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Contents

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REVIEW

	Regulatory RNAs, microRNA, long-non coding RNA and circular RNA roles in colorectal cancer stem cells
	Chao HM, Wang TW, Chern E, Hsu SH
765	Role of three-dimensional printing and artificial intelligence in the management of hepatocellular carcinoma: Challenges and opportunities
	Christou CD, Tsoulfas G
-04	
794	Role of sirtuins in esophageal cancer: Current status and future prospects
	Otsuka R, Hayano K, Matsubara H
	MINIREVIEWS
808	Vasoactive intestinal peptide secreting tumour: An overview
	Una Cidon E
820	Management of single pulmonary metastases from colorectal cancer: State of the art
	Chiappetta M, Salvatore L, Congedo MT, Bensi M, De Luca V, Petracca Ciavarella L, Camarda F, Evangelista J, Valentini V, Tortora G, Margaritora S, Lococo F
833	Current guidelines in the surgical management of hereditary colorectal cancers
000	Kudchadkar S, Ahmed S, Mukherjee T, Sagar J
	ORIGINAL ARTICLE
	Basic Study
842	
842	Basic Study
842 858	Basic Study Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4α
	 Basic Study Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4a <i>Li LL, Peng Z, Hu Q, Xu LJ, Zou X, Huang DM, Yi P</i> Bi-specific T1 positive-contrast-enhanced magnetic resonance imaging molecular probe for hepatocellular
	Basic Study Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4a Li LL, Peng Z, Hu Q, Xu LJ, Zou X, Huang DM, Yi P Bi-specific T1 positive-contrast-enhanced magnetic resonance imaging molecular probe for hepatocellular carcinoma in an orthotopic mouse model
858	 Basic Study Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4a <i>Li LL, Peng Z, Hu Q, Xu LJ, Zou X, Huang DM, Yi P</i> Bi-specific T1 positive-contrast-enhanced magnetic resonance imaging molecular probe for hepatocellular carcinoma in an orthotopic mouse model <i>Ma XH, Chen K, Wang S, Liu SY, Li DF, Mi YT, Wu ZY, Qu CF, Zhao XM</i> Xihuang pills induce apoptosis in hepatocellular carcinoma by suppressing phosphoinositide 3-

887 Effect of hepatic artery resection and reconstruction on the prognosis of patients with advanced hilar cholangiocarcinoma

Li YM, Bie ZX, Guo RQ, Li B, Wang CE, Yan F



Contor	World Journal of Gastrointestinal Oncology
Conter	Monthly Volume 14 Number 4 April 15, 2022
897	Prognostic significance of serum inflammation indices for different tumor infiltrative pattern types of gastric cancer
	Wang YF, Yin X, Fang TY, Wang YM, Zhang L, Zhang XH, Zhang DX, Zhang Y, Wang XB, Wang H, Xue YW
920	Regorafenib combined with programmed cell death-1 inhibitor against refractory colorectal cancer and the platelet-to-lymphocyte ratio's prediction on effectiveness
	Xu YJ, Zhang P, Hu JL, Liang H, Zhu YY, Cui Y, Niu P, Xu M, Liu MY
	Clinical Trials Study
935	Genome-wide methylation profiling of early colorectal cancer using an Illumina Infinium Methylation EPIC BeadChip
	Wu YL, Jiang T, Huang W, Wu XY, Zhang PJ, Tian YP



Contents

World Journal of Gastrointestinal Oncology

Monthly Volume 14 Number 4 April 15, 2022

ABOUT COVER

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AIMS AND SCOPE

The primary aim of World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

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ORIGINAL ARTICLE

Basic Study Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4α

Ling-Li Li, Ze Peng, Qian Hu, Li-Jun Xu, Xin Zou, Dong-Mei Huang, Ping Yi

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Abstract

BACKGROUND

Gastric cancer is the third deadliest cancer in the world and ranks second in incidence and mortality of cancers in China. Despite advances in prevention, diagnosis, and therapy, the absolute number of cases is increasing every year due to aging and the growth of high-risk populations, and gastric cancer is still a leading cause of cancer-related death. Gastric cancer is a consequence of the complex interaction of microbial agents, with environmental and host factors, resulting in the dysregulation of multiple oncogenic and tumor-suppressing signaling pathways. Global efforts have been made to investigate in detail the genomic and epigenomic heterogeneity of this disease, resulting in the identification of new specific and sensitive predictive and prognostic biomarkers. Trastuzumab, a monoclonal antibody against the HER2 receptor, is approved in the first-line treatment of patients with HER2+ tumors, which accounts for 13%-23% of the gastric cancer population. Ramucirumab, a monoclonal antibody against VEGFR2, is currently recommended in patients progressing after first-line treatment. Several clinical trials have also tested novel agents for advanced gastric cancer but mostly with dis-appointing results, such as anti-EGFR and anti-MET monoclonal antibodies. Therefore, it is still of great significance to screen specific molecular targets for gastric cancer and drugs directed against the molecular targets.

AIM

To investigate the effect and mechanism of berberine against tumor growth in gastric cancer xenograft models and to explore the role of hepatocyte nuclear



factor 4α (HNF4 α)-WNT5 a/β -catenin pathways played in the antitumor effects of berberine.

METHODS

MGC803 and SGC7901 subcutaneous xenograft models were established. The control group was intragastrically administrated with normal saline, and the berberine group was administrated intragastrically with 100 mg/kg/d berberine. The body weight of nude mice during the experiment was measured to assess whether berberine has any adverse reaction. The volume of subcutaneous tumors during this experiment was recorded to evaluate the inhibitory effect of berberine on the growth of MGC803 and SGC7901 subcutaneous transplantation tumors. Polymerase chain reaction assays were conducted to evaluate the alteration of transcriptional expression of HNF4 α , WNT5a and β -catenin in tumor tissues and liver tissues from the MGC803 and SGC7901 xenograft models. Western blotting and IHC were performed to assess the protein expression of HNF4 α , WNT5a and β -catenin in tumor tissues and liver tissues from the MGC803 and SGC7901 xenograft models.

RESULTS

In the both MGC803 and SGC7901 xenograft tumor models, berberine significantly reduced tumor volume and weight and thus retarded the growth rate of tumors. In the SGC7901 and MGC803 subcutaneously transplanted tumor models, berberine down-regulated the expression of HNF4 α , WNT5a and β -catenin in tumor tissues from both transcription and protein levels. Besides, berberine also suppressed the protein expression of HNF4 α , WNT5a and β -catenin in liver tissues.

CONCLUSION

Berberine retarded the growth of MGC803 and SGC7901 xenograft model tumors, and the mechanism behind these anti-growth effects might be the downregulation of the expression of HNF4 α -WNT5 a/β -catenin signaling pathways both in tumor tissues and liver tissues of the xenograft models.

Key Words: Berberine; Gastric cancer; Xenograft models; Hepatocyte nuclear factor 4α ; WNT5a

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Core Tip: Hepatocyte nuclear factor 4α (HNF4 α) is a member of the nuclear receptor transcription factor family, which is at the center of a complex transcriptional regulatory network and regulates the biological effects of different pathways via transcriptional regulation of differential target genes. The role of HNF4α in gastric cancer is poorly understood, so the study of the berberine targeting HNF4 α in gastric cancer cell xenograft models is of great significance. In this study, we showed that the inhibition of HNF4 α genes/proteins by berberine was correlated with the tumor inhibition in gastric cancer xenografts. Further experiments also indicated that berberine downregulated HNF4a to exert its antineoplastic activity in vivo.

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INTRODUCTION

Gastric cancer is the third deadliest cancer in the world and ranks second in the incidence and mortality of cancers in China[1,2]. Despite advances in prevention, diagnosis, and therapy, the absolute number of cases is increasing every year due to aging and the growth of high-risk populations, and gastric cancer is still a leading cause of cancer-related death[1,3]. Gastric cancer is a consequence of the complex interaction of microbial agents, with environmental and host factors, resulting in the dysregulation of multiple oncogenic and tumor-suppressing signaling pathways[1]. Global efforts have been made to investigate in detail the genomic and epigenomic heterogeneity of this disease, resulting in the identification of new specific and sensitive predictive and prognostic biomarkers[4]. Trastuzumab, a monoclonal antibody against the HER2 receptor, is approved in the first-line treatment of patients with HER2+ tumors, which accounts for 13%-23% of the gastric cancer population[5]. Ramucirumab, a monoclonal antibody against VEGFR2, is currently recommended in patients progressing after first-line treatment[6]. Several clinical trials have also tested novel agents, such as anti-EGFR and anti-MET



monoclonal antibodies, to treat advanced gastric cancer but mostly with disappointing results[3]. Therefore, it is still of great significance to screen specific molecular targets for gastric cancer and drugs directed against the molecular targets.

Several epidemiological studies have found that diabetes is linked with an increased risk of gastric cancer and can increase the risk of death in patients with gastric cancer, resulting in a worse prognosis [7,8]. Recently, many studies have focused on diabetes and its causal relation with neoplasms[9]. The emergence of cancer-related properties was likely a change in biological characteristics caused by the hyperglycemic environment created by diabetes, which provides the opportunity to trigger abnormal expression of the healthy genome and promote changes in biological characteristics[10]. It was demonstrated that glucose increased ATP levels to inhibit AMPK and induce the increased expression of hepatocyte nuclear factor 4α (HNF4 α)[1]. Hepatocyte nuclear factor 4α is a member of the nuclear receptor transcription factor family, which has a conserved DNA-binding domain and a ligand-binding domain [12]. HNF4 α is at the center of a complex transcriptional regulatory network and regulates the biological effects of different pathways *via* transcriptional regulation of differential target genes[13]. HNF4 α is involved in a variety of human diseases. HNF4 α plays an important role in the development and treatment of diabetes. Neonatal patients with diabetes or young adults with diabetes with mutations in the HNF4 α gene usually respond better to oral treatment with sulfonylureas[14]. Furthermore, many studies also suggest that HNF4a might be related to the occurrence and development of tumors. HNF4a was found to be highly expressed in various tumor tissues, such as lung cancers and liver cancers[15,16]. Compared with normal gastric mucosa, the protein and transcript levels of HNF4 α in gastric cancer tissues and gastric cancer cell lines are also at higher levels[17]. The role of HNF4 α in gastric cancer has gradually received attention. HNF4 α can promote the intestinal metaplasia of the gastric mucosa[18]. HNF4α can also be used as a gold standard marker to distinguish gastric metastasis from primary gastric cancer and breast cancer. Studies showed that $HNF4\alpha$ was positively expressed in all patients diagnosed with primary gastric adenocarcinoma and negatively expressed in all cases of primary breast cancer[19,20].

The hyperglycemic environment created by diabetes provides the opportunity to trigger abnormal expression of the healthy genome and promote changes in biological characteristics[10]. The hyperglycemic state observed in the diabetic milieu is predicted to enhance the gastric cancer risk in prediabetic and diabetic individuals[9]. Studies have shown that some diabetes treatment drugs can effectively reduce the risk of cancer and improve prognosis, which is of great significance for understanding the relationship between diabetes and tumorigenesis and provides a new idea for the research of cancer prevention and treatment^[21]. Berberine, an extract from the Chinese herbal medicine Coptis, has been widely used in clinical treatment and has shown excellent efficacy in the treatment of intestinal infections. Multiple studies have displayed the potential therapeutic effect of berberine on diabetes [22]. Berberine can target HNF4 α to inhibit gluconeogenesis in the treatment of type 2 diabetes and its complications^[23]. Berberine improvs insulin resistance associated not only with gut microbiota alteration in branched-chain amino acid biosynthesis, but also with branched-chain amino acid catabolism in liver and adipose tissues[24]. Berberine alleviates tau hyperphosphorylation and axonopathy-associated with diabetic encephalopathy by restoring the PI3K/Akt/GSK3β pathway[25]. Meanwhile, several studies have also showned the potential therapeutic effects of berberine against tumor growth [26-28]. Berberine exerts an inhibitory effect on cell growth and induces apoptosis by inhibiting the activation of EFGR signaling^[27]. Berberine can also suppress the growth and induce apoptosis of colorectal cancer cell lines through regulation of the Long Non-Coding RNA (lncRNA) Cancer Susceptibility Candidate 2 (CASC2)/AU-Binding Factor 1 (AUF1)/B-Cell CLL/Lymphoma 2 (Bcl-2) Axis[28]. The structural interaction of berberine with Aldo-keto reductase family 1 member C3 is attributed to the suppression of Aldo-keto reductase family 1 member C3 enzyme activity and the inhibition of 22Rv1 prostate cancer cell growth by decreasing the intracellular androgen synthesis[29]. Berberine could effectively affect both tumor outgrowth and spontaneous metastasis in triple-negative breast cancer, which was associated with the inhibition of NLRP3 inflammasome pathway[30].

Whether HNF4 α , which plays an important role in regulating the progression of diabetes and tumors, is just the right target for berberine to exert anti-gastric cancer effects remains to be elucidated. Our previous experiments demonstrated that berberine could induce cycle arrest of gastric cancer cells by targeting AMPK/HNF4α/WNT5a pathways in vitro[31]. In this study, berberine was administrated intragastrically to MGC803 and SGC7901 subcutaneous gastric cancer xenograft models. We investigated the effects of berberine against tumor growth and the role of HNF4 α -WNT5 a/β -catenin pathways involved in the antitumor effects of berberine.

MATERIALS AND METHODS

Chemicals and materials

Berberine chloride with a purity of 98% was purchased from the Sigma-Aldrich, United States. Antibodies against β-catenin were purchased from Cell Signaling Technologies (Boston, MA, United States), WNT5a was purchased from Abgent and HNF4a was purchased from Santa Cruz Biotechnology



(Santa Cruz, CA, United States).

Cell lines and cell culture

The gastric cancer cell lines MGC803 and SGC7901 were supplied by Hubei Biossci Biological Co., Ltd. Both cell lines were routinely cultured in RPMI1640(HyClone, China), supplemented with 10% fetal bovine serum (FBS) (SiJiQing, China), and 1% penicillin-streptomycin solution at 37 °C in a 5% CO2 humidified atmosphere. All experiments were performed when cells were in a logarithmic phase.

In vivo tumor xenograft model

Female BALB/c nude mice approximately 4 weeks old (13 g-17 g) were purchased from Hunan Slake Jingda Experimental Co., Ltd. (Hunan, China). Upon arrival, these animals were maintained under specific-pathogen-free conditions ($23 \pm 2 \degree C$ and $55 \pm 5\%$ humidity) with an automatically controlled 12h light/dark cycle and with a free access to sterilized flood and autoclaved water. After mice were acclimatized for about seven day, MGC803 or SGC7901 gastric cancer cells (10⁷ cells per mouse) were subcutaneously injected into the right flank of mice. Animals bearing tumors were randomly divided into two groups, and treatment was initiated after injection for 72 h when a mass of more than 6 mm in maximal diameter was identified in each mouse with daily intragastric administration of normal saline (control group) and 100 mg/kg berberine (BBR group). All animals were sacrificed on Day 18 after treatment and blood as well as tissues from all animals were collected. Tumor xenograft length (L) and width (W) were measured at two-day intervals with Vernier calipers, and tumor volume (V) was calculated using the Formula (V = $W^2 \times L/2$). Mouse body weight was monitored every two d. Animal experiments were performed according to the institutional guidelines and regulations approved by the Huazhong University of Science and Technology Institutional Animal Care and Use Committee.

Immunohistochemistry staining assay

The tumor tissues removed from mice were fixed with 4% formaldehyde and then embedded in paraffin, and then 4µm thick tissue sections were made. After heating in an oven at 60 °C for 1 h, the sections was dewaxed in a dewaxing agent and rehydrated by a concentration-gradient of alcohol (100%, 95%, 85%, 75%). Tissue sections were boiled in 10 mmol/L sodium citrate buffer (pH 6.0) at high temperature to recover the antigen and incubated with 3% hydrogen peroxide for 10 minutes to block endogenous peroxidase activity. Sections were incubated with primary antibodies against HNF4a (1: 200), WNT5a (1: 80) and β -catenin (1: 100) at 4 °C overnight. Then the tissue was incubated with a secondary antibody and observed under a microscope.

Total RNA Extraction and real-time polymerase chain reaction

Total RNA was extracted using the TRIzol Reagent (Magent, Wuhan) according to the manufacturer's instructions, and then cDNA was synthesized from 2 µg of total RNA using the 5X All-In-One RT MasterMix (ABP) at 25 °C for 10 min, 42 °C for 15 min and 85 °C for 5 min. Real-time polymerase chain reaction (PCR) was used to detect the mRNA expression levels of HNF4 α , WNT5a and β -catenin. PCR reactions were performed using LightCycler 480 (Roche Applied Science) in a total volume of 20 µL, including 10 µL of 2X SYBR Green qPCR Master Mix (ABP) at 95 °C for 10 min, 95 °C for 15 s and 60 °C for 60 s, 40 cycles according to the manufacturer's instructions. The relative gene expression was calculated with normalization to an endogenous reference (GAPDH).

The primers were as follows: GENE; Forward Primer (5'-3'); Reverse Primer (5'-3'); HNF4 α ; ATGCGACTCTCTAAAACCCTTG; ACCTTCAGATGGGGACGTGT; WNT5a; CAACTGGCAG-GACTTTCTCAA; CCTTCTCCAATGTACTGCATGTG; β-catenin; ATGGAGCCGGACAGAAAAGC; TGGGAGGTGTCAACATCTTCTT; GAPDH; TGGCCTTCCGTGTTCCTAC; GAGTTGCTGT-TGAAGTCGCA.

Western blotting

Total protein was extracted from the tumor tissues and liver tissues, and its concentration was determined by the BCA assay. Then, equal amounts of proteins (30ug/Lane) were loaded for electrophoresis, seperated by SDS-polyacrylamide electrophoresis gel (SDS-PAGE) and subsequently transferred to nitrocellulose membranes (NC membrane) (Millipore, United States). After blocking with 5% nonfat milk at room teamperature for 1h, the NC membranes were incubated with the primary antibodies overnight with gentle agitation at 4 °C. After washing with TBST for three times, the NC membranes were incubated with the secondary antibodies for 1h at room temperature, followed by visualization with a near-infrared double color laser imaging system (Odyssey, Lincoln, NE, United States).

Statistical analysis

All experiments were repeated at least three times. All data are expressed as the mean ± standard deviation. All data were plotted by GraphPad Prism 6 software. Statistical analysis was performed using SPSS (version 20.0) software. A t test was also performed to determine differences between groups. P < 0.05 was considered statistically significant.



RESULTS

Berberine inhibited the growth of MGC803 and SGC7901 gastric cancer xenograft tumors

To explore the effect of berberine on gastric cancer xenograft models, we established MGC803 and SGC7901 xenograft tumor models. As shown in Figure 1A, intragastric administration of berberine significantly reduced the tumor weight of MGC803 and SGC7901 subcutaneously transplanted tumors, and the reduction rates were approximately 50.0% and 60.9% of the control group respectively (P < P0.05). In addition, as shown in Figure 1B, berberine led to 48.6% inhibition of tumor size in MGC803 xenograft models and 51.3% inhibition of tumor volume in SGC7901 xenograft models when compared to the control group. Berberine significantly inhibited the growth rate of MGC803 and SGC7901 xenograft tumors. Moreover, berberine did not affect the body weight of MGC803 and SGC7901 xenograft models in this experiment (Figure 1C, P > 0.05). These results indicated that berberine could significantly retard the growth of MGC803 and SGC7901 xenograft tumors, which was coordinated with the experimental results of Yi et al[32].

Berberine inhibited HNF4α expression in tumor and liver tissues in both MGC803 and SGC7901 xenograft tumor models.

HNF4 α is at the center of a complex transcriptional regulatory network and is implicated in the initiation and progression of gastric cancer. To investigate the role of $HNF4\alpha$ in the growth of gastric cancer and the underlying mechanism, we examined the effect of berberine on the expression of HNF4 α in tumor tissues and liver tissues of MGC803 and SGC7901 xenograft tumor models. Immunohistochemical staining showed that berberine downregulated the expression of $HNF4\alpha$ in tumor tissues from MGC803 and SGC7901 xenograft tumor models. Besides, in the tumor tissues from the MGC803 and SGC7901 xenograft tumor models, berberine significantly reduced both the transcriptional expression and protein expression of HNF4 α compared with the control group. Moreover, berberine inhibited the mRNA level of HNF4 α in the liver tissues from the MGC803 and SGC7901 xenograft models. Furthermore, berberine also reduced the protein expression of HNF4 α in liver tissue from the MGC803 and SGC7901 xenograft models. Therefore, berberine inhibited the expression of HNF4 α in MGC803 and SGC7901 xenograft models at both the transcriptional and posttranscriptional levels, and the reduction of HNF4α significantly inhibited the growth of tumor cells (Figure 2).

Berberine inhibited WNT5a expression in tumor and liver tissues in both MGC803 and SGC7901 xenograft tumor models.

WNT5a has been proven to be involved in the development and progression of gastric cancers. WNT5a was the downstream of HNF4 α and HNF4 α regulated WNT signaling through its target gene WNT5a, a potential prognostic marker of diffuse-type gastric tumours[33]. Therefore, we examined the alteration of the expression levels of WNT5a in tumor tissues and liver tissues from xenograft tumor models. Immunohistochemical staining results showed that compared with the control group, the expression level of WNT5a in tumor tissues from both the MGC803 and SGC7901 xenograft tumor models in the berberine group was significantly down-regulated. Furthermore, berberine also significantly reduced the mRNA and protein levels of WNT5a in the tumor tissues from both the MGC803 and SGC7901 xenograft models compared to the control group. Whereas, in the liver tissues from xenograft models, berberine reduced the mRNA level of WNT5a in MGC803 xenograft models, but did not affect the transcription level of WNT5a in SGC7901 xenograft models. However, berberine reduced the protein expression of WNT5a in the liver tissues from MGC803 as well as SGC7901 xenograft model. Therefore, berberine inhibited WNT5a protein expression in MGC803 and SGC7901 xenograft models, and the downregulation of WNT5a protein expression was associated with the retardation of the tumor growth rate (Figure 3).

Berberine inhibited β-catenin expression in tumor and liver tissues in both MGC803 and SGC7901 xenograft tumor models.

However, as an extracellular ligand, WNT5a activates different intracellular signaling cascades: βcatenin-dependent and β -catenin-independent pathways[34]. In the classical WNT pathway, β -catenin acts as a major mediator and functions as a protein organizer by interacting with numerous partners at the membrane, in the cytosol, and in the nucleus [35,36]. To further illustrate the mechanism of the antitumor effects of berberine, we also detected the expression level of β -catenin in tumor tissues and liver sections upon treatment with berberine. Immunohistochemical staining results showed that in the SGC7901 and MGC803 xenograft models, berberine significantly reduced the expression of β -catenin in tumor tissues. In addition, in both the MGC803 and SGC7901 xenograft tumor models, both the transcriptional and the protein levels of β -catenin in tumor tissue in the berberine group were significantly reduced. Besides, berberine reduced the mRNA level of β-catenin in the liver tissues from SGC7901 xenograft tumor models but did not affect the transcription level of β-catenin in MGC803 xenograft tumor models. Simultaneously, berberine also downregulated the protein expression of β catenin in the liver tissues of MGC803 xenograft tumor models and SGC7901 xenograft tumor models.



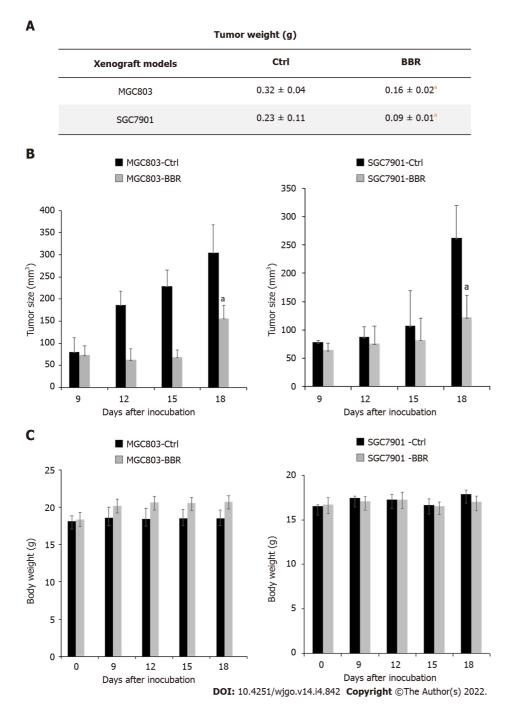
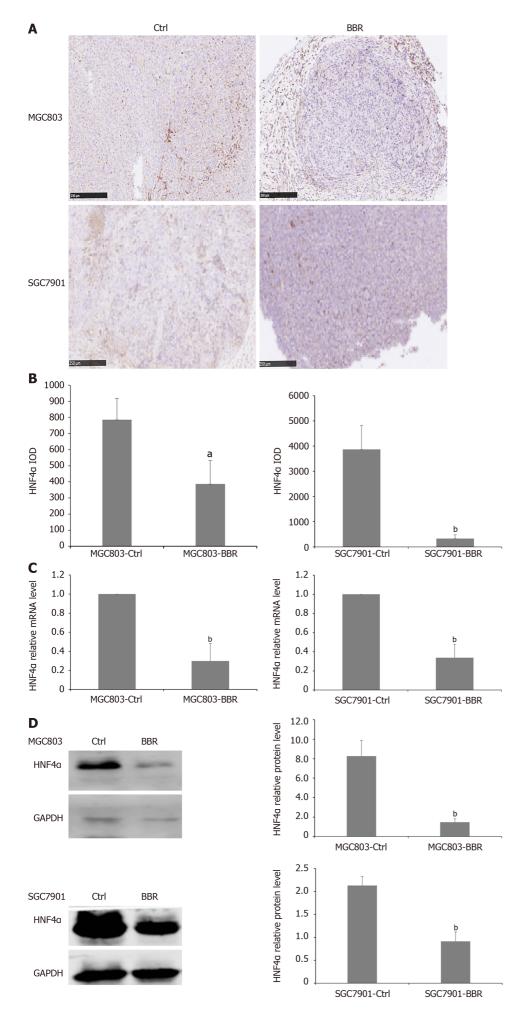


Figure 1 Berberine inhibited the growth of MGC803 and SGC7901 xenograft tumors. A: After 18 d of drug treatment, tumors were removed from the mice and weighed; B: Changes in tumor size of mice during the experiment; C: Changes in body weight of mice during the experiment. aP < 0.05. Ctrl: The control group; BBR: The berberine group.

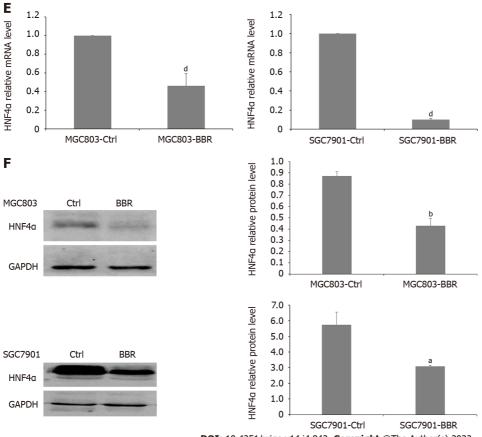
> Therefore, berberine downregulated the protein expression of β-catenin in both MGC803 and SGC7901 xenograft models, and berberine targeted the WNT5a/β-catenin signaling pathways to exert its growth inhibition effects to MGC803 and SGC7901 xenograft model tumors (Figure 4).

DISCUSSION

Berberine is a benzyl tetraisoquinoline alkaloid extracted from Chinese herbal medicines such as Coptis, Phellodendron chinense and Hydrastis canadensis. Berberine has been used clinically for many years and is effective in counteracting numerous diseases such as gastroenteritis, abdominal pain and diarrhea, with antimicrobial and anti-inflammatory properties[37]. Over the past decades, growing evidence clearly indicates that berberine can improve insulin-resistance and the levels of plasma lipids and glucose in type 2 diabetes animal models and humans[38-40]. Berberine could modulate the



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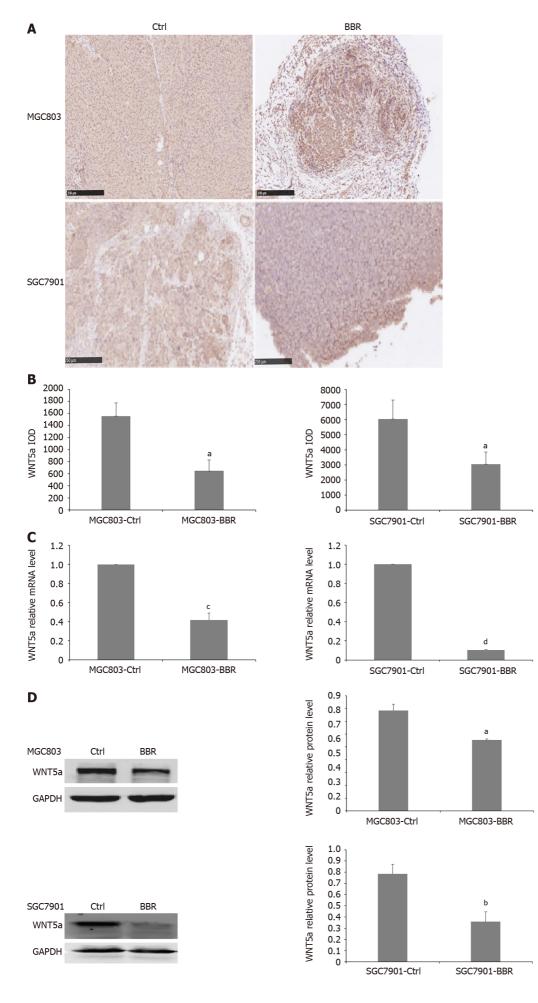
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Figure 2 Berberine inhibited hepatocyte nuclear factor 4 α expression in MGC803 and SGC7901 xenograft tumor models. A: Immunohistochemistry showed that berberine reduced the expression of hepatocyte nuclear factor 4 α (HNF4 α) in tumor tissues of MGC803 and SGC7901 xenograft tumor models; B: The quantification of immunohistochemistry of the expression of HNF4 α in tumor tissues of MGC803 and SGC7901 xenograft tumor models; C: The effect of berberine on HNF4 α mRNA expression in tumor tissues of MGC803 and SGC7901 xenograft tumor models; D: The effect of berberine on HNF4 α protein expression in tumor tissues of MGC803 and SGC7901 xenograft tumor models; E: The effect of berberine on HNF4 α mRNA level in liver tissues of MGC803 and SGC7901 xenografts; F: The effect of berberine on HNF4 α protein expression in liver tissues of MGC803 and SGC7901 xenografts. ^a*P* < 0.05, ^b*P* < 0.01, ^d*P* < 0.0001. Ctrl: The control group; BBR: The berberine group.

> composition of the gut microbiome and reduce body weight, blood glucose levels, and intestinal inflammation in diabetic mice, which demonstrates its effectiveness in the reduction of diabetic complications in this model[38]. Berberine also attenuated intestinal mucosal barrier dysfunction and immune barrier damage in type 2 diabetic rats[39]. Berberine stimulated insulin secretion in impaired glucose tolerance rats[40]. Recently, berberine has shown potential therapeutic effects in treating cancers[26]. Berberine inhibited the proliferation of SGC-7901 cells and induced apoptosis[41]. Berberine also inhibited EGFR signaling and enhanced the antitumor effects of EGFR inhibitors in gastric cancer[27]. Berberine could also inactivate MAPK signaling pathways to exert growth inhibition effects on MGC803 cells[42]. In this experiment, berberine was discovered to induce a 48.6% inhibition of tumor growth in MGC803 xenograft models and 51.3% retardation of tumor growth in SGC7901 xenograft models without any influence on body weight[42]. Therefore, berberine might be an effective and safe drug candidate for treating gastric cancer. However, whether there exists a shared molecular target which is responsible for the antitumor effects of berberine remains unclear?

> HNF4 α is a transcription factor with important roles in liver and gastrointestinal tract development, hepatocyte differentiation, and lipid and glucose metabolism[43]. HNF4 α is at the center of a complex transcriptional regulatory network where its disruption is directly linked to glucose metabolism and lipid metabolism[44]. The activation of HNF4 α may represent a novel single agent for the treatment of insulin resistance[44]. The activation of HNF4 α stimulates glucose metabolism in acute myeloid leukemia cells[45]. What's more, the regulation of HNF4 α expression and activity is highly complex, reflecting on the downstream transcriptional networks with diverse functional roles including drug metabolism, lipid homeostasis, gluconeogenesis, cell adhesion, proliferation, and apoptosis[46]. Our previous studies found that HNF4 α was involved in the glucose metabolism of DM[47]. Recently, one further study illustrated that inhibition of HNF4 α expression could induce cell cycle arrest and apoptosis in gastric cancer cells, thereby exerting antitumor growth effects[31], corresponding to the experiments in which inhibitory RNA and pharmacological inhibition of HNF4 α demonstrated antineoplastic activity *in vitro* and *in vivo via* downregulation of cyclins, cell cycle arrest and apoptosis in gastric





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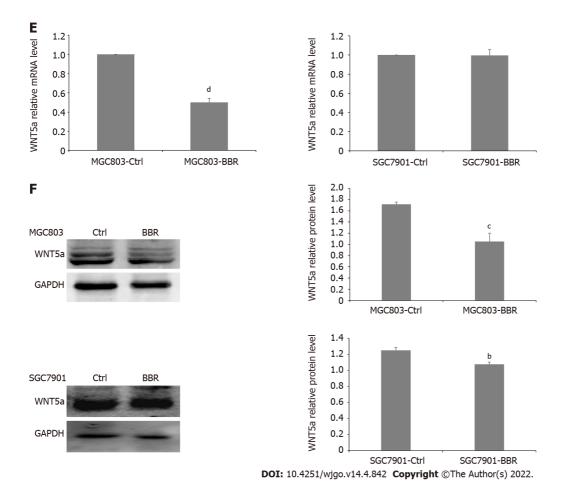
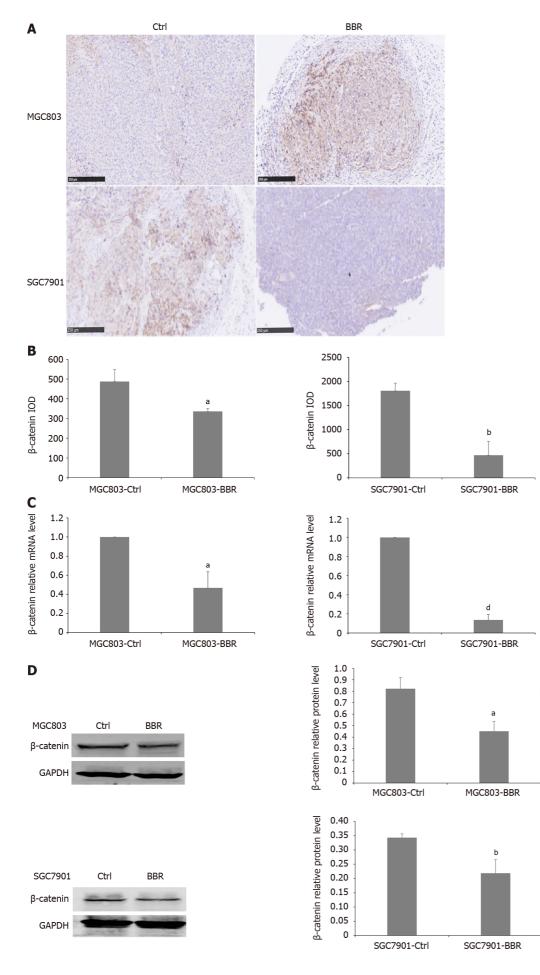


Figure 3 Berberine inhibited WNT5a expression in MGC803 and SGC7901 xenograft tumor models. A: Immunohistochemistry showed that berberine reduced the expression of WNT5a in tumor tissues of MGC803 and SGC7901 xenograft tumor models; B: The quantification of immunohistochemistry of the expression of WNT5a in tumor tissues of MGC803 and SGC7901 xenograft tumor models; C: The effect of berberine on WNT5a mRNA level in tumor tissues of MGC803 and SGC7901 xenograft tumor models; C: The effect of berberine on WNT5a mRNA level in tumor tissues of MGC803 and SGC7901 xenograft tumor models; E: The effect of berberine on WNT5a mRNA expression in liver tissues of MGC803 and SGC7901 xenografts; F: The effect of berberine on WNT5a protein expression in liver tissues of MGC803 and SGC7901 xenografts. ^aP < 0.05, ^bP < 0.01, ^cP < 0.001, ^dP < 0.0001. Ctrl: The control group; BBR: The berberine group.

cancer[33]. HNF4 α downregulation could promote tumor migration and invasion in renal cell carcinoma[48]. HNF4 α promotes the malignant phenotype of acute myeloid leukemia cells[45]. In this study, berberine could suppressed the expression of HNF4 α in tumor tissues and liver tissues from MGC803 and SGC7901 gastric cancer xenograft tumor models. More importantly, the reduced expression of HNF4 α might to be associated with the growth retardation of tumors from gastric cancer xenograft models. Therefore, berberine might target HNF4 α to inhibit tumor growth in MGC803 and SGC7901 xenograft models.

Studies have shown that WNT5a contains several highly conserved HNF4 α binding sites in its promoter region [49]. The expression level of WNT5a decreases after HNF4 α gene knockout, which further inhibits cellular glucose uptake and cell proliferation, and also induces apoptosis[33,45]. WNT5a is a member of the WNT family and can activate classical WNT (β -catenin-dependent) signaling pathways [50]. The WNT/ β -catenin signaling pathway has been recognized as a potentially therapeutic target for cancers [51]. In the classical WNT pathway, β -catenin acts as a major mediator, facilitates signal transduction, and participates in embryogenesis, cell differentiation, and proliferation processes. When the WNT pathway is on, the cytoplasmic concentration of β -catenin increases, and then it translocates into the nucleus, which activates more than one hundred target genes of the WNT pathway, while when the WNT pathway is off, β -catenin is degraded by the proteasome [52]. In this experiment, we found that the protein expression of WNT5a and β -catenin was both downregulated by berberine, indicating that inactivation of the WNT/ β -catenin pathway resulted from berberine. The WNT/ β -catenin pathway is involved in the process of tumorigenesis, invasion and metastasis. The role of WNT5a in the occurrence and progression of gastric cancer has gradually been elucidated. It was confirmed that downregulation of WNT5a expression in gastric cancer cells could inhibit EMT onset and increased WNT5a expression could also result in poor differentiation in GC[53,54]. By activating the WNT5a signaling pathway, gastric cancer cell growth was promoted and G1-S cell cycle transition was induced [55]. Overexpression of WNT5a is also associated with enhanced tumor aggressiveness [56,57]. WNT5a was also a potential





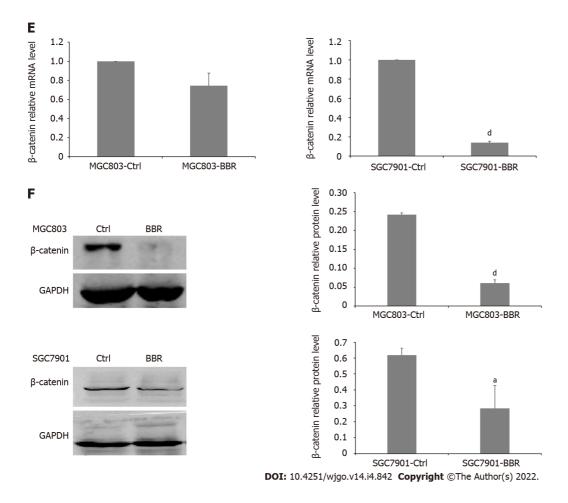


Figure 4 Berberine inhibited β-catenin expression in MGC803 and SGC7901 xenograft tumor models. A: Immunohistochemistry showed that berberine reduced the expression of β -catenin in tumor tissues of MGC803 and SGC7901 xenograft tumor models; B: The quantification of immunohistochemistry of the expression of β-catenin in tumor tissues of MGC803 and SGC7901 xenograft tumor models; C: The effect of berberine on β-catenin mRNA alteration in tumor tissues of MGC803 and SGC7901 xenograft tumor models; D: The effect of berberine on β-catenin protein expression in tumor tissues of MGC803 and SGC7901 xenograft tumor models; E: The effect of berberine on β-catenin mRNA in liver tissues of MGC803 and SGC7901 xenografts: F: The effect of berberine on β-catenin protein expression in liver tissues of MGC803 and SGC7901 xenografts. ^aP < 0.05; ^bP < 0.01; ^cP < 0.001; ^dP < 0.0001. Ctrl: The control group; BBR: The berberine group.

> suppressor of EMT and downregulation of WNT5a mRNA and protein by EGF is necessary for EGFinduced EMT in gastric cancer SGC-7901 cells[58]. The role of WNT5a in the development of gastric cancer is controversial. Recently the role of β -catenin in promoting the migration and invasion of gastric cancer has also been elucidated. CD44, which interacts with IL-13Rα2, promotes gastric cancer development and metastasis through regulation of the β -catenin signaling pathways[59]. HOXA11-AS not only promotes gastric cancer cell migration and invasion in vitro, but also promotes gastric cancer cell metastasis *in vivo*, at least in part, by regulating β -catenin[60]. Studies have also shown that the berberine combined drug HMQ1611 negatively regulates the WNT5a signaling pathway by upregulating axin and inhibiting nuclear transport of β -catenin, thereby inhibiting cancer cell proliferation and metastasis [61]. This experiment demonstrated that the reduction in the expression of WNT5a/ β -catenin signaling pathway is associated with tumor growth suppression in MGC803 and SGC7901 xenograft tumor models. This experiment displayed that berberine could downregulate the expression of WNT5a/ β -catenin in tumor tissues and liver tissues of MGC803 and SGC7901 gastric cancer xenograft tumor models at the protein level, and thereby inhibiting the growth of MGC803 and SGC7901 gastric cancer xenograft tumors.

CONCLUSION

In conclusion, we found that berberine could exert growth retardation effects on both MGC803 and SGC7901 xenograft tumors, the mechanism behind which might associate with the simultaneous reduction of both the HNF4 α and WNT5a/ β -catenin from the tumor tissues and liver tissues in the MGC803 and SGC7901 xenograft models. Therefore HNF4 α might be a potential target through which berberine exerts the effects of countering against gastric cancers.



ARTICLE HIGHLIGHTS

Research background

Gastric cancer is the third deadliest cancer in the world and ranks second in incidence and mortality of cancers in China. Despite advances in prevention, diagnosis, and therapy, the absolute number of cases is increasing every year due to aging and the growth of high-risk populations, and gastric cancer is still a leading cause of cancer-related death. Gastric cancer is a consequence of the complex interaction of microbial agents with environmental and host factors, resulting in the dysregulation of multiple oncogenic and tumor-suppressing signaling pathways. Global efforts have been undertaken to investigate in detail the genomic and epigenomic heterogeneity of this disease, resulting in the identification of new specific and sensitive predictive and prognostic biomarkers. Trastuzumab, a monoclonal antibody against the HER2 receptor, is approved in the first-line treatment of patients with HER2+ tumors, which accounts for 13%-23% of the gastric cancer population. Ramucirumab, a monoclonal antibody against VEGFR2, is currently recommended in patients progressing after first-line treatment. Several clinical trials have also tested novel agents, such as the anti-EGFR and the anti-MET monoclonal antibodies, for advanced gastric cancer but mostly with disappointing results. Therefore, screening specific molecular targets for gastric cancer and drugs directed against the molecular targets is still urgently needed.

Research motivation

To screen specific molecular targets for gastric cancer and drugs directed against the molecular targets is still urgently needed.

Research objectives

To investigate the effect and mechanism of berberine against tumor growth in gastric cancer xenograft models and to explore the role of hepatocyte nuclear factor 4α (HNF4 α)-WNT5a/ β -catenin pathway played in the antitumor effects of berberine.

Research methods

MGC803 and SGC7901 subcutaneous xenograft models were established. The control group was intragastrically administrated with normal saline, and the berberine group was administrated intragastrically with 100mg/kg/d berberine. The body weight of nude mice during the experiment was measured to assess whether berberine has any adverse reaction. The volume of subcutaneous tumors during this experiment was recorded to evaluate the inhibitory effect of berberine on the growth of MGC803 and SGC7901 subcutaneous transplantation tumors. Polymerase chain reaction assays were conducted to evaluate the alteration of transcriptional expression of HNF4 α , WNT5a and β -catenin in tumor tissues and liver tissues from the MGC803 and SGC7901 xenograft models. Western blotting and IHC were performed to assess the protein expression of HNF4 α , WNT5a and β -catenin in tumor tissues and liver tissues from the MGC803 and SGC7901 xenograft models.

Research results

In both MGC803 and SGC7901 xenograft tumor models, berberine significantly reduced tumor volume and weight and retarded the growth rate of tumors. In the SGC7901 and MGC803 subcutaneously transplanted tumor models, berberine downregulated the expression of HNF4 α , WNT5a and β -catenin in tumor tissues from both transcription and protein levels. Besides, berberine also suppressed the protein expression of HNF4 α , WNT5a and β -catenin in liver tissues.

Research conclusions

Berberine retarded the growth of MGC803 and SGC7901 xenograft model tumors, and the mechanism behind this antigrowth effects might be the downregulation of the expression of $HNF4\alpha$ -WNT5a/ β catenin signaling pathways both in tumor tissues and liver tissues of the xenograft models.

Research perspectives

 $HNF4\alpha$ might be a potential target through which berberine exerts effects of both improving diabetes and countering gastric cancers.

FOOTNOTES

Author contributions: Yi P designed the research; Li LL, Peng Z, Xu LJ and Zou X performed the research; Xu LJ, and Zou X contributed analytic tools; Li LL and Peng Z analyzed the data; Li LL wrote the manuscript; Yi P, Hu Q and Huang DM revised the manuscript.

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REFERENCES

- 1 Camargo MC, Figueiredo C, Machado JC. Review: Gastric malignancies: Basic aspects. Helicobacter 2019; 24 Suppl 1: e12642 [PMID: 31486241 DOI: 10.1111/hel.12642]
- 2 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- Serra O, Galán M, Ginesta MM, Calvo M, Sala N, Salazar R. Comparison and applicability of molecular classifications for 3 gastric cancer. Cancer Treat Rev 2019; 77: 29-34 [PMID: 31195213 DOI: 10.1016/j.ctrv.2019.05.005]
- Pellino A, Riello E, Nappo F, Brignola S, Murgioni S, Djaballah SA, Lonardi S, Zagonel V, Rugge M, Loupakis F, Fassan M. Targeted therapies in metastatic gastric cancer: Current knowledge and future perspectives. World J Gastroenterol 2019; 25: 5773-5788 [PMID: 31636471 DOI: 10.3748/wjg.v25.i38.5773]
- Bang YJ, Van Cutsem E, Feyereislova A, Chung HC, Shen L, Sawaki A, Lordick F, Ohtsu A, Omuro Y, Satoh T, Aprile 5 G, Kulikov E, Hill J, Lehle M, Rüschoff J, Kang YK; ToGA Trial Investigators. Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. Lancet 2010; 376: 687-697 [PMID: 20728210 DOI: 10.1016/s0140-6736(10)61121-x]
- Fuchs CS, Tomasek J, Yong CJ, Dumitru F, Passalacqua R, Goswami C, Safran H, Dos Santos LV, Aprile G, Ferry DR, 6 Melichar B, Tehfe M, Topuzov E, Zalcberg JR, Chau I, Campbell W, Sivanandan C, Pikiel J, Koshiji M, Hsu Y, Liepa AM, Gao L, Schwartz JD, Tabernero J; REGARD Trial Investigators. Ramucirumab monotherapy for previously treated advanced gastric or gastro-oesophageal junction adenocarcinoma (REGARD): an international, randomised, multicentre, placebo-controlled, phase 3 trial. Lancet 2014; 383: 31-39 [PMID: 24094768 DOI: 10.1016/S0140-6736(13)61719-5]
- 7 Zheng J, Xie SH, Santoni G, Lagergren J. Population-based cohort study of diabetes mellitus and mortality in gastric adenocarcinoma. Br J Surg 2018; 105: 1799-1806 [PMID: 30051463 DOI: 10.1002/bjs.10930]
- Yang HJ, Kang D, Chang Y, Ahn J, Ryu S, Cho J, Guallar E, Sohn CI. Diabetes mellitus is associated with an increased risk of gastric cancer: a cohort study. Gastric Cancer 2020; 23: 382-390 [PMID: 31853749 DOI: 10.1007/s10120-019-01033-8
- Abudawood M. Diabetes and cancer: A comprehensive review. J Res Med Sci 2019; 24: 94 [PMID: 31741666 DOI: 9 10.4103/jrms.jrms_242_19]
- 10 Wu D, Hu D, Chen H, Shi G, Fetahu IS, Wu F, Rabidou K, Fang R, Tan L, Xu S, Liu H, Argueta C, Zhang L, Mao F, Yan G, Chen J, Dong Z, Lv R, Xu Y, Wang M, Ye Y, Zhang S, Duquette D, Geng S, Yin C, Lian CG, Murphy GF, Adler GK, Garg R, Lynch L, Yang P, Li Y, Lan F, Fan J, Shi Y, Shi YG. Glucose-regulated phosphorylation of TET2 by AMPK reveals a pathway linking diabetes to cancer. Nature 2018; 559: 637-641 [PMID: 30022161 DOI: 10.1038/s41586-018-0350-5]
- 11 Li T, Chanda D, Zhang Y, Choi HS, Chiang JY. Glucose stimulates cholesterol 7alpha-hydroxylase gene transcription in human hepatocytes. J Lipid Res 2010; 51: 832-842 [PMID: 19965590 DOI: 10.1194/jlr.m002782]
- Chandra V, Huang P, Potluri N, Wu D, Kim Y, Rastinejad F. Multidomain integration in the structure of the HNF-4α 12 nuclear receptor complex. Nature 2013; 495: 394-398 [PMID: 23485969 DOI: 10.1038/nature11966]
- 13 Jiang S, Tanaka T, Iwanari H, Hotta H, Yamashita H, Kumakura J, Watanabe Y, Uchiyama Y, Aburatani H, Hamakubo T, Kodama T, Naito M. Expression and localization of P1 promoter-driven hepatocyte nuclear factor-4α (HNF4α) isoforms in human and rats. Nucl Recept 2003; 1: 5 [PMID: 12952540 DOI: 10.1186/1478-1336-1-5]
- 14 Barbetti F, D'Annunzio G. Genetic causes and treatment of neonatal diabetes and early childhood diabetes. Best Pract Res Clin Endocrinol Metab 2018; 32: 575-591 [PMID: 30086875 DOI: 10.1016/j.beem.2018.06.008]



- 15 Nakajima N, Yoshizawa A, Nakajima T, Hirata M, Furuhata A, Sumiyoshi S, Rokutan-Kurata M, Sonobe M, Menju T, Miyamoto E, Chen-Yoshikawa TF, Date H, Haga H. GATA6-positive lung adenocarcinomas are associated with invasive mucinous adenocarcinoma morphology, hepatocyte nuclear factor 4a expression, and KRAS mutations. Histopathology 2018; 73: 38-48 [PMID: 29469192 DOI: 10.1111/his.13500]
- Zhang X, Xu Y, Qian Z, Zheng W, Wu Q, Chen Y, Zhu G, Liu Y, Bian Z, Xu W, Zhang Y, Sun F, Pan Q, Wang J, Du L, 16 Yu Y. circRNA_104075 stimulates YAP-dependent tumorigenesis through the regulation of HNF4a and may serve as a diagnostic marker in hepatocellular carcinoma. Cell Death Dis 2018; 9: 1091 [PMID: 30361504 DOI: 10.1038/s41419-018-1132-6]
- 17 Orsolini L, Papanti GD, Francesconi G, Schifano F. Mind navigators of chemicals' experimenters? Cyberpsychol Behav Soc Netw 2015; 18: 296-300 [PMID: 25965863 DOI: 10.18632/oncotarget.3753]
- 18 Kojima K, Kishimoto T, Nagai Y, Tanizawa T, Nakatani Y, Miyazaki M, Ishikura H. The expression of hepatocyte nuclear factor-4alpha, a developmental regulator of visceral endoderm, correlates with the intestinal phenotype of gastric adenocarcinomas. Pathology 2006; 38: 548-554 [PMID: 17393984 DOI: 10.1080/00313020601024011]
- Koyama T, Sekine S, Taniguchi H, Tsuda H, Ikegami M, Hano H, Kushima R. Hepatocyte nuclear factor 4A expression 19 discriminates gastric involvement by metastatic breast carcinomas from primary gastric adenocarcinomas. Hum Pathol 2011; 42: 1777-1784 [PMID: 21733563 DOI: 10.1016/j.humpath.2011.04.002]
- Jucá PCFC, Corrêa S, Vignal GM, Accioly MTS, Lustosa SAS, Abdelhay E, Matos D. HNF4A expression as a potential 20 diagnostic tool to discriminate primary gastric cancer from breast cancer metastasis in a Brazilian cohort. Diagn Pathol 2017; 12: 43 [PMID: 28583188 DOI: 10.1186/s13000-017-0635-2]
- Ranc K, Jørgensen ME, Friis S, Carstensen B. Mortality after cancer among patients with diabetes mellitus: effect of 21 diabetes duration and treatment. Diabetologia 2014; 57: 927-934 [PMID: 24633676 DOI: 10.1007/s00125-014-3186-z]
- Kumar A, Ekavali, Chopra K, Mukherjee M, Pottabathini R, Dhull DK. Current knowledge and pharmacological profile of berberine: An update. Eur J Pharmacol 2015; 761: 288-297 [PMID: 26092760 DOI: 10.1016/j.ejphar.2015.05.068]
- 23 Wei S, Zhang M, Yu Y, Lan X, Yao F, Yan X, Chen L, Hatch GM. Berberine Attenuates Development of the Hepatic Gluconeogenesis and Lipid Metabolism Disorder in Type 2 Diabetic Mice and in Palmitate-Incubated HepG2 Cells through Suppression of the HNF-4α miR122 Pathway. *PLoS One* 2016; **11**: e0152097 [PMID: 27011261 DOI: 10.1371/journal.pone.0152097
- 24 Yue SJ, Liu J, Wang AT, Meng XT, Yang ZR, Peng C, Guan HS, Wang CY, Yan D. Berberine alleviates insulin resistance by reducing peripheral branched-chain amino acids. Am J Physiol Endocrinol Metab 2019; 316: E73-E85 [PMID: 30422704 DOI: 10.1152/ajpendo.00256.2018]
- Wang S, He B, Hang W, Wu N, Xia L, Wang X, Zhang Q, Zhou X, Feng Z, Chen Q, Chen J. Berberine Alleviates Tau 25 Hyperphosphorylation and Axonopathy-Associated with Diabetic Encephalopathy via Restoring PI3K/Akt/GSK3ß Pathway. J Alzheimers Dis 2018; 65: 1385-1400 [PMID: 30175975 DOI: 10.3233/JAD-180497]
- 26 Wang N, Tan HY, Li L, Yuen MF, Feng Y. Berberine and Coptidis Rhizoma as potential anticancer agents: Recent updates and future perspectives. J Ethnopharmacol 2015; 176: 35-48 [PMID: 26494507 DOI: 10.1016/j.jep.2015.10.028]
- 27 Wang J, Yang S, Cai X, Dong J, Chen Z, Wang R, Zhang S, Cao H, Lu D, Jin T, Nie Y, Hao J, Fan D. Berberine inhibits EGFR signaling and enhances the antitumor effects of EGFR inhibitors in gastric cancer. Oncotarget 2016; 7: 76076-76086 [PMID: 27738318 DOI: 10.18632/oncotarget.12589]
- Dai W, Mu L, Cui Y, Li Y, Chen P, Xie H, Wang X. Berberine Promotes Apoptosis of Colorectal Cancer via Regulation of the Long Non-Coding RNA (IncRNA) Cancer Susceptibility Candidate 2 (CASC2)/AU-Binding Factor 1 (AUF1)/B-Cell CLL/Lymphoma 2 (Bcl-2) Axis. Med Sci Monit 2019; 25: 730-738 [PMID: 30681073 DOI: 10.12659/msm.912082]
- 29 Tian Y, Zhao L, Wang Y, Zhang H, Xu D, Zhao X, Li Y, Li J. Berberine inhibits androgen synthesis by interaction with aldo-keto reductase 1C3 in 22Rv1 prostate cancer cells. Asian J Androl 2016; 18: 607-612 [PMID: 26698234 DOI: 10.4103/1008-682x.169997
- Yao M, Fan X, Yuan B, Takagi N, Liu S, Han X, Ren J, Liu J. Berberine inhibits NLRP3 Inflammasome pathway in human 30 triple-negative breast cancer MDA-MB-231 cell. BMC Complement Altern Med 2019; 19: 216 [PMID: 31412862 DOI: 10.1186/s12906-019-2615-4
- 31 Hu Q, Li L, Zou X, Xu L, Yi P. Berberine Attenuated Proliferation, Invasion and Migration by Targeting the AMPK/HNF4 α/WNT5A Pathway in Gastric Carcinoma. Front Pharmacol 2018; 9: 1150 [PMID: 30405404 DOI: 10.3389/fphar.2018.01150
- 32 Yi T, Zhuang L, Song G, Zhang B, Li G, Hu T. Akt signaling is associated with the berberine-induced apoptosis of human gastric cancer cells. Nutr Cancer 2015; 67: 523-531 [PMID: 25837881 DOI: 10.1080/01635581.2015.1004733]
- Chang HR, Nam S, Kook MC, Kim KT, Liu X, Yao H, Jung HR, Lemos R Jr, Seo HH, Park HS, Gim Y, Hong D, Huh I, 33 Kim YW, Tan D, Liu CG, Powis G, Park T, Liang H, Kim YH. HNF4α is a therapeutic target that links AMPK to WNT signalling in early-stage gastric cancer. Gut 2016; 65: 19-32 [PMID: 25410163 DOI: 10.1136/gutjnl-2014-307918]
- 34 Kikuchi A, Yamamoto H, Sato A, Matsumoto S. Wnt5a: its signalling, functions and implication in diseases. Acta Physiol (Oxf) 2012; 204: 17-33 [PMID: 21518267 DOI: 10.1111/j.1748-1716.2011.02294.x]
- 35 Xu W, Kimelman D. Mechanistic insights from structural studies of beta-catenin and its binding partners. J Cell Sci 2007; 120: 3337-3344 [PMID: 17881495 DOI: 10.1242/jcs.013771]
- 36 Kim W, Kim M, Jho EH. Wnt/β-catenin signalling: from plasma membrane to nucleus. Biochem J 2013; 450: 9-21 [PMID: 23343194 DOI: 10.1042/bj20121284]
- 37 Tillhon M, Guamán Ortiz LM, Lombardi P, Scovassi AI. Berberine: new perspectives for old remedies. Biochem Pharmacol 2012; 84: 1260-1267 [PMID: 22842630 DOI: 10.1016/j.bcp.2012.07.018]
- 38 Zhang W, Xu JH, Yu T, Chen QK. Effects of berberine and metformin on intestinal inflammation and gut microbiome composition in db/db mice. Biomed Pharmacother 2019; 118: 109131 [PMID: 31545226 DOI: 10.1016/j.biopha.2019.109131]
- Gong J, Hu M, Huang Z, Fang K, Wang D, Chen Q, Li J, Yang D, Zou X, Xu L, Wang K, Dong H, Lu F. Berberine 39 Attenuates Intestinal Mucosal Barrier Dysfunction in Type 2 Diabetic Rats. Front Pharmacol 2017; 8: 42 [PMID: 28217099 DOI: 10.3389/fphar.2017.00042]



- 40 Leng SH, Lu FE, Xu LJ. Therapeutic effects of berberine in impaired glucose tolerance rats and its influence on insulin secretion. *Acta Pharmacol Sin* 2004; 25: 496-502 [PMID: 15066220]
- 41 Yang Y, Zhang N, Li K, Chen J, Qiu L, Zhang J. Integration of microRNA-mRNA profiles and pathway analysis of plant isoquinoline alkaloid berberine in SGC-7901 gastric cancers cells. *Drug Des Devel Ther* 2018; 12: 393-408 [PMID: 29535501 DOI: 10.2147/DDDT.S155993]
- 42 Li HL, Wu H, Zhang BB, Shi HL, Wu XJ. MAPK pathways are involved in the inhibitory effect of berberine hydrochloride on gastric cancer MGC 803 cell proliferation and IL-8 secretion *in vitro* and *in vivo*. *Mol Med Rep* 2016; 14: 1430-1438 [PMID: 27278862 DOI: 10.3892/mmr.2016.5361]
- 43 **Babeu JP**, Boudreau F. Hepatocyte nuclear factor 4-alpha involvement in liver and intestinal inflammatory networks. *World J Gastroenterol* 2014; **20**: 22-30 [PMID: 24415854 DOI: 10.3748/wjg.v20.i1.22]
- 44 Huang KW, Reebye V, Czysz K, Ciriello S, Dorman S, Reccia I, Lai HS, Peng L, Kostomitsopoulos N, Nicholls J, Habib RS, Tomalia DA, Sætrom P, Wilkes E, Cutillas P, Rossi JJ, Habib NA. Liver Activation of Hepatocellular Nuclear Factor-4 α by Small Activating RNA Rescues Dyslipidemia and Improves Metabolic Profile. *Mol Ther Nucleic Acids* 2020; 19: 361-370 [PMID: 31877412 DOI: 10.1016/j.omtn.2019.10.044]
- 45 Shi J, Dai R, Chen Y, Guo H, Han Y, Zhang Y. LncRNA LINP1 regulates acute myeloid leukemia progression *via* HNF4α /AMPK/WNT5A signaling pathway. *Hematol Oncol* 2019; **37**: 474-482 [PMID: 31325181 DOI: 10.1002/hon.2651]
- 46 Lu H. Crosstalk of HNF4α with extracellular and intracellular signaling pathways in the regulation of hepatic metabolism of drugs and lipids. *Acta Pharm Sin B* 2016; **6**: 393-408 [PMID: 27709008 DOI: 10.1016/j.apsb.2016.07.003]
- 47 Yan ZQ, Leng SH, Lu FE, Lu XH, Dong H, Gao ZQ. [Effects of berberine on expression of hepatocyte nuclear factor
 4alpha and glucokinase activity in mouse primary hepatocytes]. *Zhongguo Zhong Yao Za Zhi* 2008; 33: 2105-2109 [PMID: 19160796 DOI: 10.11569/wcjd.v15.i36.3842]
- 48 Gao Y, Yan Y, Guo J, Zhang Q, Bi D, Wang F, Chang Z, Lu L, Yao X, Wei Q. HNF4α downregulation promotes tumor migration and invasion by regulating Ecadherin in renal cell carcinoma. *Oncol Rep* 2019; 42: 1066-1074 [PMID: 31322246 DOI: 10.3892/or.2019.7214]
- 49 Nam S, Chang HR, Kim KT, Kook MC, Hong D, Kwon CH, Jung HR, Park HS, Powis G, Liang H, Park T, Kim YH. PATHOME: an algorithm for accurately detecting differentially expressed subpathways. *Oncogene* 2014; 33: 4941-4951 [PMID: 24681952 DOI: 10.1038/onc.2014.80]
- 50 Endo M, Nishita M, Fujii M, Minami Y. Insight into the role of Wnt5a-induced signaling in normal and cancer cells. Int Rev Cell Mol Biol 2015; 314: 117-148 [PMID: 25619716 DOI: 10.1016/bs.ircmb.2014.10.003]
- 51 Pai SG, Carneiro BA, Mota JM, Costa R, Leite CA, Barroso-Sousa R, Kaplan JB, Chae YK, Giles FJ. Wnt/beta-catenin pathway: modulating anticancer immune response. *J Hematol Oncol* 2017; 10: 101 [PMID: 28476164 DOI: 10.1186/s13045-017-0471-6]
- 52 Kleszcz R. The canonical Wnt pathway. Postepy Biochem 2019; 65: 183-192 [PMID: 31643165 DOI: 10.18388/pb.2019_268]
- 53 Xue Y, Zhang L, Zhu Y, Ke X, Wang Q, Min H. Regulation of Proliferation and Epithelial-to-Mesenchymal Transition (EMT) of Gastric Cancer by ZEB1 via Modulating Wnt5a and Related Mechanisms. Med Sci Monit 2019; 25: 1663-1670 [PMID: 30829316 DOI: 10.12659/MSM.912338]
- 54 Zhang C, Wang T, Wu H, Zhang L, Li K, Wang F, Chen Y, Jin J, Hua D. HEF1 regulates differentiation through the Wnt5a/β-catenin signaling pathway in human gastric cancer. *Biochem Biophys Res Commun* 2019; 509: 201-208 [PMID: 30579603 DOI: 10.1016/j.bbrc.2018.12.104]
- 55 Zhao L, Liu Y, Tong D, Qin Y, Yang J, Xue M, Du N, Liu L, Guo B, Hou N, Han J, Liu S, Liu N, Zhao X, Wang L, Chen Y, Huang C. MeCP2 Promotes Gastric Cancer Progression Through Regulating FOXF1/Wnt5a/β-Catenin and MYOD1/Caspase-3 Signaling Pathways. *EBioMedicine* 2017; 16: 87-100 [PMID: 28131747 DOI: 10.1016/j.ebiom.2017.01.021]
- 56 Shojima K, Sato A, Hanaki H, Tsujimoto I, Nakamura M, Hattori K, Sato Y, Dohi K, Hirata M, Yamamoto H, Kikuchi A. Wnt5a promotes cancer cell invasion and proliferation by receptor-mediated endocytosis-dependent and -independent mechanisms, respectively. *Sci Rep* 2015; 5: 8042 [PMID: 25622531 DOI: 10.1038/srep08042]
- 57 Ara H, Takagishi M, Enomoto A, Asai M, Ushida K, Asai N, Shimoyama Y, Kaibuchi K, Kodera Y, Takahashi M. Role for Daple in non-canonical Wnt signaling during gastric cancer invasion and metastasis. *Cancer Sci* 2016; 107: 133-139 [PMID: 26577606 DOI: 10.1111/cas.12848]
- 58 Zhang Y, Du J, Zheng J, Liu J, Xu R, Shen T, Zhu Y, Chang J, Wang H, Zhang Z, Meng F, Wang Y, Chen Y, Xu Y, Gu L. EGF-reduced Wnt5a transcription induces epithelial-mesenchymal transition *via* Arf6-ERK signaling in gastric cancer cells. Oncotarget 2015; 6: 7244-7261 [PMID: 25779663 DOI: 10.18632/oncotarget.3133]
- 59 Geng B, Pan J, Zhao T, Ji J, Zhang C, Che Y, Yang J, Shi H, Li J, Zhou H, Mu X, Xu C, Wang C, Xu Y, Liu Z, Wen H, You Q. Chitinase 3-like 1-CD44 interaction promotes metastasis and epithelial-to-mesenchymal transition through βcatenin/Erk/Akt signaling in gastric cancer. *J Exp Clin Cancer Res* 2018; **37**: 208 [PMID: 30165890 DOI: 10.1186/s13046-018-0876-2]
- 60 Liu Z, Chen Z, Fan R, Jiang B, Chen X, Chen Q, Nie F, Lu K, Sun M. Over-expressed long noncoding RNA HOXA11-AS promotes cell cycle progression and metastasis in gastric cancer. *Mol Cancer* 2017; 16: 82 [PMID: 28441948 DOI: 10.1186/s12943-017-0651-6]
- 61 Dai B, Ma Y, Yang T, Fan M, Yu R, Su Q, Wang H, Liu F, Yang C, Zhang Y. Synergistic effect of berberine and HMQ1611 impairs cell proliferation and migration by regulating Wnt signaling pathway in hepatocellular carcinoma. *Phytother Res* 2019; 33: 745-755 [PMID: 30565332 DOI: 10.1002/ptr.6267]

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