World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2024 April 15; 16(4): 1091-1675





Published by Baishideng Publishing Group Inc

World Journal of Gastrointestinal Oncology

Contents

Monthly Volume 16 Number 4 April 15, 2024

EDITORIAL

1091	Parallel pathways: A chronicle of evolution in rectal and breast cancer surgery
1097	Hepatitis B virus genotypes in precision medicine of hepatitis B-related hepatocellular carcinoma: Where we are now Sukowati CHC, Jayanti S, Turyadi T, Muljono DH, Tiribelli C
	REVIEW
1104	Novel milestones for early esophageal carcinoma: From bench to bed
	Qi JH, Huang SL, Jin SZ
1119	Colorectal cancer screening: A review of current knowledge and progress in research
	Lopes SR, Martins C, Santos IC, Teixeira M, Gamito É, Alves AL
1134	New avenues for the treatment of immunotherapy-resistant pancreatic cancer
	Silva LGO, Lemos FFB, Luz MS, Rocha Pinheiro SL, Calmon MDS, Correa Santos GL, Rocha GR, de Melo FF
	MINIREVIEWS
1154	Present situation of minimally invasive surgical treatment for early gastric cancer
	Li CY, Wang YF, Luo LK, Yang XJ
1166	Mixed neuroendocrine non-neuroendocrine neoplasms in gastroenteropancreatic tract
	Díaz-López S, Jiménez-Castro J, Robles-Barraza CE, Ayala-de Miguel C, Chaves-Conde M
1180	Esophageal cancer screening, early detection and treatment: Current insights and future directions
1100	Qu HT, Li Q, Hao L, Ni YJ, Luan WY, Yang Z, Chen XD, Zhang TT, Miao YD, Zhang F
	ORIGINAL ARTICLE
	Retrospective Cohort Study
1192	Pre-operative enhanced magnetic resonance imaging combined with clinical features predict early
	Chen IP, Yang RH, Zhang TH, Liao I, A, Guan YT, Dai HY
	Cren 91, Tung MI, Enung 111, Eluo EA, Ouun 11, Dui 111
1204	Clinical analysis of multiple primary gastrointestinal malignant tumors: A 10-year case review of a single-

Zhu CL, Peng LZ

center



Contents

World Journal of Gastrointestinal Oncology

Monthly Volume 16 Number 4 April 15, 2024

Retrospective Study

1213 Predictive model for non-malignant portal vein thrombosis associated with cirrhosis based on inflammatory biomarkers

Nie GL, Yan J, Li Y, Zhang HL, Xie DN, Zhu XW, Li X

1227 Predictive modeling for postoperative delirium in elderly patients with abdominal malignancies using synthetic minority oversampling technique

Hu WJ, Bai G, Wang Y, Hong DM, Jiang JH, Li JX, Hua Y, Wang XY, Chen Y

Efficacy and predictive factors of transarterial chemoembolization combined with lenvatinib plus 1236 programmed cell death protein-1 inhibition for unresectable hepatocellular carcinoma

Ma KP, Fu JX, Duan F, Wang MQ

1248 Should we perform sigmoidoscopy for colorectal cancer screening in people under 45 years? Leong W, Guo JQ, Ning C, Luo FF, Jiao R, Yang DY

1256 Computed tomography-based radiomics diagnostic approach for differential diagnosis between early- and late-stage pancreatic ductal adenocarcinoma

Ren S, Qian LC, Cao YY, Daniels MJ, Song LN, Tian Y, Wang ZQ

1268 Prognostic analysis of related factors of adverse reactions to immunotherapy in advanced gastric cancer and establishment of a nomogram model

He XX, Du B, Wu T, Shen H

Clinical Trials Study

1281 Safety and efficacy of a programmed cell death 1 inhibitor combined with oxaliplatin plus S-1 in patients with Borrmann large type III and IV gastric cancers

Bao ZH, Hu C, Zhang YQ, Yu PC, Wang Y, Xu ZY, Fu HY, Cheng XD

Observational Study

1296 Computed tomography radiogenomics: A potential tool for prediction of molecular subtypes in gastric stromal tumor

Yin XN, Wang ZH, Zou L, Yang CW, Shen CY, Liu BK, Yin Y, Liu XJ, Zhang B

1309 Application of texture signatures based on multiparameter-magnetic resonance imaging for predicting microvascular invasion in hepatocellular carcinoma: Retrospective study

Nong HY, Cen YY, Qin M, Qin WQ, Xie YX, Li L, Liu MR, Ding K

- 1319 Causal roles of gut microbiota in cholangiocarcinoma etiology suggested by genetic study Chen ZT, Ding CC, Chen KL, Gu YJ, Lu CC, Li QY
- 1334 Is recovery enhancement after gastric cancer surgery really a safe approach for elderly patients? Li ZW, Luo XJ, Liu F, Liu XR, Shu XP, Tong Y, Lv Q, Liu XY, Zhang W, Peng D
- 1344 Establishment of a cholangiocarcinoma risk evaluation model based on mucin expression levels Yang CY, Guo LM, Li Y, Wang GX, Tang XW, Zhang QL, Zhang LF, Luo JY



Contor	World Journal of Gastrointestinal Oncology
Conten	Monthly Volume 16 Number 4 April 15, 2024
1361	Effectiveness of fecal DNA syndecan-2 methylation testing for detection of colorectal cancer in a high-risk Chinese population
	Luo WF, Jiao YT, Lin XL, Zhao Y, Wang SB, Shen J, Deng J, Ye YF, Han ZP, Xie FM, He JH, Wan Y
	Clinical and Translational Research
1374	Clinical and socioeconomic determinants of survival in biliary tract adenocarcinomas
	Sahyoun L, Chen K, Tsay C, Chen G, Protiva P
1384	Risk factors, prognostic factors, and nomograms for distant metastasis in patients with diagnosed duodenal cancer: A population-based study
	Shang JR, Xu CY, Zhai XX, Xu Z, Qian J
1421	NOX4 promotes tumor progression through the MAPK-MEK1/2-ERK1/2 axis in colorectal cancer
	Xu YJ, Huo YC, Zhao QT, Liu JY, Tian YJ, Yang LL, Zhang Y
	Basic Study
1437	Curcumin inhibits the growth and invasion of gastric cancer by regulating long noncoding RNA AC022424.2
	Wang BS, Zhang CL, Cui X, Li Q, Yang L, He ZY, Yang Z, Zeng MM, Cao N
1453	MicroRNA-298 determines the radio-resistance of colorectal cancer cells by directly targeting human dual- specificity tyrosine(Y)-regulated kinase 1A
	Shen MZ, Zhang Y, Wu F, Shen MZ, Liang JL, Zhang XL, Liu XJ, Li XS, Wang RS
1465	Human β -defensin-1 affects the mammalian target of rapamycin pathway and autophagy in colon cancer cells through long non-coding RNA TCONS_00014506
	Zhao YX, Cui Y, Li XH, Yang WH, An SX, Cui JX, Zhang MY, Lu JK, Zhang X, Wang XM, Bao LL, Zhao PW
1479	FAM53B promotes pancreatic ductal adenocarcinoma metastasis by regulating macrophage M2 polarization
	Pei XZ, Cai M, Jiang DW, Chen SH, Wang QQ, Lu HM, Lu YF
1500	Transcriptome sequencing reveals novel biomarkers and immune cell infiltration in esophageal tumori- genesis
	Sun JR, Chen DM, Huang R, Wang RT, Jia LQ
1514	Construction of CDKN2A-related competitive endogenous RNA network and identification of GAS5 as a prognostic indicator for hepatocellular carcinoma
	Pan Y, Zhang YR, Wang LY, Wu LN, Ma YQ, Fang Z, Li SB
1532	Two missense <i>STK11</i> gene variations impaired LKB1/adenosine monophosphate-activated protein kinase signaling in Peutz-Jeghers syndrome
	Liu J, Zeng SC, Wang A, Cheng HY, Zhang QJ, Lu GX
1547	Long noncoding RNAs HAND2-AS1 ultrasound microbubbles suppress hepatocellular carcinoma progression by regulating the miR-873-5p/tissue inhibitor of matrix metalloproteinase-2 axis
	Zou Q, Wang HW, Di XL, Li Y, Gao H

Conton	World Journal of Gastrointestinal Oncology
Conten	Monthly Volume 16 Number 4 April 15, 2024
1564	Upregulated lncRNA PRNT promotes progression and oxaliplatin resistance of colorectal cancer cells by regulating HIPK2 transcription
	Li SN, Yang S, Wang HQ, Hui TL, Cheng M, Zhang X, Li BK, Wang GY
	SYSTEMATIC REVIEWS
1578	Prognosis value of heat-shock proteins in esophageal and esophagogastric cancer: A systematic review and meta-analysis
	Nakamura ET, Park A, Pereira MA, Kikawa D, Tustumi F
1596	Risk factors for hepatocellular carcinoma associated with hepatitis C genotype 3 infection: A systematic review
	Farooq HZ, James M, Abbott J, Oyibo P, Divall P, Choudhry N, Foster GR
	META-ANALYSIS
1613	Effectiveness and tolerability of programmed cell death protein-1 inhibitor + chemotherapy compared to chemotherapy for upper gastrointestinal tract cancers
	Zhang XM, Yang T, Xu YY, Li BZ, Shen W, Hu WQ, Yan CW, Zong L
1626	Success rate of current human-derived gastric cancer organoids establishment and influencing factors: A systematic review and meta-analysis
	Jiang KL, Wang XX, Liu XJ, Guo LK, Chen YQ, Jia QL, Yang KM, Ling JH
	CASE REPORT
1647	Pathologically successful conversion hepatectomy for advanced giant hepatocellular carcinoma after multidisciplinary therapy: A case report and review of literature
	Chu JH, Huang LY, Wang YR, Li J, Han SL, Xi H, Gao WX, Cui YY, Qian MP
1660	Clinical pathological characteristics of "crawling-type" gastric adenocarcinoma cancer: A case report
	Xu YW, Song Y, Tian J, Zhang BC, Yang YS, Wang J
1668	Primary pancreatic peripheral T-cell lymphoma: A case report
	Bai YL, Wang LJ, Luo H, Cui YB, Xu JH, Nan HJ, Yang PY, Niu JW, Shi MY



Contents

World Journal of Gastrointestinal Oncology

Monthly Volume 16 Number 4 April 15, 2024

ABOUT COVER

Peer Reviewer of World Journal of Gastrointestinal Oncology, Lie Zheng, Director, Professor, Department of Gastroenterology, Shaanxi Provincial Hospital of Traditional Chinese Medicine, Xi'an 730000, Shaanxi Province, China. xinliwen696@126.com

AIMS AND SCOPE

The primary aim of World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

INDEXING/ABSTRACTING

The WJGO is now abstracted and indexed in PubMed, PubMed Central, Science Citation Index Expanded (SCIE, also known as SciSearch®), Journal Citation Reports/Science Edition, Scopus, Reference Citation Analysis, China Science and Technology Journal Database, and Superstar Journals Database. The 2023 edition of Journal Citation Reports® cites the 2022 impact factor (IF) for WJGO as 3.0; IF without journal self cites: 2.9; 5-vear IF: 3.0; Journal Citation Indicator: 0.49; Ranking: 157 among 241 journals in oncology; Quartile category: Q3; Ranking: 58 among 93 journals in gastroenterology and hepatology; and Quartile category: Q3. The WJGO's CiteScore for 2022 is 4.1 and Scopus CiteScore rank 2022: Gastroenterology is 71/149; Oncology is 197/366.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Xiang-Di Zhang; Production Department Director: Xiang Li; Cover Editor: Jia-Ru Fan.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Gastrointestinal Oncology	https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1948-5204 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
February 15, 2009	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Monthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Monjur Ahmed, Florin Burada	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/1948-5204/editorialboard.htm	https://www.wignet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
April 15, 2024	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2024 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2024 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: office@baishideng.com https://www.wjgnet.com



COWÛ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2024 April 15; 16(4): 1578-1595

DOI: 10.4251/wjgo.v16.i4.1578

ISSN 1948-5204 (online)

SYSTEMATIC REVIEWS

Prognosis value of heat-shock proteins in esophageal and esophagogastric cancer: A systematic review and meta-analysis

Eric Toshiyuki Nakamura, Amanda Park, Marina Alessandra Pereira, Daniel Kikawa, Francisco Tustumi

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): D Grade E (Poor): 0

P-Reviewer: Wang SY, China; Zhang JW, China

Received: October 8, 2023 Peer-review started: October 8, 2023

First decision: December 15, 2023 Revised: December 24, 2023 Accepted: January 23, 2024 Article in press: January 23, 2024 Published online: April 15, 2024



Eric Toshiyuki Nakamura, Marina Alessandra Pereira, Francisco Tustumi, Department of Gastroenterology, Instituto do Câncer, Hospital das Clínicas da Universidade de São Paulo, Faculdade de Medicina, Universidade de São Paulo, São Paulo 01246000, Brazil

Eric Toshiyuki Nakamura, Daniel Kikawa, Department of Scientific Initiation, Universidade Mogi das Cruzes, São Paulo 08780911, Brazil

Amanda Park, Department of Evidence-Based Medicine, Centro Universitário Lusíada, Centre for Evidence-Based Medicine, Centro Universitário Lusíada (UNILUS), Santos, Brazil

Francisco Tustumi, Department of Surgery, Hospital Israelita Albert Einstein, São Paulo 05652900, Brazil

Corresponding author: Francisco Tustumi, MD, PhD, Surgeon, Surgical Oncologist, Department of Surgery, Hospital Israelita Albert Einstein, Avenue Albert Einstein, 627/701 - Morumbi, São Paulo 05652900, Brazil. franciscotustumi@gmail.com

Abstract

BACKGROUND

Heat shock proteins (HSPs) are molecular chaperones that play an important role in cellular protection against stress events and have been reported to be overexpressed in many cancers. The prognostic significance of HSPs and their regulatory factors, such as heat shock factor 1 (HSF1) and CHIP, are poorly understood.

AIM

To investigate the relationship between HSP expression and prognosis in esophageal and esophagogastric cancer.

METHODS

A systematic review was conducted in accordance with PRISMA recommendations (PROSPERO: CRD42022370653), on Embase, PubMed, Cochrane, and LILACS. Cohort, case-control, and cross-sectional studies of patients with esophagus or esophagogastric cancer were included. HSP-positive patients were compared with HSP-negative, and the endpoints analyzed were lymph node metastasis, tumor depth, distant metastasis, and overall survival (OS). HSPs were stratified according to the HSP family, and the summary risk difference (RD) was calculated using a random-effect model.

RESULTS



The final selection comprised 27 studies, including esophageal squamous cell carcinoma (21), esophagogastric adenocarcinoma (5), and mixed neoplasms (1). The pooled sample size was 3465 patients. HSP40 and 60 were associated with a higher 3-year OS [HSP40: RD = 0.22; 95% confidence interval (CI): 0.09-0.35; HSP60: RD = 0.33; 95%CI: 0.17-0.50], while HSF1 was associated with a poor 3-year OS (RD = -0.22; 95% CI: -0.32 to -0.12). The other HSP families were not associated with long-term survival. HSF1 was associated with a higher probability of lymph node metastasis (RD = -0.16; 95%CI: -0.29 to -0.04). HSP40 was associated with a lower probability of lymph node dissemination (RD = 0.18; 95%CI: 0.03-0.33). The expression of other HSP families was not significantly related to tumor depth and lymph node or distant metastasis.

CONCLUSION

The expression levels of certain families of HSP, such as HSP40 and 60 and HSF1, are associated with long-term survival and lymph node dissemination in patients with esophageal and esophagogastric cancer.

Key Words: Heat-shock proteins; Heat-shock response; Prognosis; Esophageal neoplasms; Meta-analysis

©The Author(s) 2024. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Heat shock proteins (HSPs) and their regulatory factors, such as heat shock factor 1 (HSF1) and CHIP, play an important role in cellular protection against stress events, and are overexpressed in some types or cancer. However, the prognostic significance of HSPs remains unclear. In the present study, we conducted a systematic review and meta-analysis to investigate the relationship between HSP expression and prognosis in esophageal and esophagogastric cancer that included 27 studies. Our findings demonstrated that the expression levels of some families of HSP, such as HSP40 and 60 and HSF1, are associated with long-term survival and lymph node dissemination in esophageal cancer.

Citation: Nakamura ET, Park A, Pereira MA, Kikawa D, Tustumi F. Prognosis value of heat-shock proteins in esophageal and esophagogastric cancer: A systematic review and meta-analysis. *World J Gastrointest Oncol* 2024; 16(4): 1578-1595 URL: https://www.wjgnet.com/1948-5204/full/v16/i4/1578.htm DOI: https://dx.doi.org/10.4251/wjgo.v16.i4.1578

INTRODUCTION

Esophageal cancer (EC) is the sixth most common cancer worldwide, with 600000 new cases reported annually, and is the eighth deadliest cancer[1]. Despite strides in therapeutics and screening methods, the prognosis remains grim, with a 5-year survival rate of approximately 15%-25%[2]. Conventional treatments such as chemotherapy, radiation, and surgery often fall short in halting disease progression and recurrence, leaving patients with limited options[3].

Regrettably, practical prognostic factors for EC, crucial for precise diagnosis and therapy, are underexplored[4]. This underscores the urgent need to develop novel prognostic markers for EC, enabling enhanced risk stratification and targeted therapy. While significant headway has been made in diagnosing EC, the pursuit of reliable prognostic indicators persists.

Numerous studies in cancer biology have unveiled a plethora of potential targets, with heat shock proteins (HSPs) emerging as crucial players in protein folding and apoptotic modulation, reported to be overexpressed in esophageal tumors[5]. Initially discovered as proteins robustly induced in response to heat shock and various stressors, HSPs are highly conserved in all mammalian cells[5]. They contribute to protein quality control by ensuring the accurate folding of newly synthesized proteins and the refolding of denatured proteins under various intracellular and extracellular stressor conditions.

HSPs, categorized into six families based on their relative molecular sizes (HSP 20, HSP 40, HSP 60, HSP 70, and HSP 90)[5,6], are regulated by a complex interplay of factors. The swift induction of HSP expression in response to stressors constitutes the heat shock response (HSR)[7], regulated at the transcriptional level by heat shock factors (HSFs), crucial upstream transcriptional regulators of HSPs[8]. HSF1 is recognized as the primary regulator of the HSR, activated by the denaturation of intracellular proteins due to proteotoxic exposures. States of hypoxia, acidosis, and inflammation, for instance, may trigger proteotoxic effects and HSF1 activation.

Several clinical conditions linked to different families of HSP, such as acute and chronic renal diseases[9], psoriasis[10], and neurodegenerative diseases[11], disrupt normal cell functions. Abnormal expression levels of certain HSPs have been revealed in various cancer types, including prostate, bladder, breast, ovarian, colorectal, and lung cancers[12-14]. Some HSP families exhibit a significant correlation with the prognosis of different cancer types[15,16].

The varying expression levels of HSPs during malignant transformation prompt the question of whether HSPs can serve as prognostic indicators in routine clinical settings for EC[17,18]. The objective of our study was to assess the prognostic value of HSPs and their regulatory factors in the context of esophageal and esophagogastric cancer through a comprehensive meta-analysis.

Gaishideng® WJGO | https://www.wjgnet.com

MATERIALS AND METHODS

The systematic review and meta-analysis adhered to the PRISMA guidelines^[19]. The research protocol underwent registration on PROSPERO^[20] (International Prospective Register of Systematic Reviews) with the registration number CRD42022370653.

Eligibility criteria

This meta-analysis considered studies evaluating HSP in patients with esophageal squamous cell carcinoma (ESCC) or esophagogastric adenocarcinoma. Inclusion criteria comprised cohort, case-control, and cross-sectional studies, while case reports, editorials, abstracts, and those without full-text availability were excluded.

Information sources and search strategy

The search spanned PubMed, Embase, LILACS (BVS), Cochrane Library Central, and references from included articles, previous systematic reviews, and meta-analyses. A combination of MeSH terms and keywords, such as "Heat Shock Proteins", "HSP", "Heat Shock", "Esophagus", "Esophageal", and others, formed the basis of the search. The period covered was from the inception of the databases to March 2023.

Study selection

Conducted by two independent authors (Nakamura ET and Park A), the systematic literature search followed predetermined eligibility criteria. Discrepancies in study inclusion were resolved by a third reviewer. No language or publication date restrictions were imposed, and selection filters were not applied.

Data extraction

Reviewers (Nakamura ET and Park A) manually retrieved baseline characteristics and outcomes, including author name, publication year, study design, sample size, histologic cancer type, treatment type, HSP family, age, sex, stage, and follow-up.

Assessment of risk of bias

The Newcastle-Ottawa Scale (NOS) facilitated bias risk assessment, executed by two authors (Nakamura ET and Kikawa D), with adjudication by a third author (Tustumi F) in the case of disagreements.

Outcomes

The analysis encompassed postoperative mortality, postoperative complications, grade of cellular differentiation, lymph node dissemination, tumor depth, metastasis, and survival.

Statistical analysis and data synthesis

Meta-analysis was performed using STATA 16.1 software (StataCorp LLC). Categorical variables were reported as risk differences (RD), and continuous variables as mean differences (MD), both with a 95% confidence interval (CI). The I² test assessed statistical heterogeneity, and a random-effect model was employed to balance statistical and clinical heterogeneity. Forest plots were utilized for the meta-analysis compilation. Subgroup analysis was used to control histological cancer type as potential confounding variables. We assessed the subgroup of cohorts that evaluated exclusively ESCC.

RESULTS

Characteristics of the included studies

As illustrated in Figure 1, the search flow diagram delineates the identification process, initially yielding 266 primary studies. Following the removal of duplicates and exclusion of articles irrelevant to the meta-analysis, 110 candidate studies underwent full-text review. Subsequently, 27 studies aligning with the inclusion criteria were deemed suitable for the study's objectives [21-47]. Within this subset, 21 studies focused on ESCC, five on esophagogastric adenocarcinoma, and one on a mixed neoplasm. The cumulative sample size encompassed 3465 patients, with an average age of 60.7 years (range: 46-69) and a predominant male representation (76.9%; range: 38%-91%). A comprehensive summary of the included studies is presented in Table 1.

Overall survival

Elevated expression levels of HSP40 (RD = 0.22; 95%CI: 0.09-0.35) and HSP60 (RD = 0.33; 95%CI: 0.17-0.50) were associated with a heightened 3-year overall survival (OS). Conversely, the presence of HSF1 was linked to a poorer 3-year OS (RD = -0.22; 95% CI: -0.32 to -0.12) (Figure 2). Similar findings were observed for ESCC cohorts (Table 2).

Grade of cellular differentiation

No significant correlation was found between the expression of CHIP (RD = -0.03; 95%CI: -0.15 to 0.09), HSF1 (RD = 0.04; 95% CI: -0.09 to 0.17), HSP20 (RD = -0.09; 95% CI: -0.34 to 0.16), HSP40 (RD = 0.03; 95% CI: -0.10 to 0.16), HSP60 (RD = -0.21; 95% CI: -0.57 to 0.15), HSP70 (RD = -0.14; 95% CI: -0.33 to 0.04), and HSP90 (RD = -0.08; 95% CI: -0.18 to 0.02) with grade of



Table 1 Characteristics of included studies										
Ref.	Cancer type	N	HSP	Age (yr)	Male (%)	Upper third cancer location (%)	Stage III/IV (%)	Follow-up (months)		
Akutsu <i>et al</i> [21], 2011	ESCC	78	HSP90	62	89	19	51	24		
Berezowska <i>et al</i> [22], 2013	EA and G	347	HSP90	69	64	0	NI	NI		
Berg et al[23], 2011	EA	87	HSP20	63	91	0	NI	NI		
Doak <i>et al</i> [24], 2004	EA	4	HSP20	63	83	0	NI	NI		
Faried <i>et al</i> [25], 2004	ESCC	123	HSP60, 90	61	86	14	38	NI		
Huang et al[26], 2014	ESCC	81	HSP90	58	38	0	30	NI		
Iqbal <i>et al</i> [27], 2016	ESCC	46	HSP20, 70, 90	58	65	0	16	NI		
Kawanishi <i>et al</i> [<mark>28</mark>], 1999	ESCC	102	HSP20, 70	62	82	NI	36	25		
Liao <i>et al</i> [<mark>29</mark>], 2015	ESCC	134	HSF1	61	81	NI	46	NI		
Luz et al[30], 2017	ESCC	28	HSP20, 70	60	82	NI	NI	60		
Lv et al[<mark>31</mark>], 2022	ESCC and EA	87	HSP60	NI	NI	NI	52	NI		
Miyazaki <i>et al</i> [<mark>32</mark>], 2005	ESCC	61	HSP20, 70	65	87	21	49	23		
Nakajima <i>et al</i> [<mark>33</mark>], 2002	ESCC	62	HSP20, 70	61	85	13	42	NI		
Nakajima <i>et al</i> [<mark>34</mark>], 2009	EC	125	HSP70	62	86	14	38	NI		
Noguchi <i>et al</i> [35], 2002	ESCC	71	HSP70	64	89	11	45			
Ou et al[36], 2014	ESCC	328	HSP90	59	82	NI	NI	51		
Söderström <i>et al</i> [37], 2019	EA	151	HSP20, 70	65	80	0	83	NI		
Tsukao et al <mark>[38</mark>], 2017	ESCC	212	HSF1, HSP20, 70, 90	65	87	14	54	NI		
Wang et al[39], 2007	G	60	HSP70, HSP90	46	53	0	NI	NI		
Wang et al[40], 2010	ESCC	120	HSP70, 90	57	77	NI	NI	NI		
Wen et al[41], 2013	ESCC	234	CHIP	58	83	10	NI	18		
Xue <i>et al</i> [42], 2014	ESCC	112	HSP20	60	54	NI	NI	NI		
Yu et al[43], 2015	ESCC	72	HSP40	65	82	NI	66	NI		
Zhang et al[44], 2013	ESCC	120	HSP70	53	75	29	36	60		
Zhang et al[45], 2017	ESCC	162	HSP20	63	67	NI	NI	NI		
Zhang et al[46], 2020	ESCC	345	HSP20	NI	69	NI	NI	NI		
Zhao <i>et al</i> [47], 2015	ESCC	113	HSP70	58	82	20	74	NI		

HSP: Heat shock protein; ESCC: Esophageal squamous cell carcinoma; G: Gastric; EC: Esophageal cancer; EA: Esophageal adenocarcinoma; NI: Not informed; HSF: Heat shock factor.

cellular differentiation (Figure 3A).

Lymph node dissemination

The results suggested that overexpression of HSF1 (RD = -0.16; 95%CI: -0.29 to -0.04) was significantly associated with positive lymph node metastasis. High HSP40 values were associated with less risk for lymph node dissemination (RD = 0.18; 95%CI: 0.03-0.33). There was no significant difference observed for CHIP (RD = 0.00; 95%CI: 0.00-0.00), HSP20 (RD = 0.05; 95%CI: -0.15-0.24), HSP60 (RD = -0.14; 95%CI: -0.36 to 0.08), HSP70 (RD = -0.20; 95%CI: -0.48 to 0.07), and HSP90 (RD = 0.26; 95%CI: -0.62 to 0.10) (Figure 3B). Similar findings were observed for ESCC cohorts (Table 2).

Saishideng® WJGO https://www.wjgnet.com

Table 2 Subgroup analysis, the impact of heat shock protein on 3-year overall survival, grade of cellular differentiation, and tumornode-metastasis stage in esophageal squamous cell carcinoma

HSP		Overall survivall	Grade of cellular differentiation	т	N	Μ
CHIP	RD (95%CI)	0.03 (-0.09 to 0.15)	-	-	0.00 (0.00-0.00)	-
	Studies	1	-	-	1	-
HSF1	RD (95%CI)	-0.22 (-0.32 to -0.12) ¹	0.04 (-0.09 to 0.17)	-0.22 (-0.51 to 0.06)	-0.16 (-0.29 to -0.04) ¹	0.01 (-0.06 to 0.08)
	Studies	2	2	1	2	1
HSP20	RD (95%CI)	0.16 (-0.12 to 0.45)	0.03 (-0.05 to 0.11)	0.01 (-0.13 to 0.16)	-0.04 (-0.22 to 0.14)	0.00 (0.15 to -0.15)
	Studies	6	7	6	7	4
HSP40	RD (95%CI)	0.22 (0.09-0.35) ¹	0.03 (-0.10 to 0.16)	-0.04 (-0.14 to 0.06)	0.18 (0.03-0.33) ¹	0.03 (-0.05 to 0.11)
	Studies	1	1	1	1	1
HSP60	RD (95%CI)	0.33 (0.17-0.50) ¹	-0.03 (-0.20 to 0.13)	0.09 (-0.09 to 0.26)	-0.04 (-0.21 to 0.14)	0.07 (-0.06 to 0.20)
	Studies	1	1	1	1	1
HSP70	RD (95%CI)	0.07 (-0.18 to 0.31)	-0.08 (-0.17 to 0.00)	-0.07 (-0.25 to 0.11)	-0.17 (-0.45 to 0.12)	-0.10 (-0.33 to 0.12)
	Studies	8	8	6	8	5
HSP90	RD (95%CI)	0.03 (-0.23 to 0.29)	-0.02 (-0.06 to 0.03)	-0.10 (-0.20 to 0.01)	-0.24 (-0.74 to 0.26)	0.18 (-0.18 to 0.54)
	Studies	3	2	4	3	3

¹Significant values.

RD: Risk difference; CI: Confidence interval; HSP: Heat shock protein; HSF: Heat shock factor.

Tumor depth

There was no significant association between tumor depth and HSF1 (RD = -0.22; 95%CI: -0.51 to 0.06), HSP20 (RD = -0.03; 95% CI: -0.01 to 0.16), HSP40 (RD = -0.04; 95% CI: -0.14 to 0.06), HSP60 (RD = 0.01; 95% CI: -0.17 to 0.18), HSP70 (RD = 0.00; 95% CI: -0.21 to 0.22), and HSP90 (RD = -0.04; 95% CI: -0.19 to 0.12) (Figure 4A).

Distant metastasis

There was no significant association between distant metastasis and HSF1 (RD = -0.01; 95%CI: -0.06 to 0.08), HSP20 (RD = 0.04; 95% CI: -0.09 to 0.17), HSP40 (RD = 0.03; 95% CI: -0.05 to 0.11), HSP60 (RD = 0.00; 95% CI: -0.12 to 0.13), HSP70 (RD = -0.14; 95% CI: -0.38 to 0.09), and HSP90 (RD = -0.22; 95% CI: -0.55 to 0.10) (Figure 4B).

Risk of bias assessment

All eligible studies underwent a risk of bias assessment with the NOS. Two independent reviewers assessed the quality and risk of bias. In the event of a tie, the decision was determined by a third reviewer after a group discussion in which all sides were taken into account. Points given to each study are shown in Table 3.

DISCUSSION

The association between HSPs and cancer prognosis has generated significant interest, offering potential implications for clinical decision-making in cancer management. Prognostication, a pivotal aspect of cancer care, can be significantly enhanced through the exploration of HSPs in EC. This investigation holds promise for refining prognostic predictions, tailoring treatment approaches, and ultimately improving patient outcomes[48].

The tumor microenvironment, characterized by conditions such as low glucose, pH, and oxygen levels, induces the expression of HSPs[15]. These molecular chaperones, crucial in apoptosis regulation[49], respond to stressors during carcinogenesis, triggered by the emergence of oncoproteins. However, the diverse functions of HSPs within tumors are influenced by the complex genetic and epigenetic alterations characterizing carcinogenesis[50]. HSPs may play a protective role in the early stages of cancer initiation, such as in chronic esophagitis, yet exhibit different patterns as cancer progresses^[5]. For instance, HSP70 exhibits differential expression following thermal injury to the esophageal epithelium, with reduced levels post-injury and subsequent recovery-induced upregulation[51]. This finding highlights the distinct roles of certain HSPs in the context of esophageal injury and recovery in gastroesophageal reflux, known risk factors for Barrett's esophagus and esophagogastric adenocarcinoma^[52]. This nuanced understanding of HSP behavior contributes to the heterogeneous differentiation [53,54] observed within HSPs, with some members associated with aggressive cancer phenotypes and others playing a protective role in cancer development[55].

Table 3 Risk of bias assessment scores based on the Newcastle-Ottawa Scale of studies

	Selection of cohorts				Comparability of cohorts	Outcome		
Ref.	Representativeness of the exposed cohort	Selection of the non- exposed cohort	Ascertainment of exposure	Demonstration that outcome of interest was not present at start of study	Comparability of cohorts on the basis of the design or analysis	Assessment of outcome	Was follow up long enough for outcomes to occur	Adequacy of follow up of cohorts
Akutsu <i>et al</i> [21], 2011	\$	☆	\$	*	**	\$	☆	
Berezowska <i>et al</i> [22], 2013	й	*	\$	ж	**	\$	\$	
Berg <i>et al</i> [23], 2011	Å.	☆	*	*	**	\$	☆	
Doak <i>et al</i> [<mark>24</mark>], 2004	\$	\$	*	\$	☆☆	☆	☆	
Faried <i>et al</i> [25], 2004	\$	☆	\$	*	\$	\$		
Huang <i>et al</i> [<mark>26], 2014</mark>	Å	\$	*	\$	**	\$	☆	\$
Iqbal <i>et al</i> [<mark>27</mark>], 2016	\$	\$	\$	Å	**	첪		
Kawanishi <i>et al</i> [28], 1999	☆	\$	\$	\$	**	\$	\$	\$
Liao <i>et al</i> [<mark>29]</mark> , 2015	\$	☆	\$	Å	**	\$	☆	☆
Luz et al [<mark>30</mark>], 2017	\$	\$	\$	Å	**	첪		
Lv et al <mark>[31</mark>], 2022	*	\$	\$	Å	**	\$	☆	☆
Miyazaki et al <mark>[32</mark>], 2005	Å	☆	☆	*	**	\$	\$	\$
Nakajima et al <mark>[33</mark>], 2002	Å	☆	☆	*	**	\$	\$	\$
Nakajima et al[<mark>34</mark>], 2009	Å	\$	*	\$	**	\$	☆	\$
Noguchi <i>et</i> al[<mark>35</mark>], 2002	*	\$	\$	\$	**	\$	☆	☆
Ou et al [<mark>36</mark>], 2014	*	\$	\$	\$	**	\$	☆	☆
Söderström <i>et al</i> [37], 2019	☆	*	\$	\$	\$z \$z	\$	\$	☆
Tsukao <i>et al</i> [<mark>38</mark>], 2017	\$	☆	*	Å	**	\$	云	☆
Wang et al [<mark>39</mark>], 2007	Å	☆	*	\$	**	☆		
Wang <i>et al</i> [40], 2010	\$	☆		\$	**	*		
Wen <i>et al</i> [<mark>41</mark>], 2013	\$	☆	☆	\$	公公	*	☆	☆
Xue <i>et al</i> [42], 2014	\$	☆	☆	Å	**	*	☆	☆
Yu et al <mark>[43]</mark> ,	*	☆	${\simeq}$	\$	**	Å	☆	\$

2015							
Zhang <i>et al</i> [44], 2013	☆	*	\$	\$ **	*	*	☆
Zhang et al [<mark>45</mark>], 2017	*	*	\$	\$ **	*		
Zhang <i>et al</i> [<mark>46]</mark> , 2020	☆	\$	☆	\$ \Box	\$		
Zhao <i>et al</i> [47], 2015	*	\$	\$	\$ **	*	☆	☆



Figure 1 PRISMA flowchart of study inclusions and exclusions.

The present study highlights the overexpression of HSP40 and HSP60, which correlates with higher 3-year OS in EC. Moreover, these HSP families are found overexpressed in various human cancer types beyond EC, including cervical cancer, glioma, skin cancer, lung cancer, colorectal cancer, kidney cancer, and gastric cancer[56-62].

DNAJ, a HSP40 family member[63], plays a crucial role in cellular functions, including stimulating ATPase activity and performing chaperone functions such as protein folding, unfolding, translation, translocation, and degradation[64]. The research conducted by Yu *et al*[43] on DNAJB6, a nuclear-localized member of the HSP40 family, establishes its independence as a factor associated with better OS in ESCC. Elevated DNAJB6 levels were linked to down regulated AKT signaling and decreased sensitivity to AKT inhibition, providing insights for molecular targeted therapy focusing on oncogene addiction[43]. The prognosis related to HSP40 is, in part, explained by its connection to lymphatic dissemination, as HSP40 overexpression is linked to a lower probability of lymph node metastasis, suggesting a potential association with host immunity and immune-promoting functions[25,33,35].

The HSP60 family serves as an antigen for both B and T-lymphocytes, acting as a ligand for toll-like receptors and playing a pivotal role in immunity[65]. The significance of this family is highlighted by the observation that HSP60 inactivation in mice results in embryonic lethality[66]. Xanthoudakis *et al*[67] demonstrated that HSP60 facilitates procaspase-3 maturation, initiating apoptosis through a Fas-independent pathway. Additionally, HSP60 regulates mitochondrial permeability transition, establishing a cytoprotective network that counters CypD-associated cell death in tumor contexts, where CypD is a component of the mitochondrial permeability transition pore. Furthermore, HSP60 plays a crucial role in protein import and quality control machinery[68,69].

Study	HSP Alive De	+ ead	H Aliv	ISP- e Dead	Risk Diff W with 95%CI	Veight (%)
CHIP						
Wen, 2013	42	97	26	69		3.91
Heterogeneity: $\tau^2 =$	= 0.00, l ² =	.%,	H² =		0.03 [-0.09, 0.15]	
Test of $\theta_i = \theta_j$: Q(0)	= -0.00, p) = .				
HSF1						
Liao, 2015	25	49	31	29	-0.18 [-0.34, -0.01]	3.74
Tsukao, 2017	52	57	74	29	-0.24 [-0.37, -0.11]	3.88
Heterogeneity: τ ² =	0.00, l ² =	0.01	1%, ⊦	¹² = 1.00	-0.22 [-0.32, -0.12]	
Test of $\theta_i = \theta_j$: Q(1)	= 0.34, p	= 0.	56		•	
HSP20						
Berg, 2011	13	3	26	45	0.45 [0.22, 0.67]	3.50
Kawanishi, 1999	27	12	0	10	- 0.69 [0.55, 0.84]	3.82
Luz, 2017	19	4	3	2	0.23 [-0.23, 0.68]	2.38
Nakajima, 2002	16	15	16	16	0.02 [-0.23, 0.26]	3.38
Söderström, 2019	8	45	14	22	-0.24 [-0.42, -0.05]	3.66
Tsukao, 2017	17	13	20	6	-0.20 [-0.44, 0.04]	3.42
Xue. 2014	36	11	27	38		3.72
Zhang X. 2020	31	87	12	19		3.65
Heterogeneity: $\tau^2 =$: 0.11. l ² =	90.9	98%.	H ² = 11.09	0.15 [-0.10, 0.39]	0.00
Test of $\theta_i = \theta_j$: Q(7)	= 97.56, p	o = C	0.00			
HSP40						
Yu. 2015	23	45	11	81		3.87
Heterogeneity: $T^2 =$	= 0 00 l ² =	%	H ² =	0.		0.07
Test of $\theta_i = \theta_j$: Q(0)	= 0.00, p	= .				
HSP60						
Faried 2004	42	21	20	40		3 74
Heterogeneity: $T^2 =$	0.00 12 =	%	H ² =	10		0.74
Test of $\theta_i = \theta_j$: Q(0)	= 0.00, r =	= .				
HSP70						
Kawanishi, 1999	13	1	8	23		3.58
Luz, 2017	14	4	5	5	0.28 [-0.09, 0.64]	2.80
Miyazaki, 2005	3	26	11	21	-0.24 [-0.44, -0.04]	3.61
Nakajima, 2002	18	13	13	18	0.16 [-0.08, 0.41]	3.39
Nakajima, 2009	45	19	27	34	0.26 [0.09, 0.43]	3.74
Noguchi, 2002	29	19	8	15	0.26 [0.02, 0.50]	3.42
Söderström, 2019	3	25	20	45	-0.20 [-0.36, -0.04]	3.76
Tsukao, 2017	11	4	26	14	0.08 [-0.18, 0.35]	3.28
Zhang H, 2013	38	52	22	8	-0.31 [-0.50, -0.12]	3.65
Zhao, 2015	17	50	25	21	-0.29 [-0.47, -0.11]	3.69
Heterogeneity: $\tau^2 =$	0.09, l ² =	89.5	51%,	H ² = 9.53	0.06 [-0.14, 0.26]	
Test of $\theta_i = \theta_j$: Q(9)	= 91.27, p	0 = 0	0.00		•	
HSP90						
Akutsu, 2011	33	24	7	14	0.25 [0.01, 0.48]	3.42
Berezowska, 2013	55	70	61	137	- 0.13 [0.02, 0.24]	3.94
Faried, 2004	34	27	30	32	0.07 [-0.10, 0.25]	3.70
Ou, 2014	139 1	14	57	18	-0.21 [-0.33, -0.10]	3.92
Tsukao, 2017	20	15	16	4	-0.23 [-0.47, 0.01]	3.42
Heterogeneity: τ ² =	0.03, l ² =	84.3	31%,	H ² = 6.37	0.00 [-0.18, 0.18]	
Test of $\theta_i = \theta_j$: Q(4)	= 26.85, p	0 = 0	0.00		~	
Overall					0.07 [-0.04 0.18]	
Heterogeneity: T ² =	0.07. l ² =	90.4	10%	H ² = 10.41	▼	
Test of $\theta = \theta$: $\Omega(27)$	() = 284.27	7. n :	= 0.0	0		
Test of group differe	ences: Q.((6) =	44.8	6, p = 0.00		
0	-DV				-0.5 0 0.5 1	
Random-effects REM	ML model					

Figure 2 Forest plot of the eligible studies assessing the relationship between heat shock protein expression and overall survival. HSP: Heat shock protein; CI: Confidence interval.

Baishideng® WJGO https://www.wjgnet.com

April 15, 2024 Volume 16 Issue 4

A Study	HSP-	⊢ Н с1/	SP-				Risk Diff	v	Veight
CHIP	01/2 00	01/	2 05				with 55 /00		(/0)
Wen, 2013	95 44	68	27				-0.03 [-0.15.	0.091	3.54
Heterogeneity: $\tau^2 =$	0.00, l ² = .	%. H ² :	=.			Ā	-0.03 [-0.15.	0.091	
Test of $\theta_i = \theta_i$: Q(0)	= 0.00, p =	·.	- •			•	0.00 [0.10,	0.00]	
HSF1									
Liao 2015	55 21	44	14			_	-0.03[-0.18	0 111	3 44
Tsukao 2017	84 16	70	24			-	0.10[-0.02	0.211	3 56
Heterogeneity: $\tau^2 =$		10	$H^2 = 1.85$			-	0.04 [-0.02,	0.171	0.00
Test of $\theta_i = \theta_i$: Q(1)	= 1.85, p =	: 0.17	s, 11º = 1.00				0.04 [-0.03,	0.17]	
HEDDO									
Borg 2011	17 16	25	20			_	0.05 [0.16	0 071	2 10
Deels 2004	1/ 10	25	29				0.05 [-0.16,	1.001	3.10
Doak, 2004	50 00	2	0			-	-1.00 [-1.00, -	0.01	3.72
Kawanishi, 1999	56 36		3			-	-0.09 [-0.39,	0.21]	2.79
Luz, 2017	19 4	4	1			_	— 0.03 [-0.36,	0.41]	2.42
Miyazaki, 2005	27 13	14	7			_	0.01 [-0.24,	0.26]	3.03
Nakajima, 2002	34 7	22	9			_	- 0.12 [-0.08,	0.32]	3.26
Xue, 2014	35 12	36	29				— 0.19 [0.02,	0.36]	3.35
Zhang Y, 2017	88 27	35	12				0.02 [-0.13,	0.17]	3.45
Zhang X, 2020	149 59	103	32			-	-0.05 [-0.14,	0.05]	3.61
Heterogeneity: $\tau^2 =$	0.13, l ² = 9	3.09%	o, H² = 14.47				-0.09 [-0.34,	0.16]	
Test of $\theta_i = \theta_j$: Q(8)	= 7.24, p =	0.51				•			
HSP40									
Yu, 2015	54 14	70	22				0.03 [-0.10,	0.16]	3.51
Heterogeneity: $\tau^2 =$	$0.00, I^2 = .$	%, H² :	=.				0.03 [-0.10,	0.16]	
Test of $\theta_i = \theta_j$: Q(0)	= 0.00, p =								
HSP60									
Faried, 2004	16 43	17	39			_	-0.03 [-0.20,	0.13]	3.38
Lv, 2022	13 34	27	13		_		-0.40 [-0.59, -	0.20]	3.27
Heterogeneity: $\tau^2 =$ Test of $\theta_i = \theta_i$: Q(1)	0.06, l² = 8 = 7.94, p =	87.41% 0.00	o, H² = 7.94		<		-0.21 [-0.57,	0.15]	
HSP70									
Kawanishi 1999	46 25	17	14			_	- 0.10[-0.11	0.311	3 21
	20 5	3	0		-		-0.20[-0.36]-	0.041	3.42
Miyazaki 2005	18 11	23	9				-0.10[-0.33	0.141	3.09
Nakajima 2002	23 8	23	8				0.00[-0.22	0.221	3 17
Nakajima, 2002	44 15	20	10				0.06 [-0.22,	0.22]	2 20
Nakajina, 2009	44 15 20 16	10	4				0.00[-0.10,	0.23	0.00
	32 10	19	4	-			-0.16[-0.36,	0.04]	3.23
Wang, 2007	0 13	41	0				-0.87 [-0.97, -	0.78]	3.60
Wang, 2010	88 24	8	-		1		-0.21 [-0.29, -	0.14]	3.65
Zhang H, 2013	70 20	23	/				0.01 [-0.16,	0.19]	3.35
Zhao, 2015	58 9	40	6				-0.00 [-0.13,	0.12]	3.52
Heterogeneity: $\tau^2 =$ Test of $\theta_i = \theta_i$: Q(9)	0.08, l² = 9 = 219.65,	93.73% o = 0.0	5, H² = 15.94 0				-0.14 [-0.33,	0.04]	
113P90	60 0		170				0.447.000	0.403	0.00
Berezowska, 2013	29 96	25	173				0.11 [0.02,	0.19]	3.62
Faried, 2004	19 40	14	42			_	0.07 [-0.09,	0.24]	3.39
Ou, 2014	193 60	56	19				0.02 [-0.10,	0.13]	3.56
Wang, 2007	38 11	9	2		_		-0.04 [-0.30,	0.21]	3.00
Wang, 2010	81 21	15	3		-		-0.04 [-0.23,	0.15]	3.29
Heterogeneity: $\tau^2 =$	$0.00, I^2 = 8$	3.61%,	H ² = 1.09			•	0.05 [-0.01,	0.12]	
rest of $\Theta_i = \Theta_j$: Q(4)	= 3.32, p =	0.51							
Overall						•	-0.08 [-0.18,	0.02]	
Heterogeneity: $\tau^2 =$ Test of $\theta_1 = \theta_2$: $\Omega/29$	0.07, l ² = 9	92.62%	o, H² = 13.55 00			•			
Test of group differe	nces: Q _b (6	6) = 7.3	81, p = 0.29						
Bandom-effects DEM	1 model			-1	-0.5	0	0.5		

 Jaisbideng®
 WJGO
 https://www.wjgnet.com

Study	H: NO	SP+) N+	N	HSP- 0 N+			Risk Diff with 95%CI	Weig (%
CHIP								
Wen, 2013	0	139	0	95			0.00 [0.00, 0.00] 3.4
Heterogeneity: $\tau^2 =$	0.00, I	² = .%	5, H ² :	=,				
Test of $\theta_i = \theta_j$: Q(0)	= ., p =	= .						
HSF1								
Liao, 2015	29	47	36	22	-	F	-0.24 [-0.41, -0.07] 3.2
Tsukao, 2017	31	77	41	62	-	-	-0.11 [-0.24, 0.02	3.3
Heterogeneity: $\tau^2 =$	0.00, I	l ² = 30	.61%	o, H² = 1.44			-0.16 [-0.29, -0.04]
Test of $\theta_i = \theta_j$: Q(1)	= 1.44	, p = 0	0.23					
HSP20								
Berg. 2011	26	7	18	36		-	0.45 0.27. 0.64	1 3.2
Doak, 2004	1	1	0	1				1 1.7
Kawanishi 1999	42	50	3	7		_	0.16[-0.15_0.46	1 29
Luz 2017	42	7	4	0	_	-	-0.44 [-0.68 -0.19	1 30
Luz, 2017	10	01	4	10		-	-0.44 [-0.00, -0.18	1 0.0
Miyazaki, 2005	19	21	9	12	-	_	0.05[-0.22, 0.31] 3.0
Nakajima, 2002	16	15	11	20		-	0.16[-0.08, 0.40	3.0
Xue, 2014	27	20	23	42		-	0.22 [0.04, 0.40] 3.2
Zhang Y, 2017	67	48	40	7	-		-0.27 [-0.40, -0.13	3.3
Zhang X, 2020	111	88	93	47	4		-0.11 [-0.21, -0.00] 3.3
Heterogeneity: $\tau^2 =$	0.07, I	l² = 88	8.04%	o, H ² = 8.36		•	0.05 [-0.15, 0.24]
Test of $\theta_i = \theta_j$: Q(8)	= 63.8	2, p =	0.00			Ţ		
HSP40								
Yu, 2015	33	35	28	64		-	0.18 [0.03, 0.33] 3.2
Heterogeneity: $\tau^2 =$	0.00, I	² = .%	5, H ² :	=.			0.18 [0.03, 0.33	1
Test of $\theta_i = \theta_j$: Q(0)	= 0.00), p = .				•	-	
HSP60								
Faried, 2004	27	36	28	32	-	-	-0.04 [-0.21, 0.14] 3.2
Lv, 2022	17	30	25	15	-	-	-0.26 [-0.47, -0.06	3.1
Test of $\theta_i = \theta_j$: Q(1) HSP70	= 2.70), p = (0.10					
Kawanishi, 1999	37	34	8	23			0.26 [0.07, 0.46] 3.1
Luz, 2017	12	6	1	1		-	0.17 [-0.56, 0.89] 1.6
Miyazaki, 2005	7	22	21	11			-0.41 [-0.64, -0.19] 3.1
Nakajima, 2002	13	18	14	17	_	_	-0.03 [-0.28, 0.21] 3.0
Nakajima, 2009	34	30	22	39		-	0.17 [-0.00, 0.34	3.2
Noguchi, 2002	27	21	4	19			0.39 [0.18, 0.60	- 13.1
Wang, 2007	9	45	6	0	-		-0.83 [-0.93 -0.73	1 3 3
Wang 2010	11	08	Q	0			-0.88 [-0.04 -0.94	1 2 3
7hang U 0019	F4	20	25	5			-0.00 [-0.94, -0.01	1 0.0
Zhang H, 2013	01	39	25	5			-0.27 [-0.43, -0.10	3.2
znao, 2015	20	47	31	15			-0.38 [-0.55, -0.20	J 3.2
Heterogeneity: $\tau^2 = 0$ Test of $\theta_i = \theta_i$: Q(9) =	0.18, l ^a = 386.9	² = 96 97, p	.83% = 0.0	, H ² = 31.56 0			-0.20 [-0.48, 0.07]
10000								
HSP90	~~		00	50		-	0.041.030.000	1
Akutsu, 2011	23	34	28	50			0.04 [-0.12, 0.21] 3.2
Berezowska, 2013	39	86	41	157			0.10 [0.01, 0.20	3.3
Faried, 2004	22	40	33	28	-	F	-0.19 [-0.36, -0.01] 3.2
Huang, 2014	27	44	2	8			0.18 [-0.09, 0.45] 2.9
Ou, 2014	117	136	38	37			-0.04 [-0.17, 0.08] 3.3
Wang, 2007	4	45	11	0			-0.92 [-1.00, -0.84] 3.3
Wang, 2010	4	98	18	0			-0.96 [-1.00, -0.92] 3.4
Heteroaeneity: $\tau^2 = 0$	0.23. 1	² = 98	.93%	, H ² = 93.37			-0.26 [-0.62. 0 10	1
Test of $\theta_i = \theta_j$: Q(6) =	= 714.4	44, p	= 0.0	0				
Overall							-0.12 [-0.26. 0.01	1
Heterogeneity: T ² - 4	0.13	2 <u>-</u> 06	81%	$H^2 = 31.34$				
Test of $\theta_i = \theta_j$: Q(31)	= 179	- 90 92.79,	p = 0).00				
Test of group differe	nces: (Q _b (6)	= 16.	40, p = 0.01				
					-1 -0.5	0 0.5	1	
Random-effects Hedg	ges mo	odel			_ 0.5			

Figure 3 Forest plot of the eligible studies evaluating the association of the heat shock proteins with the grade of cellular differentiation and lymph node dissemination. A: Forest plot of the eligible studies evaluating the association of the heat shock proteins with the grade of cellular differentiation; B: Forest plot of the eligible studies evaluating the association of the heat shock protein s with lymph node dissemination. HSP: Heat shock protein; CI: Confidence interval.

Baishideng® WJGO | https://www.wjgnet.com

The HSP60 family holds potential as a novel prognostic biomarker in esophageal and esophagogastric cancer[25,31]. The consequences of HSP60 knockdown are substantial, compromising the integrity of respiratory complex I and inducing an excessive production of reactive oxygen species (ROS). This surplus ROS production fuels tumor progression by activating AMP-activated protein kinase, facilitating the acquisition of the Warburg phenotype in HSP60 knockdown cells. Elevated ROS levels may lead to the fragmentation of iron-sulfur clusters, consequently upregulating the expression of ADHFe1. This, in turn, triggers an increase in the production of 2-hydroxyglutarate, impacting DNA methylation and influencing the tumor's epigenetic landscape[50].

The investigation delved into HSF1 expression and its correlation with lymph node metastasis and diminished 3-year OS in EC. HSF1, a participant in the HSR, plays a multifaceted role in orchestrating molecular changes contributing to malignancy progression [70]. Its activation transforms the tumor microenvironment, promoting processes such as angiogenesis, extracellular matrix (ECM) organization, adhesion, and migration. Elevated HSF1 expression in both tumor and stromal cells significantly correlates with worse disease-free survival and OS in ESCC. Conversely, lower levels of HSF1 activation indicate a more favorable outcome, suggesting its potential as a biomarker for ESCC patient prognosis. In vivo experiments demonstrate that the absence of HSF1 reduces tumor formation, further supporting its role in malignant growth[71]. This transformation occurs through the upregulation of genes promoting the malignant phenotype and the downregulation of genes that might trigger an anticancer immune response[50]. Moreover, HSF1 activation drives specific beneficial pathways within the malignant elements, fostering processes such as angiogenesis, ECM organization, adhesion, and migration[72]. It is plausible that HSF1 is involved in the ESCC microenvironment through the same molecular mechanism. Additionally, stress-damaged proteins, when accumulated in the cytoplasm, recruit HSP70 and HSP90, which bind to HSF1, impeding its activation. Upon activation, HSF1 translocates to the nucleus, binding to the heat shock element sequence in the promoter regions of HSP genes, thereby inducing the expression of inducible HSPs like HSP27, HSP60, HSP70, HSP90, and multidrug resistance 1[73]. Furthermore, HSF1 activation plays a pivotal role in the tumor stroma, especially within cancer-associated fibroblasts. In this context, HSF1 triggers the activation of genes associated with processes such as angiogenesis, ECM remodeling, cellular adhesion, and migration. These molecular alterations collectively contribute to the promotion of malignant growth [74]. In vivo experiments have substantiated this, demonstrating that the absence of HSF1 reduces tumor formation in a mouse model lacking p53[71]. This multifaceted role underscores the potential significance of HSF1 in orchestrating molecular changes contributing to the progression of malignancy, particularly within the context of ESCC. Liao et al[29] demonstrated that the high level of HSF1 expression in both tumor cells and stromal cells was significantly associated with worse disease-free survival and OS of ESCC patients. It was also demonstrated that lower levels of HSF1 activation in both stromal and tumor cells are indicative of a more favorable outcome for patients with ESCC, suggesting the potential of HSF1 activation as a biomarker for ESCC patient prognosis^[29]. These findings align with prior research indicating heightened levels of HSF1 expression across diverse cancer types. In a study involving over 1800 participants, nuclear HSF1 levels were elevated in 80% of in situ invasive breast carcinomas^[75].

Aligned to the current finding in ESCC, HSF1 has been linked to poor prognosis in various cancer types. Engerud *et al* [76] established an association between HSF1 overexpression and poor survival after analyzing 823 endometrial cancer lesions. Ishiwata *et al* [77] similarly demonstrated an association between HSF1 expression and lower OS in oral squamous cell carcinoma. Santagata *et al* [75] revealed that high HSF1 expression was associated with lymph node invasion in breast cancer patients. Evidence related to cytoskeleton suggests that HSF1 regulates cell motility in esophagogastric adenocarcinoma by binding to the ArgBP2 promoter with the sequence nGAAn[78]. The interaction of HSF1 with MORC2 further mediates invasion and migration in esophagogastric cancer cells by inhibiting ArgBP2, a crucial regulator of cytoskeleton and cellular motility[78]. These findings indicate that the presence of HSF1 influences cell motility, thereby impacting invasion and migration in esophagogastric cancer cells.

The biomarker profile of HSPs has the potential to enhance prognostic stratification accuracy in EC, offering a pathway for personalized medicine and precision therapy-essential components of modern oncology[79]. Targeted therapy, linked to extended OS and reduced treatment costs[80-82], can be optimized by understanding the role of HSPs in cancer development and progression. Breakthroughs in HSP inhibitors and HSP cancer vaccines have been proposed, with studies suggesting their capacity to induce therapeutic resistance against radiotherapy and chemotherapy. HSPs may emerge as crucial targeting molecules for cancer therapy, particularly in esophageal oncology[5]. HSP inhibitors, by targeting key pathways regulated by HSP, have the potential to revolutionize the treatment landscape, inhibiting prosurvival pathways and altering HSP receptor expression[83], thereby reducing malignant transformation and tumor growth[84,85]. Strategies such as genetic removal, stress pathway inhibitors, RNA aptamer insertion, and gene silencing with short hairpin RNA (shRNA) exhibit promising results in inhibiting HSF1 and impeding cancer progression[86,87]. Another practical possibility is the silencing of the HSF1 gene with a shRNA, as shown by Nakamura *et al*[88], with promising results regarding cancer cell proliferation and activation of apoptotic pathways.

HSPs may also serve as adjuvants for vaccines, as evidenced in experimental models where HSP60-containing exosomes induce a substantial antitumor CD8(+) T cell response[89]. The proinflammatory response elicited by HSP60 in macrophages triggers an adaptive cellular immune reaction, suggesting its potential in enhancing immunotherapy for cancer[72]. Furthermore, HSPs could assist in indicating specific palliative, adjuvant, or neoadjuvant chemotherapy or radiotherapy regimens. Profiling HSPs has the potential to improve precision in EC management, enabling the categorization of patients based on their likelihood of responding to chemotherapy[48]. This knowledge could spare some patients from unnecessary treatment and enhance OS. However, it is crucial to acknowledge that most studies involving HSP inhibitors are limited to preclinical analysis and early-stage trials, and only future research will provide robust evidence for the efficacy of HSP therapies in clinical practice.

Zaishidena® WJGO | https://www.wjgnet.com

Α	H	SP+	н	P-						Risk Di	ff	Weight
Study	T1/2	2 ТЗ/4	T1/2	т3/4						with 95%	СІ	(%)
HSF1												
Liao, 2015	15	61	33	25			_			-0.37 [-0.53,	-0.22]	4.59
Tsukao, 2017	43	66	49	54			_			-0.08 [-0.21,	0.05]	4.81
Heterogeneity: $\tau^2 =$	0.04,	l ² = 87	.04%	, H ² = 1	7.72	<				-0.22 [-0.51,	0.06]	
Test of $\theta_i = \theta_j$: Q(1)	= 7.72	2, p = 0	0.01									
HSP20												
Berg, 2011	21	12	27	27			_	_	_	0.14 [-0.08,	0.35]	4.00
Kawanishi, 1999	48	44	4	6						0.12 [-0.20,	0.44]	2.95
Miyazaki, 2005	6	34	4	17			_	—		-0.04 [-0.24,	0.16]	4.11
Nakajima, 2002	19	12	14	17			_			0.16 [-0.08,	0.41]	3.66
Xue, 2014	27	20	22	43				_		0.24 [0.05,	0.42]	4.31
Zhang Y, 2017	12	103	8	39			-	_		-0.07 [-0.19,	0.06]	4.92
Zhang X, 2020	34	157	52	86		-	-			-0.20 [-0.30,	-0.10]	5.12
Heterogeneity: $\tau^2 = 0$	0.02,	l ² = 73	.64%	, H ² = 3	3.79					0.03 [-0.10,	0.16]	
Test of $\theta_i = \theta_j$: Q(6) =	= 25.4	ł2, p =	0.00									
Vu 2015	7	61	13	70			_	L		-0.04[-0.14	0.061	5 00
Heterogeneity: $\tau^2 = 1$	0 00	1 ² = %	H ² =							-0.04 [-0.14,	0.061	0.00
Test of $\theta_{i} = \theta_{i}; Q(0);$	= 0.00). n = .	,							0.01[0.11,	0.00]	
HSP60								_				
Faried, 2004	37	26	30	30			_	-		0.09 [-0.09,	0.26]	4.38
Lv, 2022	25	22	25	15		-				-0.09 [-0.30,	0.11]	4.05
Heterogeneity: $\tau^2 =$	0.01,	$l^2 = 41$.08%	$H^{2} = 1$	1.70		<			0.01 [- 0. 1 7,	0.18]	
Test of $\theta_i = \theta_j$: Q(1) =	= 1.70), p = (0.19									
HSP70												
Kawanishi, 1999	39	32	13	18			_			0.13 [-0.08,	0.34]	4.03
Miyazaki, 2005	1	28	9	23						-0.25 [-0.42,	-0.08]	4.45
Nakajima, 2002	19	12	14	17			_			0.16 [-0.08,	0.41]	3.66
Nakajima, 2009	48	16	20	41				-		- 0.42 [0.26,	0.58]	4.56
Noguchi, 2002	24	24	9	14				_	_	0.11 [-0.14,	0.35]	3.66
Wang, 2010	36	76	6	4						-0.28 [-0.59,	0.04]	2.99
Zhang H, 2013	39	51	22	8			-			-0.30 [-0.49,	-0.11]	4.25
Heterogeneity: $\tau^2 =$	0.07,	l ² = 8	6.05%	, H² =	7.17		<			0.00 [-0.21,	0.22]	
Test of $\theta_i = \theta_j$: Q(6)	= 51.	58, p =	= 0.00									
HSP90												
Akutsu, 2011	31	26	43	35			-	—		-0.01 [-0.18,	0.16]	4.44
Berezowska, 2013	33	92	20	178						0.16 [0.08,	0.25]	5.20
Faried, 2004	30	32	37	24		-	-	_		-0.12 [-0.30,	0.05]	4.39
Huang, 2014	29	42	5	5						-0.09 [-0.42,	0.24]	2.86
Wang, 2010	31	71	9	7				-		-0.26 [-0.52,	0.00]	3.52
Heterogeneity: $\tau^2 =$	0.02,	l ² = 7	1.84%	, H² =	3.55					-0.04 [-0.19,	0.12]	
Test of $\theta_i = \theta_j$: Q(4)	= 16.	68, p =	= 0.00									
Overall										-0.02 [-0.10,	0.06]	
Heterogeneity: $\tau^2 =$	0.03,	l ² = 8	1.25%	, H² =	5.33						-	
Test of $\theta_i = \theta_j$: Q(23)	8) = 12	22.91,	p = 0.	00								
Test of group differe	ences	: Q _b (5)	= 2.8	2, p =	0.73	_						
						-0.5		0	0.5	-		
Random-effects REM	ML mo	del										

Baisbideng® WJGO | https://www.wjgnet.com

B Study	HSP+ M0 M1	н ма	SP- M1						Risk Diff with 95%CI		Weight (%)
HSF1							_				
Liao, 2015	73 3	55	3						0.01 [-0.06,	0.08]	4.99
Heterogeneity: $\tau^2 = 0$				•		0.01 [-0.06,	0.08]				
Test of $\theta_i = \theta_j$: Q(0) =	0.00, p	=.									
HSP20											
Berg, 2011	33 0	46	8						0.15 [0.05,	0.24]	4.92
Kawanishi, 1999	92 0	10	0						0.00 [0.00,	0.00]	5.08
Miyazaki, 2005	28 12	13	8						0.08 [-0.17,	0.33]	4.10
Nakajima, 2002	29 2	24	7				_	-	0.16 [-0.01,	0.33]	4.58
Xue, 2014	22 25	45	20			_	-	-	0.22 [-0.41,	-0.04]	4.52
Heterogeneity: $\tau^2 = 0$.02, l ² =	64.36%	%, H² = 2.	81				,	0.04 [-0.09,	0.17]	
Test of $\theta_i = \theta_j$: Q(4) =	13.62, p	0.0	1				•				
HSP40											
Yu, 2015	64 4	84	8				-	,	0.03 [-0.05,	0.11]	4.96
Heterogeneity: $\tau^2 = 0$.00, l ² =	.%, H²	=.				•		0.03 [-0.05,	0.11]	
Test of $\theta_i = \theta_j$: Q(0) = 0.00, p = .											
HSP60											
Faried, 2004	55 8	48	12						0.07 [-0.06,	0.20]	4.78
Lv. 2022	42 5	38	2				-	-	0.06 [-0.17.	0.05]	4.86
Heterogeneity: $\tau^2 = 0$.00. l ² =	54.39%	6. H² = 2.	19					0.00 [-0.12.	0.131	
Test of $\theta_i = \theta_j$: Q(1) =	2.19, p	= 0.14	-,				•		,		
HSP70											
Kawanishi, 1999	71	0 31	0						0.00 [0.00,	0.00]	5.08
Miyazaki, 2005	17 1	2 24	8					-	0.16 [-0.40,	0.07]	4.22
Nakajima, 2002	28	3 25	6					-	0.10 [-0.08,	0.27]	4.56
Nakajima, 2009	58	6 47	14				-		0.14 [0.01,	0.26]	4.79
Noguchi, 2002	39	9 17	6					-	0.07 [-0.14,	0.28]	4.36
Wang, 2007	20 3	4 θ	0		-	-		-	0.63 [-0.76,	-0.50]	4.78
Wang, 2010	56 5	6 8	0					-	0.50 [-0.59,	-0.41]	4.92
Heterogeneity: $\tau^2 = 0$	0.09, l² =	93.34	%, H² = 1	5.01				-	0.14 [-0.38,	0.09]	
Test of $\theta_i = \theta_j$: Q(6) =	= 120.69	, p = 0	00				-				
HSP90											
Berezowska, 2013	103 2	2 138	60				-		0.13 [0.03,	0.22]	4.93
Faried, 2004	52 1	0 51	10						0.00 [-0.13,	0.13]	4.78
Huang, 2014	71	0 10	0						0.00 [0.00,	0.00]	5.08
Wang, 2007	15 3	4 11	0		-	_		-	0.69 [-0.82,	-0.56]	4.78
Wang, 2010	46 5	6 18	0		-	-		-	0.55 [-0.65,	-0.45]	4.91
Heterogeneity: $\tau^2 = 0$	0.13, l² =	96.98	%, H ² = 3	3.11				-	0.22 [-0.55,	0.10]	
Test of $\theta_i = \theta_j$: Q(4) =	= 160.53	, p = 0	00								
Overall							•	-	0.09 [-0.21,	0.03]	
Heterogeneity: $\tau^2 = 0$	7.32			Ŧ							
Test of $\theta_i = \theta_i$: Q(20)	= 412.5	5, p =	0.00								
Test of aroun differen	nces: O	(5) – 4	01 n - 0	55							
lost of group differen		(3) = 4		-1		-0.5	0	0.5	5		

Random-effects REML model

Figure 4 Forest plot of the eligible studies evaluating the association of heat shock proteins with tumor depth and distant metastasis. A: Forest plot of the eligible studies evaluating the association of heat shock proteins with tumor depth; B: Forest plot of the eligible studies evaluating the association of heat shock proteins with distant metastasis. HSP: Heat shock protein; CI: Confidence interval.

 Jaisbideng®
 WJGO
 https://www.wjgnet.com

Despite these insights, the meta-analysis is subject to limitations. The number of studies conducted for HSP40, HSP60, and HSF1 was relatively small, potentially impacting the overall robustness of the findings. Besides, most of the studies included only ESCC, and few studies assessed the impact of HSP on esophageal adenocarcinoma and esophagogastric junction neoplasms. Although we performed subgroup analysis for ESCC, the same subgroup analysis was not possible for adenocarcinoma due to the small number of articles. Consequently, future studies should investigate the potential value of HSP in cancer prognostication and therapy.

CONCLUSION

Our systematic review and meta-analysis highlight a significant correlation between the overexpression of HSP40 and 60, and low HSF1 expression, and favorable outcomes, including prolonged survival and diminished lymph node dissemination in individuals with esophageal and esophagogastric cancer. These results underscore the noteworthy prognostic implications of HSPs within the realm of cancer research, suggesting potential avenues for therapeutic interventions. The ongoing exploration of this field offers the prospect of furthering precision medicine and developing targeted strategies for the management of esophageal and esophagogastric cancer.

ARTICLE HIGHLIGHTS

Research background

The association between heat shock proteins (HSPs) and cancer prognosis has generated significant interest, offering potential implications for clinical decision-making in cancer management. HSPs and their regulatory factors, such as heat shock factor (HSF)1 and CHIP, play an important role in cellular protection against stress events, and are overexpressed in some types of cancer.

Research motivation

The prognostic significance of HSPs and their regulatory factors, such as HSF1 and CHIP, are poorly understood in esophageal and esophagogastric cancer.

Research objectives

We conducted a systematic review and meta-analysis to investigate the relationship between HSP expression and prognosis in esophageal and esophagogastric cancer.

Research methods

A systematic review was conducted in accordance with PRISMA recommendations, on Embase, PubMed, Cochrane, and LILACS. Cohort, case-control, and cross-sectional studies of patients with esophagus or esophagogastric cancer were included. HSP-positive patients were compared with HSP-negative, and the endpoints analyzed were lymph node metastasis, tumor depth, distant metastasis, and overall survival (OS). HSPs were stratified according to the HSP family, and the summary risk difference (RD) was calculated using a random-effect model.

Research results

The final selection comprised 27 studies, including esophageal squamous cell carcinoma (21), esophagogastric adenocarcinoma (5), and mixed neoplasms (1). The pooled sample size was 3465 patients. HSP40 and 60 were associated with a higher 3-year OS, while HSF1 was associated with a poor 3-year OS. The other HSP families were not associated with long-term survival. HSF1 was associated with a higher probability of lymph node metastasis. HSP40 was associated with a lower probability of lymph node dissemination. The expression of other HSP families was not significantly related to tumor depth and lymph node or distant metastasis.

Research conclusions

Our findings demonstrated that the expression levels of some families of HSP, such as HSP40 and 60 and HSF1, are associated with long-term survival and lymph node dissemination in esophageal and esophagogastric cancer.

Research perspectives

The results of this study underscore the noteworthy prognostic implications of HSPs within the realm of cancer research, suggesting potential avenues for therapeutic interventions. The ongoing exploration of this field offers the prospect of furthering precision medicine and developing targeted strategies for the management of esophageal and esophagogastric cancer.

Raisbidena® WJGO https://www.wjgnet.com

FOOTNOTES

Author contributions: Tustumi F designed the research, analyzed the data; Nakamura ET, Park A, and Tustumi F wrote the paper; Nakamura ET and Park A performed the research; Pereira MA and Kikawa D assisted in the research and reviewed the final manuscript.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

PRISMA 2009 Checklist statement: The authors have read the PRISMA 2009 Checklist, and the manuscript was prepared and revised according to the PRISMA 2009 Checklist.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: Brazil

ORCID number: Eric Toshiyuki Nakamura 0000-0002-9903-7553; Amanda Park 0000-0001-5142-7723; Marina Alessandra Pereira 0000-0002-6865-0988; Daniel Kikawa 0009-0001-3724-7774; Francisco Tustumi 0000-0001-6695-0496.

S-Editor: Wang JJ L-Editor: Webster JR P-Editor: Zhang XD

REFERENCES

- Böhme F, Racz K, Sebesta C Jr, Sebesta C. [Esophageal Cancer]. Wien Med Wochenschr 2023; 173: 209-215 [PMID: 36318394 DOI: 1 10.1007/s10354-022-00972-9
- Pennathur A, Gibson MK, Jobe BA, Luketich JD. Oesophageal carcinoma. Lancet 2013; 381: 400-412 [PMID: 23374478 DOI: 2 10.1016/S0140-6736(12)60643-6
- Zhou N, Hofstetter WL. Prognostic and therapeutic molecular markers in the clinical management of esophageal cancer. Expert Rev Mol Diagn 3 2020; 20: 401-411 [PMID: 32067548 DOI: 10.1080/14737159.2020.1731307]
- 4 Mao Y, Wang Y, Dong L, Zhang Y, Wang C, Zhang Q, Yang S, Cao L, Zhang X, Li X, Fu Z. Hypoxic exosomes facilitate angiogenesis and metastasis in esophageal squamous cell carcinoma through altering the phenotype and transcriptome of endothelial cells. J Exp Clin Cancer Res 2019; 38: 389 [PMID: 31488217 DOI: 10.1186/s13046-019-1384-8]
- Yun CW, Kim HJ, Lim JH, Lee SH. Heat Shock Proteins: Agents of Cancer Development and Therapeutic Targets in Anti-Cancer Therapy. 5 Cells 2019; 9 [PMID: 31878360 DOI: 10.3390/cells9010060]
- Wu J, Liu T, Rios Z, Mei Q, Lin X, Cao S. Heat Shock Proteins and Cancer. Trends Pharmacol Sci 2017; 38: 226-256 [PMID: 28012700 DOI: 6 10.1016/j.tips.2016.11.009
- 7 Shamovsky I, Nudler E. New insights into the mechanism of heat shock response activation. Cell Mol Life Sci 2008; 65: 855-861 [PMID: 18239856 DOI: 10.1007/s00018-008-7458-y]
- 8 Garrido C, Brunet M, Didelot C, Zermati Y, Schmitt E, Kroemer G. Heat shock proteins 27 and 70: anti-apoptotic proteins with tumorigenic properties. Cell Cycle 2006; 5: 2592-2601 [PMID: 17106261 DOI: 10.4161/cc.5.22.3448]
- Razzaque MS, Taguchi T. Involvement of stress proteins in renal diseases. Contrib Nephrol 2005; 148: 1-7 [PMID: 15912023 DOI: 9 10.1159/000086033]
- Wang WM, Jin HZ. Heat shock proteins and psoriasis. Eur J Dermatol 2019; 29: 121-125 [PMID: 30998191 DOI: 10.1684/ejd.2019.3526] 10
- Hunt AP, Minett GM, Gibson OR, Kerr GK, Stewart IB. Could Heat Therapy Be an Effective Treatment for Alzheimer's and Parkinson's 11 Diseases? A Narrative Review. Front Physiol 2019; 10: 1556 [PMID: 31998141 DOI: 10.3389/fphys.2019.01556]
- Zou X, Zhang G, Cheng Z, Yin D, Du T, Ju G, Miao S, Liu G, Lu M, Zhu Y. Microvesicles derived from human Wharton's Jelly mesenchymal 12 stromal cells ameliorate renal ischemia-reperfusion injury in rats by suppressing CX3CL1. Stem Cell Res Ther 2014; 5: 40 [PMID: 24646750 DOI: 10.1186/scrt428]
- Ledford H. Cancer theory faces doubts. Nature 2011; 472: 273 [PMID: 21512545 DOI: 10.1038/472273a] 13
- Calderwood SK, Khaleque MA, Sawyer DB, Ciocca DR. Heat shock proteins in cancer: chaperones of tumorigenesis. Trends Biochem Sci 14 2006; **31**: 164-172 [PMID: 16483782 DOI: 10.1016/j.tibs.2006.01.006]
- Ciocca DR, Calderwood SK. Heat shock proteins in cancer: diagnostic, prognostic, predictive, and treatment implications. Cell Stress 15 Chaperones 2005; 10: 86-103 [PMID: 16038406 DOI: 10.1379/CSC-99r.1]
- Lebret T, Watson RW, Molinié V, O'Neill A, Gabriel C, Fitzpatrick JM, Botto H. Heat shock proteins HSP27, HSP60, HSP70, and HSP90: 16 expression in bladder carcinoma. Cancer 2003; 98: 970-977 [PMID: 12942564 DOI: 10.1002/cncr.11594]
- Wang XW, Shi XH, Tong YS, Cao XF. The Prognostic Impact of Heat Shock Proteins Expression in Patients with Esophageal Cancer: A 17 Meta-Analysis. Yonsei Med J 2015; 56: 1497-1502 [PMID: 26446629 DOI: 10.3349/ymj.2015.56.6.1497]
- Kaigorodova EV, Bogatyuk MV. Heat shock proteins as prognostic markers of cancer. Curr Cancer Drug Targets 2014; 14: 713-726 [PMID: 18 25258164 DOI: 10.2174/1568009614666140926122846]
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, Shamseer L, Tetzlaff JM, Akl EA, Brennan SE, Chou R, 19 Glanville J, Grimshaw JM, Hróbjartsson A, Lalu MM, Li T, Loder EW, Mayo-Wilson E, McDonald S, McGuinness LA, Stewart LA, Thomas J, Tricco AC, Welch VA, Whiting P, Moher D. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. Int J Surg 2021; 88: 105906 [PMID: 33789826 DOI: 10.1016/j.ijsu.2021.105906]



- Nakamura ET, Tustumi F, Kikawa D, Santos AGE, Silva EIR. Heat-Shock Proteins in Esophageal Cancer: A systematic review and meta-20 analisis. [cited 27 August 2023]. Available from: https://www.crd.york.ac.uk/prospero/display_record.php?ID=CRD42022370653
- Akutsu Y, Matsubara H, Kano M, Usui A, Yoneyama Y, Ikeda N, Komatsu A, Yusup G. Correlation between gp96 expression and the surgical 21 outcome in patients with esophageal squamous cell carcinoma. Ann Surg Oncol 2011; 18: 832-837 [PMID: 20839070 DOI: 10.1245/s10434-010-1128-7]
- Berezowska S, Novotny A, Bauer K, Feuchtinger A, Slotta-Huspenina J, Becker K, Langer R, Walch A. Association between HSP90 and Her2 22 in gastric and gastroesophageal carcinomas. PLoS One 2013; 8: e69098 [PMID: 23874879 DOI: 10.1371/journal.pone.0069098]
- Berg D, Wolff C, Langer R, Schuster T, Feith M, Slotta-Huspenina J, Malinowsky K, Becker KF. Discovery of new molecular subtypes in 23 oesophageal adenocarcinoma. PLoS One 2011; 6: e23985 [PMID: 21966358 DOI: 10.1371/journal.pone.0023985]
- 24 Doak SH, Jenkins GJ, Parry EM, Griffiths AP, Baxter JN, Parry JM. Differential expression of the MAD2, BUB1 and HSP27 genes in Barrett's oesophagus-their association with aneuploidy and neoplastic progression. Mutat Res 2004; 547: 133-144 [PMID: 15013707 DOI: 10.1016/j.mrfmmm.2003.12.009
- Faried A, Sohda M, Nakajima M, Miyazaki T, Kato H, Kuwano H. Expression of heat-shock protein Hsp60 correlated with the apoptotic index 25 and patient prognosis in human oesophageal squamous cell carcinoma. Eur J Cancer 2004; 40: 2804-2811 [PMID: 15571964 DOI: 10.1016/j.ejca.2004.08.013]
- Huang T, Chen S, Han H, Li H, Huang Z, Zhang J, Yin Q, Wang X, Ma X, Dai P, Duan D, Zou F, Chen X. Expression of Hsp90a and cyclin 26 B1 were related to prognosis of esophageal squamous cell carcinoma and keratin pearl formation. Int J Clin Exp Pathol 2014; 7: 1544-1552 [PMID: 24817950]
- Iqbal MK, Zargar MA, Mudassar S, Lone GN, Yaseen SB, Andrabi KI. Expression Profiling and Cellular Localization of Stress Responsive 27 Proteins in Squamous Cell Carcinoma of Human Esophagus. Cancer Invest 2016; 34: 237-245 [PMID: 27351523 DOI: 10.1080/07357907.2016.1178760]
- Kawanishi K, Shiozaki H, Doki Y, Sakita I, Inoue M, Yano M, Tsujinaka T, Shamma A, Monden M. Prognostic significance of heat shock 28 proteins 27 and 70 in patients with squamous cell carcinoma of the esophagus. Cancer 1999; 85: 1649-1657 [PMID: 10223556 DOI: 10.1002/(SICI)1097-0142(19990415)85:8<1649::AID-CNCR2>3.0.CO;2-V]
- Liao Y, Xue Y, Zhang L, Feng X, Liu W, Zhang G. Higher heat shock factor 1 expression in tumor stroma predicts poor prognosis in 29 esophageal squamous cell carcinoma patients. J Transl Med 2015; 13: 338 [PMID: 26511079 DOI: 10.1186/s12967-015-0703-x]
- 30 Luz CC, Noguti J, Borges de Araújo L, Gianni MS, Simão Gomes T, Ricardo AN. Hsp27 and Hsp70 Expression in Esophageal Squamous. Asian Pac J Cancer Prev 2017; 18: 789-794 [PMID: 28441788 DOI: 10.22034/APJCP.2017.18.3.789]
- 31 Lv J, Wang XW, Sun XK, Yang JR, Chen PR. High Expression of Heat Shock Protein Family D Member 1 Predicts Poor Prognosis of Esophageal Cancer. J Clin Med Res 2022; 14: 273-281 [PMID: 35974809 DOI: 10.14740/jocmr4424]
- Miyazaki T, Kato H, Faried A, Sohda M, Nakajima M, Fukai Y, Masuda N, Manda R, Fukuchi M, Ojima H, Tsukada K, Kuwano H. 32 Predictors of response to chemo-radiotherapy and radiotherapy for esophageal squamous cell carcinoma. Anticancer Res 2005; 25: 2749-2755 [PMID: 16080521]
- Nakajima M, Kuwano H, Miyazaki T, Masuda N, Kato H. Significant correlation between expression of heat shock proteins 27, 70 and 33 lymphocyte infiltration in esophageal squamous cell carcinoma. Cancer Lett 2002; 178: 99-106 [PMID: 11849747 DOI: 10.1016/S0304-3835(01)00825-4]
- Nakajima M, Kato H, Miyazaki T, Fukuchi M, Masuda N, Fukai Y, Sohda M, Ahmad F, Kuwano H. Tumor immune systems in esophageal 34 cancer with special reference to heat-shock protein 70 and humoral immunity. Anticancer Res 2009; 29: 1595-1606 [PMID: 19443372]
- Noguchi T, Takeno S, Shibata T, Uchida Y, Yokoyama S, Müller W. Expression of heat shock protein 70 in grossly resected esophageal 35 squamous cell carcinoma. Ann Thorac Surg 2002; 74: 222-226 [PMID: 12118763 DOI: 10.1016/S0003-4975(02)03641-X]
- Ou Y, Liu L, Xue L, Zhou W, Zhao Z, Xu B, Song Y, Zhan Q. TRAP1 shows clinical significance and promotes cellular migration and 36 invasion through STAT3/MMP2 pathway in human esophageal squamous cell cancer. J Genet Genomics 2014; 41: 529-537 [PMID: 25438697 DOI: 10.1016/j.jgg.2014.08.004]
- Söderström HK, Kauppi JT, Oksala N, Paavonen T, Krogerus L, Räsänen J, Rantanen T. Overexpression of HSP27 and HSP70 is associated 37 with decreased survival among patients with esophageal adenocarcinoma. World J Clin Cases 2019; 7: 260-269 [PMID: 30746368 DOI: 10.12998/wjcc.v7.i3.260]
- Tsukao Y, Yamasaki M, Miyazaki Y, Makino T, Takahashi T, Kurokawa Y, Miyata H, Nakajima K, Takiguchi S, Mimori K, Mori M, Doki Y. 38 Overexpression of heat-shock factor 1 is associated with a poor prognosis in esophageal squamous cell carcinoma. Oncol Lett 2017; 13: 1819-1825 [PMID: 28454329 DOI: 10.3892/ol.2017.5637]
- Wang XP, Wang QX, Ying XP. Correlation between clinicopathology and expression of heat shock protein 72 and glycoprotein 96 in human 39 gastric adenocarcinoma. Tohoku J Exp Med 2007; 212: 35-41 [PMID: 17464101 DOI: 10.1620/tjem.212.35]
- 40 Wang X, Wang Q, Lin H. Correlation between clinicopathology and expression of heat shock protein 72 and glycoprotein 96 in human esophageal squamous cell carcinoma. Clin Dev Immunol 2010; 2010: 212537 [PMID: 20300187 DOI: 10.1155/2010/212537]
- Wen J, Luo KJ, Hu Y, Yang H, Fu JH. Metastatic lymph node CHIP expression is a potential prognostic marker for resected esophageal 41 squamous cell carcinoma patients. Ann Surg Oncol 2013; 20: 1668-1675 [PMID: 23429937 DOI: 10.1245/s10434-012-2733-4]
- 42 Xue L, Yang L, Jin ZA, Gao F, Kang JQ, Xu GH, Liu B, Li H, Wang XJ, Liu LJ, Wang BL, Liang SH, Ding J. Increased expression of HSP27 inhibits invasion and metastasis in human esophageal squamous cell carcinoma. Tumour Biol 2014; 35: 6999-7007 [PMID: 24748206 DOI: 10.1007/s13277-014-1946-5
- Yu VZ, Wong VC, Dai W, Ko JM, Lam AK, Chan KW, Samant RS, Lung HL, Shuen WH, Law S, Chan YP, Lee NP, Tong DK, Law TT, Lee 43 VH, Lung ML. Nuclear Localization of DNAJB6 Is Associated With Survival of Patients With Esophageal Cancer and Reduces AKT Signaling and Proliferation of Cancer Cells. Gastroenterology 2015; 149: 1825-1836.e5 [PMID: 26302489 DOI: 10.1053/j.gastro.2015.08.025]
- Zhang H, Chen W, Duan CJ, Zhang CF. Overexpression of HSPA2 is correlated with poor prognosis in esophageal squamous cell carcinoma. 44 World J Surg Oncol 2013; 11: 141 [PMID: 23777267 DOI: 10.1186/1477-7819-11-141]
- Zhang Y, Feng Z, Wang W, Dong J, Gong X, Pu H, Chen X. Expression of Heat Shock Protein-27 (Hsp27) and P38MAPK in Esophageal 45 Squamous Cell Carcinoma. Med Sci Monit 2017; 23: 5246-5253 [PMID: 29099815 DOI: 10.12659/MSM.904912]
- Zhang X, Liu T, Zheng S, Liu Q, Shen T, Han X, Zhang Q, Yang L, Lu X. SUMOylation of HSP27 regulates PKM2 to promote esophageal 46 squamous cell carcinoma progression. Oncol Rep 2020; 44: 1355-1364 [PMID: 32945483 DOI: 10.3892/or.2020.7711]
- Zhao G, Kang J, Jiao K, Xu G, Yang L, Tang S, Zhang H, Wang Y, Nie Y, Wu K, Fan D, Zhang D. High Expression of GRP78 Promotes 47 Invasion and Metastases in Patients with Esophageal Squamous Cell Carcinoma. Dig Dis Sci 2015; 60: 2690-2699 [PMID: 25976624 DOI:



10.1007/s10620-015-3689-6]

- 48 Tustumi F, Agareno GA, Galletti RP, da Silva RBR, Quintas JG, Sesconetto LA, Szor DJ, Wolosker N. The Role of the Heat-Shock Proteins in Esophagogastric Cancer. Cells 2022; 11 [PMID: 36078072 DOI: 10.3390/cells11172664]
- Jolly C, Morimoto RI. Role of the heat shock response and molecular chaperones in oncogenesis and cell death. J Natl Cancer Inst 2000; 92: 49 1564-1572 [PMID: 11018092 DOI: 10.1093/jnci/92.19.1564]
- Tang H, Chen Y, Liu X, Wang S, Lv Y, Wu D, Wang Q, Luo M, Deng H. Downregulation of HSP60 disrupts mitochondrial proteostasis to 50 promote tumorigenesis and progression in clear cell renal cell carcinoma. Oncotarget 2016; 7: 38822-38834 [PMID: 27246978 DOI: 10.18632/oncotarget.9615]
- 51 Dutta SM, Mustafi SB, Raha S, Chakraborty SK. Assessment of thermal stress adaptation by monitoring Hsp70 and MnSOD in the freshwater gastropod, Bellamya bengalensis (Lamark 1882). Environ Monit Assess 2014; 186: 8961-8967 [PMID: 25240497 DOI: 10.1007/s10661-014-4057-2]
- Hamel C, Ahmadzai N, Beck A, Thuku M, Skidmore B, Pussegoda K, Bjerre L, Chatterjee A, Dennis K, Ferri L, Maziak DE, Shea BJ, Hutton 52 B, Little J, Moher D, Stevens A. Screening for esophageal adenocarcinoma and precancerous conditions (dysplasia and Barrett's esophagus) in patients with chronic gastroesophageal reflux disease with or without other risk factors: two systematic reviews and one overview of reviews to inform a guideline of the Canadian Task Force on Preventive Health Care (CTFPHC). Syst Rev 2020; 9: 20 [PMID: 31996261 DOI: 10.1186/s13643-020-1275-21
- Villaseca MA, Roa I, Araya JC, Roa JC, Flores P. Double immunostaining for p53 and molecular chaperone hsp72/73 in gastric carcinoma. 53 Mol Pathol 1997; 50: 317-321 [PMID: 9536282 DOI: 10.1136/mp.50.6.317]
- Dorsey WC, Tchounwou PB. CYP1a1, HSP70, P53, and c-fos expression in human liver carcinoma cells (HepG2) exposed to 54 pentachlorophenol. Biomed Sci Instrum 2003; 39: 389-396 [PMID: 12724925]
- Uno Y, Kanda M, Miwa T, Umeda S, Tanaka H, Tanaka C, Kobayashi D, Suenaga M, Hattori N, Hayashi M, Yamada S, Nakayama G, 55 Fujiwara M, Kodera Y. Increased Expression of DNAJC12 is Associated with Aggressive Phenotype of Gastric Cancer. Ann Surg Oncol 2019; 26: 836-844 [PMID: 30617870 DOI: 10.1245/s10434-018-07149-y]
- Castle PE, Ashfaq R, Ansari F, Muller CY. Immunohistochemical evaluation of heat shock proteins in normal and preinvasive lesions of the 56 cervix. Cancer Lett 2005; 229: 245-252 [PMID: 16112431 DOI: 10.1016/j.canlet.2005.06.045]
- Canamasas I, Debes A, Natali PG, Kurzik-Dumke U. Understanding human cancer using Drosophila: Tid47, a cytosolic product of the DnaJ-57 like tumor suppressor gene 12Tid, is a novel molecular partner of patched related to skin cancer. J Biol Chem 2003; 278: 30952-30960 [PMID: 12783860 DOI: 10.1074/jbc.M304225200]
- Trentin GA, He Y, Wu DC, Tang D, Rozakis-Adcock M. Identification of a hTid-1 mutation which sensitizes gliomas to apoptosis. FEBS Lett 58 2004; 578: 323-330 [PMID: 15589840 DOI: 10.1016/j.febslet.2004.11.034]
- 59 Oka M, Sato S, Soda H, Fukuda M, Kawabata S, Nakatomi K, Shiozawa K, Nakamura Y, Ohtsuka K, Kohno S. Autoantibody to heat shock protein Hsp40 in sera of lung cancer patients. Jpn J Cancer Res 2001; 92: 316-320 [PMID: 11267942 DOI: 10.1111/j.1349-7006.2001.tb01097.x]
- 60 Kanazawa Y, Isomoto H, Oka M, Yano Y, Soda H, Shikuwa S, Takeshima F, Omagari K, Mizuta Y, Murase K, Nakagoe T, Ohtsuka K, Kohno S. Expression of heat shock protein (Hsp) 70 and Hsp 40 in colorectal cancer. Med Oncol 2003; 20: 157-164 [PMID: 12835518 DOI: 10.1385/MO:20:2:157
- Teng R, Liu Z, Tang H, Zhang W, Chen Y, Xu R, Chen L, Song J, Liu X, Deng H. HSP60 silencing promotes Warburg-like phenotypes and 61 switches the mitochondrial function from ATP production to biosynthesis in ccRCC cells. Redox Biol 2019; 24: 101218 [PMID: 31112866 DOI: 10.1016/j.redox.2019.101218]
- Isomoto H, Oka M, Yano Y, Kanazawa Y, Soda H, Terada R, Yasutake T, Nakayama T, Shikuwa S, Takeshima F, Udono H, Murata I, 62 Ohtsuka K, Kohno S. Expression of heat shock protein (Hsp) 70 and Hsp 40 in gastric cancer. Cancer Lett 2003; 198: 219-228 [PMID: 12957361 DOI: 10.1016/S0304-3835(03)00305-7]
- 63 Chatterjee S, Burns TF. Targeting Heat Shock Proteins in Cancer: A Promising Therapeutic Approach. Int J Mol Sci 2017; 18 [PMID: 28914774 DOI: 10.3390/ijms18091978]
- Kaida A, Iwakuma T. Regulation of p53 and Cancer Signaling by Heat Shock Protein 40/J-Domain Protein Family Members. Int J Mol Sci 64 2021; 22 [PMID: 34948322 DOI: 10.3390/ijms222413527]
- Lv LH, Wan YL, Lin Y, Zhang W, Yang M, Li GL, Lin HM, Shang CZ, Chen YJ, Min J. Anticancer drugs cause release of exosomes with 65 heat shock proteins from human hepatocellular carcinoma cells that elicit effective natural killer cell antitumor responses in vitro. J Biol Chem 2012; 287: 15874-15885 [PMID: 22396543 DOI: 10.1074/jbc.M112.340588]
- Christensen JH, Nielsen MN, Hansen J, Füchtbauer A, Füchtbauer EM, West M, Corydon TJ, Gregersen N, Bross P. Inactivation of the 66 hereditary spastic paraplegia-associated Hspd1 gene encoding the Hsp60 chaperone results in early embryonic lethality in mice. Cell Stress Chaperones 2010; 15: 851-863 [PMID: 20393889 DOI: 10.1007/s12192-010-0194-x]
- Xanthoudakis S, Roy S, Rasper D, Hennessey T, Aubin Y, Cassady R, Tawa P, Ruel R, Rosen A, Nicholson DW. Hsp60 accelerates the 67 maturation of pro-caspase-3 by upstream activator proteases during apoptosis. EMBO J 1999; 18: 2049-2056 [PMID: 10205159 DOI: 10.1093/emboj/18.8.2049]
- Prangenberg J, Doberentz E, Mawick A, Madea B. Mini Review: The Forensic Value of Heat Shock Proteins. Front Med (Lausanne) 2021; 8: 68 800100 [PMID: 35083250 DOI: 10.3389/fmed.2021.800100]
- 69 Tang H, Li J, Liu X, Wang G, Luo M, Deng H. Down-regulation of HSP60 Suppresses the Proliferation of Glioblastoma Cells via the ROS/ AMPK/mTOR Pathway. Sci Rep 2016; 6: 28388 [PMID: 27325206 DOI: 10.1038/srep28388]
- 70 Wang G, Cao P, Fan Y, Tan K. Emerging roles of HSF1 in cancer: Cellular and molecular episodes. Biochim Biophys Acta Rev Cancer 2020; 1874: 188390 [PMID: 32653364 DOI: 10.1016/j.bbcan.2020.188390]
- Min JN, Huang L, Zimonjic DB, Moskophidis D, Mivechi NF. Selective suppression of lymphomas by functional loss of Hsfl in a p53-71 deficient mouse model for spontaneous tumors. Oncogene 2007; 26: 5086-5097 [PMID: 17310987 DOI: 10.1038/sj.onc.1210317]
- Chen W, Syldath U, Bellmann K, Burkart V, Kolb H. Human 60-kDa heat-shock protein: a danger signal to the innate immune system. J 72 Immunol 1999; 162: 3212-3219 [PMID: 10092772 DOI: 10.4049/jimmunol.162.6.3212]
- Vilaboa NE, Galán A, Troyano A, de Blas E, Aller P. Regulation of multidrug resistance 1 (MDR1)/P-glycoprotein gene expression and 73 activity by heat-shock transcription factor 1 (HSF1). J Biol Chem 2000; 275: 24970-24976 [PMID: 10816597 DOI: 10.1074/jbc.M909136199]
- Scherz-Shouval R, Santagata S, Mendillo ML, Sholl LM, Ben-Aharon I, Beck AH, Dias-Santagata D, Koeva M, Stemmer SM, Whitesell L, 74 Lindquist S. The reprogramming of tumor stroma by HSF1 is a potent enabler of malignancy. Cell 2014; 158: 564-578 [PMID: 25083868 DOI:



10.1016/j.cell.2014.05.045]

- 75 Santagata S, Hu R, Lin NU, Mendillo ML, Collins LC, Hankinson SE, Schnitt SJ, Whitesell L, Tamimi RM, Lindquist S, Ince TA. High levels of nuclear heat-shock factor 1 (HSF1) are associated with poor prognosis in breast cancer. Proc Natl Acad Sci USA 2011; 108: 18378-18383 [PMID: 22042860 DOI: 10.1073/pnas.1115031108]
- 76 Engerud H, Tangen IL, Berg A, Kusonmano K, Halle MK, Oyan AM, Kalland KH, Stefansson I, Trovik J, Salvesen HB, Krakstad C. High level of HSF1 associates with aggressive endometrial carcinoma and suggests potential for HSP90 inhibitors. Br J Cancer 2014; 111: 78-84 [PMID: 24853175 DOI: 10.1038/bjc.2014.262]
- Ishiwata J, Kasamatsu A, Sakuma K, Iyoda M, Yamatoji M, Usukura K, Ishige S, Shimizu T, Yamano Y, Ogawara K, Shiiba M, Tanzawa H, 77 Uzawa K. State of heat shock factor 1 expression as a putative diagnostic marker for oral squamous cell carcinoma. Int J Oncol 2012; 40: 47-52 [PMID: 21879256 DOI: 10.3892/ijo.2011.1178]
- 78 Tong Y, Li Y, Gu H, Wang C, Liu F, Shao Y, Li F. HSF1, in association with MORC2, downregulates ArgBP2 via the PRC2 family in gastric cancer cells. Biochim Biophys Acta Mol Basis Dis 2018; 1864: 1104-1114 [PMID: 29339121 DOI: 10.1016/j.bbadis.2018.01.011]
- Malone ER, Oliva M, Sabatini PJB, Stockley TL, Siu LL. Molecular profiling for precision cancer therapies. Genome Med 2020; 12: 8 [PMID: 79 31937368 DOI: 10.1186/s13073-019-0703-1]
- Chawla A, Janku F, Wheler JJ, Miller VA, Ryan J, Anhorn R, Zhou Z, Signorovitch J. Estimated Cost of Anticancer Therapy Directed by 80 Comprehensive Genomic Profiling in a Single-Center Study. JCO Precis Oncol 2018; 2 [PMID: 32913996 DOI: 10.1200/PO.18.00074]
- Zhang Q, Fu Q, Bai X, Liang T. Molecular Profiling-Based Precision Medicine in Cancer: A Review of Current Evidence and Challenges. 81 Front Oncol 2020; 10: 532403 [PMID: 33194591 DOI: 10.3389/fonc.2020.532403]
- 82 Subbiah V, Kurzrock R. Challenging Standard-of-Care Paradigms in the Precision Oncology Era. Trends Cancer 2018; 4: 101-109 [PMID: 29458960 DOI: 10.1016/j.trecan.2017.12.004]
- 83 Cappello F, Conway de Macario E, Marasà L, Zummo G, Macario AJ. Hsp60 expression, new locations, functions and perspectives for cancer diagnosis and therapy. Cancer Biol Ther 2008; 7: 801-809 [PMID: 18497565 DOI: 10.4161/cbt.7.6.6281]
- Soga S, Akinaga S, Shiotsu Y. Hsp90 inhibitors as anti-cancer agents, from basic discoveries to clinical development. Curr Pharm Des 2013; 84 19: 366-376 [PMID: 22920907 DOI: 10.2174/138161213804143617]
- Sidera K, Patsavoudi E. HSP90 inhibitors: current development and potential in cancer therapy. Recent Pat Anticancer Drug Discov 2014; 9: 85 1-20 [PMID: 23312026]
- Cheeseman MD, Chessum NE, Rye CS, Pasqua AE, Tucker MJ, Wilding B, Evans LE, Lepri S, Richards M, Sharp SY, Ali S, Rowlands M, 86 O'Fee L, Miah A, Hayes A, Henley AT, Powers M, Te Poele R, De Billy E, Pellegrino L, Raynaud F, Burke R, van Montfort RL, Eccles SA, Workman P, Jones K. Discovery of a Chemical Probe Bisamide (CCT251236): An Orally Bioavailable Efficacious Pirin Ligand from a Heat Shock Transcription Factor 1 (HSF1) Phenotypic Screen. J Med Chem 2017; 60: 180-201 [PMID: 28004573 DOI: 10.1021/acs.jmedchem.6b01055]
- Salamanca HH, Antonyak MA, Cerione RA, Shi H, Lis JT. Inhibiting heat shock factor 1 in human cancer cells with a potent RNA aptamer. 87 PLoS One 2014; 9: e96330 [PMID: 24800749 DOI: 10.1371/journal.pone.0096330]
- Nakamura Y, Fujimoto M, Hayashida N, Takii R, Nakai A, Muto M. Silencing HSF1 by short hairpin RNA decreases cell proliferation and 88 enhances sensitivity to hyperthermia in human melanoma cell lines. J Dermatol Sci 2010; 60: 187-192 [PMID: 21044828 DOI: 10.1016/j.jdermsci.2010.09.009]
- 89 Chen W, Wang J, Shao C, Liu S, Yu Y, Wang Q, Cao X. Efficient induction of antitumor T cell immunity by exosomes derived from heatshocked lymphoma cells. Eur J Immunol 2006; 36: 1598-1607 [PMID: 16708399 DOI: 10.1002/eji.200535501]



WJGO | https://www.wjgnet.com



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: office@baishideng.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

