Online Submissions: http://www.wjgnet.com/1948-5204office wjgo@wjgnet.com doi:10.4251/wjgo.v2.i1.19

World J Gastrointest Oncol 2010 January 15; 2(1): 19-30 ISSN 1948-5204 (online) © 2010 Baishideng. All rights reserved.

TOPIC HIGHLIGHT

Antonio Macrì, MD, Professor, Series Editor

Pharmacologic rationale for treatments of peritoneal surface malignancy from colorectal cancer

Paul H Sugarbaker, Kurt Van der Speeten, O Anthony Stuart

Paul H Sugarbaker, O Anthony Stuart, Washington Cancer Institute, Washington Hospital Center, Washington, DC 20010, United States

Kurt Van der Speeten, Department of Surgical Oncology, Ziekenhuis Oost-Limburg, Schiepse Bos 6, 3600 Genk, Belgium Author contributions: Sugarbaker PH, Stuart OA and Van der Speeten K performed the research; Sugarbaker PH, Stuart OA and Van der Speeten K analyzed the data; Sugarbaker PH, Stuart OA and Van der Speeten K wrote the paper.

Correspondence to: Paul H Sugarbaker, MD, FACS, FRCS, Washington Cancer Institute, Washington Hospital Center, 106 Irving St., NW, Suite 3900, Washington, DC 20010,

United States. paul.sugarbaker@medstar.net
Telephone: +1-202-8773908 Fax: +1-202-8778602
Received: July 2, 2009 Revised: August 28, 2009

Accepted: September 5, 2009 Published online: January 15, 2010 expose all peritoneal surfaces is crucial to success.

© 2010 Baishideng. All rights reserved.

Key words: Intraperitoneal chemotherapy; 5-fluorouracil; Doxorubicin; Mitomycin C; Peritoneal-plasma barrier; Appendiceal cancer; Colorectal cancer; Peritoneal mesothelioma; Ovarian cancer

of carrier solution, based on body surface area, allows

prediction of systemic drug exposure and systemic

toxicity. Timing of the hyperthermic chemotherapy as

a scheduled part of the surgical procedure to uniformly

Peer reviewer: Michael J Wargovich, PhD, Professor of Cell and Molecular Pharmacology, Director, Cancer Chemoprevention Program Hollings Cancer Center, HO712H, Medical University of South Carolina, 86 Jonathan Lucas Street, PO Box 250955, Charleston, SC 29425, United States

Sugarbaker PH, Van der Speeten K, Stuart OA. Pharmacologic rationale for treatments of peritoneal surface malignancy from colorectal cancer. *World J Gastrointest Oncol* 2010; 2(1): 19-30 Available from: URL: http://www.wjgnet.com/1948-5204/full/v2/i1/19.htm DOI: http://dx.doi.org/10.4251/wjgo.v2.i1.19

Abstract

The peritoneal surfaces of the abdomen and pelvis are important sites for the dissemination of gastrointestinal and gynecologic malignancy. Transcoelomic dissemination of cancer cells gives rise to carcinomatosis, which, without special treatment, is a fatal manifestation of these diseases. To treat peritoneal carcinomatosis, cytoreductive surgery removes gross disease plus perioperative intraperitoneal and perioperative intravenous chemotherapy eradicates microscopic residual disease and chemical compatibilities. Chemotherapy agents are administered either by the intraperitoneal or intravenous route, based on their pharmacologic properties. A peritoneal-plasma barrier, which retards the clearance of high molecular weight chemotherapy from the peritoneal cavity, results in a large exposure of small cancer nodules on abdominal and pelvic surfaces. Tissue penetration of the intraperitoneal chemotherapy is facilitated by moderate hyperthermia (41-42 $^{\circ}$). Targeting of intravenous chemotherapy to the peritoneal surface is facilitated by the intraperitoneal heat. A constant dose of chemotherapy agent and volume

INTRODUCTION

The peritoneal surface remains a prominent failure site for patients with gastrointestinal and gynecological cancer. In addition to the lymphatic and hematogenous routes of dissemination, transcoelomic spread of malignant cells is an acknowledged phenomenon that ultimately gives rise to peritoneal carcinomatosis (PC). This intraperitoneal spread can occur before surgery as a direct consequence of full-thickness invasion of the involved organ by tumor and exfoliation of cancer cells into the peritoneal cavity. In addition, intraperitoneal spread might be the result of surgical trauma that causes release of tumor cells from



transected lymph and blood vessels and manipulation of the primary tumor. A review of 2756 patients by Jayne *et al*^[1] reported the incidence of PC at the time of initial surgery to be 7.7%. Brodsky *et al*^[2] suggested that in colonic cancer patients who have recurrences, peritoneal seeding occurred in 25%-35% of patients.

NATURAL HISTORY OF PERITONEAL CARCINOMATOSIS

Nevertheless, little was done to clarify the impact of peritoneal seeding upon survival until the report of Chu and colleagues was published^[3]. These investigators studied 100 patients with non-gynecologic malignancy who had biopsy-proven peritoneal carcinomatosis. The mean survival of 45 colorectal cancer patients was 8.5 mo. The presence or absence of ascites was an important poor prognostic variable in these patients.

In 2000, Sadeghi and coworkers reported on 370 patients with peritoneal carcinomatosis from non-gyne-cologic malignancies who were enrolled in a European prospective multicenter trail [Evolution of Peritoneal Carcinomatosis 1 (EVOCAPE 1)]^[4]. These patients had the benefit of fluorouracil (5-FU)-based systemic chemotherapy, but the results were remarkably similar to those reported by Chu^[3] a decade earlier. The mean survival of 118 patients with carcinomatosis from colorectal cancer was 6.0 mo.

In 2002, Jayne and colleagues from Singapore used a database of 3019 colorectal cancer patients to identify 349 (13%) with peritoneal carcinomatosis^[1]. Of special interest were the 125 patients (58%) who had synchronous primary colorectal cancer and peritoneal implants. The median survival of those patients was only 7 mo. The authors reported that survival was adversely affected by the extent of the peritoneal carcinomatosis and the stage of the primary cancer.

REVISED HYPOTHESIS REGARDING CARCINOMATOSIS AS LOCAL-REGIONAL CANCER DISSEMINATION

More recent chemotherapy, based on the use of oxaliplatin, irinotecan, and biological agents, has improved survival in patients with metastatic colon cancer to 16-20 mo^[5-7]. Unfortunately, none of the trials using modern systemic chemotherapy provide data of results in patients with isolated PC, and only address results in a collection of patients with metastatic colonic cancer at many different anatomical sites. In the past, oncologists have assumed that PC is equal to distant metastases and as such regarded it as beyond current treatment modalities. Over the last two decades a group of oncologists have worked using a revised hypothesis. Novel therapeutic approaches have emerged for patients with isolated peritoneal metastases of gastrointestinal cancer, ovarian cancer, and primary peritoneal malignancies. Early investigators considered PC as a local-regional disease warranting a localregional chemotherapy (intraperitoneal) treatment. Spratt *et al*^[8] reported for the first time in 1980 the use of heated triethylenethiophophoramide (thiotepa) in a patient with pseudomyxoma peritonei^[8]. Koga *et al*^[9] reported the use of intraperitoneal chemotherapy in 23 gastric cancer patients with PC. Speyer in 1980 used normothermic intraperitoneal 5-fluorouracil and methotrexate in 16 patients with PC^[10].

ROLE FOR CYTOREDUCTIVE SURGERY

A second essential component of the modern management of PC is cytoreductive surgery with peritonectomy procedures. The rationale for a combined approach is as follows: an aggressive surgical approach combining visceral resections and peritonectomy procedures should address the macroscopic peritoneal surface disease; the perioperative intraperitoneal chemotherapy treats any residual microscopic disease^[11]. The perioperative intraperitoneal chemotherapy includes hyperthermic intraperitoneal chemotherapy (HIPEC) and/or early postoperative intraperitoneal chemotherapy (EPIC). More recent protocols advocate bidirectional (simultaneous intraperitoneal and intravenous chemotherapy) intraoperative chemohyperthermia. This two-component approach to PC treatment requires that chemotherapy be used as a planned part of the surgical procedure. Critical to the success is proper timing of the chemotherapy in its relation to the surgery. This innovation of combined treatment of cytoreductive surgery plus perioperative intraperitoneal chemotherapy might be the paradigm shift responsible for recent successes vs prior failures in treating PC patients.

RESULTS IN TREATING CARCINOMATOSIS FROM COLORECTAL CANCER

Several phase II studies have explored the intraperitoneal route of drug delivery and show promising results in a variety of PC patients. In colorectal cancer with PC, overall survival rates between 25% and 47% are reported[12-17]. These reports, from many different institutions, stand in strong contrast with historical control groups and patients treated with systemic chemotherapy, where the mean and median overall survival were 6.0 and 3.1 mo, respectively^[4]. Verwaal et al^[18] in a phase III trial randomized treated patients with colorectal PC by palliative surgery followed by fluorouracil-leucovorin systemic chemotherapy vs maximal cytoreduction plus HIPEC with mitomycin C and subsequent systemic chemotherapy. This report showed a significant survival benefit for the HIPEC arm, with a median survival of 22.3 mo vs 12.6 mo in the control group. Glehen et al [19] collected data retrospectively on 506 PC patients from 28 institutions treated with cytoreductive surgery and perioperative intraperitoneal chemotherapy. He reported an overall median survival of 19.2 mo. Patients in whom cytoreduction was complete had a median survival of

32.4 mo vs 8.4 mo in the patients with incomplete cytoreduction.

The clinical evidence in the medical literature supporting the combined approach of cytoreductive surgery and perioperative intraperitoneal chemotherapy is growing [20-24]. What is lacking is clear understanding of the pharmacology of the perioperative use of chemotherapy in the treatment of peritoneal surface malignancy. It is possible that increased safety and important treatment innovations might originate from analyzing the pharmacologic data. This review aims to clarify the pharmacokinetic and pharmacodynamic data currently available regarding the intraperitoneal delivery of cancer chemotherapy agents in patients with PC.

THE PERITONEAL PLASMA BARRIER

The rationale for administering chemotherapeutic drugs directly into the peritoneal cavity is supported by the relative transport barrier that is formed by the tissue surrounding the peritoneal space. The peritoneum is a complex three-dimensional organ covering the abdomino-pelvic organs and the abdominal wall. It contains a large potential space. The most elaborate description of the ultra structure of the peritoneum in man goes back to 1941 by Baron et al²⁵. The peritoneum consists of a monolayer of mesothelial cells supported by a basement membrane and five layers of connective tissue, which account for a total thickness of 90 µm. The connective tissue layers include interstitial cells and a matrix of collagen, hyaluron, and proteoglycans. The cellular component consists of pericytes, parenchymal cells, and blood capillaries. The complex is often referred to as the peritoneal membrane. This description is a working model derived from research regarding the peritoneum as a dialysis membrane.

The accepted function of the peritoneum is twofold. First, it reduces friction between intraabdominal organs and the abdominal wall by producing a lubricant solution made of glycosaminoglycans and phospholipids^[26]. Secondly, it is of major importance, together with lymphoid aggregates dispersed on the visceral and parietal peritoneum, in the host defense against intraabdominal infections. A third suggested function of the peritoneum in malignancy might be its role as a first line of defense against peritoneal carcinomatosis^[27]. Any disruption in the peritoneal lining facilitates a transition from adhesion to invasion of cancer cells, resulting in the development of peritoneal tumor nodules on the abdominal or pelvic surface^[27,28].

Contrary to intuitive thinking, the elimination of the mesothelial lining, as performed during peritonectomy procedures, does not significantly alter the pharmacokinetic properties of the peritoneum in the transport of chemotherapeutic agents from the peritoneal cavity to the plasma compartment. Flessner *et al*^[29] demonstrated in a rodent model that neither removal of the stagnant fluid layer on the mesothelium nor removal of the mesothelial lining influenced the mass transfer

coefficient over the barrier. Evidence supporting this hypothesis in humans is supported by the fact that the extent of the peritonectomy in PC patients does little to alter the intraperitoneal chemotherapy pharmacokinetics of mitomycin C or 5-fluorouracil^[30,31]. The current thinking regards not only the mesothelial lining, but also the blood capillary wall and the surrounding interstitial matrix as the principal barrier for clearance of molecules from the abdominopelvic space^[32].

Much of the laboratory research concerning the pharmacokinetic properties of the peritoneum is derived from the peritoneal dialysis literature^[33]. A simplified mathematical diffusion model considers the plasma to be a single compartment separated by an effective membrane from another single compartment, the peritoneal cavity. Although this offers a conceptual model of transport and states the importance of the effective exposure area, it only offers quantitative predictability once permeability is empirically determined for each drug. It also does not offer insight into the actual tissue penetration at the level of the peritoneal membrane. Neither does it predict penetration of chemotherapy into the tumor nodules, which is the single most important factor determining response to cancer treatment.

PHARMACOKINETIC RATIONALE OF PERIOPERATIVE INTRAPERITONEAL CANCER CHEMOTHERAPY

Intraperitoneal administration of chemotherapeutic agents gives high response rates in PC patients because retention of chemotherapy in the peritoneal plasma barrier provides dose-intensive therapy. Dedrick et al^[34] in 1978 concluded that the peritoneal permeability of a number of hydrophilic anticancer drugs might be considerably slower than the plasma clearance of that same drug. This results in a significantly greater concentration in the peritoneal cavity, as compared to the plasma, after intraperitoneal administration. This concentration difference causes increased exposure of small residual tumor cells after cytoreduction to high doses of chemotherapeutic agents, with reduced systemic concentrations and lower systemic toxicity. This advantage is expressed by the Area Under the Curve (AUC) ratios of intraperitoneal vs plasma exposure. Table 1 presents molecular weight and AUC ip/iv for drugs in clinical or experimental use in PC patients^[35].

An important consideration is that high intraperitoneal concentration or AUC ip/iv does not automatically confer a greater efficacy. Even with greatly elevated intraperitoneal cancer chemotherapy concentrations, there may be limited penetration of the chemotherapeutic agent into the peritoneal tumor target. The ideal drug for intraperitoneal chemotherapy has not only a high peritoneal tissue concentration as a result of direct intraperitoneal administration, but also a high penetration into the cancer nodule. Retention of cancer chemotherapy in preperitoneal tissue and cancer nodules is augmented by slow diffusion through the capillary endothelium, deep

Table 1 Molecular weight and area under the curve ratios of intraperitoneal exposure to systemic exposure of chemotherapeutic agents used to treat peritoneal carcinomatosis

Drug	Molecular weight (Daltons)	Area under the curve ratio
5-fluorouracil	130.08	250
Carboplatin	371.25	10
Cisplatin	300.1	7.8
Docetaxel	861.9	552
Doxorubicin	579.99	230
Etoposide	588.58	65
Floxuridine	246.2	75
Gemcitabine	299.5	500
Irinotecan	677.19	N/A
Melphalan	305.2	93
Mitomycin C	334.3	23.5
Mitoxantrone	517.41	115-255
Oxaliplatin	397.3	16
Paclitaxel	853.9	1000
Pemetrexed	597.49	40.8

N/A: Not available.

in the subperitoneal space of the cancer chemotherapy solution. Low systemic concentrations and reduced systemic toxicity are maintained by rapid metabolism and excretion of drug within the body compartment.

The marked increase in exposure of peritoneal surfaces to chemotherapy solution, as compared to plasma, is illustrated in Figure 1. The chemotherapy agent, paclitaxel, has a high molecular weight (853.9 Daltons) and is slow to cross the peritoneal cavity to plasma barrier. The naked molecule of paclitaxel is highly lipophilic. It is unique in cancer chemotherapy in that the intravascular or intraperitoneal administration of the drug requires it to be suspended in a detergent that maintains the drug in solution. The detergent molecules surround the paclitaxel molecule giving it additional size and a hydrophilic character. This large molecular size and hydrophilic behavior of the complex molecule result in slow passage across the peritoneal to plasma barrier. The AUC ratio for paclitaxel is approximately 1000 (Table 1).

PHARMACOKINETIC RATIONALE OF PERIOPERATIVE INTRAVENOUS COLORECTAL CANCER CHEMOTHERAPY

New pharmacological data suggests altered pharmacodynamics of intravenously administered cancer chemotherapy drugs when used intraoperatively during a HIPEC procedure. Figure 2 shows 5-fluorouracil concentrations in the plasma, peritoneal fluid, and tumor nodules after intravenous administration at the beginning of the HIPEC procedure. Almost immediately after intravenous administration, the 5-fluorouracil is transported from the plasma compartment to the expanded peritoneal cavity. There it is retained in the artificial ascites created by the HIPEC for a substantial amount of time, before reabsorption into the systemic compartment occurs. The metabolism of 5-fluorouracil

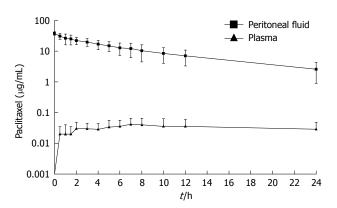


Figure 1 Pharmacokinetic study of concentration vs time for intraperitoneal paclitaxel. The chemotherapy agent at 30 mg/m^2 was instilled directly into the peritoneal cavity as rapidly as possible in a 1.5% dextrose peritoneal dialysis solution. The concentration of paclitaxel was determined in peritoneal fluid and in plasma for 24 h.

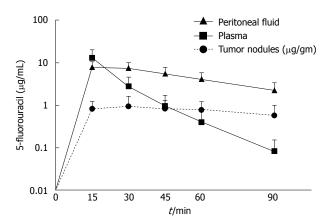


Figure 2 Pharmacodynamics during HIPEC after intravenous administration of 400 mg/m² of 5-fluorouracil given simultaneously with intraperitoneal chemotherapy in 3 L of chemotherapy solution. (Van der Speeten K, Stuart OA, Sugarbaker PH. Pharmacology of perioperative 5-fluorouracil. Submitted for publication).

in the ascites fluid is greatly reduced as compared to metabolism in the plasma. Our data suggests that the artificial ascites created by the HIPEC provides a reservoir for the intravenously administered drug. The intraoperative administration of intravenous cancer chemotherapy might offer a pharmacologic advantage in killing residual tumor cells after cytoreduction. Timing of intravenous cancer chemotherapy (intraoperative *w* preor postoperative) emerges as a new variable, which could affect efficacy of cancer chemotherapy drugs in treating patients with a peritoneal surface malignancy.

Tissue distribution and penetration depth

The simplified two-compartment model described by Dedrick and colleagues might not provide an adequate theoretical model for penetration of the intraoperatively administered (either intravenous or intraperitoneal) chemotherapy into the preperitoneal tissues and into the tumor nodules. Dedrick *et al*^{36,371} proposed a mathematical model (Figure 3) addressing the tissue penetration of low-molecular weight molecules. The drug diffuses from its peritoneal



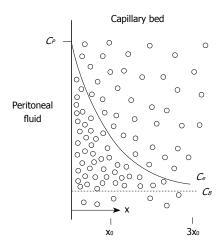


Figure 3 Conceptual diagram of tissue adjacent to the peritoneal cavity. $C_{\it p}$ = the free drug concentration in the peritoneal fluid; $C_{\it B}$ = the free drug concentration in the blood (or plasma). Solid line shows the exponential decrease in the free tissue interstitial concentration, $C_{\it B}$, as the drug diffuses down the concentration gradient and is removed by loss to the blood perfusing the tissue. Also shown are the characteristic diffusion length, x_0 , at which the concentration difference between the tissue and the blood has decreased to 37% of its maximum value, and $3x_0$, at which the difference has decreased to 5% of its maximum value^[36].

concentration, C_P , to its blood concentration, C_B , along an exponential concentration gradient over the peritoneum and preperitoneal tissues. The extracellular "deep" concentration, C_e , can then be calculated according to the formula: $C_e = C_B + (C_P - C_B) \exp[-(k/D)^{1/2}x]$. In this formula, k (min⁻¹) is the rate constant for removal of the active drug from the tissue. Movement through the tissue is characterized by the diffusivity, D (cm²/min) and x is the distance from the serosal surface (cm). This model implies that there is an exponential concentration decrease of the drug from abdominopelvic cavity across the membrane to the plasma compartment. Consequently, the depth of penetration of the intraperitoneal concentration is very limited and is in the order of 1 to 2 mm^[38,39]. Ozols et al^{40]} confirmed adriamycin penetrating only four to six cell layers of a tumor on the diaphragm in a rodent model. In all likelihood, there is variable penetration for each drug and type of tumor.

This has important consequences for implementing perioperative chemotherapy in PC patients. The cytoreduction needs to resect all tumor deposits greater than 1-2 mm for the subsequent intraperitoneal chemotherapy to be effective. Clinical data to support this pharmacologic prediction is strong; in univariate and multivariate analysis, complete cytoreduction (cancer nodules ≤ 2.5 mm) is the single most important prognostic factor [15-19]. To describe the pharmacology of both intravenous and intraperitoneal chemotherapy used after cytoreduction to treat PC, a revised theoretical model is needed. Figure 4 shows a threecompartment model constructed of a body compartment, intermediate compartment (peritoneal and preperitoneal tissues with tumor nodules) and peritoneal fluid. The movement of chemotherapy molecules from blood to peritoneal fluid through the intermediate compartment is rapid because of the extensive arterial and portal venous blood delivery to the intermediate tissues. The movement

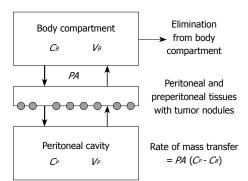


Figure 4 Three-compartment model of peritoneal transport, in which transfer of a drug from the peritoneal cavity to the blood occurs across the peritoneal membrane and preperitoneal tissues. The peritoneal surface cancer nodules are located in these tissues. The permeability-area product (PA) governs this transfer and can be calculated by measuring the rate of drug disappearance from the cavity and dividing by the overall concentration difference between the peritoneal cavity and the blood (B). C_B = the free drug concentration in the blood (or plasma); V_B = volume of distribution of the drug in the body; C_P = the free drug concentration in the peritoneal fluid; V_P = volume of the peritoneal cavity^[36].

of chemotherapy molecules from peritoneal fluid back to portal blood would be less rapid.

There are a large number of variations in intraperitoneal chemotherapy treatment protocols. All these variations reflect attempts to improve diffusivity D, decrease the rate constant k, permeability P, or effective membrane area A. A non-exhaustive list of factors influencing these values, and thus the response of the peritoneal metastases to perioperative chemotherapy, is listed in Table 2. Some of these will be explored in greater detail.

TEMPERATURE

Adding hyperthermia to intraperitoneal chemotherapy might increase the tumor response to cancer chemotherapy drugs by several mechanisms. First, heat alone has a direct anti-tumor effect. Hyperthermia above 41°C induces selective cytotoxicity of malignant cells. Several mechanisms for this effect have been proposed: impaired DNA repair, protein denaturation, and inhibition of oxidative metabolism in the microenvironment of malignant cells, leading to increased acidity, lysosomal activation, and increased cell death^[41,42]. Cells are known to react to protein denaturation by up-regulation of heat shock proteins. This induces the development of thermal tolerance in tumor cells^[43]. Thermal tolerance might cause the clinical importance of hyperthermia to be limited. Second, the cytotoxic effects of some chemotherapeutic agents are augmented by applying mild hyperthermia. Such augmented effects were postulated for doxorubicin^[44], platinum complexes^[45,46], mitomycin $C^{[46]}$, melphalan^[47], docetaxel, irinotecan, and gemcitabine^[48]. Third, this increased response might be the consequence of an increased penetration depth of the chemotherapeutic agent^[49,50].

Jacquet *et al*^{50]} studied the changes in the penetration of intraperitoneal chemotherapy that are induced by moderate hyperthermia. The heat utilized within the peritoneal cavity

Table 2 Variables influencing the response of peritoneal carcinomatosis to perioperative chemotherapy

Temperature	Increase in temperature above 37°C augments the cytotoxicity of cancer chemotherapy
Dose of intraperitoneal chemotherapy	As the dose increases the penetration of chemotherapy into cancerous tissue increases because of an increasing
	diffusion gradient
Distribution of chemotherapy solution	The open technique allows more uniform distribution of heat and chemotherapy solution because of the
and heat (open vs closed technique)	manual mixing of the abdominal and pelvic contents with the warm fluid
Timing of chemotherapy in relation to	Using the chemotherapy with surgery allows complete distribution immediately after the total lysis of
the timing of the surgical intervention	abdominal adhesions. This allows complete treatment of all peritoneal surfaces
Type of carrier solution	High molecular weight carrier solutions remain within the peritoneal cavity for a longer time period. The
	artificial ascites maintains the cancer chemotherapy in a large volume of fluid for an extended time period
Pressure	Pressure will increase the penetration of fluid and chemotherapy solution into normal and cancerous tissue
Volume of carrier solution	Increasing the volume of carrier solution without increasing the amount of chemotherapy will decrease
	the effectiveness of the treatment by lowering the diffusion gradient between the peritoneal space and the
	surrounding normal and cancerous tissue
Duration of instillation	Increasing the time period over which cancer chemotherapy is present within the peritoneal cavity will increase
	the cytotoxic effect
Vasoactive agents	Vasoactive agents will cause constriction of normal capillaries, but will not cause constriction of vessels within
	cancerous tissue. This will cause the chemotherapy to remain longer in the peritoneal space
Macromolecular vehicles	Coating of cancer chemotherapy by macromolecules may preferentially direct their entrance into cancerous
	tissue as compared to normal tissue
Drug sensitivity of the tumor	Increased responses are expected if the cancer is sensitive to the chemotherapy
Size of residual tumor nodules	The penetration of cancer chemotherapy is limited to approximately 1 mm. Therefore, large nodules greater
	than 1 or 2 mm in diameter will not be penetrated by intraperitoneal chemotherapy and should not be expected
	to be eliminated

was 41.5°C to 42°C in this rat model. Large and statistically significant increases in the amount of doxorubicin within tissues were demonstrated. The tissues in direct contact with the intraperitoneal chemotherapy were most affected. The concentration of doxorubicin in small bowel was doubled using intraperitoneal doxorubicin plus heat; a similar finding occurred for omental tissue and bladder tissue. Tissues at a distance such as heart muscle, did not show an increased concentration with heat.

PRESSURE

Dedrick *et al*³⁶ postulated that the penetration distance is equal to the square root of the ratio of the tissue diffusivity and the rate constant for drug removal from the tissue (D/k)^{1/2}. Unpublished observations by Flessner in a rat model showed a doubling of the extracellular space in the anterior abdominal wall of rats when the pressure of intra-abdominal peritoneal dialysis solution was raised from 0 to 4 cmH₂O^[29]. An increased effective diffusivity was postulated.

Animal experiments^[51,52] confirmed the increased intratumoral accumulation and antitumor effect of intraperitoneal doxorubicin and cisplatin when the intraabdominal pressure was raised. Increased intraabdominal pressure is thought to generate a convective flux that forces the drug from the peritoneal cavity into the subperitoneal tissue. At the same time, intraabdominal pressure might counteract the hydraulic capillary pressure and slow the outflow of the drug to the body compartment. Measurement of local cisplatin concentrations along the radii of peritoneal tumor nodules showed platinum penetration far beyond the 1 mm limit advocated by Los *et al*^[39]. The clinical limit of usable intraabdominal pressure enhancement is dictated by respiratory and hemodynamic

tolerance. Clinical applications of HIPEC in intraabdominal pressure settings so far has been limited to palliating debilitating malignant ascites with laparoscopic HIPEC at 10-15 mmHg^[53,54].

CARRIER SOLUTION

A variety of carrier solutions have been applied in different treatment protocols. Hypotonic, isotonic, and hypertonic solutions were explored with both low and high molecular weight chemotherapy molecules. The ideal carrier solution should enhance the exposure of the peritoneal surface and residual tumor cells to the chemotherapeutic agent. This is especially important in the setting of EPIC where maintenance of a high dwell volume of perfusate over a prolonged time period improves the distribution of the drug and the effectiveness of the treatment. Chemotherapy solutions of isotonic and hypertonic salt, dextrose, hetastarch, or icodextrin solutions have been explored. In an EPIC setting a high molecular weight solution that maintains artificial ascites showed higher drug availability because the total drug exposure depends not only on the active drug, but also on the carrier solution that maintains the peritoneal fluid volume^[55-57].

In a HIPEC setting with a relatively short dwell time, one could theoretically expect a pharmacodynamic advantage of a hypotonic carrier through the mechanism of increased tissue and tumor absorption. Contrary to experimental studies supporting this hypothesis, Elias *et al*⁵⁸ showed no increase in tumor penetration in humans. A concomitant high incidence (50%) of postoperative peritoneal bleeding and severe thrombocytopenia has contraindicated the further clinical use of hypotonic carriers.

VOLUME OF CHEMOTHERAPY SOLUTION

Peritoneal metastases and free-floating tumor cells can be present anywhere on the peritoneal surface; therefore the entire surface of the abdominal and pelvic cavity is the target. Substantial differences in body composition of patients and differences in the actual HIPEC technique (open vs closed abdomen) will result in a wide variety of perfusate volumes. In current practice, the volume of the perfusate is chosen quite arbitrarily. Following the above stated equitation concerning mass transfer over the peritoneal-plasma membrane, increasing the solution contact area A improves the mass transfer. Keshaviah demonstrated a linear rise in mass transfer in 10 patients who were dialyzed with different volumes ranging from 0.5 up to 3 L^[59]. Elias first published the importance of volume of chemotherapy in determining systemic exposure to the drug^[60]. Sugarbaker et al^[61] carried out a clinical investigation where 2 L vs 4 L vs 6 L of chemotherapy solution were administered. The dose of chemotherapy solution in these studies was constant. They showed that a more dilute intraperitoneal chemotherapy concentration retarded the clearance of chemotherapy and lead to a lesser systemic toxicity. In addition, it must be assumed that the less concentrated chemotherapy would, by the diffusion model, penetrate less into the cancer nodules and into normal tissues. These authors determined it necessary to regulate not only the chemotherapy dose but also the volume of chemotherapy solution by the patient's body surface area.

A consistent drug dose and chemotherapy solution volume might be the optimal method to predict a maximal treatment in the abdomen with predictable bone marrow toxicity. Sugarbaker and colleagues suggested that variable volume is a dangerous practice with unpredictable systemic toxicities^[61]. If a chemotherapy solution is administered until the abdomen is full, the contact area will be variable because there is a great difference in the size of the peritoneal space for different patients. If the contact area is variable the total absorption of the drug cannot be predicted.

VASOACTIVE AGENTS

The literature concerning the effects of vasoactive substances in regulating peritoneal blood flow and tumor blood flow is extensive. These agents might contribute to a delayed clearance from the peritoneal cavity, because the blood flow in the peritoneal and subperitoneal vascular network will control, to a large extent, the movement of molecules across the peritoneal and subperitoneal tissues. General statements regarding the effects of vasoactive agents are confusing and sometimes contradictory, due to the variety of experimental systems, complex interactions of local-regional and systemic effects of vasopressive agents, and large differences between the neovasculature of tumor nodules and normal capillaries. Both intravenous and intraperitoneal administration of vasoactive molecules in combination with chemotherapeutic drugs has been

explored [36,62,63]. A preclinical study of the use of an intraperitoneal epinephrine plus intraperitoneal cisplatin in a rat model with PC, showed a direct correlation between the intraperitoneal epinephrine concentration and cisplatin accumulation in rat peritoneal tumor nodules^[64]. Recently, Molucon-Chabrot et al^[65] demonstrated for the first time the safe use of intraperitoneal epinephrine with intraperitoneal cisplatin in 18 patients with advanced peritoneal carcinomatosis. Tumor responses were obtained in some patients that were resistant to intravenous platinum compounds. Lindnér et all66 reported that concurrent intravenous administration of vasopressin can increase the pharmacokinetic advantage of intraperitoneal administered carboplatin and etoposide, but not 5-FU. Further studies on the use of vasoactive agents to improve cancer chemotherapy responses in PC are required.

DOSE OF INTRAPERITONEAL CANCER CHEMOTHERAPY

Many different chemotherapy agents, drug concentrations, drug doses, and drugs schedules have evolved at many institutions for perioperative cancer chemotherapy. Most authors use a drug dose based on calculated body surface area (mg/m²), although Rubin *et al*^{67]} demonstrated that there is an imperfect correlation between actual peritoneal surface area and calculated body surface area. The same study suggests sex differences in peritoneal surface areas that in turn might affect the absorption characteristics. Females have a larger peritoneal surface in proportion to body size than males by approximately 10%. Estimates of the functional peritoneal surface area by applying stereological methods to CT scans have been attempted^[68].

Body surface area is an accurate predictor of drug metabolism and in this regard is useful for estimating systemic drug toxicity. As discussed by Sugarbaker *et al*^[61], the accuracy of this prediction is increased if the volume of chemotherapy solution is also determined by the body surface area. With a constant total dose of chemotherapy and chemotherapy solution, the bone marrow exposure to cytotoxic drugs can be most accurately predicted. If these predictions are not available, the danger of overdosing some patients and underdosing others will occur.

A significant number of institutions use a closed method for intraoperative hyperthermic chemotherapy administration. In this technique, the dose of cancer chemotherapy is calculated per liter by body surface area. The total amount of cancer chemotherapy is mixed in a large volume of carrier solution (usually 6 L) that is placed in a reservoir. For example, Deraco *et al*^[69,70] have used doxorubicin at 15.25 mg/m² per liter and cisplatin at 43 mg/m² per liter with the total volume of 6 L. Gilly and coworkers have used mitomycin C at 0.5 mg/kg and cisplatin at 0.7 mg/kg in a total volume of 4 to 6 L^[71,72]. In this method, the amount of chemotherapy solution in contact with the peritoneal surface is determined by multiple variables, in addition to the body surface area; the amount of distention (between 2 and 6 L) of the



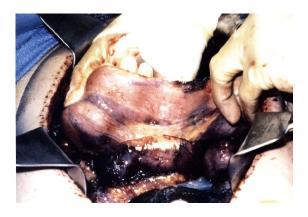


Figure 5 Spatial distribution of intraperitoneal methylene blue, using the closed abdomen technique. Although the subcutaneous tissues were uniformly stained, the small bowel loops showed variable staining caused by adherence of adjacent bowel loops.

abdominal cavity induced by the chemotherapy solution, the patient's sex, the amount of ascites present preoperatively and the extent of the visceral resection. This means that the peritoneal surface in contact with the chemotherapy solution is variable; consequently, the systemic absorption and the plasma concentration of the chemotherapy are variable. In this situation the systemic toxicity is not predictable.

In summary, a system that allows a variable amount of chemotherapy solution to contact peritoneal surfaces might result in a less accurate prediction of plasma AUC, because the total volume of intraperitoneal chemotherapy can vary widely betweens individuals. Increases in the volume of intraperitoneal chemotherapy solution cause an increase in diffusion surface and an increase in the amount of drug moving from peritoneal space to plasma.

Distribution of intraperitoneal chemotherapy

Both open and closed intraperitoneal chemotherapy methods of administration have been described^[73-77]. The closed abdomen techniques have the theoretical advantage of less heat loss during perfusion. Also, a reduction in environmental risk for the operative team has been suggested. However, to date no study has been able to detect any occupational risk for the staff^[78-80]. Open abdomen techniques will provide a better spatial distribution of the chemotherapy solution over the entire peritoneal surface. In addition, a more uniform heat distribution is expected if the chemotherapy solution is manually stirred throughout the HIPEC. Elias et al^[76] performed a prospective phase I - II study comparing seven HIPEC perfusion techniques. Judgments regarding adequate spatial distribution were made by measurement of temperature at multiple sites. Using the closed system, temperature varied greatly throughout the abdomen and pelvis. The temperature became more uniform with an open abdomen and manual distribution. Spatial diffusion, studied by adding methylene blue to the perfusate, was incomplete in the closed abdomen techniques (Figure 5). However, these differences in the delivery of HIPEC might be more theoretical than real. In studies to date, only the extent of PC, the absence of extraperitoneal disease, and the completeness of cytoreduction correlated with survival. Comparison of data from centers performing either closed or open abdomen techniques, suggest that the chemotherapy effects by open or closed methods might be similar. Long follow-up will be required to make an accurate comparison.

TIMING OF CANCER CHEMOTHERAPY IN RELATION TO TIMING OF THE SURGICAL INTERVENTION

Neoadjuvant bidirectional chemotherapy

Considering the clinical application of chemotherapy in PC patients, one can intervene at four possible points in the timeline. First, neoadjuvant bidirectional chemotherapy uses both the intraperitoneal and intravenous routes of chemotherapy administration. It has been explored as an option to reduce the extent of small PC nodules. Theoretically, it might facilitate definitive cytoreductive surgery after initial exploratory laparoscopy shows PC. Radiological and clinical responses have been reported by several groups [81-83].

Although this strategy might reduce the tumor load to be addressed by cytoreductive surgery, it has several disadvantages. Adhesions from prior surgical interventions might interfere with adequate intraperitoneal drug distribution. Complete responses are very unusual, so that further cytoreduction-chemotherapy is necessary if the approach is curative. Neoadjuvant intraperitoneal chemotherapy has been reported to add to morbidity and mortality of further surgical treatment^[84]. Extensive fibrosis as a response to chemotherapy might occur and make judgments concerning the extent of peritoneal carcinomatosis difficult, or even impossible, to assess.

Intraoperative intraperitoneal chemotherapy

Intraoperative intraperitoneal chemotherapy has been the most widely explored modality with consistent clinical improved outcomes in many phase II trials and several phase III trials [22-24].

Early postoperative intraperitoneal chemotherapy

Early postoperative intraperitoneal chemotherapy has some conceptual advantages. It is administered after cytoreductive surgery at the time of minimal residual tumor burden. Additionally, intraperitoneal treatments initiated before wound healing occurs can minimize non-uniform drug distribution and eliminate residual cancer cell entrapment in postoperative fibrin deposits. Proper selection of chemotherapy agents based on pharmacologic principles suggests the use of cell-cycle specific drugs such as 5-fluorouracil and the taxanes. Most EPIC regimens are administered postoperatively day 1 to 5 or day 1 to 4 through an inflow catheter and outflow drains placed at the time of cytoreductive surgery. EPIC can be applied with or without HIPEC^[85].



Long-term combined intraperitoneal and systemic chemotherapy

Markman *et al*⁸⁶, Alberts *et al*⁸⁷, and Armstrong *et al*⁸⁸ demonstrated in a phase III trials that intravenous plus intraperitoneal chemotherapy improves survival in patients with optimally debulked stage III ovarian cancer, as compared to intravenous chemotherapy alone. This approach might be used as "chemotherapeutic bridging" between incomplete initial surgery and definitive cytoreduction or second look surgery. This type of chemotherapy is an adjuvant and not a perioperative use of chemotherapy. Failure analysis reported for cytoreductive surgery plus perioperative chemotherapy determined recurrent cancer most frequently occurs within the abdominal and pelvic cavity^[89,90]. To optimize the treatment of patients with PC, it is likely that the greatest benefit will occur from a combination of these four treatment strategies.

DURATION

A wide variety of durations for HIPEC have been reported ranging from 30 to 120 min. The duration is not arbitrary and selection should proceed according to the pharmacologic clearance of the chemotherapeutic drug.

MACROMOLECULAR VEHICLES

In recent years an increased interest in macromolecular vehicles and other modulations of chemotherapeutic agents as a means of exploiting the regional dose intensity has emerged. The results of this research are conflicting. Contrary to intuitive thinking, macromolecules might penetrate more deeply in the subperitoneal space despite their lower diffusivities. The nature of the capillary permeability is probably the major factor responsible for this higher concentration in the subperitoneal space, together with an increased role of convection [91]. One should be cautious in concluding that this increased penetration into the subperitoneal space results in increased drug absorption into tumor nodules. That is, one should not assume that the neovascularity of tumor nodules has the same selectivity for macromolecules as normal capillaries [92]. A second obstacle to cancer chemotherapy penetration into tumor nodules concerns the interstitial pressures in tumor nodules; it is significantly higher than that of the surrounding tissue space^[93]. Convection might reduce tumor penetration by macromolecules.

INDIVIDUAL DRUG SENSITIVITY OF TUMORS WITHOUT AND WITH HYPERTHERMIA

The selection of chemotherapeutic agents used in perioperative chemotherapy protocols has been based on research in chemotherapeutic responses in systemic administration, on pharmacodynamic and pharmacokinetic properties of the drug in intraperitoneal administration, increased cytotoxicity with hyperthermia, and synergy between chemotherapeutic agents. There is solid evidence supporting a tumor-specific heterogeneous activity of cytotoxic drugs in cell cultures of different tumors^[94,95].

Mahteme et al⁹⁶ recently stated the same heterogeneous cytotoxic response of cytotoxic drugs in PC samples in a variety of tumors. The clinical implication of these data justifies further research towards an individualized selection of drugs in PC patients. However, it should be acknowledged that as yet, there is no prospective data supporting an improved clinical outcome from drug selection based on *in-vitro* drug sensitivity testing.

BIDIRECTIONAL INTRAOPERATIVE CHEMOTHERAPY

The three-compartment model described above for peritoneal transport predicts transport by diffusion from the peritoneal compartment through a peritoneal and preperitoneal tissue layer to the plasma. Drugs move from the plasma compartment through the preperitoneal tissue layer to the peritoneal compartment. By combining intraoperative intravenous and intraoperative intraperitoneal cancer chemotherapy, a bidirectional diffusion gradient is created through the intermediate tissue layer that contains the cancer nodules. This offers opportunities for optimizing cancer chemotherapy delivery to the target peritoneal tumor nodules. Elias *et al*⁶⁰ were the first to utilize this approach. Further pharmacologic studies are required to clarify the most effective chemotherapy agents for this bidirectional approach.

CONCLUSION

The administration of perioperative chemotherapy in patients with peritoneal carcinomatosis should be governed by pharmacological principles. Patients who have minimal residual disease as a result of cytoreductive surgery are candidates for perioperative chemotherapy by the intraperitoneal and intravenous route. Hyperthermia of the intraperitoneal chemotherapy solution will increase the cytotoxicity of the drug within the peritoneal cavity. In addition, heating of the peritoneal and preperitoneal tissues will maximize the systemic chemotherapy effects on carcinomatosis, a phenomenon known as heat targeting. Perioperative chemotherapy has become an important part of cancer treatment and should become a standard modality for prevention and treatment of a wide variety of cancers that involve the peritoneal surfaces.

REFERENCES

- Jayne DG, Fook S, Loi C, Seow-Choen F. Peritoneal carcinomatosis from colorectal cancer. Br J Surg 2002; 89: 1545-1550
- Brodsky JT, Cohen AM. Peritoneal seeding following potentially curative resection of colonic carcinoma: implications for adjuvant therapy. *Dis Colon Rectum* 1991; 34: 723-727
- Chu DZ, Lang NP, Thompson C, Osteen PK, Westbrook KC.



- Peritoneal carcinomatosis in nongynecologic malignancy. A prospective study of prognostic factors. *Cancer* 1989; **63**: 364-367
- 4 Sadeghi B, Arvieux C, Glehen O, Beaujard AC, Rivoire M, Baulieux J, Fontaumard E, Brachet A, Caillot JL, Faure JL, Porcheron J, Peix JL, François Y, Vignal J, Gilly FN. Peritoneal carcinomatosis from non-gynecologic malignancies: results of the EVOCAPE 1 multicentric prospective study. *Cancer* 2000; 88: 358-363
- 5 de Gramont A, Figer A, Seymour M, Homerin M, Hmissi A, Cassidy J, Boni C, Cortes-Funes H, Cervantes A, Freyer G, Papamichael D, Le Bail N, Louvet C, Hendler D, de Braud F, Wilson C, Morvan F, Bonetti A. Leucovorin and fluorouracil with or without oxaliplatin as first-line treatment in advanced colorectal cancer. J Clin Oncol 2000; 18: 2938-2947
- 6 Douillard JY, Cunningham D, Roth AD, Navarro M, James RD, Karasek P, Jandik P, Iveson T, Carmichael J, Alakl M, Gruia G, Awad L, Rougier P. Irinotecan combined with fluorouracil compared with fluorouracil alone as first-line treatment for metastatic colorectal cancer: a multicentre randomised trial. *Lancet* 2000; 355: 1041-1047
- 7 Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, Berlin J, Baron A, Griffing S, Holmgren E, Ferrara N, Fyfe G, Rogers B, Ross R, Kabbinavar F. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med 2004; 350: 2335-2342
- 8 Spratt JS, Adcock RA, Muskovin M, Sherrill W, McKeown J. Clinical delivery system for intraperitoneal hyperthermic chemotherapy. *Cancer Res* 1980; 40: 256-260
- 9 Koga S, Shimizu N, Maeta M, Hamazoe R, Izumi A. [Application of heat combined with antineoplastic agent administration in the treatment of cancer (with special reference to malignancy of the digestive system)] Gan To Kagaku Ryoho 1983; 10: 358-365
- 10 Speyer JL, Myers CE. The use of peritoneal dialysis for delivery of chemotherapy to intraperitoneal malignancies. Recent Results Cancer Res 1980; 74: 264-269
- 11 **Sugarbaker PH**. Peritonectomy procedures. *Surg Oncol Clin N Am* 2003; **12**: 703-727, xiii
- 12 Glehen O, Mithieux F, Osinsky D, Beaujard AC, Freyer G, Guertsch P, Francois Y, Peyrat P, Panteix G, Vignal J, Gilly FN. Surgery combined with peritonectomy procedures and intraperitoneal chemohyperthermia in abdominal cancers with peritoneal carcinomatosis: a phase II study. J Clin Oncol 2003; 21: 799-806
- Elias D, Blot F, El Otmany A, Antoun S, Lasser P, Boige V, Rougier P, Ducreux M. Curative treatment of peritoneal carcinomatosis arising from colorectal cancer by complete resection and intraperitoneal chemotherapy. *Cancer* 2001; 92: 71-76
- 14 **Glehen O**, Gilly FN, Sugarbaker PH. New perspectives in the management of colorectal cancer: what about peritoneal carcinomatosis? *Scand J Surg* 2003; **92**: 178-179
- 15 Shen P, Levine EA, Hall J, Case D, Russell G, Fleming R, McQuellon R, Geisinger KR, Loggie BW. Factors predicting survival after intraperitoneal hyperthermic chemotherapy with mitomycin C after cytoreductive surgery for patients with peritoneal carcinomatosis. Arch Surg 2003; 138: 26-33
- 16 Pestieau SR, Sugarbaker PH. Treatment of primary colon cancer with peritoneal carcinomatosis: comparison of concomitant vs. delayed management. Dis Colon Rectum 2000; 43: 1341-1346; discussion 1347-1348
- 17 **Witkamp AJ**, de Bree E, Kaag MM, Boot H, Beijnen JH, van Slooten GW, van Coevorden F, Zoetmulder FA. Extensive cytoreductive surgery followed by intra-operative hyperthermic intraperitoneal chemotherapy with mitomycin-C in patients with peritoneal carcinomatosis of colorectal origin. *Eur J Cancer* 2001; **37**: 979-984
- 18 Verwaal VJ, van Ruth S, de Bree E, van Sloothen GW, van Tinteren H, Boot H, Zoetmulder FA. Randomized

- trial of cytoreduction and hyperthermic intraperitoneal chemotherapy versus systemic chemotherapy and palliative surgery in patients with peritoneal carcinomatosis of colorectal cancer. *J Clin Oncol* 2003; 21: 3737-3743
- 19 Glehen O, Kwiatkowski F, Sugarbaker PH, Elias D, Levine EA, De Simone M, Barone R, Yonemura Y, Cavaliere F, Quenet F, Gutman M, Tentes AA, Lorimier G, Bernard JL, Bereder JM, Porcheron J, Gomez-Portilla A, Shen P, Deraco M, Rat P. Cytoreductive surgery combined with perioperative intraperitoneal chemotherapy for the management of peritoneal carcinomatosis from colorectal cancer: a multi-institutional study. J Clin Oncol 2004; 22: 3284-3292
- 20 Yan TD, Black D, Sugarbaker PH, Zhu J, Yonemura Y, Petrou G, Morris DL. A systematic review and meta-analysis of the randomized controlled trials on adjuvant intraperitoneal chemotherapy for resectable gastric cancer. *Ann Surg Oncol* 2007; 14: 2702-2713
- 21 **Bijelic L**, Jonson A, Sugarbaker PH. Systematic review of cytoreductive surgery and heated intraoperative intraperitoneal chemotherapy for treatment of peritoneal carcinomatosis in primary and recurrent ovarian cancer. *Ann Oncol* 2007; **18**: 1943-1950
- Yan TD, Welch L, Black D, Sugarbaker PH. A systematic review on the efficacy of cytoreductive surgery combined with perioperative intraperitoneal chemotherapy for diffuse malignancy peritoneal mesothelioma. *Ann Oncol* 2007; 18: 827-834
- Yan TD, Black D, Savady R, Sugarbaker PH. A systematic review on the efficacy of cytoreductive surgery and perioperative intraperitoneal chemotherapy for pseudomyxoma peritonei. Ann Surg Oncol 2007; 14: 484-492
- 24 Yan TD, Black D, Savady R, Sugarbaker PH. Systematic review on the efficacy of cytoreductive surgery combined with perioperative intraperitoneal chemotherapy for peritoneal carcinomatosis from colorectal carcinoma. *J Clin Oncol* 2006; 24: 4011-4019
- 25 **Baron MA**. Structure of the intestinal peritoneum in man. *Am J Anat* 1941; **69**: 439-497
- 26 Dobbie JW. Ultrastructure and pathology of the peritoneum in peritoneal dialysis. In: Gokal R, Nolph KD, editors. Textbook of peritoneal dialysis. Dordrecht: Kluwer Academic, 1994: 17-45
- 27 Sugarbaker PH. Peritoneum as the first-line of defense in carcinomatosis. J Surg Oncol 2007; 95: 93-96
- 28 Oosterling SJ, van der Bij GJ, van Egmond M, van der Sijp JR. Surgical trauma and peritoneal recurrence of colorectal carcinoma. Eur J Surg Oncol 2005; 31: 29-37
- 29 Flessner MF, Henegar J, Bigler S, Genous L. Is the peritoneum a significant transport barrier in peritoneal dialysis? *Perit Dial Int* 2003; 23: 542-549
- 30 de Lima Vazquez V, Stuart OA, Mohamed F, Sugarbaker PH. Extent of parietal peritonectomy does not change intraperitoneal chemotherapy pharmacokinetics. Cancer Chemother Pharmacol 2003; 52: 108-112
- 31 Jacquet P, Averbach A, Stephens AD, Stuart OA, Chang D, Sugarbaker PH. Heated intraoperative intraperitoneal mitomycin C and early postoperative intraperitoneal 5-fluorouracil: pharmacokinetic studies. Oncology 1998; 55: 130-138
- 32 **Stelin G**, Rippe B. A phenomenological interpretation of the variation in dialysate volume with dwell time in CAPD. *Kidney Int* 1990; **38**: 465-472
- 33 Flessner MF. The transport barrier in intraperitoneal therapy. Am J Physiol Renal Physiol 2005; 288: F433-F442
- 34 Dedrick RL, Myers CE, Bungay PM, DeVita VT Jr. Pharmacokinetic rationale for peritoneal drug administration in the treatment of ovarian cancer. Cancer Treat Rep 1978; 62: 1-11
- 35 Sugarbaker PH, Mora JT, Carmignani P, Stuart OA, Yoo D. Update on chemotherapeutic agents utilized for perioperative intraperitoneal chemotherapy. *Oncologist* 2005; 10: 112-122
- 36 **Dedrick RL**, Flessner MF. Pharmacokinetic problems in



- peritoneal drug administration: tissue penetration and surface exposure. J Natl Cancer Inst 1997; 89: 480-487
- 37 Dedrick RL, Flessner MF, Collins JM, Schultz JS. Is the peritoneum a membrane? Am Soc Artificial Internal Organs J 1982; 5: 1-8
- 38 Los G, Mutsaers PH, Lenglet WJ, Baldew GS, McVie JG. Platinum distribution in intraperitoneal tumors after intraperitoneal cisplatin treatment. Cancer Chemother Pharmacol 1990; 25: 389-394
- 39 Los G, Verdegaal EM, Mutsaers PH, McVie JG. Penetration of carboplatin and cisplatin into rat peritoneal tumor nodules after intraperitoneal chemotherapy. *Cancer Chemother Pharmacol* 1991; 28: 159-165
- 40 Ozols RF, Locker GY, Doroshow JH, Grotzinger KR, Myers CE, Young RC. Pharmacokinetics of adriamycin and tissue penetration in murine ovarian cancer. *Cancer Res* 1979; 39: 3209-3214
- 41 Sticca RP, Dach BW. Rationale for hyperthermia with intraoperative intraperitoneal chemotherapy agents. Surg Oncol Clin N Am 2003; 12: 689-701
- 42 **Dahl O**, Dalene R, Schem BC, Mella O. Status of clinical hyperthermia. *Acta Oncol* 1999; **38**: 863-873
- 43 **Lepock JR**. How do cells respond to their thermal environment? *Int J Hyperthermia* 2005; **21**: 681-687
- 44 **Hahn GM**, Braun J, Har-Kedar I. Thermochemotherapy: synergism between hyperthermia (42-43 degrees) and adriamycin (of bleomycin) in mammalian cell inactivation. *Proc Natl Acad Sci USA* 1975; **72**: 937-940
- 45 Kusumoto T, Holden SA, Ara G, Teicher BA. Hyperthermia and platinum complexes: time between treatments and synergy in vitro and in vivo. *Int J Hyperthermia* 1995; 11: 575-586
- 46 Barlogie B, Corry PM, Drewinko B. In vitro thermochemotherapy of human colon cancer cells with cis-dichlorodiam-mineplatinum(II) and mitomycin C. Cancer Res 1980; 40: 1165-1168
- 47 Urano M, Ling CC. Thermal enhancement of melphalan and oxaliplatin cytotoxicity in vitro. *Int J Hyperthermia* 2002; 18: 307-315
- 48 Mohamed F, Marchettini P, Stuart OA, Urano M, Sugarbaker PH. Thermal enhancement of new chemotherapeutic agents at moderate hyperthermia. Ann Surg Oncol 2003; 10: 463-468
- 49 Benoit L, Duvillard C, Rat P, Chauffert B. [The effect of intraabdominal temperature on the tissue and tumor diffusion of intraperitoneal cisplatin in a model of peritoneal carcinomatosis in rats] Chirurgie 1999; 124: 375-379
- 50 Jacquet P, Averbach A, Stuart OA, Chang D, Sugarbaker PH. Hyperthermic intraperitoneal doxorubicin: pharmacokinetics, metabolism, and tissue distribution in a rat model. Cancer Chemother Pharmacol 1998; 41: 147-154
- 51 Esquis P, Consolo D, Magnin G, Pointaire P, Moretto P, Ynsa MD, Beltramo JL, Drogoul C, Simonet M, Benoit L, Rat P, Chauffert B. High intra-abdominal pressure enhances the penetration and antitumor effect of intraperitoneal cisplatin on experimental peritoneal carcinomatosis. *Ann Surg* 2006; 244: 106-112
- 52 Jacquet P, Stuart OA, Chang D, Sugarbaker PH. Effects of intra-abdominal pressure on pharmacokinetics and tissue distribution of doxorubicin after intraperitoneal administration. Anticancer Drugs 1996; 7: 596-603
- 53 Garofalo A, Valle M, Garcia J, Sugarbaker PH. Laparoscopic intraperitoneal hyperthermic chemotherapy for palliation of debilitating malignant ascites. Eur J Surg Oncol 2006; 32: 682-685
- 54 Facchiano E, Scaringi S, Kianmanesh R, Sabate JM, Castel B, Flamant Y, Coffin B, Msika S. Laparoscopic hyperthermic intraperitoneal chemotherapy (HIPEC) for the treatment of malignant ascites secondary to unresectable peritoneal carcinomatosis from advanced gastric cancer. Eur J Surg Oncol 2008; 34: 154-158
- 55 Pestieau SR, Schnake KJ, Stuart OA, Sugarbaker PH. Impact

- of carrier solutions on pharmacokinetics of intraperitoneal chemotherapy. Cancer Chemother Pharmacol 2001; 47: 269-276
- 56 Mohamed F, Marchettini P, Stuart OA, Sugarbaker PH. Pharmacokinetics and tissue distribution of intraperitoneal paclitaxel with different carrier solutions. Cancer Chemother Pharmacol 2003; 52: 405-410
- 57 Mohamed F, Sugarbaker PH. Carrier solutions for intraperitoneal chemotherapy. Surg Oncol Clin N Am 2003; 12: 813-824
- 58 Elias D, El Otmany A, Bonnay M, Paci A, Ducreux M, Antoun S, Lasser P, Laurent S, Bourget P. Human pharmacokinetic study of heated intraperitoneal oxaliplatin in increasingly hypotonic solutions after complete resection of peritoneal carcinomatosis. Oncology 2002; 63: 346-352
- 59 Keshaviah P, Emerson PF, Vonesh EF, Brandes JC. Relationship between body size, fill volume, and mass transfer area coefficient in peritoneal dialysis. J Am Soc Nephrol 1994; 4: 1820-1826
- 60 Elias DM, Sideris L. Pharmacokinetics of heated intraoperative intraperitoneal oxaliplatin after complete resection of peritoneal carcinomatosis. Surg Oncol Clin N Am 2003; 12: 755-769, xiv
- 61 **Sugarbaker PH**, Stuart OA, Carmignani CP. Pharmacokinetic changes induced by the volume of chemotherapy solution in patients treated with hyperthermic intraperitoneal mitomycin C. *Cancer Chemother Pharmacol* 2006; **57**: 703-708
- 62 Chauffert B, Favoulet P, Polycarpe E, Duvillard C, Beltramo JL, Bichat F, Rat P, Genne P, Benoit L. Rationale supporting the use of vasoconstrictors for intraperitoneal chemotherapy with platinum derivatives. Surg Oncol Clin N Am 2003; 12: 835-848
- 63 Ensminger WD, Gyves JW. Regional chemotherapy of neoplastic diseases. *Pharmacol Ther* 1983; 21: 277-293
- 64 Favoulet P, Magnin G, Guilland JC, Beltramo JL, Osmak L, Benoit L, Rat P, Douvier S, Duvillard C, Chauffert B. Preclinical study of the epinephrine-cisplatin association for the treatment of intraperitoneal carcinomatosis. *Eur J Surg Oncol* 2001; 27: 59-64
- Molucon-Chabrot C, Isambert N, Benoit L, Zanetta S, Fraisse J, Guilland JC, Royer B, Monin-Baroille P, Flesch M, Fargeot P, Coudert B, Mayer F, Fumoleau P, Chauffert B. Feasibility of using intraperitoneal epinephrine and cisplatin in patients with advanced peritoneal carcinomatosis. Anticancer Drugs 2006; 17: 1211-1217
- 66 Lindnér P, Heath D, Howell S, Naredi P, Hafström L. Vasopressin modulation of peritoneal, lymphatic, and plasma drug exposure following intraperitoneal administration. Clin Cancer Res 1996; 2: 311-317
- 67 Rubin J, Clawson M, Planch A, Jones Q. Measurements of peritoneal surface area in man and rat. Am J Med Sci 1988; 295: 453-458
- 68 Chagnac A, Herskovitz P, Weinstein T, Elyashiv S, Hirsh J, Hammel I, Gafter U. The peritoneal membrane in peritoneal dialysis patients: estimation of its functional surface area by applying stereologic methods to computerized tomography scans. J Am Soc Nephrol 1999; 10: 342-346
- 69 Rossi CR, Deraco M, De Simone M, Mocellin S, Pilati P, Foletto M, Cavaliere F, Kusamura S, Gronchi A, Lise M. Hyperthermic intraperitoneal intraoperative chemotherapy after cytoreductive surgery for the treatment of abdominal sarcomatosis: clinical outcome and prognostic factors in 60 consecutive patients. *Cancer* 2004; 100: 1943-1950
- 70 Baratti D, Kusamura S, Martinetti A, Seregni E, Laterza B, Oliva DG, Deraco M. Prognostic value of circulating tumor markers in patients with pseudomyxoma peritonei treated with cytoreductive surgery and hyperthermic intraperitoneal chemotherapy. Ann Surg Oncol 2007; 14: 2300-2308
- 71 Glehen O, Schreiber V, Cotte E, Sayag-Beaujard AC, Osinsky D, Freyer G, François Y, Vignal J, Gilly FN. Cytoreductive surgery and intraperitoneal chemohyperthermia for peritoneal carcinomatosis arising from gastric cancer. Arch Surg



- 2004; 139: 20-26
- 72 **Gilly FN**, Carry PY, Sayag AC, Brachet A, Panteix G, Salle B, Bienvenu J, Burgard G, Guibert B, Banssillon V. Regional chemotherapy (with mitomycin C) and intra-operative hyperthermia for digestive cancers with peritoneal carcinomatosis. *Hepatogastroenterology* 1994; **41**: 124-129
- 73 Fujimoto S, Shrestha RD, Kokubun M, Kobayashi K, Kiuchi S, Konno C, Ohta M, Takahashi M, Kitsukawa Y, Mizutani M. Positive results of combined therapy of surgery and intraperitoneal hyperthermic perfusion for far-advanced gastric cancer. *Ann Surg* 1990; 212: 592-596
- 74 **Fujimura** T, Yonemura Y, Fushida S, Urade M, Takegawa S, Kamata T, Sugiyama K, Hasegawa H, Katayama K, Miwa K. Continuous hyperthermic peritoneal perfusion for the treatment of peritoneal dissemination in gastric cancers and subsequent second-look operation. *Cancer* 1990; **65**: 65-71
- 75 **Elias D**, Antoun S, Raynard B, Puizillout JM, Sabourin JC, Ducreux M, Lasser P. [Treatment of peritoneal carcinomatosis using complete excision and intraperitoneal chemohyperthermia. A phase I-II study defining the best technical procedures] *Chirurgie* 1999; **124**: 380-389
- 76 Elias D, Antoun S, Goharin A, Otmany AE, Puizillout JM, Lasser P. Research on the best chemohyperthermia technique of treatment of peritoneal carcinomatosis after complete resection. *Int J Surg Investig* 2000; 1: 431-439
- 77 Sugarbaker PH. An instrument to provide containment of intraoperative intraperitoneal chemotherapy with optimized distribution. J Surg Oncol 2005; 92: 142-146
- 78 Guerbet M, Goullé JP, Lubrano J. Evaluation of the risk of contamination of surgical personnel by vaporization of oxaliplatin during the intraoperative hyperthermic intraperitoneal chemotherapy (HIPEC). Eur J Surg Oncol 2007; 33: 623-626
- 79 Stuart OA, Stephens AD, Welch L, Sugarbaker PH. Safety monitoring of the coliseum technique for heated intraoperative intraperitoneal chemotherapy with mitomycin C. Ann Surg Oncol 2002; 9: 186-191
- 80 **Schmid K**, Boettcher MI, Pelz JO, Meyer T, Korinth G, Angerer J, Drexler H. Investigations on safety of hyperthermic intraoperative intraperitoneal chemotherapy (HIPEC) with Mitomycin C. *Eur J Surg Oncol* 2006; **32**: 1222-1225
- 81 Yonemura Y, Bandou E, Sawa T, Yoshimitsu Y, Endou Y, Sasaki T, Sugarbaker PH. Neoadjuvant treatment of gastric cancer with peritoneal dissemination. Eur J Surg Oncol 2006; 32: 661-665
- 82 Sugarbaker PH. Treatment of peritoneal carcinomatosis from colon or appendiceal cancer with induction intraperitoneal chemotherapy. Cancer Treat Res 1996; 82: 317-325
- 83 **Zylberberg B**, Dormont D, Janklewicz S, Darai E, Bretel JJ, Poncelet C, Guillet JL, Madelenat P. Response to neo-adjuvant intraperitoneal and intravenous immunochemotherapy followed by interval secondary cytoreduction in stage IIIc ovarian cancer. Eur J Gynaecol Oncol 2001; **22**: 40-45
- 84 **Esquivel J**, Vidal-Jove J, Steves MA, Sugarbaker PH. Morbidity and mortality of cytoreductive surgery and intraperitoneal chemotherapy. *Surgery* 1993; **113**: 631-636
- 85 Sugarbaker PH, Graves T, DeBruijn EA, Cunliffe WJ, Mullins RE, Hull WE, Oliff L, Schlag P. Early postoperative intraperitoneal chemotherapy as an adjuvant therapy to

- surgery for peritoneal carcinomatosis from gastrointestinal cancer: pharmacological studies. *Cancer Res* 1990; **50**: 5790-5794
- Markman M, Bundy BN, Alberts DS, Fowler JM, Clark-Pearson DL, Carson LF, Wadler S, Sickel J. Phase III trial of standard-dose intravenous cisplatin plus paclitaxel versus moderately high-dose carboplatin followed by intravenous paclitaxel and intraperitoneal cisplatin in small-volume stage III ovarian carcinoma: an intergroup study of the Gynecologic Oncology Group, Southwestern Oncology Group, and Eastern Cooperative Oncology Group. J Clin Oncol 2001; 19: 1001-1007
- 87 Alberts DS, Liu PY, Hannigan EV, O'Toole R, Williams SD, Young JA, Franklin EW, Clarke-Pearson DL, Malviya VK, DuBeshter B. Intraperitoneal cisplatin plus intravenous cyclophosphamide versus intravenous cisplatin plus intravenous cyclophosphamide for stage III ovarian cancer. N Engl J Med 1996; 335: 1950-1955
- 88 Armstrong DK, Bundy B, Wenzel L, Huang HQ, Baergen R, Lele S, Copeland LJ, Walker JL, Burger RA. Intraperitoneal cisplatin and paclitaxel in ovarian cancer. N Engl J Med 2006; 354: 34-43
- 89 **Yan TD**, Bijelic L, Sugarbaker PH. Critical analysis of treatment failure after complete cytoreductive surgery and perioperative intraperitoneal chemotherapy for peritoneal dissemination from appendiceal mucinous neoplasms. *Ann Surg Oncol* 2007; **14**: 2289-2299
- 90 Bijelic L, Yan TD, Sugarbaker PH. Failure analysis of recurrent disease following complete cytoreduction and perioperative intraperitoneal chemotherapy in patients with peritoneal carcinomatosis from colorectal cancer. *Ann Surg* Oncol 2007; 14: 2281-2288
- 91 Rippe B, Haraldsson B. Transport of macromolecules across microvascular walls: the two-pore theory. *Physiol Rev* 1994; 74: 163-219
- 92 **Yuan F**, Dellian M, Fukumura D, Leunig M, Berk DA, Torchilin VP, Jain RK. Vascular permeability in a human tumor xenograft: molecular size dependence and cutoff size. *Cancer Res* 1995; **55**: 3752-3756
- 93 **Flessner MF**, Choi J, Credit K, Deverkadra R, Henderson K. Resistance of tumor interstitial pressure to the penetration of intraperitoneally delivered antibodies into metastatic ovarian tumors. *Clin Cancer Res* 2005; **11**: 3117-3125
- 94 Nygren P, Fridborg H, Csoka K, Sundström C, de la Torre M, Kristensen J, Bergh J, Hagberg H, Glimelius B, Rastad J. Detection of tumor-specific cytotoxic drug activity in vitro using the fluorometric microculture cytotoxicity assay and primary cultures of tumor cells from patients. *Int J Cancer* 1994; 56: 715-720
- 95 Larsson R, Kristensen J, Sandberg C, Nygren P. Laboratory determination of chemotherapeutic drug resistance in tumor cells from patients with leukemia, using a fluorometric microculture cytotoxicity assay (FMCA). *Int J Cancer* 1992; 50: 177-185
- 96 Mahteme H, von Heideman A, Grundmark B, Tholander B, Påhlman L, Glimelius B, Larsson R, Graf W, Nygren P. Heterogeneous activity of cytotoxic drugs in patient samples of peritoneal carcinomatosis. *Eur J Surg Oncol* 2008; 34: 547-552

S- Editor Li LF L- Editor Stewart GJ E- Editor Lin YP

