World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2021 September 15; 13(9): 980-1212





Published by Baishideng Publishing Group Inc

World Journal of Gastrointestinal Oncology

Contents

Monthly Volume 13 Number 9 September 15, 2021

REVIEW

980	Involvement of integrin-activating peptides derived from tenascin-C in colon cancer progression
	Fujita M, Suzuki H, Fukai F
995	MicroRNA expression in inflammatory bowel disease-associated colorectal cancer Grillo TG, Quaglio AEV, Beraldo RF, Lima TB, Baima JP, Di Stasi LC, Sassaki LY
1017	Association between intestinal neoplasms and celiac disease: A review
	Wang M, Yu M, Kong WJ, Cui M, Gao F
1029	Real-time fluorescence image-guided gastrointestinal oncologic surgery: Towards a new era Martínez-López E, Martínez-Pérez A, Navarro-Martínez S, Sebastián-Tomás JC, de'Angelis N, García-Granero E
1043	Neoadjuvant chemotherapy for colorectal liver metastases: A contemporary review of the literature
	Guo M, Jin N, Pawlik T, Cloyd JM
	MINIREVIEWS

1062 Review of incomplete macroscopic resections (R2) in rectal cancer: Treatment, prognosis and future perspectives Pérez Lara FJ, Hebrero Jimenez ML, Moya Donoso FJ, Hernández Gonzalez JM, Pitarch Martinez M, Prieto-Puga Arjona Т

1073 Potential utility of liquid biopsies in the management of patients with biliary tract cancers: A review Shotton R, Lamarca A, Valle J, McNamara MG

1086 Conservative management of malignant gastric outlet obstruction syndrome-evidence based evaluation of endoscopic ultrasound-guided gastroentero-anastomosis Cominardi A, Tamanini G, Brighi N, Fusaroli P, Lisotti A

1099 Overgrowth of Lactobacillus in gastric cancer Li ZP, Liu JX, Lu LL, Wang LL, Xu L, Guo ZH, Dong QJ

Evidence based tools to improve efficiency of currently administered oncotherapies for tumors of the 1109 hepatopancreatobiliary system

Herold Z, Szasz AM, Dank M

1121 Screening strategy for gastrointestinal and hepatopancreatobiliary cancers in cystic fibrosis Hoskins B, Wasuwanich P, Scheimann AO, Karnsakul W

Immune aspects of hepatocellular carcinoma: From immune markers for early detection to 1132 immunotherapy

Mattos ÂZ, Debes JD, Boonstra A, Vogel A, Mattos AA



I

World Journal of Gastrointestinal Oncology Contents Monthly Volume 13 Number 9 September 15, 2021 1144 Characterization of metabolic landscape in hepatocellular carcinoma Wu J, Xue R, Jiang RT, Meng QH 1157 Effect of oncometabolic surgery on gastric cancer: The remission of hypertension, type 2 diabetes mellitus, and beyond Cheng YX, Peng D, Tao W, Zhang W **ORIGINAL ARTICLE Basic Study** Scoparone inhibits pancreatic cancer through PI3K/Akt signaling pathway 1164 Li N, Yang F, Liu DY, Guo JT, Ge N, Sun SY **Retrospective Study** Prognostic value of modified Lauren classification in gastric cancer 1184 Ning FL, Zhang NN, Wang J, Jin YF, Quan HG, Pei JP, Zhao Y, Zeng XT, Abe M, Zhang CD

META-ANALYSIS

Neoadjuvant chemotherapy without radiation as a potential alternative treatment for locally advanced 1196 rectal cancer: A meta-analysis

Wu P, Xu HM, Zhu Z

LETTER TO THE EDITOR

1210 Use of liquid biopsies in gastrointestinal cancers

Khachfe HH

Contents

Monthly Volume 13 Number 9 September 15, 2021

ABOUT COVER

Editorial Board Member of World Journal of Gastrointestinal Oncology, Rossana Berardi, MD, PhD, Director, Full Professor, Medical Oncology, Università Politecnica delle Marche, Ancona 60126, Italy. r.berardi@staff.univpm.it

AIMS AND SCOPE

The primary aim of World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

INDEXING/ABSTRACTING

The WJGO is now indexed in Science Citation Index Expanded (also known as SciSearch®), PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Reports® cites the 2020 impact factor (IF) for WJGO as 3.393; IF without journal self cites: 3.333; 5-year IF: 3.519; Journal Citation Indicator: 0.5; Ranking: 163 among 242 journals in oncology; Quartile category: Q3; Ranking: 60 among 92 journals in gastroenterology and hepatology; and Quartile category: Q3. The WJGO's CiteScore for 2020 is 3.3 and Scopus CiteScore rank 2020: Gastroenterology is 70/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ying-Yi Yuan; Production Department Director: Xiang Li; Editorial Office Director: Ya-Juan Ma.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Gastrointestinal Oncology	https://www.wignet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1948-5204 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
February 15, 2009	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Monthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Rosa M Jimenez Rodriguez, Pashtoon Kasi, Monjur Ahmed, Florin Burada	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/1948-5204/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
September 15, 2021	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



0 W J

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 September 15; 13(9): 995-1016

DOI: 10.4251/wjgo.v13.i9.995

ISSN 1948-5204 (online)

REVIEW

MicroRNA expression in inflammatory bowel disease-associated colorectal cancer

Thais Gagno Grillo, Ana Elisa Valencise Quaglio, Rodrigo Fedatto Beraldo, Talles Bazeia Lima, Julio Pinheiro Baima, Luiz Claudio Di Stasi, Ligia Yukie Sassaki

ORCID number: Thais Gagno Grillo 0000-0002-4351-5034; Ana Elisa Valencise Quaglio 0000-0002-5998-2382; Rodrigo Fedatto Beraldo 0000-0002-8398-5014; Talles Bazeia Lima 0000-0001-6523-7541; Julio Pinheiro Baima 0000-0002-4035-3113; Luiz Claudio Di Stasi 0000-0002-7864-1073; Ligia Yukie Sassaki 0000-0002-7319-8906.

Author contributions: All authors contributed to this manuscript; Grillo TG, Quaglio AEV, Beraldo RF, Lima TB, Baima JP, Di Stasi LC and Sassaki LY contributed to the conception of the study, acquisition, analysis and interpretation of data, drafting the article, revising it critically for important intellectual content, and final approval of the version to be submitted.

Conflict-of-interest statement: The authors state that they have no conflicts of interest regarding this review.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build

Thais Gagno Grillo, Rodrigo Fedatto Beraldo, Talles Bazeia Lima, Julio Pinheiro Baima, Ligia Yukie Sassaki, Department of Internal Medicine, São Paulo State University (Unesp), Medical School, Botucatu 18618-686, São Paulo, Brazil

Ana Elisa Valencise Quaglio, Luiz Claudio Di Stasi, Department of Biophysics and Pharmacology, São Paulo State University (Unesp), Institute of Biosciences, Botucatu 18618-689, São Paulo, Brazil

Corresponding author: Ligia Yukie Sassaki, MD, PhD, Academic Research, Assistant Professor, Department of Internal Medicine, São Paulo State University (Unesp), Medical School, s/n. Bairro: Rubião Junior, Botucatu 18618-686, São Paulo, Brazil. ligia.sassaki@unesp.br

Abstract

MicroRNAs (miRNAs) are non-coding RNA molecules composed of 19-25 nucleotides that regulate gene expression and play a central role in the regulation of several immune-mediated disorders, including inflammatory bowel diseases (IBD). IBD, represented by ulcerative colitis and Crohn's disease, is characterized by chronic intestinal inflammation associated with an increased risk of colorectal cancer (CRC). CRC is one of the most prevalent tumors in the world, and its main risk factors are obesity, physical inactivity, smoking, alcoholism, advanced age, and some eating habits, in addition to chronic intestinal inflammatory processes and the use of immunosuppressants administered to IBD patients. Recent studies have identified miRNAs associated with an increased risk of developing CRC in this population. The identification of miRNAs involved in this tumorigenic process could be useful to stratify cancer risk development for patients with IBD and to monitor and assess prognosis. Thus, the present review aimed to summarize the role of miRNAs as biomarkers for the diagnosis and prognosis of IBD-associated CRC. In the future, therapies based on miRNA modulation could be used both in clinical practice to achieve remission of the disease and restore the quality of life for patients with IBD, and to identify the patients with IBD at high risk for tumor development.

Key Words: MicroRNA; Colorectal cancer; Inflammatory bowel disease; Crohn's disease; Ulcerative colitis; Cancer; Diagnosis; Prognosis; Targets



upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Brazil

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: February 21, 2021 Peer-review started: February 21, 2021 First decision: May 8, 2021 Revised: May 30, 2021

Accepted: July 27, 2021 Article in press: July 27, 2021 Published online: September 15, 2021

P-Reviewer: Raoul W S-Editor: Ma YJ L-Editor: A P-Editor: Li JH



©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Inflammatory bowel diseases (IBD), represented by ulcerative colitis and Crohn's disease, are characterized by recurrent chronic intestinal inflammation associated with an increased risk of colorectal cancer. MicroRNAs (miRNAs) are small non-coding RNA molecules, composed of 19 to 25 nucleotides, which can regulate gene expression and play an important role in regulating cellular processes. Altered expression of these molecules is related to the progression of inflammation and an increased risk of colorectal cancer (CRC). Thus, the aim of the present review was to evaluate the role of miRNAs as biomarkers for the diagnosis and prognosis of IBDassociated CRC.

Citation: Grillo TG, Quaglio AEV, Beraldo RF, Lima TB, Baima JP, Di Stasi LC, Sassaki LY. MicroRNA expression in inflammatory bowel disease-associated colorectal cancer. World J Gastrointest Oncol 2021; 13(9): 995-1016

URL: https://www.wjgnet.com/1948-5204/full/v13/i9/995.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i9.995

INTRODUCTION

MicroRNAs (miRNAs) are non-coding RNA molecules composed of approximately 19–25 nucleotides that are capable of regulating gene expression[1]. MiRNAs are involved in biological functions such as embryonic development, proliferation, cell differentiation, metabolism, apoptosis, and stress response[2,3]. The isolation of miRNAs is possible using various biological materials such as tissues, cells, and body fluids (tears, urine, serum, and plasma)[4]. Polymerase chain reaction, in situ hybridization, microarrays, and RNA sequencing are the main methods used to detect miRNA expression[1].

MiRNAs can downregulate mRNA^[5] by binding to the 3' untranslated region of the target mRNAs[6]. As only one miRNA can regulate many mRNA targets, any minimal structural change can lead to major changes in cell homeostasis[7], disease evolution [8], and predisposition to neoplastic and inflammatory conditions[9-11]. Alterations in miRNA expression have been described in several tumors, including colorectal cancer (CRC) acting as oncogenes or tumor suppressors[12]. This expression is specific according to the tumor type and the surrounding tissue; thus, the study of tumor miRNAs helps to differentiate normal and tumor tissues and, in addition, reflects the degree of tumor differentiation[13].

Inflammatory bowel diseases (IBD), such as ulcerative colitis (UC) and Crohn's disease (CD), are characterized by chronic intestinal inflammation associated with an increased risk of CRC. Tumor development is one of the most feared long-term disease complications, accounting for 15% of deaths in patients with IBD[14]. UC patients are approximately 30 times more likely to develop CRC than the general population[15], the main risk factors being the extension and duration of the disease[16], family history of CRC[17], and the presence of primary sclerosing cholangitis (PSC)[18]. Likewise, patients with CD are at a higher risk of small bowel cancer^[19].

In IBD studies, miRNAs have been found to be involved in pathogenesis and can serve as diagnostic biomarkers and therapeutic targets. Recent studies have identified miRNAs associated with an increased risk of developing CRC in this population. The identification of miRNAs involved in this tumorigenic process could be useful to stratify cancer risk development for patients with IBD and to monitor and assess prognosis. Thus, this review article aimed to characterize the miRNAs expressed in IBD, CRC, and IBD-related CRC to better understand their role in the diagnosis and prognosis of these diseases, in addition to analyzing their potential as therapeutic targets.

INFLAMMATORY BOWEL DISEASE AND MICRORNA EXPRESSION

IBD is a chronic disease involving the gastrointestinal tract, the pathophysiology of



which is complex, encompassing environmental, genetic factors and the immune response[20], and comprises two specific diseases, UC and CD[20-22]. There is no specific examination to establish the diagnosis of CD and UC, which requires the association of clinical, biochemical, endoscopic, radiological, and histologic data[23]. Thus, a distinction between these two diseases can be challenging when inflammation is limited to the colon, and 10%–15% of IBD patients are classified as having indeterminate colitis[24].

In recent years, new biomarkers associated with easy application have been identified, such as miRNAs, to facilitate disease diagnosis and prognosis[25]. However, the real role of miRNAs in IBD is not fully understood, and it is believed that some miRNAs might be common to both diseases, whereas others are unique to each, depending on the severity of disease activity and the tissue analyzed[26]. Changes in miRNA expression in patients with IBD were first described by Wu et al [27] in 2008. The authors showed an increase in the expression of eight miRNAs (Let-7f, miR-16, miR-21, miR-23a, miR-24, miR-29a, miR-126, and miR-195) and a decrease in the expression of three miRNAs (miR-192, miR-375, and miR-422b) in colonic tissue samples from patients with active UC. In 2010, the same group[28] observed increased expression of miR-23b, miR106a, and miR-191 and decreased expression of miR-19b and miR-629 in colonic samples of patients with colonic CD and increased expression of miR-16, miR-21, miR-223, and miR-594 in terminal ileum samples of CD patients with active disease. In UC patients, miR-19a, miR-21, miR-31, miR-146a and miR-375 levels were found increased when compared with CD patients indicating these miRNAs as potential differential biomarkers for CD and UC[29,30]. In addition to these studies, others studies have demonstrated changes in miRNA expression in the colonic tissue of patients with IBD[31,32].

Guo *et al*[31] evaluated the differential expression of miRNA in inflamed or noninflamed ileum mucosa of patients with CD and found decreased expression of miR-192-5p in those with inflammation. Among the alterations in the expression of miRNAs already observed, miR-10a, miR-192, and miR-320 seem to negatively regulate the inflammatory response by inhibiting the expression of *NOD2* mRNA (domain 2 of nucleotide-binding oligomerization)[33-35]. In contrast, miR-155, by activating the nuclear factor kappa B (NF- κ B) signaling pathway, plays an important role in the progression of intestinal inflammation[36]. Besides that, Lu *et al*[37] showed that miR-155 was capable to decrease SH-2 containing inositol 5' polyphosphatase 1 (SHIP-1), an important phosphatase correlated with membrane trafficking, contributing to inflammation pathogenesis.

Shi *et al*[38] demonstrated that the miR-31/interleukin (IL)-25 signaling axis can regulate the Th1/Th17 IL-12/23-mediated inflammatory response in experimental colitis, indicating that a decrease in miR-31 expression with a consequent increase in IL-25 levels could be an alternative treatment for IBD. This pathway is also intricately linked to miR-155 and miR-223[39,40].

Pre-clinical studies have also been performed to evaluate the role of miRNAs in intestinal inflammation. Nata et al[41] administered miR-146b intraperitoneally to mice with dextran sodium sulfate (DSS)-induced intestinal inflammation and observed an improvement in the inflammatory process and intestinal barrier, demonstrating a potential use of miRNAs for IBD treatment[41]. Additionally, Huang et al[42] observed the regulation of leukocyte infiltration and consequently a reduction in the inflammatory process with a intracolonic injection of miR-141 in mice[42]. On the other hand, the use of an antagomir for miR-155, a small synthetic RNA complementary to miR-155 used to silence this miRNA, in the DSS-induced intestinal inflammation model improved intestinal inflammation indicating miR-155 as a possible target for IBD treatment[37]. Jin et al[43] using miRNA mimics, namely miR-133a to target UCP2 (mitochondrial uncoupling protein 2) observed a reduction in the severity of DSSinduced intestinal inflammation, suggesting that miRNA mimics are another therapeutic option for IBD patients[43]. In another study with the DSS-model of intestinal inflammation, Tian et al[44] found a super expression of miR-31, results similar to what was found in inflamed mucosa from patients with CD or UC.

Besides that, levels of miR-301a were also increased in intestinal epithelial cells from patients with active IBD reducing the expression of BTG anti-proliferation factor 1 (BTG1) and promoting Th17 cell differentiation through downregulation of Smad Nuclear Interacting Protein 1 (SNIP1). BTG1 reduces epithelial integrity and promote inflammation in mouse colon and leading to tumorigenesis. This way, blockade of miR-301a *in vivo* may serve as a novel therapeutic approach in the treatment of IBD and colitis associated-CRC[45,46].

Zaishidena® WJGO | https://www.wjgnet.com

Some miRNAs act on the same inflammatory pathways as drugs currently used to treat IBD. MiR-29 has been described to comprise a family of miRNAs with the ability to decrease IL-23 levels [47,48], effects similar to ustekinumab and others antiinterleukin 12/23 used for the treatment of moderate to severe IBD. The blockade of integrin $\alpha 4\beta 7$ in T helper lymphocytes has anti-inflammatory activity through the inhibition of leukocyte adhesion to endothelial cells. Harris et al [49] observed that endogenous miR-126 could inhibit this adhesion through the regulation of VCAM-1 adherence^[49,50], effects similar to those found with the use of vedolizumab, a monoclonal antibody that blocks integrin $\alpha 4\beta 7$. Likewise, Pathak *et al*[51] demonstrated that miR-155 targets suppressor of cytokine signaling 1 (SOCS1), a regulatory protein of the JAK signaling pathway[51], mimicking the use of JAK inhibitors currently available for UC treatment.

In addition to their possible application in clinical practice, some miRNAs can be used as predictors of the response to clinical treatment. Morilla *et al*[52] evaluated patients with severe UC and their response to corticosteroids, or infliximab and cyclosporine in those corticosteroid-refractory UC patients. The authors identified 15 miRNAs associated with the response to corticosteroids, six with the response to infliximab, and four with the response to cyclosporine, indicating that miRNAs can be used to screen patients according to the probability of responding to a specific medication. Cordes et al[53] evaluated the potential of miR-320a to monitor disease activity and predict the course of disease in patients with IBD, and found that blood levels of miR-320a were significantly increased in patients with active CD and UC when compared to healthy controls assessing the role of miRNA in monitoring inflammatory activity. Moreover, miR-320a levels were strongly correlated with endoscopic disease activity in both CD and UC patients, highlighting the use of miRNA as a noninvasive tool useful in monitoring disease activity in these patients.

New studies have evaluated the role miRNAs in fecal samples of IBD patients. Verdier et al[54] analyzed more than 800 miRNAs in fecal samples from control individuals and patients with IBD with levels of miR-223 and miR-1246 significantly increased in fecal samples from patients with active IBD. Furthermore, the miRNAs were correlated with clinical disease activity scores, such as the Mayo score, and fecal calprotectin levels in these patients, indicating that miRNAs from fecal samples might be a new noninvasive and easy fecal biomarker for monitoring IBD activity. MiR-223 was also identified as a fecal marker in a study by Schönauen et al^[25], in addition to miR-155 and miR-16. A summary of miRNAs found altered in each disease is shown in Table 1[55-61] and Table 2[62-69].

The future treatment of IBD involves the application of pharmacological strategies to control or even stop the progression of inflammation and to improve sensitivity to the therapy. This could occur, for example, through anti-miRNA oligonucleotides to inactivate miRNAs association with increased expression in the inflammatory process or increase the expression of suppressor miRNAs[70]. The alteration of immune system cells by miRNAs is also a factor for inflammation[71]. Thus, tracking the immune status in IBD based on miRNA alterations may be powerful for designing individualized therapies[72].

Taken together, these differences in the expression of miRNAs in UC and CD patients are relevant from the moment they lead to the emergence of biomarkers for diagnosis and therapeutic targets, aiming to improve the management of IBD; however, larger and more consistent studies are necessary for their implementation in clinical practice[60,72].

CRC AND MICRORNA EXPRESSION

CRC is the second most common cancer in women and the third in men, with higher rates in developed countries, and is responsible for approximately 900000 deaths each year[73]. An increase in the global incidence to 2.5 million new cases is expected in 2035[74,75], mainly due to an increase in exposure to risk factors. Obesity, physical inactivity, smoking, alcoholism, aging, and eating habits are some of the main risk factors for the appearance of tumors[73]. Genetic factors are also involved, such as the presence of a positive family history in 10%-20% of the cases [76] and hereditary syndromes in 5%-7% of the cases[77]. Patients with long-standing IBD constitute a risk group due to the presence of the inflammatory processes and the use of immunosuppressive drugs, with CRC being responsible for 15% of deaths in this population[14].

The development of CRC results from an evolutionary period of approximately 10-15 years and originates, in most cases, from alterations to the crypt pattern that



Table 1 Expression of altered miRNAs in colonic tissue in patients with Crohn's disease			
MiRNA	Expression	Ref.	
let-7i	Increased	[55]	
miR-7	Increased	[32]	
miR-9	Increased	[32]	
miR-16	Increased	[27,28]	
miR-19a	Increased	[29]	
miR-21	Increased	[9,28,32]	
miR-22	Increased	[32]	
miR-23b	Increased	[28]	
miR-29a	Increased	[27]	
miR-29b	Increased	[32]	
miR-29c	Increased	[32]	
miR-30a	Increased	[32]	
miR-30b	Increased	[32]	
miR-30c	Increased	[32]	
miR-31	Increased	[32,56]	
miR-34c-5p	Increased	[32]	
miR-101	Increased	[29]	
miR-106a	Increased	[28,32]	
miR-126	Increased	[32]	
miR-127-3p	Increased	[32]	
miR-130a	Increased	[32]	
miR-133b	Increased	[32]	
miR-141	Increased	[42]	
miR-146a	Increased	[29,32]	
miR-146b-5p	Increased	[32]	
miR-150	Increased	[32]	
miR-155	Increased	[32,56]	
miR-181c	Increased	[32]	
miR-191	Increased	[28]	
miR-196	Increased	[57]	
miR-196a	Increased	[32]	
miR-206	Increased	[58]	
miR-223	Increased	[28,32]	
miR-301a	Increased	[45]	
miR-324-3p	Increased	[32]	
miR-328	Increased	[59]	
miR-422a	Increased	[59]	
miR-449b	Increased	[55]	
miR-594	Increased	[28]	
miR 663	Increased	[58]	
miR 885-5p	Increased	[59]	



Grillo TG et al. MiRNA expression in IBD-associated CRC

let-7b	Decreased	[59]
miR-10a	Decreased	
		[33]
miR-18a	Decreased	[59]
miR-19b	Decreased	[28]
miR-26a	Decreased	[32]
miR-140-3p	Decreased	[59]
miR-143	Decreased	[60]
miR-192-5p	Decreased	[31]
miR-194b	Decreased	[58]
miR-203	Decreased	[61]
miR-216b	Decreased	[58]
miR-320a	Decreased	[34]
miR-320b	Decreased	[34]
miR-320c	Decreased	[34]
miR-375	Decreased	[27]
miR-548e	Decreased	[58]
miR-559	Decreased	[58]
miR-629	Decreased	[28]

evolves to a pre-neoplastic lesion (polyp) and later to a tumor [78,79]. Precursor lesions appear in two ways. The first is via adenoma-carcinoma, responsible for 70%–90% of tumors and related to an adenomatous polyposis coli (APC) gene mutation with the subsequent activation of RAS and loss of tumor suppressor p53 (TP53) function; the second is through a serrated neoplasia, which is responsible for 10%-20% of cases[73, 80] and is mainly related to RAS and RAF mutations[73].

It has also been noted that changes in cell homeostasis due to genetic changes lead to the activation of oncogenes and inactivation of tumor suppressor genes [78,79]. WNT signaling pathways, epidermal growth factor (EGFR), the TP53 complex, and transforming growth factor beta (TGF-β) are implicated in the carcinogenesis of CRC [81]. The WNT pathway is related to the regulation of stem cell activity in intestinal crypts, and inadvertent signaling by this pathway leads to the inhibition of cell differentiation and death, leading to the development of polyps and consequently carcinoma^[82,83].

It was noted that marked expression of miR-135b, which activates the WNT pathway is involved in sporadic and inflammatory CRC and is related to the tumor stage and a worse prognosis[84]. The EGFR signaling pathway is also responsible for cellular activities and is related to certain oncogenes, predominantly Kirsten rat sarcoma viral oncogene homolog (KRAS), for which mutations are present in approximately 30%–40% of CRC cases, resulting in worse prognosis[85]. It is also important to note that KRAS seems to be regulated by isoforms of the let-7 family, such as let-7a, which when deregulated, contributes to colorectal carcinogenesis[86].

Senescence, cell cycle arrest, apoptosis, invasion, and metastasis are related to TP53 when cells are subjected to stress^[87]. Some of the miRNAs that participate in the TP53 pathway include let-7i, miR-20a, miR-21, miR-25, miR-34a/b/c, miR-145, miR-181b, miR183, miR-195, miR-215, and miR-451 with a special attention with miR-34 family. Activation of TP53 by miR-34a has already been observed in several types of cancer, especially CRC, with a overexpression of this miRNA in those patients [88]. Finally, the TGF- β pathway regulates activities such as proliferation, differentiation, and apoptosis, and miRNAs that regulate the TGF- β receptor 2 (TGFBR2) have been identified, including miR-17-5p, miR-20a, miR-21, miR-23b, miR-106a, and miR-301a [89].

There are specific molecular expression profiles in CRC cells compared to those in non-tumor cells. Among the overexpressed miRNAs, miR-106, miR-31, miR-21, miR-25, miR-20a, miR-93, miR-183, and miR-203 have been identified, whereas those with reduced expression include miR-1, miR-126, miR-30a, miR-143, miR-145, miR-191, and miR-192[81]. A reduced expression of miR-192 seems to be related to an increase in



Table 2 Expression of altered miRNAs in the colonic tissue of patients with ulcerative colitis			
MiRNA	Expression	Ref.	
let-7e	Increased	[62]	
let-7f	Increased	[27]	
miR-7	Increased	[32]	
miR-16	Increased	[27,28]	
miR-19a	Increased	[29]	
miR-20b	Increased	[62]	
miR-21	Increased	[9,32]	
miR-23a	Increased	[27]	
miR-24	Increased	[27]	
miR-29a	Increased	[27]	
miR-29b	Increased	[32]	
miR-31	Increased	[32,56]	
miR-98	Increased	[62]	
miR-101	Increased	[29]	
miR-125b-1	Increased	[62]	
miR-126	Increased	[27,63]	
miR-127-3p	Increased	[32]	
miR-135b	Increased	[32]	
miR-146a	Increased	[29,32]	
miR-150	Increased	[64]	
miR-155	Increased	[32,56]	
miR-195	Increased	[27]	
miR-196a	Increased	[32]	
miR-206	Increased	[58]	
miR-214	Increased	[65]	
miR-223	Increased	[28,32]	
miR-301a	Increased	[45]	
miR-324-3p	Increased	[32]	
miR-548a-3p	Increased	[59]	
miR-650	Increased	[59]	
miR-663	Increased	[58]	
miR-10a	Decreased	[33]	
miR-26a	Decreased	[32]	
miR-124	Decreased	[66]	
miR-143	Decreased	[60]	
miR-145	Decreased	[60]	
miR-188-5p	Decreased	[32]	
miR-192	Decreased	[27]	
miR-194	Decreased	[67]	
miR-194b	Decreased	[58]	
miR-196b	Decreased	[59]	



Grillo TG et al. MiRNA expression in IBD-associated CRC

miR-200b	Decreased	[68]
miR-215	Decreased	[32]
miR-216b	Decreased	[58]
miR-320a	Decreased	[34]
miR-320b	Decreased	[34]
miR-320c	Decreased	[34]
miR-346	Decreased	[32]
miR-375	Decreased	[27]
miR-422b	Decreased	[27]
miR-489	Decreased	[59]
miR-548e	Decreased	[58]
miR-559	Decreased	[58]
miR-630	Decreased	[59]
miR-4284	Decreased	[69]

tumor size[90], and a reduced expression of miR-145 was determined to be related to invasion, metastasis, degree of differentiation, and tumor size[91], demonstrating the relationship between specific miRNAs and tumor behavior.

It is believed that there is a difference in miRNA expression based on the stage of the tumor. For example, overexpression of miR-92a can be a biomarker for the early diagnosis of CRC[92], whereas the overexpression of miR-21 and miR-31 is associated with advanced CRC[93,94]. According to that, Tsukamoto et al[95], demonstrated that the overexpression of exosomal miR-21 showed a significant association with liver metastasis and TNM stage in CRC patients being associated with a decrease in the overall survival and disease-free survival rates. Besides that, as a proangiogenic miRNA, miR-21 targets the programmed cell death protein 4 (PDCD4) gene enhancing invasion, intravasation and metastasis[96].

Giráldez et al[97], in turn, showed a positive correlation of appearance of distant metastasis in advanced CRC patients and miR-103 overexpression. Besides that, miR-29a also presented overexpressed in metastatic CRC patients when compared to nonmetastatic ones[98], and plasmatic expression of miR-203 and miR-141 could help in the differentiation of early and advanced CRC as demonstrated by Sun et al[99] In addition to these alterations correlated with stage of the tumor, there are also changes related to the response to treatment. MiR-1914-3p and miR-1915-3p were found downregulated in plasma samples from patients with chemo resistant CRC. This way, up-regulation of miR-1914-3p and -1915-3p reduces the chemoresistance abilities of chemo resistant CRC cells and may represent a possible therapy and diagnosis tool in CRC[100].

Alteration in angiogenesis is a contributing factor to tumor development, supporting proliferation, growth, dissemination, and metastasis[101,102]. MiRNAs are thought to participate as regulators of angiogenesis, acting both as antiangiogenic and proangiogenic[103,104], directly influencing endothelial cells or indirectly modulating protein expression [104], which makes them an interesting pathway in antiangiogenic therapies [105]. Non-responding bevacizumab (antibody anti-vascular endothelial growth factor A (VEGF-A) patients had increased levels of miR-126 correlated with an increase in tumor size[106]. On the other hand, miR-140-5p showed a tumor suppression role in CRC, targeting VEGF-A/MMP-2 pathway, and leading to inhibition of tumor progression and angiogenesis[107]. Additionally, miR-497 also blocks VEGF-A/ERK/MMP-9 pathway with reduction on angiogenesis, invasion, and metastasis in CRC[108].

Tumor growth depends on angiogenesis and the formation of new blood vessels, to ensure a continuous supply of oxygen and nutrients. This way, antiangiogenic agents are used to treat cancers, either alone or in combination[109]. However, the mechanistic details of how these combination therapies work are far from clear, and the lack of validated prognostic and predictive biomarkers represents one of the greatest obstacles in determining treatment outcomes and optimal response[109,110]. Based on that, miRNAs could serve as new predictive biomarker, therapeutic targets as an antiangiogenic therapy, or screening tools for immune-based therapies[111].



MiRNAs stability and predictive property make them ideal serum and plasma biomarkers in cancer patients, and they may be useful in predicting patterns of sensitivity and resistance to anti-cancer drugs[112]. This has brought attention to future personalized treatment strategies targeting miRNA expression in these patients. Thus, the identification of miRNAs as a tool for the early detection, prognostic evaluation, and treatment of CRC has gained clinical importance in recent years[81].

A summary of miRNAs found altered in CRC is shown in Table 3[113-141].

IBD, CRC, AND MICRORNA EXPRESSION

Chronic inflammation is a contributing factor to carcinogenesis; therefore, patients with IBD, and especially those with colonic involvement, are at an increased risk for CRC, which is responsible for approximately 15% of deaths in this population[14]. However, the exact mechanisms underlying this relationship have not yet been fully elucidated[20,142]. Among the risk factors involved, we can emphasize a positive family history of CRC, long disease duration, colonic involvement, the presence of PSC, and the presence of disease activity^[142]. For this reason, screening colonoscopy should be performed 8 years after the onset of symptoms in all patients with IBD to detect dysplasia in the early stages, with subsequent surveillance ranging from 1-5 years according to individual risk stratification[16]. Patients with IBD-associated PSC must undergo annual endoscopic surveillance after the diagnosis of PSC because of the increased risk of developing CRC[16].

The accumulation of reactive oxidative species from continuous cycles of inflammation and tissue repair results in damage to DNA, proteins, and lipids, leading to tumor development in patients with UC[143]. The progression of colitis-associated CRC occurs through processes that are perpetuated by the absence of dysplasia, undefined dysplasia, and low-and high-grade dysplasia until it progresses definitively to cancer^[144]. Therefore, despite adequate surveillance, detecting dysplasia in these patients is difficult owing to difficulties in distinguishing lesions from the adjacent inflamed mucosa and foci of constant tissue regeneration and repair caused by the inflammatory process^[145]. In 2010, O' Connor et al^[146] observed that chronic inflammation in these patients exposed them to tumorigenic traits based on the NF-KB pathways and inflammatory mediators such IL-6 and tumor necrosis factor-alpha $(TNF-\alpha)$. Oxidative stress, the activation of survival pathways, apoptosis, and the formation of a tumorigenic environment have been found to be involved in carcinogenesis. Changes in P53 gene occur early and can be detected before the onset of dysplasia, being identified in 47–85% of patients with colitis-associated CRC[146,147].

The release of cytokines resulting from chronic inflammation is related to the development of all stages of cancer, such as initiation, promotion, angiogenesis, and metastasis, and the transcription factor NF-KB is a primary factor in the inflammation/cancer cascade[146]. In addition to the involvement of molecular mediators in the inflammation/cancer relationship, such as cytokines, growth factors, Toll-like receptors (TLRs), Pl3K/MAPK signaling, and transcription factors (NF-KB/STAT3, P53, c-Myc, and Wnt/ β -catenin), among others[148], it is worth noting that cellular changes caused by a chronic inflammatory status in patients with IBD also contribute to the development of the tumor[149].

MiRNAs are believed to participate in the intestinal inflammation of IBD patients and contribute to the inflammation/tumor process. The identification of miRNAs involved in this tumorigenic process could be useful to stratify the risk of tumor development in patients with IBD and monitor and assess the prognosis of cases. Figure 1 and Table 4 shows the miRNAs found altered in UC, CD, CRC, and IBDassociated CRC summarizing the alterations described for each condition and the relationship between them.

The first study on the differential expression of miRNAs in IBD and CRCprogression from non-neoplastic mucosa to dysplasia and invasive cancer was published in 2012[150]. Using naïve immunotherapy patients from CD or UC and tree types of tissue (non-neoplastic, dysplastic and neoplastic) from each patient, the authors observed that five miRNAs (miR-193b, miR-373, let-7e, miR-15b, and miR-372) were significantly downregulated in both diseases, correlated with the progression from non-neoplastic tissue to dysplasia and from dysplasia to cancer[150]. In CD patients, during non-neoplastic to dysplasia progression, miR-181a, miR-146b-5p, let-7e, and miR-17 were found to be upregulated, on the other hand, during the progression from dysplasia to cancer, let-7e, miR-17, and miR-143 were downregulated. From the deregulated miRNAs, let-7e, miR-15b, miR-17, miR-122, miR-124, and

Mirna	Ref.
Let-7f	[27,113]
miR-1	[114,115]
miR-101	[29,116]
miR-103	[117]
miR-106a	[28,32,117]
miR-106b	[118,119]
miR-1000	[117,120]
miR-10a	[33,117]
miR-122	[115,121]
miR-126	[27,32,122]
miR-1288	[114,121]
miR-1305	[114,123]
miR-133b	[32,124]
miR-135b	[32,124]
miR-138	[116,123,125]
miR-139	[117]
miR-139-5p	[117]
miR-141	[42,121]
miR-143	[42,121]
miR-145	[60,128]
miR-147	[117]
miR-147	[113,117]
miR-140a	[32,114]
miR-155	[32,56,120]
	[27,28,116,124]
miR-16 miR-17	[124]
miR-17	[124]
miR-181a	[117]
miR-181	
	[117,121,124] [59,129]
miR-18a	
miR-191	[28,117]
miR-192	[27,90]
miR-194	[67,117,121]
miR-195	[27,117,130]
miR-196a	[32,124]
miR-19a	[29,117,118]
miR-19b	[28,113]
miR-200a	[117,121]
miR-200b	[68,121]
miR-200c	[117,121]
miR-203	[61,124]



miR-20a	[113,119,131]
miR-21	[9,32,132]
miR-210	[133]
miR-212	[133]
miR-214	[65,123]
miR-215	[32,90,121]
miR-218	[115]
miR-222	[117]
miR-223	[28,32,120,121]
miR-224	[117,121]
miR-23b	[28,114]
miR-25	[117,124]
miR-27a	[117]
miR-27b	[115]
miR-29a	[27,114,117]
miR-29b	[29,114]
miR-30a	[32,115]
miR-30c	[32,115,120]
miR-31	[32,56,128]
miR-32	[120,134]
miR-320a	[34,135]
miR-328	[59,134]
miR-34a	[136]
miR-375	[27,137]
miR-422a	[59,113,116]
miR-422b	[27,138]
miR-424	[118,123]
miR-451	[114,116]
miR-490-3p	[115,121,125]
miR-497	[115]
miR-506	[139]
miR-552	[121,134,140]
miR-650	[59]
miR-7	[32,123,140]
miR-885-5p	[59]
miR-892b	[115,121]
miR-92a	[119,141]
miR-93	[113,124]
miR-95	[117,123]
miR-96	[121]
miR-98	[62,113,117]

Baisbideng® WJGO | https://www.wjgnet.com

Table 4 MicroRNAs differentially expressed in ulcerative colitis, Crohn's disease and colorectal cancer, and shared by ulcerative colitis and Crohn's disease, ulcerative colitis and colorectal cancer, Crohn's disease and colorectal cancer and inflammatory bowel disease and colorectal cancer

Disease	Altered miRNAs
UC	miR-489 miR-630 miR-125b-1 let-7f miR-188-5p miR-196b miR-20b miR-346 miR-24 miR-548a-3p let-7e miR-23a miR-124 miR-4284
CD	let-7i miR-34c-5p miR-22 miR-9 miR-140-3p miR-196 miR-130a miR-30b miR-594 let-7b miR-29c miR-629 miR-449b miR-192-5p miR- 146b-5p
CRC	miR-497 miR-27b miR-92a miR-451 miR-20a miR-1 miR-506 Let-7f miR-181a miR-218 miR-32 miR-34a miR-106b miR-210 miR-93 miR- 148a miR-17 miR-25 miR-222 miR-103 miR-147 miR-212 miR-27a miR-107 miR-139
UC – CD	miR-320c miR-559 miR-216b miR-301a miR-320b miR-548e miR-146a miR-127-3p miR-324-3p miR-663 miR-194b miR-206 miR-26a
UC - CRC	miR-200b miR-145 miR-98 miR-214 miR-194 miR-195 miR-135b miR-215 miR-422b miR-650 miR-192
CD - CRC	miR-18a miR-19b miR-122 miR-328 miR-30a miR-106a miR-30c miR-23b miR-181c miR-422a miR-203 miR-141 miR-191 miR-133b miR- 885-5p
IBD - CRC	miR-21 miR-155 miR-200c miR-139-5p miR-223 miR-892b miR-7 miR-143 miR-150 miR-320a miR-10a miR-29b miR-196a miR-95 miR- 490-3p miR-16 miR-19a miR-126 miR-101 miR-1305 miR-424 miR-1288 miR-183 miR-96 miR-138 miR-375 miR-200a miR-29a miR-31 miR-224 miR-552

UC: Ulcerative colitis; CD: Crohn's disease; CRC: Colorectal cancer; IBD: Inflammatory bowel disease.

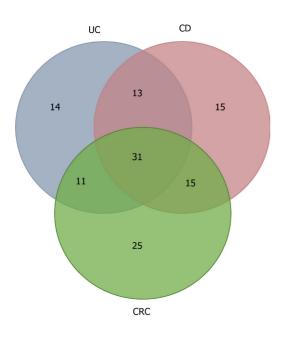


Figure 1 Venn diagram showing microRNAs differentially expressed in patients with ulcerative colitis (14), Crohn's disease (15), colorectal cancer (25), and, in the intersections, shared microRNA by ulcerative colitis and Crohn's disease (13), ulcerative colitis and colorectal cancer (11), Crohn's disease and colorectal cancer (15) and inflammatory bowel disease-associated colorectal cancer (31). UC: Ulcerative colitis; CD: Crohn's disease; CRC: Colorectal cancer.

miR-372 had a tumorigenic effect on the TP53 pathway[150].

Bai et al[151], integrated genome-wide gene expression profiles and biological pathway information to explore the associations among UC, CD and CRC at function and gene level, and found 34, 20 and 47 risk pathways for UC, CD and CRC, respectively. Furthermore, the authors found that UC and CD share 16 pathways, indicating that the two inflammatory diseases are strikingly linked with each other at the biological pathway level. On the other hand, more pathways were shared between CRC and UC compared to CRC and CD, which might suggest that UC has a potential functional link with CRC. Pathways for UC and CRC were mainly related to the immune system and metabolism like the Intestinal immune network for IgA production^[151].

When analyzing the correlation between miRNA and the risk pathways, four miRNAs participate in all three diseases (miR-146a, miR-335, miR-26b and miR-124). Targets of these four miRNAs were mainly associated with "signaling transduction",



Table 5 Enrichment analysis of the target genes of the altered microRNAs in inflammatory bowel disease-associated colorectal cancer			
Pathway	P value	Adjusted <i>P</i> value	
VEGFA-VEGFR2 signaling pathway	2.577e-31	1.203e-28	
TGF-beta signaling pathway	1.823e-23	4.257e-21	
EGF/EGFR signaling pathway	6.399e-20	9.961e-18	
DNA damage response (only ATM dependent)	8.953e-20	1.045e-17	
Oncostatin M signaling pathway	1.426e-18	1.322e-16	
Integrated breast cancer pathway	1.699e-18	1.322e-16	
Insulin signaling	4.429e-18	2.955e-16	
Focal adhesion	2.120e-17	1.238e-15	
Signaling pathways in glioblastoma	4.047e-17	2.100e-15	
ErbB signaling pathway	4.559e-16	2.129e-14	

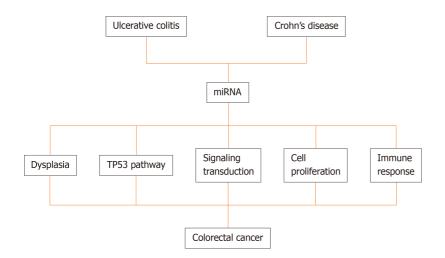


Figure 2 A schematic diagram showing the pathways modulated by miRNAs in inflammatory bowel disease-related colorectal cancer progression.

> "cell proliferation" and "immune responses". The authors concluded that miRNAs, genes and pathways are connected and there is a crosstalk between different pathways, and the miRNAs might mediate pathway crosstalk via regulating the corresponding gene^[151].

> A study conducted by Olaru *et al*^[123] evaluated fragments of colon tissue from healthy people and compared with fragments of non-inflamed mucosa, inflamed mucosa of patients with IBD, IBD-dysplasia tissue and IBD-associated cancer tissue. The authors identified five upregulated miRNAs in IBD-related cancer, namely, miR-31, miR-135b, miR-200a, miR-224, and miR-552. MiR-224 was expressed at the highestlevel during differentiation between IBD patients with tumors and patients with IBD without tumors.

> Several miRNAs are being correlated with the transition of normal tissue to dysplasia or neoplasia. In CD, miR-196 is a marker of dysplasia[57]. Further, miR-124a, a tumor-suppressive miRNA, undergoes methylation during exposure to chronic inflammation leading to the emergence of dysplasia and then neoplastic tissues in UC patients[152]. Wan et al[153] revealed that miR-155 is related to the involvement of cancer cells and worse prognosis. Additionally, Fang et al [154] analyzed patients with colorectal disease and healthy controls and determined that miR-24, miR-320a, and miR-423-5p, which were aberrantly expressed, were associated with high sensitivity for the detection of early CRC and could be promising biomarkers for IBD. In a recent study, Al-Mustanjid et al[155] used a system biology approach to identify common molecular signatures and pathways that interact between IBD and CRC and found that mir-335-5p, mir-26b-5p, mir-124-3p, mir-16-5p, mir-192-5p, mir-548c-3p, mir-29b-3p, mir-155-5p, mir-21-5p, mir-15a-5p are related with the 177 common differentially



expressed genes between IBD and CRC. A schematic diagram showing the pathways modulated by miRNAs in IBD-related CRC progression is shown in Figure 2.

Due to the role of inflammation in CRC carcinogenesis, prevention methods targeting pro-inflammatory pathways have been studied. Among these pathways are the NF- κ B pathway, which regulates innate and adaptive immune functions [156], and the phosphatidylinositol 3 kinase pathways (Pl3K), TLR, Janus kinase, and the activating factor transcript 3 (JAK/STAT3), which are also involved in the inflammation/cancer cascade [157-159]. Thus, the development of drugs that regulate miRNAs and these pro-inflammatory pathways has become an important field to prevent the development of CRC in these patients^[20].

After predicting the target genes for the IBD-associated CRC miRNAs[160] were found 8939 possible targets for the selected miRNAs (Supporting material). The enrichment analysis performed showed large modulation of signaling pathways that participate in the pathophysiology of both IBD and CRC, pathways such as the VEGFA, TGF- β and EGFR pathways that have already been discussed in this review [161-163] (Table 5). The participation of these pathways in both diseases could help to explain the correlation between these two conditions and why IBD patients are more likely to develop CRC.

CONCLUSION

Inflammatory processes and immunosuppressive drugs are the main risk factors for the development of tumors in patients with IBD. The identification of miRNAs as diagnostic biomarkers can revolutionize the screening of high-risk patients, allowing for personalized surveillance according to individual risk and the early diagnosis of lesions, directly affecting the treatment and prognosis of the patient. Further studies are needed to clarify the role of miRNAs in disease pathogenesis and evolution in patients with IBD, in addition to the identification of miRNAs related to the therapeutic response and disease prognosis. In the future, therapies based on miRNA modulation could be used in clinical practice to achieve remission of the disease and restore the quality of life for patients with IBD.

REFERENCES

- Lu TX, Rothenberg ME. MicroRNA. J Allergy Clin Immunol 2018; 141: 1202-1207 [PMID: 29074454 DOI: 10.1016/j.jaci.2017.08.034]
- 2 Iorio MV, Croce CM. MicroRNAs in cancer: small molecules with a huge impact. J Clin Oncol 2009; 27: 5848-5856 [PMID: 19884536 DOI: 10.1200/JCO.2009.24.0317]
- Huang Y, Shen XJ, Zou Q, Wang SP, Tang SM, Zhang GZ. Biological functions of microRNAs: a 3 review. J Physiol Biochem 2011; 67: 129-139 [PMID: 20981514 DOI: 10.1007/s13105-010-0050-6]
- 4 Weber JA, Baxter DH, Zhang S, Huang DY, Huang KH, Lee MJ, Galas DJ, Wang K. The microRNA spectrum in 12 body fluids. Clin Chem 2010; 56: 1733-1741 [PMID: 20847327 DOI: 10.1373/clinchem.2010.147405]
- 5 Saito Y, Suzuki H, Hibi T. The role of microRNAs in gastrointestinal cancers. J Gastroenterol 2009; 44 Suppl 19: 18-22 [PMID: 19148788 DOI: 10.1007/s00535-008-2285-3]
- Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell 2009; 136: 215-233 6 [PMID: 19167326 DOI: 10.1016/j.cell.2009.01.002]
- 7 Baz M, Ibrahim T. Role of microRNAs in the predisposition to gastrointestinal malignancies. World J Clin Cases 2020; 8: 1580-1585 [PMID: 32420299 DOI: 10.12998/wjcc.v8.i9.1580]
- Link A, Balaguer F, Shen Y, Nagasaka T, Lozano JJ, Boland CR, Goel A. Fecal MicroRNAs as 8 novel biomarkers for colon cancer screening. Cancer Epidemiol Biomarkers Prev 2010; 19: 1766-1774 [PMID: 20551304 DOI: 10.1158/1055-9965.EPI-10-0027]
- Shi C, Liang Y, Yang J, Xia Y, Chen H, Han H, Yang Y, Wu W, Gao R, Qin H. MicroRNA-21 knockout improve the survival rate in DSS induced fatal colitis through protecting against inflammation and tissue injury. PLoS One 2013; 8: e66814 [PMID: 23826144 DOI: 10.1371/journal.pone.0066814]
- 10 Gracias DT, Stelekati E, Hope JL, Boesteanu AC, Doering TA, Norton J, Mueller YM, Fraietta JA, Wherry EJ, Turner M, Katsikis PD. The microRNA miR-155 controls CD8(+) T cell responses by regulating interferon signaling. Nat Immunol 2013; 14: 593-602 [PMID: 23603793 DOI: 10.1038/ni.2576
- Singh UP, Murphy AE, Enos RT, Shamran HA, Singh NP, Guan H, Hegde VL, Fan D, Price RL, 11 Taub DD, Mishra MK, Nagarkatti M, Nagarkatti PS. miR-155 deficiency protects mice from experimental colitis by reducing T helper type 1/type 17 responses. Immunology 2014; 143: 478-489 [PMID: 24891206 DOI: 10.1111/imm.12328]



- 12 Yang L, Belaguli N, Berger DH. MicroRNA and colorectal cancer. World J Surg 2009; 33: 638-646 [PMID: 19123024 DOI: 10.1007/s00268-008-9865-5]
- 13 Cowland JB, Hother C, Grønbaek K. MicroRNAs and cancer. APMIS 2007; 115: 1090-1106 [PMID: 18042145 DOI: 10.1111/j.1600-0463.2007.apm_775.xml.x]
- 14 Munkholm P. Review article: the incidence and prevalence of colorectal cancer in inflammatory bowel disease. Aliment Pharmacol Ther 2003; 18 Suppl 2: 1-5 [PMID: 12950413 DOI: 10.1046/j.1365-2036.18.s2.2.x
- Viscido A, Bagnardi V, Sturniolo GC, Annese V, Frieri G, D'Arienzo A, Papi C, Riegler G, Corrao 15 G, Caprilli R; GISC: Italian Group for the Study of the Colon and Rectum. Survival and causes of death in Italian patients with ulcerative colitis. A GISC nationwide study. Dig Liver Dis 2001; 33: 686-692 [PMID: 11785715 DOI: 10.1016/s1590-8658(01)80046-3]
- 16 Magro F, Gionchetti P, Eliakim R, Ardizzone S, Armuzzi A, Barreiro-de Acosta M, Burisch J, Gecse KB, Hart AL, Hindryckx P, Langner C, Limdi JK, Pellino G, Zagórowicz E, Raine T, Harbord M, Rieder F; European Crohn's and Colitis Organisation [ECCO]. Third European Evidence-based Consensus on Diagnosis and Management of Ulcerative Colitis. Part 1: Definitions, Diagnosis, Extra-intestinal Manifestations, Pregnancy, Cancer Surveillance, Surgery, and Ileo-anal Pouch Disorders. J Crohns Colitis 2017; 11: 649-670 [PMID: 28158501 DOI: 10.1093/ecco-jcc/jjx008]
- 17 Askling J, Dickman PW, Karlén P, Broström O, Lapidus A, Löfberg R, Ekbom A. Family history as a risk factor for colorectal cancer in inflammatory bowel disease. Gastroenterology 2001; 120: 1356-1362 [PMID: 11313305 DOI: 10.1053/gast.2001.24052]
- 18 Zheng HH, Jiang XL. Increased risk of colorectal neoplasia in patients with primary sclerosing cholangitis and inflammatory bowel disease: a meta-analysis of 16 observational studies. Eur J Gastroenterol Hepatol 2016; 28: 383-390 [PMID: 26938805 DOI: 10.1097/MEG.000000000000576
- 19 Burisch J, Munkholm P. The epidemiology of inflammatory bowel disease. Scand J Gastroenterol 2015; 50: 942-951 [PMID: 25687629 DOI: 10.3109/00365521.2015.1014407]
- Feng Y, Zhang Y, Zhou D, Chen G, Li N. MicroRNAs, intestinal inflammatory and tumor. Bioorg 20 Med Chem Lett 2019; 29: 2051-2058 [PMID: 31213403 DOI: 10.1016/j.bmcl.2019.06.013]
- 21 Magro F, Langner C, Driessen A, Ensari A, Geboes K, Mantzaris GJ, Villanacci V, Becheanu G, Borralho Nunes P, Cathomas G, Fries W, Jouret-Mourin A, Mescoli C, de Petris G, Rubio CA, Shepherd NA, Vieth M, Eliakim R; European Society of Pathology (ESP); European Crohn's and Colitis Organisation (ECCO). European consensus on the histopathology of inflammatory bowel disease. J Crohns Colitis 2013; 7: 827-851 [PMID: 23870728 DOI: 10.1016/j.crohns.2013.06.001]
- 22 de Souza HS, Fiocchi C. Immunopathogenesis of IBD: current state of the art. Nat Rev Gastroenterol Hepatol 2016; 13: 13-27 [PMID: 26627550 DOI: 10.1038/nrgastro.2015.186]
- 23 Maaser C, Sturm A, Vavricka SR, Kucharzik T, Fiorino G, Annese V, Calabrese E, Baumgart DC, Bettenworth D, Borralho Nunes P, Burisch J, Castiglione F, Eliakim R, Ellul P, González-Lama Y, Gordon H, Halligan S, Katsanos K, Kopylov U, Kotze PG, Krustinš E, Laghi A, Limdi JK, Rieder F, Rimola J, Taylor SA, Tolan D, van Rheenen P, Verstockt B, Stoker J; European Crohn's and Colitis Organisation [ECCO] and the European Society of Gastrointestinal and Abdominal Radiology [ESGAR]. ECCO-ESGAR Guideline for Diagnostic Assessment in IBD Part 1: Initial diagnosis, monitoring of known IBD, detection of complications. J Crohns Colitis 2019; 13: 144-164 [PMID: 30137275 DOI: 10.1093/ecco-jcc/jjy113]
- Guindi M, Riddell RH. Indeterminate colitis. J Clin Pathol 2004; 57: 1233-1244 [PMID: 15563659 24 DOI: 10.1136/jcp.2003.015214]
- Schönauen K, Le N, von Arnim U, Schulz C, Malfertheiner P, Link A. Circulating and Fecal 25 microRNAs as Biomarkers for Inflammatory Bowel Diseases. Inflamm Bowel Dis 2018; 24: 1547-1557 [PMID: 29668922 DOI: 10.1093/ibd/izy046]
- Whiteoak SR, Felwick R, Sanchez-Elsner T, Fraser Cummings JR. MicroRNAs in inflammatory 26 bowel diseases: paradoxes and possibilities. Inflamm Bowel Dis 2015; 21: 1160-1165 [PMID: 25844960 DOI: 10.1097/MIB.000000000000288]
- 27 Wu F, Zikusoka M, Trindade A, Dassopoulos T, Harris ML, Bayless TM, Brant SR, Chakravarti S, Kwon JH. MicroRNAs are differentially expressed in ulcerative colitis and alter expression of macrophage inflammatory peptide-2 alpha. Gastroenterology 2008; 135: 1624-1635.e24 [PMID: 18835392 DOI: 10.1053/j.gastro.2008.07.068]
- Wu F, Zhang S, Dassopoulos T, Harris ML, Bayless TM, Meltzer SJ, Brant SR, Kwon JH. 28 Identification of microRNAs associated with ileal and colonic Crohn's disease. Inflamm Bowel Dis 2010; 16: 1729-1738 [PMID: 20848482 DOI: 10.1002/ibd.21267]
- 29 Schaefer JS, Attumi T, Opekun AR, Abraham B, Hou J, Shelby H, Graham DY, Streckfus C, Klein JR. MicroRNA signatures differentiate Crohn's disease from ulcerative colitis. BMC Immunol 2015; 16: 5 [PMID: 25886994 DOI: 10.1186/s12865-015-0069-0]
- 30 Thorlacius-Ussing G, Schnack Nielsen B, Andersen V, Holmstrøm K, Pedersen AE. Expression and Localization of miR-21 and miR-126 in Mucosal Tissue from Patients with Inflammatory Bowel Disease. Inflamm Bowel Dis 2017; 23: 739-752 [PMID: 28426456 DOI: 10.1097/MIB.0000000000001086
- 31 Guo Z, Wu R, Gong J, Zhu W, Li Y, Wang Z, Li N, Li J. Altered microRNA expression in inflamed and non-inflamed terminal ileal mucosa of adult patients with active Crohn's disease. J Gastroenterol Hepatol 2015; 30: 109-116 [PMID: 24910152 DOI: 10.1111/jgh.12644]



- 32 Fasseu M, Tréton X, Guichard C, Pedruzzi E, Cazals-Hatem D, Richard C, Aparicio T, Daniel F, Soulé JC, Moreau R, Bouhnik Y, Laburthe M, Groyer A, Ogier-Denis E. Identification of restricted subsets of mature microRNA abnormally expressed in inactive colonic mucosa of patients with inflammatory bowel disease. PLoS One 2010; 5 [PMID: 20957151 DOI: 10.1371/journal.pone.0013160]
- 33 Wu W, He C, Liu C, Cao AT, Xue X, Evans-Marin HL, Sun M, Fang L, Yao S, Pinchuk IV, Powell DW, Liu Z, Cong Y. miR-10a inhibits dendritic cell activation and Th1/Th17 cell immune responses in IBD. Gut 2015; 64: 1755-1764 [PMID: 25281418 DOI: 10.1136/gutjnl-2014-307980]
- 34 Pierdomenico M, Cesi V, Cucchiara S, Vitali R, Prete E, Costanzo M, Aloi M, Oliva S, Stronati L. NOD2 Is Regulated By Mir-320 in Physiological Conditions but this Control Is Altered in Inflamed Tissues of Patients with Inflammatory Bowel Disease. Inflamm Bowel Dis 2016; 22: 315-326 [PMID: 26752466 DOI: 10.1097/MIB.00000000000659]
- Chuang AY, Chuang JC, Zhai Z, Wu F, Kwon JH. NOD2 expression is regulated by microRNAs in 35 colonic epithelial HCT116 cells. Inflamm Bowel Dis 2014; 20: 126-135 [PMID: 24297055 DOI: 10.1097/01.MIB.0000436954.70596.9b
- 36 Min M, Peng L, Yang Y, Guo M, Wang W, Sun G. MicroRNA-155 is involved in the pathogenesis of ulcerative colitis by targeting FOXO3a. Inflamm Bowel Dis 2014; 20: 652-659 [PMID: 24583476 DOI: 10.1097/MIB.000000000000009]
- Lu ZJ, Wu JJ, Jiang WL, Xiao JH, Tao KZ, Ma L, Zheng P, Wan R, Wang XP. MicroRNA-155 37 promotes the pathogenesis of experimental colitis by repressing SHIP-1 expression. World J Gastroenterol 2017; 23: 976-985 [PMID: 28246471 DOI: 10.3748/wjg.v23.i6.976]
- Shi T, Xie Y, Fu Y, Zhou Q, Ma Z, Ma J, Huang Z, Zhang J, Chen J. The signaling axis of 38 microRNA-31/interleukin-25 regulates Th1/Th17-mediated inflammation response in colitis. Mucosal Immunol 2017; 10: 983-995 [PMID: 27901018 DOI: 10.1038/mi.2016.102]
- Wang H, Chao K, Ng SC, Bai AH, Yu Q, Yu J, Li M, Cui Y, Chen M, Hu JF, Zhang S. Pro-39 inflammatory miR-223 mediates the cross-talk between the IL23 pathway and the intestinal barrier in inflammatory bowel disease. Genome Biol 2016; 17: 58 [PMID: 27029486 DOI: 10.1186/s13059-016-0901-8
- 40 Hou J, Hu X, Chen B, Chen X, Zhao L, Chen Z, Liu F, Liu Z. miR-155 targets Est-1 and induces ulcerative colitis via the IL-23/17/6-mediated Th17 pathway. Pathol Res Pract 2017; 213: 1289-1295 [PMID: 28888763 DOI: 10.1016/j.prp.2017.08.001]
- 41 Nata T, Fujiya M, Ueno N, Moriichi K, Konishi H, Tanabe H, Ohtake T, Ikuta K, Kohgo Y. MicroRNA-146b improves intestinal injury in mouse colitis by activating nuclear factor- κ B and improving epithelial barrier function. J Gene Med 2013; 15: 249-260 [PMID: 23813877 DOI: 10.1002/jgm.2717]
- 42 Huang Z, Shi T, Zhou Q, Shi S, Zhao R, Shi H, Dong L, Zhang C, Zeng K, Chen J, Zhang J. miR-141 Regulates colonic leukocytic trafficking by targeting CXCL12β during murine colitis and human Crohn's disease. Gut 2014; 63: 1247-1257 [PMID: 24000293 DOI: 10.1136/gutjnl-2012-304213]
- 43 Jin X, Chen D, Zheng RH, Zhang H, Chen YP, Xiang Z. miRNA-133a-UCP2 pathway regulates inflammatory bowel disease progress by influencing inflammation, oxidative stress and energy metabolism. World J Gastroenterol 2017; 23: 76-86 [PMID: 28104982 DOI: 10.3748/wjg.v23.i1.76]
- Tian Y, Xu J, Li Y, Zhao R, Du S, Lv C, Wu W, Liu R, Sheng X, Song Y, Bi X, Li G, Li M, Wu X, 44 Lou P, You H, Cui W, Sun J, Shuai J, Ren F, Zhang B, Guo M, Hou X, Wu K, Xue L, Zhang H, Plikus MV, Cong Y, Lengner CJ, Liu Z, Yu Z. MicroRNA-31 Reduces Inflammatory Signaling and Promotes Regeneration in Colon Epithelium, and Delivery of Mimics in Microspheres Reduces Colitis in Mice. Gastroenterology 2019; 156: 2281-2296.e6 [PMID: 30779922 DOI: 10.1053/j.gastro.2019.02.023]
- 45 He C, Shi Y, Wu R, Sun M, Fang L, Wu W, Liu C, Tang M, Li Z, Wang P, Cong Y, Liu Z. miR-301a promotes intestinal mucosal inflammation through induction of IL-17A and TNF-α in IBD. Gut 2016; 65: 1938-1950 [PMID: 26338824 DOI: 10.1136/gutjnl-2015-309389]
- He C, Yu T, Shi Y, Ma C, Yang W, Fang L, Sun M, Wu W, Xiao F, Guo F, Chen M, Yang H, Qian J, Cong Y, Liu Z. MicroRNA 301A Promotes Intestinal Inflammation and Colitis-Associated Cancer Development by Inhibiting BTG1. Gastroenterology 2017; 152: 1434-1448.e15 [PMID: 28193514 DOI: 10.1053/j.gastro.2017.01.049]
- Bae HJ, Noh JH, Kim JK, Eun JW, Jung KH, Kim MG, Chang YG, Shen Q, Kim SJ, Park WS, Lee 47 JY, Nam SW. MicroRNA-29c functions as a tumor suppressor by direct targeting oncogenic SIRT1 in hepatocellular carcinoma. Oncogene 2014; 33: 2557-2567 [PMID: 23728341 DOI: 10.1038/onc.2013.216
- 48 Chapman CG, Pekow J. The emerging role of miRNAs in inflammatory bowel disease: a review. Therap Adv Gastroenterol 2015; 8: 4-22 [PMID: 25553076 DOI: 10.1177/1756283X14547360]
- 49 Harris TA, Yamakuchi M, Ferlito M, Mendell JT, Lowenstein CJ. MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. Proc Natl Acad Sci USA 2008; 105: 1516-1521 [PMID: 18227515 DOI: 10.1073/pnas.0707493105]
- 50 Fedyk ER, Wyant T, Yang LL, Csizmadia V, Burke K, Yang H, Kadambi VJ. Exclusive antagonism of the $\alpha 4 \beta 7$ integrin by vedolizumab confirms the gut-selectivity of this pathway in primates. Inflamm Bowel Dis 2012; 18: 2107-2119 [PMID: 22419649 DOI: 10.1002/ibd.22940]
- 51 Pathak S, Grillo AR, Scarpa M, Brun P, D'Incà R, Nai L, Banerjee A, Cavallo D, Barzon L, Palù G, Sturniolo GC, Buda A, Castagliuolo I. MiR-155 modulates the inflammatory phenotype of intestinal



myofibroblasts by targeting SOCS1 in ulcerative colitis. Exp Mol Med 2015; 47: e164 [PMID: 25998827 DOI: 10.1038/emm.2015.21]

- 52 Morilla I, Uzzan M, Laharie D, Cazals-Hatem D, Denost Q, Daniel F, Belleannee G, Bouhnik Y, Wainrib G, Panis Y, Ogier-Denis E, Treton X. Colonic MicroRNA Profiles, Identified by a Deep Learning Algorithm, That Predict Responses to Therapy of Patients With Acute Severe Ulcerative Colitis. Clin Gastroenterol Hepatol 2019; 17: 905-913 [PMID: 30223112 DOI: 10.1016/j.cgh.2018.08.068]
- 53 Cordes F, Demmig C, Bokemeyer A, Brückner M, Lenze F, Lenz P, Nowacki T, Tepasse P, Schmidt HH, Schmidt MA, Cichon C, Bettenworth D. MicroRNA-320a Monitors Intestinal Disease Activity in Patients With Inflammatory Bowel Disease. Clin Transl Gastroenterol 2020; 11: e00134 [PMID: 32352717 DOI: 10.14309/ctg.00000000000134]
- 54 Verdier J, Breunig IR, Ohse MC, Roubrocks S, Kleinfeld S, Roy S, Streetz K, Trautwein C, Roderburg C, Sellge G. Faecal Micro-RNAs in Inflammatory Bowel Diseases. J Crohns Colitis 2020; 14: 110-117 [PMID: 31209454 DOI: 10.1093/ecco-jcc/jjz120]
- 55 Ben-Shachar S, Yanai H, Sherman Horev H, Elad H, Baram L, Isakov O, Tulchinsky H, Pasmanik-Chor M, Shomron N, Dotan I. Correction: MicroRNAs Expression in the Ileal Pouch of Patients with Ulcerative Colitis Is Robustly Up-Regulated and Correlates with Disease Phenotypes. PLoS One 2016; 11: e0165220 [PMID: 27755607 DOI: 10.1371/journal.pone.0165220]
- Gwiggner M, Martinez-Nunez RT, Whiteoak SR, Bondanese VP, Claridge A, Collins JE, 56 Cummings JRF, Sanchez-Elsner T. MicroRNA-31 and MicroRNA-155 Are Overexpressed in Ulcerative Colitis and Regulate IL-13 Signaling by Targeting Interleukin 13 Receptor a-1. Genes (Basel) 2018; 9 [PMID: 29438285 DOI: 10.3390/genes9020085]
- Brest P, Lapaquette P, Souidi M, Lebrigand K, Cesaro A, Vouret-Craviari V, Mari B, Barbry P, 57 Mosnier JF, Hébuterne X, Harel-Bellan A, Mograbi B, Darfeuille-Michaud A, Hofman P. A synonymous variant in IRGM alters a binding site for miR-196 and causes deregulation of IRGMdependent xenophagy in Crohn's disease. Nat Genet 2011; 43: 242-245 [PMID: 21278745 DOI: 10.1038/ng.762
- 58 Lin J, Welker NC, Zhao Z, Li Y, Zhang J, Reuss SA, Zhang X, Lee H, Liu Y, Bronner MP. Novel specific microRNA biomarkers in idiopathic inflammatory bowel disease unrelated to disease activity. Mod Pathol 2014; 27: 602-608 [PMID: 24051693 DOI: 10.1038/modpathol.2013.152]
- 59 Iborra M, Bernuzzi F, Correale C, Vetrano S, Fiorino G, Beltrán B, Marabita F, Locati M, Spinelli A, Nos P, Invernizzi P, Danese S. Identification of serum and tissue micro-RNA expression profiles in different stages of inflammatory bowel disease. Clin Exp Immunol 2013; 173: 250-258 [PMID: 23607522 DOI: 10.1111/cei.12104]
- 60 Pekow JR, Dougherty U, Mustafi R, Zhu H, Kocherginsky M, Rubin DT, Hanauer SB, Hart J, Chang EB, Fichera A, Joseph LJ, Bissonnette M. miR-143 and miR-145 are downregulated in ulcerative colitis: putative regulators of inflammation and protooncogenes. Inflamm Bowel Dis 2012; 18: 94-100 [PMID: 21557394 DOI: 10.1002/ibd.21742]
- 61 Peck BC, Weiser M, Lee SE, Gipson GR, Iyer VB, Sartor RB, Herfarth HH, Long MD, Hansen JJ, Isaacs KL, Trembath DG, Rahbar R, Sadiq TS, Furey TS, Sethupathy P, Sheikh SZ. MicroRNAs Classify Different Disease Behavior Phenotypes of Crohn's Disease and May Have Prognostic Utility. Inflamm Bowel Dis 2015; 21: 2178-2187 [PMID: 26164662 DOI: 10.1097/MIB.000000000000478]
- 62 Coskun M, Bjerrum JT, Seidelin JB, Troelsen JT, Olsen J, Nielsen OH. miR-20b, miR-98, miR-125b-1*, and let-7e* as new potential diagnostic biomarkers in ulcerative colitis. World J Gastroenterol 2013; 19: 4289-4299 [PMID: 23885139 DOI: 10.3748/wjg.v19.i27.4289]
- 63 Feng X, Wang H, Ye S, Guan J, Tan W, Cheng S, Wei G, Wu W, Wu F, Zhou Y. Up-regulation of microRNA-126 may contribute to pathogenesis of ulcerative colitis via regulating NF-kappaB inhibitor IkBa. PLoS One 2012; 7: e52782 [PMID: 23285182 DOI: 10.1371/journal.pone.0052782]
- 64 Bian Z, Li L, Cui J, Zhang H, Liu Y, Zhang CY, Zen K. Role of miR-150-targeting c-Myb in colonic epithelial disruption during dextran sulphate sodium-induced murine experimental colitis and human ulcerative colitis. J Pathol 2011; 225: 544-553 [PMID: 21590770 DOI: 10.1002/path.2907]
- Polytarchou C, Hommes DW, Palumbo T, Hatziapostolou M, Koutsioumpa M, Koukos G, van der 65 Meulen-de Jong AE, Oikonomopoulos A, van Deen WK, Vorvis C, Serebrennikova OB, Birli E, Choi J, Chang L, Anton PA, Tsichlis PN, Pothoulakis C, Verspaget HW, Iliopoulos D. MicroRNA214 Is Associated With Progression of Ulcerative Colitis, and Inhibition Reduces Development of Colitis and Colitis-Associated Cancer in Mice. Gastroenterology 2015; 149: 981-92.e11 [PMID: 26055138 DOI: 10.1053/j.gastro.2015.05.057]
- 66 Koukos G, Polytarchou C, Kaplan JL, Morley-Fletcher A, Gras-Miralles B, Kokkotou E, Baril-Dore M, Pothoulakis C, Winter HS, Iliopoulos D. MicroRNA-124 regulates STAT3 expression and is down-regulated in colon tissues of pediatric patients with ulcerative colitis. Gastroenterology 2013; 145: 842-52.e2 [PMID: 23856509 DOI: 10.1053/j.gastro.2013.07.001]
- Zahm AM, Hand NJ, Tsoucas DM, Le Guen CL, Baldassano RN, Friedman JR. Rectal microRNAs are perturbed in pediatric inflammatory bowel disease of the colon. J Crohns Colitis 2014; 8: 1108-1117 [PMID: 24613022 DOI: 10.1016/j.crohns.2014.02.012]
- Shen Y, Zhou M, Yan J, Gong Z, Xiao Y, Zhang C, Du P, Chen Y. miR-200b inhibits TNF-α-68 induced IL-8 secretion and tight junction disruption of intestinal epithelial cells in vitro. Am J Physiol Gastrointest Liver Physiol 2017; 312: G123-G132 [PMID: 27979826 DOI:



10.1152/ajpgi.00316.2016

- 69 Koukos G, Polytarchou C, Kaplan JL, Oikonomopoulos A, Ziring D, Hommes DW, Wahed R, Kokkotou E, Pothoulakis C, Winter HS, Iliopoulos D. A microRNA signature in pediatric ulcerative colitis: deregulation of the miR-4284/CXCL5 pathway in the intestinal epithelium. Inflamm Bowel Dis 2015; 21: 996-1005 [PMID: 25738378 DOI: 10.1097/MIB.00000000000339]
- 70 Fisher K, Lin J. MicroRNA in inflammatory bowel disease: Translational research and clinical implication. World J Gastroenterol 2015; 21: 12274-12282 [PMID: 26604636 DOI: 10.3748/wjg.v21.i43.12274]
- 71 Xu XM, Zhang HJ. miRNAs as new molecular insights into inflammatory bowel disease: Crucial regulators in autoimmunity and inflammation. World J Gastroenterol 2016; 22: 2206-2218 [PMID: 26900285 DOI: 10.3748/wjg.v22.i7.2206]
- James JP, Riis LB, Malham M, Høgdall E, Langholz E, Nielsen BS. MicroRNA Biomarkers in 72 IBD-Differential Diagnosis and Prediction of Colitis-Associated Cancer. Int J Mol Sci 2020; 21 [PMID: 33114313 DOI: 10.3390/ijms21217893]
- 73 Dekker E, Tanis PJ, Vleugels JLA, Kasi PM, Wallace MB. Colorectal cancer. Lancet 2019; 394: 1467-1480 [PMID: 31631858 DOI: 10.1016/S0140-6736(19)32319-0]
- 74 Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. Gut 2017; 66: 683-691 [PMID: 26818619 DOI: 10.1136/gutjnl-2015-310912]
- 75 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 76 Henrikson NB, Webber EM, Goddard KA, Scrol A, Piper M, Williams MS, Zallen DT, Calonge N, Ganiats TG, Janssens AC, Zauber A, Lansdorp-Vogelaar I, van Ballegooijen M, Whitlock EP. Family history and the natural history of colorectal cancer: systematic review. Genet Med 2015; 17: 702-712 [PMID: 25590981 DOI: 10.1038/gim.2014.188]
- Syngal S, Brand RE, Church JM, Giardiello FM, Hampel HL, Burt RW; American College of 77 Gastroenterology. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol 2015; 110: 223-262; quiz 263 [PMID: 25645574 DOI: 10.1038/ajg.2014.435]
- 78 Medema JP. Cancer stem cells: the challenges ahead. Nat Cell Biol 2013; 15: 338-344 [PMID: 23548926 DOI: 10.1038/ncb2717]
- Nassar D, Blanpain C. Cancer Stem Cells: Basic Concepts and Therapeutic Implications. Annu Rev 79 Pathol 2016; 11: 47-76 [PMID: 27193450 DOI: 10.1146/annurev-pathol-012615-044438]
- 80 Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer. Nature 2012; 487: 330-337 [PMID: 22810696 DOI: 10.1038/nature11252]
- 81 Mohammadi A, Mansoori B, Baradaran B. The role of microRNAs in colorectal cancer. Biomed Pharmacother 2016; 84: 705-713 [PMID: 27701052 DOI: 10.1016/j.biopha.2016.09.099]
- 82 Hofacker IL. How microRNAs choose their targets. Nat Genet 2007; 39: 1191-1192 [PMID: 17898777 DOI: 10.1038/ng1007-1191]
- 83 Chivukula RR, Shi G, Acharya A, Mills EW, Zeitels LR, Anandam JL, Abdelnaby AA, Balch GC, Mansour JC, Yopp AC, Maitra A, Mendell JT. An essential mesenchymal function for miR-143/145 in intestinal epithelial regeneration. Cell 2014; 157: 1104-1116 [PMID: 24855947 DOI: 10.1016/j.cell.2014.03.055]
- 84 Nagel R, le Sage C, Diosdado B, van der Waal M, Oude Vrielink JA, Bolijn A, Meijer GA, Agami R. Regulation of the adenomatous polyposis coli gene by the miR-135 family in colorectal cancer. Cancer Res 2008; 68: 5795-5802 [PMID: 18632633 DOI: 10.1158/0008-5472.CAN-08-0951]
- 85 Dinu D, Dobre M, Panaitescu E, Bîrlă R, Iosif C, Hoara P, Caragui A, Boeriu M, Constantinoiu S, Ardeleanu C. Prognostic significance of KRAS gene mutations in colorectal cancer--preliminary study. J Med Life 2014; 7: 581-587 [PMID: 25713627]
- 86 Akao Y, Nakagawa Y, Naoe T. let-7 microRNA functions as a potential growth suppressor in human colon cancer cells. Biol Pharm Bull 2006; 29: 903-906 [PMID: 16651716 DOI: 10.1248/bpb.29.903]
- Iliopoulos D, Jaeger SA, Hirsch HA, Bulyk ML, Struhl K. STAT3 activation of miR-21 and miR-87 181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer. Mol Cell 2010; 39: 493-506 [PMID: 20797623 DOI: 10.1016/j.molcel.2010.07.023]
- Bieging KT, Mello SS, Attardi LD. Unravelling mechanisms of p53-mediated tumour suppression. 88 Nat Rev Cancer 2014; 14: 359-370 [PMID: 24739573 DOI: 10.1038/nrc3711]
- 89 Moustakas A, Heldin CH. The regulation of TGFbeta signal transduction. Development 2009; 136: 3699-3714 [PMID: 19855013 DOI: 10.1242/dev.030338]
- 90 Chiang Y, Song Y, Wang Z, Liu Z, Gao P, Liang J, Zhu J, Xing C, Xu H. microRNA-192, -194 and -215 are frequently downregulated in colorectal cancer. Exp Ther Med 2012; 3: 560-566 [PMID: 22969930 DOI: 10.3892/etm.2011.436]
- Liu Q, Yang W, Luo Y, Hu S, Zhu L. Correlation between miR-21 and miR-145 and the incidence 91 and prognosis of colorectal cancer. J BUON 2018; 23: 29-35 [PMID: 29552756]
- 92 Huang Z, Huang D, Ni S, Peng Z, Sheng W, Du X. Plasma microRNAs are promising novel biomarkers for early detection of colorectal cancer. Int J Cancer 2010; 127: 118-126 [PMID: 19876917 DOI: 10.1002/ijc.25007]
- 93 Wu Y, Song Y, Xiong Y, Wang X, Xu K, Han B, Bai Y, Li L, Zhang Y, Zhou L. MicroRNA-21



(Mir-21) Promotes Cell Growth and Invasion by Repressing Tumor Suppressor PTEN in Colorectal Cancer. Cell Physiol Biochem 2017; 43: 945-958 [PMID: 28957811 DOI: 10.1159/000481648]

- 94 Wang CJ, Zhou ZG, Wang L, Yang L, Zhou B, Gu J, Chen HY, Sun XF. Clinicopathological significance of microRNA-31, -143 and -145 expression in colorectal cancer. Dis Markers 2009; 26: 27-34 [PMID: 19242066 DOI: 10.3233/DMA-2009-0601]
- 95 Tsukamoto M, Iinuma H, Yagi T, Matsuda K, Hashiguchi Y. Circulating Exosomal MicroRNA-21 as a Biomarker in Each Tumor Stage of Colorectal Cancer. Oncology 2017; 92: 360-370 [PMID: 28376502 DOI: 10.1159/000463387]
- 96 Asangani IA, Rasheed SA, Nikolova DA, Leupold JH, Colburn NH, Post S, Allgayer H. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pdcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. Oncogene 2008; 27: 2128-2136 [PMID: 17968323 DOI: 10.1038/sj.onc.1210856]
- 97 Giráldez MD, Lozano JJ, Ramírez G, Hijona E, Bujanda L, Castells A, Gironella M. Circulating microRNAs as biomarkers of colorectal cancer: results from a genome-wide profiling and validation study. Clin Gastroenterol Hepatol 2013; 11: 681-8.e3 [PMID: 23267864 DOI: 10.1016/j.cgh.2012.12.009
- 98 Wang LG, Gu J. Serum microRNA-29a is a promising novel marker for early detection of colorectal liver metastasis. Cancer Epidemiol 2012; 36: e61-e67 [PMID: 22018950 DOI: 10.1016/j.canep.2011.05.002]
- Sun Y, Liu Y, Cogdell D, Calin GA, Sun B, Kopetz S, Hamilton SR, Zhang W. Examining plasma microRNA markers for colorectal cancer at different stages. Oncotarget 2016; 7: 11434-11449 [PMID: 26863633 DOI: 10.18632/oncotarget.7196]
- Hu J, Cai G, Xu Y, Cai S. The Plasma microRNA miR-1914* and -1915 Suppresses Chemoresistant 100 in Colorectal Cancer Patients by Down-regulating NFIX. Curr Mol Med 2016; 16: 70-82 [PMID: 26695693 DOI: 10.2174/1566524016666151222144656]
- 101 Lee SL, Rouhi P, Dahl Jensen L, Zhang D, Ji H, Hauptmann G, Ingham P, Cao Y. Hypoxia-induced pathological angiogenesis mediates tumor cell dissemination, invasion, and metastasis in a zebrafish tumor model. Proc Natl Acad Sci USA 2009; 106: 19485-19490 [PMID: 19887629 DOI: 10.1073/pnas.0909228106
- 102 Viallard C, Larrivée B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. Angiogenesis 2017; 20: 409-426 [PMID: 28660302 DOI: 10.1007/s10456-017-9562-9]
- Goradel NH, Mohammadi N, Haghi-Aminjan H, Farhood B, Negahdari B, Sahebkar A. Regulation 103 of tumor angiogenesis by microRNAs: State of the art. J Cell Physiol 2019; 234: 1099-1110 [PMID: 30070704 DOI: 10.1002/jcp.27051]
- 104 Wang Y, Wang L, Chen C, Chu X. New insights into the regulatory role of microRNA in tumor angiogenesis and clinical implications. Mol Cancer 2018; 17: 22 [PMID: 29415727 DOI: 10.1186/s12943-018-0766-4
- 105 Leone P, Buonavoglia A, Fasano R, Solimando AG, De Re V, Cicco S, Vacca A, Racanelli V. Insights into the Regulation of Tumor Angiogenesis by Micro-RNAs. J Clin Med 2019; 8 [PMID: 31757094 DOI: 10.3390/jcm8122030]
- 106 Hansen TF, Carlsen AL, Heegaard NH, Sørensen FB, Jakobsen A, Changes in circulating microRNA-126 during treatment with chemotherapy and bevacizumab predicts treatment response in patients with metastatic colorectal cancer. Br J Cancer 2015; 112: 624-629 [PMID: 25584492 DOI: 10.1038/bjc.2014.652
- Zhang W, Zou C, Pan L, Xu Y, Qi W, Ma G, Hou Y, Jiang P. MicroRNA-140-5p inhibits the 107 progression of colorectal cancer by targeting VEGFA. Cell Physiol Biochem 2015; 37: 1123-1133 [PMID: 26402430 DOI: 10.1159/000430237]
- 108 Qiu Y, Yu H, Shi X, Xu K, Tang Q, Liang B, Hu S, Bao Y, Xu J, Cai J, Peng W, Cao Q, Yin P. microRNA-497 inhibits invasion and metastasis of colorectal cancer cells by targeting vascular endothelial growth factor-A. Cell Prolif 2016; 49: 69-78 [PMID: 26840372 DOI: 10.1111/cpr.12237]
- 109 Casanovas O. Cancer: Limitations of therapies exposed. Nature 2012; 484: 44-46 [PMID: 22481354 DOI: 10.1038/484044a]
- 110 Jászai J, Schmidt MHH. Trends and Challenges in Tumor Anti-Angiogenic Therapies. Cells 2019; 8 [PMID: 31540455 DOI: 10.3390/cells8091102]
- 111 Romano G, Kwong LN. Diagnostic and therapeutic applications of miRNA-based strategies to cancer immunotherapy. Cancer Metastasis Rev 2018; 37: 45-53 [PMID: 29270700 DOI: 10.1007/s10555-017-9716-7]
- 112 Ma R, Jiang T, Kang X. Circulating microRNAs in cancer: origin, function and application. J Exp Clin Cancer Res 2012; 31: 38 [PMID: 22546315 DOI: 10.1186/1756-9966-31-38]
- Chang KH, Miller N, Kheirelseid EA, Lemetre C, Ball GR, Smith MJ, Regan M, McAnena OJ, 113 Kerin MJ. MicroRNA signature analysis in colorectal cancer: identification of expression profiles in stage II tumors associated with aggressive disease. Int J Colorectal Dis 2011; 26: 1415-1422 [PMID: 21739196 DOI: 10.1007/s00384-011-1279-4]
- 114 Slattery ML, Wolff E, Hoffman MD, Pellatt DF, Milash B, Wolff RK. MicroRNAs and colon and rectal cancer: differential expression by tumor location and subtype. Genes Chromosomes Cancer 2011; 50: 196-206 [PMID: 21213373 DOI: 10.1002/gcc.20844]
- 115 Mosakhani N, Sarhadi VK, Borze I, Karjalainen-Lindsberg ML, Sundström J, Ristamäki R, Osterlund P, Knuutila S. MicroRNA profiling differentiates colorectal cancer according to KRAS



status. Genes Chromosomes Cancer 2012; 51: 1-9 [PMID: 21922590 DOI: 10.1002/gcc.20925]

- 116 Faltejskova P, Svoboda M, Srutova K, Mlcochova J, Besse A, Nekvindova J, Radova L, Fabian P, Slaba K, Kiss I, Vyzula R, Slaby O. Identification and functional screening of microRNAs highly deregulated in colorectal cancer. J Cell Mol Med 2012; 16: 2655-2666 [PMID: 22469014 DOI: 10.1111/j.1582-4934.2012.01579.x]
- Monzo M, Navarro A, Bandres E, Artells R, Moreno I, Gel B, Ibeas R, Moreno J, Martinez F, Diaz 117 T, Martinez A, Balagué O, Garcia-Foncillas J. Overlapping expression of microRNAs in human embryonic colon and colorectal cancer. Cell Res 2008; 18: 823-833 [PMID: 18607389 DOI: 10.1038/cr.2008.81]
- 118 Wang YX, Zhang XY, Zhang BF, Yang CQ, Chen XM, Gao HJ. Initial study of microRNA expression profiles of colonic cancer without lymph node metastasis. J Dig Dis 2010: 11: 50-54 [PMID: 20132431 DOI: 10.1111/j.1751-2980.2009.00413.x]
- Nishida N, Nagahara M, Sato T, Mimori K, Sudo T, Tanaka F, Shibata K, Ishii H, Sugihara K, Doki Y, Mori M. Microarray analysis of colorectal cancer stromal tissue reveals upregulation of two oncogenic miRNA clusters. Clin Cancer Res 2012; 18: 3054-3070 [PMID: 22452939 DOI: 10.1158/1078-0432.CCR-11-1078
- Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, 120 Ferracin M, Prueitt RL, Yanaihara N, Lanza G, Scarpa A, Vecchione A, Negrini M, Harris CC, Croce CM. A microRNA expression signature of human solid tumors defines cancer gene targets. Proc Natl Acad Sci USA 2006; 103: 2257-2261 [PMID: 16461460 DOI: 10.1073/pnas.0510565103]
- 121 Olaru AV, Selaru FM, Mori Y, Vazquez C, David S, Paun B, Cheng Y, Jin Z, Yang J, Agarwal R, Abraham JM, Dassopoulos T, Harris M, Bayless TM, Kwon J, Harpaz N, Livak F, Meltzer SJ. Dynamic changes in the expression of MicroRNA-31 during inflammatory bowel disease-associated neoplastic transformation. Inflamm Bowel Dis 2011; 17: 221-231 [PMID: 20848542 DOI: 10.1002/ibd.21359
- 122 Li XM, Wang AM, Zhang J, Yi H. Down-regulation of miR-126 expression in colorectal cancer and its clinical significance. Med Oncol 2011; 28: 1054-1057 [PMID: 20680522 DOI: 10.1007/s12032-010-9637-6
- Olaru AV, Yamanaka S, Vazquez C, Mori Y, Cheng Y, Abraham JM, Bayless TM, Harpaz N, 123 Selaru FM, Meltzer SJ. MicroRNA-224 negatively regulates p21 expression during late neoplastic progression in inflammatory bowel disease. Inflamm Bowel Dis 2013; 19: 471-480 [PMID: 23399735 DOI: 10.1097/MIB.0b013e31827e78eb]
- 124 Earle JS, Luthra R, Romans A, Abraham R, Ensor J, Yao H, Hamilton SR. Association of microRNA expression with microsatellite instability status in colorectal adenocarcinoma. J Mol Diagn 2010; 12: 433-440 [PMID: 20413677 DOI: 10.2353/jmoldx.2010.090154]
- Balaguer F, Moreira L, Lozano JJ, Link A, Ramirez G, Shen Y, Cuatrecasas M, Arnold M, Meltzer 125 SJ, Syngal S, Stoffel E, Jover R, Llor X, Castells A, Boland CR, Gironella M, Goel A. Colorectal cancers with microsatellite instability display unique miRNA profiles. Clin Cancer Res 2011; 17: 6239-6249 [PMID: 21844009 DOI: 10.1158/1078-0432.CCR-11-1424]
- 126 Kulda V, Pesta M, Topolcan O, Liska V, Treska V, Sutnar A, Rupert K, Ludvikova M, Babuska V, Holubec L Jr, Cerny R. Relevance of miR-21 and miR-143 expression in tissue samples of colorectal carcinoma and its liver metastases. Cancer Genet Cytogenet 2010; 200: 154-160 [PMID: 20620599 DOI: 10.1016/j.cancergencyto.2010.04.015]
- 127 Akao Y, Nakagawa Y, Naoe T. MicroRNAs 143 and 145 are possible common onco-microRNAs in human cancers. Oncol Rep 2006; 16: 845-850 [PMID: 16969504]
- Slaby O, Svoboda M, Fabian P, Smerdova T, Knoflickova D, Bednarikova M, Nenutil R, Vyzula R. 128 Altered expression of miR-21, miR-31, miR-143 and miR-145 is related to clinicopathologic features of colorectal cancer. Oncology 2007; 72: 397-402 [PMID: 18196926 DOI: 10.1159/000113489]
- Diosdado B, van de Wiel MA, Terhaar Sive Droste JS, Mongera S, Postma C, Meijerink WJ, 129 Carvalho B, Meijer GA. MiR-17-92 cluster is associated with 13q gain and c-myc expression during colorectal adenoma to adenocarcinoma progression. Br J Cancer 2009; 101: 707-714 [PMID: 19672269 DOI: 10.1038/sj.bjc.6605037]
- 130 Wang X, Wang J, Ma H, Zhang J, Zhou X. Downregulation of miR-195 correlates with lymph node metastasis and poor prognosis in colorectal cancer. Med Oncol 2012; 29: 919-927 [PMID: 21390519 DOI: 10.1007/s12032-011-9880-5]
- Motoyama K, Inoue H, Takatsuno Y, Tanaka F, Mimori K, Uetake H, Sugihara K, Mori M. Over-131 and under-expressed microRNAs in human colorectal cancer. Int J Oncol 2009; 34: 1069-1075 [PMID: 19287964 DOI: 10.3892/ijo 00000233]
- Xu XM, Qian JC, Deng ZL, Cai Z, Tang T, Wang P, Zhang KH, Cai JP. Expression of miR-21, 132 miR-31, miR-96 and miR-135b is correlated with the clinical parameters of colorectal cancer. Oncol Lett 2012; 4: 339-345 [PMID: 22844381 DOI: 10.3892/ol.2012.714]
- 133 Chen X, Guo X, Zhang H, Xiang Y, Chen J, Yin Y, Cai X, Wang K, Wang G, Ba Y, Zhu L, Wang J, Yang R, Zhang Y, Ren Z, Zen K, Zhang J, Zhang CY. Role of miR-143 targeting KRAS in colorectal tumorigenesis. Oncogene 2009; 28: 1385-1392 [PMID: 19137007 DOI: 10.1038/onc.2008.474]
- 134 Sarver AL, French AJ, Borralho PM, Thayanithy V, Oberg AL, Silverstein KA, Morlan BW, Riska SM, Boardman LA, Cunningham JM, Subramanian S, Wang L, Smyrk TC, Rodrigues CM, Thibodeau SN, Steer CJ. Human colon cancer profiles show differential microRNA expression



depending on mismatch repair status and are characteristic of undifferentiated proliferative states. BMC Cancer 2009; 9: 401 [PMID: 19922656 DOI: 10.1186/1471-2407-9-401]

- 135 Zhang Y, He X, Liu Y, Ye Y, Zhang H, He P, Zhang Q, Dong L, Dong J. microRNA-320a inhibits tumor invasion by targeting neuropilin 1 and is associated with liver metastasis in colorectal cancer. Oncol Rep 2012; 27: 685-694 [PMID: 22134529 DOI: 10.3892/or.2011.1561]
- 136 Roy S, Levi E, Majumdar AP, Sarkar FH. Expression of miR-34 is lost in colon cancer which can be re-expressed by a novel agent CDF. J Hematol Oncol 2012; 5: 58 [PMID: 22992310 DOI: 10.1186/1756-8722-5-58
- 137 Dai X, Chiang Y, Wang Z, Song Y, Lu C, Gao P, Xu H. Expression levels of microRNA-375 in colorectal carcinoma. Mol Med Rep 2012; 5: 1299-1304 [PMID: 22377847 DOI: 10.3892/mmr.2012.815
- Arndt GM, Dossey L, Cullen LM, Lai A, Druker R, Eisbacher M, Zhang C, Tran N, Fan H, Retzlaff 138 K, Bittner A, Raponi M. Characterization of global microRNA expression reveals oncogenic potential of miR-145 in metastatic colorectal cancer. BMC Cancer 2009; 9: 374 [PMID: 19843336 DOI: 10.1186/1471-2407-9-374]
- Zhang Y, Lin C, Liao G, Liu S, Ding J, Tang F, Wang Z, Liang X, Li B, Wei Y, Huang Q, Li X, 139 Tang B. MicroRNA-506 suppresses tumor proliferation and metastasis in colon cancer by directly targeting the oncogene EZH2. Oncotarget 2015; 6: 32586-32601 [PMID: 26452129 DOI: 10.18632/oncotarget.5309]
- 140 Hamfjord J, Stangeland AM, Hughes T, Skrede ML, Tveit KM, Ikdahl T, Kure EH. Differential expression of miRNAs in colorectal cancer: comparison of paired tumor tissue and adjacent normal mucosa using high-throughput sequencing. PLoS One 2012; 7: e34150 [PMID: 22529906 DOI: 10.1371/journal.pone.0034150]
- Wu CW, Ng SS, Dong YJ, Ng SC, Leung WW, Lee CW, Wong YN, Chan FK, Yu J, Sung JJ. 141 Detection of miR-92a and miR-21 in stool samples as potential screening biomarkers for colorectal cancer and polyps. Gut 2012; 61: 739-745 [PMID: 21930727 DOI: 10.1136/gut.2011.239236]
- 142 Lichtenstein GR, Loftus EV, Isaacs KL, Regueiro MD, Gerson LB, Sands BE. ACG Clinical Guideline: Management of Crohn's Disease in Adults. Am J Gastroenterol 2018; 113: 481-517 [PMID: 29610508 DOI: 10.1038/ajg.2018.27]
- Roessner A, Kuester D, Malfertheiner P, Schneider-Stock R. Oxidative stress in ulcerative colitis-143 associated carcinogenesis. Pathol Res Pract 2008; 204: 511-524 [PMID: 18571874 DOI: 10.1016/j.prp.2008.04.011]
- Bressenot A, Cahn V, Danese S, Peyrin-Biroulet L. Microscopic features of colorectal neoplasia in 144 inflammatory bowel diseases. World J Gastroenterol 2014; 20: 3164-3172 [PMID: 24696602 DOI: 10.3748/wjg.v20.i12.3164]
- Collins PD. Strategies for detecting colon cancer and dysplasia in patients with inflammatory bowel 145 disease. Inflamm Bowel Dis 2013; 19: 860-863 [PMID: 23446340 DOI: 10.1097/MIB.0b013e3182802c6a
- 146 O'Connor PM, Lapointe TK, Beck PL, Buret AG. Mechanisms by which inflammation may increase intestinal cancer risk in inflammatory bowel disease. Inflamm Bowel Dis 2010; 16: 1411-1420 [PMID: 20155848 DOI: 10.1002/ibd.21217]
- 147 Itzkowitz SH. Molecular biology of dysplasia and cancer in inflammatory bowel disease. Gastroenterol Clin North Am 2006; 35: 553-571 [PMID: 16952740 DOI: 10.1016/j.gtc.2006.07.002]
- Josse C, Bours V. MicroRNAs and Inflammation in Colorectal Cancer. Adv Exp Med Biol 2016; 148 937: 53-69 [PMID: 27573894 DOI: 10.1007/978-3-319-42059-2 3]
- 149 Choi CR, Bakir IA, Hart AL, Graham TA. Clonal evolution of colorectal cancer in IBD. Nat Rev Gastroenterol Hepatol 2017; 14: 218-229 [PMID: 28174420 DOI: 10.1038/nrgastro.2017.1]
- 150 Kanaan Z, Rai SN, Eichenberger MR, Barnes C, Dworkin AM, Weller C, Cohen E, Roberts H, Keskey B, Petras RE, Crawford NP, Galandiuk S. Differential microRNA expression tracks neoplastic progression in inflammatory bowel disease-associated colorectal cancer. Hum Mutat 2012; 33: 551-560 [PMID: 22241525 DOI: 10.1002/humu.22021]
- Bai J, Li Y, Shao T, Zhao Z, Wang Y, Wu A, Chen H, Li S, Jiang C, Xu J, Li X. Integrating analysis 151 reveals microRNA-mediated pathway crosstalk among Crohn's disease, ulcerative colitis and colorectal cancer. Mol Biosyst 2014; 10: 2317-2328 [PMID: 24949825 DOI: 10.1039/c4mb00169a]
- Ueda Y, Ando T, Nanjo S, Ushijima T, Sugiyama T. DNA methylation of microRNA-124a is a 152 potential risk marker of colitis-associated cancer in patients with ulcerative colitis. Dig Dis Sci 2014; 59: 2444-2451 [PMID: 24825593 DOI: 10.1007/s10620-014-3193-4]
- Wan J, Xia L, Xu W, Lu N. Expression and Function of miR-155 in Diseases of the Gastrointestinal 153 Tract. Int J Mol Sci 2016; 17 [PMID: 27187359 DOI: 10.3390/ijms17050709]
- 154 Fang Z, Tang J, Bai Y, Lin H, You H, Jin H, Lin L, You P, Li J, Dai Z, Liang X, Su Y, Hu Q, Wang F, Zhang ZY. Plasma levels of microRNA-24, microRNA-320a, and microRNA-423-5p are potential biomarkers for colorectal carcinoma. J Exp Clin Cancer Res 2015; 34: 86 [PMID: 26297223 DOI: 10.1186/s13046-015-0198-6
- Al-Mustanjid M, Mahmud SMH, Royel MRI, Rahman MH, Islam T, Rahman MR, Moni MA. 155 Detection of molecular signatures and pathways shared in inflammatory bowel disease and colorectal cancer: A bioinformatics and systems biology approach. Genomics 2020; 112: 3416-3426 [PMID: 32535071 DOI: 10.1016/j.ygeno.2020.06.001]
- 156 Hayden MS, Ghosh S. Shared principles in NF-kappaB signaling. Cell 2008; 132: 344-362 [PMID: 18267068 DOI: 10.1016/j.cell.2008.01.020]



- 157 Wang SW, Sun YM. The IL-6/JAK/STAT3 pathway: potential therapeutic strategies in treating colorectal cancer (Review). Int J Oncol 2014; 44: 1032-1040 [PMID: 24430672 DOI: 10.3892/ijo.2014.2259
- 158 Shi M, Chen X, Ye K, Yao Y, Li Y. Application potential of toll-like receptors in cancer immunotherapy: Systematic review. Medicine (Baltimore) 2016; 95: e3951 [PMID: 27336891 DOI: 10.1097/MD.00000000003951]
- Liu X, Xu Y, Zhou Q, Chen M, Zhang Y, Liang H, Zhao J, Zhong W, Wang M. PI3K in cancer: its 159 structure, activation modes and role in shaping tumor microenvironment. Future Oncol 2018; 14: 665-674 [PMID: 29219001 DOI: 10.2217/fon-2017-0588]
- Huang HY, Lin YC, Li J, Huang KY, Shrestha S, Hong HC, Tang Y, Chen YG, Jin CN, Yu Y, Xu 160 JT, Li YM, Cai XX, Zhou ZY, Chen XH, Pei YY, Hu L, Su JJ, Cui SD, Wang F, Xie YY, Ding SY, Luo MF, Chou CH, Chang NW, Chen KW, Cheng YH, Wan XH, Hsu WL, Lee TY, Wei FX, Huang HD. miRTarBase 2020: updates to the experimentally validated microRNA-target interaction database. Nucleic Acids Res 2020; 48: D148-D154 [PMID: 31647101 DOI: 10.1093/nar/gkz896]
- Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, Clark NR, Ma'ayan A. Enrichr: 161 interactive and collaborative HTML5 gene list enrichment analysis tool. BMC Bioinformatics 2013; 14: 128 [PMID: 23586463 DOI: 10.1186/1471-2105-14-128]
- 162 Kuleshov MV, Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, Koplev S, Jenkins SL, Jagodnik KM, Lachmann A, McDermott MG, Monteiro CD, Gundersen GW, Ma'ayan A. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. Nucleic Acids Res 2016; 44: W90-W97 [PMID: 27141961 DOI: 10.1093/nar/gkw377]
- Xie Z, Bailey A, Kuleshov MV, Clarke DJB, Evangelista JE, Jenkins SL, Lachmann A, 163 Wojciechowicz ML, Kropiwnicki E, Jagodnik KM, Jeon M, Ma'ayan A. Gene Set Knowledge Discovery with Enrichr. Curr Protoc 2021; 1: e90 [PMID: 33780170 DOI: 10.1002/cpz1.90]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

