression of these genes is reduced in normal hepatocytes, they are also named "SUZ12 repressed" genes. During WHV-induced hepatocarcinogenesis, the SUZ12 repressed genes encoding BMP, activin membrane-bound inhibitor homolog (BAMBI), and EpCAM, as well as the proliferation gene PLK1, are selectively upregulated in woodchuck tumor cells. Furthermore, metastatic tumor antigen 1 (MTA1), a component of the nucleosome remodeling histone deacetylase complex involved in regulating transcription and chromatin remodeling, is associated with tumor invasiveness and poor prognosis in HBV-induced HCC[132]. Comparable to human HCC, the presence of MTA1 is increased in woodchuck liver tumors, its expression is regulated by the WHV X antigen (WHxAg), and the protein is essential for nuclear factor-kappa B (NF-KB) signaling and tumor progression induced by WHV [133].

Altered expression of vascular endothelial growth factor (VEGF) in the liver is used as a prognostic marker for human HCC[134] and therapeutic interventions targeting this protein or its receptors (VEGFR1/R2) can improve the clinical outcome of HCC in patients[135]. In woodchucks, WHV-induced hepatocarcinogenesis is associated with elevated VEGFR2 expression and increased ligation of VEGF to VEGFR2[136]. This VEGF-driven angiogenesis is accompanied by changes in the liver vasculature, extracellular matrix, and basement membrane, as the number of vessels positive for laminin and platelet endothelial cell adhesion molecule (PECAM1) increased while the number of collagen IV-positive blood vessels declined. This suggests that woodchucks with liver tumors can be utilized in the preclinical evaluation of VEGF-directed therapies for human HCC.

Matrix metalloproteinases (MMPs) play a central role in tumor invasion and metastasis during HBV-induced hepatocarcinogenesis[137]. For obtaining insight in the mechanisms involved in extracellular matrix remodeling in human HCCs, the expression of MMPs was investigated in woodchuck liver tumors[138]. High levels of several MMP transcripts were detected, and especially the transcript and protein levels of MMP-9 correlated with liver disease progression and tumor differentiation in woodchucks, while the protein's gelatinase activity increased during hepatocarcinogenesis. These results are comparable to findings in human liver tumors where the MMP-9 protein level was used for characterizing a more invasive and metastatic type of HCC with poor prognosis[139,140]. Since the gelatinase activities of woodchuck MMP-2 and MMP-9 could be inhibited by a commercially available drug, the use of MMP inhibitors for treatment of human HCC may be a possible treatment option and could be evaluated in woodchucks.



Hepatitis delta virus (HDV), a natural subviral agent of HBV, is known to contribute to HBV-induced hepatocarcinogenesis and to increase the overall risk of HCC in patients during concomitant infection[141-143]. Since HDV only needs the HBsAg for virion envelopment[143], persistence of HDV infection may be independent of HBV replication if integration-derived viral surface antigen can be used, as demonstrated in cell culture[118]. The contribution of HDV to HCC induction and development remains to be elucidated; however, one possible mechanism was revealed in woodchucks[144]. Intravenous inoculation of woodchucks with liver tumors using WHsAg-enveloped HDV demonstrated that malignant hepatocytes are susceptible to HDV infection. Thus, it appears likely that HDV may influence the fate of HCCs by actively replicating in tumor cells and changing the expression of host genes.

Overall, these studies demonstrated that WHV-induced hepatocarcinogenesis in woodchucks has strong similarity to HBV-induced liver carcinogenesis in humans. The features of HCC that are associated with chronic hepatitis virus infection make the woodchuck animal model unique. It further distinguishes woodchucks from other animal models, in which HCC is induced by either a chemical carcinogen, a transgene, or by transplantation of established tumor cell lines into immune-deficient or immunecompatible hosts. Additional advantages of the woodchuck model are the outbred nature of the animals and the heterogeneity of liver tumors that resemble the situation of HBV-infected patients with HCC. These studies further indicated an important role of viral DNA integration, activation of protooncogenes, microRNAs, and the viral X antigen in the malignant transformation of hepatocytes.

#### Development of woodchucks as an animal model for HCC

As described above, liver tumors develop in woodchucks with chronic WHV infection and HCC is fatal in 100% of cases. Tumor progression is usually monitored by serial ultrasonography (US)[86,145,146] and to a lesser degree by repeated magnetic resonance imaging (MRI)[147,148]. Changes in liver enzymes are also used for determining the degree of liver injury due to tumor development[104]. Especially, gamma-glutamyl transferase was validated as an oncogenic biomarker in woodchucks, as increases in this liver enzyme correlate with the onset of HCC[149]. In addition, elevated levels of AFP were linked with WHV-induced hepatocarcinogenesis in woodchucks[150].

#### Improvements in imaging techniques for human HCC

The woodchuck model of HCC has been utilized in the development of new imaging agents for enhancing the detection of hepatic neoplasms by different imaging techniques (Table 1). In the beginning, several contrast agents were evaluated for both gray scale and color Doppler US, including those that use microbubble technology, alone and in combination with hypobaric activation, a vascular imaging agent, or an agent taken up by the reticuloendothelial (RE) system. These agents facilitated tumor localization in the liver and improved measurements of tumor growth and regression in untreated versus treated woodchucks by increasing the sensitivity of US. Furthermore, iron oxide as a contrast agent for the detection of HCC by MRI was tested in woodchucks, either following parenteral administration for uptake by the RE system or intravenous injection as an arabinogalactan conjugate for targeting the asialoglycoprotein receptor that is highly upregulated on normal hepatocytes but not on liver tumor cells. Hepatic imaging with 99mTc-sulfur colloid also detected HCCs after uptake by the RE system and concentration in woodchuck liver. More recently, woodchucks were applied in the improvement of positron-emission tomography (PET) techniques for the early detection of human HCC by comparing radiotracers for uptake into liver tumors and surrounding hepatic tissues. HCC localization and response to radiotherapy was also assessed with MRI by applying contrast agents typically used in patients for visualizing lesions with abnormal vascularity. HCC detection and response to anticancer treatment was further tested by computed tomography (CT) with contrast agents for human use. MRI and/or CT techniques were also applied for generating a virtual three-dimensional (3D) model of the woodchuck hepatic vascular tree[151], as well as for producing virtual and printable 3D models of the woodchuck liver containing tumors that allowed accurate colocalization of imaging with histopathology[152].

## Improvements in techniques for accessing human HCC and treatment by embolization

The woodchuck model was further applied in the evaluation of new techniques developed for gaining less-invasive access to liver tumors for the treatment of HCC in



Table 1 Imaging techniques and contrast agents applied for the detection of woodchuck hepatocellular carcinoma						
Imaging technique	Contrast agent	Brand name	Ref.			
Ultrasonography	Air-filled albumin microspheres		[195]			
	Cyanacrylate polymer microparticles	SHU563A	[196]			
	Dodecafluoropentane emulsion	EchoGen	[197,198]			
	Galactose microparticles/palmitic acid	Levovist	[199-201]			
	Perflexane-filled lipid microspheres	Imagent	[202]			
	Perfluoropropane-filled albumin microspheres	Optison/FS069	[95,200,201,203, 204]			
Scintigraphy	99mTc-sulfur colloid		[205]			
Positron-emission tomography	[1-11C]acetate		[206-208]			
	[1-14C]acetate		[207]			
	[N-methyl-11C]choline		[206,209,210]			
	[18F]clofarabine		[211]			
	[18F]fluoro-ethylcholine		[210]			
	Anti-1-amino-3-[18F]fluoro-cyclobutyl-1-carboxylic acid		[212]			
	2-deoxy-2-[18F]fluoro-D-glucose		[208,209,213]			
	6-deoxy-6[18F]fluoro-D-glucose		[206,209,210]			
	L-[S-methyl-11C]methionine		[212]			
	3-deoxy-3-[18F]fluoro-thymidine		[214]			
Magnetic resonance imaging	Gadolinium	Gadavist or Omniscan	[157,179,185]			
	Gadopentetate dimeglumine	Magnevist	[158,189]			
	Iron oxide		[215,216]			
Computed tomography	Biodegradable radiopaque fiducial markers based on polymers and iodine	Ioversol, Isovue-370, Optiray300, or Optiray350	[152,153,159,186, 206]			
	Diatrizoic acid	Angiografin	[190]			
	Meglumine iotroxate	Biliscopin	[151]			
	Iohexol	Omnipaque or Omnipaque350	[157,188]			

patients. For improving percutaneous liver biopsy techniques, needle-based diffuse optical spectroscopy (DOS) was tested in woodchucks[153]. This established that tissue blood and lipid content and oxygenation level declined, while tissue density increased, when the needle crossed the margin from healthy hepatic parenchyma to liver tumors, indicating that these measurements could be used in real-time as a primary discriminator of normal liver and HCC.

For the treatment of human HCC, chemoembolization and radioembolization via intra-arterial therapies (IAT), alone and in combination with immunotherapy, hold great promise. For the testing of IAT, rather diverse animal species, including mice, rats, rabbits, and pigs, are commonly used as preclinical models of HCC[154-156]. Translation of IAT from these animal models into patients, however, is limited due to the dissimilarity in liver disease development and the size of the vascular system that make arterial access either impossible or challenging, and often requires a surgical cut down for the use of human-size products[155]. This situation is different in woodchucks, because the size of the animals greatly facilitates IAT and other experimental approaches of intratumoral injection. Woodchucks also possess a hepatic arterial anatomy that can be accessed via the femoral artery and allows catheterization with clinically used microcatheters[151]. Accordingly, three studies explored IAT in woodchucks with or without liver tumors[157-159]. In these studies, arterial access via the femoral artery with human standard catheters allowed delivery of contrast agents for the localization of HCCs by CT or MRI. Catheterization further permitted delivery



of embolic particles routinely used in patients into liver tumors by angiography. Lobar embolization with 355-500 µm polyvinyl alcohol (PVA) particles (Boston Scientific) was successful in woodchucks without liver tumors[158]. In addition, liver tumor embolization for the targeted delivery of 100-300 µm PVA microspheres (LC- Bead; BTG, London, United Kingdom) produced a heterogeneous distribution of embolic particles in the hepatic neoplasms[157]. Moreover, chemoembolization with drugeluting embolic 70-150 µm radiopaque PVA microspheres (LC Bead LUMI; BGT) loaded with doxorubicin resulted in a targeted drug delivery into liver tumors[159]. Doxorubicin is an anticancer drug that stops the growth of tumor cells by blocking topoisomerase II and that generates reactive oxygen species for the induction of apoptosis[160].

There is also interest in assessing the biomedical utility of nanomaterials in immunocompetent animal models for the treatment of human HCC. In particular, tumor-associated macrophages within the environment of solid tumors are a preferred target of nanoparticle-based applications, as the balance of inflammatory (tumoricidal) and immunoregulatory (tumor promoting) macrophages controls tumor development, progression, and metastasis[161]. One study evaluated the distribution and clearance of 60 nanometer gold particles into woodchuck liver and tumors after a single intravenous injection at a dose of 14 mg/kg[162]. Although these nanoparticles accumulated to some degree in the spleen after systemic administration, they were mainly found in the lysosome of immunoregulatory macrophages within the liver, as well as in liver resident macrophages. Nanoparticles were further detected in liver tumors and their accumulation in immunoregulatory macrophages was significantly greater in the periphery than in the tumor core. The study concluded that nanoparticle-based delivery of immunomodulators into tumors for treatment of HCC is feasible, especially by targeting tumor-associated macrophages and repolarizing these cells into a more inflammatory phenotype to promote anticancer immunity.

Overall, these studies established that woodchucks with liver tumors are a useful preclinical animal model for the evaluation of transarterial embolotherapies for the treatment of human HCC. They further demonstrated the feasibility of nanoparticlebased delivery of chemotherapeutics or immunomodulators into tumors and assessment of anticancer effects by CT, MRI, or PET imaging.

#### HCC treatment approaches in woodchucks

Woodchucks have been utilized in the evaluation of anticancer effects mediated indirectly by treatment with antiviral drugs or immunomodulators and directly by radiotherapy, tumor excision and ablation, gene therapy, or anticancer drugs (Figure 3).

#### Indirect treatment by antiviral drugs or immunomodulators

Woodchucks with chronic WHV infection were applied in the preclinical evaluation of antiviral drugs being developed for the treatment of HBV-infected patients [30,35,37, 68,163]. Among these drugs, nucleos(t)ide analogs that suppress viral replication in the liver, and thus reduce viremia levels in the periphery, were assessed in woodchucks mainly as a single agent but also in combination (Table 2). Many of these nucleos(t)ide analogs are now approved by national regulatory agencies for administration to patients. While most woodchuck studies were focused on testing nucleos(t)ide analogs for safety and antiviral efficacy during short-term treatment, some studies were extended for the additional evaluation of effects against liver disease progression.

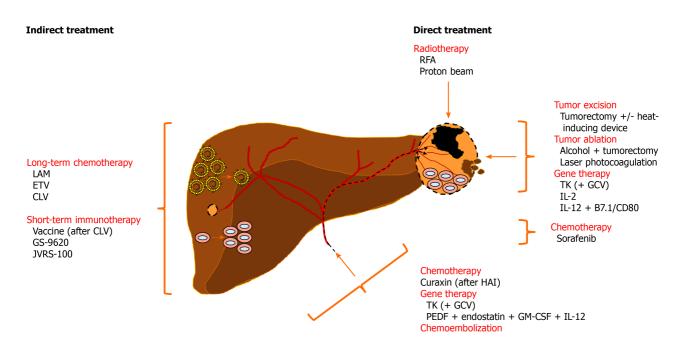
Lifelong, oral treatment of woodchucks with lamivudine, starting at an age of 8 mo and by applying two separate drug doses (*i.e.*, 5 mg/kg/d for approximately 10 mo and then 15 mg/kg/d for up to 50 mo in surviving animals), produced a 4-5 Log<sub>10</sub> reduction in viremia and the antiviral effect was sustained for 1<sup>2</sup>/<sub>3</sub> years while treatment continued[164]. Woodchucks experienced a significant delay in the onset of HCC and death due to severe liver cancer. In particular, lamivudine treatment delayed the development of liver tumors by 24 mo (until an animal age of 32 mo) and extended HCC-free survival by 12 mo (until an animal age of 44 mo). However, when oral lamivudine treatment was initiated in older woodchucks at an age of 13-19 mo and with relatively high doses (i.e., 40 mg/kg/d for 3 mo and/or 200 mg/kg/d for up to 15 mo), the shorter treatment duration and the less pronounced antiviral effect (~2.5 Log<sub>10</sub> decline in viremia) failed to delay hepatocarcinogenesis[165]. Almost all woodchucks developed liver tumors while receiving lamivudine and needed to be euthanized between 12 and 19 mo of treatment due to end-stage HCC (at an animal age of 26-38 mo). Complicating in both studies was the selection of lamivudineresistant WHV mutants during treatment. These mutations occurred frequently in the B domain of the WHV polymerase gene[166,167] and were identical to those reported



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Table 2 Nucleos(t)ide analogs evaluated in woodchucks for safety and antiviral efficacy against hepatitis B virus						
Antiviral drug	Abbreviation	Brand name	Ref.			
Adefovir dipivoxil	ADV	Hepsera	[217-219]			
Clevudine <sup>1</sup>	CLV or L-FMAU	Levovir and Revovir	[172,220-222]			
Emtricitabine	FTC	Coviracil	[219,221,223,224]			
Entecavir <sup>1</sup>	ETV	Baraclude	[171,225-228]			
Lamivudine <sup>1</sup>	LAM or 3TC	Epivir	[164,165,194,219,229]			
Telbivudine	LdT	Tyzeka	[230-232]			
Tenofovir alafenamide	TAF	Vemlidy	[233]			
Tenofovir disoproxil fumarate	TDF	Viread	[219]			
Valtorcitabine	LdC		[230-232]			

<sup>1</sup>Long-term treatment with these drugs delayed hepatocellular carcinoma (HCC) onset and extended HCC-free survival in woodchucks. See text for more details.



#### Figure 3 Overview of therapeutic interventions assessed in woodchucks with liver tumors for the treatment of human hepatocellular

carcinoma. Indirect treatment of chronic WHV carrier woodchucks with nucleos(t)ide analogs or immunomodulators reduces viremia or activates antiviral and anticancer immune responses, respectively, that delay or prevent HCC onset. Direct treatment of hepatic neoplasms by radiotherapy, excision and ablation, gene therapy, or chemotherapy induce apoptosis or necrosis of tumor cells and/or activate an intratumoral, anticancer immune response that result in partial tumor remission. Chemoembolization-mediated anticancer effects have not been evaluated in woodchucks so far. See text for more details. B7.1/CD80: Costimulatory molecule; CLV: Clevudine; ETV: Entecavir; GCV: Ganciclovir; GM-CSF: Granulocyte-macrophage colony-stimulating factor; GS-9620: Toll-like receptor 7 agonist; HAI: Hepatic artery infusion; IL-12: Interleukin 12; JVRS-100: Complex of non-coding plasmid DNA and cationic liposomes; LAM: Lamivudine; PEDF: Pigment epithelium-derived factor; RFA: Radiofrequency ablation; TK: Thymidine kinase.

> in lamivudine-treated patients, in addition to mutations in the C domain of the HBV polymerase gene[168-170].

> Long-term, oral treatment of woodchucks with entecavir for 14 or 36 mo, starting at an animal age of 8 mo and then continuing with a lower dosing frequency from 10 mo of age onward (i.e., 0.5 mg/kg/d for two months and then 0.5 mg/kg/wk for 12 or 34 mo), resulted in a 5-8 Log<sub>10</sub> reduction in serum WHV DNA in 60% or 80% of animals, respectively[171]. The levels of serum WHsAg and intrahepatic WHV cccDNA declined alongside and in parallel with the marked reductions in viremia. An emergence of entecavir-resistant mutants was not observed during the study. Since woodchucks with a sustained antiviral effect stayed negative for signs of liver tumors for up to 21/3 years after drug withdrawal, entecavir treatment prevented the

development of liver cancer in a majority of animals (i.e., up to 80% HCC-free survival).

Delayed HCC onset and prolonged survival was also achieved during long-term, oral treatment of woodchucks with clevudine for 32 wk at a dose of 10 mg/kg/d, starting at an animal age of 1-2 years[37,172]. Thereafter, half of the placebo- or clevudine-treated woodchucks received intramuscularly four doses of a conventional, alum-adsorbed WHsAg vaccine that was administered monthly after drug withdrawal. Combination treatment with clevudine and vaccine reduced viremia by up to 9 Log<sub>10</sub> with undetectable serum WHV DNA in many animals. The antiviral effect was sustained for more than 1 year after treatment cessation in 75% of woodchucks and prevented HCC onset in 38% of animals. However, once HCC was established, the growth rates (*i.e.*, volume doubling times) of liver tumors were similar to those of control animals. Importantly, initiation of clevudine treatment at an animal age of 1 year, and independent of vaccination, produced a more pronounced anticancer effect than a treatment begin at an animal age of 2 years. The development of liver tumors in the younger cohort of woodchucks was further delayed and HCCfree survival increased after 3 (50% vs 0%) and 4 years (25% vs 0%). Moreover, vaccination of these animals without initial clevudine treatment improved the B- and T-cell responses to WHsAg, the protein on which the vaccine was based, but had no effect on viral replication or liver enzyme levels. In combination with clevudine, however, vaccination enhanced these B- and T-cell responses based on the higher titers of virus-neutralizing antibodies and the greater proliferation capability to stimulation with WHsAg. In addition, combination treatment broadened the antiviral immunity to include T-cell responses to other viral antigens, such as WHcAg, WHeAg, and WHxAg, while liver enzyme levels normalized.

Woodchucks were further applied in the preclinical evaluation of immunomodulating compounds being developed for the treatment of HBV-infected patients. The immunomodulators tested so far in woodchucks suppressed WHV replication in the liver and reduced viremia and antigenemia in the periphery at varying degrees. In some instances, the antiviral effect was sustained after the end of treatment, and seroconversion to antibodies against WHsAg and/or WHeAg was achieved in a subset of animals, indicating that a functional cure was induced. Immunomodulators were administered as single agents but more often in combination with nucleos(t)ide analogs and/or inhibitors of viral gene expression and immune checkpoint markers (Table 3). Comparable to the chemotherapy studies, only two immunotherapy studies were designed or extended to include the assessment of anticancer effects.

Short-term, oral administration of the small molecule GS-9620 targeting toll-like receptor (TLR) 7 induced durable antiviral efficacy in woodchucks treated with different doses and dosing frequencies[173]. In the group with the greatest antiviral effect, animals at an age of 12-14 mo received the agonist every other day for approximately 4 wk, initially at 5 mg/kg and then at 2.5 mg/kg after a treatment interruption for 9-10 d due to liver enzyme increase and thrombocytopenia that both reversed during the dose holiday. Treatment in this group induced a rapid reduction in serum WHV DNA of 6.2 Log<sub>10</sub> that was accompanied by declines in intrahepatic WHV cccDNA and undetectable serum WHsAg. Suppressed WHV replication was sustained in all woodchucks during the 31-week follow-up period, and a subset of animals also seroconverted to antibodies against WHsAg during this time. At the end of the study in week 35, all animals were found to be HCC-free during postmortem examination. When combining all woodchucks enrolled in the various treatment groups of this study, and by including only animals that completed treatment and experienced sustained viral suppression, TLR7 agonism reduced the HCC incidence from 71% in placebo-treated control woodchucks to 8% in GS-9620-treated animals. The antiviral and anticancer effects were attributed to the activation of an immune response based on the induction CD8+ T-cells, NK-cells and B-cells, and the production of type I and II interferons in the liver. A follow-up study further indicated that GS-9620 not only targets TLR7 but also TLR8 when administered at high doses[174], possibly explaining the most superior antiviral effect observed so far in the woodchuck animal model with a single agent during short duration treatment.

Intravenous administration of JVRS-100, a complex of non-coding plasmid DNA and cationic liposomes, every second week for 12 wk at two separate doses to woodchucks with liver tumors at an age of 2 years resulted in antiviral and anticancer effects<sup>[175]</sup>. Since the high serum loads of viral DNA and antigens typically present during chronic WHV infection mediated immune suppression, and thus resistance to treatment, only animals with rather low levels of viremia and antigenemia were enrolled in the study. Serum WHV DNA declined by 0.9 Log<sub>10</sub> during JVRS-100 treatment and during the 12-week follow-up period, especially in animals that



Table 3 Immunomodulators evaluated in woodchucks for safety and antiviral efficacy against hepatitis B virus						
Immunomodulator	Compound name	Brand name	Ref.			
IFN-a			[192,226,228,229]			
RIG-I/NOD2 agonist	SB 9200	Inarigivir	[227]			
TLR7 agonist	GS-9620 <sup>1</sup>	Vesatolimod	[173]			
	APR002		[225]			
	RG7854		[38]			
TLR8 agonist	GS-9688	Selgantolimod	[174]			
TLR9 agonist	CpG-ODN		[234]			
TLR9-dependent and -independent pathways	AIC649		[235]			
	JVRS-100 <sup>2</sup>		[175]			
Viral gene expression inhibitor	RG7834		[226]			
Immune checkpoint inhibitor	Anti-PD-L1		[236]			

<sup>1</sup>Treatment delayed hepatocellular carcinoma onset in woodchucks.

<sup>2</sup>Treatment inhibited formation of new liver tumors in woodchucks.

See text for more details. Anti-PD-L1: Antibody against programmed death-ligand 1; CpG-ODN: CpG oligodeoxynucleotide, a short single-stranded synthetic DNA molecule containing unmethylated deoxycytosine-deoxyguanosine (CpG) motifs; IFN- $\alpha$ : Interferon alpha; JVRS-100: Complex of non-coding plasmid DNA and cationic liposomes; NOD2: Nucleotide-binding oligomerization domain-containing protein 2; RIG-I: Retinoic acid-inducible gene I; TLR: Toll-like receptor.

received the higher dose, but the antiviral effect was transient and less pronounced for WHsAg. Although treatment did not induce a regression of preexisting liver tumors, the higher dose prevented the formation of new tumors for 6 mo. These effects were associated with the activation of immune responses that involved CD4+ and CD8+ T-cells and T helper cell type I (Th1) cytokines, such as IFN- $\alpha$ , tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), and interleukin (IL) 2 and 12 in liver and blood, and that apparently blocked the conversion of virus-induced chronic liver disease into HCC.

Overall, these studies demonstrated that long-term treatment with nucleos(t)ide analogs primarily delays but sometimes prevents liver tumor development in woodchucks. Since these studies established a correlation between suppressed viral replication and reduced liver disease progression, early initiation and prolonged duration of conventional antiviral treatment appear most critical for the prevention of hepatitis virus-induced HCC. Since the applied treatment regimens resulted in less cellular damage and liver injury, they most likely deferred the transformation of altered hepatocytes into liver tumors. Short-term immunomodulation, either rather broad or more targeted, mediated lasting protection against formation of new liver tumors or HCC onset. In two studies, immunomodulation was associated with improved or newly elicited humoral and cellular immune responses to viral antigens that were reduced by treatment, and thus could no longer act as endogenous tolerogens.

#### Direct treatment by chemotherapy, radiotherapy, or gene therapy

Since liver tumors obtain their nutrient blood supply from the hepatic artery[176], hepatic artery infusion- (HAI) supported chemotherapy has been applied for the treatment of both primary and metastatic liver cancers in patients and shown to be an effective treatment for unresectable advanced HCC[177]. Effectiveness of this intervention relates to the concentration of chemotherapeutics in HCCs by direct delivery to the tumors, with limited systemic exposure in the liver[178]. In one woodchuck study, HAI ports were placed in the gastroduodenal artery and infused with a curaxin-based experimental anticancer drug, once per week for 3 wk at a dose of 17 mg/kg[179]. Curaxin targets a histone chaperon expressed at high levels in cancer[180] and activates the p53 tumor suppressor gene, while it simultaneously suppresses inhibition of NF-kB[181]. Tumor growth in woodchucks was suppressed after repeated treatment and the anticancer effect was associated with increases in intratumoral T-cell infiltration and tumor cell apoptosis.

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Woodchucks were also applied for testing the preventive effect of long-term, oral treatment with sorafenib[182]. Sorafenib, a small molecule receptor inhibitor of several surface tyrosine kinases, is a standard first-line therapy approved for the treatment of human HCC. Although this drug has both proapoptotic and antiangiogenic properties, the treatment benefit of sorafenib is modest, as only a 3-mo improvement in the overall survival is achieved and its indication is restricted to patients with well-preserved liver function[183]. The underlying mechanism of sorafenib-mediated anticancer activity has not been fully elucidated. Sorafenib was administered daily to woodchucks at two separate doses (i.e., 2.5 mg/kg and 5 mg/kg) using a 5-d-on and 2d-off schedule until tumor development was observed. Although all animals presented with liver tumors independent of the sorafenib dose applied, the lower dose was associated with smaller initial tumor volumes and delayed tumor growth that was associated with an increase in intratumoral CD3+ T-cell infiltration. An effect of sorafenib on chronic WHV infection was not noted. Interestingly, short-term, oral, daily sorafenib administration for 90 d was unable to reciprocate the anticancer effect obtained during long-term treatment. The study concluded that sorafenib has immunomodulatory activity that is dependent on the dose and treatment duration. Caution, however, is warranted when applying higher doses of sorafenib, because of its immunosuppressive function that relates to an increased activity of nuclear factor of activated T-cells 1 (NFAT1) and results in the in vitro inhibition of T-cell proliferation and in an increase in programmed cell death protein 1 (PD-1) expression of CD8+ T-cells, as demonstrated in woodchucks.

Woodchucks with liver tumors were further used to evaluate different ablation techniques for human HCC. One study demonstrated the feasibility of tumor excision, percutaneous alcohol ablation followed by tumorectomy, and laser photocoagulation in this animal model[86]. Extended survival for up to 16-18 mo was achieved with the first two modalities, but multiple tumor recurrence distant from the resection area occurred ultimately in all animals. A second study investigated the effect of a salinelinked dissecting sealer on the remaining tumor beds (i.e., in situ margins) after initial removal of neoplasms by tumorectomy [184]. Surface application of this device induced a heat zone area of up to 5 mm in depth, inside which residual tumor cells, if present, were efficiently destroyed, suggesting that this approach could be beneficial in reducing marginal recurrence after tumor resection. A third study tested radiofrequency ablation (RFA) using a low energy protocol and a 1.0 cm probe that produced a consistent burn area within liver tumors, as determined by necrosis of tumor cells, but was unable to fully ablate larger lesions[185]. A final study assessed the effectiveness of passive scattering proton beams with high dose fractionation[186]. Three fractionations were applied every other day within one week to the hepatic neoplasm. A partial regression of the treated liver tumor was noticed at week 3 post-treatment, which continued until the nodule disappeared at week 9, as also confirmed during postmortem evaluation one week later. The study concluded that a delayed but complete imaging response to proton beam treatment of HCC was achieved in woodchucks without visible gastrointestinal toxicity.

Gene therapeutic strategies based on the induction of apoptosis, antiangiogenesis, or anticancer immune response were assessed in woodchucks for the treatment of human HCC. In one study, an adenoviral vector encoding for the thymidine kinase (TK) of herpes simplex virus under the control of the ubiquitous cytomegalovirus promoter for conferring sensitivity to ganciclovir (GCV) treatment was administered to liver tumors either directly or indirectly *via* the hepatic artery [187]. Transduction of tumor cells and subsequent drug administration resulted in an anticancer effect in two woodchucks that was mediated by GCV-induced apoptosis; however, a third animal died due to acute liver failure that was attributed to the transduction of adjacent, nonneoplastic hepatocytes. Although tumor regression was not achieved, necrotic areas were present in tumors one week after treatment. The study emphasized the need to make vector transduction more specific to liver tumor cells by controlling TK expression with HCC-specific promoters, such as the AFP promoter.

Two other studies tested the anticancer activity mediated by the cytokine IL-12. In the first study, murine IL-12 was expressed from a replication-competent Semliki Forest virus (SFV) vector[188]. Use of this vector has the advantage that the antitumor effect mediated by the cytokine is enhanced *via* the induction of apoptosis in tumor cells that replicate SFV. A single, intratumoral injection of the vector at increasing doses during laparotomy produced a dose-dependent tumor regression that was 80% with the highest dose. Correlating with the temporary IL-12 expression, partial tumor remission was transient and neoplasms began to regrow between 6 and 14 wk after treatment. In addition, all animals experienced a temporary reduction in serum viremia and/or antigenemia. The anticancer and antiviral effects were associated with



augmented T-cell responses to tumor and viral antigens, as well as increased expression of CD4 and CD8 markers and IFN-γ and TNF-α in peripheral blood mononuclear cells. In the second study, a single dose of an adenoviral vector encoding for murine IL-12 and the costimulatory B7.1/CD80 molecule for activating T-cells was injected into liver tumors during laparotomy or under MRI guidance[189]. Transduction of tumor cells resulted in a tumor regression of 80% on average, with one animal experiencing an almost complete tumor elimination within 7 wk. Regression was associated with the induction of an anticancer immune response, as demonstrated by a massive infiltration of CD4+ and CD8+ T-cells into tumors and an increase in intratumoral IFN-γ production. The long-term anticancer effect could not be evaluated, as almost all animals were euthanized two weeks after treatment.

A final study investigated the anticancer effect mediated by antiangiogenic proteins and cytokines in woodchucks [190]. Single dose treatment via the hepatic artery with an adenoviral vector encoding for human pigment epithelium-derived factor (PEDF) and endostatin in combination with an adenoviral vector for the expression of woodchuck granulocyte-macrophage colony-stimulating factor (GM-CSF) and murine IL-12 induced a tumor regression of 90%. The partial tumor remission obtained by combination treatment was superior to the 56% and 76% reduction in tumor volume that was achieved by treatment with antiangiogenic proteins or cytokines alone, respectively. An antiviral effect was not noted during the study and serum viremia and antigenemia remained unchanged in all animals. The tumor regression induced by combination treatment was attributed to several factors, including increased infiltration of CD3+ T-cells into tumors, high intratumoral levels of NK-cells, apoptosis of tumor cells, reductions in tumor vasculature (i.e., reduced microvessel density), and declines in immune checkpoint markers [i.e., PD-1 and cytotoxic T-lymphocyteassociated protein 4 (CTLA-4)] most likely present on regulatory or immunotolerant Tcells within tumors. Since animals were only followed for two weeks after treatment, the durability of the anticancer effect could not be evaluated.

Overall, these studies established that chemotherapy, radiotherapy, and gene therapy of liver tumors are effective means for the treatment of hepatitis virus-induced HCC in woodchucks. Since some studies established a correlation between anticancer immune response and partial tumor remission, approaches which are based on immunomodulation or checkpoint inhibition for inducing functional cure of chronic HBV infection, appear promising and should further be evaluated in woodchucks for treatment of human HCC.

## CONCLUSION

WHV-infected, immunocompetent woodchucks are used to model chronic HBV infection and HCC in humans. Over the past four decades, woodchucks have been applied in the investigation of mechanisms involved in viral immunopathogenesis and hepatocarcinogenesis, in the development of new contrast agents to enhance the detection of hepatic neoplasms by various imaging techniques, in the improvement of tumor ablation strategies based on transarterial embolization and radiotherapy, and in the evaluation of therapeutic interventions directed against the severe outcome of hepatitis virus-induced liver disease. Although the latter was only assessed in a limited number of studies, in which liver tumors were targeted by indirect and direct means, the continued application of woodchucks will support not only the many efforts to cure chronic HBV infection by new antivirals and immunomodulators, but also to treat the associated disease sequelae. Future studies can take advantage of the recently identified woodchuck transcriptome[79,191,192] and genome[193] for generating all needed protein-based markers and assays, as well as of the translational value of woodchucks in predicting therapeutic efficacy against chronic HBV infection in patients[174,192,194]. Thus, chronic WHV carrier woodchucks progressing to HCC within a reasonable time will greatly aid the development and evaluation of the safety and efficacy of new anticancer prophylaxis or therapy in a relevant animal model. Increased testing of anticancer approaches in the woodchuck animal model will ultimately improve the chances for prevention and therapy of HBV-induced HCC.

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## REFERENCES

- Chang JJ, Lewin SR. Immunopathogenesis of hepatitis B virus infection. Immunol Cell Biol 2007; 1 85: 16-23 [PMID: 17130898 DOI: 10.1038/sj.icb.7100009]
- 2 Tan A, Koh S, Bertoletti A. Immune Response in Hepatitis B Virus Infection. Cold Spring Harb Perspect Med 2015; 5: a021428 [PMID: 26134480 DOI: 10.1101/cshperspect.a021428]
- Yuen MF, Chen DS, Dusheiko GM, Janssen HLA, Lau DTY, Locarnini SA, Peters MG, Lai CL. 3 Hepatitis B virus infection. Nat Rev Dis Primers 2018; 4: 18035 [PMID: 29877316 DOI: 10.1038/nrdp.2018.35]
- Revill PA, Chisari FV, Block JM, Dandri M, Gehring AJ, Guo H, Hu J, Kramvis A, Lampertico P, 4 Janssen HLA, Levrero M, Li W, Liang TJ, Lim SG, Lu F, Penicaud MC, Tavis JE, Thimme R; Members of the ICE-HBV Working Groups; ICE-HBV Stakeholders Group Chairs; ICE-HBV Senior Advisors, Zoulim F. A global scientific strategy to cure hepatitis B. Lancet Gastroenterol Hepatol 2019; 4: 545-558 [PMID: 30981686 DOI: 10.1016/S2468-1253(19)30119-0]
- 5 WHO. Hepatitis B. [accessed January 23, 2021] Available from: https://www.hoint/en/newsroom/fact-sheets/detail/hepatitis-b
- 6 Golabi P, Fazel S, Otgonsuren M, Sayiner M, Locklear CT, Younossi ZM. Mortality assessment of patients with hepatocellular carcinoma according to underlying disease and treatment modalities. Medicine (Baltimore) 2017; 96: e5904 [PMID: 28248853 DOI: 10.1097/MD.000000000005904]
- 7 Fanning GC, Zoulim F, Hou J, Bertoletti A. Therapeutic strategies for hepatitis B virus infection: towards a cure. Nat Rev Drug Discov 2019; 18: 827-844 [PMID: 31455905 DOI: 10.1038/s41573-019-0037-0
- Terrault NA, Lok ASF, McMahon BJ, Chang KM, Hwang JP, Jonas MM, Brown RS Jr, Bzowej 8 NH, Wong JB. Update on prevention, diagnosis, and treatment of chronic hepatitis B: AASLD 2018 hepatitis B guidance. *Hepatology* 2018; 67: 1560-1599 [PMID: 29405329 DOI: 10.1002/hep.29800]
- Alter H, Block T, Brown N, Brownstein A, Brosgart C, Chang KM, Chen PJ, Chisari FV, Cohen C, El-Serag H, Feld J, Gish R, Glenn J, Greten T, Guo H, Guo JT, Hoshida Y, Hu J, Kowdley KV, Li W, Liang J, Locarnini S, Lok AS, Mason W, McMahon B, Mehta A, Perrillo R, Revill P, Rice CM, Rinaudo J, Schinazi R, Seeger C, Shetty K, Tavis J, Zoulim F. A research agenda for curing chronic hepatitis B virus infection. Hepatology 2018; 67: 1127-1131 [PMID: 28877549 DOI: 10.1002/hep.29509]
- Lok AS, Zoulim F, Dusheiko G, Ghany MG. Hepatitis B cure: From discovery to regulatory 10 approval. J Hepatol 2017; 67: 847-861 [PMID: 28778687 DOI: 10.1016/j.jhep.2017.05.008]
- 11 Lau WY, Lai EC. Hepatocellular carcinoma: current management and recent advances. Hepatobiliary Pancreat Dis Int 2008; 7: 237-257 [PMID: 18522878]
- 12 Paul SB, Manjunatha YC, Acharya SK. Palliative treatment in advanced hepatocellular carcinoma: has it made any difference? Trop Gastroenterol 2009; 30: 125-134 [PMID: 20306740]
- 13 Villanueva A, Minguez B, Forner A, Reig M, Llovet JM. Hepatocellular carcinoma: novel molecular approaches for diagnosis, prognosis, and therapy. Annu Rev Med 2010; 61: 317-328 [PMID: 20059340 DOI: 10.1146/annurev.med.080608.100623]
- 14 Daher S, Massarwa M, Benson AA, Khoury T. Current and Future Treatment of Hepatocellular Carcinoma: An Updated Comprehensive Review. J Clin Transl Hepatol 2018; 6: 69-78 [PMID: 29607307 DOI: 10.14218/JCTH.2017.00031]
- Ghavimi S, Apfel T, Azimi H, Persaud A, Pyrsopoulos NT. Management and Treatment of 15 Hepatocellular Carcinoma with Immunotherapy: A Review of Current and Future Options. J Clin Transl Hepatol 2020; 8: 168-176 [PMID: 32832397 DOI: 10.14218/JCTH.2020.00001]
- 16 Saffo S, Taddei TH. Systemic Management for Advanced Hepatocellular Carcinoma: A Review of the Molecular Pathways of Carcinogenesis, Current and Emerging Therapies, and Novel Treatment Strategies. Dig Dis Sci 2019; 64: 1016-1029 [PMID: 30887150 DOI: 10.1007/s10620-019-05582-x]
- 17 Lu LC, Cheng AL, Poon RT. Recent advances in the prevention of hepatocellular carcinoma recurrence. Semin Liver Dis 2014; 34: 427-434 [PMID: 25369304 DOI: 10.1055/s-0034-1394141]
- 18 Tsai WL, Chung RT. Viral hepatocarcinogenesis. Oncogene 2010; 29: 2309-2324 [PMID: 20228847 DOI: 10.1038/onc.2010.36]
- 19 Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-E386 [PMID: 25220842 DOI: 10.1002/ijc.29210]
- 20 El-Serag HB. Epidemiology of viral hepatitis and hepatocellular carcinoma. Gastroenterology 2012; 142: 1264-1273. e1 [PMID: 22537432 DOI: 10.1053/j.gastro.2011.12.061]
- 21 Nam SW, Jung JJ, Bae SH, Choi JY, Yoon SK, Cho SH, Han JY, Han NI, Yang JM, Lee YS. Clinical outcomes of delayed clearance of serum HBsAG in patients with chronic HBV infection. Korean J Intern Med 2007; 22: 73-76 [PMID: 17616021 DOI: 10.3904/kjim.2007.22.2.73]
- 22 Bellezza CA, Sexton S, Curtin LI, Concannon PW, Baldwin BH, Graham LA, Hornbuckle WE, Roth L, Tennant BC. The laboratory woodchuck (Marmota monax). In: Fox JG, Anderson LC, Otto GM, Pritchett-Corning KR, Whary MT (eds). American College of Laboratory Animal Medicine,



Laboratory Animal Medicine. 3rd ed. Cambridge (MA): Academic Press, 2015: 351-386 [DOI: 10.1016/B978-0-12-409527-4.00008-0]

- 23 Summers J, Smolec JM, Snyder R. A virus similar to human hepatitis B virus associated with hepatitis and hepatoma in woodchucks. Proc Natl Acad Sci USA 1978; 75: 4533-4537 [PMID: 212758 DOI: 10.1073/pnas.75.9.4533]
- Summers J. Three recently described animal virus models for human hepatitis B virus. Hepatology 24 1981; 1: 179-183 [PMID: 7286898 DOI: 10.1002/hep.1840010215]
- Galibert F, Chen TN, Mandart E. Nucleotide sequence of a cloned woodchuck hepatitis virus 25 genome: comparison with the hepatitis B virus sequence. J Virol 1982; 41: 51-65 [PMID: 7086958 DOI: 10.1128/JVI.41.1.51-65.1982]
- 26 Girones R, Cote PJ, Hornbuckle WE, Tennant BC, Gerin JL, Purcell RH, Miller RH. Complete nucleotide sequence of a molecular clone of woodchuck hepatitis virus that is infectious in the natural host. Proc Natl Acad Sci USA 1989; 86: 1846-1849 [PMID: 2928306 DOI: 10.1073/pnas.86.6.1846]
- 27 Mason WS. Animal models and the molecular biology of hepadnavirus infection. Cold Spring Harb Perspect Med 2015; 5 [PMID: 25833941 DOI: 10.1101/cshperspect.a021352]
- Seeger C, Mason WS. Molecular biology of hepatitis B virus infection. Virology 2015; 479-480: 28 672-686 [PMID: 25759099 DOI: 10.1016/j.virol.2015.02.031]
- Gust ID, Burrell CJ, Coulepis AG, Robinson WS, Zuckerman AJ. Taxonomic classification of 29 human hepatitis B virus. Intervirology 1986; 25: 14-29 [PMID: 3516924 DOI: 10.1159/000149651]
- 30 Menne S, Cote PJ. The woodchuck as an animal model for pathogenesis and therapy of chronic hepatitis B virus infection. World J Gastroenterol 2007; 13: 104-124 [PMID: 17206759 DOI: 10.3748/wjg.v13.i1.104
- 31 Popper H, Roth L, Purcell RH, Tennant BC, Gerin JL. Hepatocarcinogenicity of the woodchuck hepatitis virus. Proc Natl Acad Sci USA 1987; 84: 866-870 [PMID: 3468514 DOI: 10.1073/pnas.84.3.866]
- 32 Snyder RL, Tyler G, Summers J. Chronic hepatitis and hepatocellular carcinoma associated with woodchuck hepatitis virus. Am J Pathol 1982; 107: 422-425 [PMID: 6282133]
- Cote PJ, Korba BE, Miller RH, Jacob JR, Baldwin BH, Hornbuckle WE, Purcell RH, Tennant BC, 33 Gerin JL. Effects of age and viral determinants on chronicity as an outcome of experimental woodchuck hepatitis virus infection. Hepatology 2000; 31: 190-200 [PMID: 10613745 DOI: 10.1002/hep.510310128
- 34 Roggendorf M, Kosinska AD, Liu J, Lu M. The Woodchuck, a Nonprimate Model for Immunopathogenesis and Therapeutic Immunomodulation in Chronic Hepatitis B Virus Infection. Cold Spring Harb Perspect Med 2015; 5 [PMID: 26511761 DOI: 10.1101/cshperspect.a021451]
- 35 Tennant BC, Gerin JL. The woodchuck model of hepatitis B virus infection. ILAR J 2001; 42: 89-102 [PMID: 11406711 DOI: 10.1093/ilar.42.2.89]
- 36 Cote PJ, Korba BE, Tennant BC, Gerin JL. Immunopathogenesis and immunomodulation of woodchuck hepatitis virus infection. In: Hollinger FB, Lemon SM, Margolis HS (eds). Viral hepatitis and liver disease. Baltimore (MD): Lippincott Williams & Wilkins, 1991: 483-486
- Tennant BC, Toshkov IA, Peek SF, Jacob JR, Menne S, Hornbuckle WE, Schinazi RD, Korba BE, 37 Cote PJ, Gerin JL. Hepatocellular carcinoma in the woodchuck model of hepatitis B virus infection. Gastroenterology 2004; 127: S283-S293 [PMID: 15508096 DOI: 10.1053/j.gastro.2004.09.043]
- 38 Suslov A, Wieland S, Menne S. Modulators of innate immunity as novel therapeutics for treatment of chronic hepatitis B. Curr Opin Virol 2018; 30: 9-17 [PMID: 29444493 DOI: 10.1016/j.coviro.2018.01.008]
- 39 Kosinska AD, Liu J, Lu M, Roggendorf M. Therapeutic vaccination and immunomodulation in the treatment of chronic hepatitis B: preclinical studies in the woodchuck. Med Microbiol Immunol 2015; 204: 103-114 [PMID: 25535101 DOI: 10.1007/s00430-014-0379-5]
- 40 Michalak TI. Diverse Virus and Host-Dependent Mechanisms Influence the Systemic and Intrahepatic Immune Responses in the Woodchuck Model of Hepatitis B. Front Immunol 2020; 11: 853 [PMID: 32536912 DOI: 10.3389/fimmu.2020.00853]
- Cote PJ, Toshkov I, Bellezza C, Ascenzi M, Roneker C, Ann Graham L, Baldwin BH, Gaye K, 41 Nakamura I, Korba BE, Tennant BC, Gerin JL. Temporal pathogenesis of experimental neonatal woodchuck hepatitis virus infection: increased initial viral load and decreased severity of acute hepatitis during the development of chronic viral infection. Hepatology 2000; 32: 807-817 [PMID: 11003627 DOI: 10.1053/jhep.2000.17681]
- 42 Wong DC, Shih JW, Purcell RH, Gerin JL, London WT. Natural and experimental infection of woodchucks with woodchuck hepatitis virus, as measured by new, specific assays for woodchuck surface antigen and antibody. J Clin Microbiol 1982; 15: 484-490 [PMID: 7076821 DOI: 10.1128/JCM.15.3.484-490.1982]
- 43 Millman I, Southam L, Halbherr T, Simmons H, Kang CM. Woodchuck hepatitis virus: experimental infection and natural occurrence. Hepatology 1984; 4: 817-823 [PMID: 6383996 DOI: 10.1002/hep.1840040503]
- 44 Tyler GV, Snyder RL, Summers J. Experimental infection of the woodchuck (Marmota monax monax) with woodchuck hepatitis virus. Lab Invest 1986; 55: 51-55 [PMID: 3724063]
- 45 Kajino K, Jilbert AR, Saputelli J, Aldrich CE, Cullen J, Mason WS. Woodchuck hepatitis virus infections: very rapid recovery after a prolonged viremia and infection of virtually every hepatocyte. J Virol 1994; 68: 5792-5803 [PMID: 7914548 DOI: 10.1128/JVI.68.9.5792-5803.1994]



- 46 Guy CS, Mulrooney-Cousins PM, Churchill ND, Michalak TI. Intrahepatic expression of genes affiliated with innate and adaptive immune responses immediately after invasion and during acute infection with woodchuck hepadnavirus. J Virol 2008; 82: 8579-8591 [PMID: 18596101 DOI: 10.1128/JVI.01022-08]
- 47 Wieland S, Thimme R, Purcell RH, Chisari FV. Genomic analysis of the host response to hepatitis B virus infection. Proc Natl Acad Sci USA 2004; 101: 6669-6674 [PMID: 15100412 DOI: 10.1073/pnas.0401771101
- Suresh M, Czerwinski S, Murreddu MG, Kallakury BV, Ramesh A, Gudima SO, Menne S. Innate 48 and adaptive immunity associated with resolution of acute woodchuck hepatitis virus infection in adult woodchucks. PLoS Pathog 2019; 15: e1008248 [PMID: 31869393 DOI: 10.1371/journal.ppat.1008248]
- 49 Lucifora J, Xia Y, Reisinger F, Zhang K, Stadler D, Cheng X, Sprinzl MF, Koppensteiner H, Makowska Z, Volz T, Remouchamps C, Chou WM, Thasler WE, Hüser N, Durantel D, Liang TJ, Münk C, Heim MH, Browning JL, Dejardin E, Dandri M, Schindler M, Heikenwalder M, Protzer U. Specific and nonhepatotoxic degradation of nuclear hepatitis B virus cccDNA. Science 2014; 343: 1221-1228 [PMID: 24557838 DOI: 10.1126/science.1243462]
- 50 Wieland SF, Eustaquio A, Whitten-Bauer C, Boyd B, Chisari FV. Interferon prevents formation of replication-competent hepatitis B virus RNA-containing nucleocapsids. Proc Natl Acad Sci USA 2005; 102: 9913-9917 [PMID: 15994231 DOI: 10.1073/pnas.0504273102]
- 51 Xia Y, Stadler D, Lucifora J, Reisinger F, Webb D, Hösel M, Michler T, Wisskirchen K, Cheng X, Zhang K, Chou WM, Wettengel JM, Malo A, Bohne F, Hoffmann D, Eyer F, Thimme R, Falk CS, Thasler WE, Heikenwalder M, Protzer U. Interferon-y and Tumor Necrosis Factor-a Produced by T Cells Reduce the HBV Persistence Form, cccDNA, Without Cytolysis. Gastroenterology 2016; 150: 194-205 [PMID: 26416327 DOI: 10.1053/j.gastro.2015.09.026]
- 52 Xu C, Guo H, Pan XB, Mao R, Yu W, Xu X, Wei L, Chang J, Block TM, Guo JT. Interferons accelerate decay of replication-competent nucleocapsids of hepatitis B virus. J Virol 2010; 84: 9332-9340 [PMID: 20610715 DOI: 10.1128/JVI.00918-10]
- 53 Liu Y, Nie H, Mao R, Mitra B, Cai D, Yan R, Guo JT, Block TM, Mechti N, Guo H. Interferoninducible ribonuclease ISG20 inhibits hepatitis B virus replication through directly binding to the epsilon stem-loop structure of viral RNA. PLoS Pathog 2017; 13: e1006296 [PMID: 28399146 DOI: 10.1371/journal.ppat.1006296
- Wieland SF, Guidotti LG, Chisari FV. Intrahepatic induction of alpha/beta interferon eliminates 54 viral RNA-containing capsids in hepatitis B virus transgenic mice. J Virol 2000; 74: 4165-4173 [PMID: 10756029 DOI: 10.1128/jvi.74.9.4165-4173.2000]
- Wieland SF, Spangenberg HC, Thimme R, Purcell RH, Chisari FV. Expansion and contraction of 55 the hepatitis B virus transcriptional template in infected chimpanzees. Proc Natl Acad Sci USA 2004; 101: 2129-2134 [PMID: 14764900 DOI: 10.1073/pnas.0308478100]
- 56 Anderson AL, Banks KE, Pontoglio M, Yaniv M, McLachlan A. Alpha/beta interferon differentially modulates the clearance of cytoplasmic encapsidated replication intermediates and nuclear covalently closed circular hepatitis B virus (HBV) DNA from the livers of hepatocyte nuclear factor 1alpha-null HBV transgenic mice. J Virol 2005; 79: 11045-11052 [PMID: 16103155 DOI: 10.1128/JVI.79.17.11045-11052.2005]
- 57 Hong X, Luckenbaugh L, Perlman D, Revill PA, Wieland SF, Menne S, Hu J. Characterization and Application of Precore/Core-Related Antigens in Animal Models of Hepatitis B Virus Infection. Hepatology 2021 [PMID: 33458844 DOI: 10.1002/hep.31720]
- 58 Hodgson PD, Michalak TI. Augmented hepatic interferon gamma expression and T-cell influx characterize acute hepatitis progressing to recovery and residual lifelong virus persistence in experimental adult woodchuck hepatitis virus infection. Hepatology 2001; 34: 1049-1059 [PMID: 11679978 DOI: 10.1053/jhep.2001.29004]
- Guo JT, Zhou H, Liu C, Aldrich C, Saputelli J, Whitaker T, Barrasa MI, Mason WS, Seeger C. 59 Apoptosis and regeneration of hepatocytes during recovery from transient hepadnavirus infections. J Virol 2000; 74: 1495-1505 [PMID: 10627561 DOI: 10.1128/jvi.74.3.1495-1505.2000]
- 60 Hodgson PD, Grant MD, Michalak TI. Perforin and Fas/Fas ligand-mediated cytotoxicity in acute and chronic woodchuck viral hepatitis. Clin Exp Immunol 1999; 118: 63-70 [PMID: 10540161 DOI: 10.1046/j.1365-2249.1999.01010.x]
- Korba BE, Cote PJ, Wells FV, Baldwin B, Popper H, Purcell RH, Tennant BC, Gerin JL. Natural 61 history of woodchuck hepatitis virus infections during the course of experimental viral infection: molecular virologic features of the liver and lymphoid tissues. J Virol 1989; 63: 1360-1370 [PMID: 2915383 DOI: 10.1128/JVI.63.3.1360-1370.1989]
- Korba BE, Cote PJ, Gerin JL. Mitogen-induced replication of woodchuck hepatitis virus in cultured peripheral blood lymphocytes. Science 1988; 241: 1213-1216 [PMID: 3261887 DOI: 10.1126/science.32618871
- 63 Coffin CS, Michalak TI. Persistence of infectious hepadnavirus in the offspring of woodchuck mothers recovered from viral hepatitis. J Clin Invest 1999; 104: 203-212 [PMID: 10411550 DOI: 10.1172/JCI5048
- Michalak TI, Pardoe IU, Coffin CS, Churchill ND, Freake DS, Smith P, Trelegan CL. Occult 64 lifelong persistence of infectious hepadnavirus and residual liver inflammation in woodchucks convalescent from acute viral hepatitis. Hepatology 1999; 29: 928-938 [PMID: 10051500 DOI: 10.1002/hep.510290329]



- 65 Mason WS, Jilbert AR, Summers J. Clonal expansion of hepatocytes during chronic woodchuck hepatitis virus infection. Proc Natl Acad Sci USA 2005; 102: 1139-1144 [PMID: 15657132 DOI: 10.1073/pnas.0409332102
- Summers J, Jilbert AR, Yang W, Aldrich CE, Saputelli J, Litwin S, Toll E, Mason WS. Hepatocyte 66 turnover during resolution of a transient hepadnaviral infection. Proc Natl Acad Sci USA 2003; 100: 11652-11659 [PMID: 14500915 DOI: 10.1073/pnas.1635109100]
- Chauhan R, Churchill ND, Mulrooney-Cousins PM, Michalak TI. Initial sites of hepadnavirus 67 integration into host genome in human hepatocytes and in the woodchuck model of hepatitis Bassociated hepatocellular carcinoma. Oncogenesis 2017; 6: e317 [PMID: 28414318 DOI: 10.1038/oncsis.2017.221
- Tennant BC. Animal models of hepadnavirus-associated hepatocellular carcinoma. Clin Liver Dis 68 2001; 5: 43-68 [PMID: 11218919 DOI: 10.1016/s1089-3261(05)70153-7]
- Hong X, Luckenbaugh L, Mendenhall M, Walsh R, Cabuang L, Soppe S, Revill PA, Burdette D, 69 Feierbach B, Delaney W, Hu J. Characterization of Hepatitis B Precore/Core-Related Antigens. J Virol 2021; 95 [PMID: 33148795 DOI: 10.1128/JVI.01695-20]
- 70 Ponzetto A, Forzani B. Animal models of hepatocellular carcinoma: hepadnavirus-induced liver cancer in woodchucks. Ital J Gastroenterol 1991; 23: 491-493 [PMID: 1751825]
- 71 Seeger C, Baldwin B, Hornbuckle WE, Yeager AE, Tennant BC, Cote P, Ferrell L, Ganem D, Varmus HE. Woodchuck hepatitis virus is a more efficient oncogenic agent than ground squirrel hepatitis virus in a common host. J Virol 1991; 65: 1673-1679 [PMID: 2002538 DOI: 10.1128/JVI.65.4.1673-1679.1991
- 72 Korba BE, Wells FV, Baldwin B, Cote PJ, Tennant BC, Popper H, Gerin JL. Hepatocellular carcinoma in woodchuck hepatitis virus-infected woodchucks: presence of viral DNA in tumor tissue from chronic carriers and animals serologically recovered from acute infections. Hepatology 1989; 9: 461-470 [PMID: 2465987 DOI: 10.1002/hep.1840090321]
- Cote PJ, Gerin JL. The woodchuck as a model of hepadnavirus infection, pathogenesis and therapy. 73 Forum Trends Exp Clin Med 1996; 6: 131-159
- Tennant BC, Gerin JL. The woodchuck model of hepatitis B virus infection. In: Arias IM, Boyer J, 74 Fausto N, Jakoby WB, Schachter D, Shafritz DA (eds). The liver: Biology and pathology. 3rd ed. New York (NY): Raven Press, 1994: 1455-1466
- 75 Nakamura I, Nupp JT, Cowlen M, Hall WC, Tennant BC, Casey JL, Gerin JL, Cote PJ. Pathogenesis of experimental neonatal woodchuck hepatitis virus infection: chronicity as an outcome of infection is associated with a diminished acute hepatitis that is temporally deficient for the expression of interferon gamma and tumor necrosis factor-alpha messenger RNAs. Hepatology 2001; 33: 439-447 [PMID: 11172347 DOI: 10.1053/jhep.2001.21748]
- 76 Wang Y, Menne S, Jacob JR, Tennant BC, Gerin JL, Cote PJ. Role of type 1 versus type 2 immune responses in liver during the onset of chronic woodchuck hepatitis virus infection. Hepatology 2003; 37: 771-780 [PMID: 12668969 DOI: 10.1053/jhep.2003.50154]
- Bertoletti A, Ferrari C. Innate and adaptive immune responses in chronic hepatitis B virus 77 infections: towards restoration of immune control of viral infection. Gut 2012; 61: 1754-1764 [PMID: 22157327 DOI: 10.1136/gutjnl-2011-301073]
- Ferrari C. HBV and the immune response. Liver Int 2015; 35 Suppl 1: 121-128 [PMID: 25529097 78 DOI: 10.1111/liv.12749]
- 79 Fletcher SP, Chin DJ, Ji Y, Iniguez AL, Taillon B, Swinney DC, Ravindran P, Cheng DT, Bitter H, Lopatin U, Ma H, Klumpp K, Menne S. Transcriptomic analysis of the woodchuck model of chronic hepatitis B. Hepatology 2012; 56: 820-830 [PMID: 22431061 DOI: 10.1002/hep.25730]
- 80 Michalak TI, Hodgson PD, Churchill ND. Posttranscriptional inhibition of class I major histocompatibility complex presentation on hepatocytes and lymphoid cells in chronic woodchuck hepatitis virus infection. J Virol 2000; 74: 4483-4494 [PMID: 10775584 DOI: 10.1128/jvi.74.10.4483-4494.2000]
- 81 Guy CS, Wang J, Michalak TI. Hepatocytes as cytotoxic effector cells can induce cell death by CD95 ligand-mediated pathway. *Hepatology* 2006; **43**: 1231-1240 [PMID: 16729304 DOI: 10.1002/hep.21201]
- 82 Guy CS, Rankin SL, Wang J, Michalak TI. Hepatocytes can induce death of contacted cells via perforin-dependent mechanism. Hepatology 2008; 47: 1691-1701 [PMID: 18393317 DOI: 10.1002/hep.22228]
- 83 Zhang E, Zhang X, Liu J, Wang B, Tian Y, Kosinska AD, Ma Z, Xu Y, Dittmer U, Roggendorf M, Yang D, Lu M. The expression of PD-1 ligands and their involvement in regulation of T cell functions in acute and chronic woodchuck hepatitis virus infection. PLoS One 2011; 6: e26196 [PMID: 22022563 DOI: 10.1371/journal.pone.0026196]
- Sitia G, Isogawa M, Kakimi K, Wieland SF, Chisari FV, Guidotti LG. Depletion of neutrophils blocks the recruitment of antigen-nonspecific cells into the liver without affecting the antiviral activity of hepatitis B virus-specific cytotoxic T lymphocytes. Proc Natl Acad Sci USA 2002; 99: 13717-13722 [PMID: 12368481 DOI: 10.1073/pnas.172521999]
- 85 Zhang JY, Zou ZS, Huang A, Zhang Z, Fu JL, Xu XS, Chen LM, Li BS, Wang FS. Hyper-activated pro-inflammatory CD16 monocytes correlate with the severity of liver injury and fibrosis in patients with chronic hepatitis B. PLoS One 2011; 6: e17484 [PMID: 21390263 DOI: 10.1371/journal.pone.0017484]
- Gouillat C, Manganas D, Zoulim F, Vitrey D, Saguier G, Guillaud M, Ain JF, Duque-Campos R, 86



Jamard C, Praves M, Trepo C. Woodchuck hepatitis virus-induced carcinoma as a relevant natural model for therapy of human hepatoma. J Hepatol 1997; 26: 1324-1330 [PMID: 9210620 DOI: 10.1016/s0168-8278(97)80468-0

- 87 Popper H, Shih JW, Gerin JL, Wong DC, Hoyer BH, London WT, Sly DL, Purcell RH. Woodchuck hepatitis and hepatocellular carcinoma: correlation of histologic with virologic observations. Hepatology 1981; 1: 91-98 [PMID: 6269981 DOI: 10.1002/hep.1840010202]
- 88 Mi LJ, Patil J, Hornbuckle WE, Cote PJ, Gerin JL, Tennant BC, Paronetto F. DNA ploidy analysis of hepatic preneoplastic and neoplastic lesions in woodchucks experimentally infected with woodchuck hepatitis virus. Hepatology 1994; 20: 21-29 [PMID: 8020890 DOI: 10.1016/0270-9139(94)90129-5
- Fourel G, Trepo C, Bougueleret L, Henglein B, Ponzetto A, Tiollais P, Buendia MA. Frequent 89 activation of N-myc genes by hepadnavirus insertion in woodchuck liver tumours. Nature 1990; 347: 294-298 [PMID: 2205804 DOI: 10.1038/347294a0]
- Kaneko S, Oshima T, Kodama K, Aoyama S, Yoshikawa H, Unoura M, Fukuoka K, Matsushita F, 90 Morimoto H, Kobayashi K. Stable integration of woodchuck hepatitis virus DNA in transplanted tumors and established tissue culture cells derived from a woodchuck primary hepatocellular carcinoma. Cancer Res 1986; 46: 3608-3613 [PMID: 3011252]
- Fuchs K, Heberger C, Weimer T, Roggendorf M. Characterization of woodchuck hepatitis virus 91 DNA and RNA in the hepatocellular carcinomas of woodchucks. Hepatology 1989; 10: 215-220 [PMID: 2545590 DOI: 10.1002/hep.1840100216]
- 92 Popper H, Shafritz DA, Hoofnagle JH. Relation of the hepatitis B virus carrier state to hepatocellular carcinoma. Hepatology 1987; 7: 764-772 [PMID: 3038725 DOI: 10.1002/hep.1840070425]
- 93 Chisari FV, Klopchin K, Moriyama T, Pasquinelli C, Dunsford HA, Sell S, Pinkert CA, Brinster RL, Palmiter RD. Molecular pathogenesis of hepatocellular carcinoma in hepatitis B virus transgenic mice. Cell 1989; 59: 1145-1156 [PMID: 2598264 DOI: 10.1016/0092-8674(89)90770-8]
- Roth L, King JM, Hornbuckle WE, Harvey HJ, Tennant BC. Chronic hepatitis and hepatocellular 94 carcinoma associated with persistent woodchuck hepatitis virus infection. Vet Pathol 1985; 22: 338-343 [PMID: 2994274 DOI: 10.1177/030098588502200407]
- 95 Nada T, Moriyasu F, Kono Y, Suginoshita Y, Matsumura T, Kobayashi K, Nakamura T, Chiba T. Sonographic detection of tumor blood flow using a new contrast agent in woodchuck hepatomas. J Ultrasound Med 1997; 16: 485-491 [PMID: 9315200 DOI: 10.7863/jum.1997.16.7.485]
- 96 Chavanupatkul M, Omino R, Mittal S, Kramer JR, Richardson P, Thrift AP, El-Serag HB, Kanwal F. Hepatocellular carcinoma in the absence of cirrhosis in patients with chronic hepatitis B virus infection. J Hepatol 2017; 66: 355-362 [PMID: 27693539 DOI: 10.1016/j.jhep.2016.09.013]
- Gaddikeri S, McNeeley MF, Wang CL, Bhargava P, Dighe MK, Yeh MM, Dubinsky TJ, 97 Kolokythas O, Lalwani N. Hepatocellular carcinoma in the noncirrhotic liver. AJR Am J Roentgenol 2014; 203: W34-W47 [PMID: 24951228 DOI: 10.2214/AJR.13.11511]
- 98 Abe K, Kurata T, Shikata T, Tennant BC. Enzyme-altered liver cell foci in woodchucks infected with woodchuck hepatitis virus. Jpn J Cancer Res 1988; 79: 466-472 [PMID: 2898465 DOI: 10.1111/j.1349-7006.1988.tb01615.x]
- 99 Toshkov I, Hacker HJ, Roggendorf M, Bannasch P. Phenotypic patterns of preneoplastic and neoplastic hepatic lesions in woodchucks infected with woodchuck hepatitis virus. J Cancer Res Clin Oncol 1990; 116: 581-590 [PMID: 2152341 DOI: 10.1007/BF01637078]
- Radaeva S, Li Y, Hacker HJ, Burger V, Kopp-Schneider A, Bannasch P. Hepadnaviral 100 hepatocarcinogenesis: in situ visualization of viral antigens, cytoplasmic compartmentation, enzymic patterns, and cellular proliferation in preneoplastic hepatocellular lineages in woodchucks. J Hepatol 2000; **33**: 580-600 [PMID: 11059863 DOI: 10.1034/j.1600-0641.2000.033004580.x]
- 101 Tu T, Mason WS, Clouston AD, Shackel NA, McCaughan GW, Yeh MM, Schiff ER, Ruszkiewicz AR, Chen JW, Harley HA, Stroeher UH, Jilbert AR. Clonal expansion of hepatocytes with a selective advantage occurs during all stages of chronic hepatitis B virus infection. J Viral Hepat 2015; 22: 737-753 [PMID: 25619231 DOI: 10.1111/jvh.12380]
- Mason WS, Liu C, Aldrich CE, Litwin S, Yeh MM. Clonal expansion of normal-appearing human 102 hepatocytes during chronic hepatitis B virus infection. J Virol 2010; 84: 8308-8315 [PMID: 20519397 DOI: 10.1128/JVI.00833-10]
- Cullen JM, Linzey DW, Gebhard DH. Nuclear ploidy of normal and neoplastic hepatocytes from 103 woodchuck hepatitis virus-infected and uninfected woodchucks. Hepatology 1994; 19: 1072-1078 [PMID: 8175128]
- 104 Jacob JR, Sterczer A, Toshkov IA, Yeager AE, Korba BE, Cote PJ, Buendia MA, Gerin JL, Tennant BC. Integration of woodchuck hepatitis and N-myc rearrangement determine size and histologic grade of hepatic tumors. Hepatology 2004; 39: 1008-1016 [PMID: 15057905 DOI: 10.1002/hep.20106]
- Lee JS, Chu IS, Heo J, Calvisi DF, Sun Z, Roskams T, Durnez A, Demetris AJ, Thorgeirsson SS. 105 Classification and prediction of survival in hepatocellular carcinoma by gene expression profiling. Hepatology 2004; 40: 667-676 [PMID: 15349906 DOI: 10.1002/hep.20375]
- 106 Hoshida Y, Nijman SM, Kobayashi M, Chan JA, Brunet JP, Chiang DY, Villanueva A, Newell P, Ikeda K, Hashimoto M, Watanabe G, Gabriel S, Friedman SL, Kumada H, Llovet JM, Golub TR. Integrative transcriptome analysis reveals common molecular subclasses of human hepatocellular carcinoma. Cancer Res 2009; 69: 7385-7392 [PMID: 19723656 DOI:



10.1158/0008-5472.CAN-09-1089]

- 107 Marion PL, Van Davelaar MJ, Knight SS, Salazar FH, Garcia G, Popper H, Robinson WS. Hepatocellular carcinoma in ground squirrels persistently infected with ground squirrel hepatitis virus. Proc Natl Acad Sci USA 1986; 83: 4543-4546 [PMID: 3012572 DOI: 10.1073/pnas.83.12.4543
- 108 Jayant K, Habib N, Huang KW, Warwick J, Arasaradnam R. Recent Advances: The Imbalance of Immune Cells and Cytokines in the Pathogenesis of Hepatocellular Carcinoma. Diagnostics (Basel) 2020; 10 [PMID: 32466214 DOI: 10.3390/diagnostics10050338]
- 109 Nakamoto Y, Guidotti LG, Kuhlen CV, Fowler P, Chisari FV. Immune pathogenesis of hepatocellular carcinoma. J Exp Med 1998; 188: 341-350 [PMID: 9670046 DOI: 10.1084/jem.188.2.341]
- Terradillos O, Billet O, Renard CA, Levy R, Molina T, Briand P, Buendia MA. The hepatitis B 110 virus X gene potentiates c-myc-induced liver oncogenesis in transgenic mice. Oncogene 1997; 14: 395-404 [PMID: 9053836 DOI: 10.1038/sj.onc.1200850]
- Madden CR, Finegold MJ, Slagle BL. Hepatitis B virus X protein acts as a tumor promoter in 111 development of diethylnitrosamine-induced preneoplastic lesions. J Virol 2001; 75: 3851-3858 [PMID: 11264374 DOI: 10.1128/JVI.75.8.3851-3858.2001]
- Feitelson MA, Lee J. Hepatitis B virus integration, fragile sites, and hepatocarcinogenesis. Cancer 112 Lett 2007; 252: 157-170 [PMID: 17188425 DOI: 10.1016/j.canlet.2006.11.010]
- 113 Fowler MJ, Greenfield C, Chu CM, Karayiannis P, Dunk A, Lok AS, Lai CL, Yeoh EK, Monjardino JP, Wankya BM. Integration of HBV-DNA may not be a prerequisite for the maintenance of the state of malignant transformation. An analysis of 110 liver biopsies, J Hepatol 1986; 2: 218-229 [PMID: 3958473 DOI: 10.1016/s0168-8278(86)80080-0]
- 114 Mason WS, Gill US, Litwin S, Zhou Y, Peri S, Pop O, Hong ML, Naik S, Quaglia A, Bertoletti A, Kennedy PT. HBV DNA Integration and Clonal Hepatocyte Expansion in Chronic Hepatitis B Patients Considered Immune Tolerant. Gastroenterology 2016; 151: 986-998. e4 [PMID: 27453547 DOI: 10.1053/j.gastro.2016.07.012]
- Levrero M, Zucman-Rossi J. Mechanisms of HBV-induced hepatocellular carcinoma. J Hepatol 115 2016; 64: S84-S101 [PMID: 27084040 DOI: 10.1016/j.jhep.2016.02.021]
- 116 Chauhan R, Michalak TI. Kinetics of DNA damage repair response accompanying initial hepadnavirus-host genomic integration in woodchuck hepatitis virus infection of hepatocyte. Cancer Genet 2020; 244: 1-10 [PMID: 32062411 DOI: 10.1016/j.cancergen.2020.02.001]
- 117 Freitas N, Lukash T, Gunewardena S, Chappell B, Slagle BL, Gudima SO. Relative Abundance of Integrant-Derived Viral RNAs in Infected Tissues Harvested from Chronic Hepatitis B Virus Carriers. J Virol 2018; 92 [PMID: 29491161 DOI: 10.1128/JVI.02221-17]
- Freitas N, Cunha C, Menne S, Gudima SO. Envelope proteins derived from naturally integrated 118 hepatitis B virus DNA support assembly and release of infectious hepatitis delta virus particles. J Virol 2014; 88: 5742-5754 [PMID: 24623409 DOI: 10.1128/JVI.00430-14]
- 119 Wei Y, Fourel G, Ponzetto A, Silvestro M, Tiollais P, Buendia MA. Hepadnavirus integration: mechanisms of activation of the N-myc2 retrotransposon in woodchuck liver tumors. J Virol 1992; 66: 5265-5276 [PMID: 1323693 DOI: 10.1128/JVI.66.9.5265-5276.1992]
- 120 Hansen LJ, Tennant BC, Seeger C, Ganem D. Differential activation of myc gene family members in hepatic carcinogenesis by closely related hepatitis B viruses. Mol Cell Biol 1993; 13: 659-667 [PMID: 8380230 DOI: 10.1128/mcb.13.1.659]
- 121 Bruni R, Conti I, Villano U, Giuseppetti R, Palmieri G, Rapicetta M. Lack of WHV integration nearby N-myc2 and in the downstream b3n and win loci in a considerable fraction of liver tumors with activated N-myc2 from naturally infected wild woodchucks. Virology 2006; 345: 258-269 [PMID: 16271377 DOI: 10.1016/j.virol.2005.09.061]
- 122 Bruni R, D'Ugo E, Villano U, Fourel G, Buendia MA, Rapicetta M. The win locus involved in activation of the distal N-myc2 gene upon WHV integration in woodchuck liver tumors harbors S/MAR elements. Virology 2004; 329: 1-10 [PMID: 15476869 DOI: 10.1016/j.virol.2004.08.008]
- 123 Yang D, Faris R, Hixson D, Affigne S, Rogler CE. Insulin-like growth factor II blocks apoptosis of N-myc2-expressing woodchuck liver epithelial cells. J Virol 1996; 70: 6260-6268 [PMID: 8709253 DOI: 10.1128/JVI.70.9.6260-6268.1996]
- 124 Ueda K, Ganem D. Apoptosis is induced by N-myc expression in hepatocytes, a frequent event in hepadnavirus oncogenesis, and is blocked by insulin-like growth factor II. J Virol 1996; 70: 1375-1383 [PMID: 8627653 DOI: 10.1128/JVI.70.3.1375-1383.1996]
- 125 Renard CA, Fourel G, Bralet MP, Degott C, De La Coste A, Perret C, Tiollais P, Buendia MA. Hepatocellular carcinoma in WHV/N-myc2 transgenic mice: oncogenic mutations of beta-catenin and synergistic effect of p53 null alleles. Oncogene 2000; 19: 2678-2686 [PMID: 10851067 DOI: 10.1038/sj.onc.1203617
- 126 Ura S, Honda M, Yamashita T, Ueda T, Takatori H, Nishino R, Sunakozaka H, Sakai Y, Horimoto K, Kaneko S. Differential microRNA expression between hepatitis B and hepatitis C leading disease progression to hepatocellular carcinoma. Hepatology 2009; 49: 1098-1112 [PMID: 19173277 DOI: 10.1002/hep.22749]
- 127 Connolly E, Melegari M, Landgraf P, Tchaikovskaya T, Tennant BC, Slagle BL, Rogler LE, Zavolan M, Tuschl T, Rogler CE. Elevated expression of the miR-17-92 polycistron and miR-21 in hepadnavirus-associated hepatocellular carcinoma contributes to the malignant phenotype. Am J Pathol 2008; 173: 856-864 [PMID: 18688024 DOI: 10.2353/ajpath.2008.080096]



- Chen HS, Kaneko S, Girones R, Anderson RW, Hornbuckle WE, Tennant BC, Cote PJ, Gerin JL, 128 Purcell RH, Miller RH. The woodchuck hepatitis virus X gene is important for establishment of virus infection in woodchucks. J Virol 1993; 67: 1218-1226 [PMID: 8437213 DOI: 10.1128/JVI.67.3.1218-1226.1993]
- 129 Ng SA, Lee C. Hepatitis B virus X gene and hepatocarcinogenesis. J Gastroenterol 2011; 46: 974-990 [PMID: 21647825 DOI: 10.1007/s00535-011-0415-9]
- 130 Wang WH, Studach LL, Andrisani OM. Proteins ZNF198 and SUZ12 are down-regulated in hepatitis B virus (HBV) X protein-mediated hepatocyte transformation and in HBV replication. Hepatology 2011; 53: 1137-1147 [PMID: 21480320 DOI: 10.1002/hep.24163]
- 131 Studach LL, Menne S, Cairo S, Buendia MA, Hullinger RL, Lefrançois L, Merle P, Andrisani OM. Subset of Suz12/PRC2 target genes is activated during hepatitis B virus replication and liver carcinogenesis associated with HBV X protein. Hepatology 2012; 56: 1240-1251 [PMID: 22505317 DOI: 10.1002/hep.25781]
- Ryu SH, Chung YH, Lee H, Kim JA, Shin HD, Min HJ, Seo DD, Jang MK, Yu E, Kim KW. 132 Metastatic tumor antigen 1 is closely associated with frequent postoperative recurrence and poor survival in patients with hepatocellular carcinoma. Hepatology 2008; 47: 929-936 [PMID: 18306220 DOI: 10.1002/hep.22124]
- Li YT, Liu CJ, Su TH, Cheng HR, Jeng YM, Lin HL, Wang CC, Kao JH, Chen PJ, Chen DS, Wu 133 HL. Characterization of metastatic tumor antigen 1 and its interaction with hepatitis B virus X protein in NF-κB signaling and tumor progression in a woodchuck hepatocellular carcinoma model. Oncotarget 2016; 7: 47173-47185 [PMID: 27323415 DOI: 10.18632/oncotarget.9986]
- Tseng PL, Tai MH, Huang CC, Wang CC, Lin JW, Hung CH, Chen CH, Wang JH, Lu SN, Lee CM, 134 Changchien CS, Hu TH. Overexpression of VEGF is associated with positive p53 immunostaining in hepatocellular carcinoma (HCC) and adverse outcome of HCC patients. J Surg Oncol 2008; 98: 349-357 [PMID: 18646041 DOI: 10.1002/jso.21109]
- 135 Finn RS, Zhu AX. Targeting angiogenesis in hepatocellular carcinoma: focus on VEGF and bevacizumab. Expert Rev Anticancer Ther 2009; 9: 503-509 [PMID: 19374603 DOI: 10.1586/era.09.6]
- Huang H, Salavaggione O, Rivera L, Mukherjee S, Brekken R, Tennant B, Iyer R, Adjei A. 136 Woodchuck VEGF (wVEGF) characteristics: Model for angiogenesis and human hepatocellular carcinoma directed therapies. Arch Biochem Biophys 2019; 661: 97-106 [PMID: 30439360 DOI: 10.1016/j.abb.2018.11.008
- 137 Altadill A, Rodríguez M, González LO, Junquera S, Corte MD, González-Dieguez ML, Linares A, Barbón E, Fresno-Forcelledo M, Rodrigo L, Vizoso FJ. Liver expression of matrix metalloproteases and their inhibitors in hepatocellular carcinoma. Dig Liver Dis 2009; 41: 740-748 [PMID: 19372066 DOI: 10.1016/j.dld.2009.01.016]
- 138 Ochoa-Callejero L, Toshkov I, Menne S, Martínez A. Expression of matrix metalloproteinases and their inhibitors in the woodchuck model of hepatocellular carcinoma. J Med Virol 2013; 85: 1127-1138 [PMID: 23595580 DOI: 10.1002/jmv.23571]
- Tao K, Qian N, Tang Y, Ti Z, Song W, Cao D, Dou K. Increased expression of a disintegrin and 139 metalloprotease-9 in hepatocellular carcinoma: implications for tumor progression and prognosis. Jpn J Clin Oncol 2010; 40: 645-651 [PMID: 20388695 DOI: 10.1093/jjco/hyq030]
- 140 Qin LX, Tang ZY. The prognostic molecular markers in hepatocellular carcinoma. World J Gastroenterol 2002; 8: 385-392 [PMID: 12046056 DOI: 10.3748/wjg.v8.i3.385]
- Fattovich G, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: incidence and 141 risk factors. Gastroenterology 2004; 127: S35-S50 [PMID: 15508101 DOI: 10.1053/j.gastro.2004.09.014]
- 142 Rizzetto M. Hepatitis D Virus: Introduction and Epidemiology. Cold Spring Harb Perspect Med 2015; 5: a021576 [PMID: 26134842 DOI: 10.1101/cshperspect.a021576]
- Taylor JM. Virology of hepatitis D virus. Semin Liver Dis 2012; 32: 195-200 [PMID: 22932967 143 DOI: 10.1055/s-0032-1323623]
- 144 Freitas N, Salisse J, Cunha C, Toshkov I, Menne S, Gudima SO. Hepatitis delta virus infects the cells of hepadnavirus-induced hepatocellular carcinoma in woodchucks. Hepatology 2012; 56: 76-85 [PMID: 22334419 DOI: 10.1002/hep.25663]
- 145 Shiga J, Ohnishi S, Imawari M, Yamamoto K, Koshimizu K, Sasaki N. Development and growth pattern of small hepatocellular carcinomas in woodchucks--analysis of an animal model of human hepatocellular carcinoma by ultrasonography. Jikken Dobutsu 1991; 40: 545-548 [PMID: 1748173 DOI: 10.1538/expanim1978.40.4\_545]
- 146 Lisi D, Kondili LA, Ramieri MT, Giuseppetti R, Bruni R, Della Rocca C, De Santis A, Rapicetta M. Ultrasonography in the study of hepatocellular carcinoma in woodchucks chronically infected with WHV. Lab Anim 2003; 37: 233-240 [PMID: 12869286 DOI: 10.1258/002367703766453083]
- 147 McKenzie EJ, Jackson M, Sun J, Volotovskyy V, Gruwel ML. Monitoring the development of hepatocellular carcinoma in woodchucks using 31P-MRS. MAGMA 2005; 18: 201-205 [PMID: 16133593 DOI: 10.1007/s10334-005-0120-x]
- 148 McKenzie EJ, Jackson M, Turner A, Gregorash L, Harapiak L. Chronic care and monitoring of woodchucks (Marmota monax) during repeated magnetic resonance imaging of the liver. J Am Assoc Lab Anim Sci 2006; 45: 26-30 [PMID: 16542039]
- 149 Hornbuckle WE, Graham ES, Roth L, Baldwin BH, Wickenden C, Tennant BC. Laboratory assessment of hepatic injury in the woodchuck (Marmota monax). Lab Anim Sci 1985; 35: 376-381



[PMID: 2864472]

- 150 Cote PJ, Gerin JL, Tennant BC. Alpha-fetoprotein in the woodchuck model of hepadnavirus infection and disease: normal physiological patterns and responses to woodchuck hepatitis virus infection and hepatocellular carcinoma. Cancer Res 1990; 50: 7843-7851 [PMID: 1701355]
- 151 Dahmen U, Radtke A, Schröder T, Chi H, Madrahimov N, Lu M, Schenk A, Peitgen KH, Dirsch O. Median liver lobe of woodchuck as a model to study hepatic outflow obstruction: a pilot study. Liver Int 2008; 28: 1236-1244 [PMID: 18544125 DOI: 10.1111/j.1478-3231.2008.01797.x]
- 152 Mikhail AS, Mauda-Havakuk M, Partanen A, Karanian JW, Pritchard WF, Wood BJ. Liver-specific 3D sectioning molds for correlating in vivo CT and MRI with tumor histopathology in woodchucks (Marmota monax). PLoS One 2020; 15: e0230794 [PMID: 32214365 DOI: 10.1371/journal.pone.0230794]
- 153 Nachabé R, Hendriks BH, Schierling R, Hales J, Racadio JM, Rottenberg S, Ruers TJ, Babic D. Real-Time In Vivo Characterization of Primary Liver Tumors With Diffuse Optical Spectroscopy During Percutaneous Needle Interventions: Feasibility Study in Woodchucks. Invest Radiol 2015; 50: 443-448 [PMID: 25783227 DOI: 10.1097/RLI.00000000000149]
- Aravalli RN, Golzarian J, Cressman EN. Animal models of cancer in interventional radiology. Eur 154 Radiol 2009; 19: 1049-1053 [PMID: 19137307 DOI: 10.1007/s00330-008-1263-8]
- 155 Mak IW, Evaniew N, Ghert M. Lost in translation: animal models and clinical trials in cancer treatment. Am J Transl Res 2014; 6: 114-118 [PMID: 24489990]
- Rose SC, Halstead GD, Narsinh KH. Pressure-Directed Embolization of Hepatic Arteries in a 156 Porcine Model Using a Temporary Occlusion Balloon Microcatheter: Proof of Concept. Cardiovasc Intervent Radiol 2017; 40: 1769-1776 [PMID: 28748354 DOI: 10.1007/s00270-017-1753-7]
- 157 Kim AY, Yacoub JH, Field DH, Park BU, Kallakury B, Korolowicz KE, Menne S. Suitability of the woodchuck HCC as a preclinical model for evaluation of intra-arterial therapies. Animal Model Exp Med 2020; 3: 98-102 [PMID: 32318666 DOI: 10.1002/ame2.12100]
- Wilkins LR, Stone JR, Mata J, Hawrylack A, Kubicka E, Brautigan DL. The Use of the Woodchuck 158 as an Animal Model for Evaluation of Transarterial Embolization. J Vasc Interv Radiol 2017: 28: 1467-1471 [PMID: 28941521 DOI: 10.1016/j.jvir.2017.04.005]
- Pritchard WF, Woods DL, Esparza-Trujillo JA, Starost MF, Mauda-Havakuk M, Mikhail AS, 159 Bakhutashvili I, Leonard S, Jones EC, Krishnasamy V, Karanian JW, Wood BJ. Transarterial Chemoembolization in a Woodchuck Model of Hepatocellular Carcinoma. J Vasc Interv Radiol 2020; **31**: 812-819. e1 [PMID: 32107125 DOI: 10.1016/j.jvir.2019.08.031]
- 160 Thorn CF. Oshiro C. Marsh S. Hernandez-Boussard T. McLeod H. Klein TE. Altman RB. Doxorubicin pathways: pharmacodynamics and adverse effects. Pharmacogenet Genomics 2011; 21: 440-446 [PMID: 21048526 DOI: 10.1097/FPC.0b013e32833ffb56]
- 161 Lin Y, Xu J, Lan H. Tumor-associated macrophages in tumor metastasis: biological roles and clinical therapeutic applications. J Hematol Oncol 2019; 12: 76 [PMID: 31300030 DOI: 10.1186/s13045-019-0760-3
- 162 Liu LY, Ma XZ, Ouyang B, Ings DP, Marwah S, Liu J, Chen AY, Gupta R, Manuel J, Chen XC, Gage BK, Cirlan I, Khuu N, Chung S, Camat D, Cheng M, Sekhon M, Zagorovsky K, Abdou Mohamed MA, Thoeni C, Atif J, Echeverri J, Kollmann D, Fischer S, Bader GD, Chan WCW, Michalak TI, McGilvray ID, MacParland SA. Nanoparticle Uptake in a Spontaneous and Immunocompetent Woodchuck Liver Cancer Model. ACS Nano 2020; 14: 4698-4715 [PMID: 32255624 DOI: 10.1021/acsnano.0c00468]
- Zoulim F. Evaluation of novel strategies to combat hepatitis B virus targetting wild-type and drugresistant mutants in experimental models. Antivir Chem Chemother 2001; 12 Suppl 1: 131-142 [PMID: 11594680]
- Peek SF, Toshkov IA, Erb HN, Schinazi RF, Korba BE, Cote PJ, Gerin JL, Tennant BC. 3'-164 Thiacytidine (3TC) delays development of hepatocellular carcinoma (HCC) in woodchucks with experimentally induced chronic woodchuck hepatitis virus (WHV) infection. Preliminary results of a lifetime study. Hepatology 1997; 26 Suppl: 368A
- 165 Mason WS, Cullen J, Moraleda G, Saputelli J, Aldrich CE, Miller DS, Tennant B, Frick L, Averett D, Condreay LD, Jilbert AR. Lamivudine therapy of WHV-infected woodchucks. Virology 1998; 245: 18-32 [PMID: 9614864 DOI: 10.1006/viro.1998.9150]
- Tatti KM, Korba BE, Stang HL, Peek S, Gerin JL, Tennant BC, Schinazi RF. Mutations in the 166 conserved woodchuck hepatitis virus polymerase FLLA and YMDD regions conferring resistance to lamivudine. Antiviral Res 2002; 55: 141-150 [PMID: 12076758 DOI: 10.1016/s0166-3542(02)00019-0
- Zhou T, Saputelli J, Aldrich CE, Deslauriers M, Condreay LD, Mason WS. Emergence of drug-167 resistant populations of woodchuck hepatitis virus in woodchucks treated with the antiviral nucleoside lamivudine. Antimicrob Agents Chemother 1999; 43: 1947-1954 [PMID: 10428918 DOI: 10.1128/AAC.43.8.1947
- Lau DT, Khokhar MF, Doo E, Ghany MG, Herion D, Park Y, Kleiner DE, Schmid P, Condreay LD, 168 Gauthier J, Kuhns MC, Liang TJ, Hoofnagle JH. Long-term therapy of chronic hepatitis B with lamivudine. Hepatology 2000; 32: 828-834 [PMID: 11003630 DOI: 10.1053/jhep.2000.17912]
- 169 Lok AS, Hussain M, Cursano C, Margotti M, Gramenzi A, Grazi GL, Jovine E, Benardi M, Andreone P. Evolution of hepatitis B virus polymerase gene mutations in hepatitis B e antigennegative patients receiving lamivudine therapy. Hepatology 2000; 32: 1145-1153 [PMID: 11050068 DOI: 10.1053/jhep.2000.19622]



- 170 Liu X, Schinazi RF. Hepatitis B virus resistance to lamivudine and its clinical implications. Antivir Chem Chemother 2002; 13: 143-155 [PMID: 12448687 DOI: 10.1177/095632020201300301]
- 171 Colonno RJ, Genovesi EV, Medina I, Lamb L, Durham SK, Huang ML, Corey L, Littlejohn M, Locarnini S, Tennant BC, Rose B, Clark JM. Long-term entecavir treatment results in sustained antiviral efficacy and prolonged life span in the woodchuck model of chronic hepatitis infection. J Infect Dis 2001; 184: 1236-1245 [PMID: 11679911 DOI: 10.1086/324003]
- 172 Korba BE, Cote PJ, Menne S, Toshkov I, Baldwin BH, Wells FV, Tennant BC, Gerin JL. Clevudine therapy with vaccine inhibits progression of chronic hepatitis and delays onset of hepatocellular carcinoma in chronic woodchuck hepatitis virus infection. Antivir Ther 2004; 9: 937-952 [PMID: 15651753
- 173 Menne S, Tumas DB, Liu KH, Thampi L, AlDeghaither D, Baldwin BH, Bellezza CA, Cote PJ, Zheng J, Halcomb R, Fosdick A, Fletcher SP, Daffis S, Li L, Yue P, Wolfgang GH, Tennant BC. Sustained efficacy and seroconversion with the Toll-like receptor 7 agonist GS-9620 in the Woodchuck model of chronic hepatitis B. J Hepatol 2015; 62: 1237-1245 [PMID: 25559326 DOI: 10.1016/j.jhep.2014.12.026]
- Daffis S, Balsitis S, Chamberlain J, Zheng J, Santos R, Rowe W, Ramakrishnan D, Pattabiraman D, 174 Spurlock S, Chu R, Kang D, Mish M, Ramirez R, Li L, Li B, Ma S, Hung M, Voitenleitner C, Yon C, Suresh M, Menne S, Cote P, Delaney WE 4th, Mackman R, Fletcher SP. Toll-Like Receptor 8 Agonist GS-9688 Induces Sustained Efficacy in the Woodchuck Model of Chronic Hepatitis B. Hepatology 2021; 73: 53-67 [PMID: 32246499 DOI: 10.1002/hep.31255]
- 175 Fairman J, Liu KH, Menne S. Prevention of liver tumor formation in woodchucks with established hepatocellular carcinoma by treatment with cationic liposome-DNA complexes. BMC Cancer 2017; 17: 172 [PMID: 28264666 DOI: 10.1186/s12885-017-3163-2]
- 176 BREEDIS C, YOUNG G. The blood supply of neoplasms in the liver. Am J Pathol 1954; 30: 969-977 [PMID: 13197542]
- Leal JN, Kingham TP. Hepatic artery infusion chemotherapy for liver malignancy. Surg Oncol Clin 177 N Am 2015; 24: 121-148 [PMID: 25444472 DOI: 10.1016/j.soc.2014.09.005]
- 178 Kerr DJ, Los G. Pharmacokinetic principles of locoregional chemotherapy. Cancer Surv 1993; 17: 105-122 [PMID: 8137338]
- Kim M, Powers CA, Curtin LI, Fisher DT, Sexton S, Gurova KV, Skitzki JJ, Iver RV. A 179 Translational Hepatic Artery Infusion (HAI) Model for Hepatocellular Carcinoma in Woodchucks. J Surg Res 2020; 251: 126-136 [PMID: 32143057 DOI: 10.1016/j.jss.2020.02.002]
- 180 Fleyshman D, Prendergast L, Safina A, Paszkiewicz G, Commane M, Morgan K, Attwood K, Gurova K. Level of FACT defines the transcriptional landscape and aggressive phenotype of breast cancer cells. Oncotarget 2017; 8: 20525-20542 [PMID: 28423528 DOI: 10.18632/oncotarget.15656]
- Gasparian AV, Burkhart CA, Purmal AA, Brodsky L, Pal M, Saranadasa M, Bosykh DA, 181 Commane M, Guryanova OA, Pal S, Safina A, Sviridov S, Koman IE, Veith J, Komar AA, Gudkov AV, Gurova KV. Curaxins: anticancer compounds that simultaneously suppress NF-kB and activate p53 by targeting FACT. Sci Transl Med 2011; 3: 95ra74 [PMID: 21832239 DOI: 10.1126/scitranslmed.3002530
- 182 Iver RV, Maguire O, Kim M, Curtin LI, Sexton S, Fisher DT, Schihl SA, Fetterly G, Menne S, Minderman H. Dose-Dependent Sorafenib-Induced Immunosuppression Is Associated with Aberrant NFAT Activation and Expression of PD-1 in T Cells. Cancers (Basel) 2019; 11 [PMID: 31100868 DOI: 10.3390/cancers11050681]
- 183 Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008; 359: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
- Kianmanesh R, Ogata S, Paradis V, Sauvanet A, Belghiti J. Heat-zone effect after surface 184 application of dissecting sealer on the "in situ margin" after tumorectomy for liver tumors. J Am Coll Surg 2008; 206: 1122-1128 [PMID: 18501809 DOI: 10.1016/j.jamcollsurg.2007.12.006]
- 185 Burke CT, Cullen JM, State A, Gadi S, Wilber K, Rosenthal M, Bulysheva A, Pease A, Mauro MA, Fuchs H. Development of an animal model for radiofrequency ablation of primary, virally induced hepatocellular carcinoma in the woodchuck. J Vasc Interv Radiol 2011; 22: 1613-1618. e1 [PMID: 21959057 DOI: 10.1016/j.jvir.2011.08.020]
- Cheng CW, Machtay M, Dorth J, Sergeeva O, Xia H, Manaspon C, Wu H, Iyer R, Sexton S, Xin W, 186 Exner AA, Lee Z. Delayed response to proton beam treatment of hepatocellular carcinoma. BJR Case Rep 2020; 6: 20180125 [PMID: 32201597 DOI: 10.1259/bjrcr.20180125]
- 187 Bilbao R, Gérolami R, Bralet MP, Qian C, Tran PL, Tennant B, Prieto J, Bréchot C. Transduction efficacy, antitumoral effect, and toxicity of adenovirus-mediated herpes simplex virus thymidine kinase/ ganciclovir therapy of hepatocellular carcinoma: the woodchuck animal model. Cancer Gene Ther 2000; 7: 657-662 [PMID: 10830712 DOI: 10.1038/sj.cgt.7700175]
- 188 Rodriguez-Madoz JR, Liu KH, Quetglas JI, Ruiz-Guillen M, Otano I, Crettaz J, Butler SD, Bellezza CA, Dykes NL, Tennant BC, Prieto J, González-Aseguinolaza G, Smerdou C, Menne S. Semliki forest virus expressing interleukin-12 induces antiviral and antitumoral responses in woodchucks with chronic viral hepatitis and hepatocellular carcinoma. J Virol 2009; 83: 12266-12278 [PMID: 19740992 DOI: 10.1128/JVI.01597-09]
- Pützer BM, Stiewe T, Rödicker F, Schildgen O, Rühm S, Dirsch O, Fiedler M, Damen U, Tennant 189



B, Scherer C, Graham FL, Roggendorf M. Large nontransplanted hepatocellular carcinoma in woodchucks: treatment with adenovirus-mediated delivery of interleukin 12/B7.1 genes. J Natl Cancer Inst 2001; 93: 472-479 [PMID: 11259473 DOI: 10.1093/jnci/93.6.472]

- 190 Huang KW, Wu HL, Lin HL, Liang PC, Chen PJ, Chen SH, Lee HI, Su PY, Wu WH, Lee PH, Hwang LH, Chen DS. Combining antiangiogenic therapy with immunotherapy exerts better therapeutical effects on large tumors in a woodchuck hepatoma model. Proc Natl Acad Sci USA 2010; 107: 14769-14774 [PMID: 20679198 DOI: 10.1073/pnas.1009534107]
- Liu Y, Wang B, Wang L, Vikash V, Wang Q, Roggendorf M, Lu M, Yang D, Liu J. Transcriptome 191 Analysis and Comparison of Marmota monax and Marmota himalavana, PLoS One 2016; 11: e0165875 [PMID: 27806133 DOI: 10.1371/journal.pone.0165875]
- 192 Fletcher SP, Chin DJ, Gruenbaum L, Bitter H, Rasmussen E, Ravindran P, Swinney DC, Birzele F, Schmucki R, Lorenz SH, Kopetzki E, Carter J, Triyatni M, Thampi LM, Yang J, AlDeghaither D, Murreddu MG, Cote P, Menne S. Intrahepatic Transcriptional Signature Associated with Response to Interferon-a Treatment in the Woodchuck Model of Chronic Hepatitis B. PLoS Pathog 2015; 11: e1005103 [PMID: 26352406 DOI: 10.1371/journal.ppat.1005103]
- 193 Alioto TS, Cruz F, Gómez-Garrido J, Triyatni M, Gut M, Frias L, Esteve-Codina A, Menne S, Kiialainen A, Kumpesa N, Birzele F, Schmucki R, Gut IG, Spleiss O. The Genome Sequence of the Eastern Woodchuck (Marmota monax) - A Preclinical Animal Model for Chronic Hepatitis B. G3 (Bethesda) 2019; 9: 3943-3952 [PMID: 31645421 DOI: 10.1534/g3.119.400413]
- 194 Korba BE, Cote P, Hornbuckle W, Tennant BC, Gerin JL. Treatment of chronic woodchuck hepatitis virus infection in the Eastern woodchuck (Marmota monax) with nucleoside analogues is predictive of therapy for chronic hepatitis B virus infection in humans. Hepatology 2000; 31: 1165-1175 [PMID: 10796894 DOI: 10.1053/he.2000.5982]
- 195 Goldberg BB, Hilpert PL, Burns PN, Liu JB, Newman LM, Merton DA, Witlin LA. Hepatic tumors: signal enhancement at Doppler US after intravenous injection of a contrast agent. Radiology 1990; 177: 713-717 [PMID: 2173841 DOI: 10.1148/radiology.177.3.2173841]
- 196 Forsberg F, Goldberg BB, Liu JB, Merton DA, Rawool NM, Shi WT. Tissue-specific US contrast agent for evaluation of hepatic and splenic parenchyma. Radiology 1999; 210: 125-132 [PMID: 9885597 DOI: 10.1148/radiology.210.1.r99ja11125]
- 197 Forsberg F, Liu JB, Merton DA, Rawool NM, Goldberg BB. Parenchymal enhancement and tumor visualization using a new sonographic contrast agent. J Ultrasound Med 1995; 14: 949-957 [PMID: 8583531 DOI: 10.7863/jum.1995.14.12.949]
- Forsberg F, Roy R, Merton DA, Rawool NM, Liu JB, Huang M, Kessler D, Goldberg BB. 198 Conventional and hypobaric activation of an ultrasound contrast agent. Ultrasound Med Biol 1998; 24: 1143-1150 [PMID: 9833583 DOI: 10.1016/s0301-5629(98)00062-3]
- 199 Goldberg BB, Liu JB, Burns PN, Merton DA, Forsberg F. Galactose-based intravenous sonographic contrast agent: experimental studies. J Ultrasound Med 1993; 12: 463-470 [PMID: 8411330 DOI: 10.7863/jum.1993.12.8.463
- 200 Matsumura T, Moriyasu F, Kono Y, Chiba T. [Contrast-enhanced power Doppler imaging of the liver--preliminary animal study]. Nihon Rinsho 1998; 56: 985-989 [PMID: 9577621]
- 201 Nada T, Moriyasu F, Kono Y, Matsumura T. [Sonographic depiction of woodchuck hepatomas using intravenously injected contrast agents]. Nihon Rinsho 1998; 56: 980-984 [PMID: 9577620]
- 202 Forsberg F, Liu JB, Chiou HJ, Rawool NM, Parker L, Goldberg BB. Comparison of fundamental and wideband harmonic contrast imaging of liver tumors. Ultrasonics 2000; 38: 110-113 [PMID: 10829639 DOI: 10.1016/s0041-624x(99)00149-3]
- Forsberg F, Goldberg BB, Liu JB, Merton DA, Rawool NM. On the feasibility of real-time, in vivo 203 harmonic imaging with proteinaceous microspheres. J Ultrasound Med 1996; 15: 853-60: guiz 861 [PMID: 8947861 DOI: 10.7863/jum.1996.15.12.853]
- 204 Kono Y, Moriyasu F, Nada T, Suginoshita Y, Matsumura T, Kobayashi K, Nakamura T, Chiba T. Gray scale second harmonic imaging of the liver: a preliminary animal study. Ultrasound Med Biol 1997; 23: 719-726 [PMID: 9253819 DOI: 10.1016/s0301-5629(97)00007-0]
- 205 Kallfelz FA, Hornbuckle WE, Harvey HJ, Wallace RJ, Potkewitz LG, Roth L, Tennant BC. Scintigraphic diagnosis of primary hepatocellular carcinoma in the woodchuck (Marmota monax). Am J Vet Res 1986; 47: 573-576 [PMID: 3008602]
- 206 Salem N, Kuang Y, Wang F, Maclennan GT, Lee Z. PET imaging of hepatocellular carcinoma with 2-deoxy-2[18F]fluoro-D-glucose, 6-deoxy-6[18F] fluoro-D-glucose, [1-11C]-acetate and [N-methyl-11C]-choline. Q J Nucl Med Mol Imaging 2009; 53: 144-156 [PMID: 19039303]
- 207 Salem N, Kuang Y, Corn D, Erokwu B, Kolthammer JA, Tian H, Wu C, Wang F, Wang Y, Lee Z. [(Methyl)1-(11)c]-acetate metabolism in hepatocellular carcinoma. Mol Imaging Biol 2011; 13: 140-151 [PMID: 20401538 DOI: 10.1007/s11307-010-0308-y]
- 208 Tenley N, Corn DJ, Yuan L, Lee Z. The effect of fasting on PET Imaging of Hepatocellular Carcinoma. J Cancer Ther 2013; 4: 561-567 [PMID: 24683497 DOI: 10.4236/jct.2013.42071]
- Kuang Y, Salem N, Tian H, Kolthammer JA, Corn DJ, Wu C, Wang F, Wang Y, Lee Z. Imaging 209 lipid synthesis in hepatocellular carcinoma with [methyl-11c]choline: correlation with in vivo metabolic studies. J Nucl Med 2011; 52: 98-106 [PMID: 21149484 DOI: 10.2967/jnumed.110.080366]
- Kolthammer JA, Corn DJ, Tenley N, Wu C, Tian H, Wang Y, Lee Z. PET imaging of 210 hepatocellular carcinoma with 18F-fluoroethylcholine and 11C-choline. Eur J Nucl Med Mol Imaging 2011; 38: 1248-1256 [PMID: 21344223 DOI: 10.1007/s00259-011-1743-y]



- 211 Sergeeva O, Kepe V, Zhang Y, Miller-Atkins GA, Keynon JD, Iyer R, Sexton S, Awadallah A, Xin W, Saunthararajah Y, Chan ER, Lee Z. [18F] Clofarabine for PET Imaging of Hepatocellular Carcinoma. Cancers (Basel) 2019; 11 [PMID: 31703407 DOI: 10.3390/cancers11111748]
- 212 Sergeeva O, Zhang Y, Kenyon JD, Miller-Atkins GA, Wu C, Iyer R, Sexton S, Wojtylak P, Awadallah A, Xin W, Chan ER, O'Donnel JK, Lee Z. PET imaging of hepatocellular carcinoma with anti-1-amino-3-[18F]fluorocyclobutanecarboxylic acid in comparison with L-[S-methyl-11 C]methionine. EJNMMI Res 2019; 9: 47 [PMID: 31119488 DOI: 10.1186/s13550-019-0519-4]
- 213 Salem N, MacLennan GT, Kuang Y, Anderson PW, Schomisch SJ, Tochkov IA, Tennant BC, Lee Z. Quantitative evaluation of 2-deoxy-2[F-18]fluoro-D-glucose-positron emission tomography imaging on the woodchuck model of hepatocellular carcinoma with histological correlation. Mol Imaging Biol 2007; 9: 135-143 [PMID: 17308952 DOI: 10.1007/s11307-007-0092-5]
- 214 Sergeeva O, Zhang Y, Kenyon J, Miller-Atkins G, Sergeev M, Verbus E, Iyer R, Sexton S, Kepe V, Avril N, Saunthararajah Y, Chan ER, Lee Z. Liver background uptake of [18F]FLT in PET imaging. Am J Nucl Med Mol Imaging 2020; 10: 212-225 [PMID: 33224617]
- 215 Ohtomo K, Shiga J, Sasaki Y, Itai Y. [Iron oxide-enhanced MR imaging of hepatocellular carcinoma of woodchuck]. Nihon Igaku Hoshasen Gakkai Zasshi 1991; 51: 433-435 [PMID: 1648709
- Reimer P, Weissleder R, Brady TJ, Yeager AE, Baldwin BH, Tennant BC, Wittenberg J. 216 Experimental hepatocellular carcinoma: MR receptor imaging. Radiology 1991; 180: 641-645 [PMID: 1871273 DOI: 10.1148/radiology.180.3.1871273]
- Cullen JM, Li DH, Brown C, Eisenberg EJ, Cundy KC, Wolfe J, Toole J, Gibbs C. Antiviral efficacy and pharmacokinetics of oral adefovir dipivoxil in chronically woodchuck hepatitis virusinfected woodchucks. Antimicrob Agents Chemother 2001; 45: 2740-2745 [PMID: 11557463 DOI: 10.1128/AAC.45.10.2740-2745.2001
- Jacob JR, Korba BE, Cote PJ, Toshkov I, Delaney WE 4th, Gerin JL, Tennant BC. Suppression of 218 lamivudine-resistant B-domain mutants by adefovir dipivoxil in the woodchuck hepatitis virus model. Antiviral Res 2004; 63: 115-121 [PMID: 15302140 DOI: 10.1016/j.antiviral.2004.03.005]
- 219 Menne S, Butler SD, George AL, Tochkov IA, Zhu Y, Xiong S, Gerin JL, Cote PJ, Tennant BC. Antiviral effects of lamivudine, emtricitabine, adefovir dipivoxil, and tenofovir disoproxil fumarate administered orally alone and in combination to woodchucks with chronic woodchuck hepatitis virus infection. Antimicrob Agents Chemother 2008; 52: 3617-3632 [PMID: 18676881 DOI: 10.1128/AAC.00654-08]
- Zhu Y, Yamamoto T, Cullen J, Saputelli J, Aldrich CE, Miller DS, Litwin S, Furman PA, Jilbert 220 AR, Mason WS. Kinetics of hepadnavirus loss from the liver during inhibition of viral DNA synthesis. J Virol 2001; 75: 311-322 [PMID: 11119601 DOI: 10.1128/JVI.75.1.311-322.2001]
- 221 Jacquard AC, Nassal M, Pichoud C, Ren S, Schultz U, Guerret S, Chevallier M, Werle B, Peyrol S, Jamard C, Rimsky LT, Trepo C, Zoulim F. Effect of a combination of clevudine and emtricitabine with adenovirus-mediated delivery of gamma interferon in the woodchuck model of hepatitis B virus infection. Antimicrob Agents Chemother 2004; 48: 2683-2692 [PMID: 15215126 DOI: 10.1128/AAC.48.7.2683-2692.2004]
- 222 Peek SF, Cote PJ, Jacob JR, Toshkov IA, Hornbuckle WE, Baldwin BH, Wells FV, Chu CK, Gerin JL, Tennant BC, Korba BE. Antiviral activity of clevudine [L-FMAU, (1-(2-fluoro-5-methyl-beta, L-arabinofuranosyl) uracil)] against woodchuck hepatitis virus replication and gene expression in chronically infected woodchucks (Marmota monax). Hepatology 2001; 33: 254-266 [PMID: 11124844 DOI: 10.1053/jhep.2001.20899]
- 223 Cullen JM, Smith SL, Davis MG, Dunn SE, Botteron C, Cecchi A, Linsey D, Linzey D, Frick L, Paff MT, Goulding A, Biron K. In vivo antiviral activity and pharmacokinetics of (-)-cis-5-fluoro-1-[2-(hydroxymethyl)-1,3-oxathiolan-5-yl]cytosine in woodchuck hepatitis virus-infected woodchucks. Antimicrob Agents Chemother 1997; 41: 2076-2082 [PMID: 9333028 DOI: 10.1128/AAC.41.10.2076
- Korba BE, Schinazi RF, Cote P, Tennant BC, Gerin JL. Effect of oral administration of 224 emtricitabine on woodchuck hepatitis virus replication in chronically infected woodchucks. Antimicrob Agents Chemother 2000; 44: 1757-1760 [PMID: 10817750 DOI: 10.1128/aac.44.6.1757-1760.2000
- Korolowizc KE, Li B, Huang X, Yon C, Rodrigo E, Corpuz M, Plouffe DM, Kallakury BV, Suresh 225 M, Wu TY, Miller AT, Menne S. Liver-Targeted Toll-Like Receptor 7 Agonist Combined With Entecavir Promotes a Functional Cure in the Woodchuck Model of Hepatitis B Virus. Hepatol Commun 2019; 3: 1296-1310 [PMID: 31592075 DOI: 10.1002/hep4.1397]
- 226 Menne S, Wildum S, Steiner G, Suresh M, Korolowicz K, Balarezo M, Yon C, Murreddu M, Hong X, Kallakury BV, Tucker R, Yang S, Young JAT, Javanbakht H. Efficacy of an Inhibitor of Hepatitis B Virus Expression in Combination With Entecavir and Interferon-a in Woodchucks Chronically Infected With Woodchuck Hepatitis Virus. Hepatol Commun 2020; 4: 916-931 [PMID: 32490326 DOI: 10.1002/hep4.1502]
- 227 Suresh M, Korolowicz KE, Balarezo M, Iyer RP, Padmanabhan S, Cleary D, Gimi R, Sheri A, Yon C, Kallakury BV, Tucker RD, Afdhal N, Menne S. Antiviral Efficacy and Host Immune Response Induction during Sequential Treatment with SB 9200 Followed by Entecavir in Woodchucks. PLoS One 2017; 12: e0169631 [PMID: 28056062 DOI: 10.1371/journal.pone.0169631]
- 228 Berraondo P, Di Scala M, Korolowicz K, Thampi LM, Otano I, Suarez L, Fioravanti J, Aranda F, Ardaiz N, Yang J, Kallakury BV, Tucker RD, Vasquez M, Menne S, Prieto J, González-



Aseguinolaza G. Liver-directed gene therapy of chronic hepadnavirus infection using interferon alpha tethered to apolipoprotein A-I. J Hepatol 2015; 63: 329-336 [PMID: 25772035 DOI: 10.1016/j.jhep.2015.02.048]

- 229 Korba BE, Cote P, Hornbuckle W, Schinazi R, Gangemi JD, Tennant BC, Gerin JL. Enhanced antiviral benefit of combination therapy with lamivudine and alpha interferon against WHV replication in chronic carrier woodchucks. Antivir Ther 2000; 5: 95-104 [PMID: 10971862]
- 230 Standring DN, Bridges EG, Placidi L, Faraj A, Loi AG, Pierra C, Dukhan D, Gosselin G, Imbach JL, Hernandez B, Juodawlkis A, Tennant B, Korba B, Cote P, Cretton-Scott E, Schinazi RF, Myers M, Bryant ML, Sommadossi JP. Antiviral beta-L-nucleosides specific for hepatitis B virus infection. Antivir Chem Chemother 2001; 12 Suppl 1: 119-129 [PMID: 11594678]
- 231 Bryant ML, Bridges EG, Placidi L, Faraj A, Loi AG, Pierra C, Dukhan D, Gosselin G, Imbach JL, Hernandez B, Juodawlkis A, Tennant B, Korba B, Cote P, Marion P, Cretton-Scott E, Schinazi RF, Sommadossi JP. Antiviral L-nucleosides specific for hepatitis B virus infection. Antimicrob Agents Chemother 2001; 45: 229-235 [PMID: 11120971 DOI: 10.1128/AAC.45.1.229-235.2001]
- 232 Bryant ML, Bridges EG, Placidi L, Faraj A, Loi AG, Pierra C, Dukhan D, Gosselin G, Imbach JL, Hernandez B, Juodawlkis A, Tennant B, Korba B, Cote P, Cretton-Scott E, Schinazi RF, Sommadossi JP. Anti-HBV specific beta-L-2'-deoxynucleosides. Nucleosides Nucleotides Nucleic Acids 2001; 20: 597-607 [PMID: 11563077 DOI: 10.1081/NCN-100002336]
- 233 Ahn B, Lee BC, Kim HJ, Park S, Lee Y, Choi Y, Kim HE, Kim YW. hzVSF, a novel HBV therapeutic candidate, shows WHsAg loss in woodchuck hepatitis model and safety in phase I clinical study. J Hepatol 2020; 73 Suppl 1: S861 [DOI: 10.1016/S0168-8278(20)32164-4]
- 234 Meng Z, Zhang X, Pei R, Zhang E, Kemper T, Vollmer J, Davis HL, Glebe D, Gerlich W, Roggendorf M, Lu M. Combination therapy including CpG oligodeoxynucleotides and entecavir induces early viral response and enhanced inhibition of viral replication in a woodchuck model of chronic hepadnaviral infection. Antiviral Res 2016; 125: 14-24 [PMID: 26585244 DOI: 10.1016/j.antiviral.2015.11.001]
- 235 Paulsen D, Weber O, Ruebsamen-Schaeff H, Tennant BC, Menne S. AIC649 Induces a Bi-Phasic Treatment Response in the Woodchuck Model of Chronic Hepatitis B. PLoS One 2015; 10: e0144383 [PMID: 26656974 DOI: 10.1371/journal.pone.0144383]
- 236 Liu J, Zhang E, Ma Z, Wu W, Kosinska A, Zhang X, Möller I, Seiz P, Glebe D, Wang B, Yang D, Lu M, Roggendorf M. Enhancing virus-specific immunity in vivo by combining therapeutic vaccination and PD-L1 blockade in chronic hepadnaviral infection. PLoS Pathog 2014; 10: e1003856 [PMID: 24391505 DOI: 10.1371/journal.ppat.1003856]





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